- <sup>1</sup> Title: DateLife: Leveraging databases and analytical tools to reveal the dated Tree of Life
- <sup>2</sup> Authors: Luna L. Sánchez-Reyes<sup>1</sup>, Brian C. O'Meara<sup>1</sup>
- <sup>3</sup> Correspondence address:
- <sup>4</sup> 1. Department of Ecology and Evolutionary Biology, University of Tennessee, Knoxville, 425 Hesler Biology
- <sup>5</sup> Building, Knoxville, TN 37996, USA
- 6 Corresponding authors: sanchez.reyes.luna@gmail.com, bomeara@utk.edu

abstract.- The combination of new analytical techniques, availability of more fossil and molecular data, and 7 better practices in data sharing has resulted in a steady accumulation of chronograms in public and open 8 databases such as TreeBASE, Dryad, and Open Tree of Life for a large quantity and diversity of organisms 9 in the last few decades. However, getting a tree with branch lengths proportional to time remains difficult 10 for many biologists and the non-academic community, despite its importance in many areas of research, 11 education, and science communication. datelife is a service implemented via an R package and a web site 12 (http://www.datelife.org/) for efficient reuse, summary and reanalysis of published data on lineage divergence 13 times. The main workflow starts with at least two taxon names as input, either as tip labels on a tree, or 14 as a simple comma separated character string. A name search is then performed across the chronogram 15 database and positively identified source trees are pruned to maintain queried taxa only and stored as a 16 named list of patristic distance matrices. Source chronogram data can be summarised using branch length 17 summary statistics or variance minimizing approaches to generate a single summary chronogram. Source 18 chronogram data can also be used as calibration points to date a tree containing some or all names from the 19 initial query. If there is no information available for any queried taxa, data can be simulated. All source 20 and summary chronograms can be saved in formats that permit easy reuse and reanalysis. Summary and 21 newly generated trees are potentially useful to evaluate evolutionary hypothesis in different areas of research 22 in biology. How well this trees work for this purpose still needs to be tested. datelife will be useful to 23 increase awereness on the existing variation in expert time of divergence data, and might foster exploration of 24 the effect of alternative divergence time hypothesis on the results of analyses, nurturing a culture of more 25 cautious interpretation of evolutionary results. 26

Keywords: Tree; Phylogeny; Scaling; Dating; Ages; Divergence times; Open Science; Congruification;
 Supertree; Calibrations

29

### INTRODUCTION

Clade ages represent a fundamental piece of information for evolutionary understanding in many areas 30 of research, from developmental to conservation biology (Felsenstein 1985; Webb 2000), from historical 31 biogeography to species diversification studies (Posadas et al. 2006; Morlon 2014). The primary information 32 needed for these time estimates comes from the fossil record. Coupled with phylogenies with branch lengths 33 based on molecular and/or morphological data, the time of divergence of extant and extinct lineages can be 34 reconstructed with molecular dating methods. The number of studies publishing phylogenies with branch 35 lengths proportional to geological time (hereafter chronograms) have constantly increased in number for the 36 last two decades (Kumar et al. 2017). Still, generating a chronogram is not an easy task unless you have 37 specialized training: it requires inferring a tree, understanding what fossil data are available and their limits, 38 and where fossils go on the tree. That is why there has been an urge for promoting and facilitating reuse 39 of the vast amount of phylogenetic and time of lineage divergence data that has been generated and made 40 available in publications, for the advantage of research relying on this information (Webb and Donoghue 2005; 41 Stoltzfus et al. 2013). 42

Wide interest from the scientific community to make information from phylogenies in general and chronograms 43 in particular available for consultation and reuse has spurred the creation of public platforms with various goals 44 and characteristics. TreeBASE (Morell 1996; Piel et al. 2002), the Dryad repository (http://datadryad.org/). 45 and the Open Tree of Life (OToL; Hinchliff et al. 2015) are platforms that store and make available published 46 phylogenies and chronograms for easy scientific reuse. Tree in all of them can be queried using automatised 47 web procedures, which permit personalized, large scale searches that are also very fast. OToL stores trees 48 with branch length information from a wide range of living organisms, implementing a metadata structure 49 that stores the branch length units (i.e., time or relative subtitution rates). Treebase and Dryad repositories 50 also contain trees from all groups of life, but the former did not store branch length information until recently 51 (and lacks consistent metadata on what any branch lengths stored mean) and Dryad stores many other types 52 of biological data using metadata that does not allow automatic distinction of types of trees and branch 53 length units, impairing the automatised access to time of lineage divergence information. 54

Besides keeping a repository to easily store and share expert phylogenetic and chronogram knowledge, OToL 55 also has the primary goal of synthesising all trees in their repository to expose to the community a single tree 56 of all life depicting the phylogenetic relationships among known lineages. All or parts of this synthetic tree 57 can be reused for any purpose. However, it currently only focus on synthesizing tree topology, meaning that 58 it does not expose branch length data of any type. The Timetree of Life project focuses on the synthesis of a 59 single chronogram of life (Hedges et al. 2006) and presents a very accessible, attractive interface. However, 60 the thousands of chronograms this NSF-funded project have compiled for synthesis are only publicly available 61 for visual examination in their website or for download as images, but large scale download remains prohibited 62 by their site. The latest version of their synthetic chronogram (Kumar et al. 2017) can be queried only 63 through their website in a non-automatised fashion, and only subsets of it can be reused for analyses with the 64 permission of the authors. Other platforms such as SuperSmart (Antonelli et al. 2017) and phylogenerator 65 (Pearse and Purvis 2013) are focused in automatised *de novo* chronogram inference, by reusing DNA sequence 66 data to reconstruct phylogenetic trees. However, expert fossil information necessary for subsequent molecular 67 dating analyses still needs to be compiled and curated by the user, rendering them a challenging tool to obtain 68 data on time of lineage divergence for the non-specialist. Moreover, these tools do not provide information 69 from already created expert chronograms. 70

