Running title: Adaptation of *Aquilegia* species

# Genetic and epigenetic characteristics associated with the rapid radiation of *Aquilegia* species

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**Abstract**

Elucidating the genetic and epigenetic bases underlying species diversification is crucial to understanding the evolution and persistence of biodiversity. As a well-known horticultural plant grown worldwide, the genus Aquilegia (columbine) is also a model system in adaptive radiation research. In this study, we surveyed the genomes and DNA methylomes of ten representative Aquilegia species from the Asian, European and North American lineages. Our inferences of the phylogenies and population structure revealed clearly high genetic and DNA methylomic divergence across the three lineages. By multi-levelled genome-wide scanning, we identified candidate genes exhibiting lineage-specific genetic or epigenetic variation patterns that are signatures of inter-specific divergence. We demonstrated that these species diversification-associated genetic variations and epigenetic variabilities were partially independent but were both functionally related to various biological processes vital to adaptation, including stress tolerance, cell reproduction and DNA repair. Our study provides an exploratory overview of how the established genetic and epigenetic signatures are associated with the rapid radiation of Aquilegia species.

**Key words:** Adaptive radiation; Aquilegia; Selection; Whole-genome sequencing; DNA methylation

## Introduction

Adaptive radiation is the rapid diversification of a single ancestral species into a vast array of common descendants that inhabit different ecological niches or use a variety of resources, but differ in phenotypic traits required to exploit diverse environments[1–4]. Disentangling the evolutionary mechanisms underpinning adaptive radiation is fundamental to understanding the evolution and persistence of biodiversity[5,6]. This has been a key focus of many studies which were investigating different animal and plant lineages that diversified through adaptive radiation, including Hawaiian silversword, Caribbean anoles, Darwin's finches, and African cichlids[7–10]. However, it remains under-investigated as to why some lineages could diversify rapidly but their close relatives or other sympatrically distributed lineages did not. In the past decades, accumulating evidence from diverse radiation lineages suggest that both the extrinsic environmental factors (e.g., resource availability) and genetic variations can determine the rate and volume of species diversification[11]. Among the environmental triggers, ecological opportunity is considered as the primary mechanism that causes rapid adaptive radiation through acquisition of key innovations, penetration of new environments and extinction of competitors[2,12]. On the other hand, new species also arise as a result of new genetic variations being preserved which could ultimately influence the phenotypic disparity, where natural selection act on, among closely related species[13]. In the rapid speciation of the African cichlid fishes, extrinsic environmental factors (e.g., ecological specialization) and genetic mechanisms (e.g., adaptive introgression) acted together to provoke the repeated adaptive radiation in geographically isolated lakes[7,11,14,15].

The genus *Aquilegia* L. (columbine) is a well-recognized model system to study the evolutionary mechanisms underlying adaptive radiation[16,17]. This genus includes approximately 70 recently diversified species that are widely distributed in the temperate zones of North America and Eurasia[18]. Phylogenetic and geographic inferences have illustrated two independent adaptive radiations of North American and European lineages from the ancestral Asian species[17,19]. For example, floral diversification of the North American *Aquilegia* species is highly correlated with the pollinator specialization[20–23]. In contrast, ecological adaptation and geographic isolation are considered as the major driving forces promoted rapid radiation of the European species[17,24]. In Asia, changes in pollinator and ecological habitats are both proposed to be the underpinning mechanisms that resulted in the diversification of more than 20 morphologically distinct species[25,26]. These Asian *Aquilegia* species constitute four highly divergent lineages corresponding to their geographic origins and have evolved relatively independently[25,26]. Despite this well-described evolutionary history and crucial role played by environmental factors, how

genetic and epigenetic factors are involved in the rapid speciation in this genus remains poorly investigated.

In this study, the main objective is to survey the genomes and DNA methylomes of 36 accessions from ten worldwide *Aquilegia* species from the Asian, European and North American lineages. Among the Asian species, four phylogenetically distinct species (*A. japonica, A. oxysepala, A. yabeana, and A. viridiflora*) were selected according to their geographic distributions and ecological habitats. *Aquilegia japonica* and *Aquilegia. oxysepala* are sister species inhabiting alpine tundra and low altitude forest niches in northeastern China, respectively[25,27]. Our previous studies have documented that natural selection during ecological specialization together with genetic drift under geographic isolation caused the rapid evolution of reproductive isolation between these two species[25,28]. Here, we further investigated how diverse evolutionary driving forces shaped genetic and epigenetic architectures of the two species in the processes of speciation and adaptation. In addition, we also evaluated patterns of nucleotide variation and cytosine methylation in the *A. yabeana* and *A. viridiflora*. The former species shares highly similar morphological traits and ecological niches with the *A. oxysepala* but is allopathically distributed in northern China. In contrast, while the *A. viridiflora* is sympatrically distributed with *A. yabeana* and *A. oxysepala* in northern and northeastern China, it often occupies rocky and sandy ecological niches. As a supplementary, we also examined nucleotide and cytosine methylation variation patterns of the North American and European lineages. Our study will provide a genome-wide view of how the specific genomic and epigenomic variation patterns are correlated with the diversification of Aquilegia species.

## Results

### Population structure and nucleotide variation pattern

Neighbor-joining (NJ) trees were reconstructed for the 36 *Aquilegia* accessions based on 689,123 homozygous SNPs. The phylogenic analysis suggested that these accessions of the ten species formed three distinct lineages corresponding to their geographic origins (**Figure 1a** and **Figure S1**). In brief, all 22 accessions of the four East Asian species, *A. japonica*, *A. oxysepala*, *A*. *yabeana* and *A*. *viridiflora*, clustered as a monophyletic lineage, with the first two species and their hybrid forming a clade and the last two species grouping as a sister clade. In contrast, the West Asian species *A. fragrans* clustered with the geographically adjoining European species. The principal component analysis (PCA) and population structure inferences also revealed distinct genetic structure of the three phylogenetic lineages (**Figure 1b** and **c**). It should be noted that one *A. alpina* var. alba accession shared the same ancestral genetic cluster with the North American lineage, while the putative hybrid of the *A. oxysepala* and *A. japonica* possessed an admixed genetic background (**Figure 1b** and **c**).

To further gain an insight into genome-wide nucleotide variation pattern of the ten *Aquilegia* species, we calculated nucleotide diversity ($\pi$) and genetic divergence ($F_{ST}$) for each chromosome and for 100-kb sliding windows, respectively. Among the three phylogenetic lineages, the Asian *Aquilegia* species harbored the highest nucleotide diversity compared to the European and North American lineages across the seven chromosomes (**Figure S2**). By comparing the nucleotide diversity for each 100-kb sliding window, we observed a moderate correlation of genome-wide variation pattern among the three lineages (Spearman R = 0.42-0.56) and a high correlation between the *A. oxysepala* and *A. japonica* (Spearman R = 0.70) (**Figure S3**). In particular, 116 of 241 low genetic diversity genomic regions (LDGRs, with 5% lowest $\pi$) were shared by at least two of the three lineages (**Figure S4**). Between the *A. oxysepala* and *A. japonica*, while we defined 148 LDGRs and 148 high divergence genomic regions (HDGRs, with 5% highest $F_{ST}$), only seven candidate genomic regions overlapped (**Table S1**).

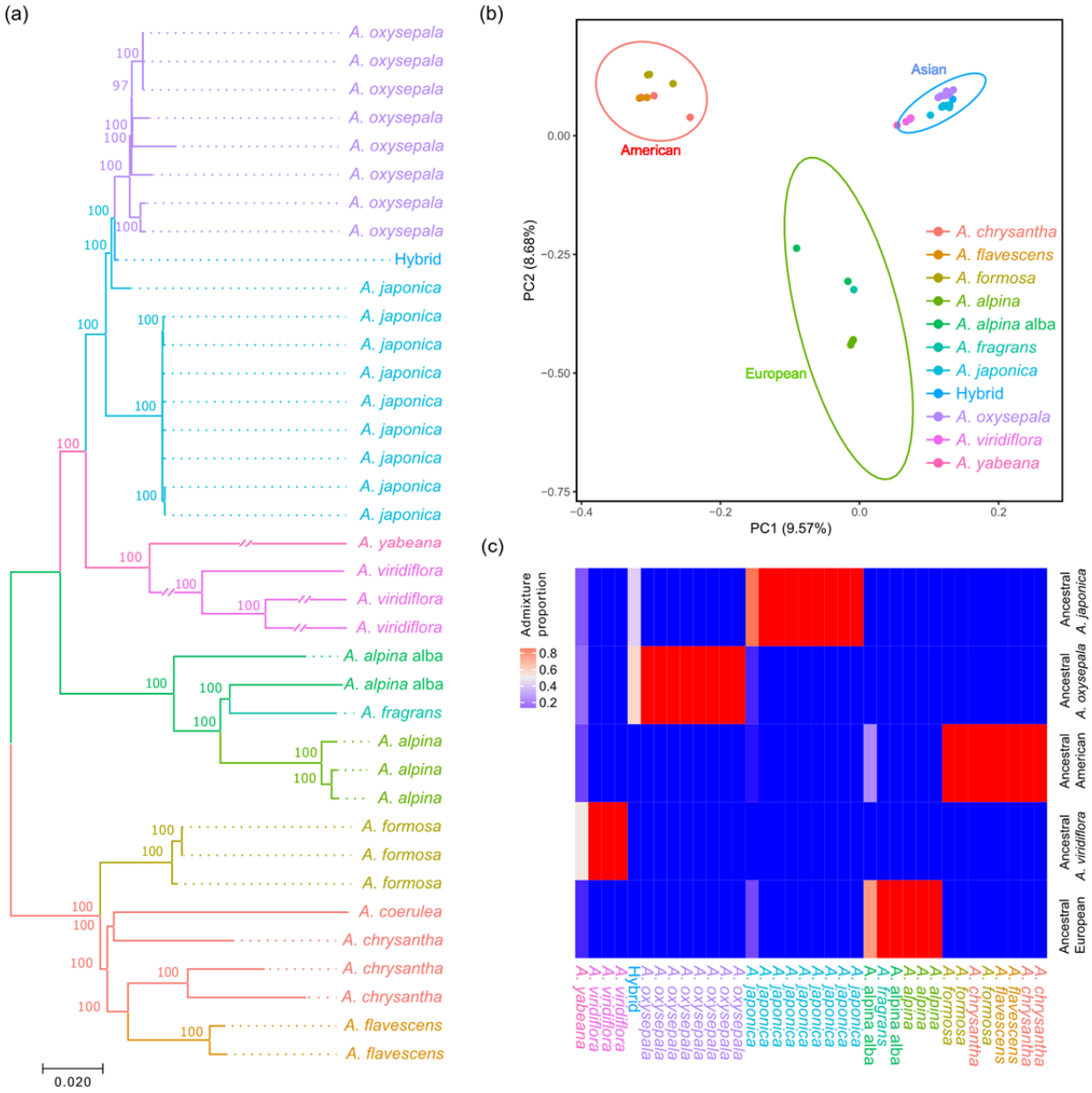


**Figure 1**. **Phylogenetic relationship and population structure of the ten worldwide *Aquilegia* species**. (a) Phylogenetic tree of the 36 accessions constructed by neighbor-joining algorithm based on 689,123 whole-genome SNPs. (b) PCA reveals genetic similarity within each of the three lineages and genetic

disparity between lineages based on 15,988 LD-pruned SNPs. Ellipses of each lineage denote 99% confidence region estimated from distribution of the first two principal components. (c) Population admixture of the 36 *Aquilegia* accessions.

**Identification of the genomic regions indicating selection pressure and highly impactful genetic variations**

Candidate genes or genomic regions associated with adaptive divergence were determined from three perspectives. First, we considered genes localized within the regions that showed low intra-specific diversity but high inter-specific divergence to be representative of intra-specific genetic differences. We thus identified 23 genes from the above seven candidate genomic regions that were both HDGRs and LDGRs shared by *A. oxysepala* and *A. japonica* (**Table S1**). Genes within these genomic regions were functionally associated with meiotic nuclear division, adenine methyltransferase and basic cellar activities.

While the first strategy mainly relied on genome-wide scanning for 100-kb non-overlapping sliding window, we also employed a functional annotation-based approach to identify highly impactful conservative clade-specific variations (CCVs) from both the within and between lineage comparisons. Our results revealed that a considerable proportion (17.9-40.5%) of the CCVs were identified in the gene body regions (**Table S2**). We then examined the potential functional impacts of genes harboring these identified CCVs. Between the *A. oxysepala* and *A. japonica*, the CCV-carrying genes were enriched in several vital biological pathways related to cell reproduction, including telomere maintenance, DNA repair, and DNA helicase activity (**Figure 2** and **Table 1**). For example, two candidate genes (*Aqcoe6G160300* and *Aqcoe7G062500*) coding for *Xklp2* (*TPX2*) were functionally correlated with spindle assembly during the mitotic process (27, 28). Among the three phylogenetic lineages, the CCVs-harboring genes were also functionally involved in the mitotic chromosome condensation, DNA ligase activity and aminopeptidase activity (**Figure 2** and **Table 1**). For instance, two CCV-containing genes (*Aqcoe2G276600* and *Aqcoe1G273400*) encoding DNA mismatch repair proteins *MutS*/*MSH* and *MutS2* (ref. 29) carried one Asian-specific-to-American frameshift variant.

Thirdly, we also derived pair-wise synonymous ($d_S$) and non-synonymous ($d_N$) mutation rate to identify genes informative of positive or purifying selection pressure. We found that species within the Asian lineage experienced significantly stronger positive ($d_N/d_S > 1$) and purifying ($d_N/d_S < 1$) selection pressures compared to the European and North American lineages (Wilcoxon rank sum test, all Bonferroni-corrected *p* values < $1.5\times10^{-16}$) (**Figure S5**). Likewise, the European species showed

significantly stronger purifying selection (Wilcoxon rank sum test, Bonferroni-corrected *p* value = $7.8 \times 10^{-8}$) compared to the North American species.

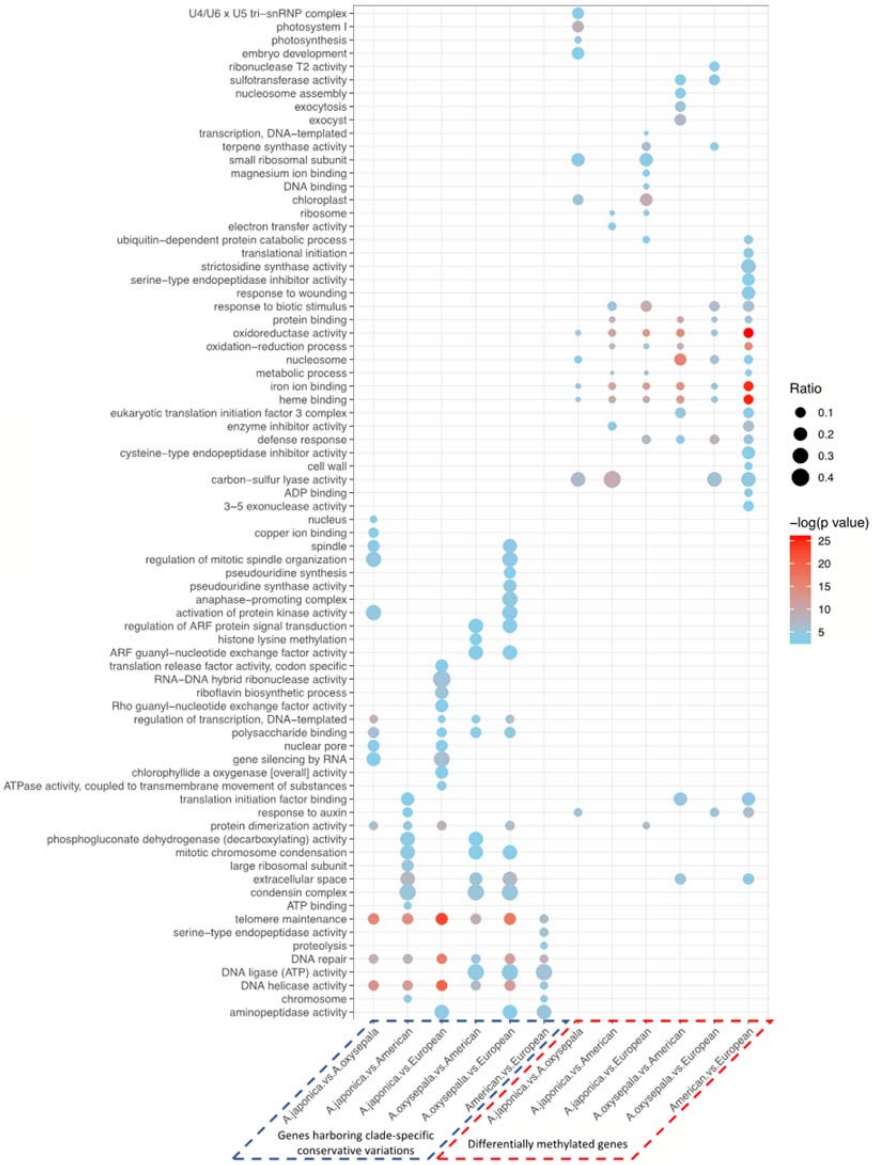


**Figure 2**. **Functional enrichment of genes harboring highly impactful CCVs and DMGs.** CCV-containing genes specific to either of the two lineages/species being compared were merged to construct a target

gene set. Ratio denotes proportion of CCV-containing genes or DMGs in the corresponding gene set of interest. Absence of dot indicates no significant enrichment.

