TITLE

Gene-by-environmental modulation of longevity and weight gain in the murine BXD family

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Abstract

Diet and environment profoundly modulate lifespan. We measured longevity as a function of diet and weight gain across a genetically diverse family of mice that segregates for over 6 million sequence variants. We followed 1348 females from both parental strains—C57BL/6J and DBA/2J—and 76 of their BXD progeny across their natural life span on a standard low fat chow diet (CD, 18% calories from fat) or on a high fat diet (HFD, 60% calories from saturated fat). On average, the HFD shortens lifespan by 85 days or 12%, roughly equivalent to an 8–10 year decrease in humans. This diet is also associated with an average two-fold higher age-adjusted risk of death compared to CD. However, variation among strains in the effect of HFD on longevity is high, ranging from a longevity loss of 54% in BXD65 to a gain of 37% in BXD8. Individual and strain-specific baseline weights and early weight gain are both associated with a mean decrease in longevity of ~4 days/g. By 500 days of age, cases on HFD gained four times as much weight as those on CD. However, strain-specific variation is again substantial and does not correlate well with lifespan. In summary, HFD has a negative effect on longevity, but genetic interactions are a much stronger factor (4% versus 30%). This highlights the unequivocal importance of genetic variation in making dietary recommendations.

Keywords

Longevity; high fat diet; body weight; genetic reference population; gene by environment (GXE)

Introduction

Longevity is among the most heterogeneous of traits. Differences in lifespan are dependent on complex innumerable gene-by-environmental (GXE) interactions (de Magalhães et al., 2012, Kuningas et al., 2008, McDaid et al., 2017, Hook et al., 2018). Nutrition, of course, has a profound influence on health and lifespan (Fontana and Partridge, 2015). Relative to an ad libitum diet, caloric restriction and intermittent fasting improve lifespan and health (reviewed by Heilbronn and Ravussin, 2003; Liang et al., 2018; Speakman et al., 2016). Effects of restricted diets and intermittent fasting are not entirely dependent on patterns of caloric intake, but depend on dietary macro- and micronutrient composition, the amount of time spent in different metabolic states, age of onset, sex, and perhaps of most importance to us, differences in genotype (Vaughan et al., 2018) and gene-by-diet interactions (Barrington et al., 2018).

The mouse is an excellent mammalian model for research at the interface of metabolism and aging, sharing most protein-coding genes with humans (Pennacchio and Rubin, 2003), but with a much shorter life cycle that enables longevity studies in controlled environments and under various experimental and dietary conditions (Miller et al., 2007), (Yuan et al., 2011), (Strong et al., 2013). However, most rodent studies do not incorporate the level of genetic complexity that is typical of human populations (Saul et al., 2019; Williams, 2006; Williams and Williams, 2017). Effects of DNA variants and dietary, drug, or environmental perturbations are usually studied on a single genome–often C57BL/6. This narrow focus greatly compromises translational utility of discoveries. To address this problem, we rely on the large family of BXD strains of mice that segregate for over 6 million variants (Peirce et al., 2004), (Wang et al., 2016). Collectively, the family also incorporates an impressive level of variation in phenotypes related to aging, metabolism, and gene expression in liver, muscle, brain, and many other tissues and cell types,(Williams et al., 2014), (Houtkooper et al., 2013), (Andreux et al., 2012), (Houtkooper et al., 2011), (Gelman et al., 1988), (De Haan and Van Zant, 1999).

Previous studies of BXDs (Gelman et al., 1988; Lang et al., 2010) demonstrate at least two-fold variation in lifespan on a standard diet—from approximately 12–15 months for the shortest lived strains to 30 months for the longest lived strains. In these studies conventional heritabilities of lifespan are as high as 25–45%, but the effective heritabilities (h^2_{RI}) that account for the depth of resampling ($n = 8$ to 12 replicates/genome) are as high as 80% (Belknap, 1998; Hook et al., 2018). The BXD family is particularly well suited to study GXE interactions because diverse but perfectly matched cohorts can be treated in parallel using different diets (Rikke et al., 2010, Hall et al., 2014, Andreux et al., 2012, Williams et al., 2016; Wu et al., 2014). The effect of genetic variation has been well studied in the context of dietary composition and caloric restriction on life span (Finkel, 2015, Keipert et al., 2011; Skorupa et al., 2008). However, key results remain

controversial. While caloric restriction is undoubtedly advantageous in boosting longevity *on average*, there is good evidence that such effects are not universal, and that certain individuals and genomes do not benefit in all environments (Barrington et al., 2018; Liao et al., 2010; Mitchell et al., 2016; Rikke et al., 2010).

In this study, we have measured longevity and body weight across a large cohort of fully sequenced, and highly diverse strains of mice that are part of the BXD family (Pierce et al., 2004; Ashbrook et al., 2019). We studied females in a well-controlled environment on two diets that differed greatly in fat content—those on a standard low fat chow diet (18% of calories from fat) and those on a high-fat diet (60% cal from fat). To the best of our knowledge this is the largest GXE experiment on the effects of high fat on longevity and weight changes, and includes matched data for 1348 cases and 76 BXD genotypes.
We address the following questions:

1. What is the average impact of a very high fat diet, otherwise well balanced for protein content, on longevity across the entire family?
2. To what extent does the strain genometype modulate effects of the high fat diet relative to the standard lower fat diet? Put another way: What is the strength of evidence in favor of GXE effects on longevity?
3. Does baseline body weight at young adulthood (~120 days) predict longevity or is the change in body weight in response to chronic high fat diet a stronger predictor of longevity?
4. To what extent is weight gain *per se* linked to a reduction in longevity and how does weight gain vary among strains?
5. Does diet itself modulate longevity after controlling for weight gain across the family or within strain?