A tool for efficient reuse of expert, published data on time of lineage divergence should have an open and 71 fully public chronogram database storing data in a format suitable for scientific reuse, an automatised way of 72 accessing the information, and straightforward means of comparing and summarizing chronogram information 73 as needed by the user. A prototype service aiming to meet this characteristics was developed over a series 74 of hackathons at the National Evolutionary Synthesis Center (Stoltzfus et al. 2013). In here we present 75 the formal description and implementation of the datelife service, constituted by an R package and a 76 web site (http://www.datelife.org/). There is still much room for improvement, and flaws and limitations 77 are addressed below. We strived for the current implementation of datelife to perform the basic tasks 78 described above, featuring a system for maintenance of an open database of chronograms pulled from public 79 repositories, methods to summarize and compare source chronograms, and new functions to visualize and 80 graphically compare source and summary chronograms. 81

### DESCRIPTION

<sup>83</sup> The basic datelife workflow is shown in figure 1 and consists of:

82

A user providing at least two taxon names as input, either as tip labels on a tree, or as a simple comma
 separated character string. The tree can be in newick or phylo format, and can be with or without
 branch lengths.

A name search is then performed across the chronogram database; source trees with at least two
matching input names are identified; all other taxa that do not match the original query are then
dropped from the positively identified source trees. These pruned chronograms are hereafter referred as
source chronograms. Finally, each source chronogram is transformed to a patristic matrix named by
the citation of the original study. This format facilitates and greatly speeds up all further analyses and
summarizing algorithms.

3. The user can obtain different summary information including: a) all source chronograms, b) maximum ages of source chronograms, c) citations of studies where source chronograms were originally published, d)
a summary table with all of the above, e) a single summary tree of all or a subset of source chronograms,
f) a report of succesful matches of input taxon names across source chronograms, and g) the single source chronogram with the greatest number of taxa. Summary information can be used to make decisions on the next steps of the workflow.

- 4. Then, source chronogram data can be used as calibration points to date a tree with or without branch
   lengths containing some or all names from the initial query.
- 5. If there is no information available for any queried taxa, users can also create both age and phylogenetic
   data for this missing taxa with a variety of algorithms described below.
- 6. Finally, users can easily save all source and summary chronograms in formats that permit easy reuse
   and reanalyses (newick and R "phylo" format), as well as view and compare results graphically, or
   construct their own graphs using inbuilt datelife graphic generation functions.

5

To gather, process, and present information, datelife builds up from functions available in several R
packages including rotl (Michonneau et al. 2016), ape (Paradis et al. 2004), geiger (Harmon et al. 2008),
paleotree (Bapst 2012), bold (Chamberlain 2018), phytools (Revell 2012), taxize (Chamberlain and Szöcs
2013; Chamberlain 2018), phyloch (Heibl 2008), phylocomr (Ooms and Chamberlain 2018) and rphylotastic
(O'Meara et al. 2019).

A datelife search currently accepts scientific names only. It can be any named clade or binomial specific. 111 The search is performed at the species level, so when input names correspond to named clades, datelife 112 pulls all accepted species names within the clade from OToL's reference taxonomy and uses all of those. 113 Searches at the infraspecies level are not currently allowed, so input names belonging to subspecies or any 114 other infraspecific category are collapsed to the species level. datelife processes input names with the taxon 115 name resolution service (TNRS; Boyle et al. 2013), which corrects potentially misspelled names and typos, 116 and standardizes spelling variations and synonyms, increasing the probability of correctly finding the queried 117 taxa in datelife's chronogram database. 118

Then, a name matching procedure is performed across datelife's chronogram database which is assembled from OToL's tree repository. Compared to other existing open tree repositories OToL's metadata rich tree store is the only one that supports search, identification, and handling of chronograms in an automatised fashion. Also, the tip names in their stored trees correspond to scientific names at or below the species level. Finally, all their chronograms come from peer-reviewed published studies generated by specialists in the targeted lineages, arguably representing expert knowledge on time of lineage divergence.

<sup>125</sup> Information from source chronograms can be summarised with a summary statistic of tree branch lengths, <sup>126</sup> such as median or mean. A much slower, but possibly more accurate Super Distance Matrix (SDM) approach <sup>127</sup> for supertree reconstruction with branch lengths (Criscuolo et al. 2006) is also implemented via the ape <sup>128</sup> package (Paradis et al. 2004). The resulting summary patristic distance matrix could be clustered with <sup>129</sup> classic algorithms to return a tree. However, the resulting trees are often non-ultrameric and do not reflect <sup>130</sup> the source chronogram data (see datelife\_examples package). Instead, we obtained a distribution of age data <sup>131</sup> from the summary matrix available for nodes on a consensus tree. The Branch Length Adjuster (BLADJ)

<sup>132</sup> algorithm (Webb et al. 2008) was then used to fix known node ages and then distribute nodes with unknown
<sup>133</sup> ages evenly over the consensus tree, minimizing age variance in the resulting chronogram.

