*RESEARCH ARTICLE [5000 words]*

**Breeding at higher latitude as measured by stable isotope is associated with higher**

**photoperiod threshold and delayed reproductive development in a songbird**

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**Running title:** Latitudinal cline in seasonal reproduction

**Key words:** Dark eyed Junco, seasonal reproduction, critical photoperiod, Life-history states,

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**Abstract**

Many organisms time reproduction to photoperiod, a constant from year to year. Predicting how

anthropogenic change will influence future timing demands greater knowledge of the current role

of photoperiod. We held two closely related bird populations in a common environment. One

population is resident; the other winters in sympatry with the resident population but migrates

north prior to reproducing. We increased photoperiod gradually and measured preparation for

migration and reproduction, using feather stable isotopes to estimate breeding latitude. We

predicted population differences in the minimum stimulatory day length to elicit a response

(CPP, critical photoperiod) and co-variation between CPP and distance migrated. We found clear

population differences in CPP and greater CPP in longer distance migrants. We conclude that

current geographic variation in reproductive timing has a genetic or early developmental basis

and recommend that future research focus on how anthropogenic changes will interact with CPP

to adjust timing of reproduction and migration.

**Introduction**

Animals across the globe follow the seasons and match their growth, development, gonadal

recrudescence, migration, and other seasonal life-history states to exploit the seasons most

favorable for survival and reproduction (Wingfield et al., 1992; Dawson 2013). Birds breeding at

different latitudes vary in duration and timing of seasonal life-history states to match breeding to

periods when resources well-suited for nesting growth are abundant (Lack 1968; Visser at al.,

2004). Different species depend on food supplies available at different times of the year, hence

optimal timing varies by species and populations within species (Dawson and Goldsmith 1983;

Wingfield et al., 1992; Dawson et al., 2001; Ball and Ketterson 2008; Watts et al., 2015). When

there is a mismatch between food availability and the timing of breeding, nestling growth and

survival can be compromised (Visser at al., 2004; Jonzén et al., 2006).

Individuals must prepare in advance to time their seasonal events to match the environment

they occupy (Menaker 1971; Bradshaw and Holzapfel 2007). Photoperiod is the only consistent

reliable cue for seasonally breeding animals and is predictable at a given latitude. Hence,

changing photoperiod (i.e., day length) acts as the primary predictive cue to time seasonal

phenological events such as migration and breeding (Rowan, 1926; Wingfield et al., 1992;

Bronson and Heideman, 1994; Dawson 2001). In general, rate of gonadal maturation appears to

be directly proportional to increasing day length (Farner and Wilson, 1957; Follett and Maung,

1978). Photoperiodic responses depend on encephalic photoreceptors perceiving light during the

stimulatory phase of a daily rhythm of sensitivity (Follett et al., 1992; Ball and Balthazart, 2003;

Yasuo et al., 2003). Seasonally breeding animals that undergo annual gonadal recrudescence and

regression in response to changing photoperiod as a primary predictive cue, also rely on

supplementary cues to initiate and regulate timing of reproductive development (Bronson and

Heideman, 1994; Dawson 2001; Wingfield 2012). Towards the end of the breeding season, many

bird species are no longer responsive to long days and are said to become photorefractory. They

show a decline in gonad volume and reduced testosterone while days are still long, well before

the return of short photoperiods during autumn (Burger, 1949; Miller, 1954). Exposure to short

days during autumn is then required to break the photorefractory period and restore a bird's

ability to undergo gonadal recrudescence in response to increasing photoperiod the following

spring (Farner and Mewaldt, 1955). In short, seasonal phenology can be referred to in terms of

the periodic appearance of life-history states consisting of a photosensitive state capable of

responding to increasing photoperiod when encountered, a photostimulatory state that is induced

by increasing day length, and a photorefractory state in which an animal is no longer responsive

to long days.

Reproductive timing is driven by the hypothalamic-pituitary-gonadal (HPG) axis.

Gonadotropin releasing hormone 1 (GnRH1) is released from the hypothalamus to stimulate

release of the gonadotropins, luteinizing hormone (LH) and follicle-stimulating hormone (FSH),

from the pituitary (Li et al., 1994; Cho et al., 1998). LH and FSH stimulate gonadal growth and

development of gametes, as well as production and release of sex steroids. Injecting controlled

doses of exogenous GnRH (i.e., a GnRH challenge) to individuals and measuring downstream

activity of the HPG axis has been a successful tool to investigate variation in animals'

physiological state and behavior (Jawor et al., 2006; Spinney et al., 2006; Grieves et al., 2016).

While much of this has been known for decades, significant knowledge gaps remain with

respect to the specific mechanisms accounting for timing differences among populations that

breed in different environments (Fudickar et al., 2016; Ramenofsky et al., 2017). We studied

dark-eyed juncos (*Junco hyemalis),* a small songbird that consists of migratory and sedentary

(i.e., resident) populations, some of which live in sympatry during the winter and early spring

(Fudickar et al., 2016; Grieves et al., 2016). Residents initiate preparation for reproduction prior

to the departure of migrants for their breeding grounds. Following spring migration, migrant and

resident juncos are geographically isolated for the remainder of the breeding season.  Hence

migrants and residents can be exposed to the same environment in spring but different

environments during summer.

In a prior study, resident and migrant male juncos were held captive in a common garden

and exposed to the same photoperiod programmed to match the natural increase in spring.

Residents were found to increase cloacal protuberance volume (CPV; a primary sperm storage

structure for male birds) earlier than migrants, i.e. at a shorter photoperiod (Fudickar et al.,

2016). Here, we extend this study to examine changes in the reproductive axis during all four life

history states and report differences in the critical photoperiodic threshold (CPP) in spring, as

well as differences in the timing of breeding termination and attaining refractoriness. We

predicted that migrants and residents held in a captive common environment in gradually

increasing photoperiod would differ in the photoperiod at which cloacal protuberance (CPV),

baseline testosterone ($T_0$), and testosterone in response to GnRH challenge (dT) would increase

in spring, with the CPP being lower in residents. We also predicted that residents would enter the

photorefractory state later than migrants, thus prolonging the time when CPV, $T_0$, dT were

elevated. We used stable hydrogen isotopes ratios ($\delta^2$H) in feathers to estimate breeding latitude,

which has been used as a proxy for determining variation in locations where feathers are grown

(Rubenstein et al., 2002; Hobson 2003).

If our predictions were supported, we would conclude that population level variation in

CPP and thus in timing has a genetic or early developmental basis, which would permit further

investigation of the locus of variation in brain, gonad or periphery. If our predictions were not

borne out, i.e., migrants and residents did not differ in CPP in a common environment, this

would suggest that timing is highly flexible regardless of migratory strategy or population of

origin.

