# Large scale differential expression analysis identifies genes associated with Bipolar Disorder

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## Abstract

### Background and purpose

Bipolar disorder (BD) remains one of the most common psychiatric disorders with high morbidity and mortality. Several polymorphisms have been found to be implicated in the pathogenesis of BD, however, these loci have small effect sizes that fail to explain the high heritability of the disease. Here, we attempt to better understand the genetic basis of bipolar disorder by identifying differentially expressed genes in post-mortem brain tissues of patients with BD versus normal controls.

### Methods and results

A total of 9 gene expression data sets with 702 samples derived from post-mortem brain were available for differential expression analysis. 7 data sets were used for the discovery of the gene signature and the other two were left for independent validation. The multi-cohort analysis was performed using a random-effects model and resulted in the identification of 13 up-regulated and 9 down-regulated genes in BD. The BD gene signature was validated in two independent data sets resulting in an Area Under the ROC Curve (AUC) of 0.74

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and 0.73 respectively and Area Under the Precision Recall Curve (AUPRC) of 0.72 and 0.66 respectively. Gene set enrichment analysis was performed and resulted in the identification of several biological processes and pathways related to BD including Ca transport, inflammation and DNA damage response.

## Conclusion

Our findings support the previous findings that link BD pathogenesis to abnormalities in glial inflammation and calcium transport and also identify several other biological processes not previously reported to be associated with the development of BD. Such findings will improve our understanding of the genetic basis underlying BD and may have future clinical implications.

**Keywords**— Bipolar Disorder, differential gene expression, inflammation, calcium transport

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## 1  Introduction

Bipolar disorder (BD) is a common psychiatric disorder characterized by alternating periods of depression and abnormally elevated mood associated with changes in the activity levels [1]. The lifetime prevalence of Bipolar disorder is approximately 2.4% with average age of onset of 25 years [2].

BD is a highly heritable disease with overall heritability reaching up to 70% based on twin studies [3, 4] with the risk being 10-fold higher in the first degree relatives of those affected with BD when compared with normal population. The biological mechanisms underlying the etiology of bipolar disorder remain unclear with recent evidence pointing toward a significant role of disrupted Ca dynamics and oxidative DNA damage in the pathogenesis of BD [5, 6, 7]. Although several susceptibility loci with small effect sizes have been found to be associated with BD [8, 9], there is no much evidence supporting the role of specific genes in the pathogenesis of the disease [10].

In this study, we attempt to identify the genes and biological pathways associated with bipolar disorder. For this purpose, we analyze microarray gene expression data derived from post-mortem brain tissue to identify the deferentially expressed genes between BD and normal cases. Detecting those genes and pathways would provide us with a better understanding of the molecular pathogenesis of this disease and may have a future therapeutic implications.

## 2 Methods

### 2.1 Data collection

We performed an extensive search of the Gene Expression Omnibus (GEO) database [11]looking for microarray gene expression data of bipolar disorder. A total of 9 data sets were identified (table 1) with 702 post-mortem brain samples. We used 7 data sets for discovery of the BD gene signature and kept 3 data sets out for independent validation of the gene signature performance.

| GEO accession | Type of experiment | Number of samples (BD cases) | Tissue source |
|---|---|---|---|
| **Discovery** | | | |
| **GSE120340** | Expression profiling by array | 20 (10) | Dorsolateral prefrontal cortex |
| **GSE87610** | Expression profiling by array | 145 (73) | Dorsolateral prefrontal cortex |
| **GSE12654** | Expression profiling by array | 26 (11) | Anterior prefrontal cortex |
| **GSE12649** | Expression profiling by array | 67 (33) | Dorsolateral prefrontal cortex |
| **GSE62191** | Expression profiling by array | 59 (29) | Frontal cortex |
| **GSE92538-GPL10526** | Expression profiling by array | 68 (12) | Dorsolateral prefrontal cortex |
| **GSE92538-GPL17027** | Expression profiling by array | 161 (42) | Dorsolateral prefrontal cortex |
| **Validation** | | | |
| **GSE5388** | Expression profiling by array | 61 (30) | Dorsolateral prefrontal cortex |
| **GSE35977** | Expression profiling by array | 95 (45) | Parietal cortex |