**Table 1**. Information of the high-impact conservative clade-specific variants (CCVs) in the cell reproduction related genes.

| Gene | Variant-carrying | Reference | Chromosome | Position | Reference allele | Variant | Annotation | Gene function |
|---|---|---|---|---|---|---|---|---|
| Aqcoe1G273400 | Asian | American | Chr1 | 18994915 | GAA | GAAA | frameshift | DNA mismatch repair protein *MutS2* |
| Aqcoe2G151500 | European | American | Chr2 | 15305837 | A | G | splicing | PIF1-like helicase |
| | European | American | | 15307442 | A | C | stop gain | |
| | European | American | | 15309865 | AATATATAT | AATATATATAT | frameshift | |
| | European | Asian | | 15307442 | A | C | stop gain | |
| | European | Asian | | 15309865 | AATATATAT | AATATATATAT | frameshift | |
| | *A. oxysepala* | *A. japonica* | | 15305837 | A | G | splicing | |
| | *A. oxysepala* | *A. japonica* | | 15309267 | AT | A | frameshift | |
| Aqcoe2G177700 | European | American | Chr2 | 21794397 | TATGCACCAAAGGTATCACGATGC | TATGC | frameshift | PIF1-like helicase |
| | European | American | | 21794979 | TT | TTGT | frameshift | |
| | European | Asian | | 21794397 | TATGCACCAAAGGTATCACGATGC | TATGC | frameshift | |
| | *A. oxysepala* | *A. japonica* | | 21795089 | CA | C | frameshift | |
| Aqcoe6G208600 | European | American | Chr6 | 15364081 | A | ATCTCTTCG | frameshift | PIF1-like helicase |
| | European | Asian | | 15364081 | A | ATCTCTTCG | frameshift | |
| | *A. japonica* | *A. oxysepala* | | 15364330 | TAA | TA | frameshift | |
| Aqcoe6G253800 | European | American | Chr6 | 22789898 | C | T | stop gain | DNA helicase |
| | European | American | | 22790012 | G | A | splicing | |
| | European | Asian | | 22789898 | C | T | stop gain | |
| | *A. japonica* | *A. oxysepala* | | 22790012 | G | A | splicing | |
| Aqcoe2G276600 | Asian | American | Chr2 | 33314422 | AGGGGG | AGGGGGG | frameshift | DNA mismatch repair protein *Msh6* |
| Aqcoe6G160300 | *A. japonica* | *A. oxysepala* | Chr6 | 9414625 | G | A | stop gain | *TPX2* |
| Aqcoe7G062500 | *A. oxysepala* | *A. japonica* | Chr7 | 3789055 | G | A | stop gain | cell cycle regulated microtubule associated protein |

**CG methylation patterns and differentially methylated genes**

In parallel with the above genomic analyses, we also investigated CG methylation pattern of the representative *Aquilegia* species. Despite variability across the 36 *Aquilegia* accessions, the North American, Asian and European species showed no distinguishable differences (*t* test, all Bonferroni-corrected *p* values > 0.01) in overall percentage of methylated cytosines (**Figure 3a**). We then performed PCA to examine the CG-cytosine methylomic diversity of all the *Aquilegia* accessions. The resulting overall methylation pattern highly resembled the above genomic inferences, with the European and American species forming two distinct groups and the four Asian species forming three separate clusters (**Figure S6**). We then assessed the CG methylation patterns for the European and North American lineages as well as the three Asian species (*A. japonica*, *A. oxysepala* and *A. viridiflora*) separately. Consistent with the described genomic features, heterogeneous pattern of the CG methylation was also observed for the seven chromosomes, with the chromosome 4 demonstrating obviously higher overall CG methylation divergence compared to the other six chromosomes (**Figure 3b**). We further quantified CG methylation level deposited in the genic regions, putative *cis*-regulatory regions and CG island, respectively. In genic and regulatory regions, all three lineages shared similar modification patterns with apparent depletion of CG methylation around the transcription start site (TSS) and transcription end site (TES) (**Figure 3c**). However, the American lineage exhibited hyper-methylation (more than 10%) around the center of CG islands and a more drastic decrease throughout the CG island shores compared to the European and Asian species (**Figure 3d**).

To examine the biological impacts of CG methylation on the species diversification, differentially methylated regions (DMRs) and differentially methylated genes (DMGs) were identified for both within- and between-lineage comparisons, respectively (**Tables S3** and **S4**). Within the Asian lineage, 3,622 DMRs in 2,899 DMGs were identified between the *A. japonica* and *A. oxysepala*. Functional enrichment of these DMGs indicated that the two species may have different activities in photosynthesis-related pathways, including photosystem I, photosynthesis and chloroplast (**Figure 2**). For example, two photosynthesis-related genes, *PsaA*/*PsaB* and *CemA*, showed significantly differential methylation between the two species in the genic regions (**Figure S7a** and **b**). At the inter-lineage level, apparently

more DMGs were identified between the North American and European species (6,087 genes) compared to those of between the two lineages and Asian species (3,308-5,003 genes) (**Table S3** and **S4**). DMGs characterized from the inter-lineage comparisons were mainly involved in the plant growth (e.g., response to auxin) and defense, response to biotic stimulus and wounding (**Figure 2**).

We then examined whether the candidate genes (CCV-carrying genes and DMGs) superimposed on the same signature of natural selection. We found while a considerable proportion of the candidate genes were shared for each of the genetics- and epigenetics-based assessments (**Figure S8**), they showed a segregated distribution pattern across all comparisons (**Figure S9**). Likewise, the Gene Ontology (GO) enrichment analyses of the candidate gene identified from the genetic and epigenetic levels were enriched in functionally complementary pathways (**Figure 2**), suggesting co-existence of different underlying evolutionary mechanisms.

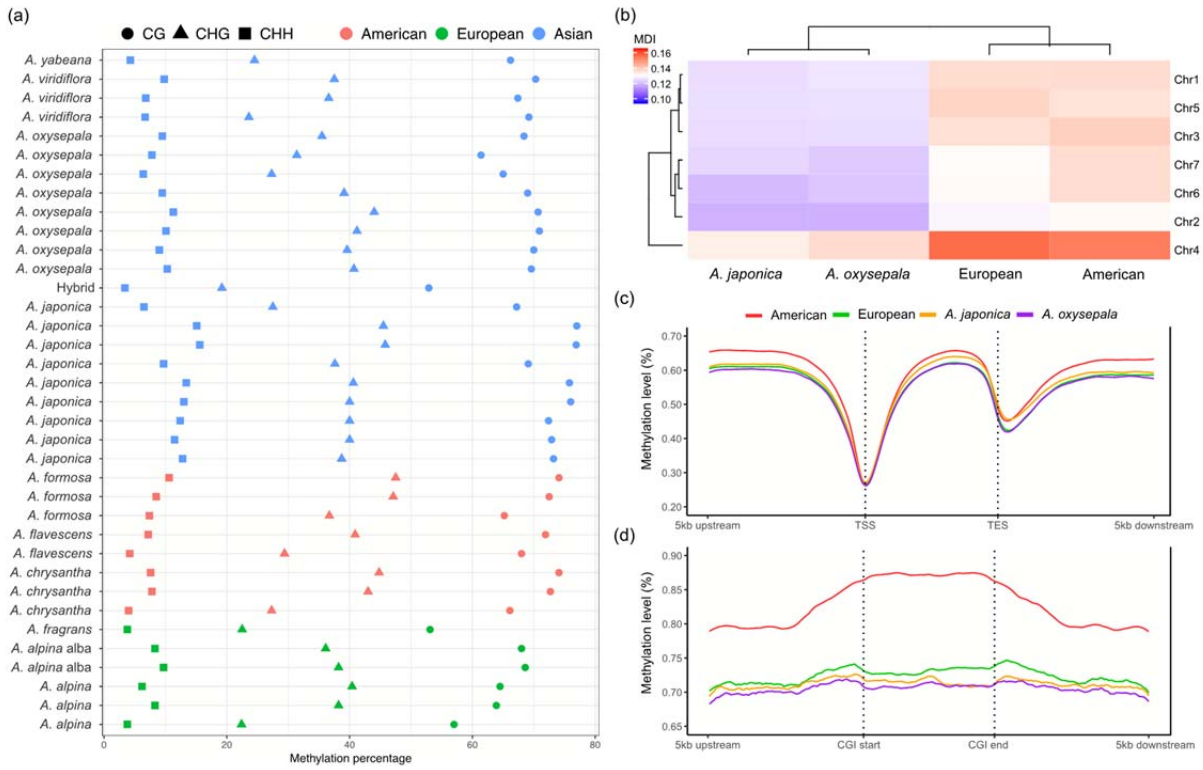


**Figure 3**. **Patterns of cytosine methylation for the ten worldwide *Aquilegia* species.** (a) Genome-wide cytosine methylation levels of 36 accessions. (b) MDI illustrates chromosome-level CG methylation similarity. *Aquilegia viridiflora* was used as the reference. (c) CG methylation profiling in genic region across the four *Aquilegia* groups. Each row represents one genic region starting at 5-kb upstream of its TSS and terminating at 5-kb downstream of its TES, sorted by mean methylation level of all analyzed CG loci. Gene body regions were scaled to have the same length. (d) CG methylation profiling in and around CG islands.

**Association between epigenetic variability and genetic variations**

Since both genetic variation and differential CG methylation seemed to have crucial and multifaceted influences on the adaptation of the ten *Aquilegia* species, we wondered whether differential epigenetic modifications were dependent on genetic variations. Among the 588,659 CG loci examined, 224,222 (38.09%) carried a CG-loss variation. We then illustrated epigenetic variability for the variation-carrying and non-variant CG loci, respectively. As shown in Figure 4, genetic-epigenetic associations of varying magnitude were observed in both types of CG loci. The variation-carrying CG loci conveyed information that highly resembled their genetic background. The overall methylation pattern was highly conserved within the same species but exhibited obvious divergence across the ten *Aquilegia* species (**Figure 4a**). In contrast, CG methylation divergence at the non-variant CG loci varied with higher variability at both the intra- and inter-specific levels (**Figure 4b**). By examining the correlation of genetic variability and cytosine methylation, we found that CG methylation divergence at variation-carrying CG-site was largely attributable to the CG-loss variations (**Figure 4c**). In particular, 75% of the CG-loss variations occurring at the most highly variable CG-methylated dinucleotides could explain at least 75% of the total epigenetic variability *per se*. Nevertheless, there was still a considerable proportion of epigenetic variability that could not be sufficiently explained by the variant-CG locus (**Figure 4d**).

We also attempted to identify *cis*-driver mutations for each of the 1,229 DMRs between the *A. japonica* and *A. oxysepala*. Our results revealed that only 568 out of the 1,229 (46.2%) DMRs were significantly associated with at least one genetic variation inside or around a 500 base-pair (bp) upstream/downstream genomic region, even under the least stringent *p* value threshold ($5\times10^{-5}$), indicating that the epigenetic changes were only partially dependent on *cis*-genetic driving mutations (**Figure 4e**). Moreover, we observed weak yet significant associations between differential CG methylation and selection pressure. In most inter-lineage comparisons, DMGs were significantly more prone to be under positive selection than non-DMGs (**Table 2**), suggesting that epigenetic modifications could probably assist selection pressure in shaping genotypes. In contrast, DMGs were significantly less prone to be under purifying selection (**Table 2**).

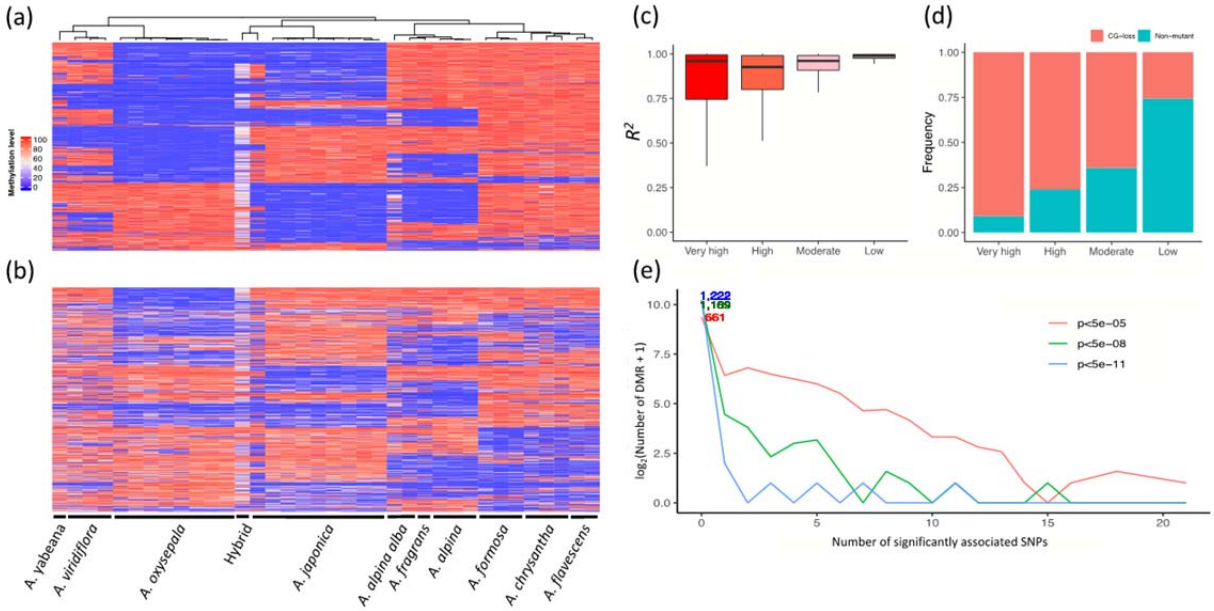


**Figure 4**. **Association between CG-loss variations and epigenetic variability**. (a) Top 3,000 most variable CG loci containing CG-loss variations. (b) Top 3,000 most variable non-variant CG loci across 36 accessions show clade-specific methylation patterns. CG methylation in the hybrid tends to be neutralized possibly due to heterozygosity. (c) Linear regression demonstrates that CG-loss variations explain a large proportion of CG methylation variation. (d) Summary of composition of each category with regard to whether each CG locus contains a CG-loss variation. Epigenetic variability was determined by standard deviation in methylation β value across all 36 accessions. CG loci with top 10,000, 10,001-50,000 and 50,001-150,000 largest standard deviation were ordinally labelled as possessing "very high", "high" and "moderate" variability respectively. The rest CG loci were labelled as possessing "low" variability. (e) Association test shows most DMRs were independent of *cis*-acting SNPs. Results under different significance levels are compared in this exploratory analysis.

