

1 A system for tracking whisker kinematics and whisker shape in three dimensions

2

3 Short title: Tracking whisker kinematics and shape in three dimensions

4

5

6 Rasmus S. Petersen ^{1¶*}, Andrea Colins Rodriguez ^{1¶}, Mathew Hywel Evans ², Dario Campagner ³ and Michaela S. E.

7 Loft ¹

8 ¹ Division of Neuroscience and Experimental Psychology, University of Manchester, Manchester, United Kingdom

9 ² School of Psychology, University of Nottingham, Nottingham, United Kingdom.

10 ³ Sainsbury Wellcome Centre for Neural Circuits and Behaviour, University College London, London, United Kingdom

11 * Corresponding author

12 E-mail: R.Petersen@manchester.ac.uk

13 ¶ These authors contributed equally to this work

14

15

16

17

18

19

20

21

22

23

24 **Abstract**

25 Quantification of behaviour is essential for systems neuroscience. Since the whisker system is a major model system for
26 investigating the neural basis of behaviour, it is important to have methods for measuring whisker movements from
27 behaving animals. Here, we developed a high-speed imaging system that measures whisker movements simultaneously
28 from two vantage points. We developed an algorithm that uses the ‘stereo’ video data to track multiple whiskers by
29 fitting 3D curves to the basal section of each target whisker. By using temporal information to constrain the fits, the
30 algorithm is able to track multiple whiskers in parallel with low error rate. We used the output of the tracker to produce a
31 3D description of each tracked whisker, including its 3D orientation and 3D shape, as well as bending-related mechanical
32 force. In conclusion, we present an automatic system to track whiskers in 3D from high-speed video, creating the
33 opportunity for comprehensive 3D analysis of sensorimotor behaviour and its neural basis.

34 **Author summary**

35 The great ethologist Niko Tinbergen described a crucial challenge in biology to measure the “total movements made by
36 the intact animal”. Advances in high-speed video and machine analysis of such data have made it possible to make
37 profound advances. Here, we target the whisker system. The whisker system is a major experimental model in
38 neurobiology and, since the whiskers are readily imageable, the system is ideally suited to machine vision. Rats and mice
39 explore their environment by sweeping their whiskers to and fro. It is important to measure whisker movements in 3D,
40 since whiskers move in 3D and since the mechanical forces that act on them are 3D. However, the problem of
41 automatically tracking whiskers in 3D from video has generally been regarded as prohibitively difficult. Our innovation
42 here is to extract 3D information about whiskers using a two-camera, high-speed imaging system and to develop
43 computational methods to infer 3D whisker state from the imaging data. Our hope is that this study will facilitate
44 comprehensive, 3D analysis of whisker behaviour and, more generally, contribute new insight into brain mechanisms of
45 perception and behaviour.

46

47

48

49 **Introduction**

50 Substantial progress towards the long-standing ambition of measuring “total movements made by the intact animal” (1) is
51 coming from the application of powerful machine vision methods to video recordings of behaving animals (2). Since the
52 whisker system is a major experimental model in neuroscience and since the whiskers are readily imageable (3,4), the
53 whisker system is ideally suited to this endeavour. Tracking the whiskers of mice/rats has already deepened our
54 understanding of active sensation and refined our capacity to relate behaviour to neural mechanisms (5–10) Our aim here
55 was to develop a method to track whisker movements and whisker shape in 3D in behaving mice at millisecond temporal
56 resolution.

57 Whisker movement is 3D. During each whisking cycle, the follicles translate with respect to the head and each whisker
58 rotates in 3D. Although only the horizontal component of this movement is typically measured, whiskers also move
59 vertically (11) and rotate around their longitudinal axes (‘roll’) (5). The mechanical forces of whisker-object contact that
60 are the primary drivers of neural activity are also 3D. When a mouse ‘whisks’ against an object, the whiskers bend.
61 Again, although only the horizontal component of bending is typically measured, bending can occur in all directions (12).
62 In the trigeminal ganglion, all directions of deflection are encoded (13–15), indicating that 3D bending information is
63 both encoded and transmitted to the brain.

64 Starting with the first “cinematographic” study of whisking by Welker in 1964, there is a 50 year history of increasingly
65 sophisticated efforts to measure whisker movement from behaving animals (4). Most studies have measured whisker
66 movement only in the horizontal plane, using either linear CCD arrays (16,17) or high-speed imaging (6,18–20)
67 However, horizontal plane imaging provides direct measurement of only one of the 3 angles that define 3D whisker
68 orientation. Moreover, estimates of whisker-object bending force (‘bending moment’) obtained by imaging apparent
69 curvature of a whisker in the horizontal plane (19,21,22) can be contaminated by roll (5). This is significant, since
70 bending moment is the primary driver of contact-related mechanotransduction (23–25). High-speed cameras sufficient to
71 form the basis of a 3D whisker imaging system have long been available: the main bottleneck to achieving 3D whisker
72 tracking has been the computational complexity of the 3D inference problem. A few studies have measured aspects of 3D
73 whisker movement in vivo (5,11,12,26) and ex vivo (27),but no automatic approach has so far been developed that
74 measures both 3D whisker orientation and 3D whisker shape from high-speed video of behaving animals. Here, we
75 obtained 3D information using a two-camera, high-speed imaging system and developed computational methods to infer
76 3D whisker state from stereo video data. We use the system to track up to 8 whiskers in parallel, and to obtain a 3D
77 description of each whisker, encompassing both its 3D orientation and 3D shape.

78 **Results**

79 **3D Imaging of whisking behaviour**

80 To obtain a video data set with which to develop 3D whisker tracking, we trained head-fixed mice to detect objects with
81 their whiskers (n=6). On each trial, a vertical pole was presented in either an anterior location out of reach of the
82 whiskers ('no-go trial') or a posterior location within reach ('go trial'). Mice learned to perform the task accurately
83 ($81 \pm 17\%$, mean \pm SD over mice) and performed 135 ± 22 trials per daily session. When the pole moved up at the onset
84 of a trial, mice would typically commence exploratory whisking. On go trials, one or more whiskers typically contacted
85 the pole; on no-go trials, there was no contact. In this way, we obtained a varied data set, which included episodes of
86 whisking both with and without contact (average behavioural session ~ 0.5 M video frames).

87 We recorded high-speed video of mice using a system of two high-speed cameras (Fig 1). One camera imaged whisking
88 in the horizontal plane. The other camera imaged in an off-vertical plane, the orientation of which (25° off coronal, 10°
89 off horizontal) was optimised to minimise occlusion of whiskers against the background of the body of the mouse (Fig
90 1). For brevity, we refer to this second image plane as 'vertical'.

91

92 **Fig 1. Experimental set-up for 3D imaging.** A) Schematic showing the camera angles and 3D head-centred xyz
93 coordinate frame. B) Horizontal and vertical views, with corresponding 2D coordinate frames.

94

95 **Bezier curve framework for representing whiskers in 3D**

96

97 Using the two-camera set-up, we imaged mice (1000 frames/s) as they performed the pole detection task. This resulted
98 in a time series of image pairs (horizontal and vertical views): we refer to each such image pair as a 'frame'. Our next
99 aim was to develop an automated algorithm to track whiskers in 3D. For two reasons, we focussed on the basal segment
100 of the whisker shafts. First, during whisker-object contact, whiskers bend and the associated mechanical force/moment
101 drives mechanoreceptors located in the follicle. Thus, changes in whisker shape at the base of the whisker are intimately
102 related to neural activity in the ascending whisker pathway (24,25). Second, tracking only the basal segment (not the
103 whole whisker) reduces the number of parameters required to describe the shape of a whisker. To accurately describe the
104 shape of a whisker across its entire length requires at least a 5th degree polynomial (22) which, in 3D, has 15 parameters.

105 However, the basal segment of a whisker is well-approximated by a quadratic curve (19,27) which, in 3D, has 9
106 parameters.

107

108 The whisker tracker describes each target whisker as a 3D Bezier curve (Fig 2). This is a parametric curve segment $\mathbf{b}(s)$
109 $= (x(s), y(s), z(s))$, where $0 \leq s \leq 1$ parameterises location along the curve. In our case, $s = 0$ marked the end closest
110 to the whisker base and $s = 1$ marked the end furthest from the base. A Bezier curve is defined by 2 or more ‘control
111 points’ (Fig 2), the number of which controls the complexity of the curve. We used quadratic Bezier curves, each of
112 which has 3 control points.

113

114 **Fig 2. Description of whiskers by quadratic 3D Bezier curves.** Left: schematic of a 3D Bezier curve representing a
115 whisker (blue line), defined by its three control points $\mathbf{cp}_0, \mathbf{cp}_1$ and \mathbf{cp}_2 (coloured dots). Middle, right: projection of the
116 3D Bezier curve, and its control points, onto horizontal and vertical image planes.

117

118 Whisker tracking algorithm

119

120 The essence of our algorithm was to track one or more target whiskers by fitting 3D Bezier curves to the image data.
121 Analogously to stereoscopic vision, two complementary planar views of a whisker provide sufficient information for its
122 full 3D location, orientation and shape to be inferred. The core principle of the tracker was, for each frame, to tune the
123 control points of the Bezier curves so that their projections onto the two image planes matched as closely as possible the
124 images of the basal segments of the target whiskers. The degree of match was quantified by a cost function (Methods,
125 Equation 4). Since whiskers imaged as black (low pixel intensity) and background as white (high pixel intensity), the
126 average image intensity along a whisker was low whereas that over a background region was high. Thus, if Bezier curve
127 $\mathbf{b}(s)$ accurately described a given whisker, then the sum of the image intensity along the projection of the curve in both
128 horizontal and vertical views was low (Fig 2) and at a local minimum with respect to local changes to the positions of the
129 control points.

130

131 In each frame, the cost function was minimised with respect to the control point positions of each Bezier curve
132 independently. It was possible to track multiple whiskers in parallel by taking advantage of prior knowledge of natural
133 whisker motion. First, since whiskers move and deform smoothly over time, the location of a whisker in a given frame
134 was (with sparse exceptions, see below) predictable from that in the previous frames. Thus, by seeding the control point
135 positions of a Bezier curve representing a given whisker using corresponding positions from previous frames (Methods),

136 it was possible to maintain accurate ‘locking’ between each Bezier curve and its target whisker. Second, we used prior
137 knowledge to constrain the cost function. As detailed in Methods, a ‘temporal contiguity’ constraint penalised
138 discontinuous temporal changes in Bezier curves (Methods, Equation 7) and a ‘shape complexity’ constraint penalised
139 unnaturally complex curve shapes (Methods, Equation 8). The full whisker tracking pipeline (Fig 3) is detailed in
140 Methods.

141

142 **Fig 3. Whisker tracking pipeline.** Left: Initialisation of control points for a given target whisker (see Methods for
143 details). Initial values for control points in horizontal (top, white circles) and vertical views (bottom, white circles). White
144 dotted lines in vertical view represent the range of z values consistent with each of the (x,y) points in horizontal view.
145 Middle: Estimation of snout contour (yellow). Right: Fitting of 3D Bezier curves to image data. Projections of the 3D
146 Bezier curve for one whisker (blue lines) and of its control points (blue dots) are shown in horizontal (top) and vertical
147 (bottom) views. Yellow dots indicate intersections between snout contour and extrapolated Bezier curves.

148 **Tracking multiple whiskers in 3D**

149 To test the algorithm, we applied it to the image data from our task. We found that we were able to track several
150 whiskers at the same time (Fig 4; Movie 1). Fig 4C shows a 12 ms sequence of whisker-pole contact from a mouse where
151 8 whiskers were intact and the others had been trimmed to the level of the fur (Fig 4A-B). The algorithm successfully
152 tracked changes in both orientation and shape of the 8 whiskers. Different types of motion were tracked: some whiskers
153 bent against the pole whilst others slipped past it (Fig 4D). The outcome of the tracker was a sequence of 3D curve
154 segments, each representing the basal segment of a given whisker in a given frame (Fig 4E).

155

156 **Fig 4. Tracking multiple whiskers in 3D.** A-B) 8 whiskers were tracked in a 3.5 s video sequence. C) A 12 ms
157 sequence of frames showing Bezier curves for all tracked whiskers, projected into horizontal and vertical views, taken
158 from the example video (Movie 1). Whiskers are colour coded as in panel A. D) Tracking solutions for 2 whiskers
159 (colour coded as in panel A) across 12 frames projected onto horizontal and vertical views. E) 3D tracking solutions for 8
160 whiskers across a sequence of 30 frames, including the sequence of panel D.

161 To assess tracking accuracy, we tracked a randomly selected set of 100 trials (50 go, 50 no-go) where a mouse was
162 performing the task with 3 whiskers, all others trimmed to the level of the fur. This dataset comprised 350,000 frames.
163 During ‘free whisking’, changes in whisker position/shape were entirely due to whisking motion, and such changes were
164 smooth as a function of time, so that the ‘temporal contiguity’ and ‘shape complexity’ constraints of the cost function

165 (Methods, Equations 4 and 7) were accurate. Such errors as did occur were mainly due to (1) occlusion and (2) whisker
166 overlap. (1) On occasion, a whisker was occluded against either the mouse's body (ear or cheek) or the experimental
167 apparatus (pole). However, by optimising the view angles of the cameras (see above) and by minimising the image
168 footprint of the apparatus, we minimised these effects. On the no-go trials, we detected 0.11 occlusion events/whisker
169 per 1000 frames. Since occlusion was rare, such events were dealt with by skipping affected frames and restarting
170 tracking afterwards. (2) On occasion, whiskers overlapped each other in either horizontal or vertical view. Because the
171 tracker applies prior knowledge of natural whisker shape/location, our algorithm was relatively robust to such events.
172 However, errors did sometimes occur. The most common overlap error was when a small whisker overlapped a large
173 one to such a degree that the Bezier curve tracking the small whisker locked onto the large whisker. Such events were
174 minimised by trimming all non-target whiskers to the level of the fur and were rectified, when they did occur, by using
175 GUI tools to nudge the control points of the errant Bezier curve back onto its target whisker. The incidence of overlap
176 errors increased with the number of intact whiskers. It depended also on their location: there was typically less overlap
177 within a row of whiskers than across an arc. On no-go trials of the test data set, there were 0.01 overlap errors/whisker
178 per 1000 frames).

179 Tracking was more challenging when videos include not only whisking motion but also whisker-object contact. During
180 contact, changes in whisker position/shape from frame to frame were most often smooth and gradual (Fig 7) but,
181 occasionally, a whisker slipped off the pole at high speed ('slip event'), generating discontinuous whisker change
182 between adjacent frames (Fig 4D-E). During such slips, tracking errors sometimes occurred, since the tracker's routine
183 for estimating the location of a whisker based on previous frames assumes smooth motion. On go-trials, high-speed slips
184 occurred in a small fraction of video frames (0.23 slips/whisker per 1000 frames). Errors resulting from slips were
185 corrected, as above, using GUI tools.