**Material and Methods**

*Study Species*

The dark-eyed junco (*Junco hyemalis*) is a broadly distributed North American songbird (Nolan

et al., 2002). Diversifying approximately 15,000 years ago, juncos subspecies vary in plumage

coloration, reproductive timing, and migratory behavior (Atwell et al., 2011; Fudickar et al.,

2016). Within this species complex, a migratory subspecies, *J.h. hyemalis,* (hereafter 'migrants'*)*

breeds in temperate coniferous and mixed forests across Canada and Alaska, whereas a sedentary

subspecies, *J.h. carolinensis*, (hereafter, 'residents') is found year-round in Appalachian

Mountains of the eastern United States. Following the fall migration, migrants overwinter in the

United States east of the Rocky Mountains. Some migrant and resident subpopulations are found

in overlapping distributions in the Appalachian Mountains during the winter; specifically, both

migrants and residents are frequently caught foraging in mixed flocks in the winter at Mountain

Lake Biological Station, in Pembroke, VA (Nolan, 2002).

*Bird Capture and Housing*

Between November 1 to December 5, 2017 male overwintering migratory dark-eyed juncos

(n=45) were captured using mist nests from their overwintering sites in Bloomington, IN (39.16

$^0$N, 86.52$^0$W). Additionally, sympatric resident (n=15) and migrants (n=15) male dark-eyed

juncos were captured at University of Virginia's Mountain Lake Biological Station in Giles

County (37.37 $^0$N, 80.52$^0$W). Scientific collecting permits were issued by the Virginia

Department of Game and Inland Fisheries (permit # 052971), the Indiana Department of Natural

Resources (permit# 1803), and the US Fish and Wildlife Service (permit # 20261). All methods

were approved under protocol (# 15-026-17) by the Indiana University Institutional Animal Care

and Use Committee.

Resident dark-eyed juncos are relatively bigger in body size, more uniformly colored, and

heavier in body mass than migrants except during migration, when migrants store fat (Pyle,

1997). We classified the subpopulations using plumage and bill coloration (pink bill, *J. h.*

*hyemalis*; blue-gray bill, *J. h. carolinesis:* Nolan 2002). Age was determined by looking

collectively at wing, plumage color, brown/black contrast in the iris of the eye and ossification of

skull (Nolan 2002; Cristol et al., 2003). Sex was determined by measuring wing length and

confirmed later by growth of cloacal protuberance. After capture, all the birds were transported

to Kent Farm Research Station in Bloomington, Indiana and housed in outdoor aviary under

natural day length, temperature and *ad libitum* food until December 15, 2017. On January 18,

2018, we moved all birds to individual cages (61 x 46 x 46 cm and 46 x 46 x 46 cm) with *ad*

*libitum* food and water. Migrants and residents were randomly distributed across three rooms for

four months. After four months the birds were free-flying until the endpoint sampling at 16L

photoperiod on July 31, 2018.

### *Feather Stable Hydrogen Isotopes*

The most distal secondary feather of the right wing was collected from each individual at the

time of capture for analysis of $\delta^2H$. After collection, feathers were cleaned, cut from most distal

end, weighed to approximately 0.5 mg, and placed into a 3 x 5-mm silver capsule, and mailed for

further quantification of $\delta^2H$ to the US Geological Survey Stable Isotope Lab in Denver, CO.

$\delta^2$H values were measured using established methods of mass spectrometry (Wunder et al., 2012;

Fudickar et al., 2016). The $\delta^2$H ratios were reported in parts per mil notation (‰) with respect to

VSMOW (Vienna Standard Mean Oceanic Water) using internal standards. We used North

American $\delta^2$H precipitation map for August (http://wateriso.utah.edu/waterisotopes/index.html)

for the schematic representation of junco_$\delta^2$H values (Fig. 1 a). The $\delta^2$H values were used as a

continuous variable against all the physiological and hormone measures. To analyze latitudinal

differences in the CPP for physiological responses, we created two subjective groups within

migrants for analysis: high latitude migrant (HLM; -141‰ to -90‰), low latitude migrants

(LLM; -88‰ to -30‰). The values for low latitude migrants were similar to those of residents

(Fig. 1 b).

***Experimental Design and Sample Collection***

In order to determine differences in gonadal recrudescence and migration-related physiological

changes between residents and migrants in response to increasing photoperiod, we artificially

regulated changes in day length. Photoperiod was increased every twelve days from January 18

to May 6 in the following schedule: 9L:15D, 10L:14D, 11L:13D, 11.4L:12.6D, 11.75L:12.25D,

12L:12D, 12.4L:11.6D, 12.75L:11.25D, 13L:11D, 15L:9D, 16L:8D. After May 6, day length

remained the same until the end of the experiment on July 31, 2018. Birds were processed for

physiological measurements and bled after experiencing three days in each photoperiod till 15L.

Under 16L of day length, all physiological measurements except bleeding were continued until

the birds regressed their CP after experiencing 40 days in 16 L. At the end of period at 16L day

length, all the birds were bled to measure $T_0$ and dT in photorefractory state.

*Morphological Measurements*

During each sampling, we measured other indicators for preparation for reproduction and

migration, including subcutaneous fat score (FS), cloacal protuberance volume (CPV) and body

mass (BM) (Fudickar et al., 2016; Greives et al., 2016). Cloacal protuberance volume (CPV) is

used as a measure of spermatogenesis, sperm storage and gonadal growth during the breeding

season in males (Wolfson 1952). Volume of the CP was estimated using the equation for the

volume of a cylinder, $V= \pi(radius)^2 Height$ (Schut et al., 2012). Postnuptial (pre-basic) molt was

scored at the end of the experiment based on primary, secondary, and head feathers in both the

populations. Each region was given a score from 1-10 depending on the extent of molting

feathers: no molt as 0 (0%), light molt (1-10%), moderate molt (11-50%) and heavy molt (51-

100%). The percentages were summed to generate a total molt score for each bird (modified

from Ramenofsky et al., 2017).

*Blood Sample Collection and Testosterone Hormone Assays*

Immediately after capturing a bird from its cage, we took a 100 µl of blood sample by puncturing

the alar wing vein for baseline testosterone ($T_0$). Birds then received an intrapectoral muscle

GnRH injection (~50 µl) (chicken GnRH, American peptide, Sunnyvale, CA) dissolved in PBS

vehicle, which is known to activate the HPG axis in juncos (Wingfield et al., 1979; Jawor et al.,

2006; Greives et al., 2016). Thirty minutes following the GnRH injection a second blood sample

(50 µl) was taken from the wing vein to measure GnRH-challenged testosterone ($T_{30}$) levels.