**Table 2.** Significant correlation between differential methylation and natural selection.

| Type of selection | Differential methylation | Jap-Oxy* | Jap-Ame | Jap-Eur | Oxy-Ame | Oxy-Eur | Ame-Eur |
|---|---|---|---|---|---|---|---|
| Positive selection | DMG | 7.2% | 7.3% | 11.9% | 6.7% | 8.4% | 8.9% |
| | non-DMG | 4.4% | 5.0% | 5.6% | 4.7% | 5.4% | 5.4% |
| | *p* value | 0.11 | 7.3e-02 | 3.9e-05 | 6.7e-02 | 1.8e-02 | 2.8e-04 |
| Purifying selection | DMG | 3.1% | 1.8% | 2.4% | 2.3% | 2.0% | 1.9% |
| | non-DMG | 4.3% | 4.3% | 4.7% | 4.9% | 5.1% | 4.0% |
| | *p* value | 0.53 | 3.2e-02 | 9.1e-02 | 1.3e-02 | 8.4e-03 | 1.0e-02 |

*: Each percentage represents the proportion of genes belonging to either DMGs or non-DMGs compared between the two corresponding clades that are under corresponding or higher strength of positive selection. For example, 7.2% indicates that 7.2% DMGs compared between *A. japonica* and *A. oxysepala* are under strong selection; 4.4% indicates that 4.4% non-DMGs compared between these two species are under strong selection. *p* values were obtained from Chi-square tests and were not adjusted for multiple testing due to dependence arising from overlapping gene sets.

Jap: *A. japonica*; Oxy: *A. oxysepala*; Ame: American; Eur: European

## Discussion

### Genetically determined mechanisms associated with the rapid diversification of *Aquilegia* species

Elucidating evolutionary mechanisms underpinning species diversification is crucial to understanding the evolution and persistence of biodiversity[2,5,6]. The genus *Aquilegia* provides an ideal system to address how the diverse evolutionary mechanisms promoted rapid adaptive radiation[16,17]. Although various environmental conditions related to ecological opportunities, such as shifts in pollinator and habitat, have been proposed to facilitate the evolution of reproductive isolation[21,25], genetic basis associated with the rapid diversification of *Aquilegia* species has still remained largely unclear. In this study, we surveyed the genomes of ten worldwide *Aquilegia* species to address whether specific genetic architectures have been involved in the rapid species diversification. Broadly consistent with previously inferred phylogeny[17,19,25,30], the ten *Aquilegia* species from Asia, Europe and North America formed three phylogenetically independent lineages corresponding to their geographic origins. This attribute renders the *Aquilegia* species a suitable system to identify genomic variations associated with the repeated adaptive speciation by extensive comparisons from different facets.

It has been proposed that if a genetic factor is the potential determinant promoting adaptive speciation, one would expect to identify specific genetic architectures in the diversified lineages[8,14,31]. In Darwin's finches, for example, polyphyletic topology was observed as a general pattern in 14 morphologically distinct species, phenotypic diversity of the beak shape was mainly determined by natural selection acting on the *ALX1* gene during the ecological specialization process[8]. A similar phenomenon was observed in the East African cichlid fish where the radiating lineages are more dynamic in terms of gene content and transcriptomic landscape compared to their non-radiating relatives[14,31]. In this study, the genome-wide nucleotide variation pattern highly reflects the evolutionary history that the Asian, European and North American *Aquilegia* species have clearly diverged and evolved allopatrically in respective geographic regions. This also suggests that a considerable proportion of the genetic variations and changes in environment are intertwined during the diversification process. As expected, our genome-wide scanning for selection signatures revealed distinct positive and purifying selection modes in the intra- and inter-lineage comparisons. More importantly, the CCV-carrying genes

identified from the three lineages are associated with cell reproduction (e.g., telomere maintenance and mitotic chromosome condensation) and other functionally important traits. Similar genomic feature was also observed in *A. japonica* and *A. oxysepala*. Our previous studies have demonstrated that natural selection and genetic drift together resulted in the rapid evolution of reproductive isolation[25,30]. Here, we further demonstrate that candidate genes involved in the adaptive speciation are functionally enriched in the pathways related to cell reproduction (e.g., telomere maintenance), stress tolerance (e.g., response to wounding) and basic cellular activities. It should be noted that although a majority of the enriched pathways are specific to each comparison, enrichment of cell reproduction-related pathways (e.g., telomere maintenance, DNA repair and DNA helicase activity) and stress tolerance are shared in the intra- and inter-lineage comparisons. Taken together, these findings indicate that specific genetic determinants might have conferred high adaptability to the Aquilegia species to cope with different environmental conditions.

**Evolutionary potential of cytosine methylation in the adaptation of *Aquilegia* species**

The role of epigenetic modification in the long-term evolutionary process has long been debated[32–34]. It has been proposed that epigenetic variations are frequently under the genetic control which can alter rapidly as a result of environmental induction and stochastic epimutation[35,36]. Nevertheless, it has also been recognized that some epigenetic variations can persist over generations and be highly correlated with phenotypic diversity[32]. As illustrated in Arabidopsis, changes in cytosine methylation can produce meiotically stable epialleles, which could eventually lead to phenotypic diversity in the absence of genetic variations[37–39]. Here, we assessed whether the epigenetic modifications were also associated with the adaptive speciation of the *Aquilegia* species. Consistent with the genomic features detailed above, high divergence of cytosine methylation was observed across the Asian, European and North American lineages. Notably, differential cytosine methylation was not only found across the seven chromosomes but also evident in the gene body of DMGs and CG island region among the three lineages. Particularly, functional enrichment analyses identified significant associations with adaptation-related traits, including plant growth, stress tolerance and basic cellular activities. For example, the candidate DMGs identified between the *A. japonica* and *A. oxysepala*, showed significant enrichment in pathways related to diverse important phenotypic traits, such as photosynthesis, embryo development and response to auxin. These features indicated that epigenetic factors might also play a role in response to diverse environmental conditions.

We noted that some candidate genes and enriched pathways had shared hotspots of both genetic and epigenetic disparities, especially those related to cell reproduction, plant growth and stress tolerance. Many studies based on human and mouse have shown that genetic variations can manipulate *cis*-CG methylation at specific loci to further influence phenotypes, where CG methylation serves as a mediator[40,41]. By analyzing the associations between genetic and epigenetic variability, we conclude that while many CG-loss variations can directly lead to depletion of CG methylation, a lot of DMRs are not manipulated by any *cis*-variations. Since gene body CG methylation in plants generally stabilizes gene expression and is positively correlated with gene expression[42–45], differential methylation in our study is indicative of likely differential amount of gene products. Based on these attributes together with the plausible associations between differential methylation (e.g., DMGs) and positive selection (e.g., $d_N/d_S$), we propose that epigenetic modification may be a complementary mechanism facilitating phenotypic diversity of the *Aquilegia* species.

**Limitations and future directions**

We realized that this study has some limitations. Fistful, the small sample size in this study may introduce bias and inflation of false positives, and we postulate that our findings should be interpreted carefully and considered exploratory. When the association between genetic divergence and evolutionary events is investigated, it is impossible to deny the roles of other evolutionary forces. We acknowledge that the lineage-specific allele frequencies are possibly consequences of genetic drift, and genetic hitchhiking may lead to identification of candidate genes residing in neighboring genomic regions representing the other driving forces. Therefore, we claim that the candidate genes identified to be associated with adaptive radiation do not necessarily point towards causal evolutionary mechanisms. They may also be by-products of the long-term process of adaptive radiation. In addition, we never than less only focused on analysis of CG methylation as puzzles persist regarding the functional roles of CHG and CHH methylation. We also expect that future studies with larger sample sizes will be able to improve the statistical power and investigate *trans*-genetic control.

## Methods and Materials

### Sample collection, DNA extraction and whole-genome sequencing

In this study, a total of 36 accessions from ten worldwide *Aquilegia* species were collected (**Table S5**). Among the Asian species, four phylogenetically distinct species (*A. japonica*, *A. oxysepala, A*. *yabeana*, and *A*. *viridiflora*) are selected according to their geographic distributions and ecological habitats. *Aquilegia japonica* and *A. oxysepala* are sister species inhabiting alpine tundra and low altitude forest niches in northeastern China, respectively[25,46]. Eighteen accessions were collected to represent these two Asian species and their putative hybrid. In addition, four accessions were collected from the other two Asian species, *A*. *yabeana* and *A*. *viridiflora*. The former species shares highly similar morphological traits and ecological niches with *A. oxysepala* but is allopatrically distributed in the northern China. In contrast, *A. viridiflora* is sympatrically distributed with *A*. *yabeana* and *A. oxysepala* in the northern and northeastern China, but often occupies rocky and sandy ecological niches. Furthermore, six and eight accessions were sampled from the European and North American lineages, respectively. All the 36 accessions were grown in green house under the same conditions (25°C/12 hours, 16°C/12 hours). Genomic DNA was extracted from fresh mature leaves using TianGen plant genomic DNA kit. Whole genome resequencing and bisulfite sequencing were performed on the extracted genomic DNA using the Illumina X-ten platform (Illumina, California, USA). Short-insert (350 bp) DNA libraries of all accessions were constructed by NovoGene (NovoGene, Tianjin, China). Genome assembly of an admixed species *A. coerulea* "Goldsmith" was obtained from Phytozome v12.1 (https://phytozome.jgi.doe.gov) as the reference genome[16].

### Sequence assembly, functional annotation and genetic diversity

Whole genome sequences of each accession were aligned against the reference genome using default settings of the BWA-MEM algorithm implemented in Burrows-Wheeler Aligner (BWA)[47]. Raw assemblies were realigned using IndelRealigner provided in the Genome Analysis Tool Kit by default settings[48].

495 Single nucleotide polymorphisms (SNPs) and insertions/deletions (INDELs) were reported using
496 SAMtools[49]. Only the high-quality variants (SNPs and INDELs) (read depth > 3, mapping quality > 20 and
497 missing allele < 1%) were retained for subsequent population genomics analyses. Genomic annotation of
498 the identified variants was reported for each of the 36 samples separately. Functional annotation of
499 each identified variant was performed using SnpEff, based on the reference genome[50].
500 To infer the phylogenetic relationship between the ten *Aquilegia* species, NJ trees were
501 reconstructed for each chromosome and the whole genome dataset using MEGA 7(ref. 51). PCA was
502 carried out to examine the genetic diversity of the 36 *Aquilegia* accessions[52]. Ancestral components
503 were estimated using ADMIXTURE[53] with different number of populations ranging from one to ten.
504 Optimal population composition with the least 5-fold cross-validation error was selected to decompose
505 ancestral admixture. To obtain the genome-wide nucleotide variation pattern, nucleotide diversity ($\pi$)
506 and genetic differentiation (Weir and Cockerham's $F_{ST}$) were calculated for each 100 kb non-overlapping
507 sliding window using VCFtools[54,55]. Pair-wise non-synonymous-to-synonymous ($d_N/d_S$) ratios of the ten
508 species were inferred by yn00 program in the Phylogenetic Analysis by Maximum Likelihood (PAML)
509 package[56]. Inter-lineage $d_N/d_S$ value for each gene was derived by averaging $d_N/d_S$ values obtained from
510 all pairwise comparisons of samples belonging to the two lineages under investigation. Candidate genes
511 with the 5% highest and 5% lowest $d_N/d_S$ values were considered to have undergone strong positive and
512 purifying selection, respectively.
513
514 **Cytosine methylation pattern and epigenetic population structure**
515 Whole genome bisulfite sequencing data were pre-processed using TrimGalore (https://www.
516 bioinformatics.babraham.ac.uk/projects/trim_galore/, accessed August 21, 2018). Paired-end reads
517 were then aligned to the reference genome using Bismark[57] with a moderately stringent minimum-score
518 function (L,0,-0.3). De-duplicated alignments of the 36 *Aquilegia* accessions were used to report cytosine
519 methylation level using "Bismark_Methylation_Extractor", on loci with a read depth ≥ 3. Genomic
520 annotations of the methylated cytosine site were identified based on the reference genome using an in-
521 house Python script. PCA was conducted for 588,659 loci which were passed the quality control to infer
522 the CG-methylomic diversity of the ten *Aquilegia* species. Differential cytosine methylation was
523 determined at the gene and chromosome levels, respectively. At the gene level, we determined DMRs
524 for each 100 bp non-overlapping sliding window using Cochran-Mantel-Haenszel (CMH) test to account
525 for imbalanced read depth (Supplementary Notes). Genomic regions that possessed a Benjamin-
526 Hochberg adjusted *p* value < 0.05 and showed inter-specific or inter-lineage methylation divergence

527 higher than 25% were defined as significant DMRs. Genes with > 20% of the genic region being DMR(s)
528 were defined as DMGs. Chromosome level methylation patterns were measured by chromosomal
529 methylation discrepancy index (MDI)[58]. Methylation patterns of the identified DMGs were visually
530 confirmed on Integrative Genomics Viewer[59] prior to downstream analyses and biological interpretation.
531 In addition, we identified CG islands from the *A. coerulea* "Goldsmith" reference genome using EMBOSS
532 cpgplot with default settings[60]. Only the identified CG-enriched genomic regions with > 200 bp were
533 defined as CG islands. We then investigated inter-specific and inter-lineage methylation patterns in and
534 around the CG islands.
535
536 **Associations between the genetic variation and cytosine methylation**
537 We tested for associations between the identified DMGs and genes under positive selection by a Chi-
538 square test. Linear regression model was adopted to measure the direct causal effect of CG-loss
539 variation on CG methylation. To further assess whether genetic variations drive the establishment of
540 DMG, driving mutations of DMRs between the *A. japonica* and *A. oxysepala* were identified using an
541 Eigenstrat-based method (See Supplementary Notes for more details)[61].
542
543 **Identification of conservative clade-specific variant**
544 CCVs were defined as variants that had a SnpEff-predicted "high" functional impact and that were
545 conserved across all samples belonging to the same species or lineage, but not present in any sample of
546 the other species/lineages. Since the biological consequences of heterozygous variants were less
547 affirmable, only the homozygous point mutations and INDELs were included in the characterization of
548 CCVs, including frameshift, stop-gain, stop-loss, start-loss and splicing-alteration variations.
549
550 **Functional analysis**
551 The above mentioned genetic and epigenetic analyses helped to identify relevant candidate genes,
552 which might be associated with the rapid diversification of the *Aquilegia* species from different
553 perspectives. These candidate genes were employed to conduct functional enrichment analyses using
554 the R package topGO with default settings[62]. Enriched GO terms that possessed a $p$ value <0.05 were
555 considered statistically significant. Since the statistical tests performed by topGO are not independent,
556 multiple testing correction does not apply here[62]. Structures of functional domains of targeted genes
557 were determined based on the InterPro database (https://www.ebi.ac.uk/interpro, accessed January 25,

2019). Distribution patterns of the identified candidate genes and their related functional pathways were visualized using the R package jvenn[63].

**Data availability**

All data generated from the study were submitted to EBI under the accession number PRJEB34182. All scripts for conducting computational analyses are available upon reasonable request to the corresponding author.

**Author Contributions**

L.F.L., J.Z and A.S.X conceived this project. Z.H.W, T.L. and M.R.L. developed statistical analysis pipeline. Z.H.W., T.L., M.R.L., N.D., L.Z.L., Y.J.H and X.G. carried out experiments and analyzed the data. T.L., M.R.L., N.D., Z.H.W., L.Z.L., X.G., and L.F.L. participated in discussion and interpreted the data. Z.H.W, T.L. and L.F.L. wrote the manuscript. All authors read and approved the manuscript.

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**Competing Interests**

The authors have declared that no competing interests exist.

**Supplementary files**

**Figure S1**. Per-chromosome phylogenetic trees reconstructed using neighbor-joining algorithm. Polymorphisms detected on each chromosome were retrieved separately to infer the phylogeny.

**Figure S2**. Distribution of nucleotide diversity ($\pi$) at the whole-genome level and the per-chromosome level. Nucleotide diversity was estimated for each lineage pooling corresponding species, as well as for *A. japonica* and *A. oxysepala*.