Methods

Animals and Diets

Animals were raised and housed in a specific pathogen-free (SPF) facility at UTHSC (Memphis, TN), at 20–24 °C in temperature on a 12-hour light cycle. During the course of our study, serum samples from sentinel mice were tested quarterly for the following pathogens–Ectromelia virus, Epizootic Diarrhea of Infant Mice (EDIM), Lymphocytic Choriomeningitis (LCM), Mycoplasma pulmonis, Mouse Hepatitis Virus (MHV), Murine Norovirus (MNV), Mouse Parvovirus (MPV), Minute Virus of Mice (MVM), Pneumonia Virus of Mice (PVM), Respiratory Enteric Virus III (REO3), Sendai, and Theiler's Murine encephalomyelitis (TMEV GDVII). Semiannual necropsies are performed to test for endoparasites by microscopic examination of intestinal contents and anal tape preparations and ectoparasites by direct pelt microscopic examination. All such tests were negative throughout the experimental course.

The focus of this study is on the overall effects of diet on weight and longevity in the BXD family, rather than on the genetic control of longevity or weight *per se*. In some aspects, our study design is more like an observational prospective cohort rather than a controlled animal experiment—the main reason being that cases were entered into the aging colony and onto the HFD limb of the study at different ages. For this reason, we used methods of observational data analysis with minor modifications.

From October 2011 through to December 2018, animals from both parental strains, C57BL/6J and DBA/2J, and ~76 BXD strains were followed from their move from a large breeding colony into the aging colony (typically around 120 ± 66 days of age but with a wide range, from 26 days to 358 days) until their death. All animals were initially raised by dams on the chow diet. Females were aged in groups of up to 10 in polypropylene cages (935 cm^2) provisioned with Envigo Teklad 7087 soft cob bedding. Animals were provided either a standard low fat chow diet (CD, Envigo Teklad Global 2018, 18.6% protein, 18% calories from fat, 6.2% fat (ether-extractable), 3.1 kcal/g), or a widely used blue high fat diet (HFD, Envigo Teklad TD06414, 18.4% protein, 60.3% calories from lard, 37% saturated, 47% monounsaturated, 16% polyunsaturated fats, 5.1 kcal/g). Animals had *ad libitum* access to food and aquifer-sourced municipal tap water.

We studied a total of 1348 individuals ($n = 663$ on CD, $n = 685$ on HFD). Animals were labeled using ear tags, and individuals were randomly assigned to chow or high fat diet. Baseline weight was measured at entry into the study. 77% ($n = 527$) of animals started on HFD at ages between 50-185 days, but some started on the diet at ages as low as 26 days or as high as 358 days. Fewer than 2% of animals ($n = 12$) were placed on HFD at an age of greater than 365 days, and these were not included in the analysis. Less than

20% of animals were retired breeders that entered the study at 180+ days of age. Each animal was weighed to the nearest 0.1 gram every other month from start of diet until death. A separate subpopulation of 662 animals (n = 333 on CD, n = 329 on HFD) from matching BXD strains were sacrificed at specific time-points (6, 12, 18 and 24 months-of-age) for tissue collection across both diet cohorts (Williams EG et al., in submission). Organ weight data at 18 months of age from these animals were included in the analysis for this study. The aging colony at UTHSC is still in operation, but for this analysis we only consider animals with deaths between April 2012 and November 2018.

Longevity data from both cohorts in our aging colony (separate, combined and difference scores) are available in GeneNetwork.org (GN) under 'BXD RI Family' in the dataset 'BXD Published Phenotypes' (GN traits 18435, 18441, 19451, 19452, 21302, 21450). Body weight data at 6, 12, 18 and 24 months of age on both diets is also documented in GN (traits 19126, 19130, 19131, 19167, 19168, 19169, 19170, and 19171). Organ weight data on both diets, including liver, heart, kidneys and brain, at 18 months of age, can be found in GN as well (traits 20156, 20157, 20158, 20159, 20353, 20354, 20148, 20149, 20150, 20151, 20146, 20147).

The colony was moved to a new vivarium (TSRB) in April 2016. Approximately 60% of the animals lived and died in the original Nash annex vivarium, ~35% were born in the Nash but lived in both vivaria, and ~5% were born and spent their entire lives in the new facility. We carefully evaluated birth and death data over all seasons from both vivaria to successfully rule out any site-specific or seasonal effect on longevity (Supplementary data available). Animals were inspected daily and deaths were recorded for each animal with a precision of one day. Moribund animals (~10%) were euthanized, and those above the age of 200 days were included in longevity calculations. Criteria for euthanasia were based on an assessment by our veterinary staff following AAALAC guidelines.

Most animals were fixed by immersion in 10% neutral buffered formalin within 24 hours after death. The body cavity was opened prior to immersion to improve tissue preservation. Evenly balanced cohorts on the diet were selected based on fixation quality for necropsy with histopathology of tissues. A board-certified veterinary pathologist (RWR) performed necropsies and judged probable cause of death and other morbidities.

All experimental procedures were in accordance with the Guidelines for the Care and Use of Laboratory Animals published by the National Institutes of Health and were approved by the UTHSC institutional Animal Care and Use Committee.

Statistics

Longevity and body weight data were stratified by diet and by strain. Effects of different diets and body weight on longevity were analyzed using a random-effects model in R using the metafor package (Viechtbauer, 2010) and a mixed-effects Cox proportional hazard model using Therneau's *coxme* R package 2.2.-10 (CRAN.R-project.org/package =coxme) (Therneau and Grambsch, 2000). Survival analyses were performed using the survival package for R and the data were right-censored (see Fig 2, censored cases CD $n = 32$, HFD $n = 80$). Survival curves were computed by ANOVA and regression analyses were performed using R. Results were also tested using Wald test, likelihood ratio test, and Wilcoxon test.