Table 1: Collected data sets showing the number of samples with the number of bipolar disease (BD) cases. 7 data sets were used for discovery of the BD gene signature and 2 data sets were used for independent validation

## 2.2 The meta-analysis

We used R Programming language [12] and "Metaintegrator" package [13] which utilizes a gene expression meta-analysis workflow described by Haynes et al [14]. This approach computes a Hedges effect size [15]for each gene in each data set. These effect sizes are then pooled across all the data sets (Supplementary figures 1-2) using a random-effect model by assuming that results from each study is drawn from a single distribution and that each inter-study difference is just a random effect. The approach computes the log sum of p-values that each gene is up/down-regulated, then combine p-values using Fisher's method and finally performs Benjamini-Hochberg FDR correction across all genes [16].

A gene would be deferentially expressed if the absolute value of its effect size is $> 0$, false discovery rate is $<= 0.15$ and it has to be significantly up/down regulated in all of the 7 training data sets with a heterogeneity P-value cutoff of 0.05 [17]. All discovery steps were conducted on the training data sets only.

## 2.3 Validation of the resulting gene signature

The performance of the resulting BD gene signature was further validated in the 2 independent cohorts with a total of 156 samples. We used the Area Under the ROC curve (AUC), Area Under the Precision Recall Curve (AUPRC) and accuracy as evaluation metrics.

## 2.4 Enrichment analysis

Functional Enrichment analysis was performed using Enrichr R package [18, 19] to identify the Gene Ontology (GO) biological processes and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways enriched in the discovered genes.

# 3 Results

The meta-analysis process resulted in the identification of 13 up-regulated and 9 down-regulated genes in the post-mortem brain samples of bipolar disorder cases compared with normal controls (Table2).

| Gene Symbol | Effect size | Effect size p-value |
|---|---|---|
| | **Up-regulated genes** | |
| *MYO6* | 0.233803528781553 | 3.92088529079236e-05 |

| Gene Symbol | Effect size | Effect size p-value |
| --- | --- | --- |
| *ERCC8* | 0.223683432497003 | 5.06105158365377e-05 |
| *SLC12A4* | 0.153840135678242 | 0.000140523441674623 |
| *CA12* | 0.249039212602444 | 0.000169533685049415 |
| *ZNF273* | 0.287473913205269 | 0.000202310305620391 |
| *TP53* | 0.166740750366376 | 0.000301337862141146 |
| *MYRIP* | 0.240157780766432 | 0.000306767694381059 |
| *FGF2* | 0.230057248064757 | 0.000314051978487104 |
| *BCL2L11* | 0.14089492142396 | 0.000369520305484846 |
| *COL11A1* | 0.279394249891102 | 0.000763160058191395 |
| *PRKG1* | 0.194638754941504 | 0.000863840829673692 |
| *RRM1* | 0.197841881310803 | 0.000948698091142044 |
| *PAQR3* | 0.18648054393229 | 0.00097136180697817 |
| **Down-regulated genes** | | |
| *CD74* | -0.314560020628663 | 3.71498955514197e-07 |
| *CNP* | -0.231525261417171 | 4.41001367269943e-06 |
| *HLA-DPB1* | -0.183506935227843 | 1.17960248475937e-05 |
| *DDX17* | -0.318235420127414 | 2.63152971729961e-05 |
| *HLA-DPA1* | -0.268023572622133 | 5.76742550284089e-05 |
| *MRPL33* | -0.260414913702845 | 0.000125094044339841 |
| *CCKBR* | -0.327943659949979 | 0.000258927502648349 |
| *ACAT2* | -0.330687000537771 | 0.000359707958498527 |
| *CD53* | -0.359439343217732 | 0.000366805911760364 |

Table 2: BD signature genes. 13 up-regulated and 9 down-regulated genes

## 3.1 Strong performance in the two validation data sets

We measured the discriminatory ability of our gene signature by applying it to the 2 independent data sets to assess its performance on unseen data.