For tree dating, the congruification algorithm described by Eastman et al. (2013) is implemented to find shared nodes between trees (congruent nodes). The ages of these nodes are then used as calibrations to date any given tree. Currently implemented methods for tree dating are BLADJ, MrBayes (Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003) and PATHd8 (Britton et al. 2007), a non-clock, rate-smoothing dating method.

139

## Benchmark

datelife's code speed was tested on an Apple iMac with one 3.4 GHz Intel Core i5 processor. We registered 140 variation in computing time of query processing and search through the database relative to number of 141 queried taxon names. Query processing increases roughly linearly with number of input taxon names, and 142 increases considerably if TNRS service is activated. Up to ten thousand names can be processed and searched 143 in less than 30 minutes. A name search through the chronogram database with an already processed query 144 can be performed in less than a minute, even with a very large number of taxon names (Fig. 2). datelife's 145 code performance was evaluated with a set of unit tests designed and implemented with the R package 146 testthat (R Core Team 2018) that were run locally –using the devtools package (R Core Team 2018), and 147 on a public server -via GitHub, using the continuous integration tool Travis CI (https://travis-ci.org). At 148 present, unit tests cover more than 50% of datelife's code (https://codecov.io/gh/phylotastic/datelife). 149

150

### EXAMPLE

In this section we demonstrate the types of outputs that can be obtained with datelife, using as an example the bird family Fringillidae of true finches. We performed a higher-taxon search to obtain all data on lineage divergence available in datelife's database for all recognised species within the Fringillidae (475 spp. according to the Open Tree of Life taxonomy). There are 13 chronograms containing at least two Fringillidae species, published in 9 different studies (Fig. 3). Data from these source chronograms was used to generate two types of summary chronograms, median and SDM. As explained in the "Description" section, data from

<sup>157</sup> source chronograms was first summarised into a single distance matrix (using the median and the SDM <sup>158</sup> method respectively) and then the available node ages were used as fixed ages over a consensus tree topology, <sup>159</sup> to obtain a fully dated tree with the program BLADJ (Fig. 4). Median summary chronograms are older <sup>160</sup> and have wider variation in maximum ages than chronograms obtained with SDM. In both cases, ages are <sup>161</sup> generally consistent with source ages, but see next section.

Data from source chronograms was also used to date tree topologies with no branch length information and 162 trees with branch lengths in relative substitution rates (Figs. 5 and 6). As a form of cross validation, we used 163 tree topologies from each study and calibrated them using information from all other source chronograms. In 164 the absence of branch length data, the ages of internal nodes were approximately recovered in almost all 165 cases (except for studies 3, and 5; Fig. 5). Maximum tree ages were only approximately recovered in one case 166 (study 2; Fig. 5). To exemplify dating a tree with branch lengths provided by the user, we obtained DNA 167 sequence data from the Barcode of Life Database (BOLD) and branch lengths were successfully generated for 168 all source chronograms. However, dating with PATHd8 (using congruified calibrations) was only successful in 169 three cases (studies 3, 5, and 9, shown in Fig. 6). From these, two trees have a different sampling than the 170 original source chronogram, mainly because DNA data for some species is absent from the BOLD. Maximum 171 ages are quite different from source chronograms, but this might be explained also by the differences in 172 sampling between source chronograms and BOLD trees. More examples and code used to generate these 173 trees can be consulted in https://github.com/LunaSare/datelife examples. 174

175

### FLAWS, LIMITATIONS AND PROSPECTS

The main goal of datelife is to make expert information on time of lineage divergence easily accesible for comparison, reuse, and reanalysis, to researchers in all areas of science and with all levels of expertise in the matter. It is a very fast tool that fulfills the quality of openness and does not require any expert biological knowledge from users –besides the names of the organisms they want to work with– for any of its functionalities. However, it has many flaws. Some of them can be overcome, some of them might represent limitations.

At the moment, datelife's chronogram database is not very large, storing 231 chronograms up to the time 182 the manuscript was written. This represents 5.79% of the largest existing chronogram database, which is 183 not open for scientific reuse nor automatised data mining (Kumar et al. 2017). OToL is the only public 184 tree repository from where datelife can currently pull chronograms to construct its database. A previous 185 version of TimeTree's synthetic chronogram (Hedges et al. 2015) was made available in the OToL repository, 186 hence the amount of lineages represented in datelife's database is at least as substantial as TimeTree's. This 187 ensures that some information will be available for any given query, but it does not ensure that the full 188 state of knowledge of time of divergence data will be available for any given lineage. Thus, incorporation of 189 more published chronograms deposited in OpenTree, or perhaps directly pulled from the Dryad repository, 190 to datelife's database is crucial to improve its services. Methods to automatically mine chronogram data 191 from Dryad could be designed and implemented. However, the unit of branch lengths would still need to be 192 determined by hand. Consequently, we would like to emphasize on the importance of sharing chronogram 193 data for the scientific community, in repositories that require expert input and manual curation, such as 194 OToL's tree repository. 195