Birds were kept in an opaque bag between injections to reduce stress. After collection, blood

samples were immediately processed to extract plasma and stored at $- 20^0 C$ until assayed for

testosterone.

We determined $T_0$ and $T_{30}$ concentration from 20 µl plasma aliquots following established

methods for our species (Jawor et al., 2006; Fudickar et al., 2016), using high sensitivity

testosterone kits (Enzo Life Sciences, ADI-900-176, Farmingdale, NY) to determine circulating

levels of $T_0$ and $T_{30}$. The GnRH induced testosterone level (dT) was calculated by subtracting $T_0$

from $T_{30}$. All samples were measured in duplicate and randomized over forty plates. The intra-

plate and inter-plate coefficient of variation were 6.77% ± 2.07% (mean ± SE) and 13.7%

respectively.

*Statistical Analysis*

Data were analyzed using R (version 3.2.0). Differences in mean hydrogen isotope ratios ($\delta^2$H)

between migrants and residents were determined using an unpaired Student's t-test of population

means. We used a Box-cox test of transformation to determine the normal distribution of all the

response variables (i.e., $T_0$, dT, CPV, FS, and BW). We used a square root transformation for

CPV and FS, a logarithmic transformation for T0 and dT, and no transformation for BW. To

quantify whether day length, population, or the interaction between day length and population

had a significant association with response variables, we used two-way analysis of variance (2-

way ANOVA) followed by Tukey's post-hoc multiple comparison tests (alpha < 0.05).

Considering repeated measures for the same individuals, we used a generalized liner mixed-

effect model (GLMM) with day length and population as main effects, and age, $\delta^2$H as a

covariate to determine effect of treatment on physiological responses. To find the critical

photoperiod at which physiological parameters started to change, we used change point analysis

(CPA) package in R (Killick and Eckley 2014; Robart et al., 2018). We used change point mean

function which is based on the likelihood ratio and cumulative sum (CUSUM) test statistics. The

CUSUM distribution does not assume data to be normally distributed and specified a single

change point.

To assess co-variation between $\delta^2H$ values as a continuous variable and morphological

and hormonal measurements, we combined migrants and residents and performed Pearson

correlations for CPV, BW, $T_0$, dT, and molt score, and Spearman correlation for FS on one

sampling date for each of four life-history states (LHSs): (1) photosensitive (9L, defined as

beginning of experiment prior to recrudescence), (2) recrudescence (defined as the date of

change point for CPV and dT), (3) photostimulatory, (defined as the date of seasonal peak values

at 15L), and (4) photorefractory, (defined as the date of lowest seasonal value, 16L endpoint).

We also calculated these correlations for migrants only on these same dates.

**Results**

**Hydrogen Isotope Values for Migrants and Residents**

There was a large range in the individual $\delta^2H$ values in migrants (lowest $\delta^2H$ = -141‰, highest

$\delta^2H$ = -33‰) in comparison to resident juncos (lowest $\delta^2H$ = -81‰, highest $\delta^2H$ = -37‰;

except one outlier that had $\delta^2H$ = -109‰). Mean $\delta^2H$ differed significantly between resident and

migrant juncos (p < 0.0001; Student's t-test). Mean $\delta^2H$ isotope was significantly lower in

migrants than in residents (migrants mean $\delta^2H$ = -104.9‰, residents mean $\delta^2H$ = -58‰; Fig. 1

c).

**CPP for Gonadal Recrudescence in Migrants and Residents**

CPV varied significantly by day length ($F_{13,\ 663.38}$ = 66.3716, p < 0.0001), population ($F_{1,}$

$_{52.34}$ = 23.0848, p < 0.0001), and the interaction between day length and population ($F_{13,\ 663.43}$ =

7.1168, p < 0.0001; Fig. 2 a; Table 1). The change point analysis showed CPP to be lower for

gonadal recrudescence in residents than migrants. Growth in CPV in residents was detected at

12.4 h of day length, whereas migrants did not exhibit significant growth of CPV until 13 h of

day length (Fig. 2 a). $T_0$ also varied significantly with day length ($F_{13, 659.60} = 12.417$, $p < 0.0001$)

and the interaction between day length and population ($F_{13, 659.64} = 1.9205$, $p = 0.02521$), but

there was no effect of population (Fig. 2 b; Table 1). Change point analysis showed no CPP for

$T_0$. The variable dT varied with day length ($F_{13, 663.29} = 62.786$, $p < 0.0001$), population ($F_{1, 52.11} =$

$45.5151$, $p < 0.0001$), and the interaction between day length and population ($F_{13, 663.34} = 5.5765$,

$p < 0.0001$; Fig. 2 c; Table 1). The effect of the co-variate $\delta^2H$ was close to significance ($F_{1, 51.94}$

$= 3.9416$, $p = 0.0524$). Similar to CPV response, residents showed earlier dT elevation at 11 h of

day length, whereas migrants were delayed by 1h to 12 h of day length (Fig. 2 c). Comparing dT

between HLM, LLM and Residents showed no difference. Interestingly, LLM elevated dT at

11.4 h of day length which differed from migrants originating from higher latitudes (Fig. 2 d).

Age did not show any variation in any physiological response.

**CPP for Fat score and Body mass**

Migrants showed increase in pre-migratory fat score with increasing day length ($F_{13, 662.52} =$

$25.8556$, $p < 0.0001$) in comparison to resident birds which did not fatten ($F_{1, 51.99} = 9.9204$, $p =$

$0.0027$). There was also a significant interaction between day length and population ($F_{13, 662.57} =$

$9.1958$, $p < 0.0001$; Fig. 1 e, Table 1). At the beginning of the experiment, residents had higher

body mass than migrants due to their larger body size. Migrant body mass increased significantly

with day length as they fattened ($F_{13, 660.99} = 16.0132$, $p < 0.0001$), and the interaction between

day length and population was significant ($F_{13, 661.02} = 16.9076$, $p < 0.0001$; Fig. 2 f, Table 1).

Change point analysis revealed CPP for body mass at 11.4 h of day length for migrants (Fig. 2 f);

resident birds did not change body mass as day length increased (Fig. 2 f).

**Life-history State Dependent Changes in Relationship of Phenology to Stable Isotope**

**Values**

We examined LHS-dependent changes in the relationships among CPV, dT, BW, FS with

respect to $\delta^2H$ values, considering residents and migrant collectively (Fig. 3) and dT/BW

relationship with $\delta^2H$ values in migrants separately (Fig. 3).