In the first validation data set (GSE5388), the signature achieved an AUC of 0.74, AUPRC of 0.72, accuracy of 74% and sensitivity of 90%. In the second validation data set (GSE35977), the signature achieved an AUC of 0.73, AUPRC of 0.664, accuracy of 68.4% and sensitivity of 89% (Figure 2).

Figure 1: ROC curves of the 2 validation data sets

The signature performance in the validation data sets showed great consistency with the performance in the discovery data sets without overfitting (Summary AUC = 0.74, 95% CI = 0.67-0.80) (figure**??**).

Figure 2: ROC curves of the 7 discovery data sets. Notice the summary AUC of 0.74 which is identical to the AUC in each of the validation data sets (no overfitting)
.

## 3.2 Enrichment analysis

Enrichment analysis has revealed that the discovered gene signature is incorporated in many significant pathways and biological processes. Several of these biological processes were previously reported to be associated with bipolar disorder and other psychiatric disease, for example: inflammation, apoptosis, regulation of DNA damage response and Ca transport. We confirm this association and report other pathways and biological processes not previously reported to be associated with BD.

# 4 Discussion

Bipolar disorder is a highly heterogeneous disease and the genetic basis underlying its pathophysiology is complex [20]. Several genome-wide association studies (GWAS) have identified several copy number variants (CNVs) and single-nucleotide polymorphisms (SNPs)

that seem to play a role in the high heritability of the disease but each of these variants has a small effect [21, 22]. Moreover, these studies failed to identify specific genes that may have a large effect in the pathogenesis of the disease, most likely due to small sample size.

Here we tried to overcome the problem of inadequate sample size by incorporating multiple gene expression studies in a meta-analysis process to achieve high statistical power. Through this large scale gene expression analysis of 702 post-mortem brain samples, we managed to identify 13 up-regulated and 9 down-regulated genes in BD. The resulting gene signature managed to achieve great performance in the two independent validation data sets. In the first validation cohort, the signature achieved an AUC of 0.74 and AUPRC of 0.72. In the second validation cohort, the signature achieved an AUC of 0.73 and AUPRC of 0.664.

The up-regulated genes were enriched in many biological processes (Supplementary Table 1) like Ca ion transport, cytoskeletal assembly, apoptotic signaling, intracellular signal transduction, cell migration, inflammation and response to DNA damage. Several of these biological processes have been reported to be associated with BD and other psychiatric disorders like Schizophrenia [23, 24, 25, 26]. These same processes are incorporated in the pathogenesis of atherosclerosis which may explain part of the association between BD and other cardiovascular diseases [27, 28, 29].

The 9 down-regulated genes were enriched in important biological processes and pathways (Supplementary Table 2) like the positive regulation of lymphocyte proliferation, antigen processing/presentation and regulation of macrophages cytokines production supporting the notion that neuronal and glial inflammation may have a role in the pathogenesis of BD.

## 5 Conclusion

In this study, we provide more insights into the genetic basis of BD through the identification of DE genes in post-mortem brain tissues of BD patients compared with normal controls. The DE genes were enriched in a number of biological processes and pathways that were previously reported to have a role in the pathogenesis of BD, most importantly the pathways regulating Ca transport and inflammatory processes. Such findings, together with previous research will provide a more clear understanding of the molecular pathophysiology of BD and may have future therapeutic potentials.

## References

[1] Ian M Anderson, Peter M Haddad, and Jan Scott. Bipolar disorder. *BMJ*, 345, 2012.

