Title: Reptile-like physiology in Early Jurassic stem-mammals

Authors: Elis Newham[1]*, Pamela G. Gill[2,3]*, Philippa Brewer[3], Michael J. Benton[2], Vincent Fernandez[4,5], Neil J. Gostling[6], David Haberthür[7], Jukka Jernvall[8], Tuomas Kankanpää[9], Aki Kallonen[10], Charles Navarro[2], Alexandra Pacureanu[5], Berit Zeller-Plumhoff[11], Kelly Richards[12], Kate Robson-Brown[13], Philipp Schneider[14], Heikki Suhonen[10], Paul Tafforeau[5], Katherine Williams[14], & Ian J. Corfe[8]*.

Affiliations:

[1]School of Physiology, Pharmacology & Neuroscience, University of Bristol, Bristol, UK.

[2]School of Earth Sciences, University of Bristol, Bristol, UK.

[3]Earth Science Department, The Natural History Museum, London, UK.

[4]Core Research Laboratories, The Natural History Museum, London, UK.

[5]European Synchrotron Radiation Facility, Grenoble, France.

[6]School of Biological Sciences, University of Southampton, Southampton, UK.

[7]Institute of Anatomy, University of Bern, Bern, Switzerland.

[8]Institute of Biotechnology, University of Helsinki, Helsinki, Finland.

[9]Department of Agricultural Sciences, University of Helsinki, Helsinki, Finland.

[10]Department of Physics, University of Helsinki, Helsinki, Finland.

[11]Helmholtz-Zentrum Geesthacht, Zentrum für Material-und Küstenforschung GmbH Germany.

[12]Oxford University Museum of Natural History, Oxford, OX1 3PW, UK.

[13]Department of Anthropology and Archaeology, University of Bristol, Bristol, UK.

[14]Faculty of Engineering and Physical Sciences, University of Southampton, Southampton, UK.

*e-mail: en12630@bristol.ac.uk; pam.gill@bristol.ac.uk; ian.corfe@helsinki.fi

5   **Abstract:** There is uncertainty regarding the timing and fossil species in which mammalian endothermy arose, with few studies of stem-mammals on key aspects of endothermy such as basal or maximum metabolic rates, or placing them in the context of living vertebrate metabolic ranges. Synchrotron X-ray imaging of incremental tooth cementum shows two Early Jurassic stem-mammals, *Morganucodon* and *Kuehneotherium*, had lifespans (a basal metabolic rate

10   proxy) considerably longer than comparably sized living mammals, but similar to reptiles, and that *Morganucodon* had femoral blood flow rates (a maximum metabolic rate proxy) intermediate between living mammals and reptiles. This shows maximum metabolic rates increased evolutionarily before basal rates, and that contrary to previous suggestions of a Triassic origin, Early Jurassic stem-mammals lacked the endothermic metabolism of living mammals.

One Sentence Summary: Surprisingly long lifespans and low femoral blood flow suggest reptile-like physiology in key Early Jurassic stem-mammals.

Main Text: Recent discoveries and analyses have revolutionized our knowledge of Mesozoic

20   mammals, revealing novel aspects of their ecology *(1, 2)* development *(3, 4)* systematics *(2, 4)* and macroevolution *(5, 6)*. However, details of physiology are more difficult to determine from fossils, and our knowledge of physiological evolution remains comparatively poor. Living mammals are endotherms, possessing the ability to control and maintain metabolically produced

heat, and have a substantially higher capacity for sustained aerobic activity than ectothermic animals *(7-9)*. The origin of endothermy is an important event in mammalian evolution, often noted as key to their success *(7-9)*. Several competing hypotheses seek to explain the selective pressures and adaptive pathways of endothermic evolution: (a) selection for higher maximum metabolic rates (MMR) enhanced sustained aerobic activity *(7, 10, 11)*; (b) selection for higher basal metabolic rates (BMR) enhanced thermoregulatory control *(12, 13)*; or (c) MMR and BMR evolved in lockstep with each other *(8, 9)*.

Direct evidence from living mammals to support these hypotheses is equivocal *(7)*. Recent analyses find no long-term evolutionary trend in BMR *(14)* contradicting earlier suggestions of increasing BMR throughout the Cenozoic *(13)*, and so implying that the Middle Jurassic (~170 Ma) most recent common ancestor (MRCA) of living mammals *(14)* possessed a BMR within the range of present-day mammals. Several indirect indicators of metabolic physiology in fossil synapsids have been suggested but provide contradictory evidence for the timing of origin of endothermy and its evolutionary tempo. These include: the presence of fibrolamellar long-bone, first seen in the Early Permian (~300 Ma) synapsid *Ophiacodon (15)*; the presence of an infraorbital canal and lack of parietal foramen, used to infer facial whiskers, fur, lactation and endothermy in Early Triassic (~245 Ma) cynodonts *(16)*; inferred maxillary nasal turbinates in the Late Permian (~255 Ma) therapsid *Glanosuchus*, used to suggest that mammalian levels of endothermy evolved by the Late Triassic (~210 Ma) *(17)*; a trend toward increased relative brain size initiated in Late Triassic non-mammaliaform cynodonts *(18)* and the mammaliaform stem-mammal *Morganucodon (19, 20)*; and acquisition of a parasagittal gait in the Early Cretaceous (~125 Ma) therian mammal *Eomaia (21)*. Several recent studies provide more quantitative links to physiological parameters. Oxygen isotopes were used to infer elevated

thermometabolism in Middle-Late Permian (~270-255 Ma) eucynodonts *(22)*; red blood cell size diminution in Late Permian (~255 Ma) eutheriodontid therapsids was qualitatively linked via two proxies to increased MMR *(23)*; and osteocyte lacuna shape correlations suggested 'mammalian' resting metabolic rates in Permo-Triassic (~250 Ma) dicynodonts *(24)*.

5 However, the inconsistency of these characters, in time and with respect to phylogeny *(25, 26)*, along with re-assessments of function in relation to endothermy *(8, 27)*, limit their use as conclusive indicators of modern mammalian levels of endothermy in fossil taxa. Such temporal and phylogenetic heterogeneity suggests the evolution of mammalian endothermy followed a complex, mosaic pattern with different physiological aspects evolving independently,

10 and at separate rates, towards current mammalian levels. Additionally, few of these physiological proxies are directly related to measurable aspects of metabolic rate.

To address these issues, we used two proxies to improve understanding of physiology at one of the most important nodes along this transition by estimating BMR and calculating a known proxy for MMR for two of the earliest mammaliaforms, *Morganucodon* and

15 *Kuehneotherium (1, 28)*.

Lifespan: a proxy for mammaliaform physiology

We used maximum lifespan estimates for fossil mammaliaform taxa as a proxy for BMR *(29)*. In extant tetrapods, negative correlations exist between lifespan and BMR *(29)* (and lifespan and

20 growth rate *(30)*, another indicator of metabolism - Supplementary Materials). In general, the longer a mammal's lifespan, the lower its size-adjusted BMR, so an accurate assessment of mammaliaform lifespan can be used to estimate metabolic potential. To do so we used cementochronology, which counts annual growth increments in tooth root cementum, and has

been widely used to record lifespans in extant mammals *(31, 32)*. Cementum is a mineralized dental tissue surrounding the tooth root (Fig 1A, B), attaching it to the periodontal ligament and anchoring the tooth within the alveolus *(31)*. Cementum growth is continuous throughout life in extant mammals and seasonally appositional in nature, forming a series of increments of differing thickness and opacity when viewed in histological thin-sections under light microscopy. The correlation between increment count and chronological age is well documented, with one thick and one thin increment deposited every year *(31)*, where the thin, hyper-mineralized opaque increments record growth rate reduction in less favourable seasons *(33)*.

Despite this potential, cementochronology has not previously been attempted for fossil mammals older than the Pleistocene (2.6 Ma) *(34)*, because histological thin-sections destroy fossils and provide only a restricted field-of-view. We overcame these problems by using propagation phase-contrast X-ray synchrotron radiation microtomography (PPC-SRμCT) to non-destructively image fossilized cementum increments *(35, 36)*. The sub-micrometre resolution, fast-throughput and three-dimensional (3D) nature of PPC-SRμCT allows large sample sizes and volumetric datasets of increments along their entire transverse and longitudinal trajectories. Cementum increments are known for lensing and coalescence, creating errors in counts based on single, or limited numbers of, two-dimensional thin sections created for each tooth *(31)* (Fig S1). PPC-SRμCT imaging and 3D segmentation of individual cementum increments across extensive vertical distances allows principal annual increments to be distinguished from accessory increments created by lensing and coalescence (Materials and Methods).

Morganucodon and *Kuehneotherium* are coexisting Early Jurassic (~200 Ma) shrew-sized insectivores1, known from thousands of teeth and bones (Materials and Methods), providing a rare opportunity to analyse the population-sized samples needed for confident

maximum lifespan estimation. Importantly, these are the earliest diphyodont taxa (Fig S2), with a single replacement of non-molar teeth and no molar tooth replacement *(28)*, and so estimates of lifespan are accurate to the time of the measured tooth root formation. We applied PPC-SRμCT to isolated teeth, and mandibles with multiple teeth or roots *in-situ*, totalling 87 *Morganucodon* specimens (52 isolated teeth, 35 dentaries), and 119 *Kuehneotherium* specimens (116 isolated teeth, 3 dentaries). From these, 34 *Morganucodon* and 27 *Kuehneotherium* specimens were sufficiently preserved for three observers to independently estimate lifespan from cementum increments and compare for accuracy and precision validation (Table S1). The remainder showed physical and/or diagenetic damage preventing increment counting (Fig S3).

The cementum of *Morganucodon* and *Kuehneotherium* (Fig 1A, B) is distinguished from dentine in our PPC-SRμCT data by a distinct boundary layer, which lies external to the dentine's granular layer of Tomes and is interpreted as the hyaline layer of Hopewell-Smith (Fig 1C-G). Synchrotron nanotomographic imaging (30 nm isotropic voxel size) of several exceptionally preserved specimens highlights individual Sharpey's fibre bundles (linking cementum to the periodontal ligament in extant mammals), extending radially through the cementum (Fig 1G). In transverse section, cementum is ~10-70 μm radially thick and displays contrasting light and dark circumferential increments representing different material densities (Figs 1E-G, 2A-D). Higher density increments (represented by greater greyscale values) average 2-3 μm radial thickness, and lower density increments 1-3 μm (Figs 1C-G, 2A-D). Individual increments can be followed continuously longitudinally and transversely through the entire tooth root (Fig 2E, F).

We tested the accuracy of cementum increment counts for predicting mammaliaform lifespan by additionally PPC-SRμCT imaging eight dentulous *Morganucodon* specimens with a range of teeth *in-situ*. We counted cementum increments for several teeth along the tooth rows,

and lines of arrested growth (LAGs) in the dentary bone periosteal region in two of these (Fig 3).