186 **Measuring 3D whisker orientation and 3D whisker shape**

187 Having tracked one or more whiskers in a set of videos, the next step was to use the tracking data to estimate 3D whisker
188 kinematics and 3D whisker shape. To this end, we measured the 3D orientation of each whisker (12,27) in terms of its
189 azimuth (θ), elevation (φ) and roll (ζ) (Fig 5; Methods; Movie 2).

190 **Fig 5. Description of a whisker in terms of 3D kinematic and 3D shape parameters.** A) Azimuth (θ), elevation (φ)
191 and roll (ζ) angles. These angles are defined with respect to the tangent to the Bezier curve $\mathbf{b}(s)$ describing the whisker,
192 at $s = 0$. Azimuth describes rotation about the vertical (dorso-ventral) axis through $s = 0$; elevation describes rotation
193 about the horizontal (medio-lateral) axis through $s = 0$; roll describes rotation about the x' axis, defined in panel B. B)

194 *Left.* Whisker-centric coordinate frame with origin at $s = 0$ (Equations 9-11). The x' axis is tangent to $\mathbf{b}(s)$ at $s = 0$; the
195 y' axis is the direction in which $\mathbf{b}(s)$ curves; the z' axis is orthogonal to the $x' - y'$ plane. *Middle.* Components of
196 moment in the whisker-centric coordinate frame. *Right.* 2D and 3D whisker curvature (Equations 13-15).

197 To illustrate the method, we first estimated 3D whisker angles during free whisking and illustrate results for a mouse
198 with whiskers C1-C3 intact (Fig 6). Azimuthal whisker angle was highly correlated across whiskers (whiskers C1-C3,
199 Pearson correlation coefficients $\rho = 0.98-0.99$), as was elevation ($\rho = 0.94-0.99$) (Fig 6A). Elevation was highly anti-
200 correlated with azimuth ($\rho = -0.89--0.96$; Fig. 6B left). Roll angle correlated with azimuth/elevation but, consistent with
201 (5), the degree of correlation was whisker-dependent ($\rho = 0.13-0.80$; Fig. 6B middle, right). Whisker-object contact
202 perturbed these angle relationships and partially decorrelated the azimuth-elevation relationship ($\rho = -0.49--0.79$).

203

204 **Fig 6. 3D whisker kinematics during free whisking.** **A)** Changes in 3D angles for whiskers C1, C2 and C3 during a 3.5
205 s episode of free whisking. **B)** Relationships between angles.

206 An important feature of video-based whisker tracking is that allows non-invasive measurement not only of kinematics but
207 also of mechanical forces/moments acting on the whiskers (4,12,19,21). The physical basis for this is that the shape of a
208 whisker contains information about these mechanical factors. In particular, there is a linear relationship between whisker
209 curvature and bending moment. Previous studies have sought to measure the bending moment related to whisker-object
210 contact by imaging in the horizontal plane and have found that changes of whisker curvature in the horizontal plane
211 predicts touch-triggered neural activity (7,20,24,25). However, there are limitations of the planar imaging approach. First,
212 it senses only the component of bending moment in the horizontal image plane and necessarily misses any out-of-plane
213 bending. Second, since whiskers roll during the whisking cycle, the shape of a whisker, as projected in the horizontal
214 plane can change purely due to roll even during free whisking (Fig 7). A benefit of 3D imaging is that it addresses these
215 limitations. First, 3D imaging enables bending in any direction to be measured. Second, it permits the intrinsic shape of a
216 whisker to be teased apart from both its position and angular orientation. We measured the intrinsic shape, which we
217 term κ_{3D} : $\kappa_{3D}(s)$ expresses the 3D curvature at each point s along the whisker shaft and has the key property of being
218 invariant to curve location and to curve orientation (Methods; Equation 13).

219 First, to verify that our system accurately measures 3D curvature we tested whether it could correctly recover the shape
220 of a rotating, rigid object. To this end, we simulated whisking (motion with both forward-backward and rolling
221 components) a rigid disk with curvature comparable to that of a typical mouse whisker (a 13 mm diameter, glass
222 coverslip), imaged it as described above and tracked its edge (Fig 7A). Due to the rolling motion, curvature in the

223 horizontal and vertical planes, κ_h and κ_v , oscillated strongly (Fig 7B). Despite this, our algorithm correctly recovered
224 that 3D curvature κ_{3D} was constant - fluctuations (SD) in κ_{3D} were 5.6% that of κ_h and 8% that of κ_v - and matched the
225 true curvature of the object (Fig 7B).

226 However, when we measured $\kappa_{3D}(s)$ of mouse whiskers from behaving mice, we found substantial fluctuations, even
227 during free whisking (Fig 7C): SD of κ_{3D} was 40-91% that of κ_h and 51-68% that of κ_v . This suggests that, in contrast to
228 rat where the proximal segment of the whisker shaft undergoes rigid motion during whisking (5), during natural
229 whisking, mouse whiskers do not behave as rigid objects. To determine whether this lack of rigidity occurs in the absence
230 of whisker movement ('static') or whether it is dependent on movement ('dynamic'), we first positioned a (stationary) ex
231 vivo mouse whisker (C3) in the apparatus at a series of roll angles, imaged it, tracked it and measured κ_{3D} . Varying roll
232 angle changed κ_{3D} by up to 6% (Fig 7D). Next, we 'whisked' the whisker backwards and forwards in the horizontal
233 plane at different velocities (Fig 7E) and measured κ_{3D} as a function of whisking phase (Fig 7F). We found that κ_{3D}
234 changed by up to 12% over a cycle. This change in κ_{3D} increased linearly with whisking velocity (Fig 7F; $R^2=0.95$).
235 Linear extrapolation to average whisking velocity observed during free whisking in mice performing our task ($0.78 \text{ }^\circ/\text{ms}$)
236 implied a κ_{3D} change of 66% (C3). In comparison, changes in κ_{3D} during free whisking were 56-150% (whiskers C1-3).
237 Overall, these data indicate that whisker motion during free whisking in mouse is non-rigid and predominantly a dynamic
238 effect.

239

240 **Fig 7. Tracking and estimating 3D curvature for a rigid test object (panels A-B), whiskers of a behaving mouse**
241 **(panel C) and an ex vivo whisker (panels D-F).** **A)** Tracking the edge of a coverslip. The cover slip was mounted, like a
242 lollipop, on a rod; the rod was oriented in the mediolateral direction and rotated around its axis. Red lines indicate
243 tracking results (300 ms period, 10 ms intervals). **B)** Top: Azimuth angle for two trials (black and grey traces). Bottom
244 shows measured curvature: horizontal curvatures (dotted lines), κ_{3D} (solid lines) and true curvature (orange). **C)**
245 Horizontal and 3D curvatures during free whisking (same trial as Fig 6). Solid lines represent κ_{3D} and dotted lines
246 indicate horizontal curvatures for C1-3 (colours coded as in Fig 6). **D)** κ_{3D} for a stationary ex vivo whisker (C3) at
247 different roll angles. **E)** Azimuth angle for ex vivo trials with simulated whisking at different speeds. **F)** κ_{3D} as a function
248 of whisking phase.

249 Estimation of κ_{3D} allowed construction of a simple proxy to the magnitude of the bending moment, which we term $\Delta\kappa_{3D}$
250 (Methods). This quantity is a generalisation of the corresponding measurement from horizontal plane imaging, referred
251 to here as $\Delta\kappa_h$. Since the mice were whisking against a vertical pole, the predominant changes in curvature were in the

252 horizontal plane and, thus, one might expect minimal benefit from the 3D approach. Even here, however, we found $\Delta\kappa_h$
253 to be markedly contaminated by roll. A simple instance of this effect is shown in Fig 8A (Movie 3). Here, the whisker
254 initially curved downwards (roll angle -90°): whisker-pole contact from time 0-45 ms rolled the whisker in the caudal
255 direction (roll angle -180°) with only minimal change in 3D curvature but with substantial effect on the curvature
256 projected in the horizontal plane. Thus, $\Delta\kappa_h$ increased by 0.05 mm^{-1} , introducing a marked mismatch between $\Delta\kappa_h$ and Δ
257 κ_{3D} (Fig 8A bottom). A more typical and complex instance of the effect of roll angle is shown in Fig 8B (movie 4).
258 Here there were large fluctuations in $\Delta\kappa_h$ (e.g., at times 310-370 ms, fifth touch) which almost entirely reflected changes
259 in roll angle in the absence of change to 3D curvature. On average, $\Delta\kappa_h$ explained 44% of the variation in $\Delta\kappa_{3D}$ (touch
260 periods from 47 trials). In this way, 3D imaging permits more accurate measurement of mechanical forces acting on
261 whiskers.

262

263 **Fig 8. Comparison of 2D and 3D curvature as mouse whisks against a pole (whisker C2): curvatures (upper**
264 **panel)-, 3D kinematics (middle panel) and curvature change (bottom panel, $\Delta\kappa_{3D}$, $\Delta\kappa_h$, and $\Delta\kappa_v$).** A) Contact
265 episode where both movement and bending of the whisker were largely restricted to the horizontal plane. In this case, Δ
266 κ_{3D} and $\Delta\kappa_h$ were highly correlated. Grey shading indicates periods of whisker-pole contact. See Movie 3. B) Example
267 with same whisker as panel A for contact episode with significant vertical component of whisker motion. See Movie 4.

268

269 Discussion

270 In order to obtain a comprehensive description of 3D whisker movements and 3D whisker-object interactions, we imaged
271 mouse whisking behaviour using a high-speed 'stereo' imaging system. We developed software, first, to fit a 3D curve
272 segment to each of one or more target whiskers and, second, to extract 3D kinematic and shape parameters from them.
273 The new method allows both the 3D orientation (azimuth, elevation and roll) of a whisker and its intrinsic 3D curvature
274 to be measured with millisecond precision, during both free whisking and whisker-object contact.

275 The vast majority of previous work on automatic whisker tracking has focussed on imaging in the horizontal plane (6,16–
276 19,28,29)(6,16–20,28) . However, as detailed in the Introduction, single-plane imaging necessarily captures only a
277 fraction of the full 3D kinematic and 3D shape parameters that characterise a whisker. The advance here is an automatic
278 method able to extract a full 3D description of both whisker kinematics and whisker shape.

279 Our work builds on previous advances in 3D whisker tracking in behaving rodents. Methods based on linear CCD arrays
280 and markers provide simple and effective means to measure some aspects of 3D whisker kinematics, but also have
281 limitations. CCD arrays (11) image whiskers at a single point along the shaft: this method confounds whisker translation
282 with whisker rotation, cannot measure whisker shape and cannot recover roll angle. Also, CCD study of 3D whisking
283 has been limited to one whisker at a time. The approach of marking a whisker with spots of dye (5,26) has been used to
284 recover all 3 orientation angles but has not been used to measure whisker shape changes during object touch. Also,
285 although application of dye marker dots was reported not to disturb rat whisking, mouse whiskers are thinner, less stiff
286 and hence more liable to perturbation. Our method addresses these limitations by leveraging the extra information
287 available from video, and achieves automatic, multi-whisker tracking without markers, as well as the ability to recover
288 not only three-angle kinematic information but also information about whisker shape changes during object contact. The
289 only previous 3D video-based study (12) required manual tracking for the vertical view.

290 The method presented here has some limitations. First, although the error rate was low, the experimental conditions were
291 designed to avoid both extensive object-whisker occlusion (by using a stimulus object with small foot-print) and
292 extensive whisker-whisker overlap (by trimming non-target whiskers). Error rates are likely to be higher in the presence
293 of experimental apparatus where there is more occlusion or if no whisker trimming is carried out. Use of additional
294 cameras provides a potential way to reduce error rates further. Second, our tracker is focussed on describing the basal
295 segment of the whisker, not the full whisker, since our focus is primarily on elucidating the fundamental mechanical
296 events that drive neural activity in the whisker system. To track a whisker across its entire length, a quadratic curve
297 description is insufficient. One direction for future work is to investigate whether the tracker can be extended to fit
298 higher order Bezier curves without excessive loss of robustness. Third, our method has been developed for head-fixed
299 mice. This has the advantage that it simplifies the tracking problem. However, it would also be useful to extend the
300 approach to freely moving animals, perhaps by combining whisker tracking with head-body tracking (30–33).

301 In conclusion, this study has presented the first automatic method for measurement of both 3D whisker kinematics and
302 whisker shape changes with millisecond resolution. The method can be combined with cellular resolution neural activity
303 measurement and thus has potential to advances our understanding of sensorimotor behaviour in a key model system.

304

305

306

307 **Movies**

308 **Movie 1: Tracking example of 8 whiskers.** Left: horizontal view of the whiskers and tracking superimposed. Colours
309 are shown as in Fig 4A. Axes are shown as in Fig 1B. Middle: Vertical view of the whiskers and tracking superimposed.
310 Right: Bezier curves in the three dimensional space. Axes are shown as in Fig 1A.

311 **Movie 2: Whisker tracking and variables.** Top: Horizontal and vertical views with tracking superimposed. Colours are
312 shown as in Fig 4A. Bottom: 3D kinematic and 3D shape parameters: Horizontal angle (Azimuth), vertical angle
313 (Elevation) , horizontal and vertical curvature, κ_{3D} and roll for each tracked whisker.

314 **Movie 3: Whisker tracking example of movement restricted to the horizontal plane (Fig 8A).** Top from left to right:
315 Horizontal and vertical views with tracking of C2 superimposed. Bezier curve in the whisker centred coordinate frame
316 isolating roll angle (Fig 5A). Comparison of whisker shape over time: Dashed line represents the whisker shape at $t = 0$
317 ms and solid line represents whisker shape of current frame. Bezier curve was rotated using azimuth, elevation and roll
318 angle to be captured in the two dimensional plane. Bottom from left to right: Horizontal angle (Azimuth), Vertical angle
319 (Elevation), Roll, horizontal and 3D curvature over time. Colours are shown as in Fig 8A.

320 **Movie 4: Whisker tracking example of movement with significant vertical components (Fig 8B).** Top from left to
321 right: Horizontal and vertical views with tracking of C2 superimposed. Bezier curve in the whisker centred coordinate
322 frame isolating roll angle (Fig 5A). Comparison of whisker shape over time: Dashed line represents the whisker shape at
323 $t = 0$ ms and solid line represents whisker shape of current frame. Bezier curve was rotated using azimuth, elevation and
324 roll angle to be captured in the two dimensional plane. Bottom from left to right: Horizontal angle (Azimuth), Vertical
325 angle (Elevation), Roll, horizontal and 3D curvature over time. Colours are shown as in Fig 8A.

