Dynamic evolution of *de novo* DNA methyltransferases in rodent and primate genomes

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Abstract

Transcriptional silencing of retrotransposons via DNA methylation is paramount for mammalian fertility and reproductive fitness. During germ cell development, most mammalian species utilize the *de novo* DNA methyltransferases DNMT3A and DNMT3B to establish DNA methylation patterns. However, many rodent species deploy a third enzyme, DNMT3C, to selectively methylate the promoters of young retrotransposon insertions in their germline. The evolutionary forces that shaped DNMT3C's unique function are unknown. Using a phylogenomic approach, we confirm here that *Dnmt3C* arose through a single duplication of *Dnmt3B* that occurred between 45-71Mya in the last common ancestor of muroid rodents. Most importantly, we reveal that DNMT3C is composed of two independently evolving segments: the latter two-thirds has undergone recurrent gene conversion with *Dnmt3B*, whereas the N-terminus has not undergone gene conversion and has evolved under strong diversifying selection. We hypothesize that positive selection of *Dnmt3C* is the result on an ongoing evolutionary arms race with young retrotransposon lineages in muroid genomes. Interestingly, although primates lack DNMT3C, we find that the N-terminus of DNMT3A has also evolved under diversifying selection. Thus, the N-termini of two independent *de novo* DNMT enzymes have evolved under diversifying selection in rodents and primates. We hypothesize that repression of young retrotransposons might be driving the recurrent innovation of a functional domain in the N-termini on germline DNMTs in mammals.

Introduction

The deposition of methylation on DNA is a deeply conserved process. In mammals, it plays crucial roles in genome stability, development, genomic imprinting and chromosome-wide epigenetic silencing such as X-inactivation (1). Mammalian DNMTs (DNA methyltransferases) are enzymes that catalyze the addition of a methyl group onto cytosines (2). Most mammals encode three catalytically active enzymes (DNMT1, DNMT3A, and DNMT3B) and one non-enzymatic germ cell-specific cofactor (DNMT3L) (2-5). While DNMT1 targets hemi-methylated cytosines (maintenance DNA methyltransferase) (6-8), DNMT3A and 3B are classified as *de novo* methyltransferases that target unmodified sites (9-12). In mice, constitutive genetic knockouts of *Dnmt1, Dnmt3A,* or *Dnmt3B* are lethal, whereas *Dnmt3L* mutations lead to sterility (10, 13, 14).

Phylogenetic analyses have suggested that the DNMT enzymes belong to the clade of 5-cytosine methyltransferases, which likely predated the origin of eukaryotes (5, 15). Although both *Dnmt1* and *Dnmt3A* were present in the common ancestor of all metazoans, *Dnmt3B* is believed to have arisen by a gene duplication event close to the origin of tetrapods (5, 16). Closer phylogenetic analyses in several taxa have revealed mammalian lineage-specific duplications, including the duplication and co-retention of several *Dnmt1* paralogs in marsupials (17) and the evolution of *Dnmt3L* from *Dnmt3A* in eutherian mammals (18). Another gene duplication from *Dnmt3B* gave rise to *Dnmt3C* in muroid rodents where it has acquired a distinct, non-redundant role in retrotransposon repression during spermatogenesis (19, 20). Thus, a series of ancient and recent gene duplications have led to the current repertoires of mammalian DNMTs.

Mammalian DNA methylation plays a critical role in retrotransposon silencing in the germline (21). Germ cell development is a particularly vulnerable stage for mammalian genomes, because many epigenetic chromatin marks that otherwise repress retrotransposons -like DNA methylation- are transiently erased. However, re-establishing silencing of retrotransposons in mammalian genomes is challenging, because of their vast sequence heterogeneity. Retrotransposons can belong to many evolutionarily distinct families that each exhibit rapid sequence divergence (22). In mice, during male fetal germ cell development in mice, two distinct waves of *de novo* methylation target retrotransposons according to their age (23). During the first wave, evolutionarily old retrotransposons gain methylation together with the rest of the genome. However, evolutionarily young and transcriptionally active retrotransposons are refractory to this wave and require the piRNA pathway - a small RNA-based defense system - to target DNA methylation to their promoters (23, 24).

Two recent studies showed that DNMT3C is the crucial enzyme for this process (19, 20). *Dnmt3C* knock-out (KO) males are sterile and their germ cell methylation profiles are similar to those of piRNA mutants, with a 1% drop in genome-wide DNA methylation content that selectively affects the promoters of young copies of LINE and ERVK retrotransposons (20, 23, 25). This contrasts with germ cell-specific *Dnmt3B* KO, which has no impact on male fertility (26), whereas constitutive *Dnmt3B* KO are embryonic lethal (10). On the other hand, germ cell-specific *Dnmt3*A KO males are infertile and display mild alteration of the methylation profiles of

a few older retrotransposon families (26, 27). This suggests that *Dnmt3A* might contribute to the first wave of *de novo* methylation and act non-redundantly of *Dnmt3C* for the silencing of mouse retrotransposons.

Catalytically active DNMT3s have three well-defined domains. The most C-terminal region encodes the methyltransferase domain (MTase) that includes highly conserved protein motifs that catalyze the addition of methyl groups (28, 29). The central portion encodes two chromatin-reading domains that play important roles in their targeting and regulation (30). The ADD (ATRX-DNMT3-DNMT3L) domain is repelled by methylated states of the histone modification H3K4 (31-34), preventing *de novo* DNA methylation at H3K4me3-enriched gene promoters. The PWWP (Pro-Trp-Trp-Pro) domain, helps anchor DNMT3 proteins to nucleosomes marked by histone modification H3K36me (35-39). The H3K36me mark is enriched in the body of actively transcribed genes and pericentromeric heterochromatin. Interestingly, mouse *Dnmt3C* lost the two exons coding for the PWWP domain, making it unique among catalytically active DNMT3s (19, 20). In contrast to the central and C-terminal segments, the N-terminal portion of DNMT3s remain completely uncharacterized.

Based on both its recent origin and its role in silencing young, potentially rapidly adapting retrotransposon families, we speculated that *Dnmt3C* might be participating in ongoing genetic conflicts in rodent genomes (22). We therefore performed a detailed phylogenetic survey of rodent genomes to investigate *Dnmt3C*'s age and the evolutionary forces that shape its unique function. Extending previous findings, we dated *Dnmt3C's* evolutionary origin in the common ancestor of muroids between 45 and 71 million years ago. Unexpectedly, we found a pattern of gene conversion between *Dnmt3B* and *Dnmt3C* paralogs throughout muroid evolution. This gene conversion recurrently homogenizes the latter two-thirds of DNMT3B and DNMT3C but does not extend to their N-terminal domains. We provide evidence of strong diversifying selection in the N-terminal tail of DNMT3C, but not DNMT3B, which we hypothesize is the result of an ongoing evolutionary arms race with active retrotransposons in these lineages. Although *Dnmt3C* is not present outside rodents, we found that the N-terminal tail of *DNMT3A* has similarly evolved under diversifying selection in primates. Thus, two distinct DNMT enzymes display hallmarks of ongoing genetic conflicts with retrotransposons in two separate mammalian lineages.

Results

Evolutionary origins and dynamics of *Dnmt3C* in rodents

To investigate the evolutionary age and dynamics of *Dnmt3C*, we retrieved and annotated DNMT3 sequences in partially or fully assembled genomes of 19 species of Glires – which include rodents and lagomorphs (Figure 1A, see methods). Like other mammals, most species of Glires encode unique *Dnmt3A, Dnmt3L* and *Dnmt3B* genes within syntenic loci present in all placental mammals (Figure 1A). However, in the sub-group of muroid species, the syntenic locus containing *Dnmt3B* also encodes *Dnmt3C* (Figure 1A) (20).

We investigated genomes from 11 muroid and 8 "outgroup" species and used available transcriptome or *de novo* gene assemblies to annotate coding sequences (CDS, see methods). In some cases, genome assemblies allowed us to tentatively assign gene orthology using shared synteny. However, in most cases, genome assemblies were too fragmented to reconstruct genomic contexts. Instead, we focused on retrieving partial or full-length sequences of putative *Dnmt* genes. We then constructed a multiple alignment and used maximum likelihood methods to build a gene phylogeny (see methods for details). Using this approach, we were able to resolve all retrieved sequences into distinct clades of DNMTs (Figure 1B).

If *Dnmt3C* arose from *Dnmt3B* in the last common ancestor of all muroids, we would expect *i) Dnmt3C* sequences to branch inside the *Dnmt3B* clade and *ii)* form two independent lineages following the split of muroids from other rodents and lagomorphs. Our first expectation was met; all putative *Dnmt3C* sequences branched within the *Dnmt3B* clade, supporting the close relatedness of these two genes relative to other *Dnmts* (Figure 1B). Moreover, a detailed phylogeny including all *Dnmt3B* and *Dnmt3C* orthologs was consistent with a single duplication event (Figure 1C). Based on the presence of *Dnmt3C* in mountain blind mole rats (*Nannospalax galili*), but not beavers or guinea pigs (*Castor canadensis* and *Cavia porcellus*), we estimate that the duplication occurred before the radiation of muroids between 45 and 71 million years ago (40).