Another potential concern comes from summary chronograms. We currently summarize by default all source 196 chronograms that overlap with at least two taxa. Users can subset source data if they have reasons to favor 197 some source chronograms over others. Strictly speaking, a good chronogram should reflect the real time 198 of lineage divergence accurately and precisely. To our knowledge, there is no objective way to determine 199 if an expert chronogram is better than other. Some criteria that have been put forward are the level of 200 lineage sampling and the number of calibrations used. Scientists usually also favor chronograms coming from 201 studies with primary calibrations to ones from secondary calibrations. It has been observed with simulations 202 that divergence times inferred with secondary calibrations are significantly younger than those inferred with 203 primary calibrations in analyses performed with bayesian inference methods when priors are implemented in 204 similar ways in both analyses (Schenk 2016). Yet, there are different ways to use secondary calibrations and 205 the bias might not be encountered with other dating methods that do not require setting priors (such as ML 206 methods). In sum, further studies are required to fully understand the effect of using secondary calibrations 207 on time estimates and downstream anlyses. 208

Furthermore, even chronograms obtained with primary fossil data can show substantial variation in time 209 estimates between clades, as observed from the comparison of source chronograms in the Fringillidae example. 210 This observation is often encountered in the literature (see, for example, the ongoing debate about crown group 211 age of angiosperms (Ramshaw et al. 1972; Sanderson and Doyle 2001; Magallón et al. 2015; Barba-Montoya 212 et al. 2018). For some studies, especially ones based on branch lengths (e.g., studies of species diversification, 213 timing of evolutionary events, phenotypic trait evolution), using a different chronogram may return different 214 results (Title and Rabosky 2016). Stitching together these chronograms can create a larger tree that uses 215 information from multiple studies, but the effect of uncertainties and errors here on downstream analyses is 216 still largely unknown. 217

Summarizing high-quality chronograms might also imply summarizing evolutionary hypothesis. This could be 218 enlightening from certain point of view, since it could help getting a single global evolutionary history for a 219 lineage. It could also be really misleading, since we are potentially loosing important parts of the evolutionary 220 history of a lineage that might only be reflected in some chronograms and not from the summary chronogram. 221 Ideally, we should still rely on time of lineage divergence data obtained from a single analysis using fossil 222 data as primary sources of calibrations, and using fossils that have already been curated as calibrations to 223 date other trees, which should reflect a more homogeneous evolutionary history (Antonelli et al. 2017). This 224 will be implemented in future datelife versions. 225

Alternatively, one could try to choose the "best" chronogram. Several characteristics of the data used for dating 226 analyses as well as from the output chronogram itself, could be used to score quality of source chronograms. 227 Some characteristics that are often cited as a measure of improved quality in dating studies addressing 228 long-studied organisms are: quality of alignment (missing data, GC content), lineage sampling (strategy 229 and proportion), phylogenetic and dating inference method, number of fossils used as calibrations, support 230 for nodes and ages, and magnitude of confidence intervals. To facilitate subsetting of source chronograms 231 following different criteria by the users, this information should be included as metadata manually entered by 232 curators in the near future. 233

<sup>234</sup> In other areas of biological research, such as ecology and conservation biology, it has been shown that at least

some data on lineage divergence represents a relevant improvement for testing alternative hypothesis using 235 phylogenetic distance. Hence, we allow accepted ways of creating branch lengths in the absence of starting 236 branch length information (such as BLADJ (Webb et al. 2008)) for several taxa lacking this information. 237 Making up branch lengths in this or other ways is accepted in scientific publications: Rabosky et al. (2018) 238 created a time-calibrated tree of 31,536 ray-finned fishes, of which only 37% had molecular data; Jetz et 230 al. (2012), created a time-calibrated tree of all 9,993 bird species, where 67% had molecular data; Smith 240 and Brown (2018) constructed a tree of 353,185 seed plants where only 23% had molecular data. Taken to 241 the extreme, one could make a fully resolved, calibrated tree of all modern and extinct taxa using a single 242 taxonomy and a single calibration with the polytomy resolution and branch imputation methods. There has 243 yet to be a thorough analysis of what can go wrong when one goes beyond the data in this way, so we urge 244 caution; we also urge readers to follow the example of many of the large tree papers cited above and make 245 sure results are substantially similar between trees fully reconstructed with molecular or other data and trees 246 that are reconstructed using taxonomy. 247

248

### CONCLUSIONS

Divergence time information is key to many areas of evolutionary studies: trait evolution, diversification, biogeography, macroecology and more. Generating this information is difficult, especially for those who want to use phylogenies but who are not systematists, or do not have the time to acquire and develop the necessary knowledge and data curation skills to produce chronograms *de novo*. Knowledge on clade ages is also crucial for science communication and education.

datelife allows an easy and fast obtention, as well as comparison of publicly available information on time of lineage divergence, providing a straightforward way to get an informed idea on the state of knowledge of the time frame of evolution of different regions of the tree of life, allowing identification of regions that require more research or that have conflicting information. Both summary and newly generated trees are potentially useful to evaluate evolutionary hypothesis in different areas of research. datelife helps with awereness on the existing variation in expert time of divergence data, and might foster exploration of the effect of alternative divergence time hypothesis on the results of analyses, nurturing a culture of more cautious

<sup>261</sup> interpretation of evolutionary results.