[2] Lakshmi N Yatham, Sidney H Kennedy, Sagar V Parikh, Ayal Schaffer, David J Bond, Benicio N Frey, Verinder Sharma, Benjamin I Goldstein, Soham Rej, Serge

Beaulieu, Martin Alda, Glenda MacQueen, Roumen V Milev, Arun Ravindran, Claire O'Donovan, Diane McIntosh, Raymond W Lam, Gustavo Vazquez, Flavio Kapczinski, Roger S McIntyre, Jan Kozicky, Shigenobu Kanba, Beny Lafer, Trisha Suppes, Joseph R Calabrese, Eduard Vieta, Gin Malhi, Robert M Post, and Michael Berk. Canadian network for mood and anxiety treatments (canmat) and international society for bipolar disorders (isbd) 2018 guidelines for the management of patients with bipolar disorder. *Bipolar Disord*, 20(2):97–170, Mar 2018.

[3] Jack Edvardsen, Svenn Torgersen, Espen Roysamb, Sissel Lygren, Ingunn Skre, Sidsel Onstad, and Per Anders Oien. Heritability of bipolar spectrum disorders. unity or heterogeneity? *J Affect Disord*, 106(3):229–240, Mar 2008.

[4] Nick Craddock and Pamela Sklar. Genetics of bipolar disorder. *Lancet*, 381(9878):1654–1662, May 2013.

[5] Michael J Berridge. Calcium signalling and psychiatric disease: bipolar disorder and schizophrenia. *Cell Tissue Res*, 357(2):477–492, Aug 2014.

[6] Xiaoyun Guo, Dengtang Liu, Tong Wang, and Xingguang Luo. Aetiology of bipolar disorder: contribution of the l-type voltage-gated calcium channels. *Gen Psychiatr*, 32(2):e100009, 2019.

[7] Klaus Munkholm, Henrik Enghusen Poulsen, Lars Vedel Kessing, and Maj Vinberg. Elevated levels of urinary markers of oxidatively generated dna and rna damage in bipolar disorder. *Bipolar Disord*, 17(3):257–268, May 2015.

[8] Seth A Ament, Szabolcs Szelinger, Gustavo Glusman, Justin Ashworth, Liping Hou, Nirmala Akula, Tatyana Shekhtman, Judith A Badner, Mary E Brunkow, Denise E Mauldin, Anna-Barbara Stittrich, Katherine Rouleau, Sevilla D Detera-Wadleigh, John I Jr Nurnberger, Howard J Edenberg, Elliot S Gershon, Nicholas Schork, Nathan D Price, Richard Gelinas, Leroy Hood, David Craig, Francis J McMahon, John R Kelsoe, and Jared C Roach. Rare variants in neuronal excitability genes influence risk for bipolar disorder. *Proc Natl Acad Sci U S A*, 112(11):3576–3581, Mar 2015.

[9] M Kataoka, N Matoba, T Sawada, A-A Kazuno, M Ishiwata, K Fujii, K Matsuo, A Takata, and T Kato. Exome sequencing for bipolar disorder points to roles of de novo loss-of-function and protein-altering mutations. *Mol Psychiatry*, 21(7):885–893, Jul 2016.

[10] Michael C O'Donovan and Michael J Owen. The implications of the shared genetics of psychiatric disorders. *Nat Med*, 22(11):1214–1219, Nov 2016.

[11] Tanya Barrett, Stephen E. Wilhite, Pierre Ledoux, Carlos Evangelista, Irene F. Kim, Maxim Tomashevsky, Kimberly A. Marshall, Katherine H. Phillippy, Patti M. Sherman, Michelle Holko, Andrey Yefanov, Hyeseung Lee, Naigong Zhang, Cynthia L. Robertson, Nadezhda Serova, Sean Davis, and Alexandra Soboleva. NCBI GEO: archive for functional genomics data sets—update. *Nucleic Acids Research*, 41(D1):D991–D995, 11 2012.

[12] Ross Ihaka and Robert Gentleman. R: A language for data analysis and graphics. 5(3):299–314, 1996.