*Migrants, residents, and stable isotope-* During the photosensitive state, $\delta^2H$ values were

significantly positively correlated with BW ($r = 0.4184$, $p = 0.0013$; Fig. 3c) and FS ($r = 0.2695$,

$p = 0.045$; Fig. 3 d), but not with CPV or dT (Fig. 3 a, b). During recrudescence, both CPV ($r =$

$0.647$, $p < 0.0001$; Fig. 3 e) and dT ($r = 0.4698$, $p = 0.0006$; Fig. 3 f) showed significant positive

correlation and BW ($r = -0.3315$, $p = 0.0126$; Fig. 3 g) and FS ($r = -0.4752$, $p = 0.0002$; Fig. 3 h)

showed negative correlation with $\delta^2H$ values at their respective change point day lengths.

When resident juncos reached their peak (the stimulatory phase), $\delta^2H$ values were positively

correlated with CPV ($r = 0.3823$, $p = 0.0043$; Fig. 3 i) and negatively correlated with BW ($r = -$

$0.273$, $p = 0.0458$; Fig. 3 k), and FS ($r = -0.4786$, $p < 0.0001$; Fig. 3 l). However, when all the

birds reached peak stimulation at 15L, $\delta^2H$ values were not correlated with dT (Fig. 3 j). When

the birds reached the refractory state, BW ($r = 0.5657$, $p < 0.0001$; Fig. 3 o) and dT ($r = 0.334$, $p$

$= 0.0403$; Fig. 3 n) were positively correlated with $\delta^2H$ values providing evidence for latitudinal

variation in timing of reaching refractoriness. CPV and FS were no longer correlated with $\delta^2H$

values (Fig. 3 m, p).

*Migrants and stable isotopes-* Migrants with different $\delta^2H$ values did not show any correlation

with dT and BW in the photosensitive state (Fig. 3 q). With respect to dT in migrants at 11.75 L

(recrudescence; CPP for LLM), gonadal recrudescence was delayed in migrants with lighter $\delta^2H$

values (r = 0.3708, p = 0.0201) and there was no correlation between $\delta^2H$ values and BW (Fig. 3

r). When the migrants reached peak photostimulation, there was no correlation between $\delta^2H$

values and dT, BW (Fig. 3 s). Under the refractory state, comparison of migrant dT (r = 0.4452,

p = 0.0155) and BW (r = 0.3264, p = 0.0426) (Fig. 3 t) reveled a positive correlation with $\delta^2H$

values, providing evidence for $\delta^2H$-dependent difference in the timing of onset of refractoriness

which occurred earlier with lower isotope values (i.e., proxy for higher latitude).

**Timing of molt, migrants and residents**

Finally, post-breeding molt score was negatively correlated with $\delta^2H$ values during the refractory

state (Primary molt: r = -0.4318, p = 0.0008, Fig. 4 a, b; head molt: r = -0.3975, p = 0.006; Fig.

4a-d). Relationship between dT levels, BW and molt score support that birds breeding at

different latitudes also differ in the timing of refractoriness (Fig. 4; Fig. 5). Migrants did not

show any significant correlation with $\delta^2H$ values during the refractory state.

**Discussion**

We found that captive migrant and resident juncos held under naturally increasing photoperiod

exhibited different critical photoperiods for gonadal recrudescence. Resident juncos exhibited

earlier CPV growth and elevation in dT than migrants. Further, timing of elevated CPV and dT

response were associated with latitude as estimated using stable $\delta^2H$ values. Our findings thus

provide evidence for population level variation in the timing of initiation and termination of

breeding in a common environment as a function of latitude, when latitude is estimated using

$\delta^2$H isotope. Termination of reproduction started earlier in migrants than residents, as indicated

by earlier timing of post-breeding molt, concurrent with earlier decline in the dT levels.

Assuming that birds return to their sites of origin, we can conclude that birds destined to breed at

higher latitudes delayed onset of gonadal growth as a correlate of breeding later in the year after

completing longer migrations. To our knowledge such continuous covariation between the

timing of annual life-history states, photosensitive - recrudescence – stimulation, and

refractoriness across different breeding latitudes has not been described previously in any bird

species (Fudickar et al., 2016; Ramenofsky et al., 2017).

*Birds breeding at lower latitudes initiate preparation for reproduction earlier*

The natural history of reproductive timing has shown that birds species breeding at higher

latitudes tend to breed later and terminate reproduction sooner. Additionally, within species,

populations breeding at higher latitudes initiate egg laying later than closely-related populations

found at lower latitudes (Baker 1938, Myers, 1955).  Two independent studies investigating

seasonal reproductive physiology in free living quail from two different locations differing by $9^0$

latitude showed a 2-3 week advance in egg laying date in birds breeding at lower latitudes

(Genelly 1955; Anthony 1970). Another study from two independent labs in England ($52^0$N) and

California ($37^0$N) showed maximum stimulated gonads earlier at lower latitudes corresponding

to first egg laying dates (Dawson and Goldsmith 1982; Rothery et al., 2001).

With respect to migratory species, previous studies have related migratory distance and

breeding latitude to reproductive timing in different bird species (Rubenstein et al., 2002,

Fudickar et al., 2016), and numerous studies have shown that within species complexes,

populations living at lower latitudes tend to have longer breeding seasons than those found at

higher latitude (Dawson and Goldsmith 1983; Dawson, 2013; Greives et al., 2016). But

preparation for breeding begins prior to the beginning of breeding season as defined by first egg

laying date. Gonadal recrudescence precedes reproduction and is associated with a rise in

circulating testosterone.

*Latitude is directly proportional to critical photoperiod response for gonad recrudescence-*

There are a very few studies testing the difference in photoperiodic threshold as a prerequisite to

initiate early gonadal recrudescence. Birds collected from three different latitudes ($45^0$, $57^0$, $70^0$

N) and maintained in a common garden set up where they were exposed to gradually increasing

photoperiod showed earlier maturation in those from the lower latitude ($45^0$N), but not in other

two groups of birds from higher latitudes (Silverin et al., 1993). A recent common garden study

of two subspecies of white crowned sparrows (*Zonotichia leucophrys*), and dark-eyed juncos

(*Junco hyemalis*) that differed in migratory strategy showed differential responses in sensitivity

to increasing day length, which influences the induction and termination of breeding (Fudickar et

al., 2016; Ramenofsky et al., 2017). Our results fill several gaps in knowledge about the

differences in the critical photoperiod threshold of dark-eyed junco subspecies as they transition

from photosensitivity to gonadal recrudescence, duration of the stimulatory phase and timing of

attaining photorefractoriness.