However, the second expectation of independent evolution of *Dnmt3B* and *Dnmt3C* genes following *Dnmt3C's* origin was not met. While most *Dnmt3B* and *Dnmt3C* genes grouped into two distinct clades according to the accepted muroid species phylogeny (41), both the prairie vole (*Microtus ochrogaster*) and the mountain blind mole rat (*Nannospalax galili*) had *Dnmt3B*

and *Dnmt3C* paralogs that were more closely related to each other than to their respective orthologs (Figure 1C, asterisks). This pattern could indicate separate origins of *Dnmt3C* in these species or alternatively, recent gene conversion. It is also possible that partial gene conversion between *Dnmt3B* and *Dnmt3C* occurred in other muroid species but was not evident in phylogenetic analyses based on full-length sequences.

We therefore used a likelihood-based method, GARD, to map putative recombination breakpoints between *Dnmt3B* and *Dnmt3C* (see methods, (42)). Such analyses aim to identify recombination breakpoints based on segments of multiple alignments that have clearly discordant phylogenetic histories from each other. We identified three high confidence breakpoints in muroid *Dnmt3B* and *Dnmt3C* sequences, partitioning the aligned sequences into four segments with distinct evolutionary histories- A, B, C, and D (Figure 2A). Upon generating phylogenies of each segments independently, we observed a discordance between these segments was not limited to prairie vole and mountain blind mole rat (Figure 2B), suggesting that gene conversion between *Dnmt3B* and *Dnmt3C* occurred in many muroid lineages.

Next, we investigated the individual evolutionary trajectories of the distinct recombination segments within *Dnmt3C*. Consistent with rampant gene conversion, nucleotide phylogenies showed that segment B and C - encoding the ADD and part of the MTase (Figure 2A) - grouped *Dnmt3C* and *Dnmt3B* paralogs by species (Figure 2B and Supplementary Figure S1) rather than by orthology groups. We also found evidence for gene conversion in Segment D, which encodes the rest of the MTase (Supplementary Figure S1B). With the exception of prairie voles and mountain blind mole rats, we found no evidence for gene conversion in segment A, which encodes the N-terminal tail of DNMT3C (Figure 2A). Indeed, a phylogeny based on segment A alone almost perfectly separated the *Dnmt3B* and *Dnmt3C* paralogs based on orthology groups, consistent with divergent evolution of the two genes following duplication (Figure 2B). Thus, in most muroids and in contrast to the rest of the gene, the 5' ends of *Dnmt3B* and *Dnmt3C* do not appear to have engaged in recent gene conversion. Consistent with these findings, DNMT3B and DNMT3C protein sequences shared much higher homology in their C-terminal compared to their N-terminal domains (Figure 2C).

To further confirm our findings, we investigated the coding and non-coding genomic sequences of *Dnmt3B* and *Dnmt3C* for signatures of high sequence identity. High nucleotide identities between mouse *Dnmt3C* and *Dnmt3B* were evident not only in coding exons but also across

many introns (Figure 2D). More specifically, all introns in segment C displayed more than 70% identity in segment C but not in segment A (Figure 2D), consistent with our recombination breakpoint analysis (Figure 2A). We found an even more evident pattern of sequence homogenization between *Dnmt3B* and *Dnmt3C* in genomes of rats and mountain blind mole rats (Figures S2C and S2D respectively). In particular, the high sequence identity between the *Dnmt3B* and *Dnmt3C* loci in mountain blind mole rats (Supplementary Figure S1D) supports the hypothesis that this species, as well as prairie voles (Figure 2B), engaged in gene conversion more recently that other muroids. Taken together, these results suggest that following duplication, *Dnmt3B* and *Dnmt3C* have been subject to extensive gene conversion, except in their 5' ends. Thus, DNMT3C N-termini evolve under distinct evolutionary trajectories from their DNMT3B counterparts, while the central domains and C-termini of Dnmt3B, and Dnmt3C exchange sequences to remain similar within each genome.

DNMT3C N-terminal domain evolve under positive selection

Gene conversion has homogenized several segments of DNMT3C and DNMTB, but not their N-terminal domains. We hypothesized that this could be to retain the functional divergence of DNMT3B and DNMT3C in their N-terminal domains. For example, loss of the ancestral PWWP domain in DNMT3C may have allowed it to specialize for functions distinct from DNMT3B. If this were the case, we might expect to find additional differences in the selective constraints that act on *Dnmt3B* versus *Dnmt3C*, especially in their N-terminal domains. We, therefore, investigated how the DNMT3B and DNMT3C N-terminal domains may have diverged in their selective constraints.

Based on branch lengths (Figure 1B), *Dnmt3C* appears to be the most divergent of all *Dnmt3* genes in muroid rodents, followed by *Dnmt3L, Dnmt3B*, and finally *Dnmt3A*, which is most highly conserved. To evaluate selective constraints, we calculated the rates of non-synonymous (amino-acid altering, dN) and synonymous (silent, dS) substitutions across orthologous sequences of all *Dnmt* genes. *Dnmt3C* displays the highest average pairwise dN/dS of all *Dnmt3* genes *(*0.88*)* compared to *Dnmt3L* (0.23), *Dnmt3B* (0.22), and *Dnmt3A* (0.02) (Figure 1B). Higher dN/dS values could reflect relaxation of selective constraint. Alternatively, these higher values could be the result of diversifying selection acting on *Dnmt3C*.

To distinguish between these possibilities, we used likelihood methods implemented in the PAML package to detect signatures of positive selection (43). Muroid species are an ideal species set for these analyses because they span a short evolutionary time (~40 million years) with low saturation of dS (41). We separately analyzed each of the four recombination segments across all orthologs identified in muroids. Because some *Dnmt3C* genes are based on incomplete gene models, each segment alignment contained between 8 to 11 species (Table 1). We then used PAML to identify site that were subject to positive selection (see methods) (43). We found no evidence of positive selection having acted on *Dnmt3B*. In contrast, we found strong support for positive selection having acted on segment A of *Dnmt3C*, but not on segments B, C, or D (Table1).

In segment A of *Dnmt3C,* PAML analyses estimated 49% of sites to have evolved with an average dN/dS greater than 1 indicative of potential diversifying selection; their average dN/dS was estimated to be 1.6. Of these, four sites were highlighted with a high posterior probability of having evolved under positive selection (Bayes Empirical Bayes [P] ≥ 90%, Table1 and Figure 3B). These sites (codons 54, 57, 95 and 96 in mouse *Dnmt3C*) all cluster within the most 5' end of the gene (first 300bp of the CDS) and display extensive diversification in both charge and hydrophobicity across muroids (Figure 3B). For site 95 and 96, rapid evolution disrupts a highly conserved arginine patch of unknown function, which is highly conserved among muroid DNMT3B proteins (Figure 3C). Thus, in addition to the loss of the PWWP domain, DNMT3B and DNMT3C differ in the selective constraints they are subject to. The signature of positive selection and loss of the PWWP domains makes *Dnmt3C* unique among all *Dnmt3* genes.

Selective constraints acting on DNMT3 proteins in primate genomes

The evolutionary birth of *Dnmt3C* afforded muroid rodents a unique opportunity to silence young, active retrotransposon families by DNA methylation. However, most mammalian genomes face a similar pressure by young retrotransposon lineages and yet do not encode *Dnmt3C*. We, therefore, hypothesized that non-rodent mammalian species might deploy alternate mechanisms, possibly other DNMT enzymes, to achieve DNMT3C-like repression of their active retrotransposons. If true, we might expect *Dnmt3* genes to be locked in these molecular arms-races, and therefore subjected to similar selective pressures as *Dnmt3C* in muroid rodents *i.e.,* diversifying selection.

To investigate this possibility, we analyzed the evolutionary constraints that act on *DNMT3* genes in primates, a distinct lineage of mammals that have substantial genomic resources across multiple species in a comparable evolutionary timespan to muroid rodents (40). Using maximum likelihood-based analyses, we found strong evidence of diversifying selection acting on *DNMT3A* and marginal evidence of positive selection in the catalytically-inactive co-factor *DNMT3L* (Table 2). In contrast, we found no evidence of diversifying selection in primate *DNMT3B* (Table 2) or muroid *DNMT3A* (Table 1).

As in muroid *Dnmt3C*, the diversifying selection signature also appeared to primarily map to the N-terminal domain of primate *DNMT3A* (codons 61 and 81, Table 2 and Figure 4). To rule out that this signature could be due to unaccounted recombination, we performed GARD analyses. This identified a single break point (within the first 1kb of the CDS), however, while a maximum likelihood phylogeny of the first segment (including the rapidly evolving sites) had strong bootstrap support, the second segment did not (Supplementary Figure S2). Further inspecting this segment showed a high rate of CpG mutations which prevents appropriate reconstruction of its evolution and accurate selection analyses. We therefore conclude that there is not sufficient evidence for gene conversion affecting *DNMT3A* evolution in primates. In spite of this, PAML analysis of *DNMT3A* putative 1st (N-terminal) segment also identifies sites 61 and 81 as evolving under positive selection (not shown).