**Figure S3**. Spearman correlation of the genome-wide nucleotide variation pattern for each 100-kb sliding window between European and Asian (a), North American and Asian (b), European and North American (d), *Aquilegia japonica* and *A. oxysepala* (d). Each dot represents a 100-kb sliding window. Values on the x- and y-axis are the nucleotide diversity ($\pi$) for each sliding window.

**Figure S4**. Overlapping of low diversity genomic region (LDGR) between the three lineages. 148 LDGRs with 5% lowest nucleotide diversity were defined as LDNRs in each lineage, totaling 241 unique regions.

**Figure S5**. Pair-wise $d_N/d_S$ ratio for all genes between the species within and between the Asian, European and North American lineages.

**Figure S6**. PCA illustrates three distinct clusters corresponding to the three lineages. Asian species further demonstrated higher inter-specific divergence than the American and the European species. PCA was performed based on 588,659 loci with sufficiently high sequencing quality.

**Figure S7**. Illustration of differential methylation in two photosynthesis genes. CG methylation pattern of two genes, *Aqcoe7G230600* photosystem I *PsaA*/*PsaB* (a) and *Aqcoe7G231300 CemA* (b) in *A. japonica* and *A. oxysepala* throughout the gene body region. Red bars indicate methylation level (0-100) at CG loci. Genomic coordinates on the chromosome 7 are annotated.

**Figure S8**. Overlapping of the CCVs (a) and DMGs (b) identified in inter-lineage/species comparisons. A considerable proportion of these CCVs (84.1%) and DMGs (51.3%) were shared by two or more inter-lineage/species comparisons.

**Figure S9**. Venn analyses of the candidate genes carrying CCVs and DMGs. Each subpanel indicates the comparison between the *A. japonica* and *A. oxysepala* (a), *A. japonica* and North American (b), *A. japonica* and European (c), *A. oxysepala* and North American (d), *A. oxysepala* and European (e), North American and European (f).

**Table S1**. Candidate genomic regions that showed high genetic divergence (top 5% highest $F_{ST}$) between *Aquilegia japonica* and *A. oxysepala* and low nucleotide diversity (top 5% lowest $\pi$) within each species.

**Table S2**. Summary of the highly impactful clade specific variations (CCVs) at both the species and lineage levels.

**Table S3**. Statistics for differentially methylated regions (DMRs) among the four *Aquilegia* lineages or species. Odds ratio estimates the relative methylation level between two lineages or species being compared in corresponding region. DMRs were sorted by genomic coordinates with hypo-methylated DMRs in the first lineage/species preceding hyper-methylated DMRs.

**Table S4**. Statistics for differentially methylated genes (DMGs) among the four *Aquilegia* lineages/ species. Only genes harboring a high density of differentially methylated regions (> 2 per kb) were considered DMGs.

**Table S5**. Information of the 36 *Aquilegia* samples used in this study.

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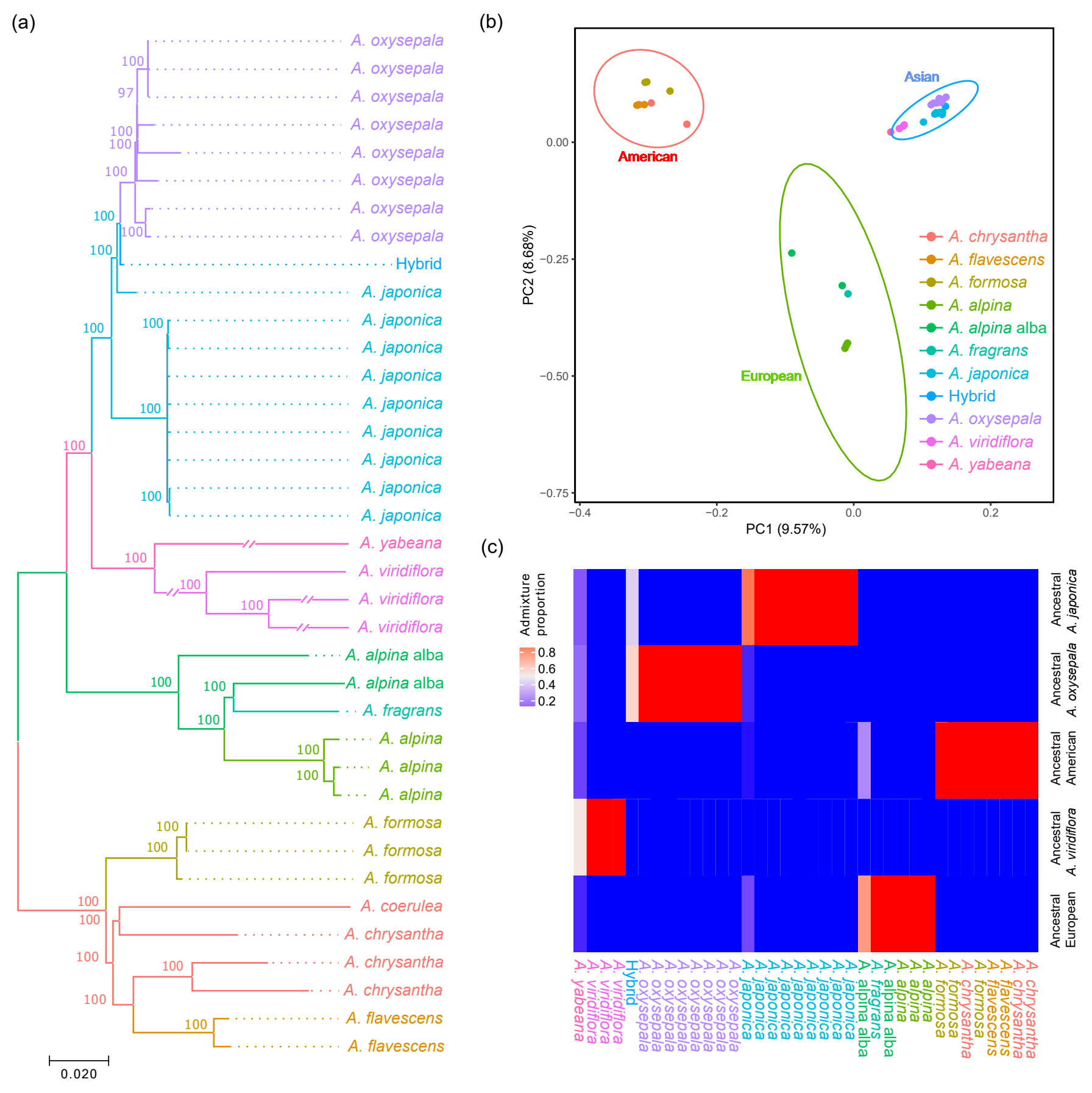
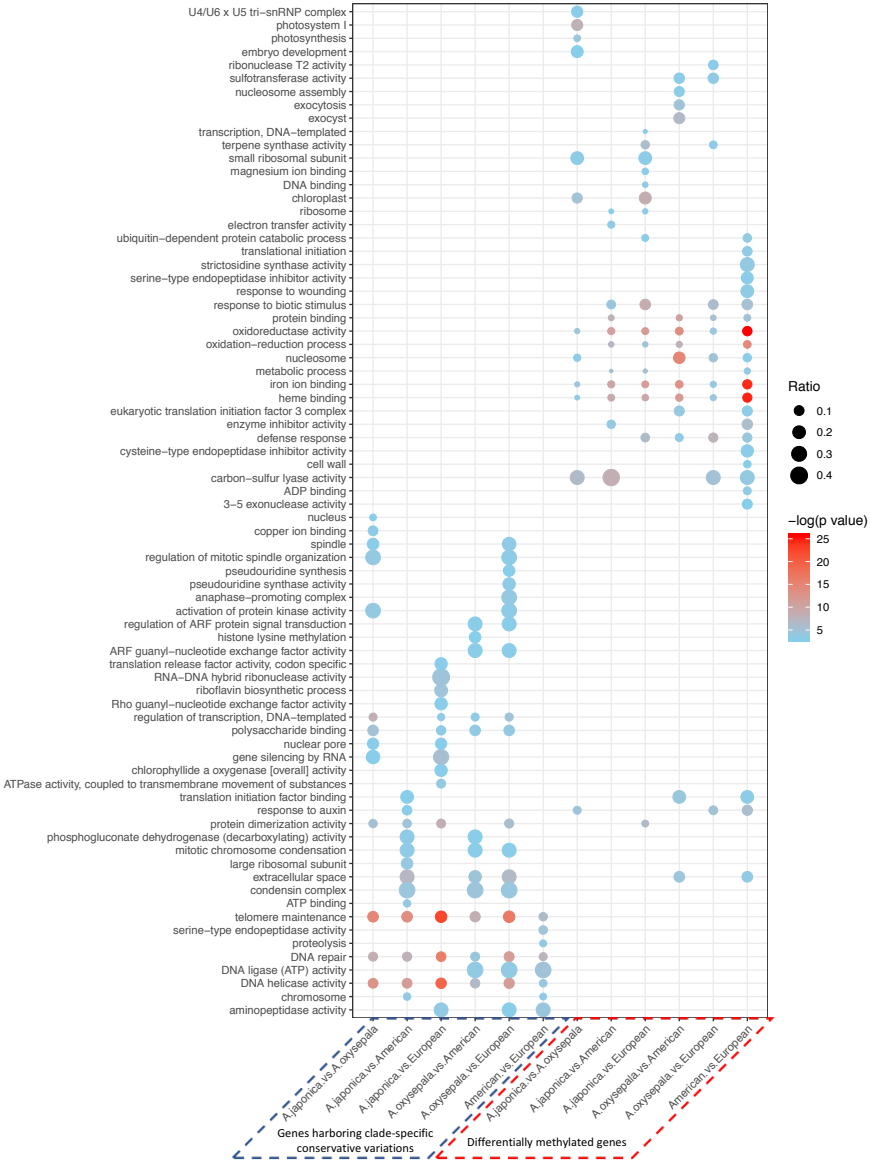
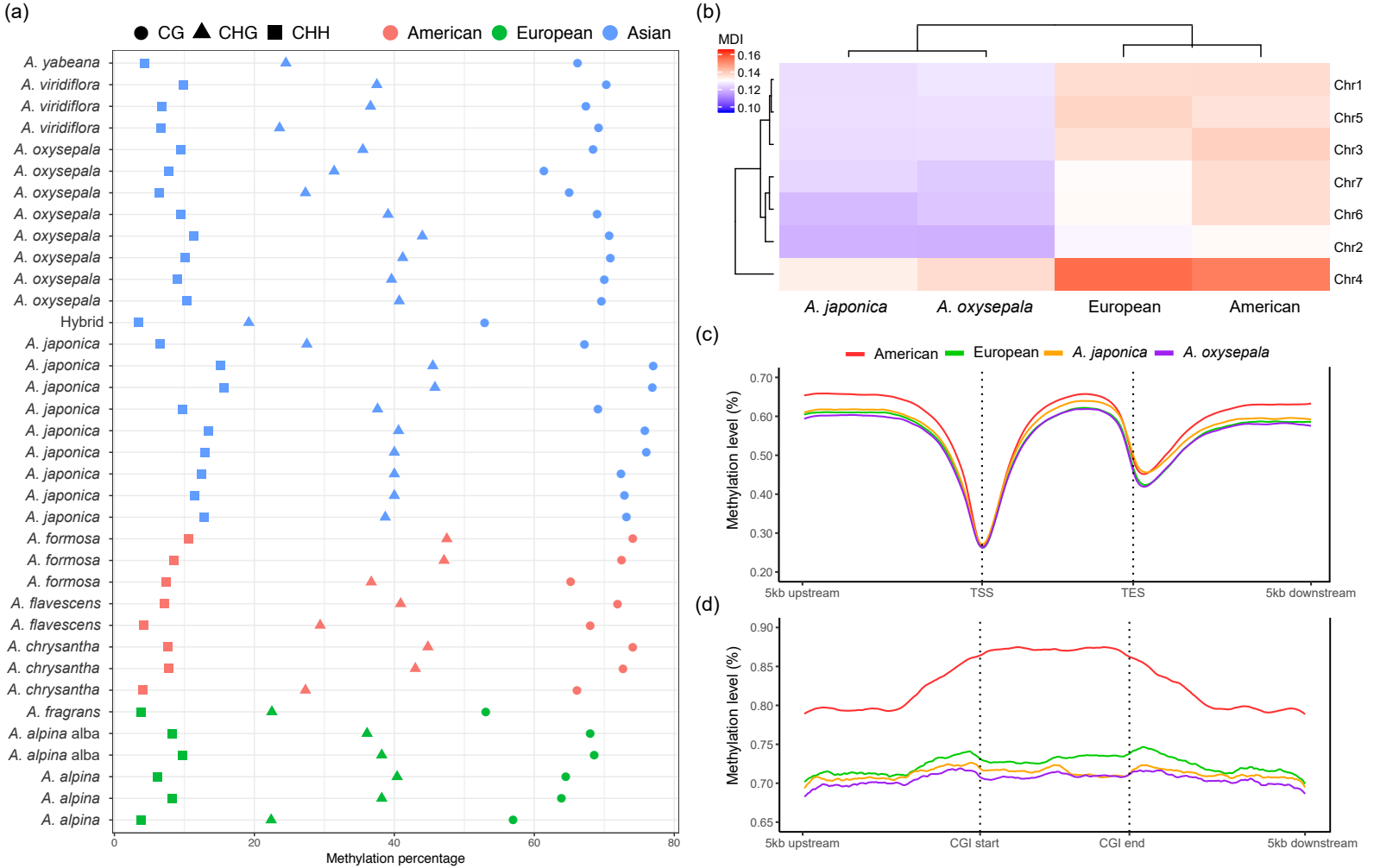
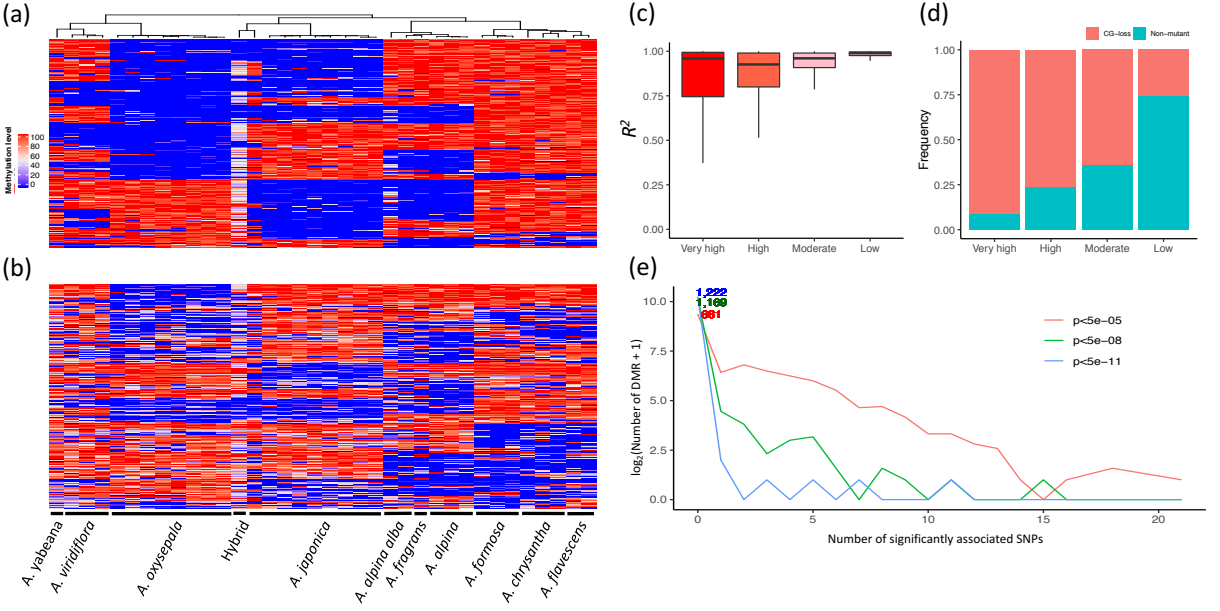
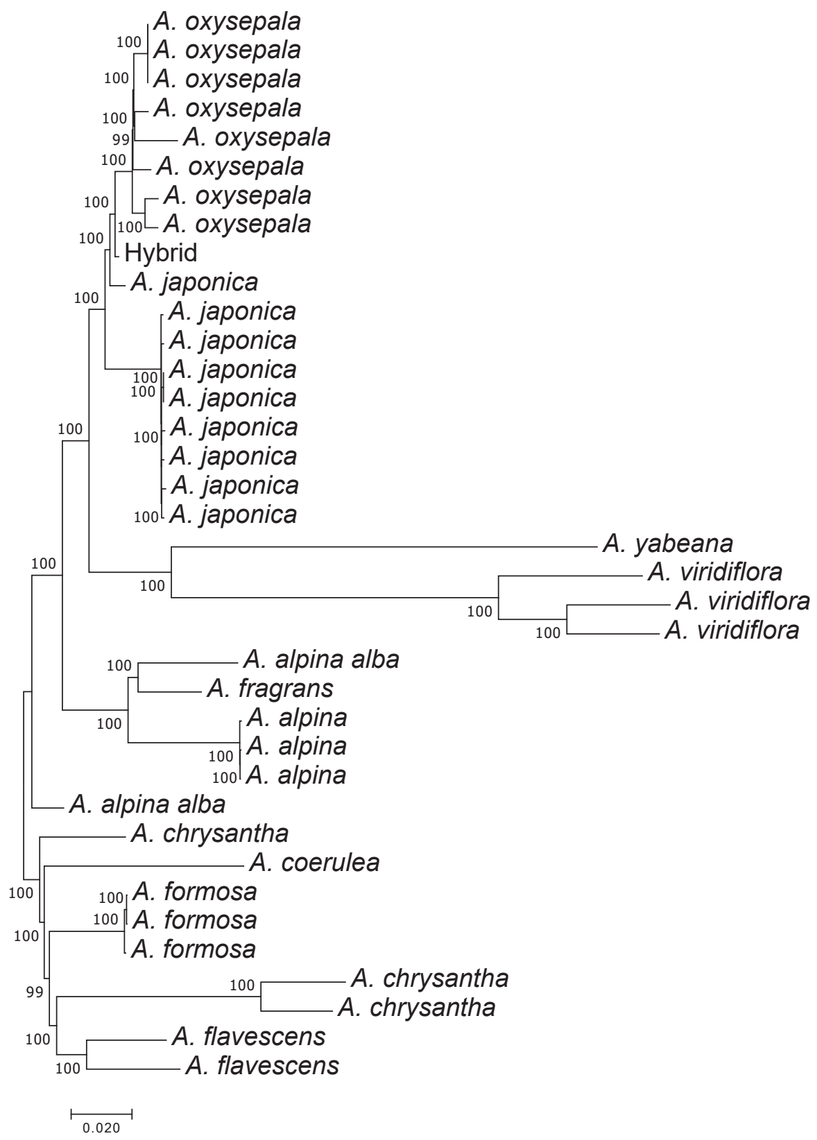
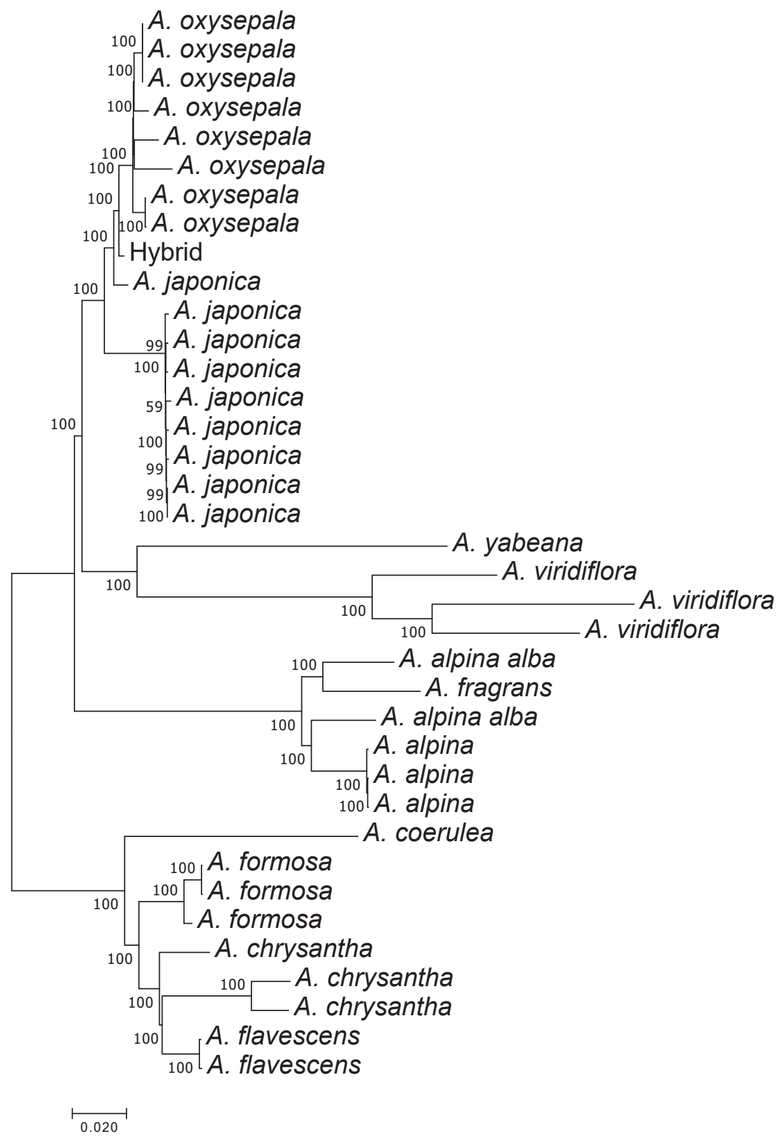