*Birds breeding at higher latitude terminate reproduction sooner-*

The transition from the stimulatory to refractory state is signaled by molt (Hall and Fransson

2000) and our results also revealed a difference between migrant and resident juncos in the

timing of refractoriness. This compares to starlings held in captivity and exposed to photoperiods

simulating annual cycles at higher ($52^0$N) and lower ($9^0$N) latitude.  The starlings showed earlier

gonadal maturation in $9^0$N, but birds from both latitudes regressed their gonads at the same time

(Dawson, 2013). In our study, towards the end of the experiment when all birds were on 16h

photoperiod, high latitude migrants no longer responded to GnRH by elevating T, while residents

and low latitude migrants continued to elevate T in response to GnRH.  Further, the relationship

between molt score and T in response to GnRH across latitude also showed that birds originating

from higher latitudes became photorefractory earlier.

The observed difference in the pattern created by latitudinal variation in CPP and seasonal life-

history states can provide a framework for testing how various climatic variables account for

variation in seasonal timing of birds from different latitudes. The juncos with lighter $\delta^2$H delayed

recrudescence, remained in breeding phase for a shorter period, and become refractory sooner,

unlike the juncos with heavier $\delta^2$H which recrudesced earlier, had longer breeding periods and

entered the refractory state later. In total our results point out towards latitudinal variation in the

pace of life-history states and the mechanisms underlying seasonal changes in the responsiveness

of HPG axis.

Seasonal life-history states and associated phenology have long been studied, but are currently

receiving renewed attention in the context of global climate change. In the temperate zone,

photoperiod, temperature, and other environmental variables often correlate with seasonal

phenology across latitude. As, a consequence, the study of species-specific seasonal phenology

has strong application in the context of global climate change (Schwartz 2003, Parmesan 2006).

Fitness for a seasonal animal involves not only the ability to synchronize behavior and

physiology to the seasons but also to anticipate, prepare, and cope with the changing seasons. For

migratory birds, there is an additional challenge of identifying the optimal time to initiate

migration and recrudesce gonads while living at locations distant from their breeding locations.

For example, warmer winters are resulting in earlier springs.  As a consequence, migrants that

arrive at times that were formerly optimal find that peak food availability for rearing offspring

has already passed, leading to mismatches between migration schedules and optimal times to

breed (Lack 1968; Visser at al., 2004). Some long-distance migratory bird species have advanced

their spring arrival dates in response to climate change, but others have not and arrived too late

for the pulse of insect food needed to nurture offspring (Visser et al., 2004; Jonzén et al., 2006).

Mismatch in the seasonal timing has significant consequences at the population level (Nussey et

al., 2005; Both et al, 2006). Thus, knowing the mechanisms of life-history state-dependent

phenology may help in predicting the effect of climate change on survival and fitness of species

(Miller-Rushing et al., 2010).

404
In order to estimate the ecological consequences of climate change we must be able to

forecast the shifts in the direction, magnitude, and phase of phenological processes under

different environmental scenarios. This forecasting is impeded due to a shortage of precise

knowledge of the mechanisms determining the pace of life-history events.  Phenology is a key

process that reflects an organism's micro-evolutionary response to a wide range of

environmental cues (van Asch et al., 2007). Animals distributed geographically across a wide

range of photoperiod, temperature, and other environmental conditions that vary with latitude are

known to express phonological events at different phases of the annual cycle. Hence, it is critical

to incorporate mechanistic and evolutionary perspective while forecasting ecological

consequences of climate change.

**Conclusions**

Our study demonstrates differences in seasonal timing across latitudes in response to changing

photoperiod and reveals some of the underlying mechanisms and their potential for adaptive

response to environmental change. Birds breeding at lower latitudes recrudesced earlier,

maintained the stimulatory phase for longer, and attained refractoriness later. Whereas, birds

originating at higher latitude delayed recrudescence, remained stimulatory for a shorter time, and

attained refractoriness sooner. That is, latitude was directly proportional to the critical

photoperiod required for recrudesce and inversely proportional to the timing of refractoriness.

Particularly informative was a group of migrants that had $\delta^2H$ similar to residents and an

intermediate response in CPP and timing of refractoriness. This may indicate that migration

delays reproduction, but the extend of the delay depends on how far a bird has to travel. The

approach used in this study can be applied to other species in which populations that differ in

where they breed are found in the same winter environment as they do or do not prepare to

migrate. It will be interesting to learn whether their patterns resemble those seen in the junco.

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**Authors' contribution**: DS and EDK conceived the idea. DS, SRR, AAK, and KAA carried out

the experiment. CS performed the hydrogen stable isotope experiment and provided data. DS

wrote the manuscript with the help of all authors. All authors approved the final draft.

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**Figure legends**

**Figure 1.** Schematic diagram of the hydrogen isotope ($\delta^2$H) precipitation map at different

latitudes in North America during the months of July and August, Juncos undergo a pre-basic

molt after breeding and their feathers incorporate stable hydrogen isotope signatures from their

local environments (Rubenstein et al., 2002). Left panel represents the latitudinal distribution of

dark-eyed juncos breeding range based on their feather $\delta^2$H (a). The bar graph represents mean

difference in resident and migrant juncos' hydrogen isotopes. Values of $\delta^2$H on the Y-axis are

represented as hollow circle for migrants, solid grey circle for LLM, and solid circle for

residents. The alpha was set at 0.05 (b). Some of the individual migrants' overlap in isotope with

residents (c).  The bluish bill residents (*J.h. carolinesis*) (R; $\delta^2$H: -37 to -81) breed at lower

latitudes and have heavier $\delta^2$H in comparison to pink bill migrants (*J.h. hyemalis*) which breed

over a wide latitudinal range from south (low latitude migrants; $\delta^2$H: -30 to -88) to farther north

(high latitude migrants; $\delta^2$H: -90 to -141).   Have you double checked the numbers recently?

**Figure 2.** Latitudinal cline in critical photoperiod threshold for different seasonal physiological

responses.  Measurement of CPV (a), $T_0$ (b), dT (c), FS (d), BW (e) in migrants (hollow circle)

and residents (solid circle), and dT (f) in HLM (hollow circle), LLM (grey solid circle) and

residents (solid circle) starting from 9 h of light to 16h of light. Y-axis represents the

physiological parameters and X-axis represents increasing photoperiod in hours of day length

exposure.  Each data point represents mean ± SEM. Data were analyzed using a mixed-effect

model with repeated measures for effects of day length, population, and interaction between day

length and population. Statistical significance was defined by alpha <0.05. The arrow in the

circle and above day length shows critical photoperiod threshold point in respective population,

determined by change point analysis. The seasonal life-history states (LHSs) were defined as

photosensitive (9L), recrudescence (CPV, 12.4L; dT, 11L), photostimulatory (15L), and

photorefractory (16L) state.