Comparisons between counts of cementum increments are identical across all four premolars

(p1-p4) and the anterior-most molars m1 and m2 in all specimens where they occur together

(Table S2). Counts are also identical between dentary LAGs and cementum increments in the

teeth present, p3-m2, of both specimens where LAGs were found (Fig 3). In three specimens

with m1-3, m3 has one less increment than m1/m2. In one specimen i4 and the canine have one

less increment than p1–3. This agreement between p1-m2 teeth and dentary increment counts

indicates growth in both teeth and jaws was following the same, circum-annual rhythm

previously reported for multiple extant mammal species *(31)*. We consider this strong support for

the accuracy of lifespan estimates based on these increment counts. Additionally, the differences

in increment counts along toothrows show eruption of the adult dentition in *Morganucodon*

occurred over more than one year, which is considerably slower than in extant small mammals

(31) (Supplementary text).

Long mammaliaform lifespans and low BMR

Cementum increment counts provide a minimum estimate of maximum lifespan of 14 years for

Morganucodon, and nine years for *Kuehneotherium* (Figs 2, 4A). These may underestimate true

maximum lifespan, as any damage to outer cementum increments would reduce estimated

maximum lifespan. One-way ANOVA comparisons of mean intra-observer coefficient of

variation (*CV*) between our study and ten previous cementochronological studies of different

extant mammal species (Table S3) with similar age ranges show values for PPC-SRµCT data of

Morganucodon (*CV* = 9.32) and *Kuehneotherium* (*CV* = 4.89) are significantly lower (*p* < 0.01)

than previous thin section-based studies (minimum extant *CV* = 14.2, mean *CV* = 21.8).

We estimated a body mass of 17.9 g for *Morganucodon* and 23.8 g for *Kuehneotherium* (Materials and Methods). Maximum lifespan and mean body mass for the mammaliaforms were compared with published data for large samples of terrestrial, non-volant wild extant mammal (n = 279) and non-avian reptile (n = 252) species (Table S4). Ordinary least squares (OLS) regression of $\log_{10}$ transformed values shows the fossil mammaliaforms fall within the range of extant reptiles and have longer maximum lifespans for their size, and are further above the mammal regression mean, than all extant mammals under 4 kg (the long-lived and secondarily dwarfed *(37)* mouse lemur *Microcebus murinus* is closest). Only the short-beaked echidna *Tachyglossus aculeatus*, a monotreme with long lifespan and low metabolic rate, exceeds the *Kuehneotherium*, but not *Morganucodon*, distance above the mammalian mean (Fig 4B). One-way ANCOVA comparisons show that regression slopes for extant mammals and reptiles are statistically similar but their means are significantly separated (*p* <0.01), with reptiles on average living 12.2 years longer than mammals of the same body mass.

To estimate BMR, we used OLS and recovered significant correlations between log10 transformed values of wild lifespan and mass-specific standard metabolic rate (msSMR; measured in mL.O2.hr-1.g-1 and analogous with BMR in extant mammals – SMR was used as BMR cannot be measured in reptiles *(38)*) from published data for 117 extant mammals and 55 extant reptiles (Table S1; Fig S4A). Using the correlation between mammal lifespan and msSMR we estimated a msSMR of 0.38 mL.O2.hr-1.g-1 for *Morganucodon* and 0.51 mL.O2.hr-1.g-1 for *Kuehneotherium.* From the correlation between reptile lifespan and msSMR we estimated a msSMR of 0.10 mL.O2.hr-1.g-1 (*Morganucodon*) and 0.15 mL.O2.hr-1.g-1 (*Kuehneotherium*). When log10 OLS is used to regress these estimates against body mass, both mammaliaforms fall outside the 95% predictor interval (PI) of the mammalian data and within the reptile range of

msSMR, whether estimated from mammalian or reptilian data (Fig 4C). This suggests the mammaliaforms had significantly lower msSMR values compared to extant mammals of similar size. The comparably sized mammal (<100 g) of lowest msSMR is the marsupial *Dasycercus cristicauda*, with wild lifespan seven years and msSMR 0.63 mL.O2.hr-1.g-1 (Fig 4C).

In summary, our estimates of maximum lifespan provided by tomographic imaging of cementum increments in *Morganucodon* and *Kuehneotherium* are significantly longer than the wild lifespan of any extant mammal of comparable body mass. These lifespans provide SMR/BMR estimates considerably lower than comparably sized extant mammals, instead corresponding to those of extant reptiles.

Femoral blood-flow shows intermediate MMR

To compare our fossil mammaliaform BMR estimates with MMR, we used a second proxy directly linked to MMR *(39)*. The ratio between nutrient foramen area and femur length has been used as an index for relative blood flow (Qi) through the femur during and after metabolically demanding exercise ($Qi = rf4/L$; where rf = foramen radius and L = femur length), previously shown to correlate well with MMR *(39)*. From micro-computed tomography (μCT) data of the six most complete *Morganucodon* femoral diaphyses available, we segmented all nutrient foramina (Fig 5A) and estimated their area by measuring their minimal radii (Materials and Methods). *Kuehneotherium* could not be included as no suitable femoral specimens are known.

We estimated a Qi of 3.829e-7 mm3 for *Morganucodon* and compared this with published and new data (Materials and Methods) for extant mammals (n = 69) and reptiles (n = 30). The latter includes varanids (n = 8) which in the absence of mammalian predators fill an active hunting niche and tend to have mammalian MMR levels while retaining reptilian BMR

levels *(39)* (Table S4). One-way ANCOVA comparisons show that means of log10 OLS regression slopes of body mass against Qi for extant mammals and non-varanid reptiles are significantly different (p <0.01), while the slopes are similar (Materials and Methods). *Morganucodon* is further above (higher Qi for its mass) the non-varanid reptile mean than all non-varanid reptiles. However, *Morganucodon* is also slightly further from the mammalian mean than the non-varanid reptile mean, and considerably closer to small non-varanid reptile species than small mammalian species (Fig 5B). This intermediate Qi, and so inferred intermediate MMR, suggests that while retaining typical reptilian BMR, *Morganucodon* had an MMR above non-varanid reptiles, but not as high as mammals or actively foraging varanid reptiles.

Discussion

We have used two quantitative proxies to determine the metabolic status of early mammaliaforms. Relatively long lifespans for both *Morganucodon* and *Kuehneotherium* result in BMR estimates equivalent to modern reptiles of comparable size, and indeed at the higher lifespan/lower BMR end of the reptile scale for *Morganucodon*. In contrast, femoral blood flow estimates (Q_i) suggest that the MMR of *Morganucodon* was intermediate between extant non-varanid reptiles and mammals. We therefore infer that in *Morganucodon* increased MMR (and so also absolute aerobic capacity, AAC = MMR - BMR) was initially selected for before BMR, and that the MMR-first hypothesis *(7)* is the best-supported model for the evolution of mammalian endothermy. We suggest that at least *Morganucodon*, if not also *Kuehneotherium*, occupied a metabolic grade approaching extant varanids; able to undergo longer bouts of aerobically demanding activity than non-varanid reptiles, but not capable of sustaining either mammalian levels of aerobic activity or the elevated thermometabolism exhibited by living endotherms.

Evidence from non-mammalian synapsids (including changes in gait *(8)*, long bone histology *(15)*, and development of secondary osteological features correlated with increased metabolic rate *(18, 19)*) indicate unquestionable changes in physiology from pelycosaur- to mammaliaform-grade taxa. Determinate growth *(40)* and reduction of dental replacement (diphyodonty) in basal mammaliaforms permitted more precise occlusion, which has been considered a key innovation in the development of mammalian endothermy through enabling increased assimilation and higher metabolism *(41)*. However, determinate growth and diphyodonty appear to have preceded the appearance of modern mammalian levels of endothermy, at least in *Morganucodon* and *Kuehneotherium*. We therefore suggest that the development of precise occlusion in basal mammaliaforms may be more associated with dietary specialization and niche partitioning *(1)*.

Comparison of our results to other recent studies of physiology in fossil synapsids supports the hypothesis of a complex, mosaic pattern of endothermic evolution with different characters being selected for at different rates through time and with respect to phylogeny. For example, the size diminution associated with the cynodont-mammaliaform transition *(42)* may have reversed the evolutionary trajectory of some previous histological proxies for endothermy *(43)*, contributing to the complex, contradictory patterns observed. Our study also suggests more work is needed to directly compare fossil and extant ecto- and endothermic taxa, to better understand their relative metabolic properties. Many previous studies rely on simple binary divisions, such as the presence/absence of fibrolamellar bone and/or respiratory nasal turbinates. These proxies cannot represent accurately the complex series of physiological characteristics that range between ectothermy and endothermy, and are frequently distributed homoplastically across the synapsid phylogeny, individually and with respect to each other. Other studies with relative

data such as preserved apatite oxygen isotopes *(22)* allow comparisons with co-habiting ectothermic taxa but cannot be directly compared to extant data and cannot indicate where fossil taxa fall in the metabolic spectrum of extant vertebrates. However, our results are compatible with recent work on living mammals that the BMR of the Middle Jurassic (~170 Ma)

5   mammalian MRCA was comparable to present-day values *(14)*. This indicates evolution towards modern-day mammalian endothermy occurred during the 25 million year-long Early Jurassic and suggests the mammalian mid-Jurassic adaptive radiation *(5, 6)* was driven by this, or vice versa.

In conclusion, our data offer a direct link to measurable aspects of endothermy, BMR and MMR, at a key point in mammalian evolution. Further work applying these methods to younger

10 Mesozoic mammaliaforms and mammals, and comparison with evidence from other physiological characteristics, will allow the evolutionary tempo and mode of multiple aspects of mammalian physiology to be determined. The early mammaliaforms *Morganucodon* and *Kuehneotherium* possessed surprisingly low, reptile-like metabolic rates, plus a mixture of plesiomorphic and derived characters *(7)* relating to life history and physiology. Ultimately, we

15 can no longer assume that the endothermic metabolism of living mammals had evolved in the earliest mammaliaforms.

References and notes:

1. P. G. Gill, M. A. Purnell, N. Crumpton, K. Robson Brown, N. J. Gostling, M.

20   Stampanoni, E. J. Rayfield, Dietary specializations and diversity in feeding ecology of the earliest stem mammals. *Nature* **512**, 303-305 (2014).

2. Z. X. Luo, Transformation and diversification in early mammal evolution. *Nature* **450**, 1011-1019 (2007).

3. C. F. Zhou, B. A. S Bhullar, A. I Neander, T Martin, Z. X. Luo. New Jurassic mammaliaform sheds light on early evolution of mammal-like hyoid bones. *Science* **365,** 276-279. (2019).

4. G. Han, F. Mao, S. Bi, Y. Wang, J. A. Meng, Jurassic gliding euharamiyidan mammal with an ear of five auditory bones. *Nature* **551**, 451-456 (2017).

5. G. P. Wilson, A. R. Evans, I. J. Corfe, P.D. Smits, M. Fortelius, J. Jernvall, Adaptive radiation of multituberculate mammals before the extinction of dinosaurs. *Nature* **483**, 457-460 (2012).

6. R. A. Close, M. Friedman, G. T. Lloyd R. B. Benson, Evidence for a mid-Jurassic adaptive radiation in mammals. *Curr. Biol.* **25**, 2137-2142 (2015).

7. A. F. Bennett, J. A. Ruben, Endothermy and activity in vertebrates. *Science* **206**, 649-654 (1979).

8. T. S. Kemp, The origin of mammalian endothermy: a paradigm for the evolution of complex biological structure. *Zool. J. Linn. Soc.* **147**, 473-488 (2006).