262

## AVAILABILITY

datelife is free and open source and it can be used through its current website http://www.datelife.org/ 263 query/, through its R package, and through Phylotastic's project web portal http://phylo.cs.nmsu.edu:3000/. 264 datelife's website is maintained using RStudio's shiny server and the shiny package open infras-265 datelife's R package stable version will be available for installatructure, as well as Docker. 266 tion from the CRAN repository (https://cran.r-project.org/package=datelife) using the command 267 install.packages(pkgs = "datelife") from within R. Development versions are available from the 268 GitHub repository (https://github.com/phylotastic/datelife) and can be installed using the command 269 devtools::install\_github("phylotastic/datelife"). 270

271

### SUPPLEMENTARY MATERIAL

<sup>272</sup> Code used to generate all versions of this manuscript, the biological examples, as well as the benchmark of
<sup>273</sup> functionalities are respectively in the datelife\_paper1, datelife\_examples, and datelife\_benchmark repositories
<sup>274</sup> in LLSR GitHub account.

275

## Funding

Funding was provided by the US National Science Foundation (NSF) grants ABI-1458603 to Datelife project and DBI-0905606 to the National Evolutionary Synthesis Center (NESCent), and the Phylotastic project Grant ABI-1458572.

279

### Acknowledgements

We thank colleagues from the O'Meara Lab at the University of Tennesse Knoxville for suggestions, discussions and software testing. The late National Evolutionary Synthesis Center (NESCent), which sponsored hackathons that led to initial work on this project. The team that assembled datelife's first proof of concept: Tracy Heath, Jonathan Eastman, Peter Midford, Joseph Brown, Matt Pennell, Mike Alfaro, and

- 284 Luke Harmon. The Open Tree of Life project that provides the open, metadata rich repository of trees
- used for datelife. The many scientists who publish their chronograms in an open, reusable form, and the
- <sup>286</sup> scientists who curate them for deposition in the Open Tree of Life repository. The NSF for funding nearly all
- <sup>287</sup> the above, in addition to the ABI grant that funded this project itself.

#### References

288

Antonelli A., Hettling H., Condamine F.L., Vos K., Nilsson R.H., Sanderson M.J., Sauquet H., Scharn
R., Silvestro D., Töpel M., Bacon C.D., Oxelman B., Vos R.A. 2017. Toward a self-updating platform for
estimating rates of speciation and migration, ages, and relationships of Taxa. Systematic Biology. 66:153–166.

Bapst D.W. 2012. Paleotree: An R package for paleontological and phylogenetic analyses of evolution.
 Methods in Ecology and Evolution. 3:803–807.

Barba-Montoya J., Reis M. dos, Schneider H., Donoghue P.C., Yang Z. 2018. Constraining uncertainty in the
timescale of angiosperm evolution and the veracity of a cretaceous terrestrial revolution. New Phytologist.
218:819–834.

Barker F.K., Burns K.J., Klicka J., Lanyon S.M., Lovette I.J. 2012. Going to extremes: Contrasting rates of
 diversification in a recent radiation of new world passerine birds. Systematic biology. 62:298–320.

Barker F.K., Burns K.J., Klicka J., Lanyon S.M., Lovette I.J. 2015. New insights into new world biogeography:
An integrated view from the phylogeny of blackbirds, cardinals, sparrows, tanagers, warblers, and allies. The
Auk: Ornithological Advances. 132:333–348.

- Boyle B., Hopkins N., Lu Z., Raygoza Garay J.A., Mozzherin D., Rees T., Matasci N., Narro M.L., Piel
  W.H., Mckay S.J., Lowry S., Freeland C., Peet R.K., Enquist B.J. 2013. The taxonomic name resolution
  service: An online tool for automated standardization of plant names. BMC Bioinformatics. 14.
- Britton T., Anderson C.L., Jacquet D., Lundqvist S., Bremer K. 2007. Estimating Divergence Times in Large
  Phylogenetic Trees. Systematic Biology. 56:741–752.
- <sup>307</sup> Burns K.J., Shultz A.J., Title P.O., Mason N.A., Barker F.K., Klicka J., Lanyon S.M., Lovette I.J. 2014.
   <sup>308</sup> Phylogenetics and diversification of tanagers (passeriformes: Thraupidae), the largest radiation of neotropical
   <sup>309</sup> songbirds. Molecular Phylogenetics and Evolution. 75:41–77.
- <sup>310</sup> Chamberlain S. 2018. bold: Interface to Bold Systems API..