326

327 **Methods**

328 All experimental protocols described in this section were approved by both United Kingdom Home Office national
329 authorities and institutional ethical review. All parameters and variables used in this section are summarised in Table 1.
330 All computer code was written in MATLAB (The MathWorks Inc., Natick, MA).

331 **Behavioural apparatus**

332 Mice (C57; males; 6 weeks at time of implant) were implanted with a titanium head-bar as detailed in (24). After surgery,
333 mice were left to recover for at least 5 days before starting water restriction (1.5 ml water/day). Training began 7-10 days
334 after the start of water restriction.

335 Mice were trained and imaged in a dark, sound-proofed enclosure using apparatus adapted from (24). A head-fixed
336 mouse was placed inside a perspex tube, from which its head emerged at one end. The stimulus object was a 0.2 mm
337 diameter, vertical carbon fibre pole which could be translated parallel to the anterior-posterior (AP) or medio-lateral
338 (ML) axes of the mouse by a pair of linear stepper motors and rotated in the horizontal plane to 'go' or 'no-go' locations
339 by a rotatory stepper motor. To allow vertical movement of the pole into and out of range of the whiskers, the apparatus
340 was mounted on a pneumatic linear slide, powered by compressed air. The apparatus was controlled from MATLAB via
341 a real-time processor. Mouse response was monitored by a lick port located anterior to the mouth. Licks were detected as
342 described in (6). Each lick port consisted of a metal tube connected to a water reservoir via a computer-controlled
343 solenoid valve. Lick port position was monitored using an infrared camera and adjusted using a micromanipulator.

344

345 **Behavioural task**

346

347 Head-fixed mice were trained to detect the presence of a metal pole using their whiskers, using behavioural procedures
348 similar to (9). On each trial, the pole was presented either within reach of the whiskers ('go trial') or out of reach ('no-go
349 trial'). At the start of each trial, the computer triggered the pole to move up (travel time ~100 ms). The pole stayed up
350 for 1 s, before moving down. On go trials, the correct response was for the mouse to lick a lick port. Correct responses
351 were rewarded by a drop of water (~10 μ l). Incorrect responses on go trials (not licking) were punished by timeout (3-5
352 s). On no-go trials, the correct response was to refrain from licking and incorrect responses (licking) were punished by
353 timeout and tone (frequency 12 kHz).

354

355 **High-speed stereo whisker imaging**

356 Whiskers were imaged, based on the methods of (24), except that, to provide 3D information, two cameras were used.
357 The whiskers were imaged using two high-speed cameras (Mikrotron LTR2, Unterschleissheim, Germany; 1000
358 frames/s, 0.4 ms exposure time) via telecentric lenses (Edmunds Optics 55-349, Barrington, NJ) as illustrated in Fig 1.
359 Illumination for each camera was provided by a high-power infrared LED array (940 nm; Roithner LED 940-66-60,
360 Vienna, Austria) via diffuser and condensing lens. The imaging planes of the two cameras were horizontal (spanning AP
361 and ML axes) and vertical respectively. The field of views were typically 480 x 480 pixels, with pixel width 0.047 mm.

362 The two cameras were synchronised by triggering data acquisition off the computer-generated TTL pulse that initiated a
363 trial. Typically, imaging data were acquired in an interval starting 0.5 s before pole onset, ending 1.8 s after pole onset.
364 To provide an independent check that data files from the two cameras came from corresponding trials, an IR LED was
365 positioned in the corner of the field of view of each camera and, starting at pole onset on each trial, flashed a binary
366 sequence that encoded the trial number. Onset of this LED signal also served to verify camera synchrony.

367 **Coordinate frame and calibration**

368

369 To describe the location of whiskers in 3D, we used a left-handed Cartesian coordinate frame, fixed with
370 respect to the head of the animal (Fig 1). The axes were x (AP, with positive x posterior); y (ML, with
371 positive y medial) and z (DV, with positive z dorsal). In standard anatomical convention, the x - y (AP-ML)
372 plane was horizontal; the x - z (AP-DV) plane sagittal and the y - z (ML-DV) plane coronal. The vector \mathbf{p}^{3D}
373 $= (x,y,z)^T$ denotes a point with coefficients along the x , y and z axes respectively. Throughout, we denote
374 vectors by lower-case bold (e.g., \mathbf{p}), scalars by lower-case italic (e.g., s) and matrices by upper-case bold (e.g.,
375 \mathbf{M}).

376

377 Analogously to stereoscopic vision, our whisker tracker infers 3D whisker location/orientation from images
378 obtained from two viewpoints - horizontal and vertical. Pixel locations in the horizontal image were defined
379 using the x and y axes of the 3D frame (Fig 1). Thus, the location of a point \mathbf{p}^H in the horizontal image was
380 described by a vector (x,y) . Pixel location \mathbf{p}^V in the vertical image was described by a vector (v,w) , defined
381 with respect to axes v and w (Fig 1). Due to the orientation of the vertical-view camera detailed above, the
382 v,w coordinate frame was rotated and translated with respect to the x,y,z frame. The relation between the
383 x,y,z and v,w coordinate frames was determined as follows.

384

385 Since the imaging was done with telecentric lenses, the mappings from 3D to the two image planes were
386 described as orthogonal projections. The projections of a 3D point \mathbf{p}^{3D} onto a 2D point \mathbf{p}^H in the horizontal
387 image plane and a 2D point \mathbf{p}^V in the vertical plane were:

388

$$\mathbf{p}^H = \mathbf{H}\mathbf{p}^{3D} + \mathbf{h} \quad (1)$$

389

$$\mathbf{p}^V = \mathbf{V}\mathbf{p}^{3D} + \mathbf{v} \quad (2)$$

390

391 \mathbf{H} and \mathbf{V} were 2x3 matrices; \mathbf{h} and \mathbf{v} , 2-element column vectors. The 3D coordinate frame and the horizontal
392 image frame had common x and y axes and a common x,y origin: hence, $\mathbf{H} = [1 \ 0 \ 0; 0 \ 1 \ 0]$ and $\mathbf{h} = [0;0]$.

393

394 To determine the mapping from 3D to the vertical image plane (\mathbf{V} and \mathbf{v}), we performed the following
395 calibration procedure. Using stepper motors, we moved an object with 2 protruding pins on a 3D path through
396 the region of the behavioural set-up where the target whiskers were typically located, and recorded a sequence
397 of 100 corresponding images on each camera. The z location of each pin was known in each image. We then
398 tracked the tips of the 2 pins in each horizontal and vertical image frame to obtain a time series consisting of
399 the (x,y,z) and (v,w) coordinates of each pin tip. Using Equation 2, \mathbf{V} and \mathbf{v} were then estimated by linear
400 regression.

401

402 **Bezier curve framework for whisker tracking**

403 Our aim was to develop ‘whisker tracker’ software to track the orientation and shape of one or more target whiskers. The
404 whisker tracker described each target whisker segment as a Bezier curve, since these have convenient mathematical
405 properties (Fig 2). A Bezier curve is a parametric curve segment $\mathbf{b}(s) = (x(s),y(s),z(s))$, where $0 \leq s \leq 1$
406 parameterises location along the curve segment: in our case, $s = 0$ marked the end closest to the whisker base and $s = 1$
407 marked the end furthest from the base. The shape, orientation and position of a Bezier curve are determined by its
408 ‘control points’, the number of which determines the complexity of the curve. We used quadratic Bezier curves, which
409 have 3 control points \mathbf{cp}_i where $i = 0,1,2$, each with coordinates (x,y,z) . These control points were termed “proximal” (
410 \mathbf{cp}_0), “middle” (\mathbf{cp}_1) and “distal” (\mathbf{cp}_2) according to their distance from the whisker base. \mathbf{cp}_0 defined the location of the
411 basal end of the whisker segment, \mathbf{cp}_2 the distal end and \mathbf{cp}_1 the shape. In terms of these control point parameters, a
412 quadratic Bezier curve $\mathbf{b}(s)$ was expressed as:

413

$$\mathbf{b}(s) = \mathbf{cp}_0(1-s)^2 + 2\mathbf{cp}_1(1-s)s + \mathbf{cp}_2s^2 \quad (3)$$

414

415 **Whisker tracking pipeline**

416 **Overview.** Whisker tracking was operated via a Graphical User Interface (GUI). The GUI allowed a user to load a
417 pair of corresponding videos (horizontal view and vertical view). The first step was to calibrate, as detailed above. Next,
418 to initialise tracking, target whiskers were specified (automatically or manually) by defining approximate locations for
419 Bezier control points. After initialisation, each video frame was processed automatically, in turn. First, the contour of the
420 snout was located in both horizontal and vertical views. Second (except in the first frame), initial estimates for the Bezier
421 control points were calculated by linear extrapolation from their locations in the previous frames. Third, each Bezier
422 curve was fitted to the image data by adjusting its control points to minimise the cost function defined below (Equation
423 4). Provided the quality of fit for a given Bezier curve met a minimum threshold, tracking of that curve proceeded
424 automatically to the next frame.

425 **Manual initialisation** (Fig 3 left panel). When tracking a video for the first time, the first step was to specify the
426 target whiskers. In the first frame of the video, the user employed a graphical user interface (GUI) to select sets of
427 control points, specifying one or more target whiskers. For each target whisker, the user defined approximate locations
428 for control points specifying a curve segment corresponding to the basal segment of the whisker, by making computer-
429 mouse clicks within the video images. For each target whisker, the user first specified (x,y) coordinates for the 3 control
430 points in the horizontal view. Since the imaging geometry was described by linear equations (Equations 1 and 2), each
431 such point corresponded to a line in the vertical view. In the vertical view, the user specified a point (v,w) along each
432 vertical line where it intersected the target whisker. From these (x,y,v,w) data, the z coordinates of the control point
433 estimates were calculated from the calibration equation (Equation 2). Once initial values for all 3 control points of a
434 given target whisker were specified, refined estimates were calculated by the fitting procedure described below. In order
435 to obtain a reference value for the length of each curve, the arc length of each Bezier curve was calculated. To obtain a
436 reference value for the distance of the proximal control point from the snout, a second order polynomial was fitted to the
437 Bezier curve and extrapolated to find its intersection with the snout contour.

438 **Automatic initialisation.** Typically, an experiment will result in many videos taken of the same mouse under
439 identical experimental conditions. Once one video was tracked using the manual initialisation procedure described
440 above, other videos could then be initialised automatically through a template-matching approach. To initialise tracking
441 of a new video, the user selected a previously tracked video (via the GUI). For each target whisker from this file, a
442 sample of Bezier curve ‘templates’ were extracted (typically, the solution in every fifth video frame) and goodness of fit
443 of each sample curve to the first frame of the new video was calculated (using the cost function, Equation 4). The lowest
444 cost template was then selected. The template was refined by optimising the fit with respect to translations along both x
445 and z axes (within the range ± 5 pixels).

446 **Snout contour detection** (Fig 3 middle panel). First, to isolate the contour of the snout in a given video frame, fine
447 structure such as the hairs of the fur and the whiskers were removed by median filtering (5 x 5 pixels, 0.23 x 0.23 mm)
448 of the images followed by smoothing with a Gaussian filter (SD = 12 pixels, 0.56 mm). Next, the spatial gradient of each
449 filtered image was calculated in a direction approximately normal to the snout contour. This gradient was small, except
450 at the edge of the snout where it had a large peak. In the horizontal image, the snout contour was estimated as a function
451 of the x coordinate by minimising the gradient with respect to y . In the vertical image, the snout contour was estimated
452 as a function of the w coordinate by minimising the gradient with respect to v .

453 **Bezier curve fitting** (Fig 3 right panel). To achieve 3D tracking, we fitted 3D Bezier curves to the horizontal and
454 vertical view image data by varying the locations of their control points so as to minimise the following cost function.
455 Control points for each target whisker were optimised independently:

$$E(f) = E_h(f) + E_v(f) + R_1(f) + R_2(f) \quad (4)$$

456
457 Here $E(f)$ is the cost (or mismatch) between the image data of frame f and the Bezier curve $\mathbf{b}(f,s) = (x(f,s), y(f,s), z$
458 $(f,s))$, defined by control points $\mathbf{cp}_0(f)$, $\mathbf{cp}_1(f)$ and $\mathbf{cp}_2(f)$. $E_h(f)$ and $E_v(f)$ quantified how well $\mathbf{b}(f,s)$ described,
459 respectively, the horizontal and vertical image data of frame f . $R_1(f)$ and $R_2(f)$ were regularising terms (defined
460 below). In the following, to keep down clutter in the notation, dependence on frame and whisker is omitted except where
461 necessary for clarity.

462 E_h and E_v were defined as line integrals over the projection of $\mathbf{b}(s)$ in the horizontal/vertical images respectively:

$$E_h = \oint_{s=0}^{s=1} ds I_h(x(s), y(s)) \quad (5)$$

463

$$E_v = \oint_{s=0}^{s=1} ds I_v(v(s), w(s)) \quad (6)$$

464

465 Here: $I_h(x,y)$ was the intensity at point (x,y) in the horizontal view image, calculated by linear interpolation between
 466 pixel values and $I_v(v,w)$ the analogous quantity for the vertical view image; $(x(s),y(s))$ was the projection of $\mathbf{b}(s)$ in the
 467 horizontal view image and $(v(s),w(s))$ its projection in the vertical view image (Equations 1 and 2).

468 Except at occasional stick-slip events, whiskers move smoothly and, when imaged at 1000 frames/s, changes in location
 469 and shape from frame to frame were usually small, particularly for the basal segment. The regularising term R_1
 470 formalised this prior knowledge of natural whisking behaviour ('temporal contiguity'):

$$R_1(f) = \frac{1}{2}\sigma_1 \sum_{i=0}^{i=2} \|\mathbf{cp}_i(f) - \hat{\mathbf{cp}}_i(f)\|^2 \quad (7)$$

471 Here: $\mathbf{cp}_i(f)$ was the location of control point i of the whisker in frame f and $\hat{\mathbf{cp}}_i(f)$ was its location estimated by
 472 extrapolation based on its location in the previous two frames; σ_1 was a variable gain that the user could set from the
 473 GUI.