(a)
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
Hybrid
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. yabeana
A. viridiflora
A. viridiflora
A. viridiflora
A. alpina alba
A. alpina alba
A. fragrans
A. alpina
A. alpina
A. alpina
A. formosa
A. formosa
A. formosa
A. coerulea
A. chrysantha
A. chrysantha
A. chrysantha
A. flavescens
A. flavescens
0.020
(b)
Asian
American
European
PC2 (8.68%)
PC1 (9.57%)
0.00
−0.25
−0.50
−0.75
−0.4
−0.2
0.0
0.2
A. chrysantha
A. flavescens
A. formosa
A. alpina
A. alpina alba
A. fragrans
A. japonica
Hybrid
A. oxysepala
A. viridiflora
A. yabeana
(c)
Admixture proportion
0.8
0.6
0.4
0.2
Ancestral A. japonica
Ancestral A. oxysepala
Ancestral American
Ancestral A. viridiflora
Ancestral European

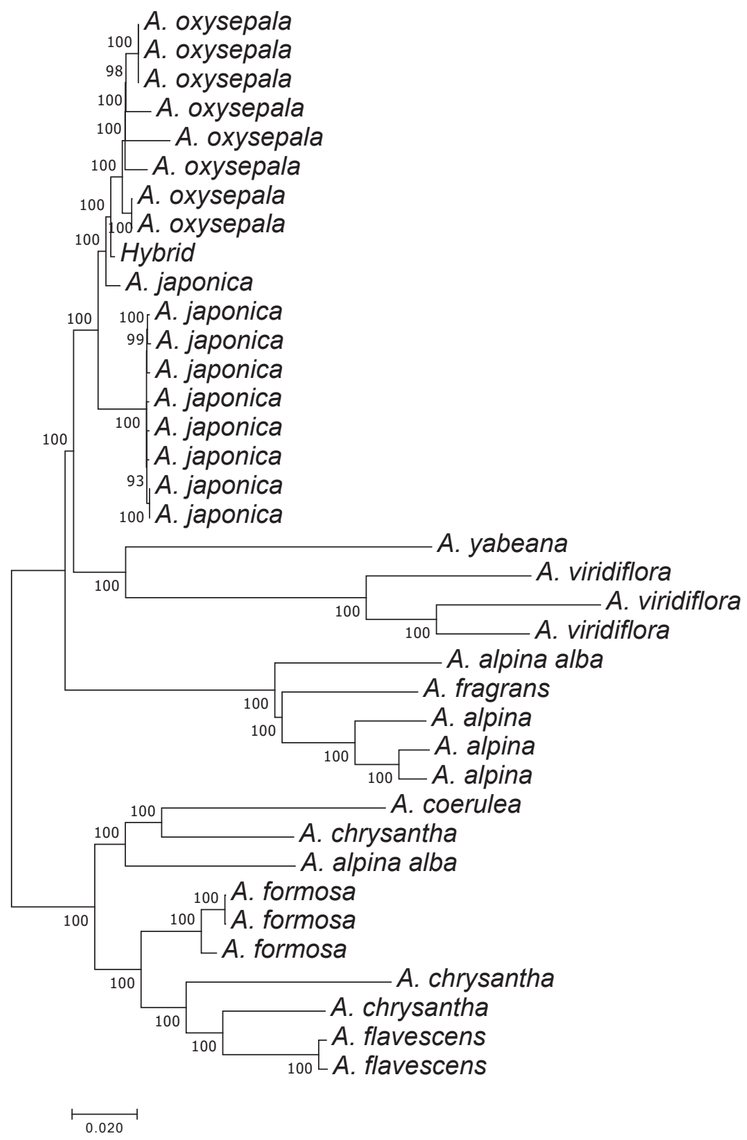

U4/U6 x U5 tri−snRNP complex
photosystem I
photosynthesis
embryo development
ribonuclease T2 activity
sulfotransferase activity
nucleosome assembly
exocytosis
exocyst
transcription, DNA−templated
terpene synthase activity
small ribosomal subunit
magnesium ion binding
DNA binding
chloroplast
ribosome
electron transfer activity
ubiquitin−dependent protein catabolic process
translational initiation
strictosidine synthase activity
serine−type endopeptidase inhibitor activity
response to wounding
response to biotic stimulus
protein binding
oxidoreductase activity
oxidation−reduction process
nucleosome
metabolic process
iron ion binding
heme binding
eukaryotic translation initiation factor 3 complex
enzyme inhibitor activity
defense response
cysteine−type endopeptidase inhibitor activity
cell wall
carbon−sulfur lyase activity
ADP binding
3−5 exonuclease activity
nucleus
copper ion binding
spindle
regulation of mitotic spindle organization
pseudouridine synthesis
pseudouridine synthase activity
anaphase−promoting complex
activation of protein kinase activity
regulation of ARF protein signal transduction
histone lysine methylation
ARF guanyl−nucleotide exchange factor activity
translation release factor activity, codon specific
RNA−DNA hybrid ribonuclease activity
riboflavin biosynthetic process
Rho guanyl−nucleotide exchange factor activity
regulation of transcription, DNA−templated
polysaccharide binding
nuclear pore
gene silencing by RNA
chlorophyllide a oxygenase [overall] activity
ATPase activity, coupled to transmembrane movement of substances
translation initiation factor binding
response to auxin
protein dimerization activity
phosphogluconate dehydrogenase (decarboxylating) activity
mitotic chromosome condensation
large ribosomal subunit
extracellular space
condensin complex
ATP binding
telomere maintenance
serine−type endopeptidase activity
proteolysis
DNA repair
DNA ligase (ATP) activity
DNA helicase activity
chromosome
aminopeptidase activity
A.japonica.vs.A.oxysepala
A.japonica.vs.American
A.japonica.vs.European
A.oxysepala.vs.American
A.oxysepala.vs.European
American.vs.European
A.japonica.vs.A.oxysepala
A.japonica.vs.American
A.japonica.vs.European
A.oxysepala.vs.American
A.oxysepala.vs.European
American.vs.European
Genes harboring clade-specific conservative variations
Differentially methylated genes
Ratio
0.1
0.2
0.3
0.4
−log(p value)
25
20
15
10
5

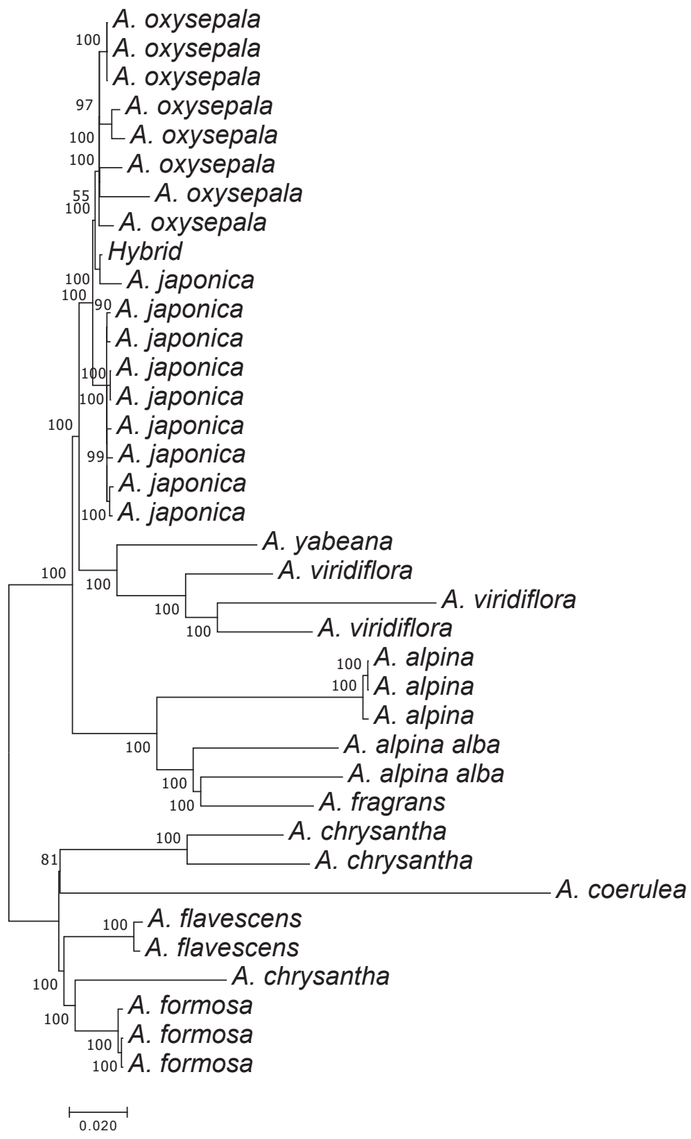

(a)
CG
CHG
CHH
American
European
Asian
A. yabeana
A. viridiflora
A. viridiflora
A. viridiflora
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
A. oxysepala
Hybrid
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. japonica
A. formosa
A. formosa
A. formosa
A. flavescens
A. flavescens
A. chrysantha
A. chrysantha
A. chrysantha
A. fragrans
A. alpina alba
A. alpina alba
A. alpina
A. alpina
A. alpina
0
20
40
60
80
Methylation percentage
(b)
MDI
0.16
0.14
0.12
0.10
Chr1
Chr5
Chr3
Chr7
Chr6
Chr2
Chr4
A. japonica
A. oxysepala
European
American
(c)
American
European
A. japonica
A. oxysepala
Methylation level (%)
0.70
0.60
0.50
0.40
0.30
0.20
5kb upstream
TSS
TES
5kb downstream
(d)
Methylation level (%)
0.90
0.85
0.80
0.75
0.70
0.65
5kb upstream
CGI start
CGI end
5kb downstream