9. A. Clarke, A. H. O. Pörtner, Temperature, metabolic power and the evolution of endothermy. *Biol. Rev.* **85**, 703-727 (2010).

10. P. Koteja, Energy assimilation, parental care and the evolution of endothermy. *Proc. Royal Soc. B.* **267**, 479-484 (2000).

11. J. A. Hopson, The role of foraging mode in the origin of therapsids: implications for the origin of mammalian endothermy. *Fieldiana Life Earth Sci.* **49**, 126-149 (2012).

12. C. G. Farmer, Reproduction: the adaptive significance of endothermy. *Am. Nat.* **162**, 826-840 (2003).

13. B. G. Lovegrove, The evolution of endothermy in Cenozoic mammals: a plesiomorphic-apomorphic continuum. *Biol. Rev.* **87**, 128-162 (2012).

14. J. Avaria-Llautureo, C. E. Hernández, E. Rodríguez-Serrano, C. Venditti. The decoupled nature of basal metabolic rate and body temperature in endotherm evolution. *Nature*, **572**, 651-654 (2019).

15. C. D. Shelton, P. M. Sander, Long bone histology of *Ophiacodon* reveals the geologically earliest occurrence of fibrolamellar bone in the mammalian stem lineage. *C. R. Palevol.* **16**, 397-424 (2017).

16. J. Benoit, P. R. Manger, B. S. Rubidge, Palaeoneurological clues to the evolution of defining mammalian soft tissue traits. *Sci. Rep.* **6**, 25604 (2016).

17. W. J. Hillenius, Turbinates in therapsids: evidence for Late Permian origins of mammalian endothermy. *Evolution* **48**, 207-229 (1994).

18. J. Benoit, V. Fernandez, P. R. Manger, B. S. Rubidge, Endocranial casts of pre-mammalian therapsids reveal an unexpected neurological diversity at the deep evolutionary root of mammals. *Brain Behav. Evolut.* **90**, 311-333 (2017).

19. T. B. Rowe, T. E. Macrini, Z. X. Luo, Z. X. Fossil evidence on origin of the mammalian brain. *Science* **332**, 955-957 (2011).

20. E. A. Hoffman, T. B. Rowe, Jurassic stem-mammal perinates and the origin of mammalian reproduction and growth. *Nature* **561**, 104-108 (2018).

21. Z. Kielan-Jaworowska, J. H. Hurum, Limb posture in early mammals: sprawling or parasagittal. *Acta Palaeontol. Pol.* **51**, 393-406 (2006).

22. K. Rey, R. Amiot, F. Fourel, F. Abdala, F. Fluteau, N-E. Jalil, J. Liu, B. S. Rubidge, R. M.H. Smith, J. S. Steyer, P. A. Viglietti, X. Wang, C. Lécuyer, Oxygen isotopes suggest

elevated thermometabolism within multiple Permo-Triassic therapsid clades. *Elife* **6**, e28589 (2017).

23. A. K. Huttenlocker, C. G. Farmer, Bone microvasculature tracks red blood cell size diminution in Triassic mammal and dinosaur forerunners. *Curr. Biol*. **27**, 48-54 (2017).

24. C. Olivier, A. Houssaye, N. E. Jalil, J. Cubo, First palaeohistological inference of resting metabolic rate in an extinct synapsid, *Moghreberia nmachouensis* (Therapsida: Anomodontia). *Biol. J. Linn. Soc*. **121**, 409-419 (2017).

25. A. Crompton, T. Owerkowicz, B. A. Bhullar, C. Musinsky, Structure of the nasal region of non-mammalian cynodonts and mammaliaforms: speculations on the evolution of mammalian endothermy. *J. Vertebrate Paleontol*. **37**, e1269116 (2017).

26. P. G. Rodrigues, A. G. Martinelli, C. L. Schultz, I. J. Corfe, P. G. Gill, M. B. Soares, E. J. Rayfield, Digital cranial endocast of *Riograndia guaibensis* (Late Triassic, Brazil) sheds light on the evolution of the brain in non-mammalian cynodonts. *Hist. Biol*. 1-18 (2018).

27. J. P. Hayes, T. Garland Jr. The evolution of endothermy: testing the aerobic capacity model. *Evolution* **49**, 836-847 (1995).

28. Z. Kielan-Jaworowska, R. L. Cifelli, Z. X. Luo, *Mammals from the age of dinosaurs: origins, evolution, and structure* (Columbia University Press, New York, United States, 2004).

29. A. J. Hulbert, R. Pamplona, R. Buffenstein, W. A. Buttemer, Life and death: metabolic rate, membrane composition, and life span of animals. *Physiol. Rev*. **87**, 1175-1213 (2007).

30. J. M. Grady, B. J. Enquist, E. Dettweiler-Robinson, N. A. Wright, F. A. Smith, Evidence for mesothermy in dinosaurs. *Science* **344**, 1268-1272 (2014).

31. G. Klevezal, *Recording structures of mammals* (CRC Press. Boca Raton, Florida, US, 1995).

32. S. Naji, T. Colard, J. Blondiaux, B. Bertrand, E. d'Incau, J. P. Bocquet-Appel. Cementochronology, to cut or not to cut? *Int. J. Paleopathol.* **15**, 113-119 (2016).

33. D. E. Lieberman, Life history variables preserved in dental cementum microstructure. *Science* **261**, 1162-1164 (1993).

34. A. J. Stutz, Polarizing microscopy identification of chemical diagenesis in archaeological cementum. *J. Archaeol. Sci*. **29**, 1327-1347 (2002).

35. E. Newham, thesis, University of Southampton (2018).

36. A. Le Cabec, N. K. Tang, V. Ruano Rubio, S. Hillson, Nondestructive adult age at death estimation: Visualizing cementum annulations in a known age historical human assemblage using synchrotron X-ray microtomography. *Am. J. Phys. Anthropol*. **168**, 25-44 (2019).

37. A. D. Yoder, C. R. Campbell, M. B. Blanco, M. Dos Reis, J. U. Ganzhorn, S. M. Goodman, K. E. Hunnicutt, P. A. Larsen, P. M. Kappeler, R. M. Rasoloarison, J. M. Ralison, D. L. Swofford, D. W. Weisrock. Geogenetic patterns in mouse lemurs (genus *Microcebus*) reveal the ghosts of Madagascar's forests past. *PNAS* **113**, 8049-8056 (2016).

38. C. R. White, N. F. Phillips, N. F. R. S. Seymour, The scaling and temperature dependence of vertebrate metabolism. *Biol. Letters* **2**, 125-127 (2005).

39. R. S. Seymour, S. L. Smith, C. R. White, D. M. Henderson, D. Schwarz-Wings, Blood flow to long bones indicates activity metabolism in mammals, reptiles and dinosaurs. *Proc. Royal Soc. B.* **279**, 451-456 (2011).

40. R. N. O'Meara, R. J. Asher, The evolution of growth patterns in mammalian versus nonmammalian cynodonts. *Paleobiology* **42**, 439-464 (2016).

41. T. S. Kemp, *The origin and evolution of mammals.* (Oxford University Press. Oxford, UK, 2005).

42. S. Lautenschlager, P. G. Gill, Z. X. Luo, M. J. Fagan, E. J. Rayfield, The role of miniaturization in the evolution of the mammalian jaw and middle ear. *Nature* **561**, 533-537 (2018).

43. J. Botha-Brink, D. Codron, A. K. Huttenlocker, K. D. Angielczyk, M. Ruta, Breeding young as a survival strategy during Earth's greatest mass extinction. *Sci. Rep.* **6**, 24053 (2016).

44. K. A. Kermack, F. Mussett, H. W. Rigney, The lower jaw of *Morganucodon*. *Zool. J. Linn. Soc.* **53**, 87–175 (1973).

45. Whiteside, D. I., Duffin, C. J., Gill, P. G., Marshall, J. E., & Benton, M. J. The Late Triassic and Early Jurassic fissure faunas from Bristol and South Wales: stratigraphy and setting. *Palaeontol. Pol.* **67**, 257–287 (2016).

46. D. M Kermack, K. A. Kermack, F. Mussett, The Welsh pantothere *Kuehneotherium praecursoris. Zool. J. Linn. Soc.* **47**, 407–423 (1968).

47. P. G. Gill, thesis, University of Bristol (2004).

48. D. Paganin, S. C. Mayo, T. E. Gureyev, P. R. Miller, S. W. Wilkins, Simultaneous phase and amplitude extraction from a single defocused image of a homogeneous object. *J. Microscopy* **206**, 33-40 (2002).

49. F. Marone, M. Stampanoni, Regridding reconstruction algorithm for real-time tomographic imaging. *J. Synchrotron Radiation* **19**, 1029-1037 (2012).

50. R. Mokso, P. Cloetens, Nanoscale zoom tomography with hard x rays using Kirkpatrick-Baez optics. *Appl. Phys. Lett.* **90**, 144104 (2007).

51. C. A. Schneider, W. S. Rasband, K. W. Eliceiri, NIH Image to ImageJ: 25 years of image analysis. *Nature Methods,* **9**, 671–675 (2001).

52. J. R. Foster, Preliminary body mass estimates for mammalian genera of the Morrison Formation (Upper Jurassic, North America). *PaleoBios* **28**, 114–122 (2009).

53. Z. X. Luo, A. W. Crompton, A. L. Sun, A new mammaliaform from the Early Jurassic and evolution of mammalian characters. *Science* **292**, 1535–1540 (2001).

54. Q. Ji, Z. X. Luo, C. X. Yuan, A. R. Tabrum, A swimming mammaliaform from the Middle Jurassic and ecomorphological diversification of early mammals. *Science* **311**, 1123–1127 (2006).

55. J. A. Mclean, J. R. Speakman, Effects of body mass and reproduction on the basal metabolic rate of brown long–eared bats (*Plecotus auratus*). *Physiol. Biochem. Zool.* **73**, 112–121 (2000).

56. A. B. Salmon, S. Leonard, V. Masamsetti, A. Pierce, A. J. Podlutsky, N. Podlutskaya, A. Richardson, S. N. Austad, A. R. Chaudhuri, The long lifespan of two bat species is correlated with resistance to protein and enhanced protein homeostasis. *FASAB J.* **23**, 2317–2326 (2009).

57. N. I. Becker, J. A. Encarnacao, M. Tschapka, E. K. V. Kalko, Energetics and life–history of bats in comparison to small mammals. *Ecol. Res.* **28**, 249–258 (2013).

58. J. Munshi–South, G. S. Wilkinson, G. S. Bats and birds: exceptional longevity despite high metabolic rates. *Ageing Res. Rev.* **9**, 12–19 (2010).

59. N. W. Kasting, S. A. Adderley, T. Safford, K. G. Hewlett, Thermoregulation in beluga (*Delphinapterus leucas*) and killer (*Orcinus orca*) whales. *Physiol. Zool.* **62**, 687–701 (1989).

60. R. M. Ortiz, D. P. Noren, B. Litz, C. L. A. Ortiz, A new perspective on adiposity in a naturally obese mammal. *Am. J. Physiol.–Endocrinol. Metab.* **281**, E1347–E1351 (2001).