- <sup>311</sup> Chamberlain S.A., Szöcs E. 2013. taxize : taxonomic search and retrieval in R [version 2; referees: 3 approved].
- $_{312}$  F1000Research. 2:1–29.
- Claramunt S., Cracraft J. 2015. A new time tree reveals earth history's imprint on the evolution of modern
- <sup>314</sup> birds. Science advances. 1:e1501005.
- <sup>315</sup> Criscuolo A., Berry V., Douzery E.J., Gascuel O. 2006. SDM: A fast distance-based approach for (super)tree
- <sup>316</sup> building in phylogenomics. Systematic Biology. 55:740–755.
- Eastman J.M., Harmon L.J., Tank D.C. 2013. Congruification: Support for time scaling large phylogenetic trees. Methods in Ecology and Evolution. 4:688–691.
- <sup>319</sup> Felsenstein J. 1985. Phylogenies and the Comparative Method. The American Naturalist. 125:1–15.
- 320 Gibb G.C., England R., Hartig G., McLenachan P.A., Taylor Smith B.L., McComish B.J., Cooper A., Penny
- D. 2015. New zealand passerines help clarify the diversification of major songbird lineages during the oligocene. Genome biology and evolution. 7:2983–2995.
- Harmon L., Weir J., Brock C., Glor R., Challenger W. 2008. GEIGER: investigating evolutionary radiations.
  Bioinformatics. 24:129–131.
- Hedges S.B., Dudley J., Kumar S. 2006. TimeTree: A public knowledge-base of divergence times among
   organisms. Bioinformatics. 22:2971–2972.
- Hedges S.B., Marin J., Suleski M., Paymer M., Kumar S. 2015. Tree of life reveals clock-like speciation and
   diversification. Molecular Biology and Evolution. 32:835–845.
- Heibl C. 2008. PHYLOCH: R language tree plotting tools and interfaces to diverse phylogenetic software
   packages..
- <sup>331</sup> Hinchliff C.E., Smith S.A., Allman J.F., Burleigh J.G., Chaudhary R., Coghill L.M., Crandall K.A., Deng J.,
- 332 Drew B.T., Gazis R., Gude K., Hibbett D.S., Katz L.A., Laughinghouse H.D., McTavish E.J., Midford P.E.,

- <sup>333</sup> Owen C.L., Ree R.H., Rees J.A., Soltis D.E., Williams T., Cranston K.A. 2015. Synthesis of phylogeny and
- taxonomy into a comprehensive tree of life. Proceedings of the National Academy of Sciences. 112:12764–12769.
- Hooper D.M., Price T.D. 2017. Chromosomal inversion differences correlate with range overlap in passerine
  birds. Nature ecology & evolution. 1:1526.
- Huelsenbeck J.P., Ronquist F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics.
   17:754–755.
- Jetz W., Thomas G., Joy J.J., Hartmann K., Mooers A. 2012. The global diversity of birds in space and
  time. Nature. 491:444–448.
- Kumar S., Stecher G., Suleski M., Hedges S.B. 2017. TimeTree: A Resource for Timelines, Timetrees, and
   Divergence Times. Molecular biology and evolution. 34:1812–1819.
- Magallón S., Gómez-Acevedo S., Sánchez-Reyes L.L., Hernández-Hernández T. 2015. A metacalibrated
   time-tree documents the early rise of flowering plant phylogenetic diversity. New Phytologist. 207:437–453.
- Michonneau F., Brown J.W., Winter D.J. 2016. rotl: an R package to interact with the Open Tree of Life data. Methods in Ecology and Evolution. 7:1476–1481.
- <sup>347</sup> Morell V. 1996. The roots of phylogeny. Science. 273:569.
- Morlon H. 2014. Phylogenetic approaches for studying diversification. Ecology Letters. 17:508–525.
- O'Meara B., Md Tayeen A.S., Sanchez Reyes L.L. 2019. Rphylotastic: An r interface to 'phylotastic' web
   services..
- <sup>351</sup> Ooms J., Chamberlain S. 2018. Phylocomr: Interface to 'phylocom'..
- Paradis E., Claude J., Strimmer K. 2004. APE: analyses of phylogenetics and evolution in R language.
   Bioinformatics. 20:289–290.

- Pearse W.D., Purvis A. 2013. PhyloGenerator: An automated phylogeny generation tool for ecologists.
  Methods in Ecology and Evolution. 4:692–698.
- <sup>356</sup> Piel W.H., Donoghue M., Sanderson M. 2002. TreeBASE : A database of phylogenetic information. In:
- <sup>357</sup> Shimura J., Wilson K., Gordon D., editors. To the interoperable "catalog of life" with partners. Tsukuba,
- <sup>358</sup> Japan: National Institute for Environmental Studies. p. 41–47.
- <sup>359</sup> Posadas P., Crisci J.V., Katinas L. 2006. Historical biogeography: A review of its basic concepts and critical
   <sup>360</sup> issues. Journal of Arid Environments. 66:389–403.
- Price T.D., Hooper D.M., Buchanan C.D., Johansson U.S., Tietze D.T., Alström P., Olsson U., Ghosh-Harihar
  M., Ishtiaq F., Gupta S.K., others. 2014. Niche filling slows the diversification of himalayan songbirds.
  Nature. 509:222.
- Rabosky D.L., Chang J., Title P.O., Cowman P.F., Sallan L., Friedman M., Kaschner K., Garilao C., Near
  T.J., Coll M., others. 2018. An inverse latitudinal gradient in speciation rate for marine fishes. Nature.
  559:392.
- Ramshaw J., Richardson D., Meatyard B., Brown R., Richardson M., Thompson E., Boulter D. 1972. The
  time of origin of the flowering plants determined by using amino acid sequence data of cytochrome c. New
  Phytologist. 71:773–779.
- R Core Team. 2018. R: a language and environment for statistical computing. Vienna, Austria: R Foundation
  for Statistical Computing.
- Revell L.J. 2012. Phytools: An r package for phylogenetic comparative biology (and other things). Methods
  in Ecology and Evolution. 3:217–223.
- <sup>374</sup> Ronquist F., Huelsenbeck J.P. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models.
  <sup>375</sup> Bioinformatics. 19:1572–1574.
- <sup>376</sup> Sanderson M.J., Doyle J.A. 2001. Sources of error and confidence intervals in estimating the age of angiosperms