Results

High fat diet shortens lifespan but with considerable variation among strains

Sets of females from 76 strains were assigned to HFD diet at an average of 120 days of age (Figure 1A). The HFD has a very significant average effect on lifespan across the family as a whole ($p<2.2E-16$, $r =$ 0.2). Mean lifespan decreases from 690 ± 8 SE (±199 SD) days on the control diet to 605 ± 6 SE (±169 SD) days on the HFD (Figure 1B). Median longevity decreases 77 days—from 703 to 626 (Figure 2A). Assuming linear scaling and that a dietary change was imposed in humans at about 20 years-of-age, this 77 to 85 days difference would scale roughly to a 8-10 year loss of longevity in humans (Flurkey K, Currer JM, Harrison DE. 2007).

Figure 1 (**A**) Balanced sets of females from 76 BXD strains were assigned to the low fat chow diet (CD) or the high fat (HFD) diet and weighed every two months. (**B**) High fat diet modulates longevity. When grouped by diet—and irrespective of genetic background—median lifespan of CD cohort exceeds by 77 days that of the HFD cohort (see *box plot* inset). Red and blue dots represent individual cases on CD and HFD.

Using a mixed-effects Cox model with diet as a fixed effect and strain as random effect, we estimated a hazard ratio of 2.0, indicating that animals on HFD have two-fold higher age-adjusted risk of death after the dietary change compared to CD-fed animals. The hazard ratio is relatively constant throughout the study and there is no crossing of the two cumulative hazard curves (Figure 2B, $p = 0.47$).

While HFD decreases lifespan at the family level, individual strains show significant differences in how they react to diet. The parents exemplify the difference—the DBA/2J paternal strain shows little or no effect of diet on longevity, whereas the C57BL/6J maternal strain loses 76 days on the HFD (Figure 2C). Some BXD progeny strains even live longer on the HFD, demonstrating that the longevity-diet relation is modulated by a GXE component (Figure 2D). Overall, longevity in 21 strains out of 67 was significantly

affected by HFD in either direction at a nominal *p* threshold of 0.05. To correct for multiple testing (*n* = 67) we computed *q* values (see Figure 2D * symbol for *q* value < 0.1) and with this correction, only 15 strains have significantly different longevity. Interestingly, BXD8 has significantly improved longevity on a high fat diet, with a median increase in longevity of 208 days (*t* = 4.0, *p* = 0.0052, q<0.05, two-tailed), with one other strain trending in the same direction—BXD172 (146 day increase on HFD, *p* = 0.073), but with a high q value of 0.85. As expected, high fat diet reduces longevity for most strains (Fig 2D, right side)—most prominently for BXD65, with a decrease in median longevity of nearly a year (345 days, *t* = 9.3 *p* = 9.0E-7, q = 6.0E-7).

Figure 2 (**A**) Diet influence on longevity. Cases on the HFD have a two-fold higher risk of death compared to those on the CD. (**B**) Cumulative hazard curves by diet do not cross and the hazards ratio of 2.0 is relatively constant throughout the study. (**C**) The lifespan of C57BL/6J, but not DBA/2J, is influenced by diet. Numbers in parentheses are median lifespans in days. (**D**) Longevity on a high fat diet depends strongly on strain. Red points represent longevities of cases on CD and blue points those on HFD. Lines represent median survival. Grey bars represent the difference in median survival on the diets. The few negative values to the left indicate higher survival on a HFD. Parental strains and F1 are denoted by bold italics font. Asterisks in bars denote significant FDR scores at a q value of 0.1. Censored cases in A– C are still alive and are marked by + signs.

Body weight at young adulthood is a strong predictor of longevity

Animals were weighed regularly throughout their lifespan. As expected, animals on the high fat diet gained more weight on average than those on the chow diet (Figure 3A). Initial body weight measured at entry into the aging colony—on average at 120 days—has a significant influence on eventual lifespan after adjusting for differences in age at baseline weight ($p <0.001$, $r = 0.1$) (Figure 3B). A one gram increase in initial baseline weight is associated with a 5-day loss of longevity across all strains. At this early stage there is by design no significant weight difference between cases assigned to the HFD (23.11 ± 0.22 SE g, $n = 685$) and those continuing on the CD for the remainder of their lives (23.26 ± 0.22 g, $n = 659$). Of interest, the slope of –5 days/g of body weight at ~120 days is not affected by the subsequent diet (note the parallel red and blue lines in Figure 3B). This demonstrates that the initial size-longevity relation is relatively insensitive to GXE.

Early body weight gain is associated with a reduction in longevity

Body weight measured after 100 days on both diets correlates negatively with longevity (Figure 3C), with a one gram increase now corresponding to a decrease of 4 days ($r = 0.3$). This effect is observed even after adjusting for strain differences. Overall, 53 out of 67 strains had significant weight gain on the HFD after 100 days ($p <0.05$, $q<0.1$, t values ranging from –3.03 to –13.43). Looking at change in body weight after 100 days on diet, early body weight gain in response to the high fat diet, but not the chow diet, is negatively correlated with longevity ($p = 0.004$, $r = 0.1$) (Figure 3D).

Diet significantly alters longevity after adjusting for weight gain

We chose to focus on two time points for body weight analyses- 100 days (early weight gain on HFD) and 400 days on diet (~highest measured weight point on both diet curves). The mean weight of the population plateaus around 500 days of age and then begins to decline, for both diets. By 500 days of age animals had been on HFD for 400 ± 44 days and gained an average of 29.5 g. Those on the CD gained only 6.2 g (mean weight on CD = 29.7 ± 0.35 SE g, $n = 447$; mean weight on HFD = 52.6 ± 0.63 SE g, $n = 447$). Surprisingly, the substantial increase in body weight on the HFD did not significantly correlate with lifespan (Figure 3E). Adjusting for strains, 10% of the effect of diet on longevity is mediated through body weight gain. Mirroring this interesting observation, sustained weight gain after 400 days on high fat diet (Figure 3F) has no predictive value, emphasizing that in our study the diet itself, rather than weight gain, modulates longevity. Overall, 45 out of 57 strains had significant weight gain upon eating a high fat diet for 400 days ($p<0.05$, $q<0.1$, t- values ranging from –3.03 to –13.14) (Figure 3G).