61. M. Chikina, J. D. Robinson, N. L. Clark, Hundreds of genes experienced convergent shifts in selective pressure in marine mammals. *Mol. Biol. Evol.* **33**, 2182–1292 (2016)

62. I. Scharf, A. Feldman, M. Novosolov, D. Pincheira-Donoso, I. Das, M Böhm, P. Uetz, O. Torres-Carvajal, A. Bauer, U. Roll, S. Meiri, Late bloomers and baby boomers: ecological drivers of longevity in squamates and the tuatara. *Global Ecol. Biogeog.* **24** 396–405 (2015).

63. F. A. Smith, S. K. Lyons, S. K. Morgan Ernest, K. E. Jones, D. M. Kaufman, T. Dayan, P. A. Marquet, J. H. Brown, J. P. Haskell, Body mass of late Quaternary mammals. *Ecol. Arch.* **E084** (2003).

64. J. P. D. Magalhães, J. Costa, G. M. Church, An analysis of the relationship between metabolism, developmental schedules, and longevity using phylogenetic independent contrasts. *J. Gerontol. Ser. A: Biol. Sci. Med. Sci.* **62**, 149–160 (2007).

65. J. Werner, E. M. Griebeler, Allometries of maximum growth rate versus body mass at maximum growth indicate that non–avian dinosaurs had growth rates typical of fast growing ectothermic sauropsids. *PLOS One* **9**, e88834 (2014).

66. T. J. Case, On the evolution and adaptive significance of postnatal growth rates in the terrestrial vertebrates. *Q. Rev. Biol*. **53**, 243-282 (1978).

67. E. M. Zullinger, R. E. Ricklefs, K. H. Redford, G. M. Mace, Fitting sigmoidal equations to mammalian growth curves. *J. Mammalogy* **65**, 607-636.

68. F. A. Jenkins, F. R. Parrington, The postcranial skeletons of the Triassic mammals *Eozostrodon*, *Megazostrodon* and *Erythrotherium*. *Phil. Trans. R. Soc. Lond. B*. **273,** 387–431 (1976).

69. Z. X. Luo, Z. Kielan-Jaworowska, R. L. Cifelli, In quest for a phylogeny of Mesozoic mammals. *Acta Palaeontol. Pol.* **47**, 1-78 (2002).

70. J. A. Schultz, B. A. S. Bhullar, Z. X. Luo, Re-examination of the Jurassic Mammaliaform *Docodon victor* by Computed Tomography and Occlusal Functional Analysis. *J. Mamm. Evol.*. **26,** 9-38. (2019).

71. T. Martin, Tooth replacement in Late Jurassic Dryolestidae (Eupantotheria, Mammalia). *J. Mamm. Evol.* **4**, 1–18. (1997).

72. Z. X. Luo, Z. Kielan-Jaworowska, R. Cifelli, Evolution of dental replacement in mammals. *Bull. Carnegie Mus. Nat. Hist.* **36**, 159-176 (2004).

73. X. Zheng, S. Bi, X. Wang, J. Meng, A new arboreal haramiyid shows the diversity of crown mammals in the Jurassic period. *Nature* **500**, 199-202.

74. S. Ray, J. Botha, A. Chinsamy, Bone histology and growth patterns of some nonmammalian therapsids. *J. Vert. Paleontol.* **24**, 634-648 (2004).

75. G. A. Grau, G. C. Sanderson, J. Rogers, Age determination of raccoons. *The J. Wildlife Management* 34, 364-372 (1970).

76. W. C. Gasawey, D. B. Harkness, R. A. Rausch, Accuracy of moose age determinations form incisor cementum layers. *J. Wildlife Management* **42**, 558-563 (1978).

77. G. A. Klevezal, Z. Pucek, Growth layers in tooth cementum and dentine of European bison and its hybrids with domestic cattle. *Acta Theriol*. **32**, 115-128 (1987).

78. R. F. Kay, J. G. Cant, Age assessment using cementum annulus counts and tooth wear in a free-ranging population of *Macaca mulatta*. *Am. J. Primatology* **15**, 1-15 (1988).

79. G. Cederlund, P. Kjellander, F. Stålfelt, Age determination of roe deer by tooth wear and cementum layers-tests with known age material. In Csányi, S. & Ernhaft, J. (Eds). *Transactions of the 20th Congress of the International Union of Game Biologists, Gödöllö* (540-545). University of Agricultural Sciences. Gödöllö, Hungary (1991).

80. J. L. Bodkin, J. A. Ames, R. J. Jameson, A. M. Johnson, G. M. Matson, Estimating age of sea otters with cementum layers in the first premolar. *J. wildlife management* **61**, 967-973 (1997).

81. D. B. Landon, C. A. Waite, R. O. Peterson, L. D. Mech, Evaluation of age determination techniques for gray wolves. *J. wildlife management* **62**, 674-682 (1998).

82. S. N. Christensen-Dalsgaard, J. Aars, M. Anderson, C. Lockyer, N. G. Yoccoz, Accuracy and precision in estimation of age of Norwegian Arctic polar bears (*Ursus maritimus*) using dental cementum layers from known-age individuals. *Polar Biology* **33**, 589-597 (2010).

83. K. Pasda, Assessment of age and season of death of West Greenland reindeer by counting cementum increments in molars. *Documenta Archaeobiologiae* **4**, 125-140 (2006).

84. F. J. Pérez-Barbería, E. I. Duff, M. J. Brewer, F. E. Guinness, Evaluation of methods to age Scottish red deer: the balance between accuracy and practicality. *J. Zool*. **294**, 180-189 (2014).

85. https://genomics.senescence.info/species/

86. https://animaldiversity.org

87. J. M. Anastasiadis, A. H. Whitaker, Longevity of free-living Hoplodactylus maculatus (Reptilia: Gekkonidae). *N. Z. J. Ecol.* **10**, 141-142 (1987).

88. R. C. Averill-Murray, A. P. Woodman, J. M. Howland, "Population ecology of the Sonoran desert tortoise in Arizona" in *The Sonoran desert tortoise: natural history, biology, and* conservation, T. R. van Devender Eds. (Arizona-Sonora Desert Museum, 2002), 109-134.

89. A. R. Britton, R. Whitaker, N. Whitaker, Here be a dragon: exceptional size in a saltwater crocodile (Crocodylus porosus) from the Philippines. *Herpetol. Rev.* **43**, 541-546 (2012).

90. L. A. Fitzgerald, Tupinambis lizards and people: a sustainable use approach to conservation and development. *Conserv. Biol.* **8**, 12-15 (1994).

91. J. L. Hickman, Observations on the skink lizard Egernia whitii (Lacepede). *Pap. Proc. R. Soc Tasmania*, **94**, 111 (1960).

92. P. R. Olofson, *ecosystem species and community profiles: Life histories and environmental requirements of key plants, fish and wildlife* (San Francisco Estuary Project c/o SF Bay Regional Water Quality Control Board. California, US, 2005).

93. B. Grzimek, N. Schlager, D. Olendorf, *Grzimek's animal life encyclopedia* (Detroit, US, 2003).

94. J. D. Riedle. thesis, University of Oklahoma (2001).

95. P. J. Stafford, J. R. Meyer, *Guide to the reptiles of Belize* (Academic Press. Cambridge, Massachusetts, US, 2000).

96. A. M. Audet, C. B. Robbins, S. Larivière, Alopex lagopus. *Mamm. Species* **713**, 1-10 (2002).

97. W. C. Pitt, J. Beasley, J. G. W. Witmer, *Ecology and management of terrestrial vertebrate invasive species in the United States* (CRC Press. Boca Raton, Florida, US, 2017).

98. A. C. Alberts, R. L. Carter, W. K. Hayes, E. P. Martins, *Iguanas: biology and conservation* (University of California Press, California, US, 2004).

99. G. Amori, S. Bertolino, S. Masciola, S. Moreno, J. Palomo, C. Rotondo, L. Luiselli, Aspects of demography in three distinct populations of garden dormouse, Eliomys quercinus, across Italy and Spain. *Rend. Lincei* **27**, 357-368 (2016).

100. Assessment, A. S. C., 2013. Bailey's Eastern Woodrat.

101. M. L. Augee, B. Gooden, A. Musser, *Echidna: extraordinary egg-laying mammal* (CSIRO publishing. Clayton, Australia, 2006).

102. D. M. Avery, G. Avery, N. G. Palmer, Micromammalian distribution and abundance in the Western Cape Province, South Africa, as evidenced by Barn owls Tyto alba (Scopoli). *J. Nat. Hist.* **39**, 2047-2071 (2005).

103. R, A. Avery, Clutch size and reproductive effort in the lizard Lacerta vivipara Jacquin. *Oecologia* **19**, 165-170 (1975).

104. G. A. Balme, L. Hunter, A. R. Braczkowski, Applicability of age-based hunting regulations for African leopards. *PloS One* **7**, p.e35209 (2012).

105. R. Barbault, Y. P. Mou, Population dynamics of the common wall lizard, Podarcis muralis, in southwestern France. *Herpetologica* **44**, 38-47 (1988).

106. M. A. Bartels, D. P. Thompson, Spermophilus lateralis. *Mamm. Species* **440**, 1-8 (1993).

107. M. Bekoff, Canis latrans. *Mamm. Species* **79**, 1-9 (1977).

108. P. Bergeron, M. Festa-Bianchet, A. Von Hardenberg, B. Bassano, Heterogeneity in male horn growth and longevity in a highly sexually dimorphic ungulate. *Oikos* **117**, 77-82 (2008).

109. J. Berry, Aspects of wildebeest Connochaetes taurinus ecology in the Etosha National Park-a synthesis for future management. *Madoqua* **20**, 137-148 (1997).

110. C. H. Bérubé, M. Festa-Bianchet, J. T. Jorgenson, Individual differences, longevity, and reproductive senescence in bighorn ewes. *Ecology* **80**, 2555-2565 (1999).

111. W. J. Bigler, J. H. Jenkins, Population characteristics of Peromyscus gossypinus and Sigmodon hispidus in tropical hammocks of south Florida. *J. Mammal.* **56**, 633-644 (1975).

112. G. Bino, T. R. Grant, R. T. Kingsford, Life history and dynamics of a platypus (Ornithorhynchus anatinus) population: four decades of mark-recapture surveys. *Sci. Rep.* **5**, 16073 (2015).

113. D. Blair, Update on the status of the San Salvador rock iguana, Cyclura rileyi rileyi. *Iguana Times* **1**, 1-3 (1991).

114. F. N. Blanchard, M. R. Gilreath, F. C. Blanchard, The eastern ring-neck snake (Diadophis punctatus edwardsii) in Northern Michigan (Reptilia, Serpentes, Colubridae). *J. Herpetol*. **13**, 377-402 (1979).

115. J. E. Bobick, M. Peffer, *Science and Technology Desk Reference. Ed. Series* (Gale Research Inc. Washington D.C., US, 1993).

116. S. A. Bouguerra, M. C. Bourdillon, Y. Dahmani, F. Bekkhoucha, Non insulin dependent diabetes in sand rat (Psammomys obesus) and production of collagen in cultured aortic smooth muscle cells. Influence of insulin. *J. Diabetes Res.* **2**, 37-46 (2001).