**Figure 3.** Life history state dependent changes in the relationship between CPV, BW, FS and dT

across latitude. Correlation between residents and migrants CPV, $T_0$, dT, BW, and FS against

$\delta^2H$ as a constant variable in photosensitive (9L: initial point; a-d), recrudescence (12.4L:

CPV/BW/FS and 11L: dT; critical photoperiod threshold, e-h), photostimulatory (15L: CPV, dT

level at highest peak; i-l), and photorefractory (16L: endpoint; m-p) states. Correlation among

migrants dT and BW against $\delta^2H$ values across different LHSs (q-t). Pearson correlation was

used for all physiological data except fat score which is an ordinal value. Spearman correlation

was used for fat score ordinal data. X-axis represents the $\delta^2H$ as a constant variable and Y axis

represents individual data points for CPV, dT (hollow circles), BW, FS (solid circle). Statistical

significance was defined by alpha <0.05. Linear regression line with 95% confidence interval

(shaded area) represents significant correlation as solid line and dotted line for no significant

correlation.

**Figure 4.** Latitudinal variation in timing of molt, dT, and BW during endpoint refractory state.

Pearson correlation between BW/ primary molt score (a), BW/ head molt score (b), dT/primary

molt score (c), and dT/head molt score (d) against $\delta^2H$ as a constant variable in 16 L

photorefractory state. X-axis represents the $\delta^2H$ as a constant variable and Y axis represents

individual data points BW, dT (hollow circles), primary and head molt score (solid circle).

Statistical significance was defined by alpha <0.05. Linear regression line with 95% confidence

interval (shaded area) represents significant correlation as solid line and dotted line for no

significant correlation.

**Figure 5**. Latitudinal cline in critical photoperiod (CPP) for gonadal recrudescence and pace of

life history states. Schematic summary Figure showing the breeding latitude dependent response

to different critical photoperiod (CPP) and difference in the pace of different life history states in

dark eyed juncos. Left panel showing the CPP for cloacal protuberance and dT response across

latitude (a). Right panel showing the latitude-dependent seasonal waveforms of different life

history states derived from common garden experiment. Photosensitive juncos overwintering at

the same latitude exposed to increasing day length in a common garden show divergence in the

development of recrudescence, photostimulation and timing of attaining photorefractoriness.

Residents (black solid line), Low Latitude Migrants (black dotted line), high latitude migrants

(red dotted line) exhibit differences in CPP to initiate gonad recrudescence, reach stimulation and

attain photorefractoriness.  Residents breeding at lower latitudes reproduce earlier, maintain

stimulatory phase for a longer period and undergo refractory state later. High and Low Latitude

migrants delay recrudescence and reach peak stimulation later and undergo refractoriness sooner

(b).

**Table 1.**

Factor (s) affecting different physiological responses (CPV, FS, BW, $T_0$, dT): Analysis of

variance table of type III with Satterthwaite approximation for degrees of freedom derived from

linear mixed effects model. Number of asterisk (*) denotes level of significance p <0.05(*), p

<0.001(**), P<0.0001(***).

**References**

Anthony, R. (1970). Ecology and reproduction of California quail in southeastern Washington.

*Condor*, 72, 276–287.

Atwell, J.W., O'Neal, D.M., & Ketterson, E.D. (2011). Scientific research agenda: animal

migrations as a moving target for conservation: intra-species variation and responses to

environmental change, as illustrated in a sometimes migratory songbird. *Environ. Law*,

41, 289–655.

Baker, J.R. (1938). The relation between latitude and breeding seasons in birds. *Proc. Zool. Soc.*

*London Series A,* 108, 557–582.

Ball, G.F., & Balthazart, J. (2003). Birds return every spring like clockwork, but where is the

clock? *Endocrinology*, 144, 3739–3741.

Ball, G.F., & Ketterson, E.D. (2008). Sex differences in the response to environmental cues

regulating seasonal reproduction in birds. *Philos. Trans. R. Soc. B Biol. Sci.,* 363, 231–

246.

Both, C., Bouwhuis, S., Lessells, C. M. & Visser, M. E. (2006). Climate change and population

declines in a longdistance migratory bird. *Nature*, 441, 81–83.

Bradshaw, W. E. & Holzapfel, C. M. (2007). Evolution of animal photoperiodism. *Annu. Rev.*

*Ecol. Evol. Syst.,* 38, 1–25.

Bronson, F. H., & P. Heideman. (1994). Seasonal regulation of reproduction in mammals.

*Physiology of Reproduction*, 2, 541–584.

Burger, J.W. (1949). A review of experimental investigations on seasonal reproduction in birds.

*Wilson Bull.,* 61, 211–230.

Cho, R.N., Hahn, T.P., MacDougall-Shackleton, S., & Ball, G.F. (1998). Seasonal variation in

brain GnRH in free-living breeding and photorefractory House finches (*Carpodacus*

*mexicanus*). *Gen. Comp. Endocrinol.,* 109, 244–250.

Cristol, D. A., Reynolds E. Leclerc B., J. E., Donner A. H., Farabaugh C. S., & C. Ziegenfus W.

S. (2003). Migratory dark-eyed juncos, *Junco hyemalis*, have better spatial memory and

denser hippocampal neurons than nonmigratory conspecifics. *Animal Behaviour,* 66,

317–328.

Dawson, A. (2013). The effect of latitude on photoperiodic control of gonadal maturation,

regression and molt in birds. *Gen. Comp. Endocrinol.,* 190, 129–133.

Dawson, A., & Goldsmith, A.R. (1983). Plasma prolactin and gonadotrophins during gonadal

development and the onset of photorefractoriness in male and female starlings (*Sturnus*

*vulgaris*) on artificial photoperiods. *J. Endocrinol.,* 97, 253–260.

Dawson, A., Goldsmith, A.R., & Nicholls, T.J. (1985). Development of photorefractoriness in

intact and castrated male starlings (*Sturnus vulgaris*) exposed to different periods of long-

day lengths. *Physiological Zoology*, 58, 253–261.

Dawson, A., King, V.M., Bentley, G.E., & Ball, G.F. (2001). Photoperiodic control of

seasonality in birds. *J. Biol. Rhythms,* 16, 365–380.

Farner, D.S., & Mewaldt, L.R. (1955). The natural termination of the refractory period in white-

crowned sparrows. *Condor,* 57, 112–116.

Farner, D.S., & Wilson, A.C. (1957). A quantitative examination of testicular growth in the

white-crowned sparrow. *Biol. Bull.,* 113, 254–267.

Follett, B.K., Kumar, V., & Juss, T.S. (1992). Circadian nature of the photoperiodic clock in

Japanese quail. *J. Compar. Physiol. A – Sensory Neur. Behav. Physiol.,* 171, 533–540.

Follett, B.K., & Maung, S.L. (1978). Rate of testicular maturation, in relation to gonadotrophin

and testosterone levels, in quail exposed to various artificial photoperiods and to natural

daylengths. *J. Endocrinol.,* 78, 267–280.