474 Additional regularisation was necessary to address degeneracy that could arise when tracking near-straight whiskers.
 475 Since a line segment is fully described by the location of its two ends, a straight whisker is fully defined by its proximal
 476 and distal control points – in this case, the middle control point is ill-defined. We found, under such situations, that the
 477 middle control point tended to migrate towards the whisker base and to generate high-curvature, unnatural shapes when
 478 extrapolating the curve to the snout contour (see above). To address this, we used a second regularising term which
 479 penalised deviations of the middle control point away from the midpoint between the proximal and distal control points:

$$R_2 = \frac{1}{2}\sigma_2 \left\| \frac{(\mathbf{cp}_1(f) - \mathbf{cp}_0(f))^T \mathbf{q}}{\|\mathbf{q}\|} - \frac{1}{2}\|\mathbf{q}\| \right\|^2 \quad (8)$$

480

481 Here $\mathbf{q} = \mathbf{cp}_2(f) - \mathbf{cp}_0(f)$ and σ_2 was a user-adjustable gain. R_2 measured deviation of the component of the middle
 482 control point $\mathbf{cp}_1(f)$ (relative to $\mathbf{cp}_0(f)$) in the direction of $\mathbf{cp}_2(f)$ (relative to $\mathbf{cp}_0(f)$) away from the midpoint of the
 483 $\mathbf{cp}_0(f) - \mathbf{cp}_2(f)$ line.

484 Nonlinear cost functions can be difficult to minimise due to local minima. However, in the present case, due to the
 485 smooth motion of whiskers referred to above, we expected control point solutions usually to be close to their values in
 486 the previous frame. Not only was it therefore effective to use a local search strategy, where the initial value for a given
 487 control point was set by extrapolating its values from the previous two frames ($\hat{\mathbf{cp}}_i(f)$), but this also made it possible to

488 track multiple whiskers independently. The cost function (Equation 4) was minimised (using MATLAB function
489 ‘fminunc’) with respect to components of the control points. To counter-act possible drift of $\mathbf{b}(s)$ along the whisker shaft
490 over time, or change in the arc-length of $\mathbf{b}(s)$ over time, we minimised the cost function with respect to components of
491 \mathbf{cp}_0 and \mathbf{cp}_2 normal to $\mathbf{b}(s)$ at $s = 0$ and $s = 1$ respectively. This procedure also had the advantage of reducing the
492 number of free parameters from 9 to 7. Furthermore, after convergence in a given frame, both the arc length of $\mathbf{b}(s)$ and
493 the distance of \mathbf{cp}_0 to the snout were normalised to equal their reference values set in the first frame (see above), whilst
494 preserving curve shape.

495 **Error correction.** As noted above, tracking of each target whisker proceeded automatically to the next frame, so long
496 as the cost E (Equation 4) remained less than a user-defined threshold (adjustable via the GUI). Should the threshold be
497 exceeded, for example when a Bezier curve was ‘left behind’ by rapid, discontinuous motion of its target whisker during
498 a slip event, tracking of that whisker ceased. To correct such an error, the GUI had tools allowing the user to nudge
499 control points back onto the target whisker, and to restart automatic tracking.

500 **Extracting 3D kinematics of the tracked whiskers**

501 The next step was to use the tracking data to estimate 3D whisker kinematics and 3D whisker shape. Since whiskers
502 bend during whisker-object contact, and this contact-induced whisker bending is a fundamental driver of neural activity
503 (see Introduction), it was important to develop a general procedure for describing 3D whisker motion, applicable to non-
504 rigid whisker movement. We separated changes to the orientation of a quadratic curve from changes to its shape in the
505 following manner.

506 Formally, we described whisker orientation by the following ‘whisker-centred’ Cartesian coordinate frame $x'y'z'$, with
507 origin at $s = 0$ (12). In contrast to the head-centred coordinate frame xyz , the $x'y'z'$ frame is time-dependent; rotating and
508 translating along with its target whisker. The x' -axis is aligned to the longitudinal axis of the whisker (tangent to $\mathbf{b}(s)$ at
509 $s = 0$). The y' -axis is orthogonal to the x' -axis, such that the $x' - y'$ plane is that within which $\mathbf{b}(s)$ curved. The z' -axis
510 is orthogonal to both x' and y' axes. Let \mathbf{i}' , \mathbf{j}' and \mathbf{k}' be unit vectors that point in the direction of the x' , y' and z' axes
511 respectively:

$$\mathbf{i}' = \left. \frac{\frac{d\mathbf{b}}{ds}}{\left| \frac{d\mathbf{b}}{ds} \right|} \right|_{s=0} \quad (9)$$

512

$$\mathbf{j}' = \frac{\left. \frac{d^2 \mathbf{b}}{ds^2} \right|_{s=0}}{\left| \left. \frac{d^2 \mathbf{b}}{ds^2} \right|_{s=0} \right|} \quad (10)$$

513

$$\mathbf{k}' = \mathbf{i}' \times \mathbf{j}' \quad (11)$$

514

515 Here $\mathbf{i}' \times \mathbf{j}'$ denotes the cross product of vectors \mathbf{i}' and \mathbf{j}' . The orientation of a whisker was then described by the 3D
 516 angle of the $x'y'z'$ coordinate frame with respect to the xyz coordinate frame. We translated the frames to have a
 517 common origin and then calculated the 3D rotation matrix that rotates the xyz frame to the $x'y'z'$ frame (34). This
 518 rotation can be described as the net effect of an ordered sequence of three elemental rotations with angles θ (azimuth), φ
 519 (elevation) and ζ (roll), and was expressed as a matrix $\mathbf{R}(\theta, \varphi, \zeta)$. Azimuth describes rotation in the horizontal ($x - y$)
 520 plane, about an axis parallel to the z axis through the whisker base; elevation describes rotation in the vertical ($x - z$)
 521 plane, about an axis parallel to the y - axis; roll describes rotation around the axis of the whisker shaft (Fig 5AB). We
 522 determined the angles θ, φ, ζ for a given whisker at a given time point by minimising the error function:

$$\sum (\mathbf{R}(\theta, \varphi, \zeta) [\mathbf{i}' \mathbf{j}' \mathbf{k}'] - [\mathbf{i} \mathbf{j} \mathbf{k}])^2 \quad (12)$$

523 Here \mathbf{i}, \mathbf{j} and \mathbf{k} are column unit vectors parallel to the x, y and z axes and the summation is over all matrix elements.

524 **Extracting 3D shape and bending moment of the tracked whiskers**

525 Having described the orientation of a whisker, the next task was to describe its shape. By ‘shape’, we intend those
 526 geometric properties of a curve that are invariant to its location and orientation. As noted above, we described whiskers
 527 by quadratic curve segments, which curve entirely within a plane (geometric torsion $\tau(s) = 0$). The intrinsic shape of a
 528 quadratic curve is fully described by a curvature function $\kappa_{3D}(s)$ (35):

$$\kappa_{3D}(s) = \frac{\left| \frac{d\mathbf{b}(s)}{ds} \times \frac{d^2\mathbf{b}(s)}{ds^2} \right|}{\left| \frac{d^3\mathbf{b}(s)}{ds^3} \right|} \quad (13)$$

529

530 Here $|\mathbf{a}|$ denotes the magnitude (2-norm) of vector \mathbf{a} . $\kappa_{3D}(s)$ has units of 1/distance and is the reciprocal of the radius of
 531 the circle that best fits the curve at point s . We computed planar curvatures as:

532

$$\kappa_h(s) = \frac{\frac{dx d^2y}{ds ds^2} - \frac{d^2x dy}{ds^2 ds}}{\left(\left(\frac{dx}{ds} \right)^2 + \left(\frac{dy}{ds} \right)^2 \right)^{\frac{3}{2}}} \bigg|_{s=0} \quad (14)$$

533

$$\kappa_v(s) = \frac{\frac{dz d^2y}{ds ds^2} - \frac{d^2z dy}{ds^2 ds}}{\left(\left(\frac{dz}{ds} \right)^2 + \left(\frac{dy}{ds} \right)^2 \right)^{\frac{3}{2}}} \bigg|_{s=0} \quad (15)$$

534

535 Here $x(s), y(s), z(s)$ are the components of $\mathbf{b}(s)$ in the x, y, z coordinate frame. Note, as detailed in Results, that, in
 536 contrast to $\kappa_{3D}(s)$, $\kappa_h(s)$ and $\kappa_v(s)$ are not invariant measures of geometric shape; they depend also on curve orientation.

537 In whisker-centric coordinates, bending corresponds to changes in shape of $\mathbf{b}(s)$ in the $x' - y'$ or $x' - z'$ planes (with,
 538 respectively, component $\mathbf{m}_{z'}$ defined in the direction of the positive z' axis and $\mathbf{m}_{y'}$ defined in the directions of the
 539 positive y' axis) (Fig 5;(36)). Since $\mathbf{b}(s)$ is a quadratic curve, it has zero torsion and its curvature is entirely confined to
 540 the $x' - y'$ plane: $\kappa_{3D}(s)$ is the curvature in this plane; the only non-zero component of bending moment is $\mathbf{m}_{z'}$.

541 Applying the standard relation between bending moment about a given axis and curvature in the plane normal to that axis
 542 (21,27), it follows that $\mathbf{m}_{z'}(s)$ is proportional to:

$$\Delta\kappa_{3D}(s) = \kappa_{3D}(f,s) - \kappa_{3D,0}(s) \quad (16)$$

543

544 where $\kappa_{3D,0}(s)$ is the curvature when the whisker is free from contact and in its resting state. All results presented here
 545 were evaluated at $s = 0$.

546 **Table 1. Parameters and variables summary.**

Axes

x	Anterior-posterior axis, with positive posterior
y	Medio-lateral axis, with positive medial
z	Dorsal-ventral axis, with positive dorsal
v	Medio-lateral axis from the vertical view, with positive medial
w	Dorsal-ventral axis from the vertical view, with positive ventral
x'	Axis from the whisker centred coordinated frame described as tangent to the whisker at $s = 0$. Positive indicates toward the tip of the whisker.
y'	Axis from the whisker centred coordinated frame defined by the second derivative of the Bezier curve at $s = 0$. Positive indicates $s > 0$
z'	Axis from the whisker centred coordinated frame defined as the cross product between x' and y'

Calibration

\mathbf{p}^H	Vector that represent a point in the horizontal plane with coefficients $(x,y)^T$
\mathbf{H}	Matrix belonging to the mapping between \mathbf{p}^{3D} and \mathbf{p}^H . $\mathbf{H} = [\mathbf{1} \ \mathbf{0} \ \mathbf{0}; \mathbf{0} \ \mathbf{1} \ \mathbf{0}]$
\mathbf{p}^{3D}	Vector that represent a point with coefficients $(x,y,z)^T$
\mathbf{h}	Vector belonging to the mapping between \mathbf{p}^{3D} and \mathbf{p}^H . $\mathbf{h} = [\mathbf{0}; \mathbf{0}]$
\mathbf{p}^V	Vector that represent a point in the horizontal plane with coefficients $(v,w)^T$
\mathbf{V}	Matrix belonging to the mapping between \mathbf{p}^{3D} and \mathbf{p}^V . Values of \mathbf{V} are fixed during the calibration procedure
\mathbf{v}	Vector belonging to the mapping between \mathbf{p}^{3D} and \mathbf{p}^V . Values of \mathbf{v} are fixed during the calibration procedure

Bezier curves and fitting process

$\mathbf{b}(s)$	Bezier curve evaluated in s where $0 \leq s \leq 1$
\mathbf{cp}_i	Control point $i = 0,1,2$ with coordinates $(x,y,z)^T$
j	whisker
f	frame
$E_v(f,j)$	Fitting of the whisker j in the frame f to the vertical image
$E_h(f,j)$	Fitting of the whisker j in the frame f to the horizontal image
$I_h(x,y)$	Intensity at the point (x,y) in the horizontal image
$I_v(v,w)$	Intensity at the point (v,w) in the horizontal image
R_1	Regularising term from the objective function that account for smooth movement of the whisker

	over time
R_2	Regularising term from the objective function that account for the shape of the whisker
σ_1	Selectable parameter that weight the first regularising factor R_1
σ_2	Selectable parameter that weight the second regularising factor R_2
\mathbf{q}	Vector produced by the subtraction of the positions of the furthest and nearest control points
Extraction of kinematic parameters	
\mathbf{i}'	Unit vector that point in the direction of x'
\mathbf{j}'	Unit vector that point in the direction of y'
\mathbf{k}'	Unit vector that point in the direction of z'
ζ	Rotation angle of the whisker respect to x'
θ	Azimuth angle defined as the angle between the x axis and the projection of tangent at the base of the whisker in the horizontal plane
φ	Elevation angle defined as the angle between the $-z$ axis and the projection of tangent at the base of the whisker in the vertical image
$\kappa_{3D}(s)$	3D Curvature evaluated in s
$\kappa_h(s)$	Curvature of the projection of the Bezier curve in the horizontal plane evaluated in s
$\kappa_v(s)$	Curvature of the projection of the Bezier curve in the vertical image evaluated in s