117. R. W. Braithwaite, Behavioural changes associated with the popu- lation cycle of Antechinus stuartii (Marsupialia). *Aust. J. Zool.* **22**, 45-62 (1974).

118. S. Broekhuizen, F. Maaskamp, Age determination in the European hare (Lepus europaeus Pallas) in the Netherlands. *Z. Säugetierkd*. **44**, 162-174 (1979).

119. A. M Bronikowski, J. Altmann, D. K. Brockman, M. Cords, L. M. Fedigan, A. Pusey, T. Stoinski, W. F. Morris, K. B. Strier, S. C. Alberts, Aging in the natural world: comparative data reveal similar mortality patterns across primates. *Science* **331**, 1325-1328 (2011).

120. L. N. Brown, Population dynamics of the western jumping mouse (Zapus princeps) during a four-year study. J. Mammal. **51**, 651-658 (1970).

121. J. L. Buckley, D. L. Spencer, P. Adams, Muskox (Ovibos moschatus) longevity. *J. Mammal.* **35**, 456-456 (1954).

122. V. de Buffrénil, G. Hémery, Variation in longevity, growth, and morphology in exploited Nile monitors (Varanus niloticus) from Sahelian Africa. *J. Herpetol*. **36**, 419-427 (2002).

123. C. Busch, A. I. Malizia, O. A. Scaglia, O. A. Reig, Spatial distribution and attributes of a population of Ctenomys talarum (Rodentia: Octodontidae). *J. Mammal.* **70**, 204-208 (1989).

124. D. Buys, H. M. Dott, Population fluctuations and breeding of eland Taurotragus oryx in a western Transvaal nature reserve. *Koedoe* **34**, 31-36 (1991).

125. G. A. Casey, W. A. Webster, Age and sex determination of striped skunks (Mephitis mephitis) from Ontario, Manitoba, and Quebec. *C. J. Zool.* **53**, 223-226 (1975).

126. J. Castanet, D. G. Newman, H. S. Girons, Skeletochronological data on the growth, age, and population structure of the tuatara, Sphenodon punctatus, on Stephens and Lady Alice Islands, New Zealand. *Herpetologica* **44**, 25-37 (1988).

127. Castro-Arellano, I., Zarza, H. and Medellín, R.A. Philander opossum. *Mamm. Species* **638**, 1-8 (2000).

128. J. Chamberlain, Stochastic life-history variation in populations of Western ribbon Snakes (Thamnophis 26ehaviou) in east Texas. *Biology Theses*, 3 (2011).

129. D. J. Chivers, Socio-ecology and Conservation of Malayan Forest Primates. *Environ. Conserv*. **4**, 102-102 (1977).

130. S. van Dyck, R. Strahan, *The Mammals of Australia* (New Holland Publishers. Sydney, Australia, 2008).

131. S. Churchfield, J. Hollier, V. K. Brown, Population dynamics and survivorship patterns in the common shrew Sorex araneus in southern England. *Acta Theriol.* **40**, 53-53 (1995).

132. B. R. Clark, E. O. Price, Sexual maturation and fecundity of wild and domestic Norway rats (Rattus norvegicus). *Reproduction* **63**, 215-220 (1981).

133. M. Clinchy, A. C. Taylor, L. Y. Zanette, C. J. Krebs, P. J. Jarman, Body size, age and paternity in common brushtail possums (Trichosurus vulpecula). *Mol. Ecol*. **13**, 195-202 (2004).

134. L. Cohen-Shlagman, Y. Yom-Tov, S. Hellwing, The biology of the Levant vole, Microtus guentheri. Israel. I. Population dynamics in the field. *Z. Saugetierkunde* **49**, 135-147 (1984).

135. L. R. Collins, *Monotremes and Marsupials* (Smithsonian Institute Press. Washington, D. C., US, 1973).

136. J. D. Congdon, R. D. Nagle, O. M. Kinney, R. C. van Loben Sels, T. Quinter, D. W. Tinkle, Testing hypotheses of aging in long-lived painted turtles (Chrysemys picta). *Exp. Gerontol.* **38**, 765-772 (2003).

137. M. B. Connior, T. S. Risch, Home range and survival of the Ozark Pocket Gopher (Geomys bursarius ozarkensis) in Arkansas. *Am. Midl. Nat.* **164**, 80-91 (2010).

138. W. E. Cooper, L. J. Vitt, Deferred agonistic 27ehaviour in a long-lived scincid lizard Eumeces laticeps. *Oecologia* **72**, 321-326 (1987).

139. C. J. Martinka, K. L. McArthur, *Bears, Their Biology and Management: A Selection of Papers from the Fourth International Conference on Bear Research and Management*

Held at Kalispell, Montana, USA, February 1977. Bear Biology Association. Missoula, Montana, US (1980).

140. A. J. Curtin, G. R. Zug, J. R. Longevity and growth strategies of the desert tortoise (Gopherus agassizii) in two American deserts. *J. Arid Environ*. **73**, 463-471 (2009).

141. A. I. Dagg, J. B. Foster, *The Giraffe: Its Biology. Behavior and Ecology* (Krieger Publishing Company. Malabar, Florida, US, 1980).

142. J. C. Daly, J. L. Patton, Growth, reproduction, and sexual dimorphism in Thomomys bottae pocket gophers. *J. Mammal*. **67**, 256-265 (1986).

143. R. W. Dapson, J. M. Irland, An accurate method of determining age in small mammals. *J. Mammal*. **53**, 100-106 (1972).

144. B. Dassonville, thesis, University of Rennes (1978).

145. J. H. M. David, thesis, University of Cape Town (1980).

146. D. P. DeMaster, I. Stirling, Ursus maritimus. *Mamm. Species* **145**, 1-7 (1981).

147. J. U, Din, M. A. Nawaz, Status of the Himalayan lynx in district Chitral, NWFP, Pakistan. *J. Anim. Plant Sci*. **20**, 17-22 (2010).

148. C. M. Drea, L. G. Frank, "The social complexity of spotted hyenas" in *Animal social complexity: intelligence, culture, and individualized societies*, F. B. M. de Wall & P. L. Tyack, Eds. (Harvard University Press, 2003), 121-148.

149. R. D. Earle, K. R. Kramm, Techniques for age determination in the Canadian porcupine. *J. Wildl. Manag*. **44**, 413-419 (1980).

150. M. J. Eaton, W. A. Link, Estimating age from recapture data: integrating incremental growth measures with ancillary data to infer age-at-length. *Ecol. Appl*. **21**, 2487-2497 (2011).

151. J. F. Eisenberg, D. E. Wilson, Relative brain size and demographic strategies in didelphid marsupials. *Am. Nat*. **118**, 1-15 (1981).

152. S. K. Eltringham, W. J. Jordan, "The Vicuna of the Pampa Galeras National Reserve the conservation issue" in *Problems in Management of Locally Abundant Wild Mammals*, P.A. Jewell Eds. (Acedemic Press, 1981), 277-289.

153. S. Erlinge, M. Sandell, Seasonal changes in the social organization of male stoats, Mustela erminea: an effect of shifts between two decisive resources. *Oikos* **47**, 57-62 (1986).

154. S. M. Ernest, Life history characteristics of placental nonvolant mammals: ecological archives. *Ecology* **84**, 402-3402 (2003).

155. B. D. Eshelman, G. N. Cameron, Baiomys taylori. *Mamm. Species* **285**, 1-7 (1987).

156. E. Nevo, A. Beiles, T. Spradling, Molecular evolution of cytochrome b of subterranean mole rats, Spalax ehrenbergi superspecies, in Israel. *J. Mol. Evol.* **49**, 215-226 (1999).

157. G. M. Fellers, C. A. Drost, Ecology of the island night lizard, Xantusia riversiana, on Santa Barbara Island, California. *Herpetol. Monogr*. **5**, 28-78 (1991).

158. G. f. Fisler, Age structure and sex ratio in populations of Reithrodontomys. *J. Mammal*. **52**, 653-662 (1971).

159. T. H. Fleming, The population ecology of two species of Costa Rican heteromyid rodents. *Ecology* **55**, 493-510 (1974).

160. S. S. Flower, Contributions to our Knowledge of the Duration of Life in Vertebrate Animals III. Reptiles. *Proc. Zool. Soc. Lond.* **95**, 911-981 (1925).

161. L. H. Frame, J. R. Malcolm, G. W. Frame, H. Van Lawick, Social Organization of African Wild Dogs (Lycaon pictus) on the Serengeti Plains, Tanzania 1967–1978 1. *Zeitschrift für Tierpsychol.* **50**, 225-249 (1979).

162. N. R. French, B. G. Maza, A. P. Aschwanden, Life spans of Dipodomys and Perognathus in the Mojave Desert. *J. Mammal.* **48**, 537-548 (1967).

163. J. W. Froehlich, R. W. Thorington, J. S. Otis, The demography of howler monkeys (Alouatta palliata) on Barro Colorado Island, Panama. *Int. J. Primatol.* **2**, 207-236 (1981).

164. D. G. Kleiman, V. Geist, M. C. McDade, *Grzimek's Animal Life Encyclopedia. Mammals I - IV* (Gale. Detroit, Michigan, US, 2004).

165. S. W. Gay, T. L. Best, Age-related variation in skulls of the puma (Puma concolor). *J. Mammal.* 77, 191-198 (1996).

166. D. J. Germano, L. R. Saslaw, P. T. Smith, B. L. Cypher, Survivorship and reproduction of translocated Tipton kangaroo rats in the San Joaquin Valley, California. *Endanger. Species Res.* **19**, 265-276 (2013).

167. N. B. Frazer, J. W. Gibbons, J. L.Greene, Exploring Fabens' growth interval model with data on a long-lived vertebrate, Trachemys scripta (Reptilia: Testudinata). *Copeia* **1990**, 112-118 (1990).

168. A. R. Glatston, M. Roberts, The current status and future prospects of the red panda (Ailurus fulgens) studbook population. *Zoo Biol.* **7**, 47-59 (1988).

169. C. J. Goin, O. B. Goin, G. R. Zug, *Introduction to herpetology* (W. H. Freeman & Co. Stuttgart, Germany, 1978).

170. T. C. Goldsmith, Mammal aging: Active and passive mechanisms and their medical implications. *Biosci. Hypotheses* **2**, 59-64 (2009).

171. L. R. Goshe, L. Avens, F. S. Scharf, A. L. Southwood, Estimation of age at maturation and growth of Atlantic green turtles (Chelonia mydas) using skeletochronology. *Mar. Biol.* **157**, 1725-1740 (2010).

172. B. Graw, M. Manser, Life history patterns and biology of the slender mongoose (Galerella sanguinea) in the Kalahari Desert. *J. Mammal.* **98**, 332-338 (2017).

173. A. D. Griffiths, Demography and home range of the frillneck lizard, Chlamydosaurus kingii (Agamidae), in northern Australia. *Copeia* **1999**, 1089-1096 (1999).

174. B. Grzimek, *Grzimek's Animal Life Encyclopedia. Mammals I - IV* (McGraw-Hill Publishing Company. New York, US, 1990).