[13] Purvesh Khatri, Silke Roedder, Naoyuki Kimura, Katrien De Vusser, Alexander A. Morgan, Yongquan Gong, Michael P. Fischbein, Robert C. Robbins, Maarten Naesens, Atul J. Butte, and Minnie M. Sarwal. A common rejection module (crm) for acute rejection across multiple organs identifies novel therapeutics for organ transplantation. *Journal of Experimental Medicine*, 210(11):2205–2221, 2013.

[14] Winston A Haynes, Francesco Vallania, Charles Liu, Erika Bongen, Aurelie Tomczak, Marta Andres-Terrè, Shane Lofgren, Andrew Tam, Cole A Deisseroth, Matthew D Li, Timothy E Sweeney, and Purvesh Khatri. Empowering multi-cohort gene expression analysis to increase reproducibility. *Pacific Symposium on Biocomputing. Pacific Symposium on Biocomputing*, 22:144–153, 2017.

[15] Larry V. Hedges. Distribution theory for glass's estimator of effect size and related estimators. *Journal of Educational Statistics*, 6(2):107–128, 1981.

[16] Yoav Benjamini and Yosef Hochberg. Controlling the false discovery rate: A practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society: Series B (Methodological)*, 57(1):289–300, 2019/07/24 1995.

[17] David C. Hoaglin. Misunderstandings about q and 'cochran's q test' in meta-analysis. *Statistics in Medicine*, 35(4):485–495, 2016.

[18] Maxim V Kuleshov, Matthew R Jones, Andrew D Rouillard, Nicolas F Fernandez, Qiaonan Duan, Zichen Wang, Simon Koplev, Sherry L Jenkins, Kathleen M Jagodnik, Alexander Lachmann, Michael G McDermott, Caroline D Monteiro, Gregory W Gundersen, and Avi Ma'ayan. Enrichr: a comprehensive gene set enrichment analysis web server 2016 update. *Nucleic Acids Res*, 44(W1):W90–7, Jul 2016.

[19] Edward Y Chen, Christopher M Tan, Yan Kou, Qiaonan Duan, Zichen Wang, Gabriela Vaz Meirelles, Neil R Clark, and Avi Ma'ayan. Enrichr: interactive and collaborative html5 gene list enrichment analysis tool. *BMC Bioinformatics*, 14:128, Apr 2013.

[20] Nick Craddock and Pamela Sklar. Genetics of bipolar disorder. *Lancet*, 381(9878):1654–1662, May 2013.