- <sup>377</sup> from rbcL and 18S rDNA data. American Journal of Botany. 88:1499–1516.
- 378 Schenk J.J. 2016. Consequences of secondary calibrations on divergence time estimates. PLoS ONE. 11.
- Smith S.A., Brown J.W. 2018. Constructing a broadly inclusive seed plant phylogeny. American Journal of
   Botany. 105:302–314.
- <sup>381</sup> Stoltzfus A., Lapp H., Matasci N., Deus H., Sidlauskas B., Zmasek C.M., Vaidya G., Pontelli E., Cranston
- <sup>382</sup> K., Vos R., Webb C.O., Harmon L.J., Pirrung M., O'Meara B., Pennell M.W., Mirarab S., Rosenberg M.S.,
- Balhoff J.P., Bik H.M., Heath T.A., Midford P.E., Brown J.W., McTavish E.J., Sukumaran J., Westneat M.,
- Alfaro M.E., Steele A., Jordan G. 2013. Phylotastic! Making tree-of-life knowledge accessible, reusable and
- 385 convenient. BMC Bioinformatics. 14.
- Title P.O., Rabosky D.L. 2016. Do Macrophylogenies Yield Stable Macroevolutionary Inferences? An
   Example from Squamate Reptiles. Systematic Biology.:syw102.
- Webb C.O. 2000. Exploring the Phylogenetic Structure of Ecological Communities : An Example for Rain
   Forest Trees. The American Naturalist. 156:145–155.
- Webb C.O., Ackerly D.D., Kembel S.W. 2008. Phylocom: Software for the analysis of phylogenetic community structure and trait evolution. Bioinformatics. 24:2098–2100.
- Webb C.O., Donoghue M.J. 2005. Phylomatic: Tree assembly for applied phylogenetics. Molecular Ecology
   Notes. 5:181–183.

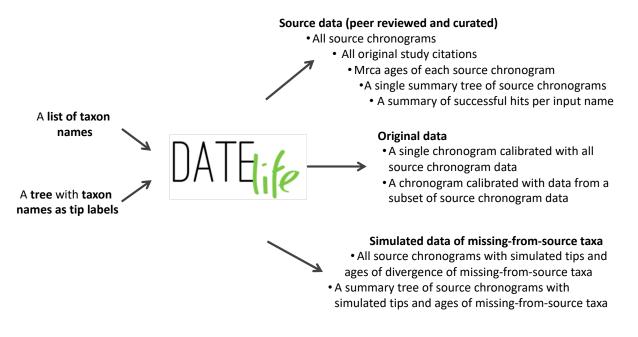


FIGURE 1

<sup>394</sup> Stylized DateLife workflow. This shows the general worflows and analyses that can be performed with

<sup>395</sup> datelife, via the R package or through the website at http://www.datelife.org/. Details on the functions

<sup>396</sup> involved on each workflow are shown in datelife's R package vignette.

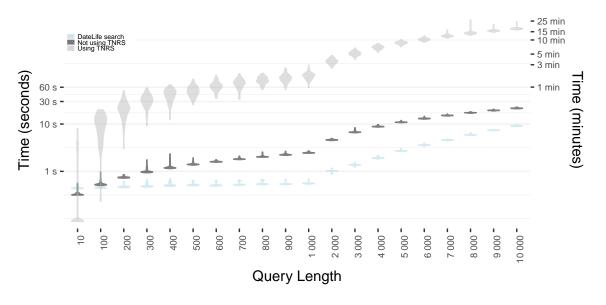
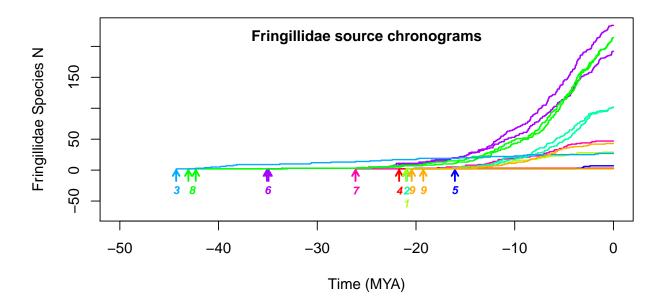


FIGURE 2

Computation time of query processing and search across datelife's chronogram database relative to number of input taxon names. We sampled N names from the class Aves for each cohort 100 times and then performed a search with query processing not using the Taxon Names Resoultion Service (TNRS; dark gray), and using TNRS (light gray). We also performed a search using the already processed query for comparison (light blue).



## FIGURE 3

Lineage through time (LTT) plots of source chronograms containing all or a subset of species from the bird family Fringillidae of true finches. Arrows indicate maximum age of each chronogram. Numbers reference to chronograms' original publications 1: Barker et al. (2012), 2: Barker et al. (2015), 3: Burns et al. (2014), 4: Claramunt and Cracraft (2015), 5: Gibb et al. (2015), 6: Hedges et al. (2015), 7: Hooper and Price (2017), 8: Jetz et al. (2012), 9: Price et al. (2014).