547

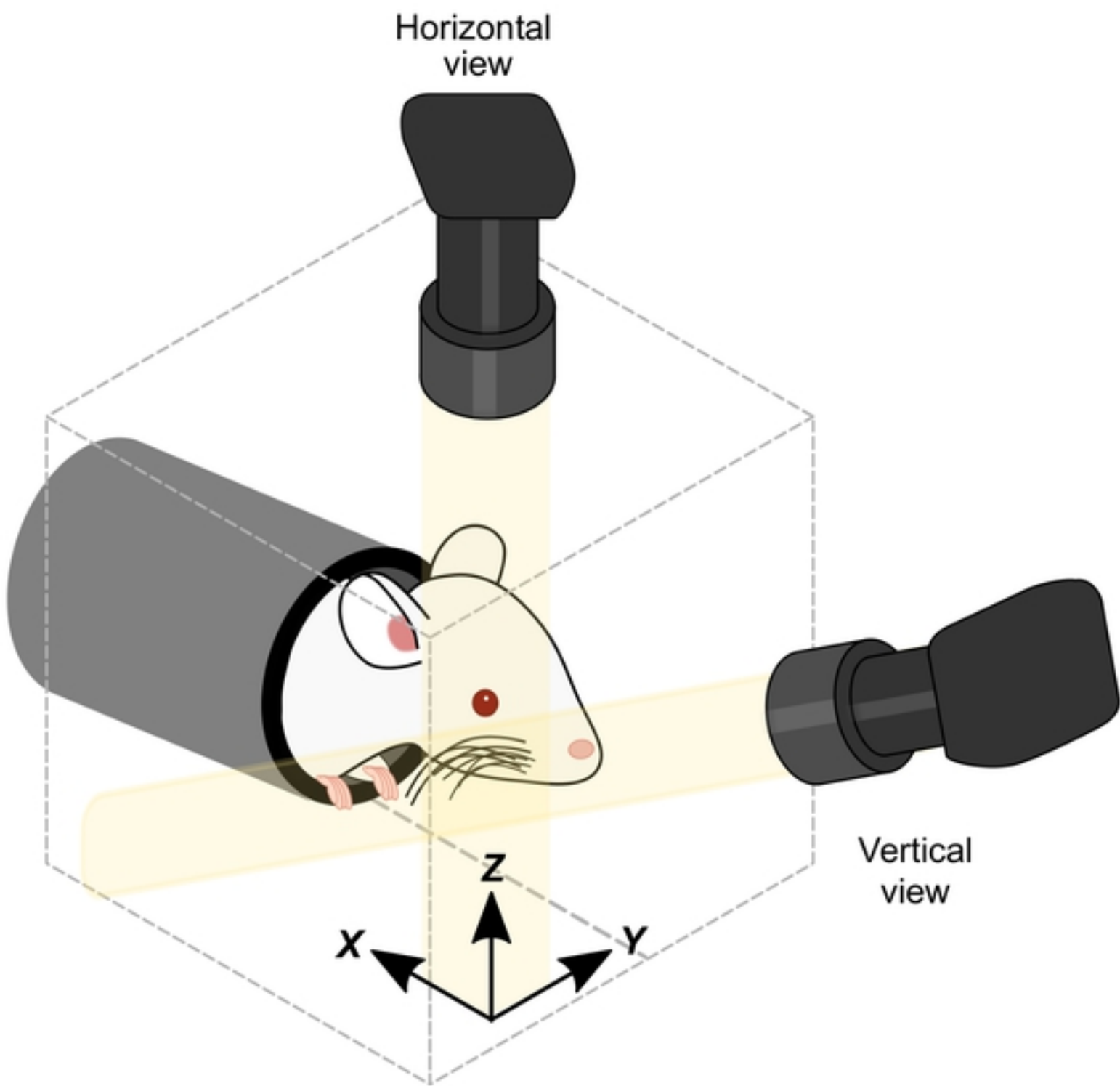
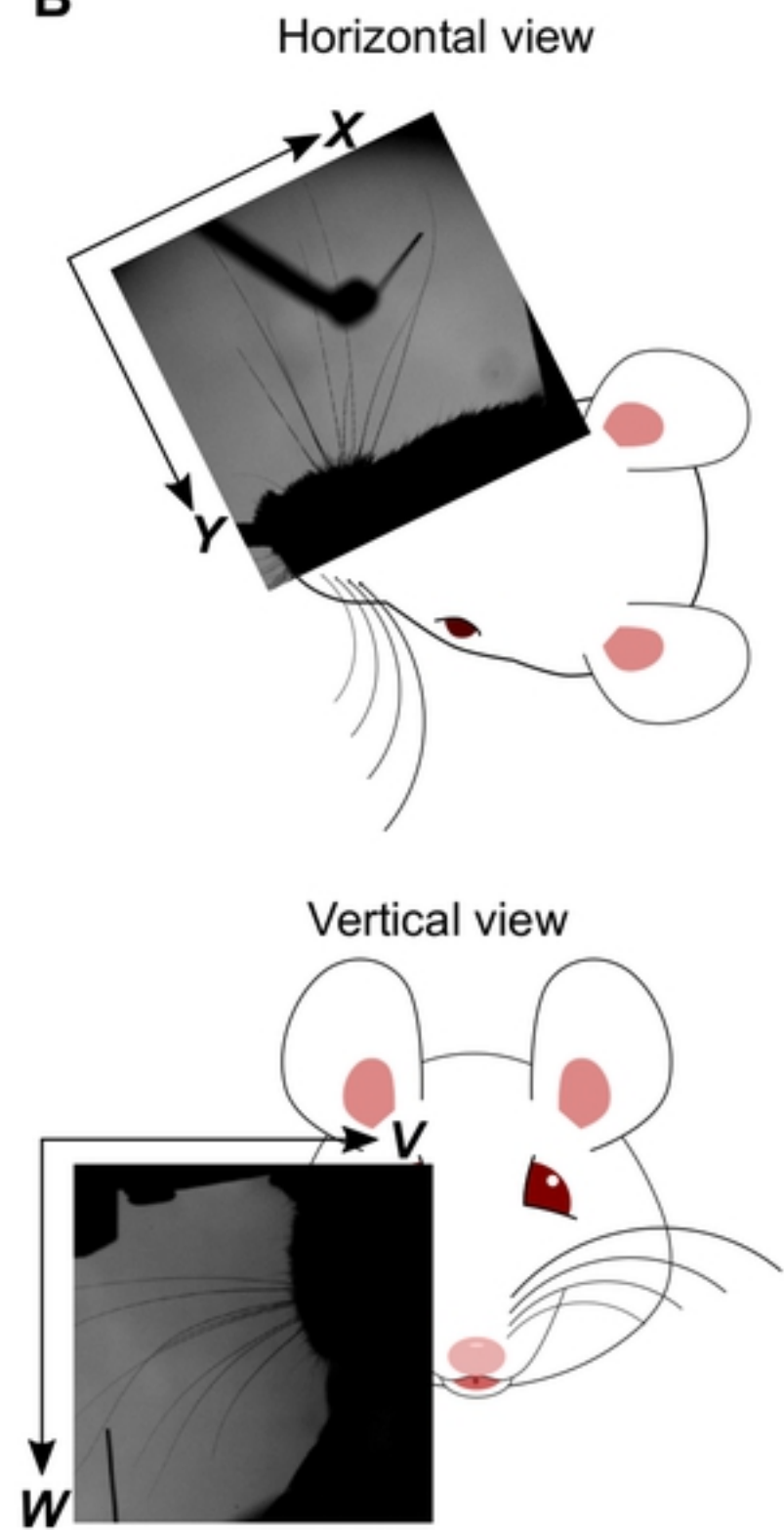
548 **References**

- 549 1. Tinbergen N. The study of instinct. The study of instinct. New York, NY, US: Clarendon Press/Oxford
550 University Press; 1951. xii, 237–xii, 237.
- 551 2. Brown AEX, De Bivort B. Ethology as a physical science. Nat Phys. 2018 Jul 9;14(7):653–7.
- 552 3. Maravall M, Diamond ME. Algorithms of whisker-mediated touch perception. Curr Opin Neurobiol.
553 2014;25:176–86.
- 554 4. Campagner D, Evans MH, Loft MSE, Petersen RS. What the whiskers tell the brain. Vol. 368, Neuroscience.
555 2018. p. 95–108.
- 556 5. Knutsen PM, Biess A, Ahissar E. Vibrissal Kinematics in 3D: Tight Coupling of Azimuth, Elevation, and
557 Torsion across Different Whisking Modes. Neuron. 2008 Jul 10;59(1):35–42.

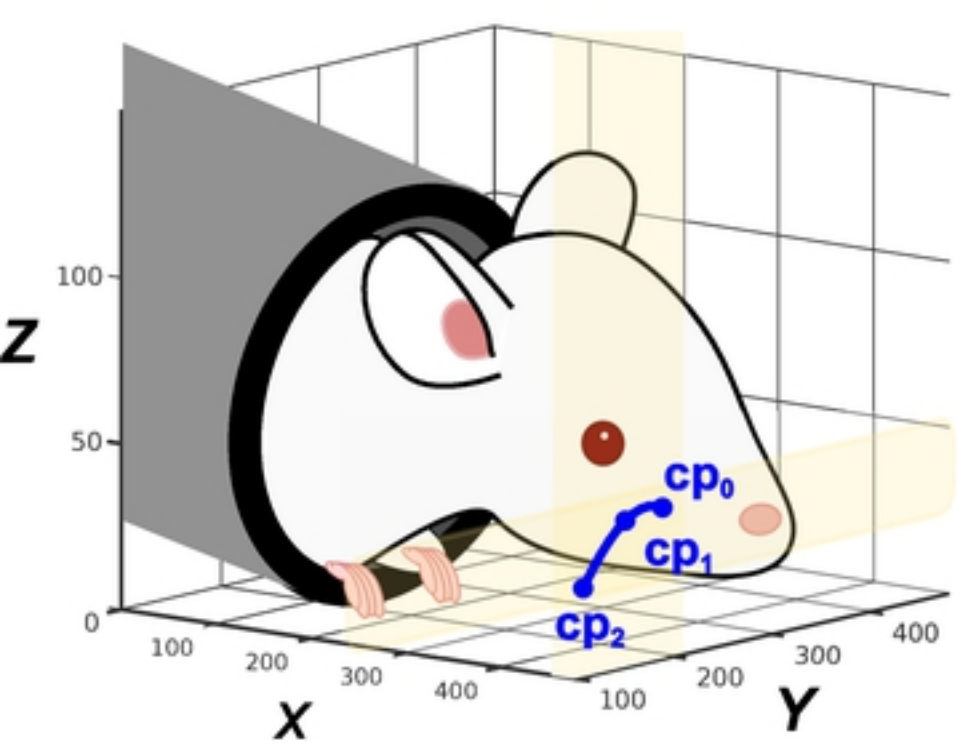
- 558 6. O'Connor DH, Peron SP, Huber D, Svoboda K. Neural activity in barrel cortex underlying vibrissa-based object
559 localization in mice. *Neuron*. 2010;67(6):1048–61.
- 560 7. Hires SA, Gutnisky DA, Yu J, O'Connor DH, Svoboda K. Low-noise encoding of active touch by layer 4 in the
561 somatosensory cortex. *Elife*. 2015 Aug 5;4(AUGUST2015):e06619.
- 562 8. Evans MH, Loft MSE, Campagner D, Petersen RS, Evans MH, Loft MSE, et al. Sensing the Environment With
563 Whiskers. In: *Oxford Research Encyclopedia of Neuroscience*. 2019. p. 1.
- 564 9. Campagner D, Evans MH, Chlebkova K, Colins-Rodriguez A, Loft MSE, Fox S, et al. Prediction of Choice
565 from Competing Mechanosensory and Choice-Memory Cues during Active Tactile Decision Making. *J Neurosci*.
566 2019 May 15;39(20):3921–33.
- 567 10. Zuo Y, Diamond ME. Rats Generate Vibrissal Sensory Evidence until Boundary Crossing Triggers a Decision.
568 *Curr Biol*. 2019 Apr 12;29(9):1415-1424.e5.
- 569 11. Bermejo R, Vyas A, Zeigler HP. Topography of rodent whisking - I. Two-dimensional monitoring of whisker
570 movements. *Somatosens Mot Res*. 2002;19(4):341–6.
- 571 12. Huet LA, Schroeder CL, Hartmann MJZ. Tactile signals transmitted by the vibrissa during active whisking
572 behavior. *J Neurophysiol*. 2015;113(10):3511–8.
- 573 13. Gibson JM, Welker WI. Quantitative Studies of Stimulus Coding in First-Order Vibrissa Afferents of Rats. 2.
574 Adaptation and Coding of Stimulus Parameters. *Somatosens Res*. 1983 Jan 10;1(2):95–117.
- 575 14. Lichtenstein SH, Carvell GE, Simons DJ. Responses of Rat Trigeminal Ganglion Neurons to Movements of
576 Vibrissae in Different Directions. *Somatosens Mot Res*. 1990 Jan 10;7(1):47–65.
- 577 15. Bale MR, Petersen RS. Transformation in the Neural Code for Whisker Deflection Direction Along the
578 Lemniscal Pathway. *J Neurophysiol*. 2009 Nov;102(5):2771–80.
- 579 16. Bermejo R, Houben D, Zeigler HP. Optoelectronic monitoring of individual whisker movements in rats. *J*
580 *Neurosci Methods*. 1998 Sep 1;83(2):89–96.
- 581 17. Wolfe J, Hill DN, Pahlavan S, Drew PJ, Kleinfeld D, Feldman DE. Texture coding in the rat whisker system:
582 Slip-stick versus differential resonance. *PLoS Biol*. 2008;6(8):1661–77.
- 583 18. Szwed M, Bagdasarian K, Blumenfeld B, Barak O, Derdikman D, Ahissar E. Responses of Trigeminal Ganglion

- 584 Neurons to the Radial Distance of Contact During Active Vibrissal Touch. *J Neurophysiol.* 2006 Feb;95(2):791–
585 802.
- 586 19. Pammer L, O’Connor DH, Hires SA, Clack NG, Huber D, Myers EW, et al. The Mechanical Variables
587 Underlying Object Localization along the Axis of the Whisker. *J Neurosci.* 2013;33(16):6726–41.
- 588 20. Peron SP, Freeman J, Iyer V, Guo C, Svoboda K. A Cellular Resolution Map of Barrel Cortex Activity during
589 Tactile Behavior. *Neuron.* 2015;86(3):783–99.
- 590 21. Birdwell JA, Solomon JH, Thajchayapong M, Taylor M a, Cheely M, Towal RB, et al. Biomechanical models for
591 radial distance determination by the rat vibrissal system. *J Neurophysiol.* 2007;98(4):2439–55.
- 592 22. Clack NG, O’Connor DH, Huber D, Petreanu L, Hires A, Peron S, et al. Automated tracking of whiskers in
593 videos of head fixed rodents. Prlic A, editor. *PLoS Comput Biol.* 2012 Jul 5;8(7):e1002591.
- 594 23. Bush NE, Schroeder CL, Hobbs JA, Yang AE, Huet LA, Solla SA, et al. Decoupling kinematics and mechanics
595 reveals coding properties of trigeminal ganglion neurons in the rat vibrissal system. *Elife.* 2016 Jun 27;5.
- 596 24. Campagner D, Evans MH, Bale MR, Erskine A, Petersen RS. Prediction of primary somatosensory neuron
597 activity during active tactile exploration. *Elife.* 2016 Feb 15;5(Feb 15):e10696.
- 598 25. Severson KS, Xu D, Van de Loo M, Bai L, Ginty DD, O’Connor DH. Active Touch and Self-Motion Encoding
599 by Merkel Cell-Associated Afferents. *Neuron.* 2017;94(3):666-676.e9.
- 600 26. Roy S, Bryant JL, Cao Y, Heck DH. High-precision, three-dimensional tracking of mouse whisker movements
601 with optical motion capture technology. *Front Behav Neurosci.* 2011;5:27.
- 602 27. Quist BW, Hartmann MJZ. Mechanical signals at the base of a rat vibrissa: the effect of intrinsic vibrissa
603 curvature and implications for tactile exploration. *J Neurophysiol.* 2012 May;107(9):2298–312.
- 604 28. Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, et al. DeepLabCut: markerless pose
605 estimation of user-defined body parts with deep learning. *Nat Neurosci.* 2018 Sep 20;21(9):1281–9.
- 606 29. Peron SP, Freeman J, Iyer V, Guo C, Svoboda K. A Cellular Resolution Map of Barrel Cortex Activity during
607 Tactile Behavior. *Neuron.* 2015 May 6;86(3):783–99.
- 608 30. Vanzella W, Grion N, Bertolini D, Perissinotto A, Zoccolan D. A passive, camera-based head-tracking system
609 for real-time, 3D estimate of head position and orientation in rodents. *bioRxiv.* 2019;599365.