Figure 3 (**A**) Body weight by diets and age. Asterisks denotes significance at p <0.05 and <0.001. Note the pronounced decline in body weight on both diets after about 500 days. (**B**) Initial body weight (baseline weight at entry into colony, mean age of ~120 days) has a modest but consistent negative slope with longevity (–5 days/g, p<0.001) that is not exacerbated by the HFD. (**C**) Body weight after 100 days on the two diets (~260 days of age) still correlates negatively with longevity (–4 days/g, p <0.001, see line labeled c in Panel A). (**D**) Early weight change in response to HFD (blue line), measured at 100 days on diet, is

negatively related with longevity (–4 days/g, p = 0.004), but this is not true of cases remaining on CD. (**E**) Surprisingly after 400 days on diet (~500 days of age) current body weight does not predict variance in longevity (see line labeled d in Panel A). (**F**) Substantial weight change after 400 days on HFD (blue line) is now not predictive of longevity. (**G**) Strain-wise changes in median weight after 100 days on diets. Red points represent longevities of cases on CD and blue points those on HFD. Lines represent median survival. Grey bars represent the difference in median survival on the diets. Parental strains and F1 are denoted by bold italics font.

Insufficient evidence of association between longevity and major metabolic organ weight

We compared organ and tissue weights of a separate subsample of animals on the two diets at 500 days. HFD mice (*n* = 63) had 84% greater fat mass, 25% greater heart mass, 19% greater liver mass, and 18% greater kidney mass at 500 days compared CD controls (*n* = 71). However, HFD did not influence brain mass. No significant correlations were found between longevity and any of these organ weights at 18 months in this small subsample (Supplemental figures S1A-F).

Major morbidities contributing to death in the aging colony

We carried out gross necropsy with or without histopathology included for 76 animals (69 with a single cause of morbidity and 7 with multiple causes) from 45 strains on CD and 79 animals (71 with a single cause of morbidity and 8 with multiple causes) from 43 strains on HFD. Likely cause of death was clear at necropsy or following histopathology in 87% of animals. Hematopoietic neoplasia—lymphomas and histiocytic sarcomas—was a leading cause of death, accounting for ~35% of deaths in both cohorts. Miscellaneous non-neoplastic conditions causing significant morbidity or death were detected in 24% of CD and 32% of HFD-fed animals.

Preliminary evidence points towards the influence of diet on causes of morbidity and mortality (Table 1). The HFD cohort displays an increased prevalence and severity of cardiovascular disease and lesions (atrial thrombosis, cardiomyopathy and cardiac dilation sometimes with hepatic steatosis and centrilobular atrophy). Nineteen percent of HFD mice (15 out of 79) have heart pathologies rated moderate to severe whereas only 1 out of 76 CD mice had a heart lesion rated at least moderately severe (2-tail Fisher's exact test *p* = 0.0003). Four HFD, but no CD, cases had systemic polyarteritis. In contrast, some pathologies appear to be more pervasive in the CD cohort. Thirty-six percent of CD mice displayed non-hematopoietic malignant neoplasia (27 of 76 CD–11 sarcomas, 15 carcinomas, and 1 teratoma) compared to 13% HFD

mice with non-hemopoietic malignancy (10 of 79 HFD–6 sarcomas and 4 carcinomas); this preliminary finding is nominally significant (Fisher $p = 0.0012$, not corrected for multiple comparisons).

Table 1. Major morbidities contributing to death in BXD family

Morbidity Type	Diet	n with single morbidity	n with multiple morbidities
Unnatural causes	CD	7	1
	HF	3	0
Non-neoplastic conditions	CD	16	2
	HF	22	2
Heart/ cardiovascular	CD	0	1
	HF	8	7
Polyarteritis (autoimmune)	CD	0	0
	HF	3	1
Lymphoma	CD	25	3
	HF	26	1
Sarcoma	CD	9	2
	HF	4	2
Carcinoma	CD	12	3
	HF	4	0
Renal	CD	0	1
	HF	1	3
Teratoma	CD	0	1
	HF	0	0

Unnatural causes = flooded cage, foot injury in wire lid, eye abrasion leading to euthanasia etc.

Miscellaneous non- neoplastic conditions = inflammation, infection, amyloidosis, ulcerative dermatitis

Heart = major contribution of cardiovascular failure other than polyarteritis

Polyarteritis = systemic multifocal arteritis suggestive of autoimmune disease

Lymphoma = hematopoietic neoplasia including lymphoma and histiocytic sarcoma

Sarcoma = nonhistiocytic sarcoma, spindle cell, hemangiosacoma, leiomyosarcoma, sarcoma NOS

Carcinoma = carcinomas (hepatocellular, squamous cell)

Renal = renal failure due to severe nephropathy

Discussion

We have studied effects of a high fat diet on aging and lifespan in the BXD family of mice. Our focus here is on overall and strain-specific effects on longevity and weight gain. To our knowledge, this is the largest GXE experimental population study of diet and longevity in any mammal. However, its outcomes are female–specific and we assume the same will be true for males (Andreux et al., 2012; Williams et al., 2016). In the introduction, we pose five questions for which we now have good answers: 1. Yes, as expected a diet that is very high in saturated fat—lard, in our case—reduces longevity by an average of 12%. As in humans, the risk of cardiovascular disease is significantly higher. 2. However, this is not a universal response, and GXE effects are strong for both longevity and weight gain. Even after correction for multiple comparisons one strain of mouse lives significantly longer and another strain gains no weight on the HFD. 3. We confirm that lower weight at an early age is linked to longer longevity, and that this effect is also true on the HFD. 4. There is at best only a modest link between weight gain after maturity and longevity. Weight gain late in life—between ages of 12–18 months—accounts for minimal variation in longevity. 5. Finally, diet modulates longevity and has a stronger effect on longevity than weight gain itself. By far the strongest effect on longevity and weight gain is genotype.