175. C. P. Groves, Ceratotherium simum. *Mamm. Species* **8**, 1-6 (1972).

176. B Grzimek, *Grzimek's Animal Life Encyclopedia. Mammals I - IV* (McGraw-Hill Publishing Company. New York, US, 1990).

177. E. R. Guiler, D. A. Kitchener, Further observations on longevity in the wild potoroo, Potorous tridactylus. *Aust. J. Sci.* **30**, 105-106 (1967).

178. D. L. Gummer, thesis, University of Calgary (1997).

179. M. Hagan, Biological Resource Assessment of APN 302-076-10 California City, California. *APN* **302**, 76-100, (2018).

180. R. J. Hall, H. S. Fitch, Further observations on the demography of the great plains skink (Eumeces obsoletus). *Trans. Kans. Acad. Sci.* **74**, 93-98 (1971).

181. A. Hämäläinen, M. Dammhahn, F. Aujard, M. Eberle, I. Hardy, P. M. Kappeler, M. Perret, S. Schliehe-Diecks, C. Kraus, Senescence or selective disappearance? Age

trajectories of body mass in wild and captive populations of a small-bodied primate. *Proc. R. Soc. B.* **281**, p.20140830 (2014).

182. P. H. Harvey, R. D. Martin, T. H. Clutton-Brock, "Life histories in comparative perspective" in *Primate Societies*, B.B. Smuts, D.L. Cheney, R.M. Seyfarth, R.W. Wrangham, T.T. Struhsaker, Eds. (University of Chicago Press, 1987).

183. B.B. Smuts, D.L. Cheney, R.M. Seyfarth, R.W. Wrangham, T.T. Struhsaker, *Primate Societies* (The University of Chicago Press. Chicago, US, 1987).

184. J. J. Heatley, G. E. Mauldin, D. Y. A. Cho, A review of neoplasia in the captive African hedgehog (Atelerix albiventris). *Semin. Avian and Exotic Pet Med.* **14**, 182-192 (2005).

185. L. R. Hill, D. R. Lee, M. E. Keeling, Surgical technique for ambulatory management of airsacculitis in a chimpanzee (Pan troglodytes). *Comparative Med.* **51**, 80-84 (2001).

186. I. E. Hoffmann, E. Millesi, S. Huber, L. G. Everts, J. P. Dittami, Population dynamics of European ground squirrels (Spermophilus citellus) in a suburban area. *J. Mammal.* **84**, 615-626 (2003).

187. M. H. Yoon, S. J. Jung, S.J. H. S. Oh, Population structure and reproductive pattern of the Korean striped field mouse, Apodemus agrarius. *Korean J. Biol. Sci.* **1**, 53-61 (1997).

188. J. L. Hoogland, Cynomys ludovicianus. *Mamm. Species* **535**, 1-10 (1996).

189. H. N. Hoeck, Demography and competition in hyrax. *Oecologia* **79**, 353-360 (1989).

190. R. A. How, J. L. Barnett, A. J. Bradley, W. F. Humphreys, R. Martin, The population biology of Pseudocheirus peregrinus in a Leptospermum laevigatum thicket. *Possums and Gliders*, 261-268 (1984).

191. J. Hoyle, R. Boonstra, Life history traits of the meadow jumping mouse, Zapus hudsonius, in southern *Ontario. Can. Field-nat.* **100**, 537-544 (1986).

192. D. Hunsaker, D. Shupe, "Behavior of new world marsupials" in *The Biology of Marsupial*s, D. Hunsaker II Ed. (Acedemic Press, 1977).

193. E. V. Ivanter, Contribution to the ecology of Sicista betulina pall. *Aquilo, Ser. Zoologica.* **13**, 103-108 (1972).

194. W. Jezierski, Longevity and mortality rate in a population of wild boar. *Acta Theriol.* **22**, 337-348 (1977).

195. M. L. Jones, V. J. A. Manton, "History in captivity" in *The Biology and Management of an Extinct Species Pere David's Deer*, B.B. Beck, C. Wemmer Eds. (Noyes Publishing, 1983) 1-14.

196. M. L. Jones, Longevity of captive mammals. *Zool. Garten* **52**, 113-28 (1982).

197. R. Juskaitis, Life tables for the common dormouse Muscardinus avellanarius in Lithuania. *Acta Theriol.* **44**, 465-470 (1999).

198. J. Kingdon, *An Atlas of Evolution in Africa III (C and D)* (Academic Press. London, UK, 1982).

199. T. H. Kirkpatrick, Studies of Macropodidae in Queensland. 2. Age estimation in the grey kangaroo, the red kangaroo, the eastern wallaroo and the red-necked wallaby, with notes on dental abnormalities. *Queensland J. Agric. Anim. Sci.* **22**, 301-317 (1965).

200. D. G. Kleiman, D. S. Mack, A peak in sexual activity during mid-pregnancy in the golden lion tamarin, Leontopithecus rosalia (Primates: Callitrichidae). *J. Mammal.* **58**, 657-660 (1977).

201. G. Klevezal, *Recording structures of mammals* (CRC Press, Boca Raton, Florida, US, 1995).

202.J. Koenig, R. Shine, G. Shea, The ecology of an Australian reptile icon: how do blue-tongued lizards (Tiliqua scincoides) survive in suburbia? *Wildlife Res*. **28**, 214-227 (2001).

203.J. L. Koprowski, J. L. Roseberry, W. D. Klimstra, Longevity records for the fox squirrel. *J. Mammal*. **69**, 383-384 (1988).

204.Z. A. Krasiński, Dynamics and structure of the European bison population in the Białowieża Primeval Forest. *Acta Theriol*. **23**, 3-48 (1978).

205.N. P. E. Langham, The ecology of the common tree shrew, Tupaia glis in peninsular Malaysia. *J. Zool*. **197**, 323-344 (1982).

206.S. Larivière, Ictonyx striatus. *Mamm. Species* **698**, 1-5 (2002).

207.J. Larkin, thesis, University of Nevada (2006).

208.L. Laursen, M. Bekoff, Loxodonta africana. *Mamm. Species* **92**, 1-8 (1978).

209.C. P. Groves, D. M. Leslie Jr, B. A. Huffman, R. Valdez, K. Habibi, P. Weinberg, J. Burton, P. Jarman, W. Robichaud, *Family Bovidae (hollow-horned ruminants)* (Lynx Edicions. Barcelona, Spain, 2015).

210.W. Leuthold, Ecological separation among browsing ungulates in Tsavo east National Park, Kenya. *Oecologia* **35**, 241-252 (1978).

211.J. G. Lewis, R. T. Wilson, R.T. The ecology of Swayne's hartebeest. *Biol. Conserv*. **15**, 1-12 (1979).

212.D. B. Lindenmayer, R. C. Lacy, Metapopulation viability of Leadbeater's possum, Gymnobelideus leadbeateri, in fragmented old-growth forests. *Ecol. Appl*. **5**, 164-182 (1995).

213. J. P. Linduska, Longevity of some Michigan farm game mammals. *J. Mammal*. **28**, 126-129 (1947).

214. J. D. Litzgus, Sex differences in longevity in the spotted turtle (Clemmys guttata). *Copeia* **2006**, 281-288 (2006).

215. B. Lobert, A. K. Lee, "Reproduction and life history of Isoodon obesulus in Victorian heathland" in *Bandicoots and Bilbies*, J. H. Seebeck, P. R. Brown, R. L. Wallis, C. M. Kemper, Eds. (Surrey Beaty & Sons, 1990), 311-318.

216. L. J. Lombaard, Age determination and growth curves in the black-backed jackal, Canis mesomelas Schreber, 1775 (Carnivora: Canidae). *Ann. Transvaal Mus*. **27**, 135-169 (1971).

217. R. R. Lopez, M. E. Vieira, N. J. Silvy, P. A. Frank, Survival, mortality, and life expectancy of Florida Key deer. *The J. Wildl. Manag*. **67**, 34-45 (2003).

218. J. H. Lotze, S. Anderson, Procyon lotor. *Mamm. Species* **119**, 1-8 (1979).

219. D. W. MacDonlad, *The New Encyclopedia of Mammals* (Oxford University Press. Oxford, UK, 2001).

220. D. W. Macdonald, C. Newman, P. M. Nouvellet, C. D. Buesching, An analysis of Eurasian badger (Meles meles) population dynamics: implications for regulatory mechanisms. *J. Mammal*. **90**, 1392-1403 (2009).

221. A. L. McLean, thesis, University of Adelaide (2015).

222. E. Magnanou, J. Attia, R. Fons, G. Boeuf, J. Falcon, The timing of the shrew: continuous melatonin treatment maintains youthful rhythmic activity in aging Crocidura russula. *PloS One* **4**, p.e5904 (2009).

223. G. M. Maynes, Distribution and aspects of the biology of the parma wallaby, Macropus parma, in New South Wales. *Wildlife Res.* **4**, 109-125 (1977).

224. V. Mazak, Panthera tigris. *Mamm. Species* **152**, 1-8 (1981).

225. A. G. McAdam, S. Boutin, A.K. Sykes, M. M. Humphries, Life histories of female red squirrels and their contributions to population growth and lifetime fitness. *Ecoscience* **14**, 362-369 (2007).

226. J. A. McAllister, R. S. Hoffmann, Phenacomys intermedius. *Mamm. Species* **305**, 1-8 (1988).

227. J. McCosker, thesis, University of Queensland (1988).

228. D. A. McFarlane, J. Blake, The late Pleistocene hutias (Geocapromys brownii) of Red Hills Fissure, Jamaica. *Geol. J.* **40**, 399-404 (2005).

229. C. R. McMahon, M. J. Buscot, N. L. Wiggins, N. Collier, J. H. Maindonald, H. I. McCallum, D. M. Bowman, A two-phase model for smoothly joining disparate growth phases in the macropodid Thylogale billardierii. *PloS One* **6**, p.e24934 (2011.

230. P. A. Medica, F. B. Turner, D. D. Smith, Effects of radiation on a fenced population of horned lizards (Phrynosoma platyrhinos) in southern Nevada. *J. Herpetol*. **7**, 79-85 (1973).

231. L. Medway, *The Wild Mammals of Malaya (Peninsular Malaysia) and Singapore* (Oxford University Press.Oxford, UK, 1978).

232. P. K. Merrett, *Edentates*. Privately published (1983).

233. G. R. Michener, J. W. Koeppl, Spermophilus richardsonii. *Mamm. Species* **243**, 1-8 (1985).

234. K. D. Micone, thesis, University of Nevada (2002).

235. M. G. L. Mills, Hyaena brunnea. *Mamm. Species* **194**, 1-5 (1982).

236. P. O. Montiglio, D. Garant, P. Bergeron, G. D. Messier, D. Réale, Pulsed resources and the coupling between life-history strategies and exploration patterns in eastern chipmunks (T amias striatus). *J. Anim. Ecol.* **83**, 720-728 (2014).

237. M. P. English, P. Morris, Trichophyton mentagrophytes var. erinacei in hedgehog nests. *Sabouraudia* **7**, 118-121 (1969).