Fudickar, A.M., Greives, T.J., Atwell, J.W., Stricker, C.A., & Ketterson, E.D. (2016). Reproductive allochrony in seasonally sympatric populations maintained by differential response to photoperiod: implications for population divergence and response to climate change. *Am. Nat.,* 187, 436–446.

Greives, T.J., Fudickar, A.M., Atwell, J.W., Meddle, S.L. & Ketterson, E.D. (2016) Early spring sex differences in luteinizing hormone response to gonadotropin releasing hormone in co-occurring resident and migrant dark-eyed juncos (*Junco hyemalis*). *General and Comparative Endocrinology,* 236, 17-23.

Hall, K. S. S. & Fransson, T. (2000). Lesser whitethroats under time-constraint moult more rapidly and grow shorter wing feathers. *J. Avian Biol.,* 31, 583–587.

Hobson K.A. (2003). Making migratory connection with stable isotopes. In: *Avian Migration (Ed. By P. Berthold et al.),* pp. 379–391. Berlin, Heidelberg: Springer-Verlag.

Jawor, J. M., McGlothlin, J.W., Casto, J. M., Greives, T. J., Snajdr, E. A., Bentley, G. E., et al. (2006). Seasonal and individual variation in response to GnRH challenge in male darkeyed juncos (*Junco hyemalis*). *General and Comparative Endocrinology,* 149, 182–189.

600 Jonzén, N., Lindén, A., Ergon, T., Knudsen, E., Vik, J.O., Rubolini, D., et al. (2006). Rapid
601            advance of spring arrival dates in long-distance migratory birds. *Science,* 312, 1959-
602            1961.

604 Killick, R., & Eckley, I. (2014). Changepoint: An R Package for Changepoint Analysis. *Journal*
605            *of statistical software.,* 58, 1-19.

607 Lack, D. (1968). Ecological Adaptations for Breeding in Birds. *Chapman & Hall, London*.

609 Li, Q., Tamarkin, L., Levantine, P., & Ottinger, M. (1994). Estradiol and androgen modulate
610            chicken luteinizing hormone-releasing hormone-I release in vitro. *Biol. Reprod.,* 51, 896–
611            903.

613 Menaker, M. (1971). Rhythms, reproduction and photoreception. *Biol. Reprod.,* 1, 295–308.

615 Miller-Rushing, A.J., Høye, T.T., Inouye, D.W. & Post, E. (2010). The effects of phenological
616            mismatches on demography. *Phil. Trans. R. Soc. B*, 365, 3177–3186.

618 Miller, A.H. (1954). The occurrence and maintenance of the refractory period in crowned
619            sparrows. *Condor,* 56, 13–20.

621 Myers, M.T. (1955). The breeding of the blackbird, song thrush and mistle thrush in Great
622            Britain: part 1, breeding seasons. *Bird Study* 2, 2–24.

Genelly, R.E. (1955). Annual cycle in a population of California quail. *Condor,* 57, 263– 285.

Nolan Jr., V., Ketterson, E.D., Cristol, D.A., Rogers, C.M., Clotfelter, E.D., Titus, R.C., et al.

(2002). Dark-eyed Junco (*Junco hyemalis*). In: Poole, A. (Ed.), The Birds of North

America Online. Cornell Lab of Ornithology, Ithaca, NY.

http://bna.birds.cornell.edu/bna.

Nussey, D. H., Postma, E., Gienapp, P. & Visser, M. E. (2005). Selection on heritable

phenotypic plasticity in a wild bird population. *Science*, 310, 304–306.

Parmesan, C. (2006). Ecological and evolutionary responses to recent climate change. *Annu. Rev.*

*Ecol. Evol. Syst.,* 37, 637–669.

Pyle, P. 1997. Identification guide to North American birds. Slate Creek Press, Bolinas. CA.

Ramenofsky, M., Campion A.W., Pérez, J.H., Krause, J.S., & Németh, Z. (2017). Behavioral and

physiological traits of migrant and resident white crowned sparrows: a common garden

approach. *Journal of Experimental Biology*, 220, 1330-1340.

Rothery, P., Wyllie, I., Newton, I., Dawson, A., & Osborn, D. (2001). The timing and duration of

moult in adult Starlings *Sturnus vulgaris* in east-central England. *Ibis*, 143, 435–441.

Rowan, W. (1926). On photoperiodism, reproductive periodicity, and the annual migrations of
certain birds and fishes. *Proc. Boston Soc. Nat. Hist.,* 38, 147-189.

Rubenstein, D.R., Chamberlain, C.P., Holmes, R.T., Ayres, M.P., Waldbauer, J.R., Graves, G.R.,
& Tuross, N.C. (2002). Linking Breeding and Wintering Ranges of a Migratory Songbird
Using Stable Isotopes. *Science,* 295, 1062-1065.

Robart, A.R., McGuire, M.M.K., & Watts, H.E. (2018). Increasing photoperiod stimulates the
initiation of spring migratory behaviour and physiology in a facultative migrant, the pine
siskin. *R. Soc. open sci.* 5, 180876.

Schut, E., Magrath, M.J.L., van Oers, K., & Komdeur, J. (2012). Volume of the cloacal
protuberance as an indication of reproductive state in male Blue Tits *Cyanistes caeruleus.*
*Ardea,* 100, 202-205.

Schwartz, M.D. (ed.) (2003). Phenology: an integrative environmental science. Norwell, MA:
Kluwer Academic Publishers.

Silverin, B., Massa, R., & Stokkan, K.A. (1993). Photoperiodic adaptation to breeding at
different latitudes in Great tits. *Gen. Comp. Endocrinol.,* 90, 14–22.

Spinney, L., Bentley, G., & Hau, M. (2006). Endocrine correlates of alternative phenotypes in
the white-throated sparrow (*Zonotrichia albicollis*). *Horm. Behav.*, 50, 762–771.

van Asch, M., Tienderen, P.H., Holleman, L.J.M., & Visser, M.E. (2007). Predicting adaptation of phenology in response to climate change, an insect herbivore example. *Global Change Biol.*, 13, 1596–1604.

Visser, M. E., Both, C., & Lambrechts, M.M. (2004). Global climate change leads to mistimed avian reproduction. *Adv. Ecol. Res.,* 35, 89–110.

Watts, H. E., MacDougall-Shackleton, S. A., & Hahn, T. P. (2015). Variation among individuals in photoperiod response: Effects of breeding schedule, photoperiod, and age-related photoperiodic experience in birds. *J Exp Zool A Ecol Genet Physiol.,* 323, 368-374.