GXE and longevity

In the BXD family, we find that a diet rich in saturated fats shortens lifespan by an average of almost three months, roughly scaling to 8–10 year decrease in humans. This diet is associated with an average two-fold higher age-adjusted risk of death compared to the chow-fed controls. Longevity under the two diets correlates moderately ($r = 0.60$, GN traits 18435 and 18441). However, strains display remarkably wide variation in responses to diet, and despite the strong effect, diet accounts for just 4% of the total variance in longevity. In comparison, strain accounts for 30% of variance. Strain longevity combined across the two diets varies from 307 ± 37 days in BXD13 ($n = 21$) to 852 ± 33 days in BXD168 ($n = 23$). Some strains are fully resistant to the negative effects of HFD on body weight and lifespan while others are strongly affected, indicating substantial gene-by-diet interaction effects. While mean longevity on high fat is shortened by an average of 10%, genetic factors account for roughly two-fold range in life span. At least one strain, BXD8, actually has significantly improved longevity on the HFD (+37% on HF). Other strains such as BXD16 and BXD73, are immune to the high fat challenge with respect to change in life span. Of course, longevity of most strains is adversely affected (n = 67) and in the case of BXD65 lifespan is cut in half. Weight gain is characterized by a similar GXE effect—at least four family members are resistant to weight gain, including BXD16, BXD77, BXD87, and BXD91, gaining at most 5% over 100 days.

Our findings can be compared to strain variation and GXE effects in response to dietary restriction. Dietary restriction without malnutrition is regarded as having an almost universal benefit on longevity (Mair and Dillin, 2008; Masoro, 2009; Weindruch et al., 1986). One exception is a pair of studies on the impact of moderately intense restriction—a 40% reduction in caloric intake—across a large family of LXS strains of mice ((Liao et al., 2010; Rikke et al., 2010); *n* of up 44 strains with 10–20 replicates per strain). The most notable finding is the remarkably high variation in strain-specific change in longevity in response to DR. Life span is shortened in some LXS strains (maximum of 671-day loss), but lengthened in others (minimum of 300-day gain; GeneNetwork LXS phenotype 10164). Both the Liao and Rikke papers generated substantial controversy (Mattson, 2010) which was mirrored to some extent in matched studies of non-human primates (Mattison et al., 2017). Given such contrast in outcomes, it would be worthwhile extending the analysis of HFD to other dietary interventions in the expanded BXD family (Ashbrook et al., 2019).

Diet and Morbidity

Major morbidities and likely causes of death among different members of the BXD family appear to be influenced by diet. Those on HFD have an increased prevalence and severity of cardiovascular disease and lesions. However, the effects of a very high fat diet on cardiovascular disease incidence is quite modest in mice, unlike that observed in human cohorts (Menotti and Puddu, 2015; Sacks Frank M. et al., 2017). Interestingly, incidence of sarcomas and carcinomas are higher on the CD than HFD. Large prospective studies have failed to detect strong associations between dietary fat and cancer risk (Willett, 2000). Evaluating a causal role of diet-induced obesity in the etiology of several chronic diseases and cancers has been difficult due to correlations with numerous lifestyle factors and resulting confounding biases. Human epidemiology is increasingly using Mendelian randomization to assess the possible causal associations between risk factors and diseases (Gao et al., 2016). Well-controlled animal experiments could similarly provide additional understanding of causal associations and mechanisms underlying such complex relationships.

GXE, Weight Gain, and Associations with Longevity

Chronically high levels of fat consumption lead to substantial weight gain, associated metabolic disorders, and shortened lifespan, but causal interactions among these critical traits remains controversial. For example, studies exploiting Mendelian randomization have not shown a compelling link of triglyceride levels on longevity in humans (Liu et al., 2017). In general, a diet high in fat leads to obesity and reduced lifespan in diverse species including *Drosophila*, *C. elegans*, and mice (Otabe et al., 2007), (Yen and Curran, 2016), (Gáliková and Klepsatel, 2018). In our study, higher youthful body weight—between 100 to 200 days-of-age—is associated with reduction in longevity of 4 to 5 days per gram. This corroborates

much previous work that also demonstrates that larger body size within species is associated with shorter life span. For example, in outcrossed mice, for each gram increase in weight at 180 days, longevity is reduced by 10 to 15 days (Miller et al., 2000, their Figure 1). Among breeds of dog, for each kilogram increase in weight, longevity is reduced by ~15 days (Kraus et al., 2013). In young adult and middle-aged humans, for each kilogram increase in body weight, longevity is reduced by 80 to 146 days (Samaras et al., 2002, their figure 6). In these three species, a 5% gain in young adult body weight is associated with a 1–3% loss in longevity. In both mice and dogs the relation between early weight gain and longevity is in part a well-known function of growth hormone (GH)–insulin-like growth factor 1(IGF1) activity, although other mechanisms and gene variants are also likely to contribute. In humans, causes of this relation are probably a result of an interplay of many factors, especially nutritional balance, mean activity levels, and health care systems. For example, tall stature protects against cardiovascular disease due to lower levels of adiposity, lipid fractions, blood pressure, and better lung function (Nüesch et al., 2016). The linkage of longevity to IGF1 levels is also more tenuous than in mouse and dog (Sanders et al.,, 2018), although short stature and lower GH/IGF1 signaling is associated with resistance to most types of cancers (Choi et al., 2019; Nunney, 2018).