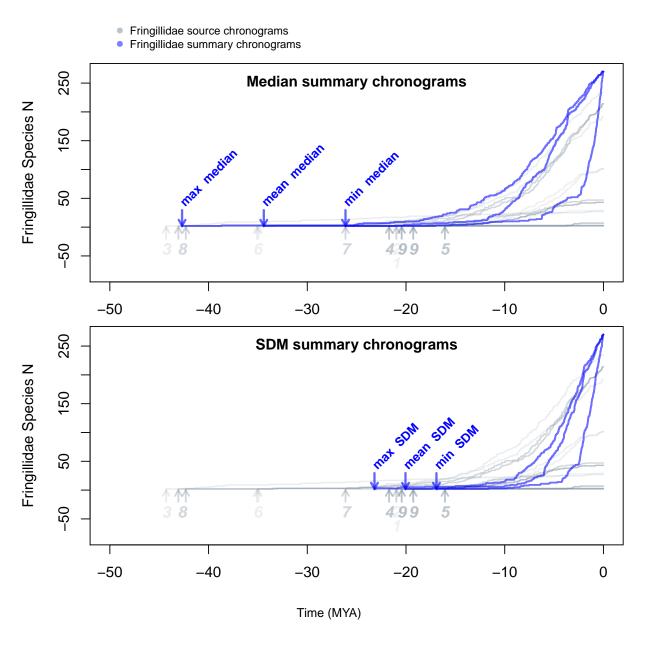
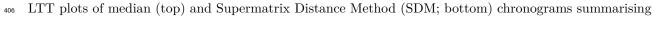


FIGURE 4



407 information from source chronograms found for the Fringillidae. Arrows indicate tree maximum age.

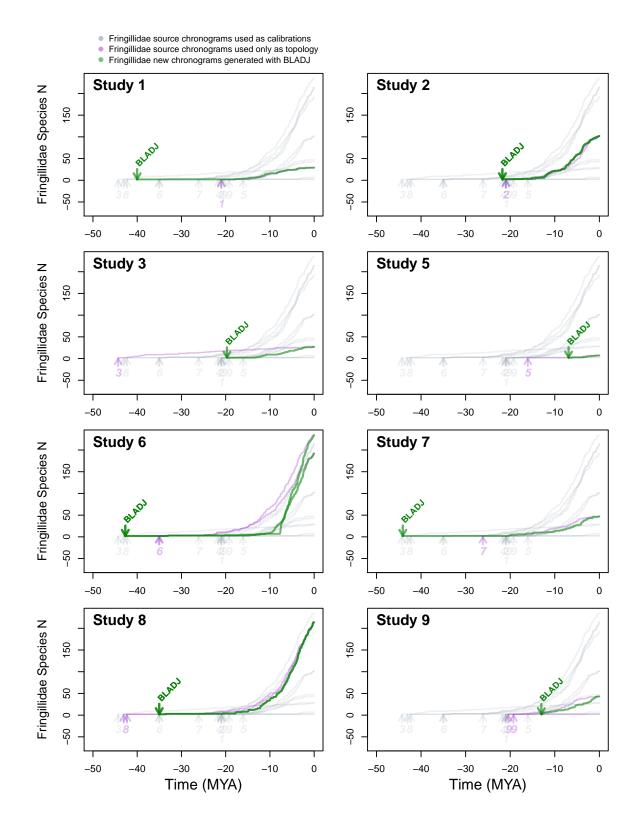


FIGURE 5

- <sup>408</sup> LTT plots showing results from the cross-validation analyses of trees without branch lengths dated using
- <sup>409</sup> BLADJ. The dating analysis can only be performed in trees with more than 2 tips, thus excluding chronogram
- 410 from study 4; its data was still used as calibration for the other source chronograms.

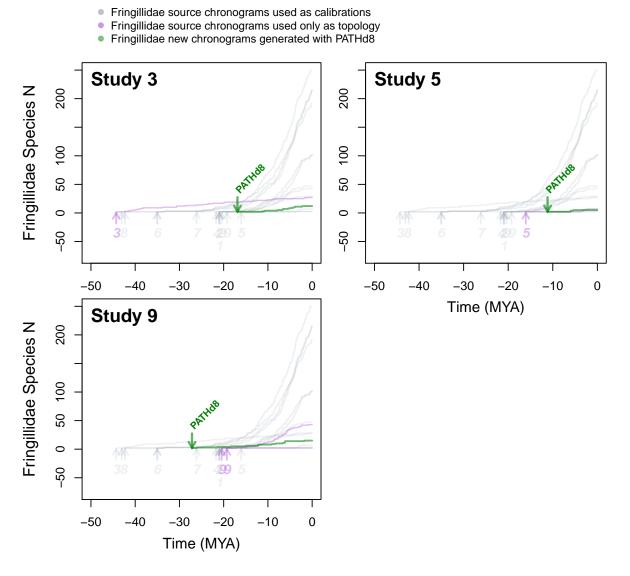


FIGURE 6

LTT plots showing results from the cross-validation analyses of trees with branch length reconstructed with data from the Barcode of Life Database (BOLD) dated using PATHd8. We could construct a tree with branch lengths for all source chronograms. However, dating with PATHd8 was only successful in three source chronograms shown here.