Thus, rather unexpectedly, we find evidence of diversifying selection on distinct *DNMT3* genes in rodent and primate genomes (Tables 1 and 2). Our findings suggest that the N-terminal portions of DNMT3 proteins wage evolutionary arms races for DNA methylation of young, active retrotransposons in different mammalian lineages. They further raise the possibility that DNMT3A, which is universal to all mammals, may be the original DNMT that represses young retrotransposons. The subsequent birth of DNMT3C in muroid rodents may have absolved DNMT3A of this role, which could be why we cannot detect any signatures of diversifying selection in *Dnmt3A* in rodent species.

Discussion

Retrotransposons pose a significant fitness challenge to host genomes. To protect themselves, host genomes deploy multipronged strategies to curb retrotransposon activity. Here we identified the selective forces shaping the function of a recently duplicated DNA

methyltransferase, DNMT3C, that specifically targets evolutionarily young retrotransposons in muroid rodents. We found that *Dnmt3C* has undergone recurrent gene conversion with its parental gene *Dnmt3B*, except for the N-terminal domain. These findings are reminiscent of previous studies of gene families subject to genetic conflicts (44). For example, the true evolutionary histories of the mammalian antiviral *IFIT1/IFIT1B* paralogs, which diverged 100 million years ago, were also confounded by recurrent gene conversion (45). Similarly, recurrent gene conversion affected the histone-fold domain but not the distinct N-terminal tails of centromeric histone paralogs in *Drosophila* species (46). In all these cases (*Dnmt3, IFIT, Cid*) as well as several additional examples (44), natural selection likely selects against gene conversion in the domain that likely drives the functional diversification of the paralogs. As a result, only the non-converted gene segment can recreate the true evolutionary history of the paralogs. In all these cases, the close proximity of the paralogs following gene duplication might have facilitated multiple episodes of gene conversion between them.

We found that the N-terminal domain of *Dnmt3C*, but not its parental gene *Dnmt3B*, has evolved under strong diversifying selection. Diversifying selection - especially in a host 'defense' gene - is a signature of an evolutionary arms race between host genomes and retrotransposons (22). As host genomes deploy repressive chromatin strategies, retrotransposons must adapt to ward off host repression, in turn spurring host adaptation. The evolutionary arms race model further makes the prediction that residues or domains that directly engage in the antagonism should be rapidly evolving. Thus, the positive selection in *Dnmt3* genes could be the result of active antagonism by a product of young retrotransposons (RNA or protein). Under this model, the positive selection would allow the DNMT3 proteins to evade binding and antagonism by young retrotransposons.

We favor the alternate model that positive selection shapes the targeting of DNMT3 proteins to young retrotransposons to mediate their silencing. This predicted activity would be similar to the KZNF (KRAB domain containing Zinc Finger) proteins, which use rapid evolution of their DNA-binding domains to keep pace with a changing nucleotide landscape of retrotransposon families (47). We hypothesize that similar evolutionary dynamics could drive the diversifying selection of the N-terminal domains in rodent DNMT3C and primate DNMT3A proteins. Interestingly, DNMT3A exists both as a long isoform, A1, and a N-terminus-less short isoform, A2 (48), as a result of alternative promoter usage. We posit that the long DNMT3A1 isoform may target young retrotransposons in male germ cells in DNMT3C-less mammalian species, such as primates.

The recurrent signature of rapid evolution within the N-termini of two different DNMT3 proteins in different mammalian lineages may highlight a novel functional domain that may be key to DNMT targeting of retrotransposons. Unlike the canonical PWWP, ADD and MTase domains, however, this domain may be characterized by its rapid evolution rather than conservation. How this domain engages with retrotransposons remains to be determined. In contrast to KZNF proteins, there is no suggestion that DNMT3 proteins have DNA sequence-binding specificity. Instead, it is possible that this region mediates interaction with components of the piRNA pathway - some of which are rapidly evolving in other animals (49, 50).

In sum, the DNMT3C N-terminal domains can be distinguished from other DNMT3 proteins in two key aspects: diversifying selection but also loss of a coding PWWP domain, which is essential for coupling *de novo* DNA methylation to local chromatin environment, via recognition of H3K36 methylated histones (36, 37, 39). In DNMT3B, the PWWP domain binds H3K36me3 marks, which are typical of transcribed gene bodies (51). In DNMT3A, the PWWP domain is intact and was recently shown to mediate DNMT3A-dependent methylation of intergenic sequences (52). We hypothesize here that DNMT3C's N-terminal domain may be required to substitute for PWWP-dependent chromatin-targeting function. However, the mode of targeting of DNMT3C to young retrotransposon promoters remains to be determined.

In conclusion, our evolutionary studies identified a new functional domain in DNMT3C, a DNA methyltransferase that mediates full silencing of the most active, rapidly-adapting retrotransposon families in rodent genomes. Furthermore, based on our findings of diversifying selection in primate DNMT3A, we suggest that diversifying selection of enzymes that methylate retrotransposons in developing germ cells might be pervasive across mammalian genomes, although this targeting may be mediated by distinct DNMT3 paralogs.

Methods

Identification of DNMT3 orthologs

To identify *Dnmt* orthologs we performed tBLASTn searches on the NCBI non-redundant nucleotide database (53, 54), using reference protein sequences of mouse DNMT3A (NP_031898.1), DNMT3B (XP_006498745.1), DNMT3L (NP_001075164.1) as well as the predicted protein sequence from the *Dnmt3C* cDNA cloned from male fetal gonads (20). While most *Dnmt3s* have predicted sequences in reference databases, *Dnmt3c* genes are not

annotated in most muroid genomes. In these cases we queried genomes directly using tBLASTn, and predicted gene models from contigs using GeneWise (55). CDSs were annotated based on the longest mouse gene model.

Queried genomes

We used the following genome assemblies to predict *Dnmt3B* and *Dnmt3C* gene models. Muroids: *Mus musculus* (UCSC mm10), *Mus spretus* (Sanger, SPRET_EiJ), *Mus caroli* (Sanger, CAROLI_EiJ), *Mus pahari* (Sanger, Pahari_EiJ), *Apodemus sylvaticus* (NCBI, GCA_001305905.1_ASM130590v1), *Rattus norvegicus* (UCSC, rn6), *Peromyscus maniculatus* (NCBI, GCF_000500345.1_Pman_1.0), *Myodes glareolus* (NCBI, GCA_001305785.1_ASM130578v1), *Microtus agrestis* (NCBI, GCA_001305995.1_ASM130599v1), *Microtus ochrogaster* (NCBI, MicOch1), *Mesocricetus auratus* (NCBI, MesAur1), *Cricetulus griseus* (UCSC, criGri1), *Nannospalax galili* (NCBI, GCF_000622305.1_S.galili_v1.0).

Glires: *Castor canadensis* (NCBI, C.can genome v1.0), *Oryctolagus cuniculus* (UCSC, oryCun2), *Marmota marmota* (NCBI, GCF_001458135.1_marMar2.1), *Ictidomys tridecemlineatus* (UCSC, speTri2), *Cavia porcellus* (Broad Institute cavPor3).

Species divergence times

Divergence time estimates were obtained from using timetree.org (40), by specifying sister taxa that belong to either Glires, rodents or muroids. Timetree outputs a range of estimated divergence times summarizing phylogenetic and fossil dating.

Synteny analysis

Shared synteny blocks were identified using the online server Genomicus (V95.1) (16). Mouse was used as a reference locus and individual synteny blocks were inspected using the UCSC genome browser (56).

Alignments and phylogenies

All sequence alignments are available as supplementary material. Alignments were generated using ClustalW v2.1 (IUB cost matrix, (57)) or MAFFT v7.388 (58). Maximum likelihood phylogenies were built using PHyML v3.0 with 100 bootstraps (59). Trees were visualized using the software Geneious Prime (Biomatters Ltd). In all cases we used nucleotide alignments of the CDS and the HKY85 substitution model.

Detection of recombination

To test for recombination, we used an alignment of *Dnmt3C* and *3B* CDS from six species with nearly complete gene models (mouse, Mus caroli, rat, prairie vole, chinese hamster, and mountain blind mole rat). Assembly gaps were removed. To detect recombination breakpoints, we used GARD with the general discrete model of site to site variation and three rate classes (42). We kept breakpoints with right and left p-values below 0.01. We subsequently segmented the *Dnmt3C* alignment according to these breakpoints. Similarly, recombination in primate *DNMT3A* was tested using an alignment of all primate CDS.

Genomic alignments

To identify region of homology between *Dnmt3C* and *Dnmt3B* genomic loci, we extracted the regions from assembled genomes of the mouse and rat and contigs of mountain blind mole rat and aligned them using mVista (60). Exon annotations were based on reference alignments with the species CDS.