In humans an increase in BMI from 27 to 42 kg/m^2 increases all-cause mortality hazard ratio 1.6-fold (Wade et al., 2018). We see an even greater effect in the BXD family—the HFD diet increases weight 1.8-fold and the mortality hazard ratio 2-fold. But these average hazard ratios mask impressive modulation by genetic differences. While 45 of 57 family members gain significant weight, the other 12 gain only modest and statistically insignificant weight. For example, even after 400 days on the HFD, BXD16 is only 1.05-fold heavier than control, whereas BXD24 is 2.1-fold heavier. Remarkably, only 10% of the effect of diet on longevity is mediated through weight gain. HFD itself exerts a stronger direct effect on longevity in female BXDs than weight gain *per se*.

Future Directions

Impressive GXE differences among the BXD family emphasize the complexity of interactions between diet, weight gain, and longevity. These effects need to be teased apart and explained at genetic, epigenetic, and mechanistic levels—work that is now in progress using the same strains and cohorts (Andreux et al., 2012; Houtkooper et al., 2013; Jha et al., 2018a, 2018b; Sandoval-Sierra et al., 2019; Williams et al., 2016; Wu et al., 2014). Substantial diversity in outcomes among close-knit members the BXD family highlights the fact that observations vary between individuals and over time due to the non-ergodic nature of biology (Nadeau and Auwerx, 2019). Population or group averages hence will obscure major GXE effects. Not much can be said with certainty from a single genometype or even a small number of diverse strains. In our

case, detecting strong GXE interactions required data from 10 mice each of more than 65 strains under both conditions. Data of this type can be a foundation for specific predictions and to design a second wave of interventional studies to extend longevity and vigor (Miller et al., 2007; Strong et al., 2013). When extrapolated to humans, our results indicate that conventional dietary recommendations will be too generalized to provide effective individual guidance. We emphasize that genotype and GXE effects are far stronger modulators of longevity and weight gain than average population effects.

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Tissue Acquisition: SR, JI, CJ, MM, AC, KM, MM, WZ, JH, SM, LW, TS, CK, LL, RWW
Data Handling: SR, MBS, PJ, EGW, AS, MH, AC, RWR, SS, RWW
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References

Andreux, P.A., Williams, E.G., Koutnikova, H., Houtkooper, R.H., Champy, M.-F., Henry, H., Schoonjans, K., Williams, R.W., and Auwerx, J. (2012). Systems Genetics of Metabolism: The Use of the BXD Murine Reference Panel for Multiscalar Integration of Traits. Cell 150, 1287–1299.

Barrington, W.T., Wulfridge, P., Wells, A.E., Rojas, C.M., Howe, S.Y.F., Perry, A., Hua, K., Pellizzon, M.A., Hansen, K.D., Voy, B.H., et al. (2018). Improving Metabolic Health Through Precision Dietetics in Mice. Genetics 208, 399–417.

Belknap, J.K. (1998). Effect of within-strain sample size on QTL detection and mapping using recombinant inbred mouse strains. Behav. Genet. 28, 29–38.

Choi, Y.J., Lee, D.H., Han, K.-D., Yoon, H., Shin, C.M., Park, Y.S., and Kim, N. (2019). Adult height in relation to risk of cancer in a cohort of 22,809,722 Korean adults. Br. J. Cancer 120, 668–674.

De Haan, G., and Van Zant, G. (1999). Genetic analysis of hemopoietic cell cycling in mice suggests its involvement in organismal life span. FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol. 13, 707–713.

Finkel, T. (2015). The metabolic regulation of aging. Nat. Med. 21, 1416–1423.

Fontana, L., and Partridge, L. (2015). Promoting Health and Longevity through Diet: From Model Organisms to Humans. Cell 161, 106–118.

Flurkey K, Currer JM, Harrison DE. (2007). The mouse in biomedical research (Amsterdam; Boston: Elsevier, AP).

Gáliková, M., and Klepsatel, P. (2018). Obesity and Aging in the Drosophila Model. Int. J. Mol. Sci. 19, 1896.

Gao, C., Patel, C.J., Michailidou, K., Peters, U., Gong, J., Schildkraut, J., Schumacher, F.R., Zheng, W., Boffetta, P., Stucker, I., et al. (2016). Mendelian randomization study of adiposity-related traits and risk of breast, ovarian, prostate, lung and colorectal cancer. Int. J. Epidemiol. 45, 896–908.

Gelman, R., Watson, A., Bronson, R., and Yunis, E. (1988). Murine chromosomal regions correlated with longevity. Genetics 118, 693–704.

Hall, R.A., Liebe, R., Hochrath, K., Kazakov, A., Alberts, R., Laufs, U., Böhm, M., Fischer, H.-P., Williams, R.W., Schughart, K., et al. (2014). Systems Genetics of Liver Fibrosis: Identification of Fibrogenic and Expression Quantitative Trait Loci in the BXD Murine Reference Population. PLoS ONE 9.

Heilbronn, L.K., and Ravussin, E. (2003). Calorie restriction and aging: review of the literature and implications for studies in humans. Am. J. Clin. Nutr. 78, 361–369.

Hook, M., Roy, S., Williams, E.G., Bou Sleiman, M., Mozhui, K., Nelson, J.F., Lu, L., Auwerx, J., and Williams, R.W. (2018). Genetic cartography of longevity in humans and mice: Current landscape and horizons. Biochim. Biophys. Acta BBA - Mol. Basis Dis. 1864, 2718–2732.

Houtkooper, R.H., Argmann, C., Houten, S.M., Cantó, C., Jeninga, E.H., Andreux, P.A., Thomas, C., Doenlen, R., Schoonjans, K., and Auwerx, J. (2011). The metabolic footprint of aging in mice. Sci. Rep. 1.