238. C. J. Moss, The demography of an African elephant (Loxodonta africana) population in Amboseli, Kenya. *J. Zool.* **255**, 145-156 (2001).

239. J. L. Mulder, Longevity records in the red fox. *Lutra* **47**, 51-52 (2004).

240. D. W. Müller, J. M. Gaillard, L. B. Lackey, J. M. Hatt, M. Clauss, Comparing life expectancy of three deer species between captive and wild populations. *Eur. J. Wildl. Res.* **56**, 205-208 (2010).

241. A. Myllymäki, Demographic mechanisms in the fluctuating populations of the field vole Microtus agrestis. *Oikos* **29**, 468-493 (1977).

242. N. P. Naumov, V. S. Lobachev, "Ecology of desert rodents of the USSR (jerboas and gerbils)" in *Rodents in desert environments*, I. Prakash, P. K. Ghosh Eds. (Springer, 1976), 465-606.

243. M. A. van Noordwijk, C. P. van Schaik, The effects of dominance rank and group size on female lifetime reproductive success in wild long-tailed macaques, Macaca fascicularis. *Primates* **40**, 105-130 (1999).

244. G. A. Novikov, *Carnivorous mammals of the fauna of the USSR*. Isreal Program for Scientific Translations (1962).

245.R. M. Nowak, J. L. Paradiso, *Mammals of the World* (John Hopkins Press, Baltimore, US, 1991).

246.H. Núñez, S. F. Fox, Liolaemus puritamensis, a new species of iguanid lizard previously confused with Liolaemus multiformis (Squamata: Iguanidae). *Copeia* **1989**, 456-460 (1989).

247.M. A. O'connell, "Ecology of didelphid marsupials from northern Venezuela" in *Vertebrate Ecology in the Northern Neotropics*, J. F. Eisenberg Ed. (Smithsonian Institution Press, 1979), 73-87.

248.T. P. O'Farrell, R. J. Olson, R. O. Gilbert, J. D. Hedlund, A Population of Great Basin Pocket Mice, Perognathus parvus, in the Shrub-Steppe of South-Central Washington. *Ecol. Monogr.* **45**, 1-28 (1975).

249.J. R. Oaks, J. M. Daul, G. H. Adler, Life span of a Tropical Forest rodent, Proechimys semispinosus. *J. Mammal.* **89**, 904-908 (2008).

250.M. Oakwood, thesis, Australian National University (1997).

251.I. Ogurlu, E. Gundogdu, I. Ceyhun Yildirim, Population status of jungle cat (Felis chaus) in Egirdir lake, Turkey. *J. Environ. Biol*. **31**, 179 (2010).

252.A. C. Ostroff, E. J. Finck, Spermophilus franklinii. *Mamm. Species*, **724**, 1-5 (2003).

253.R. N. Owen-Smith, *Megaherbivores: the influence of very large body size on ecology* (Cambridge university press, Cambridge, UK, 1988).

254.R. A. Palombit, Longitudinal patterns of reproduction in wild female siamang (Hylobates syndactylus) and white-handed gibbons (Hylobates lar). *Int. J. Primatol.* **16**, 739-760 (1995).

255. J. N. Pauli, J. P. Whiteman, B. G. Marcot, T. M. McClean, M. Ben-David, DNA-based approach to aging martens (Martes americana and M. caurina). *J. Mammal.* **92**, 500-510 (2011).

256. D. J. Pearson, B. Jones, *Lancelin Island skink recovery plan.* Department of Conservation and Land Management. Melville, Australia (2000).

257. O. P. Pearson, Longevity of the short-tailed shrew. *Am. Midl. Nat.* **34**, 531-546 (1945).

258. P. G. Pearson, A field study of Peromyscus populations in Gulf Hammock, Florida. *Ecology* **34**, 199-207 (1953).

259. R. L. Peterson, A review of the general life history of moose. *Nat. Can.* **101**, 9-21 (1974).

260. A. Pilastro, G. Tavecchia, G. Marin, Long living and reproduction skipping in the fat dormouse. *Ecology* **84**, 1784-1792 (2003).

261. R. A. Powell, R. D. Leonard, Sexual dimorphism and energy expenditure for reproduction in female fisher Martes pennanti. *Oikos* **40**, 166-174 (1983).

262. W. B. Preston, thesis, University of Oklahoma (1970).

263. M. Pucek, Polish Theriological Bibliography. *Acta Theriol.* **26**, 509-522 (1981).

264. F. Punzo, S. Chavez, Effect of aging on spatial learning and running speed in the shrew (Cryptotis parva). *J. Mammal.* **84**, 1112-1120 (2003).

265. J. Qu, W. Li, M. Yang, W. Ji, Y. Zhang, Life history of the plateau pika (Ochotona curzoniae) in alpine meadows of the Tibetan Plateau. *Z. Säugetierkd.* **78**, 68-72 (2013).

266. S. Moose, thesis, Gulbarga University (2017).

267. D. G. Read, Reproduction and breeding season of Planigale gilesi and P. tenuirostris (Marsupialia: Dasyuridae). *Aust. Mammal.* **7**, 161-173 (1984).

268. R. D. Stone, The social ecology of the Pyrenean desman (Galemys pyrenaicus)(Insectivora: Talpidae), as revealed by radiotelemetry. *J. Zool*. **212**, 117-129 (1987).

269. M. Richholt, L. Carbyn, "Using tooth sectioning to age swift fox" in *The swift fox: ecology and conservation of swift foxes in a changing world*, A. Marsha, L. Carbyn Eds. (University of Regina Press, 2003), 161-165.

270. J. P. Rood, Ecological and behavioural comparisons of three genera of Argentine cavies. *Anim. Behav. Monogr*. **5**, 3-83 (1972).

271. C. Ross, Life history patterns of New World monkeys. *Int. J. Primatol*. **12**, 481-502 (1991).

272. L. M. Rotterman, T. S. Jackson, *Sea otter, Enhydra lutris.* Marine Mammal Commission (1988).

273. E. Shargal, N. Kronfeld-Schor, T. Dayan, Population biology and spatial relationships of coexisting spiny mice (Acomys) in Israel. *J. Mammal.* **81**, 1046-1052 (2000).

274. F. Saucy, thesis, University de Neauchatel (1988).

275. I. R. Savic, Ecology of the mole rat Spalax leucodon Nordm. In Yugoslavia. *Proc. Natl. Acad. Sci. U.S.A*. **44**, 5-7 (1973).

276. F. R. Scarff, S. G. Rhind, J. S. Bradley, Diet and foraging behaviour of brush-tailed phascogales (Phascogale tapoatafa) in the jarrah forest of south-western Australia. *Wildlife Res.* **25**, 511-526 (1998).

277. I. Scharf, S. Meiri, Sexual dimorphism of heads and abdomens: different approaches to 'being large' in female and male lizards. *BioL. J. LinN. Soc*. **110**, 665-673 (2013).

278. C. M. Schmidt, J. U. Jarvis, N. C. Bennett, The long-lived queen: reproduction and longevity in female eusocial Damaraland mole-rats (Fukomys damarensis). *Afr. Zool.* **48**, 193-196 (2013).

279. C. Schradin, Comparative studies need to rely both on sound natural history data and on excellent statistical analysis. *R. Soc. Open Sci.* **4**, 170346 (2017).

280. C. W. Schwartz, E. R. Schwartz, *The wild mammals of Missouri* (University of Missouri. Missouri, US, 2002).

281. T. W. Clark, J. P. Gibbs, P. W. Goldstraw, Some demographics of the extirpation from the wild of eastern barred bandicoots (Perameles gunni) in 1988-91, near Hamilton, Victoria, Australia. *Wildlife Res*. **22**, 289-297 (1995).

282. S. Gursky, M. Shekelle, A. Nietsch, "The conservation status of Indonesia's tarsiers" in *Primates of the Oriental Night*, M. Shekelle Ed. (Indonesia Institude of Science, Research Center for Biology, 2008).

283. M. Sheridan, R. H. Tamarin, Space use, longevity, and reproductive success in meadow voles. *Behav. Ecol. Sociobiol*. **22**, 85-90 (1988).

284. J. Shield, Reproduction of the Quokka Setonix brachyurus, in captivity. *J. Zool*. **155**, 427-444 (1968).

285. A. Shima, "Life span" in *Suncus murinus*, K. Kondo Ed. (Japan Scientific Societies Press, 1986), 160-162.

286. J. Shoshani, J. F. Eisenberg, Elephas maximus. *Mamm. Species* **182**, 1-8 (1982).

287. A. W. F. Banfield, A. W. F. *The Mammals of Canada* (University of Toronto Press. Toronto, Canada, 1974).

288. P. Sinai, B. R. Krasnov, G. I. Shenbrot, I. Choshniak, Ecology and behaviour of the lesser Egyptian gerbil (Gerbillus gerbillus) (Rodentia: Gerbillidae) from the Negev Highlands and Arava valley, Israel. *Mammalia* **67**, 1-14 (2003).

289. A. R. E. Sinclair, Mammal population regulation, keystone processes and ecosystem dynamics. *Philos. Trans. Royal Soc. B.* **358**, 1729-1740 (2003).

290. E. M. Smirina, A. Y. Tsellarius, Aging, longevity, and growth of the desert monitor lizard (Varanus griseus Daud.). *Russ. J. Herpetol.* **3**, 130-142 (1996).

291. G. C. Smith, The biology of the yellow-footed antechinus, Antechinus flavipes (Marsupialia: Dasyuridae), in a swamp forest on Kinaba Island, Cooloola Quensland. *Wildlife Res.* **11**, 465-480 (1984).

292. M. J. Smolen, H.H. Genoways, R.J. Baker, Demographic and reproductive parameters of the yellow-cheeked pocket gopher (Pappogeomys castanops). *J. Mammal.* **61**, 224-236 (1980).

293. G. L. Smuts, Age determionation in Burchell's zebra (Equus burchelli antiquorum) from the Kruger National Park. *S. Afr. J. Wildlife Res.* **4**, 103-115 (1974).

294. A. T. Snider, J. K. Bowler, *Longevity of reptiles and amphibians in North American collections. Herpetological Circular, No. 21* (Society of the Study of Amphibians and Reptiles. Oxford, Ohio, US, 1992).

295. R. L. Snyder, S. C. Moore, Longevity of captive mammals in Philadelphia Zoo. *International Zoo Yearbook*, 8, 175-183 (1968).

296. C. A. Spinage, Geratodontology and horn growth of the impala (Aepyceros melampus). *J. Zool.* **164**, 209-225 (1971).

297. C. Spinage, *A territorial antelope: the Uganda waterbuck* (Academic Press. Cambridge, Massachusetts, US, 2012).

298. J. Steffen M. Krohn, K. Paarmann, C. Schwitlick, T. Brüning, R. Marreiros, A. Müller-Schiffmann, C. Korth, K. Braun, J. Pahnke, Revisiting rodent models: Octodon degus as Alzheimer's disease model? *Acta Neuropathol. Commun.* **4**, 91 (2016).

299. S. U. Stroganov, *Carnivorous mammals of Siberia* (Israel Program for Scientific Translations. Jerusalem, Israel, 1969).

300. M. E. Sunquist, S. N. Austad, F. Sunquist, Movement patterns and home range in the common opossum (Didelphis marsupialis). *J. Mammal.* **68**, 173-176 (1987).