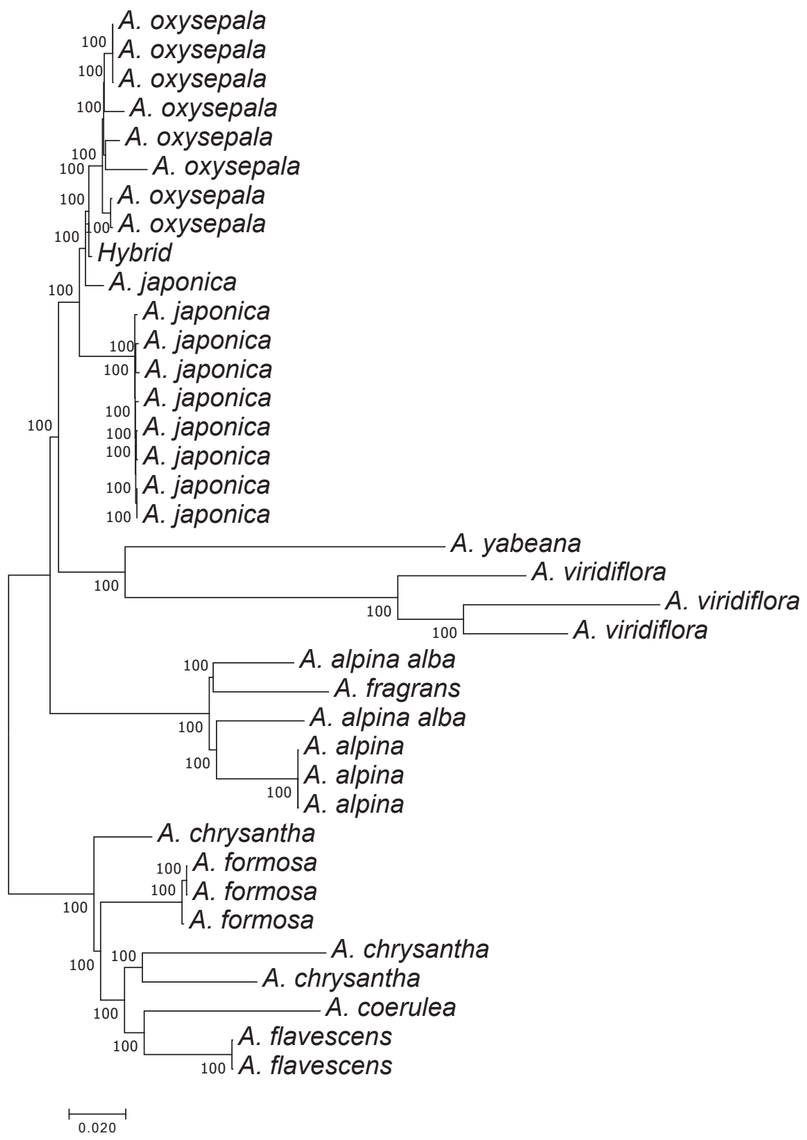

(a)
Methylation level
100
80
60
40
20
0
(b)
A. yabeana
A. viridiflora
A. oxysepala
Hybrid
A. japonica
A. alpina alba
A. fragrans
A. alpina
A. formosa
A. chrysantha
A. flavescens
(c)
$R^2$
1.00
0.75
0.50
0.25
0.00
Very high
High
Moderate
Low
(d)
CG-loss
Non-mutant
Frequency
1.00
0.75
0.50
0.25
0.00
Very high
High
Moderate
Low
(e)
1,222
1,169
581
p<5e−05
p<5e−08
p<5e−11
log2(Number of DMR + 1)
10.0
7.5
5.0
2.5
0.0
0
5
10
15
20
Number of significantly associated SNPs

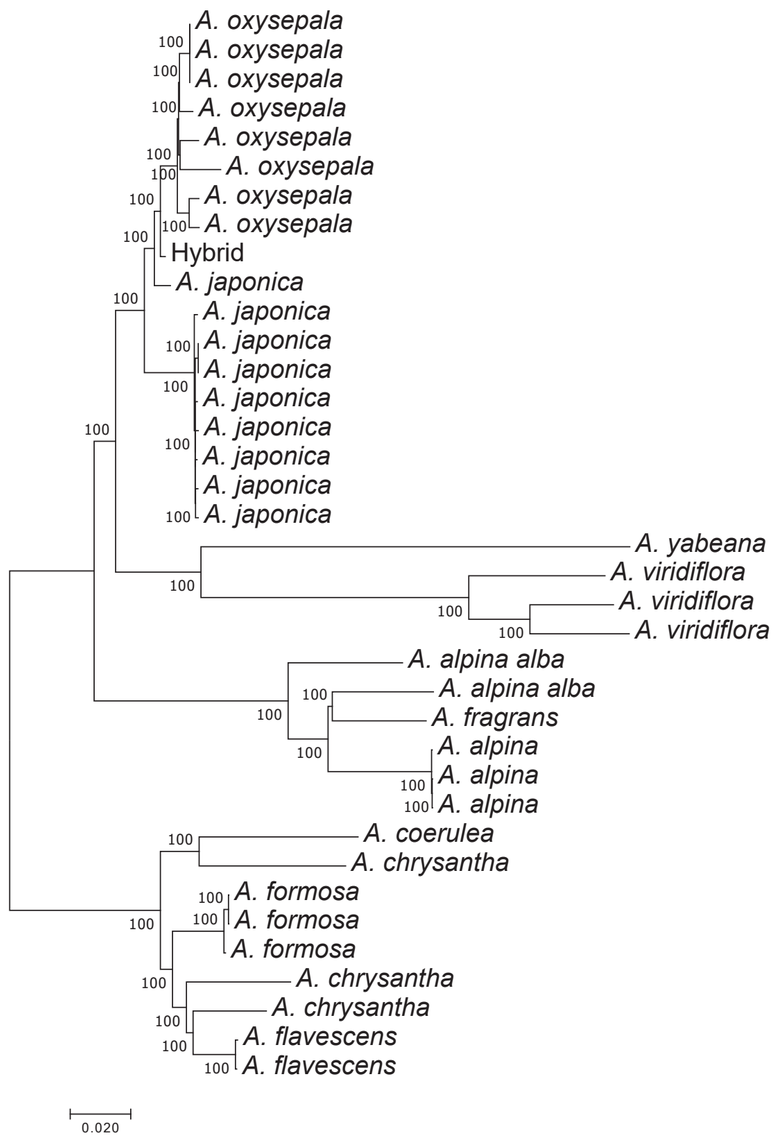

Chr1
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. coerulea
A. formosa
A. chrysantha
A. flavescens
0.020

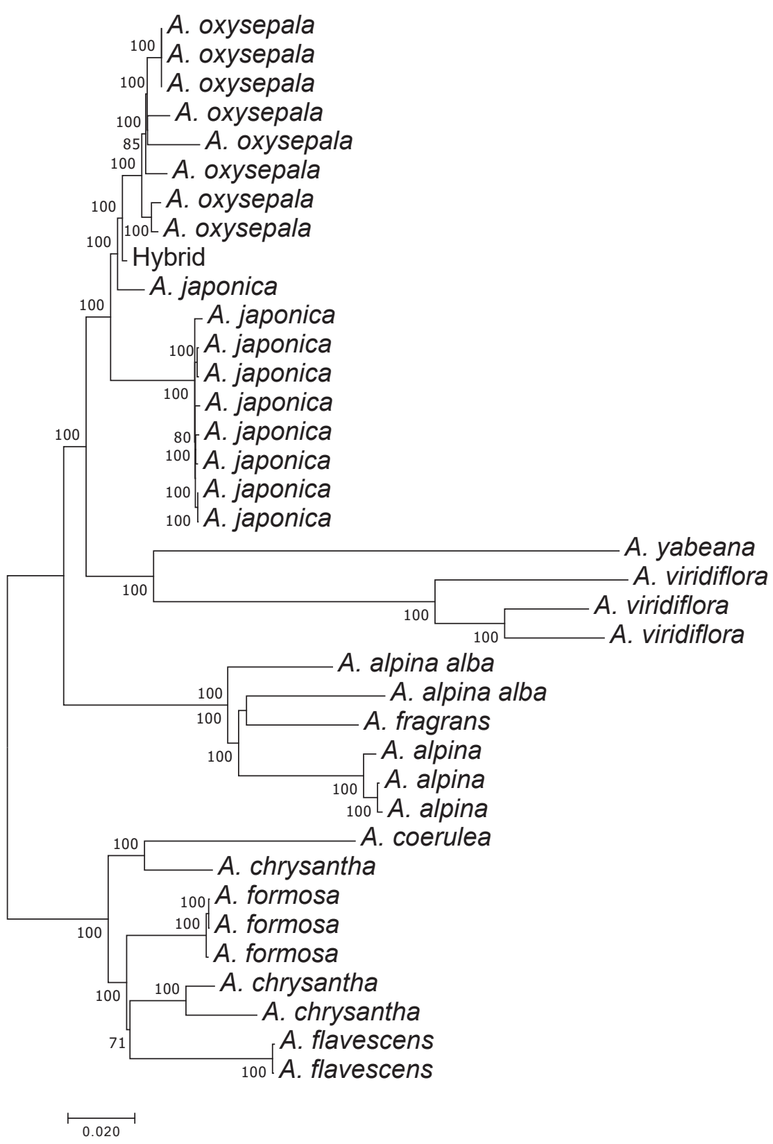

Chr2
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. coerulea
A. formosa
A. chrysantha
A. flavescens
0.020

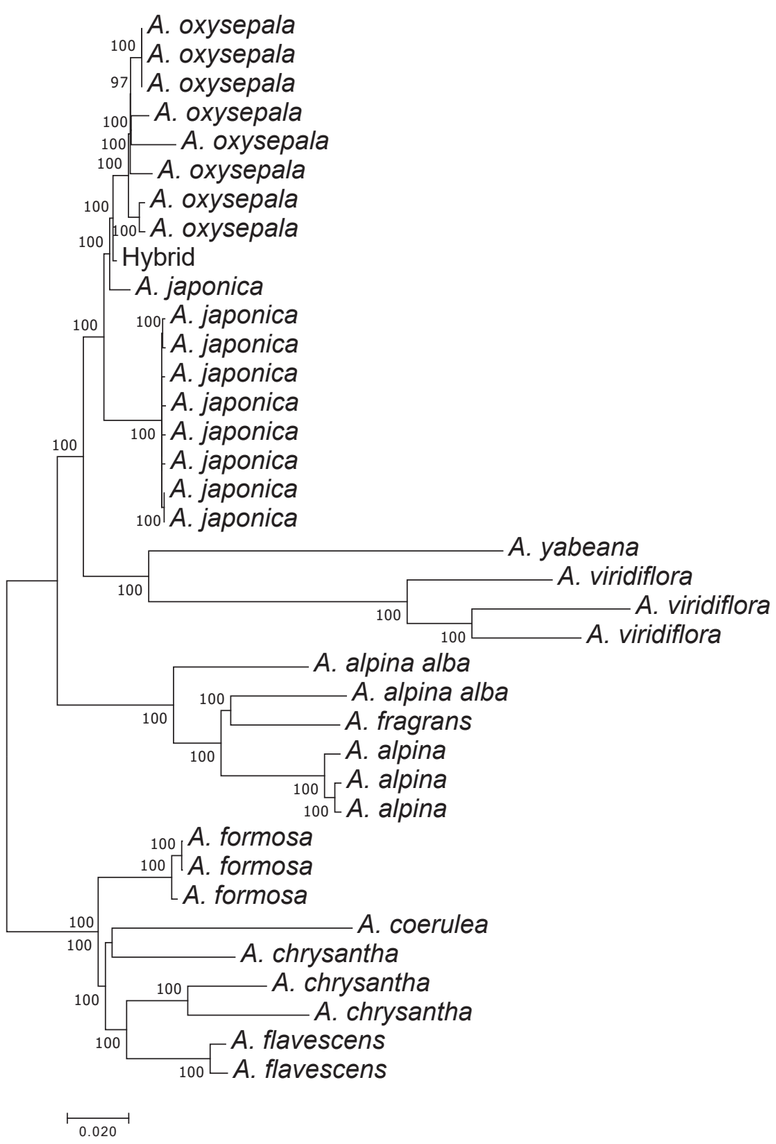

Chr3
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. coerulea
A. chrysantha
A. formosa
A. flavescens
0.020

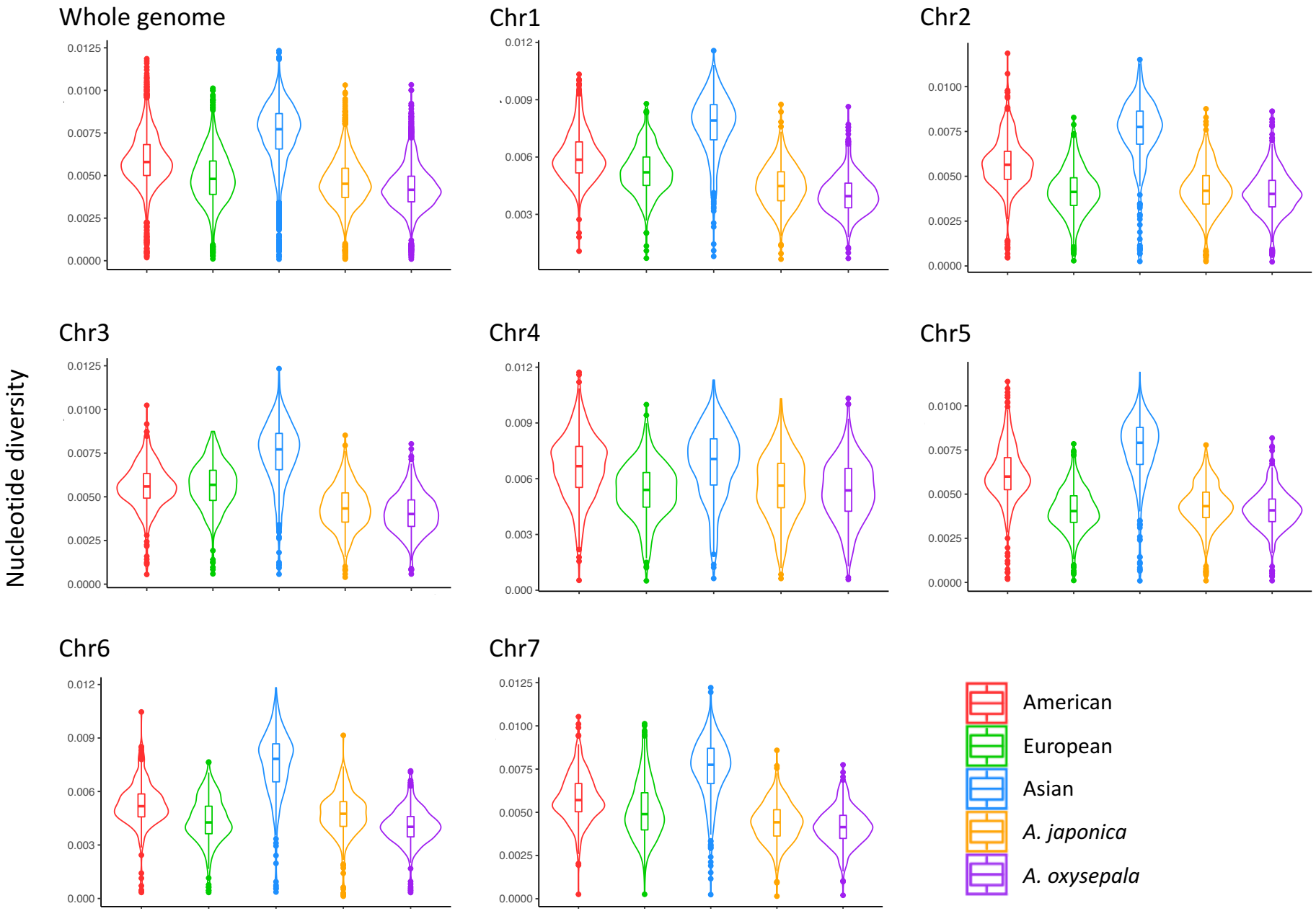

Chr4
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina
A. alpina alba
A. fragrans
A. chrysantha
A. coerulea
A. flavescens
A. formosa
0.020