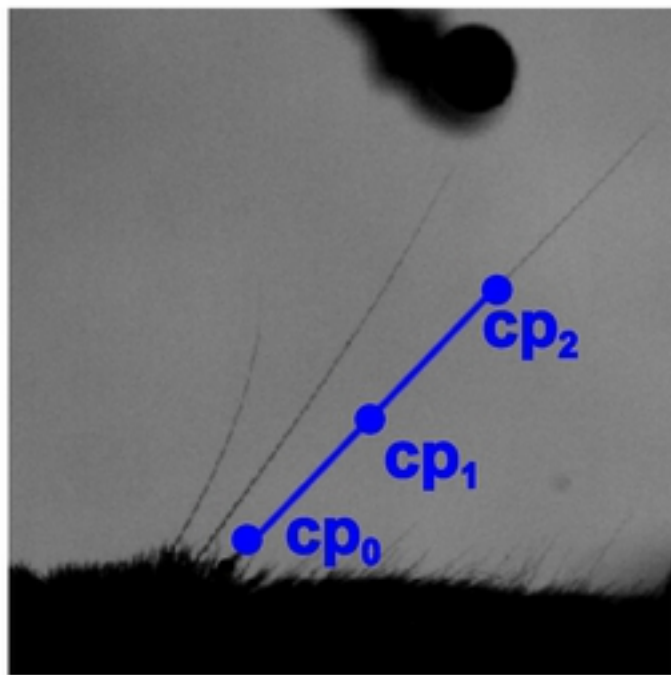
- 610 31. Knutsen PM, Derdikman D, Ahissar E. Tracking whisker and head movements in unrestrained behaving rodents.
611 J Neurophysiol. 2005;93(4):2294–301.
- 612 32. Hewitt BM, Yap MH, Hodson-Tole EF, Kennerley AJ, Sharp PS, Grant RA. A novel automated rodent tracker
613 (ART), demonstrated in a mouse model of amyotrophic lateral sclerosis. J Neurosci Methods. 2018 Apr
614 15;300:147–56.
- 615 33. Pasquet MO, Tihy M, Gourgeon A, Pompili MN, Godsil BP, Léna C, et al. Wireless inertial measurement of
616 head kinematics in freely-moving rats. Sci Rep. 2016 Dec 21;6(1):35689.
- 617 34. Goldstein, H., Poole, C. P., & Safko JL. Classical Mechanics. Vol. 28. Pearson Education; 2012. 638 p.
- 618 35. Marsh D. Applied Geometry for Computer Graphics and CAD. Springer; 2005. 350 p.
- 619 36. Yang AET, Hartmann MJZ. Whisking Kinematics Enables Object Localization in Head-Centered Coordinates
620 Based on Tactile Information from a Single Vibrissa. Front Behav Neurosci. 2016 Jul 19;10:145.
- 621

A**B**

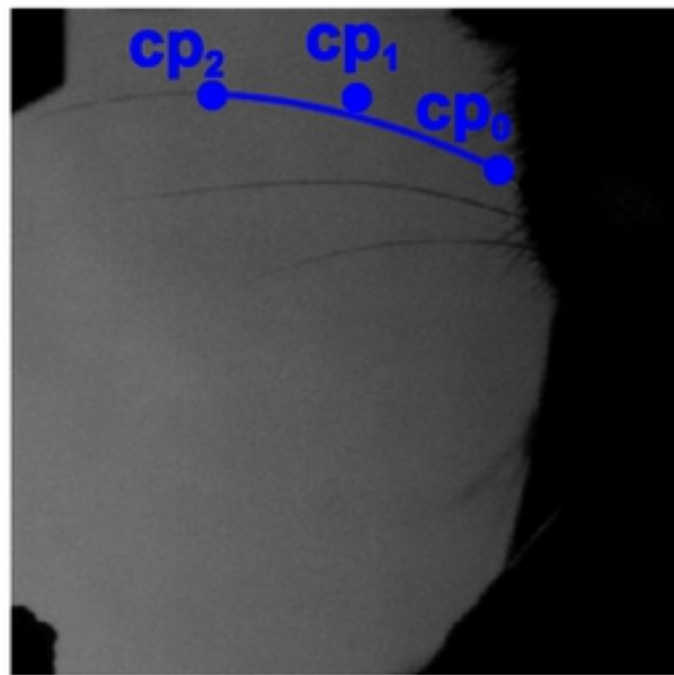
fig



Horizontal view



Vertical view



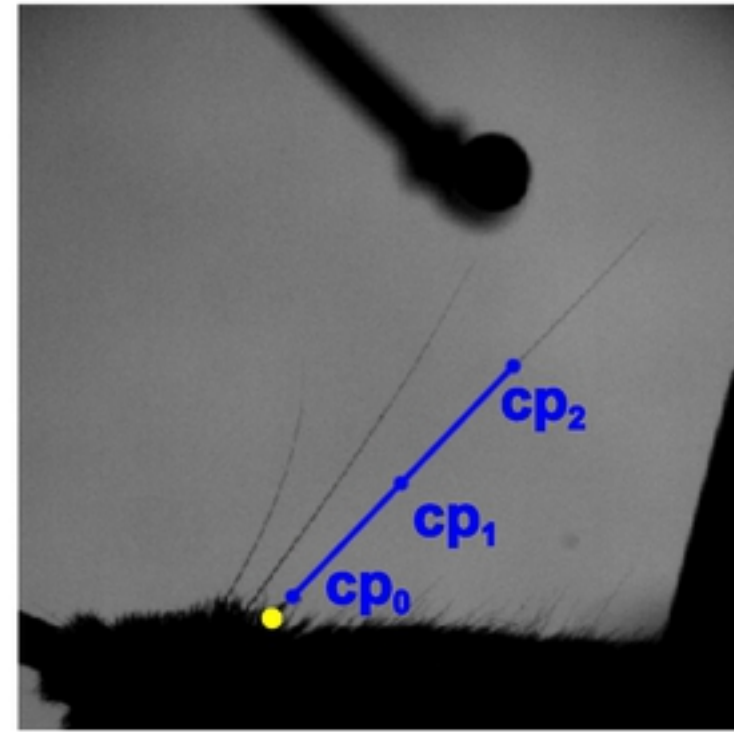
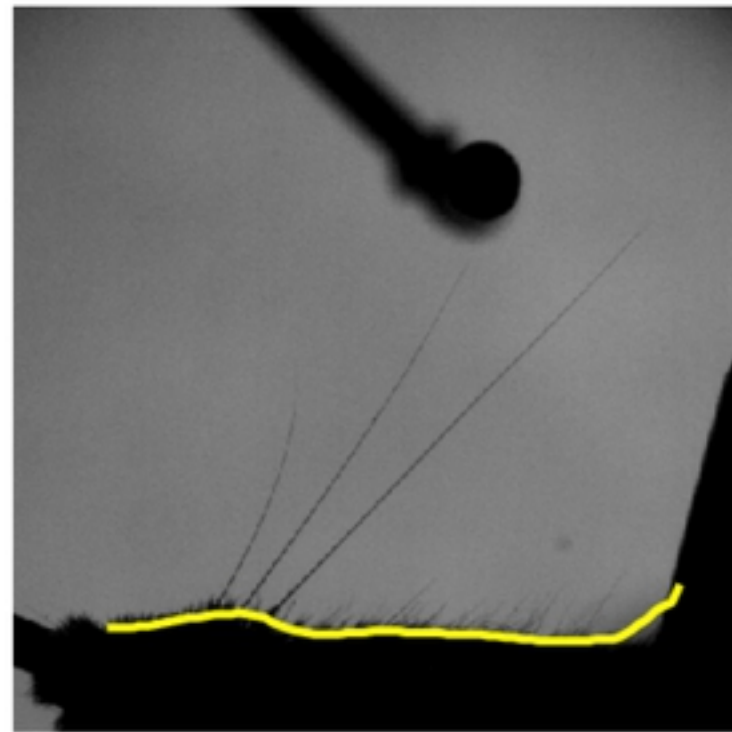
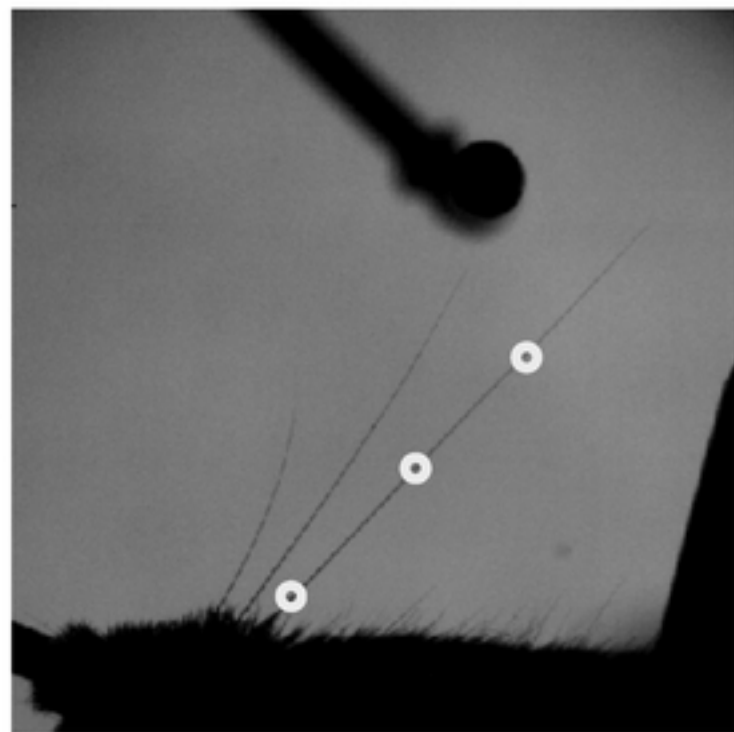
fig

Initialisation

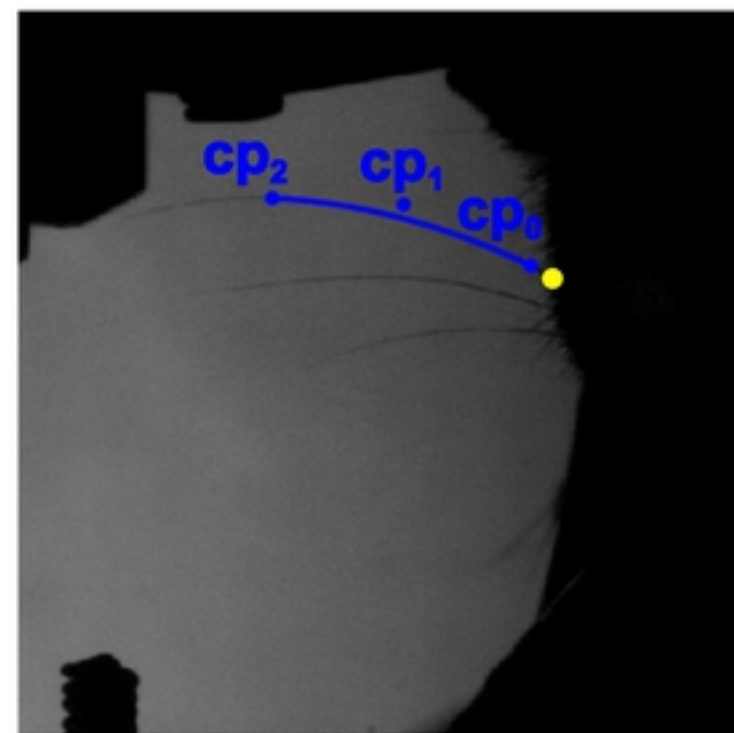
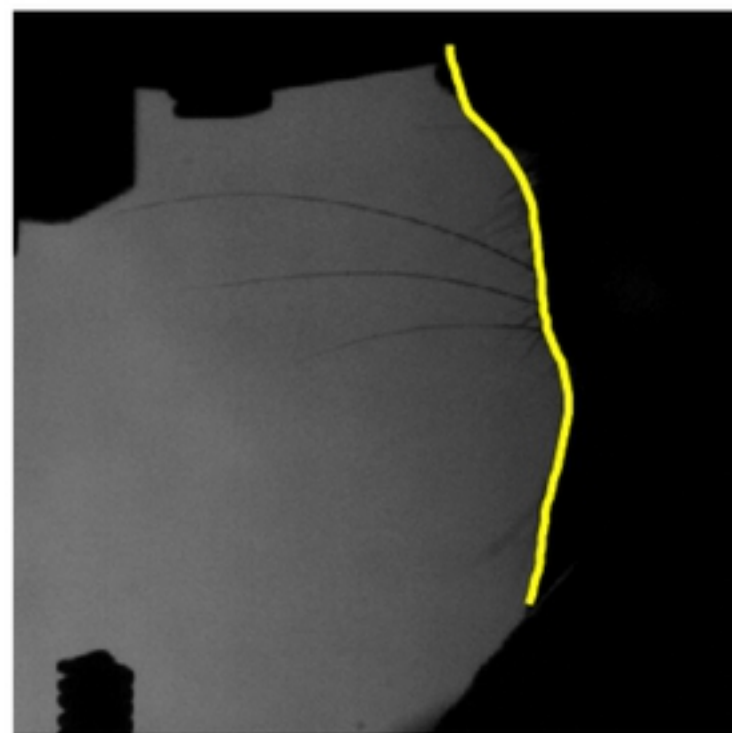
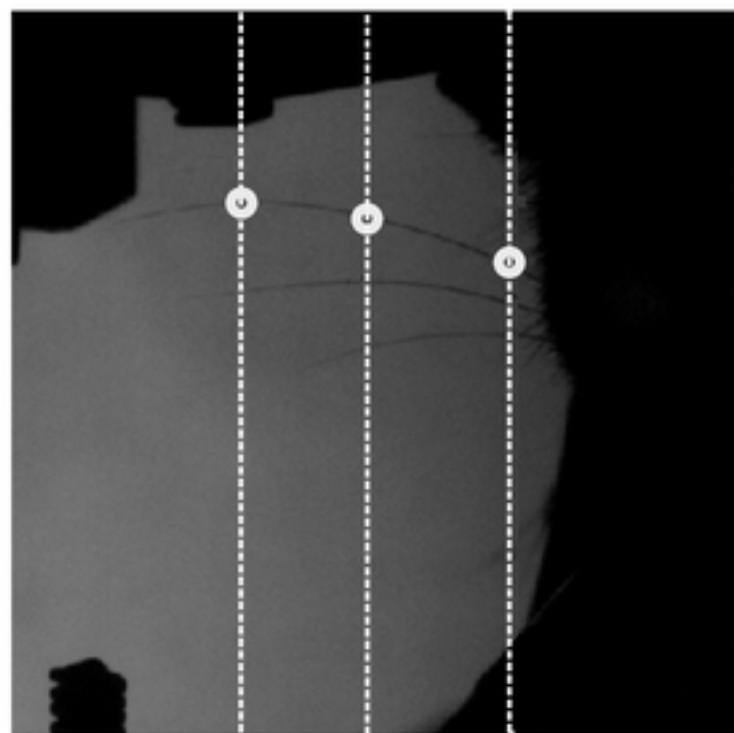
Snout contour detection