Selection analyses

We measured overall dN/dS rates with codeml, PAMLX V1.3.1 (43), under model 0 and average pairwise with SNAP V2.1.1 (61). We tested for positive selection using codon alignments generated with PAL2NAL (62) free of any gaps and stop codons and with either accepted species or gene phylogenies. We compared "NSsites" evolutionary models that either do not allow dN/dS to exceed 1 (M7 or M8a) to a model that does (M8). We tested for statistical significance using a chi-square test of the twice the difference in log-likelihoods between M8 and matched null model M7 or M8a, with the degrees of freedom reflecting the difference in number of parameters between the two models compared (43). Positively selected sites were classified as those sites with M8 Bayes Empirical Bayes posterior probability > 90%. The results we present are from codeml runs using the F3x4 codon frequency model, and initial omega 0.4. Analyses were robust to use of different starting parameters (codon frequency model F61; starting omega 1.5).

DNMT3C and DNMT3B logo plots

Logo plots were generated using weblogo.berkeley.edu (63); using all muroid species with alignable sequences over these exons: mouse (*Mus musculus*), *Mus spretus, Mus caroli*, rat, deer mouse, field vole, prairie vole, bank vole, chinese hamster, and mountain blind mole rat.

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Table1: Summary of selection tests across muroid *Dnmts* genes. Recombination segments of *Dnmt3C* were analyzed independently, while full-length sequences were used for other *Dnmts*. P-values are for likelihood ratio tests between substitution models allowing or not allowing for positive selection using codeml (PAML). See text and methods for details.

	Seg.	nb. species	Length (bp)	PAML - M7 vs M8 p-value	PAML - M8a vs M8 p-value	M(0) dN/dS	% sites dN/dS>1 (avg. dN/dS)	Sites BEB≥90%
Dnmt3C	**A**	11	471	0.002	0.004	0.886	49% (1.62)	54(T), 57(Q), 95(P), 96 (L)
	B	8	135	0.509	0.965	0.176	-	N/A
	C	8	546	1.000	0.827	0.114	-	N/A
	D	10	666	1.000	0.907	0.184	-	N/A
Dnmt3B	**all**	8	2052	0.170	0.475	0.116	1% (1.59)	N/A
Dnmt3A	**all**	9	2718	0.823	0.463	0.022	-	N/A
Dnmt3L	**all**	9	1218	0.546	0.732	0.271	-	N/A

Table2: Summary of selection tests across primate DNMTs. (A) Codeml (PAML) analyses using the accepted species phylogeny. P-values are for likelihood ratio tests between substitution models allowing or not allowing for positive selection.

	nb. species	Length (bp)	PAML - M7 vs M8 p-value	PAML - M8a vs M8 p-value	M(0) dN/dS	% sites dN/dS>1 (avg. dN/dS)	Sites BEB≥90%
DNMT3A	20	2724	0.022	≤ 0.001	0.047	2.02 (1.8)	66 (P), 81 (A)
DNMT3B	21	2481	0.149	0.430	0.080	1.81 (1.7)	N/A
DNMT3L	20	1155	0.018	0.124	0.204	5.29 (1.6)	N/A

Literature cited

1. Smith ZD, Meissner A. DNA methylation: roles in mammalian development. Nat Rev Genet. 2013;14(3):204-20. Epub 2013/02/13. doi: 10.1038/nrg3354. PubMed PMID: 23400093.
2. Lyko F. The DNA methyltransferase family: a versatile toolkit for epigenetic regulation. Nat Rev Genet. 2018;19(2):81-92. Epub 2017/10/17. doi: 10.1038/nrg.2017.80. PubMed PMID: 29033456.
3. Bestor TH. The DNA methyltransferases of mammals. Hum Mol Genet. 2000;9(16):2395-402. Epub 2000/09/27. doi: 10.1093/hmg/9.16.2395. PubMed PMID: 11005794.
4. Lees-Murdock DJ, McLoughlin GA, McDaid JR, Quinn LM, O'Doherty A, Hiripi L, Hack CJ, Walsh CP. Identification of 11 pseudogenes in the DNA methyltransferase gene family in rodents and humans and implications for the functional loci. Genomics. 2004;84(1):193-204. Epub 2004/06/19. doi: 10.1016/j.ygeno.2004.02.004. PubMed PMID: 15203217.
5. Ponger L, Li WH. Evolutionary diversification of DNA methyltransferases in eukaryotic genomes. Mol Biol Evol. 2005;22(4):1119-28. Epub 2005/02/04. doi: 10.1093/molbev/msi098. PubMed PMID: 15689527.
6. Bestor T, Laudano A, Mattaliano R, Ingram V. Cloning and sequencing of a cDNA encoding DNA methyltransferase of mouse cells. The carboxyl-terminal domain of the mammalian enzymes is related to bacterial restriction methyltransferases. J Mol Biol. 1988;203(4):971-83. Epub 1988/10/20. PubMed PMID: 3210246.
7. Gruenbaum Y, Cedar H, Razin A. Substrate and sequence specificity of a eukaryotic DNA methylase. Nature. 1982;295(5850):620-2. Epub 1982/02/18. doi: 10.1038/295620a0. PubMed PMID: 7057921.
8. Song J, Rechkoblit O, Bestor TH, Patel DJ. Structure of DNMT1-DNA complex reveals a role for autoinhibition in maintenance DNA methylation. Science. 2011;331(6020):1036-40. Epub 2010/12/18. doi: 10.1126/science.1195380. PubMed PMID: 21163962; PMCID: PMC4689315.
9. Jia D, Jurkowska RZ, Zhang X, Jeltsch A, Cheng X. Structure of Dnmt3a bound to Dnmt3L suggests a model for de novo DNA methylation. Nature. 2007;449(7159):248-51. Epub 2007/08/24. doi: 10.1038/nature06146. PubMed PMID: 17713477; PMCID: PMC2712830.
10. Okano M, Bell DW, Haber DA, Li E. DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development. Cell. 1999;99(3):247-57. Epub 1999/11/11. doi: 10.1016/s0092-8674(00)81656-6. PubMed PMID: 10555141.
11. Okano M, Xie S, Li E. Cloning and characterization of a family of novel mammalian DNA (cytosine-5) methyltransferases. Nat Genet. 1998;19(3):219-20. Epub 1998/07/14. doi: 10.1038/890. PubMed PMID: 9662389.
12. Zhang ZM, Lu R, Wang P, Yu Y, Chen D, Gao L, Liu S, Ji D, Rothbart SB, Wang Y, Wang GG, Song J. Structural basis for DNMT3A-mediated de novo DNA methylation. Nature.

2018;554(7692):387-91. Epub 2018/02/08. doi: 10.1038/nature25477. PubMed PMID: 29414941; PMCID: PMC5814352.
13. Bourc'his D, Xu GL, Lin CS, Bollman B, Bestor TH. Dnmt3L and the establishment of maternal genomic imprints. Science. 2001;294(5551):2536-9. Epub 2001/11/24. doi: 10.1126/science.1065848. PubMed PMID: 11719692.
14. Li E, Bestor TH, Jaenisch R. Targeted mutation of the DNA methyltransferase gene results in embryonic lethality. Cell. 1992;69(6):915-26. Epub 1992/06/12. doi: 10.1016/0092-8674(92)90611-f. PubMed PMID: 1606615.
15. Law JA, Jacobsen SE. Establishing, maintaining and modifying DNA methylation patterns in plants and animals. Nat Rev Genet. 2010;11(3):204-20. Epub 2010/02/10. doi: 10.1038/nrg2719. PubMed PMID: 20142834; PMCID: PMC3034103.
16. Nguyen NTT, Vincens P, Roest Crollius H, Louis A. Genomicus 2018: karyotype evolutionary trees and on-the-fly synteny computing. Nucleic Acids Res. 2018;46(D1):D816-D22. Epub 2017/11/01. doi: 10.1093/nar/gkx1003. PubMed PMID: 29087490; PMCID: PMC5753199.
17. Alvarez-Ponce D, Torres-Sanchez M, Feyertag F, Kulkarni A, Nappi T. Molecular evolution of DNMT1 in vertebrates: Duplications in marsupials followed by positive selection. PLoS One. 2018;13(4):e0195162. Epub 2018/04/06. doi: 10.1371/journal.pone.0195162. PubMed PMID: 29621315; PMCID: PMC5886458.
18. Yokomine T, Hata K, Tsudzuki M, Sasaki H. Evolution of the vertebrate DNMT3 gene family: a possible link between existence of DNMT3L and genomic imprinting. Cytogenet Genome Res. 2006;113(1-4):75-80. Epub 2006/04/01. doi: 10.1159/000090817. PubMed PMID: 16575165.
19. Jain D, Meydan C, Lange J, Claeys Bouuaert C, Lailler N, Mason CE, Anderson KV, Keeney S. rahu is a mutant allele of Dnmt3c, encoding a DNA methyltransferase homolog required for meiosis and transposon repression in the mouse male germline. PLoS Genet. 2017;13(8):e1006964. doi: 10.1371/journal.pgen.1006964. PubMed PMID: 28854222; PMCID: 5607212.
20. Barau J, Teissandier A, Zamudio N, Roy S, Nalesso V, Herault Y, Guillou F, Bourc'his D. The DNA methyltransferase DNMT3C protects male germ cells from transposon activity. Science. 2016;354(6314):909-12. Epub 2016/11/20. doi: 10.1126/science.aah5143. PubMed PMID: 27856912.
21. Yoder JA, Walsh CP, Bestor TH. Cytosine methylation and the ecology of intragenomic parasites. Trends Genet. 1997;13(8):335-40. Epub 1997/08/01. PubMed PMID: 9260521.
22. Molaro A, Malik HS. Hide and seek: how chromatin-based pathways silence retroelements in the mammalian germline. Curr Opin Genet Dev. 2016;37:51-8. doi: 10.1016/j.gde.2015.12.001. PubMed PMID: 26821364; PMCID: PMC4914476.
23. Molaro A, Falciatori I, Hodges E, Aravin AA, Marran K, Rafii S, McCombie WR, Smith AD, Hannon GJ. Two waves of de novo methylation during mouse germ cell development. Genes Dev. 2014;28(14):1544-9. doi: 10.1101/gad.244350.114. PubMed PMID: 25030694; PMCID: PMC4102761.
24. Aravin AA, Sachidanandam R, Bourc'his D, Schaefer C, Pezic D, Toth KF, Bestor T, Hannon GJ. A piRNA pathway primed by individual transposons is linked to de novo