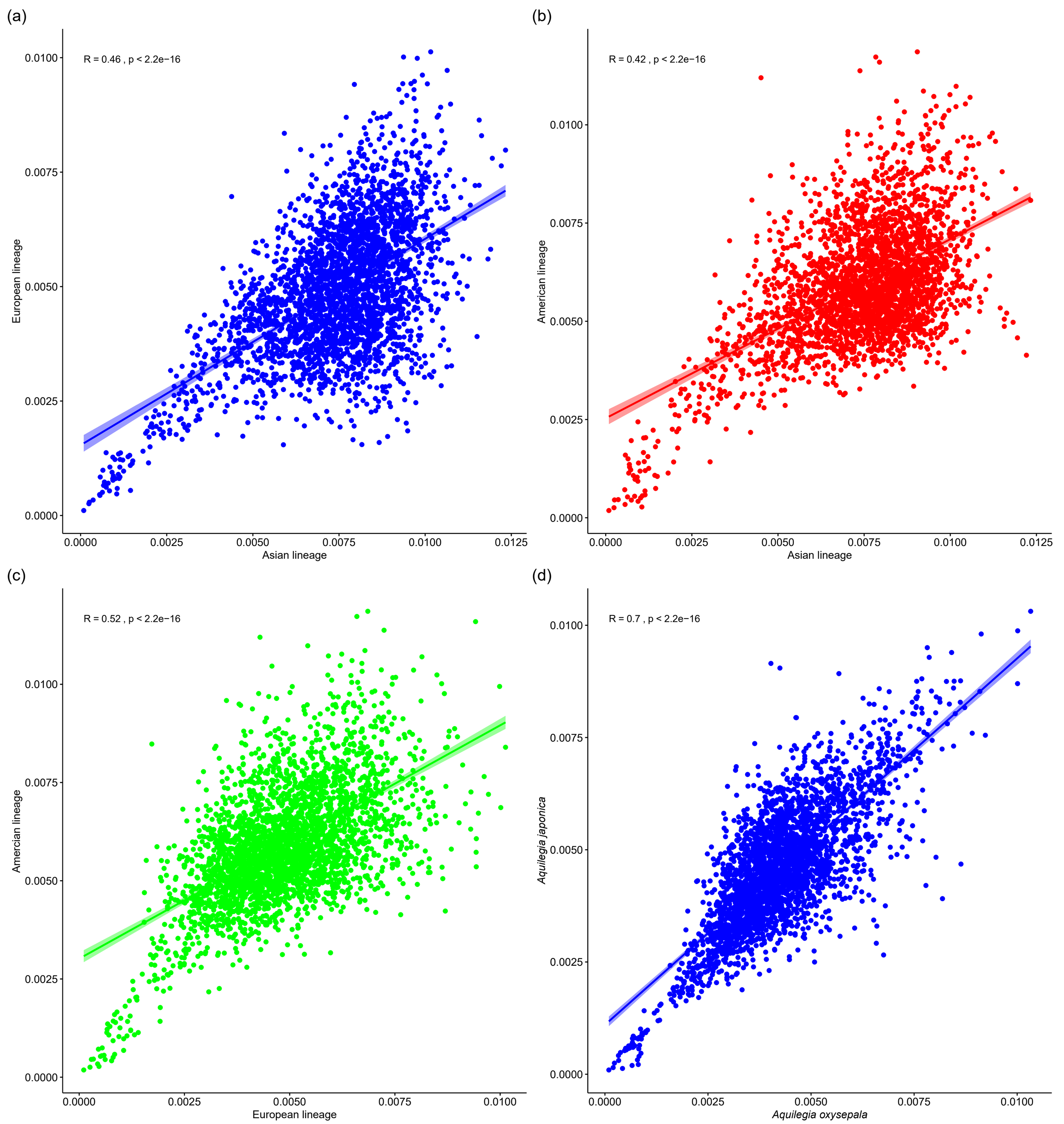

Chr5
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. chrysantha
A. formosa
A. coerulea
A. flavescens
0.020

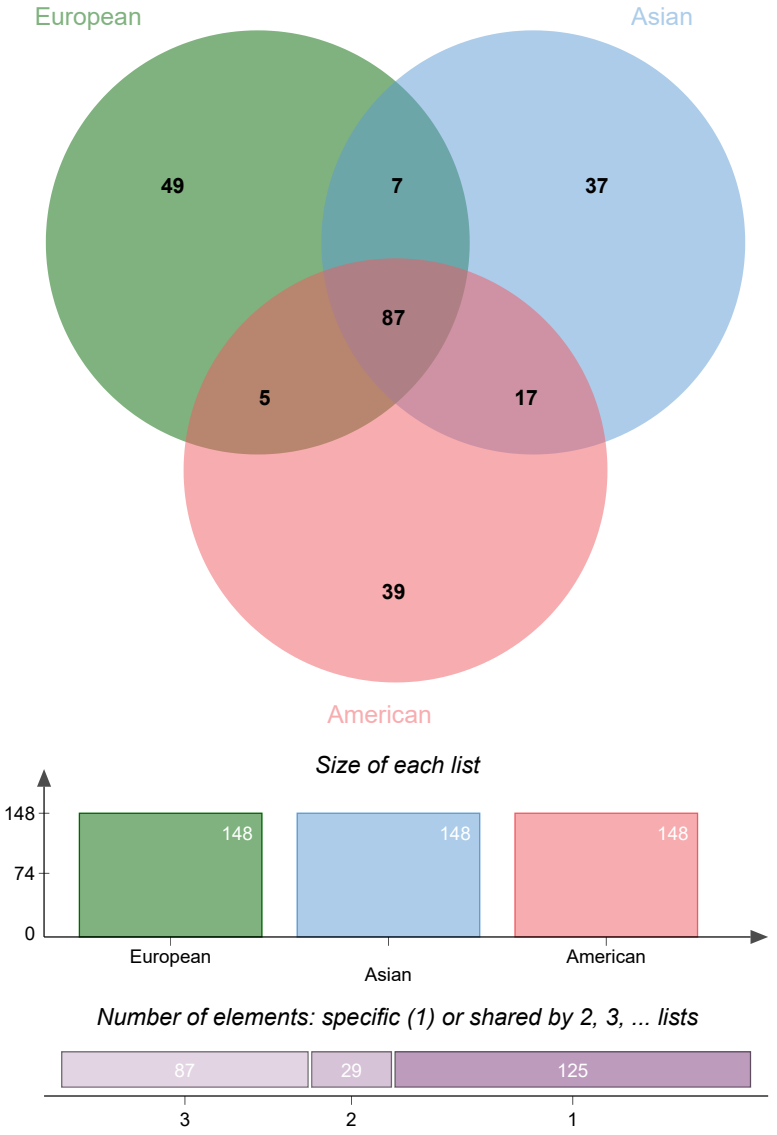

Chr6
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. coerulea
A. chrysantha
A. formosa
A. flavescens
0.020

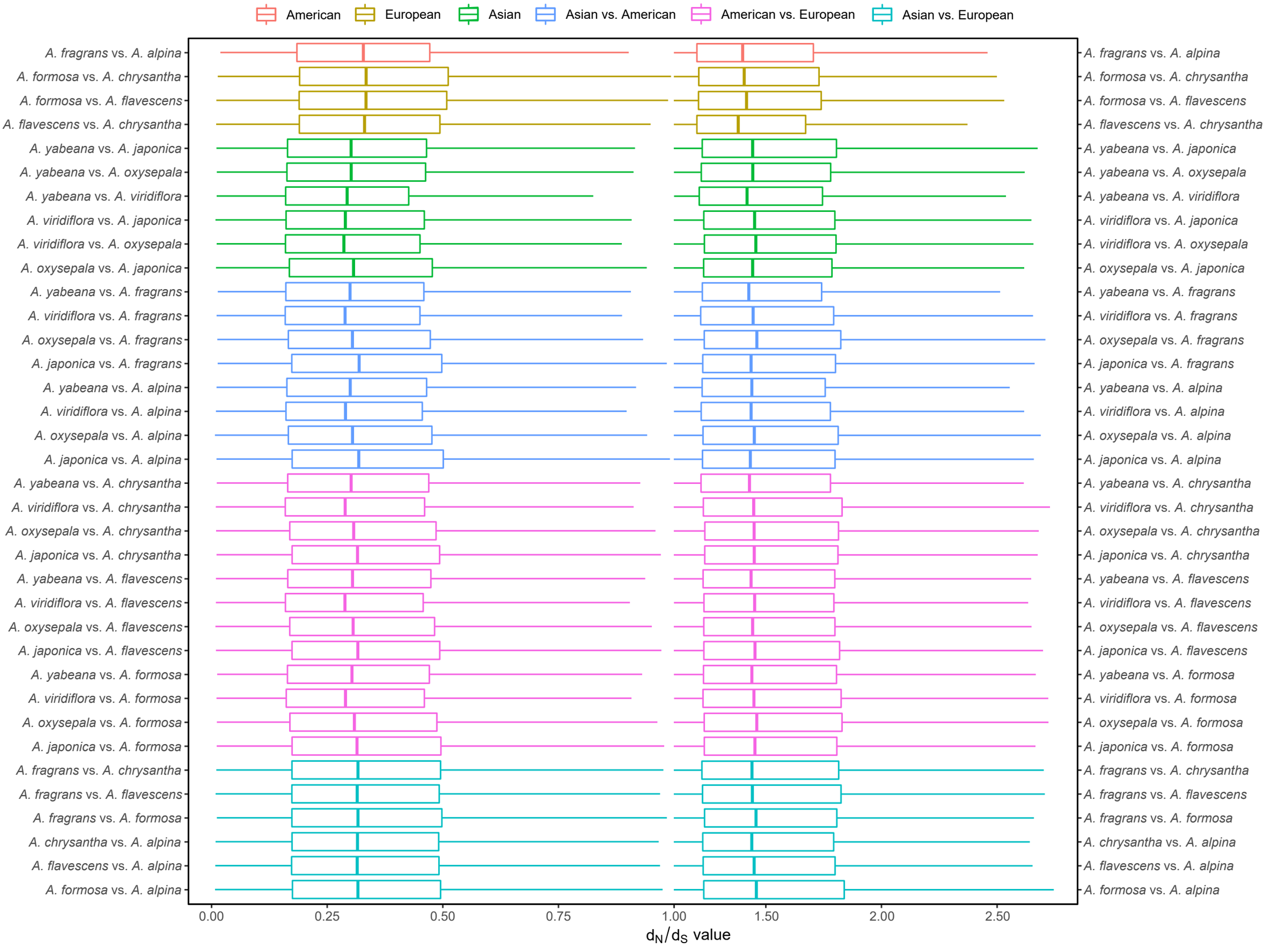

Chr7
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. coerulea
A. chrysantha
A. formosa
A. flavescens
0.020

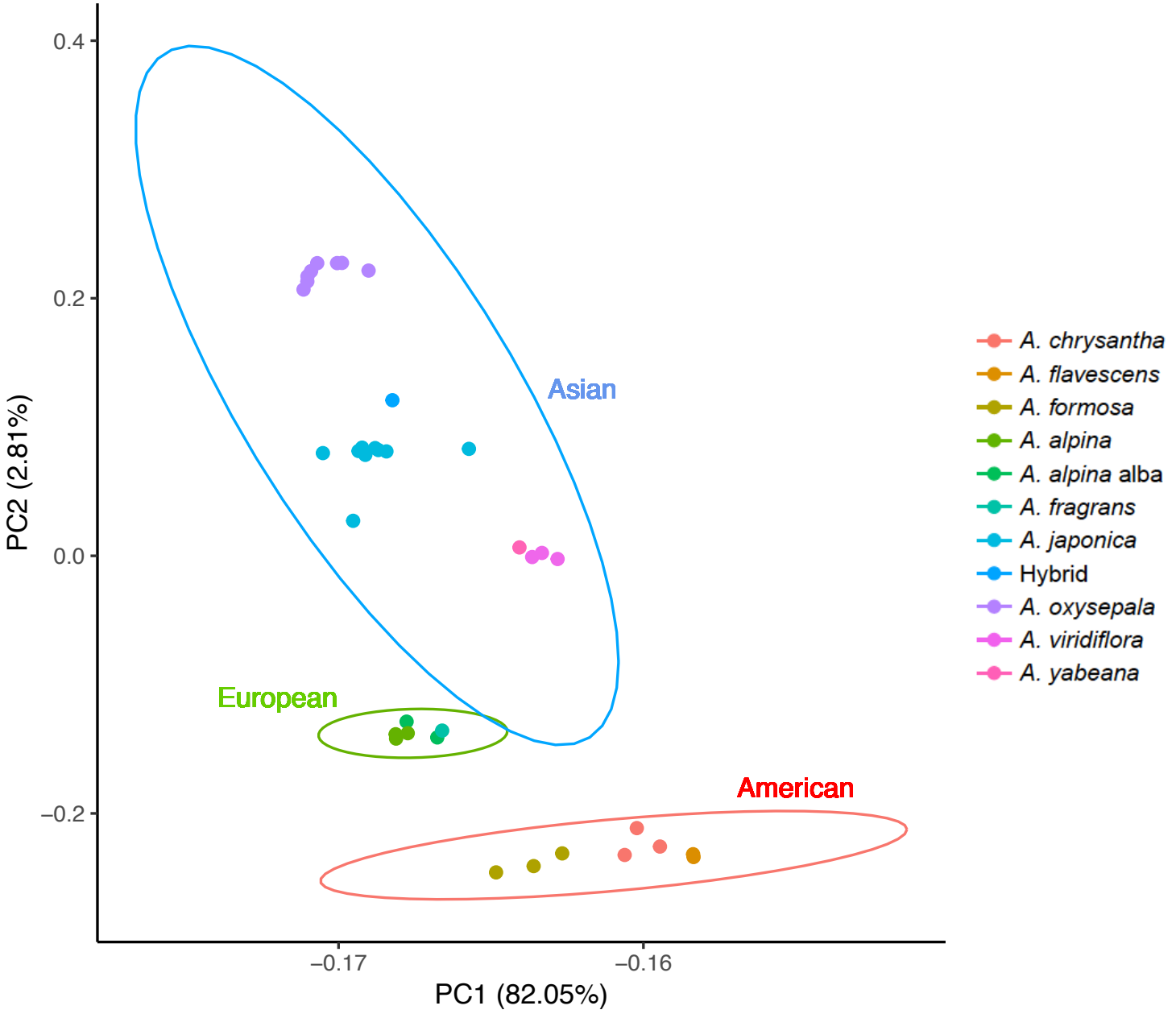

All
A. oxysepala
Hybrid
A. japonica
A. yabeana
A. viridiflora
A. alpina alba
A. fragrans
A. alpina
A. formosa
A. coerulea
A. chrysantha
A. flavescens
0.020


Whole genome
Chr1
Chr2
Chr3
Chr4
Chr5
Chr6
Chr7
Nucleotide diversity
American
European
Asian
A. japonica
A. oxysepala