Houtkooper, R.H., Mouchiroud, L., Ryu, D., Moullan, N., Katsyuba, E., Knott, G., Williams, R.W., and Auwerx, J. (2013). Mitonuclear protein imbalance as a conserved longevity mechanism. Nature 497, 451–457.

Jha, P., McDevitt, M.T., Halilbasic, E., Williams, E.G., Quiros, P.M., Gariani, K., Sleiman, M.B., Gupta, R., Ulbrich, A., Jochem, A., et al. (2018a). Genetic Regulation of Plasma Lipid Species and Their Association with Metabolic Phenotypes. Cell Syst. 6, 709-721.e6.

Jha, P., McDevitt, M.T., Gupta, R., Quiros, P.M., Williams, E.G., Gariani, K., Sleiman, M.B., Diserens, L., Jochem, A., Ulbrich, A., et al. (2018b). Systems Analyses Reveal Physiological Roles and Genetic Regulators of Liver Lipid Species. Cell Syst. 6, 722-733.e6.

Keipert, S., Voigt, A., and Klaus, S. (2011). Dietary effects on body composition, glucose metabolism, and longevity are modulated by skeletal muscle mitochondrial uncoupling in mice. Aging Cell 10, 122–136.

Kuningas, M., Mooijaart, S.P., Van Heemst, D., Zwaan, B.J., Slagboom, P.E., and Westendorp, R.G.J. (2008). Genes encoding longevity: from model organisms to humans. Aging Cell 7, 270–280.

Lang, D.H., Gerhard, G.S., Griffith, J.W., Vogler, G.P., Vandenbergh, D.J., Blizard, D.A., Stout, J.T., Lakoski, J.M., and McClearn, G.E. (2010). Quantitative trait loci (QTL) analysis of longevity in C57BL/6J by DBA/2J (BXD) recombinant inbred mice. Aging Clin. Exp. Res. 22, 8–19.

Liang, Y., Liu, C., Lu, M., Dong, Q., Wang, Z., Wang, Z., Xiong, W., Zhang, N., Zhou, J., Liu, Q., et al. (2018). Calorie restriction is the most reasonable anti-ageing intervention: a meta-analysis of survival curves. Sci. Rep. 8, 5779.

Liao, C.-Y., Rikke, B.A., Johnson, T.E., Diaz, V., and Nelson, J.F. (2010). Genetic Variation in the Murine Lifespan Response to Dietary Restriction: from Life Extension to Life Shortening. Aging Cell 9, 92–95.

Liu, Z., Burgess, S., Wang, Z., Deng, W., Chu, X., Cai, J., Zhu, Y., Shi, J., Xie, X., Wang, Y., et al. (2017). Associations of triglyceride levels with longevity and frailty: A Mendelian randomization analysis. Sci. Rep. 7, 41579.

de Magalhães, J.P., Wuttke, D., Wood, S.H., Plank, M., and Vora, C. (2012). Genome-Environment Interactions That Modulate Aging: Powerful Targets for Drug Discovery. Pharmacol. Rev. 64, 88–101.

Mattison, J.A., Colman, R.J., Beasley, T.M., Allison, D.B., Kemnitz, J.W., Roth, G.S., Ingram, D.K., Weindruch, R., de Cabo, R., and Anderson, R.M. (2017). Caloric restriction improves health and survival of rhesus monkeys. Nat. Commun. 8, 14063.

Mattson, M.P. (2010). Genes and behavior interact to determine mortality in mice when food is scarce and competition fierce. Aging Cell 9, 448–449.

McDaid, A.F., Joshi, P.K., Porcu, E., Komljenovic, A., Li, H., Sorrentino, V., Litovchenko, M., Bevers, R.P.J., Rüeger, S., Reymond, A., et al. (2017). Bayesian association scan reveals loci associated with human lifespan and linked biomarkers. Nat. Commun. 8.

Menotti, A., and Puddu, P.E. (2015). How the Seven Countries Study contributed to the definition and development of the Mediterranean diet concept: A 50-year journey. Nutr. Metab. Cardiovasc. Dis. 25, 245–252.

Miller, R.A., Chrisp, C., and Atchley, W. (2000). Differential longevity in mouse stocks selected for early life growth trajectory. J. Gerontol. A. Biol. Sci. Med. Sci. 55, B455-461.

Miller, R.A., Harper, J.M., Galecki, A., and Burke, D.T. (2002). Big mice die young: early life body weight predicts longevity in genetically heterogeneous mice. Aging Cell 1, 22–29.

Miller, R.A., Harrison, D.E., Astle, C.M., Floyd, R.A., Flurkey, K., Hensley, K.L., Javors, M.A., Leeuwenburgh, C., Nelson, J.F., Ongini, E., et al. (2007). An Aging Interventions Testing Program: study design and interim report. Aging Cell 6, 565–575.

Mitchell, S.J., Matute, J.M., Scheibye-Knudsen, M., Fang, E., Aon, M., González-Reyes, J.A., Cortassa, S., Kaushik, S., Gonzalez-Freire, M., Patel, B., et al. (2016). Effects of Sex, Strain, and Energy Intake on Hallmarks of Aging in Mice. Cell Metab. 23, 1093–1112.

Nadeau, J.H., and Auwerx, J. (2019). The virtuous cycle of human genetics and mouse models in drug discovery. Nat. Rev. Drug Discov. 18, 255–272.

Nüesch, E., Dale, C., Palmer, T.M., White, J., Keating, B.J., van Iperen, E.P., Goel, A., Padmanabhan, S., Asselbergs, F.W., Verschuren, W., et al. (2016). Adult height, coronary heart disease and stroke: a multi-locus Mendelian randomization meta-analysis. Int. J. Epidemiol. 45, 1927–1937.