Bezier curve fitting

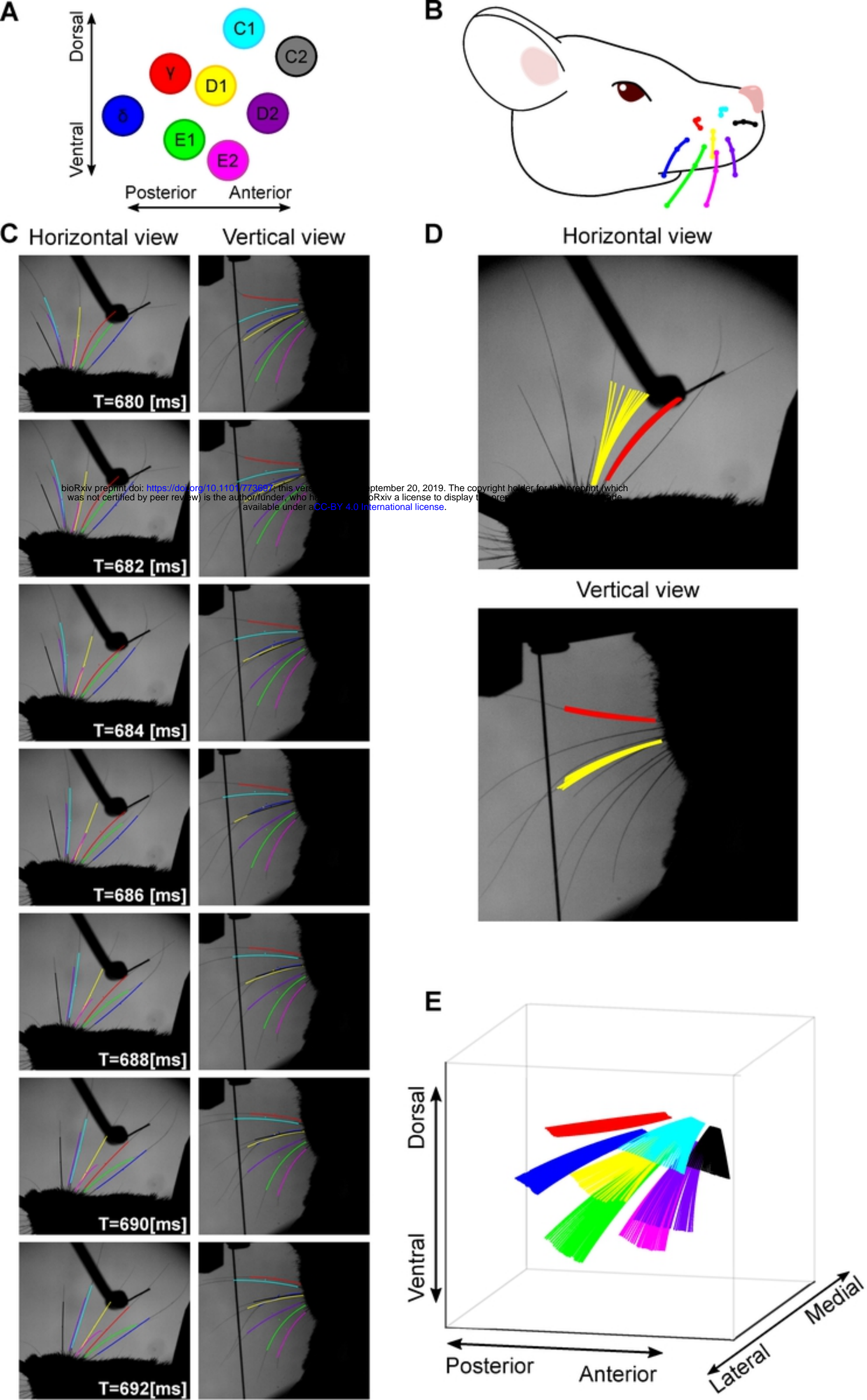
Horizontal view



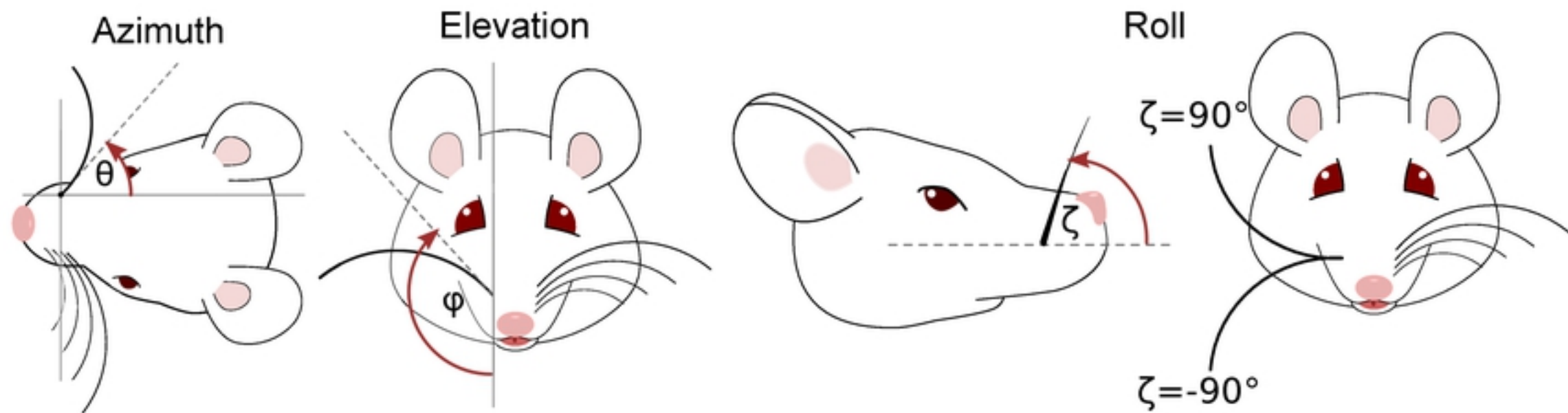
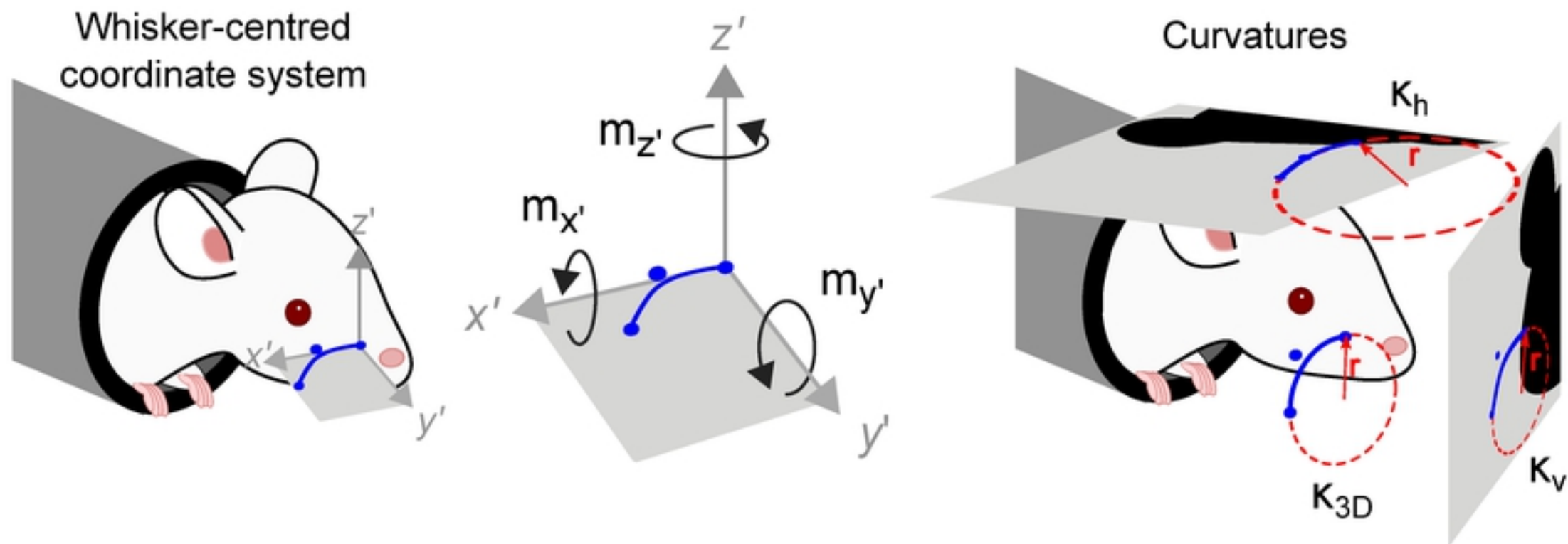
Vertical view

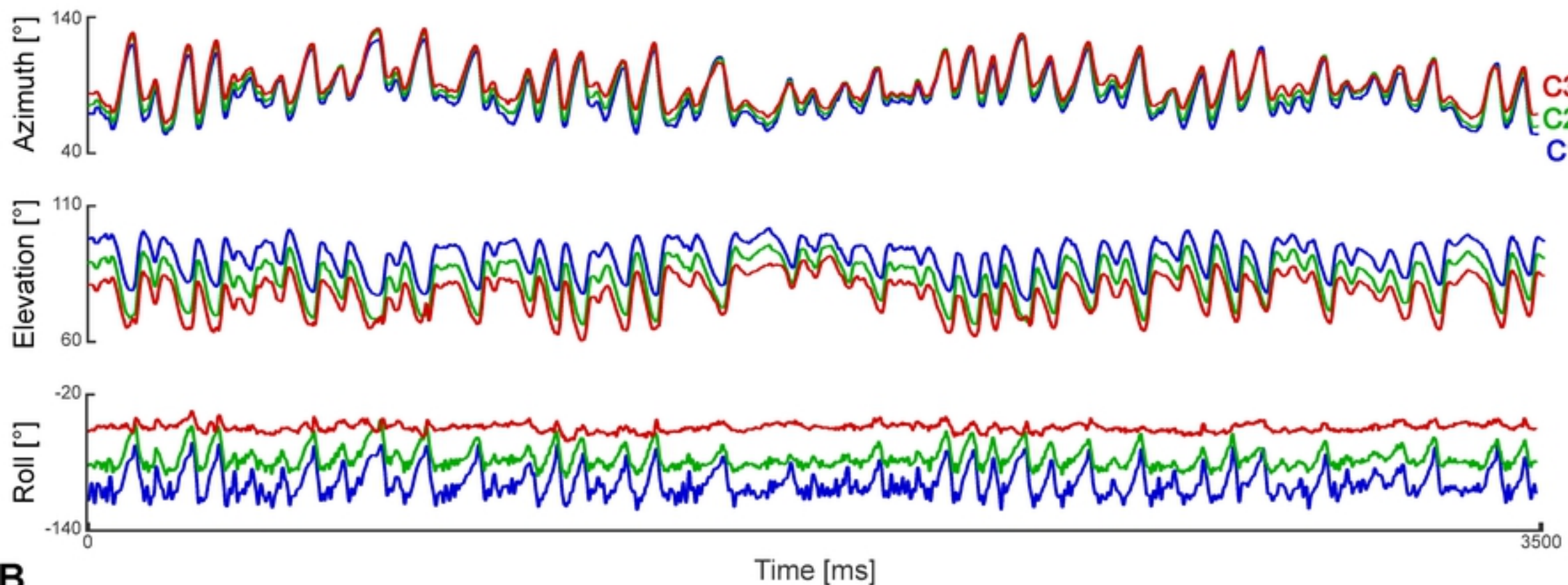
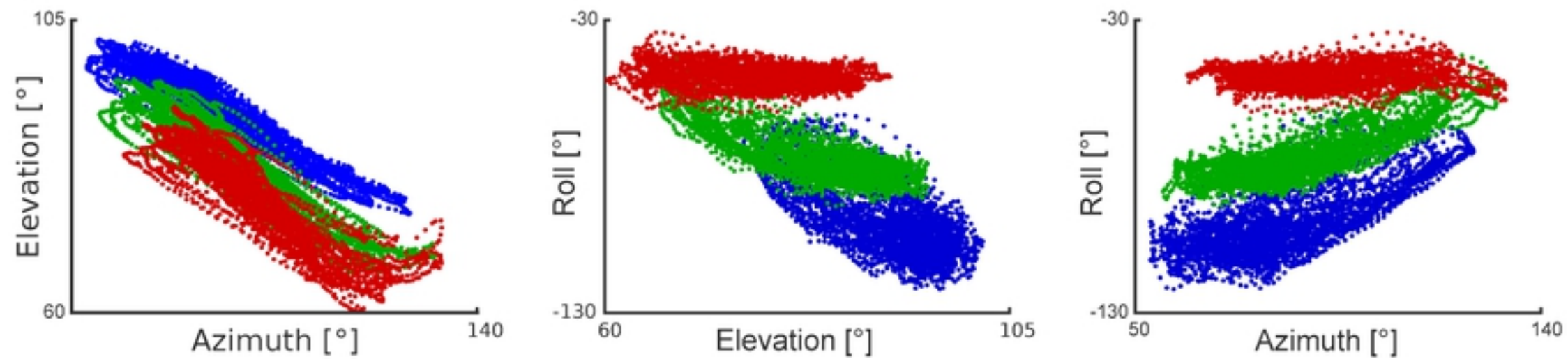