DNA methylation in mice. Mol Cell. 2008;31(6):785-99. Epub 2008/10/17. doi: 10.1016/j.molcel.2008.09.003. PubMed PMID: 18922463; PMCID: PMC2730041.

25. Manakov SA, Pezic D, Marinov GK, Pastor WA, Sachidanandam R, Aravin AA. MIWI2 and MILI Have Differential Effects on piRNA Biogenesis and DNA Methylation. Cell Rep. 2015;12(8):1234-43. Epub 2015/08/19. doi: 10.1016/j.celrep.2015.07.036. PubMed PMID: 26279574; PMCID: PMC4554733.
26. Kaneda M, Okano M, Hata K, Sado T, Tsujimoto N, Li E, Sasaki H. Essential role for de novo DNA methyltransferase Dnmt3a in paternal and maternal imprinting. Nature. 2004;429(6994):900-3. Epub 2004/06/25. doi: 10.1038/nature02633. PubMed PMID: 15215868.
27. Kato Y, Kaneda M, Hata K, Kumaki K, Hisano M, Kohara Y, Okano M, Li E, Nozaki M, Sasaki H. Role of the Dnmt3 family in de novo methylation of imprinted and repetitive sequences during male germ cell development in the mouse. Hum Mol Genet. 2007;16(19):2272-80. Epub 2007/07/10. doi: 10.1093/hmg/ddm179. PubMed PMID: 17616512.
28. Posfai J, Bhagwat AS, Posfai G, Roberts RJ. Predictive motifs derived from cytosine methyltransferases. Nucleic Acids Res. 1989;17(7):2421-35. Epub 1989/04/11. doi: 10.1093/nar/17.7.2421. PubMed PMID: 2717398; PMCID: PMC317633.
29. Timinskas A, Butkus V, Janulaitis A. Sequence motifs characteristic for DNA [cytosine-N4] and DNA [adenine-N6] methyltransferases. Classification of all DNA methyltransferases. Gene. 1995;157(1-2):3-11. Epub 1995/05/19. doi: 10.1016/0378-1119(94)00783-o. PubMed PMID: 7607512.
30. Jeltsch A, Broche J, Bashtrykov P. Molecular Processes Connecting DNA Methylation Patterns with DNA Methyltransferases and Histone Modifications in Mammalian Genomes. Genes (Basel). 2018;9(11). Epub 2018/11/25. doi: 10.3390/genes9110566. PubMed PMID: 30469440; PMCID: PMC6266221.
31. Guo X, Wang L, Li J, Ding Z, Xiao J, Yin X, He S, Shi P, Dong L, Li G, Tian C, Wang J, Cong Y, Xu Y. Structural insight into autoinhibition and histone H3-induced activation of DNMT3A. Nature. 2015;517(7536):640-4. Epub 2014/11/11. doi: 10.1038/nature13899. PubMed PMID: 25383530.
32. Ooi SK, Qiu C, Bernstein E, Li K, Jia D, Yang Z, Erdjument-Bromage H, Tempst P, Lin SP, Allis CD, Cheng X, Bestor TH. DNMT3L connects unmethylated lysine 4 of histone H3 to de novo methylation of DNA. Nature. 2007;448(7154):714-7. Epub 2007/08/10. doi: 10.1038/nature05987. PubMed PMID: 17687327; PMCID: PMC2650820.
33. Otani J, Nankumo T, Arita K, Inamoto S, Ariyoshi M, Shirakawa M. Structural basis for recognition of H3K4 methylation status by the DNA methyltransferase 3A ATRX-DNMT3-DNMT3L domain. EMBO Rep. 2009;10(11):1235-41. Epub 2009/10/17. doi: 10.1038/embor.2009.218. PubMed PMID: 19834512; PMCID: PMC2775176.
34. Zhang Y, Jurkowska R, Soeroes S, Rajavelu A, Dhayalan A, Bock I, Rathert P, Brandt O, Reinhardt R, Fischle W, Jeltsch A. Chromatin methylation activity of Dnmt3a and Dnmt3a/3L is guided by interaction of the ADD domain with the histone H3 tail. Nucleic Acids Res. 2010;38(13):4246-53. Epub 2010/03/13. doi: 10.1093/nar/gkq147. PubMed PMID: 20223770; PMCID: PMC2910041.

35. Chen T, Tsujimoto N, Li E. The PWWP domain of Dnmt3a and Dnmt3b is required for directing DNA methylation to the major satellite repeats at pericentric heterochromatin. Mol Cell Biol. 2004;24(20):9048-58. Epub 2004/10/01. doi: 10.1128/MCB.24.20.9048-9058.2004. PubMed PMID: 15456878; PMCID: PMC517890.
36. Dhayalan A, Rajavelu A, Rathert P, Tamas R, Jurkowska RZ, Ragozin S, Jeltsch A. The Dnmt3a PWWP domain reads histone 3 lysine 36 trimethylation and guides DNA methylation. J Biol Chem. 2010;285(34):26114-20. Epub 2010/06/16. doi: 10.1074/jbc.M109.089433. PubMed PMID: 20547484; PMCID: PMC2924014.
37. Ge YZ, Pu MT, Gowher H, Wu HP, Ding JP, Jeltsch A, Xu GL. Chromatin targeting of de novo DNA methyltransferases by the PWWP domain. J Biol Chem. 2004;279(24):25447-54. Epub 2004/03/05. doi: 10.1074/jbc.M312296200. PubMed PMID: 14998998.
38. Qiu C, Sawada K, Zhang X, Cheng X. The PWWP domain of mammalian DNA methyltransferase Dnmt3b defines a new family of DNA-binding folds. Nat Struct Biol. 2002;9(3):217-24. Epub 2002/02/12. doi: 10.1038/nsb759. PubMed PMID: 11836534; PMCID: PMC4035047.
39. Rondelet G, Dal Maso T, Willems L, Wouters J. Structural basis for recognition of histone H3K36me3 nucleosome by human de novo DNA methyltransferases 3A and 3B. J Struct Biol. 2016;194(3):357-67. Epub 2016/03/20. doi: 10.1016/j.jsb.2016.03.013. PubMed PMID: 26993463.
40. Hedges SB, Marin J, Suleski M, Paymer M, Kumar S. Tree of life reveals clock-like speciation and diversification. Mol Biol Evol. 2015;32(4):835-45. doi: 10.1093/molbev/msv037. PubMed PMID: 25739733; PMCID: PMC4379413.
41. Steppan SJ, Schenk JJ. Muroid rodent phylogenetics: 900-species tree reveals increasing diversification rates. PLoS One. 2017;12(8):e0183070. Epub 2017/08/17. doi: 10.1371/journal.pone.0183070. PubMed PMID: 28813483; PMCID: PMC5559066.
42. Kosakovsky Pond SL, Posada D, Gravenor MB, Woelk CH, Frost SD. GARD: a genetic algorithm for recombination detection. Bioinformatics. 2006;22(24):3096-8. doi: 10.1093/bioinformatics/btl474. PubMed PMID: 17110367.
43. Yang Z. PAML: a program package for phylogenetic analysis by maximum likelihood. Comput Appl Biosci. 1997;13(5):555-6. PubMed PMID: 9367129.
44. Daugherty MD, Zanders SE. Gene conversion generates evolutionary novelty that fuels genetic conflicts. Curr Opin Genet Dev. 2019;58-59:49-54. Epub 2019/08/30. doi: 10.1016/j.gde.2019.07.011. PubMed PMID: 31466040.
45. Daugherty MD, Schaller AM, Geballe AP, Malik HS. Evolution-guided functional analyses reveal diverse antiviral specificities encoded by IFIT1 genes in mammals. Elife. 2016;5. Epub 2016/06/01. doi: 10.7554/eLife.14228. PubMed PMID: 27240734; PMCID: PMC4887208.
46. Kursel LE, Malik HS. Recurrent Gene Duplication Leads to Diverse Repertoires of Centromeric Histones in Drosophila Species. Mol Biol Evol. 2017;34(6):1445-62. Epub 2017/03/24. doi: 10.1093/molbev/msx091. PubMed PMID: 28333217; PMCID: PMC5435080.
47. Thomas JH, Schneider S. Coevolution of retroelements and tandem zinc finger genes. Genome Res. 2011;21(11):1800-12. Epub 2011/07/26. doi: 10.1101/gr.121749.111. PubMed PMID: 21784874; PMCID: 3205565.