Nunney, L. (2018). Size matters: height, cell number and a person's risk of cancer. Proc. R. Soc. B Biol. Sci. 285, 20181743.

Otabe, S., Yuan, X., Fukutani, T., Wada, N., Hashinaga, T., Nakayama, H., Hirota, N., Kojima, M., and Yamada, K. (2007). Overexpression of human adiponectin in transgenic mice results in suppression of fat accumulation and prevention of premature death by high-calorie diet. Am. J. Physiol. - Endocrinol. Metab. 293, E210–E218.

Peirce, J.L., Lu, L., Gu, J., Silver, L.M., and Williams, R.W. (2004). A new set of BXD recombinant inbred lines from advanced intercross populations in mice. BMC Genet. 5, 7.

Pennacchio, L.A., and Rubin, E.M. (2003). Comparative genomic tools and databases: providing insights into the human genome. J. Clin. Invest. 111, 1099–1106.

Rikke, B.A., Liao, C.-Y., McQueen, M.B., Nelson, J.F., and Johnson, T.E. (2010). Genetic dissection of dietary restriction in mice supports the metabolic efficiency model of life extension. Exp. Gerontol. 45, 691–701.

Sacks Frank M., Lichtenstein Alice H., Wu Jason H.Y., Appel Lawrence J., Creager Mark A., Kris-Etherton Penny M., Miller Michael, Rimm Eric B., Rudel Lawrence L., Robinson Jennifer G., et al. (2017). Dietary Fats and Cardiovascular Disease: A Presidential Advisory From the American Heart Association. Circulation 136, e1–e23.

Samaras, T.T., Storms, L.H., and Elrick, H. (2002). Longevity, mortality and body weight. Ageing Res. Rev. 1, 673–691.

Sandoval-Sierra, J.V., Helbing, A.H.B., Williams, E.G., Ashbrook, D.G., Roy, S., Williams, R.W., and Mozhui, K. (2019). Influence of body weight at young adulthood on the epigenetic clock and lifespan in the BXD murine family. BioRxiv 791582.

Saul, M.C., Philip, V.M., Reinholdt, L.G., and Chesler, E.J. (2019). High-Diversity Mouse Populations for Complex Traits. Trends Genet. 35, 501–514.

Skorupa, D.A., Dervisefendic, A., Zwiener, J., and Pletcher, S.D. (2008). Dietary composition specifies consumption, obesity and lifespan in Drosophila melanogaster. Aging Cell 7, 478–490.

Speakman, J.R., Mitchell, S.E., and Mazidi, M. (2016). Calories or protein? The effect of dietary restriction on lifespan in rodents is explained by calories alone. Exp. Gerontol. 86, 28–38.

Strong, R., Miller, R.A., Astle, C.M., Baur, J.A., de Cabo, R., Fernandez, E., Guo, W., Javors, M., Kirkland, J.L., Nelson, J.F., et al. (2013). Evaluation of Resveratrol, Green Tea Extract, Curcumin, Oxaloacetic Acid, and Medium-Chain Triglyceride Oil on Life Span of Genetically Heterogeneous Mice. J. Gerontol. A. Biol. Sci. Med. Sci. 68, 6–16.

Therneau, T.M., and Grambsch, P.M. (2000). Modeling Survival Data: Extending the Cox Model (New York: Springer-Verlag).

Vaughan, K.L., Kaiser, T., Peaden, R., Anson, R.M., de Cabo, R., and Mattison, J.A. (2018). Caloric Restriction Study Design Limitations in Rodent and Nonhuman Primate Studies. J. Gerontol. Ser. A 73, 48–53.

Viechtbauer, W. (2010). Conducting Meta-Analyses in R with the metafor Package. J. Stat. Softw. 36, 1–48.

Wade, K.H., Carslake, D., Sattar, N., Davey Smith, G., and Timpson, N.J. (2018). BMI and Mortality in UK Biobank: Revised Estimates Using Mendelian Randomization. Obes. Silver Spring Md 26, 1796–1806.

Willett, W.C. (2000). Diet and Cancer. The Oncologist 5, 393–404.

Williams, R.W. (2006). Animal models in biomedical research: ethics, challenges and opportunities. Princ. Mol. Med. 2nd edition, 53–60.

Williams, R.W., and Williams, E.G. (2017). Resources for Systems Genetics. In Systems Genetics, K. Schughart, and R.W. Williams, eds. (New York, NY: Springer New York), pp. 3–29.

Williams, E.G., Mouchiroud, L., Frochaux, M., Pandey, A., Andreux, P.A., Deplancke, B., and Auwerx, J. (2014). An Evolutionarily Conserved Role for the Aryl Hydrocarbon Receptor in the Regulation of Movement. PLOS Genet. 10, e1004673.

Williams, E.G., Wu, Y., Jha, P., Dubuis, S., Blattmann, P., Argmann, C.A., Houten, S.M., Amariuta, T., Wolski, W., Zamboni, N., et al. (2016). Systems proteomics of liver mitochondria function. Science 352, aad0189.

Wu, Y., Williams, E.G., Dubuis, S., Mottis, A., Jovaisaite, V., Houten, S.M., Argmann, C.A., Faridi, P., Wolski, W., Kutalik, Z., et al. (2014). Multilayered Genetic and Omics Dissection of Mitochondrial Activity in a Mouse Reference Population. Cell 158, 1415–1430.

Yen, C.A., and Curran, S.P. (2016). Gene-diet interactions and aging in C. elegans. Exp. Gerontol. 86, 106–112.

Yuan, R., Peters, L.L., and Paigen, B. (2011). Mice as a Mammalian Model for Research on the Genetics of Aging. ILAR J. Natl. Res. Counc. Inst. Lab. Anim. Resour. 52, 4–15.