fig

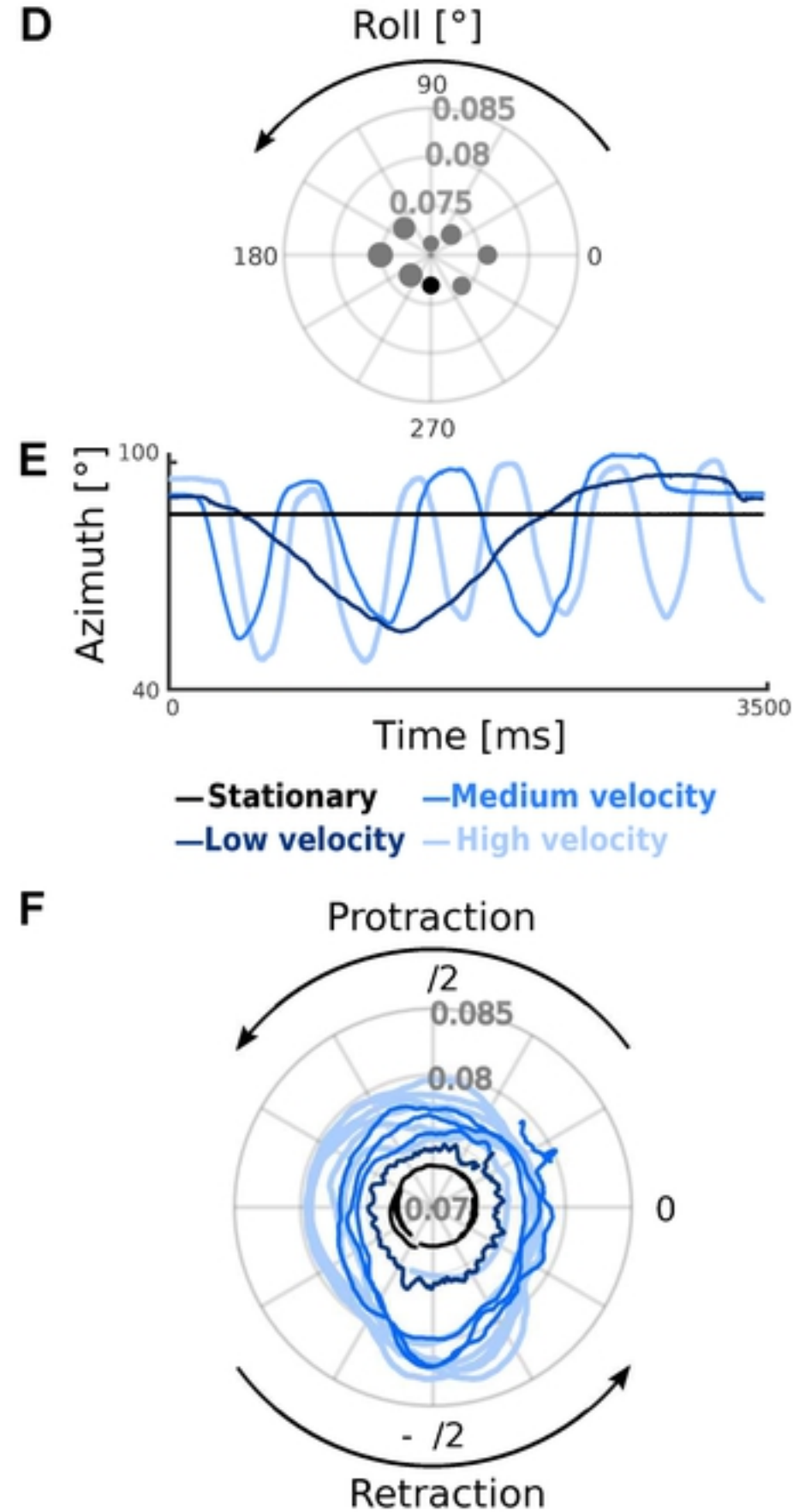
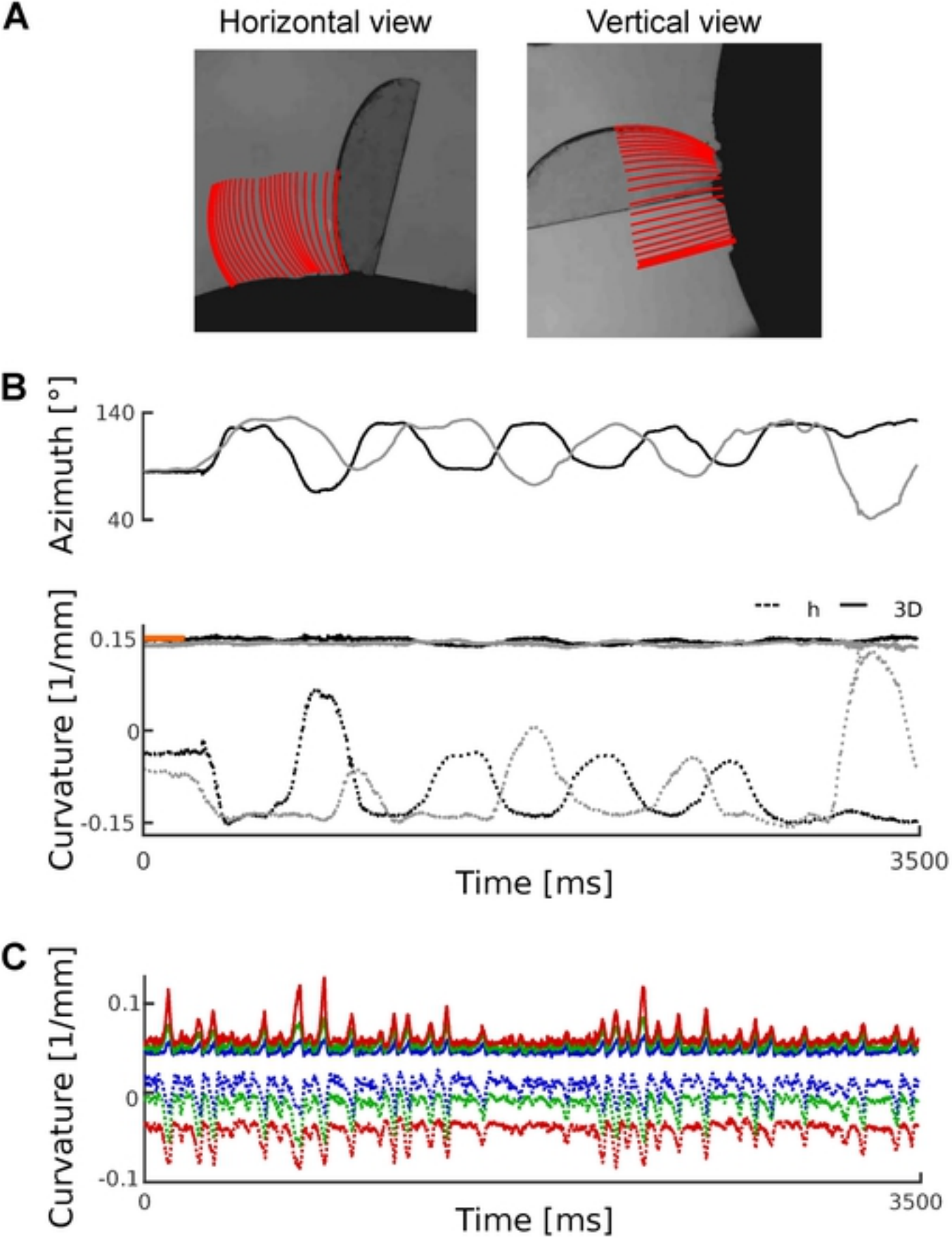


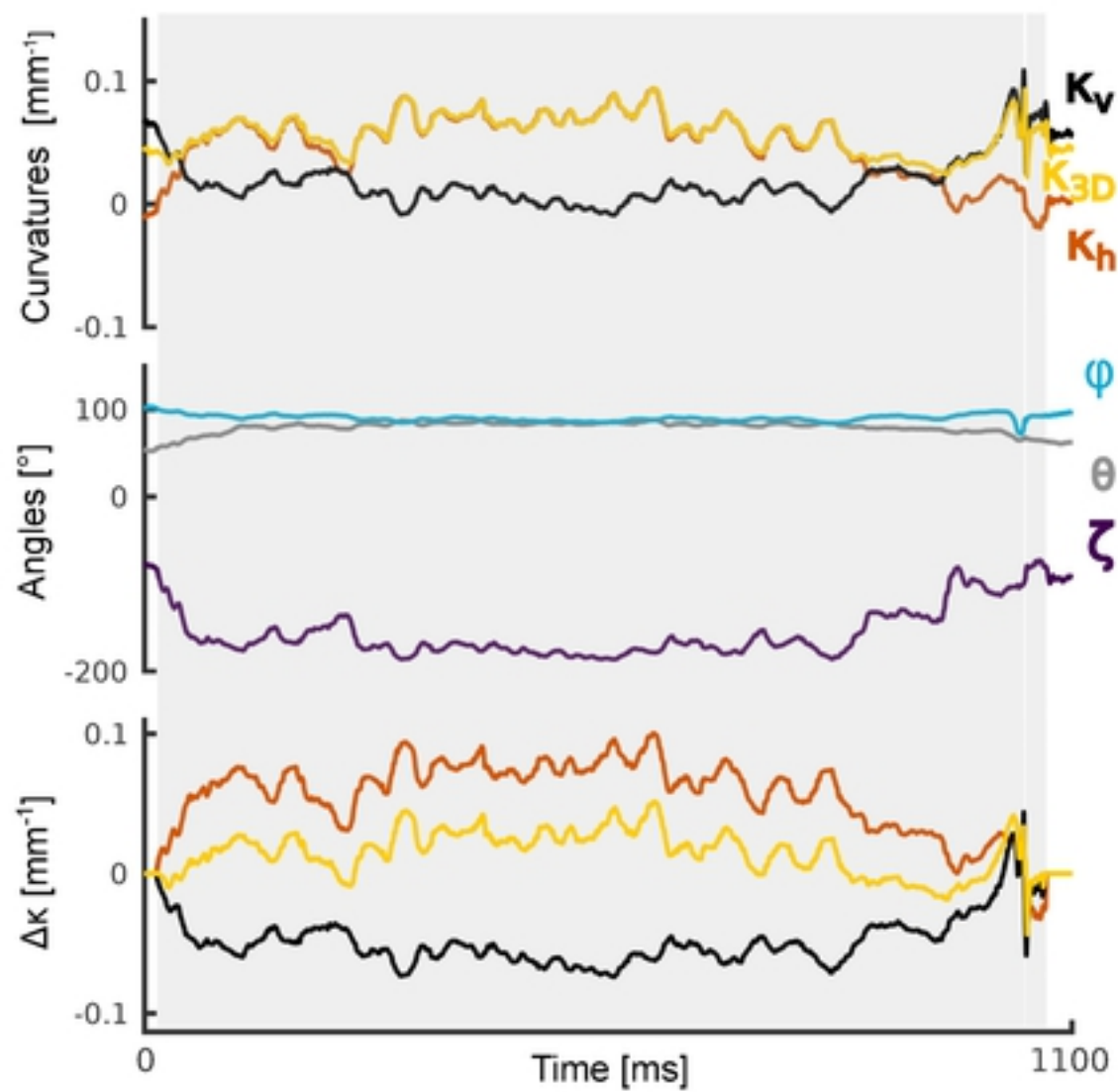
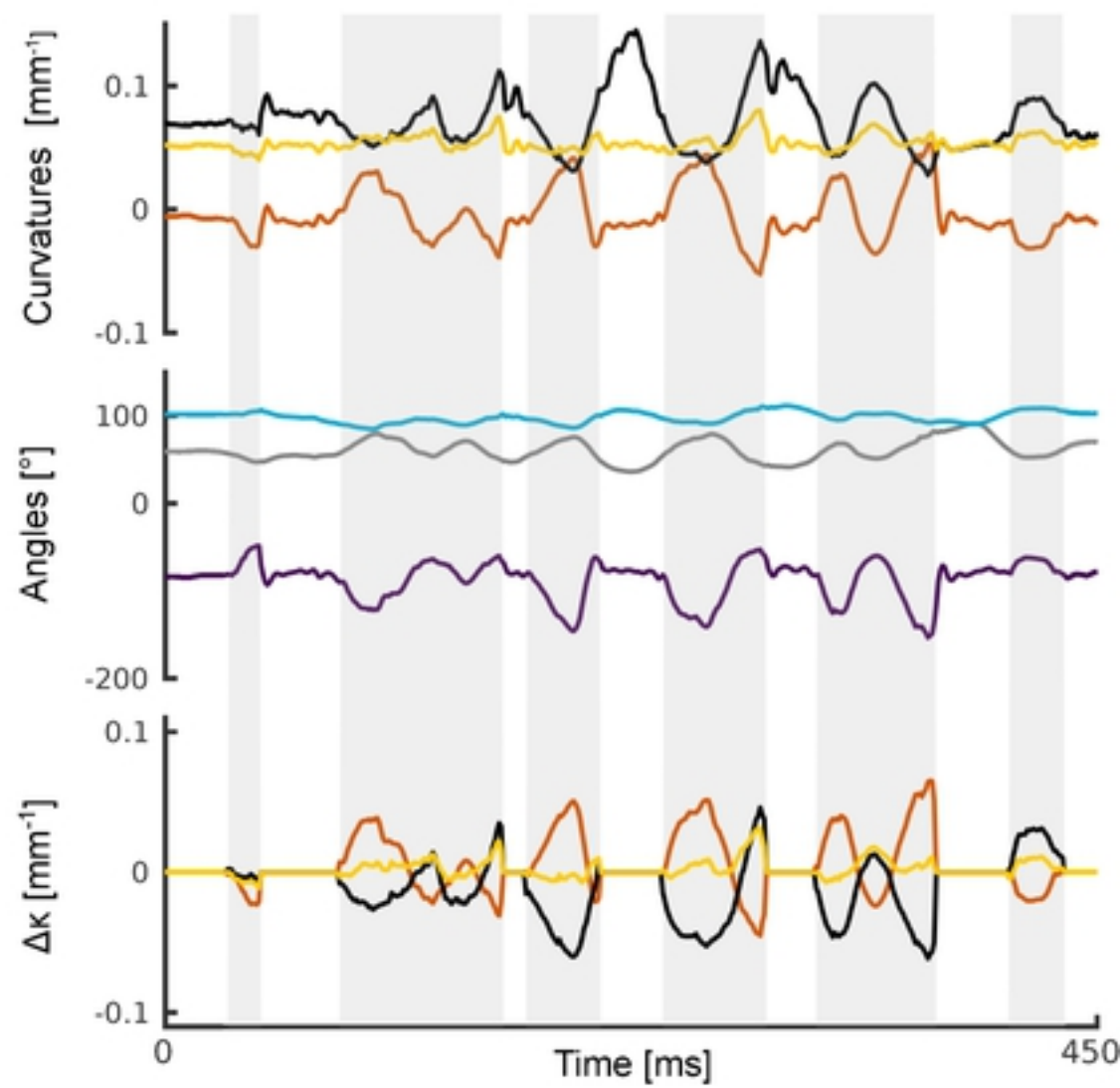
fig

A**B**

A**B**

fig



A**B**

fig