48. Chen T, Ueda Y, Xie S, Li E. A novel Dnmt3a isoform produced from an alternative promoter localizes to euchromatin and its expression correlates with active de novo methylation. J Biol Chem. 2002;277(41):38746-54. Epub 2002/07/26. doi: 10.1074/jbc.M205312200. PubMed PMID: 12138111.
49. Simkin A, Wong A, Poh YP, Theurkauf WE, Jensen JD. Recurrent and recent selective sweeps in the piRNA pathway. Evolution. 2013;67(4):1081-90. Epub 2013/04/05. doi: 10.1111/evo.12011. PubMed PMID: 23550757; PMCID: PMC3992950.
50. Yi M, Chen F, Luo M, Cheng Y, Zhao H, Cheng H, Zhou R. Rapid evolution of piRNA pathway in the teleost fish: implication for an adaptation to transposon diversity. Genome Biol Evol. 2014;6(6):1393-407. Epub 2014/05/23. doi: 10.1093/gbe/evu105. PubMed PMID: 24846630; PMCID: PMC4079211.
51. Baubec T, Colombo DF, Wirbelauer C, Schmidt J, Burger L, Krebs AR, Akalin A, Schubeler D. Genomic profiling of DNA methyltransferases reveals a role for DNMT3B in genic methylation. Nature. 2015;520(7546):243-7. Epub 2015/01/22. doi: 10.1038/nature14176. PubMed PMID: 25607372.
52. Weinberg DN, Papillon-Cavanagh S, Chen H, Yue Y, Chen X, Rajagopalan KN, Horth C, McGuire JT, Xu X, Nikbakht H, Lemiesz AE, Marchione DM, Marunde MR, Meiners MJ, Cheek MA, Keogh MC, Bareke E, Djedid A, Harutyunyan AS, Jabado N, Garcia BA, Li H, Allis CD, Majewski J, Lu C. The histone mark H3K36me2 recruits DNMT3A and shapes the intergenic DNA methylation landscape. Nature. 2019;573(7773):281-6. Epub 2019/09/06. doi: 10.1038/s41586-019-1534-3. PubMed PMID: 31485078; PMCID: PMC6742567.
53. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. J Mol Biol. 1990;215(3):403-10. doi: 10.1016/S0022-2836(05)80360-2. PubMed PMID: 2231712.
54. Coordinators NR. Database resources of the National Center for Biotechnology Information. Nucleic Acids Res. 2016;44(D1):D7-19. doi: 10.1093/nar/gkv1290. PubMed PMID: 26615191; PMCID: 4702911.
55. Birney E, Clamp M, Durbin R. GeneWise and Genomewise. Genome Res. 2004;14(5):988-95. Epub 2004/05/05. doi: 10.1101/gr.1865504. PubMed PMID: 15123596; PMCID: PMC479130.
56. Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM, Haussler D. The human genome browser at UCSC. Genome Res. 2002;12(6):996-1006. Epub 2002/06/05. doi: 10.1101/gr.229102. PubMed PMID: 12045153; PMCID: PMC186604.
57. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG. Clustal W and Clustal X version 2.0. Bioinformatics. 2007;23(21):2947-8. Epub 2007/09/12. doi: 10.1093/bioinformatics/btm404. PubMed PMID: 17846036.
58. Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 2013;30(4):772-80. Epub 2013/01/19. doi: 10.1093/molbev/mst010. PubMed PMID: 23329690; PMCID: PMC3603318.
59. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance

of PhyML 3.0. Syst Biol. 2010;59(3):307-21. doi: 10.1093/sysbio/syq010. PubMed PMID: 20525638.

60. Frazer KA, Pachter L, Poliakov A, Rubin EM, Dubchak I. VISTA: computational tools for comparative genomics. Nucleic Acids Res. 2004;32(Web Server issue):W273-9. doi: 10.1093/nar/gkh458. PubMed PMID: 15215394; PMCID: PMC441596.

61. Korber B, Muldoon M, Theiler J, Gao F, Gupta R, Lapedes A, Hahn BH, Wolinsky S, Bhattacharya T. Timing the ancestor of the HIV-1 pandemic strains. Science. 2000;288(5472):1789-96. Epub 2000/06/10. doi: 10.1126/science.288.5472.1789. PubMed PMID: 10846155.

62. Suyama M, Torrents D, Bork P. PAL2NAL: robust conversion of protein sequence alignments into the corresponding codon alignments. Nucleic Acids Res. 2006;34(Web Server issue):W609-12. doi: 10.1093/nar/gkl315. PubMed PMID: 16845082; PMCID: PMC1538804.

63. Crooks GE, Hon G, Chandonia JM, Brenner SE. WebLogo: a sequence logo generator. Genome Res. 2004;14(6):1188-90. doi: 10.1101/gr.849004. PubMed PMID: 15173120; PMCID: PMC419797.

Figure legends

Figure 1: ***Dnmt3C* duplicated in the last common ancestor of all muroids.** (A) Schematic of the genomic locus encoding *Dnmt3* genes in representative species of Glires. Estimated divergence times based on Timetree analyses (40) are indicated at each major node. *Dnmt3* genes (colors) and corresponding neighboring genes (grey) are indicated with arrows. "?" denotes incomplete assembly while crosses denote absence of coding sequence. (B) Maximum likelihood nucleotide phylogeny of *Dnmt3* genes in Glires. Bootstrap values and average pairwise dN/dS are indicated for each clade. (C) Maximum likelihood phylogeny of all identified *Dnmt3B* and *Dnmt3C* genes. Incomplete sequences are indicated with "inc" while cases where *Dnmt3B* and *Dnmt3C* orthologs from the same species unexpectedly group together are highlighted with '*'. Bootstrap support values above 50% are reported. Also included are rabbit, marmot and squirrel *Dnmt3Bs* and selected *Dnmt3A*s. Abbreviations of species names: "M.B.mole.rat": mountain blind mole rat; "D.mouse": deer mouse; "C.hamster": chineese hamster; "F.vole": field vole; "B.vole": bank vole; "P.vole": prairie vole.

Figure 2: **Gene conversion between *Dnmt3C* and *Dnmt3B* in muroids.** (A) Schematic representation of *Dnmt3C* and *3B* genomic loci (top). Arrows indicate gene orientation in the mm10 assembly. Recombination map of *Dnmt3C* CDS is shown at the bottom. Breakpoints (arrowheads) identified by GARD (see methods) and recombination segments ("A" through "D") are indicated on the *Dnmt3C* CDS with its predicted protein domains. (B) Maximum likelihood phylogenies of recombination segments A and B. Bootstrap supports above 50% are shown. (C) Average pairwise percent identities between muroid DNMT3C and DNMT3B proteins before and after the PWWP domain. (D) Map of the mouse *Dnmt3B* genomic locus with annotated exons (boxes). Percent nucleotide identity with *Dnmt3C* is plotted (y-axis) using mVISTA (see methods). Recombination segments (top) and predicted domains (bottom) are also shown.

Figure 3: DNMT3C N-terminus is subject to positive selection. (A) Schematic representation of positive selection test results for all recombination segments of DNMT3C CDS. "***" denotes the finding of positive selection in segment A (see text and methods for details), whereas "∅" indicates no support for positive selection. (B) Amino acid alignments (positions 50 to 61, and 91 to 100) of muroid DNMT3Cs showing four positively selected sites identified with PAML (red arrowheads). Sequences are arranged according to segment A phylogeny with species names on the right. (C) Amino acid logos of DNMT3C and 3B around the positively selected sites

(arrowheads). Backslashes indicates sequences not shown. The grey box denotes an alignment gap between DNMT3C and 3B.

Figure 4. Positive selection of N-terminal domain of primate DNMT3A. Amino acid alignments (position 58 to 90) of primate DNMT3As showing the two positively selected sites identified with PAML (arrowheads). Sequences are arranged according to the accepted species phylogeny with species names on the right.

Supplementary Figure S1: (A) Maximum likelihood nucleotide phylogeny of recombination segment C. (B) Maximum likelihood nucleotide phylogeny of recombination segment D. Bootstrap support is shown for each node. Scale bars show nucleotide substitutions per site. (C) Map of the rat *Dnmt3B* genomic locus with annotated exons (boxes). Percent nucleotide identity with *Dnmt3C* is plotted (y-axis). Predicted domains are shown at the bottom. (D) as in (C) but for mountain blind mole rat sequences.