Wingfield, J.C., Crim, J.W., Mattocks, P.W., & Farner, D.S. (1979). Responses of photosensitive and photo-refractory male white-crowned sparrows (*Zonotrichia-leucophrys Gambelii*) to synthetic mammalian luteinizing hormone releasing hormone (Syn-LHRH). *Biol. Reprod,* 21, 801–806.

Wingfield, J.C., Hahn, T.P., Levin, R., & Honey, P. (1992). Environmental predictability and control of gonadal cycles in birds. *J. Exp. Zool.,* 261, 214–231.

Wingfield, J.C. (2012). Regulatory mechanisms that underlie phenology, behavior, and coping with environmental perturbations: an alternative look at biodiversity. *Auk* 129, 1-7.

Wolfson, A. (1952). The cloacal protuberance: a means for determining breeding condition in

live male passerines. *Bird-Banding*. 23, 159-165.

Wunder, M.B., Jehl, J.R., & Stricker, C.A. (2012). The early bird gets the shrimp: confronting

assumptions of isotopic equilibrium and homogeneity in a wild bird population. *Journal*

*of Animal Ecology*, 81, 1223–1232.

Yasuo, S., Watanabe, M., Okabayashi, N., Ebihara, S., & Yoshimura, T. (2003). Circadian clock

genes and photoperiodism: comprehensive analysis of clock gene expression in the

mediobasal hypothalamus, the suprachiasmatic nucleus, and the pineal gland of Japanese

quail under various light schedules. *Endocrinology*, 144, 3742–3748.

**Figure 1**

b)

HLM  LLM, R

**HLM**: high latitude migrants
**LLM**: low latitude migrant
  **R**  : residents

c)

Migrant    Resident

$\delta^2$H (‰)

☐ -26.1 to -0.8
☐ -51.3 to -26.1
☐ -76.6 to -51.3
☐ -101.8 to -76.6
☐ -127.1 to -101.8
☐ -152.3 to -127.1
☐ -177.6 to -152.3
☐ -202.8 to -177.6
☐ -228.1 to -202.8

**Figure 2**

LHSs: [ *Photosensitive: 9L; Recrudescence: 12.4L (CPV), 11L (dT),*
*11.4L (dT, LLM); Photostimulatory: 15L; Photorefractory: 16L* ]

residents, migrants CPV, dT BW, and FS Vs d²H     migrants dT and BW Vs d²H

Life history states (LHSs)

photosensitive · recrudescence · photostimulatory · photorefractory

d²H (‰)

a)

(δ²H) latitude

HLM
LLM
R

12L (HLM)
11.4L (LLM)
11L (R)
dT

13L (M)
12.4L (R)
CPV

Critical photoperiod for dT and CPV response

b)

T0    T30

GnRH1

Daylength (h)

9L  10L  11L  11.4L  11.75L  12L  12.4L  12.75L  13L  15L  16L  16L

δ²H ratio: breeding latitude

photosensitive    photostimulatory    photorefractory

dT Critical photoperiod (CPP) response

R — LLM - - - - HLM —·—·—

**Table 1.**

| CPV | Sum of Square | Mean Square | Numerator DF | Denominator DF | F-value | p-value |
|---|---|---|---|---|---|---|
| Day length (F1) | 4361.7 | 335.52 | 13 | 663.38 | 66.3716 | **<0.0001**(***) |
| Population (F2) | 116.7 | 116.70 | 1 | 52.34 | 23.0848 | **<0.0001**(***) |
| Age | 2.1 | 2.13 | 1 | 266.93 | 0.4209 | 0.517 |
| $\delta^2H$ | 12.5 | 12.48 | 1 | 52.16 | 2.4683 | 0.1222 |
| F1 X F2 | 467.7 | 35.98 | 13 | 663.43 | 7.1168 | **<0.0001**(***) |

| $T_0$ | Sum of Square | Mean Square | Numerator DF | Denominator DF | F-value | p-value |
|---|---|---|---|---|---|---|
| Day length (F1) | 20.0796 | 1.54459 | 13 | 659.60 | 12.4217 | **<0.0001**(***) |
| Population (F2) | 0.4149 | 0.41487 | 1 | 50.22 | 3.3364 | 0.07371 |
| Age | 0.0422 | 0.04220 | 1 | 516.23 | 0.3394 | 0.56043 |
| $\delta^2H$ | 0.1572 | 0.15717 | 1 | 49.94 | 1.2639 | 0.26628 |
| F1 X F2 | 3.1044 | 0.23880 | 13 | 659.64 | 1.9205 | **<0.0252**(*) |

| dT | Sum of Square | Mean Square | Numerator DF | Denominator DF | F-value | p-value |
|---|---|---|---|---|---|---|
| Day length (F1) | 313.023 | 24.0787 | 13 | 663.29 | 62.7863 | **<0.0001**(***) |
| Population (F2) | 17.455 | 17.4541 | 1 | 52.11 | 45.5151 | **<0.0001**(***) |
| Age | 0.439 | 0.4393 | 1 | 252.53 | 1.1455 | 0.2855 |
| $\delta^2H$ | 1.512 | 1.5116 | 1 | 51.94 | 3.9416 | **0.0524** |
| F1 X F2 | 27.802 | 2.1386 | 13 | 663.34 | 5.5765 | **<0.0001**(***) |

| FS | Sum of Square | Mean Square | Numerator DF | Denominator DF | F-value | p-value |
|---|---|---|---|---|---|---|
| Day length(F1) | 58.464 | 4.4972 | 13 | 662.52 | 25.8556 | **<0.0001**(***) |
| Population (F2) | 1.726 | 1.7255 | 1 | 51.99 | 9.9204 | **0.00271**(**) |
| Age | 0.046 | 0.0460 | 1 | 333.71 | 0.2646 | 0.607325 |
| $\delta^2$H | 0.052 | 0.0516 | 1 | 51.77 | 0.2969 | 0.588199 |
| F1 X F2 | 20.793 | 1.5995 | 13 | 662.57 | 9.1958 | **<0.0001**(***) |

| BW | Sum of Square | Mean Square | Numerator DF | Denominator DF | F-value | p-value |
|---|---|---|---|---|---|---|
| Day length(F1) | 342.22 | 26.3247 | 13 | 660.99 | 16.0132 | **<0.0001**(***) |
| Population (F2) | 1.86 | 1.8597 | 1 | 52.05 | 1.1313 | 0.2924 |
| Age | 3.03 | 3.0307 | 1 | 594.55 | 1.8436 | 0.1750 |
| $\delta^2$H | 3.14 | 3.1420 | 1 | 51.75 | 1.9112 | 0.1728 |
| F1 X F2 | 361.34 | 27.7951 | 13 | 661.02 | 16.9076 | **<0.0001**(***) |