301. M. R. Symonds, Life histories of the Insectivora: the role of phylogeny, metabolism and sex differences. *J. Zool.* **249**, 315-337 (1999).

302. E. Taube, J. Keravec, J. C. Vié, J. M. Duplantier, Reproductive biology and postnatal development in sloths, Bradypus and Choloepus: review with original data from the field (French Guiana) and from captivity. *Mammal Rev.* **31**, 173-188 (2001).

303. J. M. Taylor, J. H. Calaby, Rattus lutreolus. *Mamm. Species* **299**, 1-7 (1988).

304. P. J. Taylor, J. U. M. Jarvis, T. M. Crowe, K. C. Davies, Age determination in the Cape molerat Georychus capensis. *Afr. Zool.* **20**, 261-267 (1985).

305. T. V. Krylova, Vorastnaya struktura priodioy poputskii Mongol' skoi peschanki (*Meriones unguiculatus*) (Age structure of a natural population of the Mongolian jird). *Zool. Z.* **57**, 1842-1847 (1978).

306. R. C.Trout, A review of studies on populations of wild harvest mice (Micromys minutus (Pallas). *Mammal Rev.* **8**, 143-158 (1978).

307. N. V. Tupikova, G. A. Sidorova, E. A. Konovalova, A method of age determination in Clethrionomys. *Acta Theriol.* **13**, 99-115 (1968).

308. M. M. Paquin, G. D. Wylie, E. J. Routman, Population structure of the giant garter snake, Thamnophis gigas. *Conserv. Gen.* **7**, 25-36 (2006).

309. M. J. Van Staaden, Suricata suricatta. *Mamm. Species* **483**, 1-8 (1994).

310. K. Vernes, L. C. Pope, Reproduction in the northern brown bandicoot (Isoodon macrourus) in the Australian Wet Tropics. *Aust. J. Zool.* **57**, 105-109 (2009).

311. B. J. Verts, L. N. Carraway, Tamias minimus. *Mamm. Species* **653**, 1-10 (2001).

312. S. J. Ward, Life-History of the Eastern Pygmy-Possum, Cercartetus-Nanus (Burramyidae, Marsupialia), in South-Eastern Australia. *Aust. J. Zool.* **38**, 287-304 (1990).

313. S. A. Wich, S. S. Utami-Atmoko, T. M. Setia, H. D. Rijksen, C. Schürmann, J. Van Hoof, C. P. van Schaik, Life history of wild Sumatran orangutans (Pongo abelii). *J. Hum. Evol.* **47**, 385-398 (2004).

314. P. M. Wilkinson, T. R. Rainwater, A. R. Woodward, E. H. Leone, C. Carter, Determinate growth and reproductive lifespan in the American alligator (Alligator mississippiensis): evidence from long-term recaptures. *Copeia* **104**, 843-852 (2016).

315. R. D. Wooller, M. B. Renfree, E. M. Russell, A. Dunning, S. W. Green, P. Duncan, Seasonal changes in a population of the nectar-feeding marsupial Tarsipes spencerae (Marsupialia: Tarsipedidae). *J. Zool.* **195**, 267-279 (1981).

316. P. A. Woolley, Reproduction in Antechinomys laniger ('spenceri'Form) (Marsupialia: Dasyuridae): Field and Laboratory. *Wildlife Res.* **11**, 481-489 (1984).

317. P. A. Woolley, The species of Dasycercus Peters, 1875 (Marsupialia: Dasyuridae). *Mem. Mus. Vic*. **62**, 213-221 (2005).

318. R. H. Yager, C.B. Frank. The nine-banded armadillo for medical research. *Institute Laboratory Animal Research News*, 15, 4-5 (1972).

319. G. R. Zug, *Herpetology – an introductory biology of amphibians and reptiles* (San Diego Academic Press, California, US, 1993).

320. R. G. Zweifel, C. H. Lowe, *The ecology of a population of Xantusia vigilis, the desert night lizard* (American Museum of Natural History. New York, US, 1966).

321. S. Zohdy, B. D. Gerber, S. Tecot, M. B. Blanco, J. M. Winchester, P. C. Wright, J. Jernvall, Teeth, sex, and testosterone: aging in the world's smallest primate. *PLoS ONE* **9**, e109528 (2014).

Acknowledgements: Many thanks to the Natural History Museum, London and University Museum of Zoology, Cambridge for loans of specimens; facilitated by Martha Richter, Rob Asher and Matt Lowe. Assistance with lab work and materials: Tom Davies, Wendy Dirks, Dani Schmidt, Remmert Schouten, Pedro Viegas, John Cunningham & Duncan Murdock. Discussions: Chris Dean, Wendy Dirks, Jim Hopson, Thomas Martin, Rachel O'Meara, Stephen Naji, Tanya Smith & Emily Rayfield.

Funding: We acknowledge the European Synchrotron Radiation Facility, Grenoble, France, for provision of synchrotron radiation facilities on beamlines ID19 and ID16A (project ES152), and we would like to thank Peter Cloetens for assistance in using beamline ID16A. We also acknowledge the Paul Scherrer Institut, Villigen, Switzerland for provision of synchrotron radiation beamtime at beamline TOMCAT of the Swiss Light Source. The research leading to

these results has received funding from the European Community's Seventh Framework

Programme (FP7/2007-2013) under grant agreement n.°312284 (for CALIPSO). This project

was part-funded by a Natural Environmental Research Council studentship, and an Engineering

and Physical Sciences Research Council studentship, awarded to EN and PS (Grant number

NE/R009783/1). Thank you to the Natural History Museum London for contributing to travel for

PB via the Departmental Investment Fund for Earth Sciences. We thank Ginko Investments Ltd

for funding for materials and travel and the University of Bristol Bob Savage memorial fund for

travel for EN.

Author Contributions: IJC & PGG conceived and designed the project. EN, PGG, PB, KR &

IJC selected, prepared and curated specimens. EN, PGG, PB, VF, DH, TK, AK, FM, AP, BP,

PS, HS, PT & IJC performed the synchrotron experiments. EN, AK & IJC performed the

microCT experiments. EN processed and EN, CN & KY analysed the synchrotron data. EN &

IJC analysed the micro-CT data. EN, PGG & IJC discussed the interpretations. EN wrote the first

draft and created all figures; EN, PGG & IJC wrote the manuscript; all authors provided a critical

review of the manuscript and approved the final draft. This article originated as a Masters thesis

(University of Bristol), then a Ph.D. thesis (University of Southampton), performed by EN and

supported and supervised by PGG, PS, NJG, JJ, KRB, MJB & IJC.

Competing interests: The authors declare no competing interests.

Data and materials availability: The tomographic data that support the findings of this study

are available from the corresponding authors upon reasonable request. Physiological source data

for charts and graphs in figures 4 and 5b are provided with the paper in Extended Data and

Supplementary Information tables.

Supplementary Materials:

Materials and Methods

Supplementary text

Figures S1-S5

5   Tables S1-S5

References (*44-321*)

Fig. 1.

Fig. 1. Cementum of *Morganucodon* and *Kuehneotherium*. A, **B.** 3D reconstructions of **A**, *Morganucodon* right lower molar tooth NHMUK PV M 104134 (voxel size 2 μm, μCT) and **B**, *Kuehneotherium* right lower molar tooth NHMUK PV M 21095 (voxel size 1.2 μm, PPC-SRμCT). Green = cementum. **C**, **D.** Transverse PPC-SRμCT virtual thin-sections (0.33 μm voxel size) of roots of **C**, NHMUK PV M 104134 and **D**, NHMUK PV M 27436 (*Kuehneotherium*). Red bracketed line highlights cementum surrounding dentine. **E**, **F.** Close-ups of boxes in **C**, **D**,

with five and four circumferential light/dark increment pairs highlighted by red arrows, respectively. **G**. Synchrotron nanotomographic virtual thin-section of *Morganucodon* (30 nm voxel size) - close-up of region near box in **E.** Vertical red arrows = cementum increments; horizontal blue arrows, dashed blue lines and Sf = radial bands of Sharpey's fibres; yellow dashed line and glT = granular layer of Tomes; green dashed line and hlH-S = hyaline layer of Hopewell-Smith.

Fig. 2.

Fig. 2. Three-dimensional segmentation of *Morganucodon* and *Kuehneotherium* specimens with the highest cementum increment counts. A, **B.** Transverse virtual thin-sections of PPC-SRμCT reconstructions (0.33 μm³ voxel size). **A**, *Morganucodon* specimen NHMUK PV M 104127 showing 55 μm thick cementum layer around root dentine. **B**, *Kuehneotherium* specimen UMZC Sy 141 showing 32 μm thick cementum. **C**, **D.** Detail of cementum of **C**, *Morganucodon* and **D**, *Kuehneotherium*. Cementum increments highlighted by 14 and nine multi-colouredarrows, respectively. **E**, **F.** 3D segmentations of cementum increments of **E**, *Morganucodon*, and **F**, *Kuehneotherium*. The colour of each increment corresponds to arrow colour in **C**, **D**.

Fig. 3.

Fig. 3. Shared increment patterns between m1 and m2 tooth-root cementum and the dentary of *Morganucodon* specimen NHMUK PV M 96413. A. Four lines of arrested growth and a fifth incipient one are visible within the dentary periosteal region, each highlighted by 3D segmented bands of differing colour corresponding to coloured arrows in accompanying transverse PPC-SRμCT slice. Only LAGs persisting through the volume are segmented/highlighted. This pattern is mirrored in **B,** the anterior root of the m1 tooth; **C,** the posterior root of the same m1; and **D,** the anterior root of the m2 tooth.

Fig. 4.

Fig. 4. Lifespan and metabolic estimates of *Morganucodon* and *Kuehneotherium*. A.

Histogram of lifespan estimates from cementum increment counts. **B**. Log_{10} biplot of mean body

mass (g) against maximum wild lifespan (years) for extant mammals (n = 279), extant non-avian

reptiles (n = 252), and fossil mammaliaforms. **C.** Log_{10} biplot of mean body mass (g) against

mass specific standard metabolic rate (msSMR; $mL.O_2.hr^{-1}.g^{-1}$) for extant mammals (n = 117)

and extant reptiles (n = 55), and estimates for fossil mammaliaforms. OLS regression lines in **B**,

C, show extant mammals (black) and extant reptiles (blue), 95% confidence intervals shown

with dashed lines, 95% predictor intervals by dotted lines.

Fig. 5.

Fig. 5. Femoral foramina and estimates of relative femoral bloodflow for _Morganucodon_. A.

3D µCT reconstruction of _Morganucodon_ femur (specimen UMZC Eo PC 19_6, voxel size 4

µm), with all identifiable foramina segmented/highlighted with colour. **B**. $\log_{10}$ biplot of mean

body mass (g) against estimated blood flow index (Qi; mm^3) for extant non-varanid reptiles (n =

22), extant varanid reptiles (n=8), extant mammals (n=69), and _Morganucodon_. OLS regression

lines in **B** are extant mammals (black), extant non-varanid reptiles (blue) and extant varanids

(purple), with 95% confidence intervals represented by dashed lines.