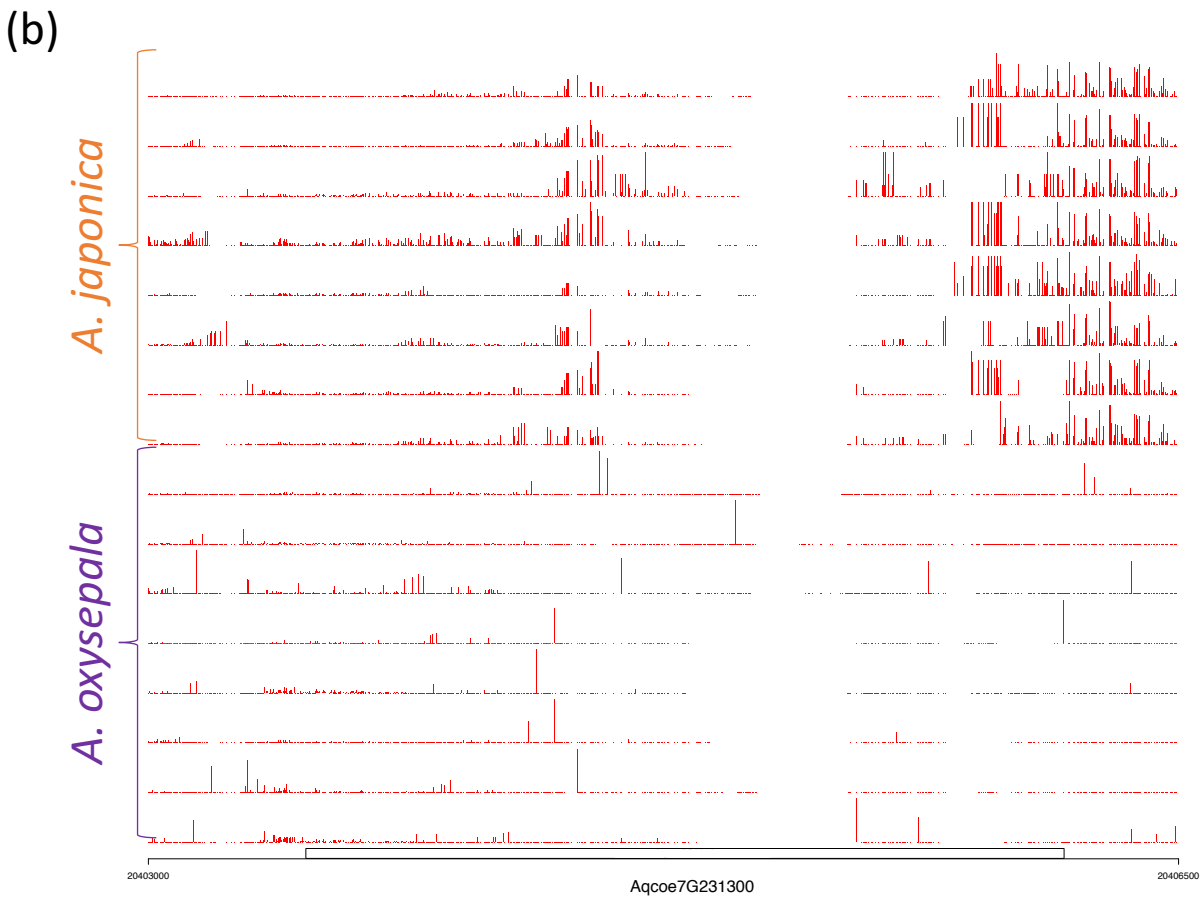

(a)
R = 0.46 , p < 2.2e−16
European lineage
Asian lineage
0.0000
0.0025
0.0050
0.0075
0.0100
0.0125
(b)
R = 0.42 , p < 2.2e−16
American lineage
Asian lineage
(c)
R = 0.52 , p < 2.2e−16
Amercian lineage
European lineage
(d)
R = 0.7 , p < 2.2e−16
Aquilegia japonica
Aquilegia oxysepala


European
Asian
American
49
7
37
87
5
17
39
Size of each list
148
74
0
148
148
148
European
Asian
American
Number of elements: specific (1) or shared by 2, 3, ... lists
87
29
125
3
2
1

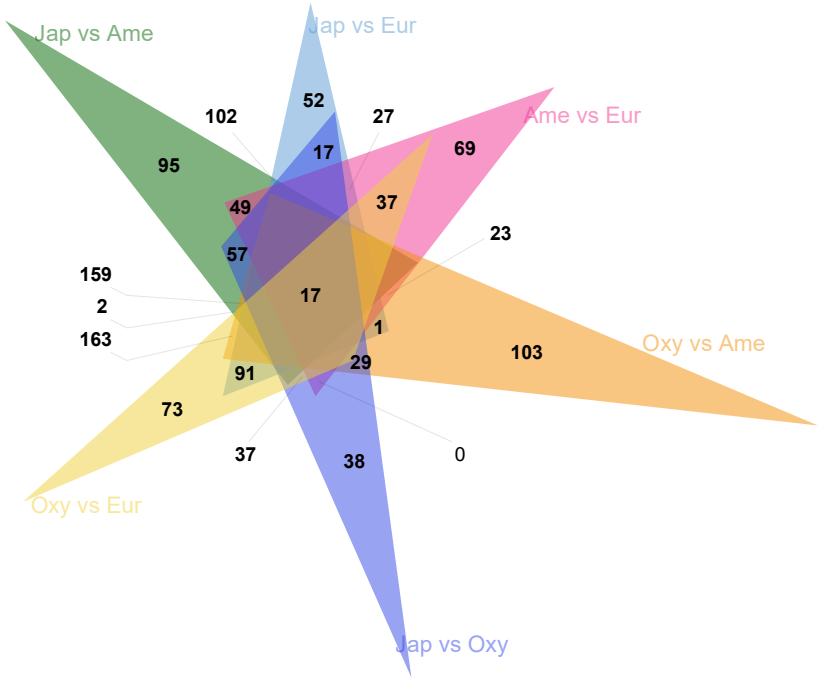

American
European
Asian
Asian vs. American
American vs. European
Asian vs. European
A. fragrans vs. A. alpina
A. formosa vs. A. chrysantha
A. formosa vs. A. flavescens
A. flavescens vs. A. chrysantha
A. yabeana vs. A. japonica
A. yabeana vs. A. oxysepala
A. yabeana vs. A. viridiflora
A. viridiflora vs. A. japonica
A. viridiflora vs. A. oxysepala
A. oxysepala vs. A. japonica
A. yabeana vs. A. fragrans
A. viridiflora vs. A. fragrans
A. oxysepala vs. A. fragrans
A. japonica vs. A. fragrans
A. yabeana vs. A. alpina
A. viridiflora vs. A. alpina
A. oxysepala vs. A. alpina
A. japonica vs. A. alpina
A. yabeana vs. A. chrysantha
A. viridiflora vs. A. chrysantha
A. oxysepala vs. A. chrysantha
A. japonica vs. A. chrysantha
A. yabeana vs. A. flavescens
A. viridiflora vs. A. flavescens
A. oxysepala vs. A. flavescens
A. japonica vs. A. flavescens
A. yabeana vs. A. formosa
A. viridiflora vs. A. formosa
A. oxysepala vs. A. formosa
A. japonica vs. A. formosa
A. fragrans vs. A. chrysantha
A. fragrans vs. A. flavescens
A. fragrans vs. A. formosa
A. chrysantha vs. A. alpina
A. flavescens vs. A. alpina
A. formosa vs. A. alpina
0.00
0.25
0.50
0.75
1.00
1.50
2.00
2.50
$d_N/d_S$ value

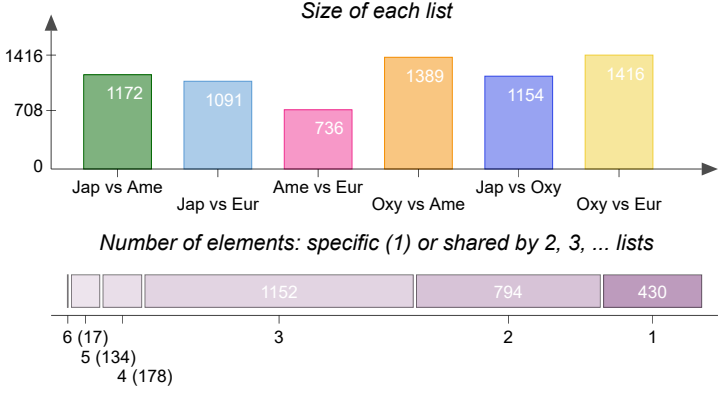

0.4
0.2
0.0
−0.2
PC2 (2.81%)
−0.17
−0.16
PC1 (82.05%)
Asian
European
American
A. chrysantha
A. flavescens
A. formosa
A. alpina
A. alpina alba
A. fragrans
A. japonica
Hybrid
A. oxysepala
A. viridiflora
A. yabeana

(a)
A. japonica
A. oxysepala
20392800
20394100
Aqcoe7G230600

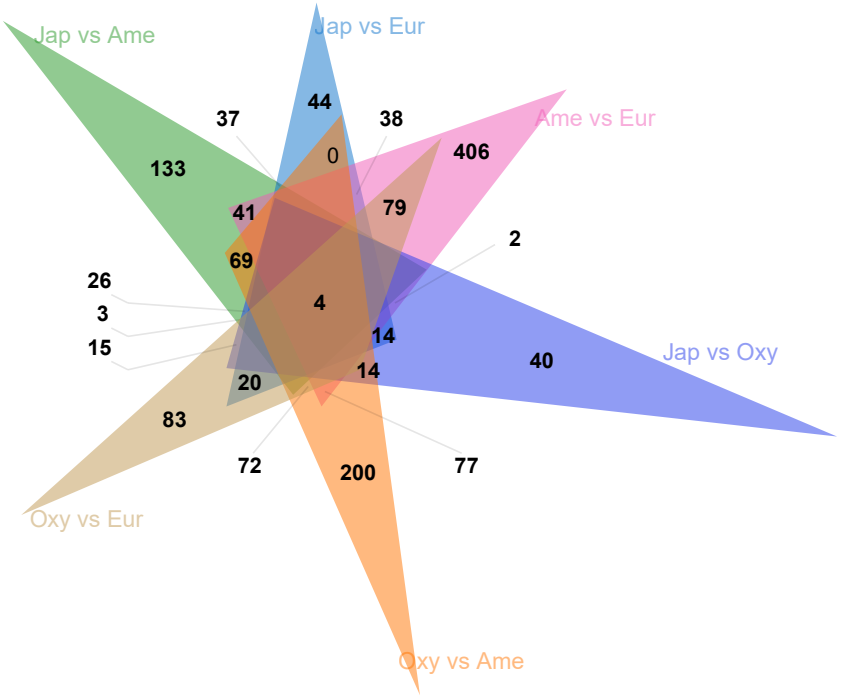

(b)
A. japonica
A. oxysepala
20403000
20406500
Aqcoe7G231300

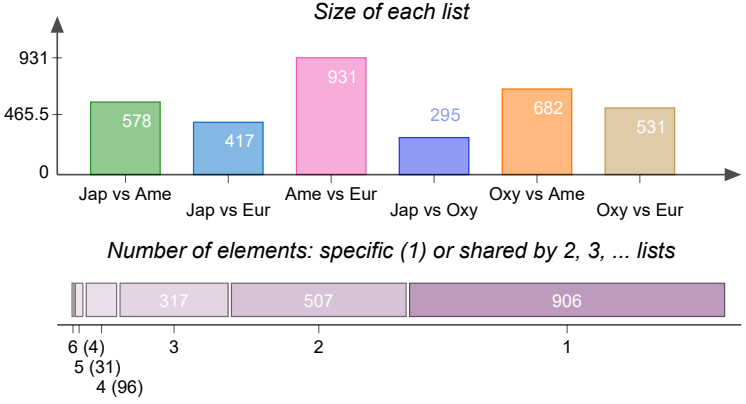
(a)


Jap vs Ame
Jap vs Eur
Ame vs Eur
Oxy vs Ame
Oxy vs Eur
Jap vs Oxy
102
52
27
95
17
69
49
37
23
57
159
17
2
1
163
103
91
29
73
37
38
0

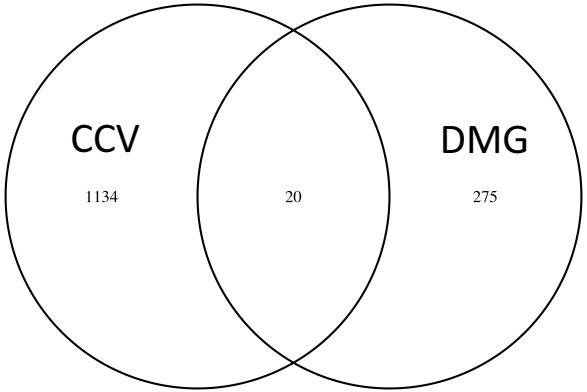

Size of each list
1416
708
0
1172
1091
736
1389
1154
1416
Jap vs Ame
Jap vs Eur
Ame vs Eur
Oxy vs Ame
Jap vs Oxy
Oxy vs Eur
Number of elements: specific (1) or shared by 2, 3, ... lists
1152
794
430
6 (17)
5 (134)
4 (178)
3
2
1

(b)

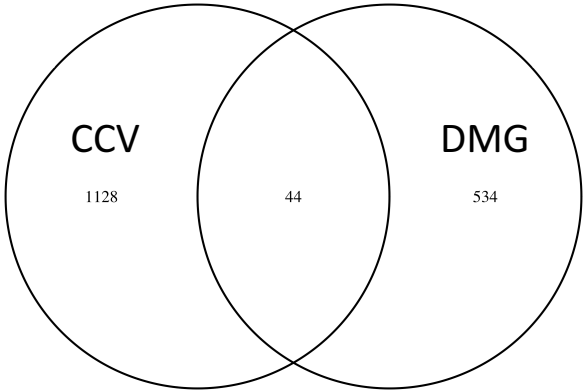

Jap vs Ame
Jap vs Eur
Ame vs Eur
Jap vs Oxy
Oxy vs Eur
Oxy vs Ame
37
44
38
133
0
406
41
79
2
69
26
4
3
14
15
40
14
20
83
72
200
77


Size of each list
931
465.5
0
578
417
931
295
682
531
Jap vs Ame
Jap vs Eur
Ame vs Eur
Jap vs Oxy
Oxy vs Ame
Oxy vs Eur
Number of elements: specific (1) or shared by 2, 3, ... lists
317
507
906
6 (4)
5 (31)
4 (96)
3
2
1

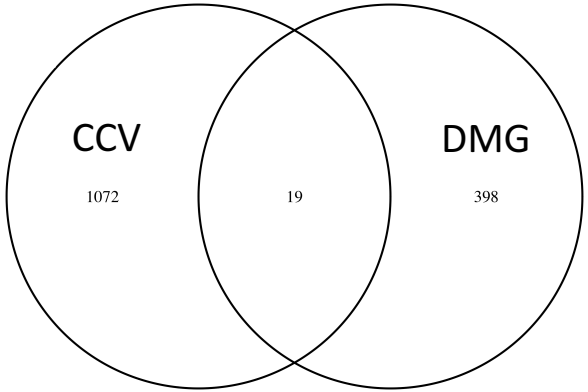
(a) *A. japonica* and *A. oxysepala*



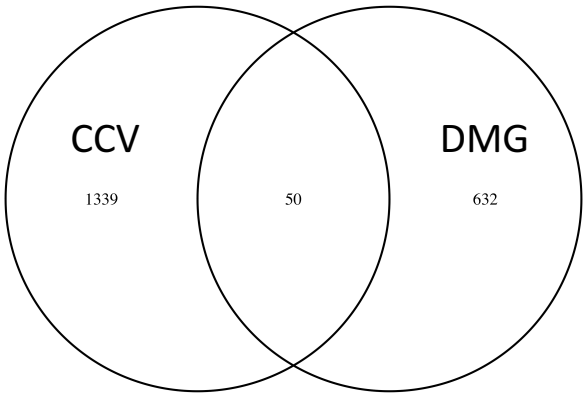

(b) *A. japonica* and North Amrican



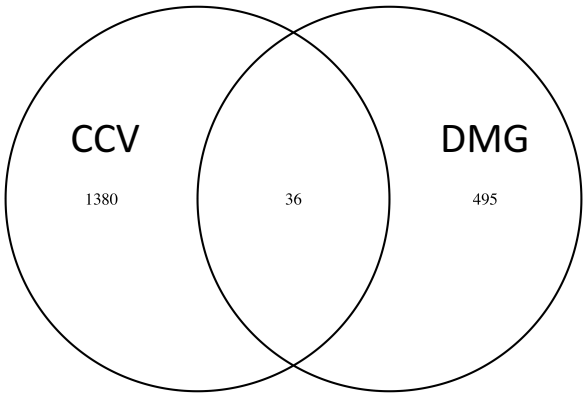

(c) *A. japonica* and European

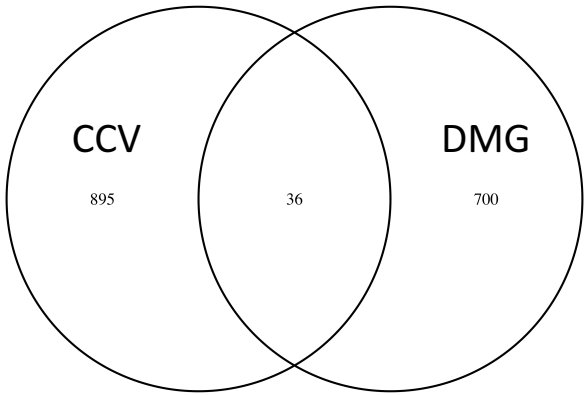



(d) *A. oxysepala* and North American




(e) *A. oxysepala* and European




(f) North American and European