Supplementary Figure S2: Maximum likelihood nucleotide phylogeny of putative recombination segment A (blue, 0 to 1041bp) and B (red, 1042 to 2727bp) of primate DNMT3A found with GARD (see methods). Bootstrap support is shown for each node. Nodes with low bootstrap support are shown in red. Scale bars show nucleotide substitutions per site.

Supplementary Files 1 to 8: Alignments of *Dnmt* sequences used in this study.

Figure 1

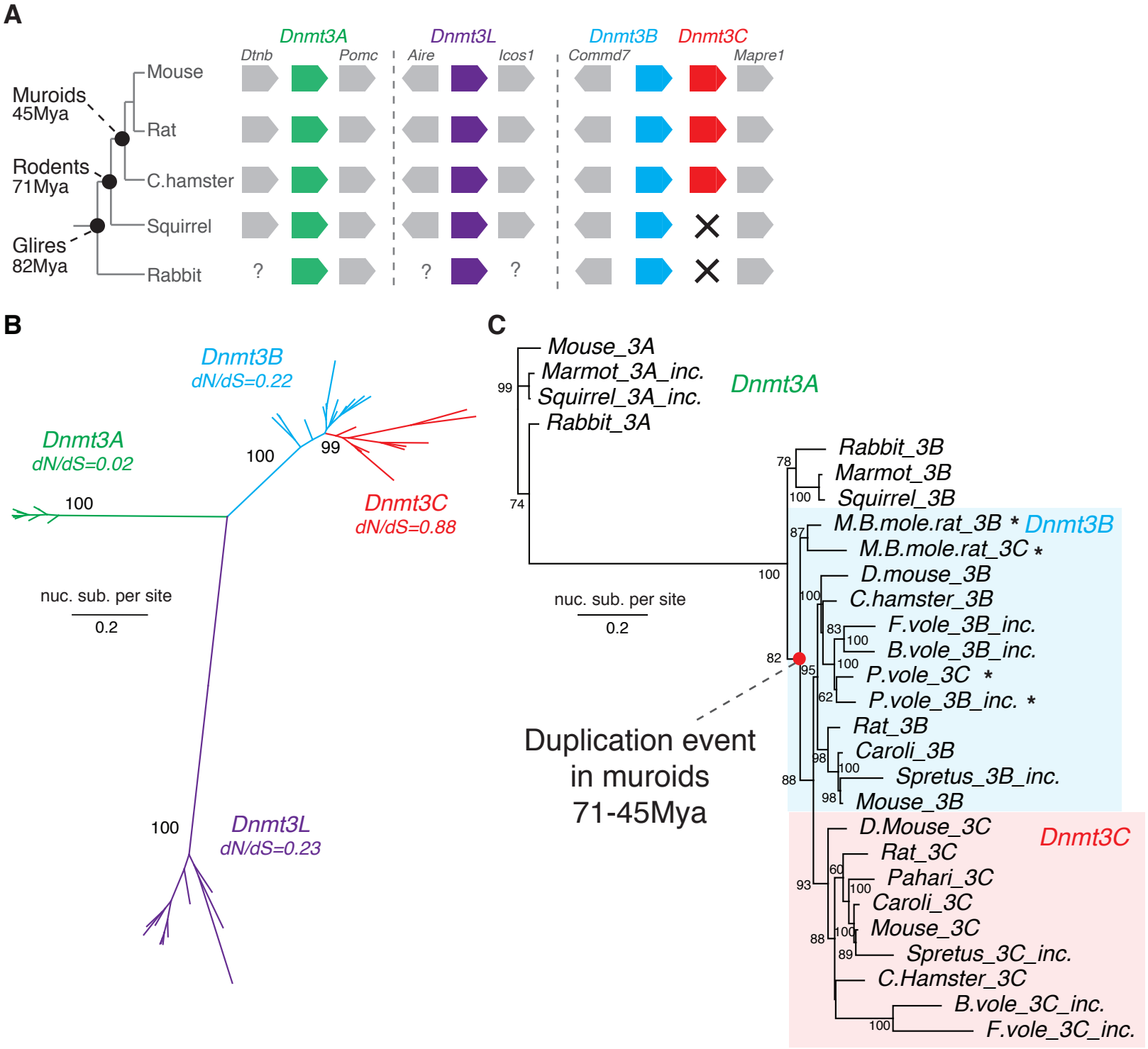
A
Dnmt3A
Dnmt3L
Dnmt3B
Dnmt3C
Dtnb
Pomc
Aire
Icos1
Commd7
Mapre1
Muroids
45Mya
Rodents
71Mya
Glires
82Mya
Mouse
Rat
C.hamster
Squirrel
Rabbit
B
Dnmt3B
dN/dS=0.22
Dnmt3A
dN/dS=0.02
Dnmt3C
dN/dS=0.88
Dnmt3L
dN/dS=0.23
nuc. sub. per site
0.2
C
Mouse_3A
Marmot_3A_inc.
Squirrel_3A_inc.
Rabbit_3A
Dnmt3A
Rabbit_3B
Marmot_3B
Squirrel_3B
M.B.mole.rat_3B *
M.B.mole.rat_3C *
Dnmt3B
D.mouse_3B
C.hamster_3B
F.vole_3B_inc.
B.vole_3B_inc.
P.vole_3C *
P.vole_3B_inc. *
Rat_3B
Caroli_3B
Spretus_3B_inc.
Mouse_3B
D.Mouse_3C
Rat_3C
Pahari_3C
Caroli_3C
Mouse_3C
Spretus_3C_inc.
C.Hamster_3C
B.vole_3C_inc.
F.vole_3C_inc.
Dnmt3C
Duplication event
in muroids
71-45Mya
nuc. sub. per site
0.2

Figure 2

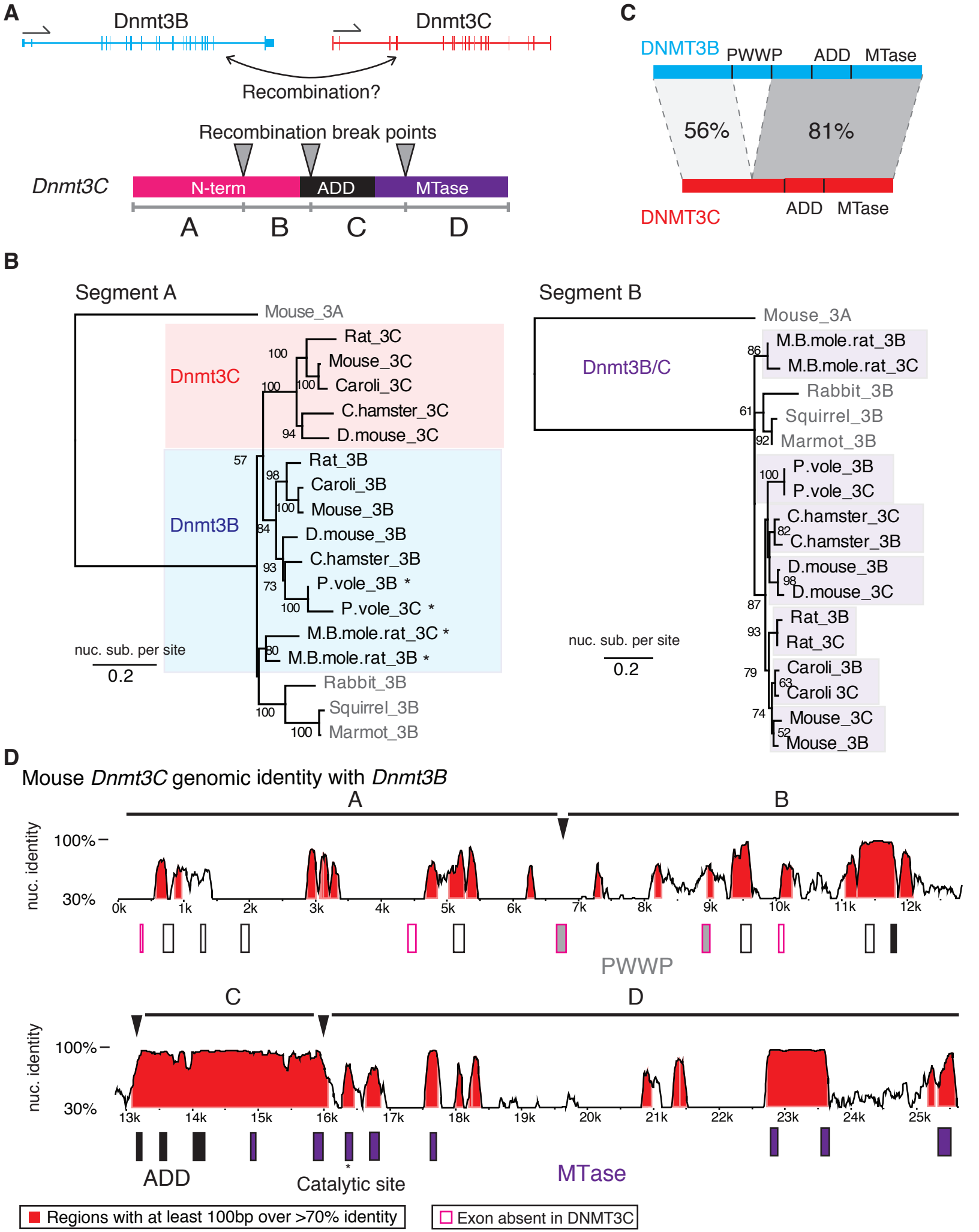
A
Dnmt3B
Dnmt3C
Recombination?
Recombination break points
Dnmt3C
N-term
ADD
MTase
A
B
C
D
C
DNMT3B
PWWP
ADD
MTase
56%
81%
DNMT3C
ADD
MTase
B
Segment A
Mouse_3A
Dnmt3C
Rat_3C
Mouse_3C
Caroli_3C
C.hamster_3C
D.mouse_3C
Dnmt3B
Rat_3B
Caroli_3B
Mouse_3B
D.mouse_3B
C.hamster_3B
P.vole_3B *
P.vole_3C *
M.B.mole.rat_3C *
M.B.mole.rat_3B *
Rabbit_3B
Squirrel_3B
Marmot_3B
nuc. sub. per site
0.2
Segment B
Mouse_3A
Dnmt3B/C
M.B.mole.rat_3B
M.B.mole.rat_3C
Rabbit_3B
Squirrel_3B
Marmot_3B
P.vole_3B
P.vole_3C
C.hamster_3C
C.hamster_3B
D.mouse_3B
D.mouse_3C
Rat_3B
Rat_3C
Caroli_3B
Caroli 3C
Mouse_3C
Mouse_3B
nuc. sub. per site
0.2
D
Mouse Dnmt3C genomic identity with Dnmt3B
A
B
nuc. identity
100%
30%
0k
1k
2k
3k
4k
5k
6k
7k
8k
9k
10k
11k
12k
PWWP
C
D
nuc. identity
100%
30%
13k
14k
15k
16k
17k
18k
19k
20k
21k
22k
23k
24k
25k
ADD
Catalytic site
MTase
Regions with at least 100bp over >70% identity
Exon absent in DNMT3C