[21] Liping Hou, Sarah E Bergen, Nirmala Akula, Jie Song, Christina M Hultman, Mikael Landén, Mazda Adli, Martin Alda, Raffaella Ardau, Bárbara Arias, Jean-Michel Aubry, Lena Backlund, Judith A Badner, Thomas B Barrett, Michael Bauer, Bernhard T Baune, Frank Bellivier, Antonio Benabarre, Susanne Bengesser, Wade H Berrettini, Abesh Kumar Bhattacharjee, Joanna M Biernacka, Armin Birner, Cinnamon S Bloss, Clara Brichant-Petitjean, Elise T Bui, William Byerley, Pablo Cervantes, Caterina Chillotti, Sven Cichon, Francesc Colom, William Coryell, David W Craig, Cristiana Cruceanu, Piotr M Czerski, Tony Davis, Alexandre Dayer, Franziska Degenhardt, Maria Del Zompo, J Raymond DePaulo, Howard J Edenberg, Bruno Étain, Peter Falkai, Tatiana Foroud, Andreas J Forstner, Louise Frisén, Mark A Frye, Janice M Fullerton, Sébastien Gard, Julie S Garnham, Elliot S Gershon, Fernando S Goes, Tiffany A Greenwood, Maria Grigoroiu-Serbanescu, Joanna Hauser, Urs Heilbronner, Stefanie Heilmann-Heimbach, Stefan Herms, Maria Hipolito, Shashi Hitturlingappa, Per Hoffmann, Andrea Hofmann, Stephane Jamain, Esther Jiménez, Jean-Pierre Kahn, Layla Kassem, John R Kelsoe, Sarah Kittel-Schneider, Sebastian Kliwicki, Daniel L Koller, Barbara König, Nina Lackner, Gonzalo Laje, Maren Lang, Catharina Lavebratt, William B Lawson, Marion Leboyer, Susan G Leckband, Chunyu Liu, Anna Maaser, Pamela B Mahon, Wolfgang Maier, Mario Maj, Mirko Manchia, Lina Martinsson, Michael J McCarthy, Susan L McElroy, Melvin G McInnis, Rebecca McKinney, Philip B Mitchell, Marina Mitjans, Francis M Mondimore, Palmiero Monteleone, Thomas W Mühleisen, Caroline M Nievergelt, Markus M Nöthen, Tomas Novák, John I Jr Nurnberger, Evaristus A Nwulia, Urban Ösby, Andrea Pfennig, James B Potash, Peter Propping, Andreas Reif, Eva Reininghaus, John Rice, Marcella Rietschel, Guy A Rouleau, Janusz K Rybakowski, Martin Schalling, William A Scheftner, Peter R Schofield, Nicholas J Schork, Thomas G Schulze, Johannes Schumacher, Barbara W Schweizer, Giovanni Severino, Tatyana Shekhtman, Paul D Shilling, Christian Simhandl, Claire M Slaney, Erin N Smith, Alessio Squassina, Thomas Stamm, Pavla Stopkova, Fabian Streit, Jana Strohmaier, Szabolcs Szelinger, Sarah K Tighe, Alfonso Tortorella, Gustavo Turecki, Eduard Vieta, Julia Volkert, Stephanie H Witt, Adam Wright, Peter P Zandi, Peng Zhang, Sebastian Zollner, and Francis J McMahon. Genome-wide association study of 40,000 individuals identifies two novel loci associated with bipolar disorder. *Hum Mol Genet*, 25(15):3383–3394, Aug 2016.

[22] Consortium Cross-Disorder Group of the Psychiatric Genomics. Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *Lancet*, 381(9875):1371–1379, Apr 2013.

[23] F M Benes, D Matzilevich, R E Burke, and J Walsh. The expression of proapoptosis genes is increased in bipolar disorder, but not in schizophrenia. *Mol Psychiatry*, 11(3):241–251, Mar 2006.

[24] Christopher J. Carter. EIF2B and Oligodendrocyte Survival: Where Nature and Nurture Meet in Bipolar Disorder and Schizophrenia? *Schizophrenia Bulletin*, 33(6):1343–1353, 02 2007.

[25] C J Carter. Multiple genes and factors associated with bipolar disorder converge on growth factor and stress activated kinase pathways controlling translation initiation: implications for oligodendrocyte viability. *Neurochem Int*, 50(3):461–490, Feb 2007.

[26] Roos C Padmos, Manon H J Hillegers, Esther M Knijff, Ronald Vonk, Anne Bouvy, Frank J T Staal, Dick de Ridder, Ralph W Kupka, Willem A Nolen, and Hemmo A Drexhage. A discriminating messenger rna signature for bipolar disorder formed by an aberrant expression of inflammatory genes in monocytes. *Arch Gen Psychiatry*, 65(4):395–407, Apr 2008.

[27] Marion Leboyer, Isabella Soreca, Jan Scott, Mark Frye, Chantal Henry, Ryad Tamouza, and David J Kupfer. Can bipolar disorder be viewed as a multi-system inflammatory disease? *J Affect Disord*, 141(1):1–10, Dec 2012.

[28] Miriam Weiner, Lois Warren, and Jess G Fiedorowicz. Cardiovascular morbidity and mortality in bipolar disorder. *Ann Clin Psychiatry*, 23(1):40–47, Feb 2011.

[29] Giuseppe Maina, Virginio Salvi, Alberto Vitalucci, Virginia D'Ambrosio, and Filippo Bogetto. Prevalence and correlates of overweight in drug-naive patients with bipolar disorder. *J Affect Disord*, 110(1-2):149–155, Sep 2008.