Figure 3

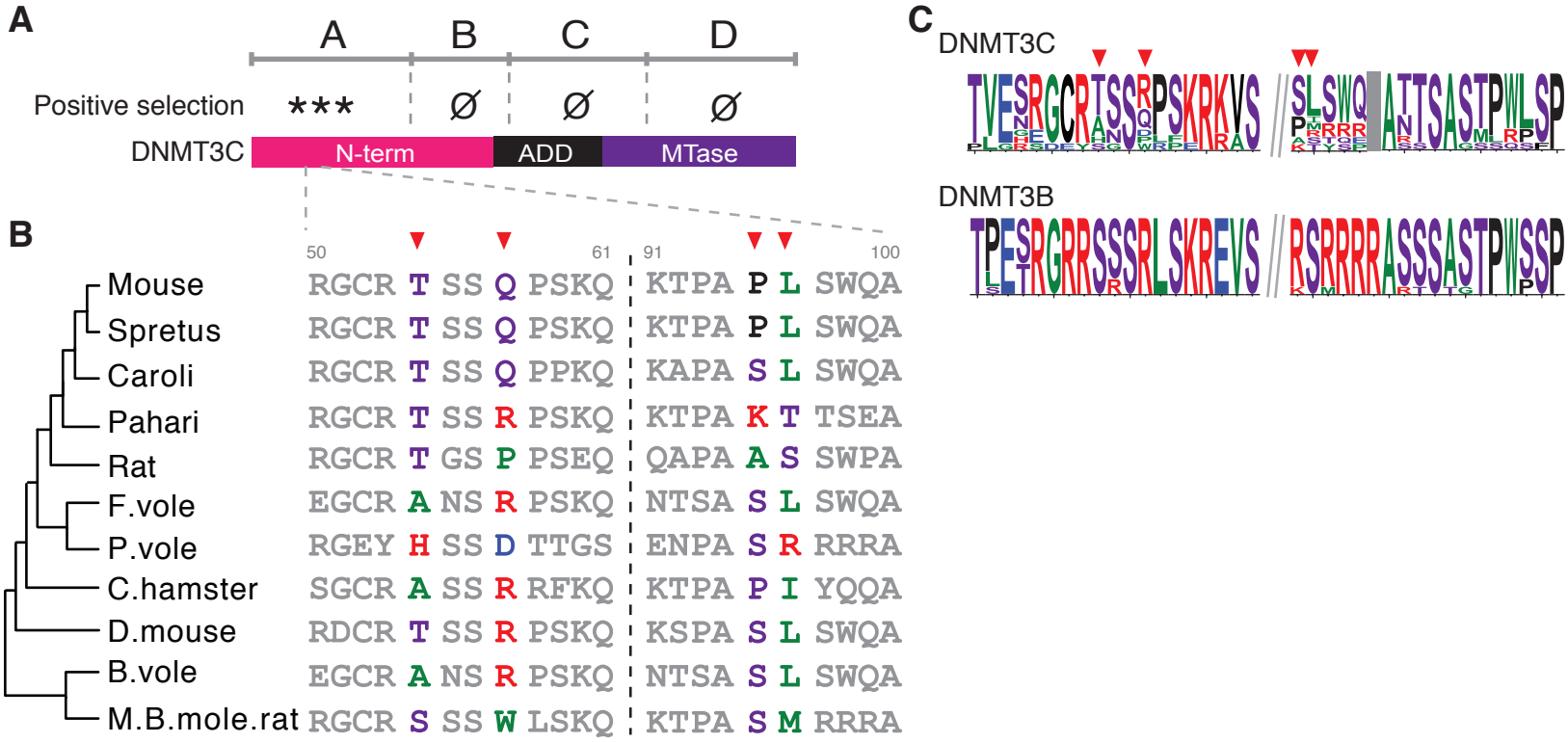
A
A
B
C
D
Positive selection
***
Ø
Ø
Ø
DNMT3C
N-term
ADD
MTase
B
50
61
91
100
Mouse RGCR T SS Q PSKQ KTPA P L SWQA
Spretus RGCR T SS Q PSKQ KTPA P L SWQA
Caroli RGCR T SS Q PPKQ KAPA S L SWQA
Pahari RGCR T SS R PSKQ KTPA K T TSEA
Rat RGCR T GS P PSEQ QAPA A S SWPA
F.vole EGCR A NS R PSKQ NTSA S L SWQA
P.vole RGEY H SS D TTGS ENPA S R RRRA
C.hamster SGCR A SS R RFKQ KTPA P I YQQA
D.mouse RDCR T SS R PSKQ KSPA S L SWQA
B.vole EGCR A NS R PSKQ NTSA S L SWQA
M.B.mole.rat RGCR S SS W LSKQ KTPA S M RRRA
C
DNMT3C
DNMT3B

Figure 4

DNMT3A
PWWP
ADD
MTase
58
90
Bonobo ESGDTPKD P AVISKSPSMAQDPG A SELLPNGDL
Chimp ESGDTPKD P AVISKSPSMAQDPG A SELLPNGDL
Human ESGDTPKD P AVISKSPSMAQDSG A SELLPNGDL
Gorilla ESGDTPKD P AVISKSPSMAQDSG A SELLPNGDL
Orangutan ESSDTPKD P AVISKSPSMAQDSG S SELLPNGDL
Rhesus macaque ESSDTPKD P AVTSKSPSMAQDSG S SELLPNGDL
Crab-eating macaque ESSDTPKD P AVTSKSPSMAQDSG S SELLPNGDL
Pig-tailed macaque ESSDTPKD P AVTSKSPSMAQDSG S SELLPNGDL
Mangabey ESSDTPKD P AVTSKSPSMAQDSG S SELLPNGDL
Drill ESSDTPKD P AVTSKSPSMAQDSG S SELLPNGDL
African green monkey ESSDTPKD P AVTSKSPSMAQDSG P SELLPNGDL
Black snub nose monkey ESSDTPKD P AVTSKSPSMAQDSG P SELLPNGDL
Red colobus monkey ESSDTPKD P AVTSKSPSMAQDSG P SELLPNGDL
Colobus monkey ESSDTPKD S VVTSKSPSMAQDSG P SELLPNGDL
Marmoset ESSDTPKD P AVTSKTPSMAQDSG P SELLPNGDL
Squirrel monkey ESSDTPKD P AVTSKSPSMAQDSG P SELLPNGDL
Sapajou ESSDTPKD A AVTSKSPSMAQDSG P SELLPNGDL
Sifaka ESSDTPKD T AVTSKSPSTAQDSG P SELLPNGDL
Gray mouse lemur ESSDTPKD T AVTSKSPSTAQDSG C SELLPNGDL
Bushbaby ESSDTPKD P AVTSKTPSTAQDSG P SELLPNGDL