1 2 3 Title: Epidemiology of bacterial contamination of inert hospital surfaces and equipment in critical and non-critical care units: a Brazilian multicenter study 4 Short title: *Epidemiology of contamination of hospital surfaces* 5 6 Dayane Otero Rodrigues^{1*}, Laís da Paixão Peixoto²¶, Erica Tatiane Mourão Barros²¶, 7 Julianne Rodrigues Guimarães^{2¶}, Bruna Clemente Gontijo^{3&}, Jaisa Leite Almeida^{3&}, 8 Lucas Guimarães de Azevedo³&, Júlia Cristina Oliveira e Lima³& and Deyse Silva 9 Câmara^{3&} 10 11 ¹ Centro de Ciências Biológicas e da Saúde, Universidade Federal do Oeste da Bahia, 12 Barreiras, Bahia, Brasil 13 ² Laboratório de Microbiologia, Curso de Biomedicina, Centro Universitário Luterano de 14 Palmas, Palmas, Tocantins, Brasil 15 ³ Curso de Medicina, Centro de Ciências Biológicas e da Saúde, Universidade Federal do 16 Oeste da Bahia, Barreiras, BA, Brasil 17 18 19 *Corresponding author 20 Email: dayotero@yahoo.com.br (DOR) 21 22 23 24 ¶These authors contributed equally to this work. 25 &These authors also contributed equally to this work.

Abstract

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

The hospital environment is an important reservoir of microorganisms, including multidrug-resistant pathogens, which can cause in-patient contamination and healthcarerelated infections. The objective of this study was to analyze the epidemiology of bacterial contamination (contaminated sites, pathogen species and their antimicrobial susceptibility, and tracking of multidrug-resistant microorganisms - MDR) of inert hospital surfaces and medical equipment in two public hospitals in Northern Brazil. This was a cross-sectional study with 243 samples (n = 208, from Hospital A; and n = 35, from Hospital B) collected by friction with swabs moistened in Brain Heart Infusion from inert surfaces and equipment. The samples were cultivated and bacterial species were identified by the classical approach and tested for their susceptibility through agar diffusion assay according to the Clinical and Laboratory Standards Institute (CLSI). Most inert surfaces and equipment analyzed presented bacterial contamination (95.5%). Staphylococcus aureus was the main pathogen of clinical significance detected both in Hospital A (61.8%) and B (68.6%). Hospital A showed higher rates of isolated MDR bacteria than Hospital B, especially in the Adult Intensive Care Unit, which included methicillin-resistant Staphylococcus aureus (MRSA) (52.7%), Enterobacteria resistant to 4th generation cephalosporins (19.4%), and multidrug-resistant *Pseudomonas aeruginosa* (2.78%). The failures in the prevention and control of infections in the two hospitals analyzed reinforce the need for a revised protocol for cleaning and disinfection of inert surfaces and medical equipment, and for regulation of antibiotic dispensing, mainly in the AICU of Hospital A, which was found to be a reservoir of MDR pathogens. This study is innovative because it is the pioneer in Western Bahia that describes the epidemiology of contamination of hospital surfaces, opportuning futures studies in this field.

Introduction

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

Healthcare-associated infections (HAI) are a major public health concern commonly associated with extended length of hospital stay. HAI account for high hospital costs and contribute to increased morbidity and mortality of infected patients [1]. HAI are usually caused by pathogenic bacteria that may emerge from the patient's endogenous microflora during antibiotic therapy in approximately 70% of the cases [2,3]. HAI may also be acquired from the exogenous environment (30% of the cases) in that the hospital setting plays a significant role in contagion and transmission outbreaks [3,4]. In the hospital setting, patients, staff and visitors represent the main reservoir of microorganisms, whereas secondary reservoirs include all environments where nutrients, moisture, and temperature are suitable for microbial survival, such as air humidifiers and nebulizers [5,6]. In addition, dry and inanimate surfaces can also serve as a reservoir of pathogens [3,5,7-9], as in mattresses and bed frames [4,10,11], door knobs [11,12], and even in medical equipment such as stethoscopes and ultrasound devices [7-11]. Contamination of these surfaces contributes to pathogen spreading and, as a result, development of horizontal infections [13-14]. Overall, surfaces can be directly contaminated by bacteria from colonized and/or infected patients or from the hands of health professionals [5,7]. Highly touched surfaces (e.g. bed frames, stethoscopes, bedside tables, and door knobs) [11-13] may be contaminated by common bacteria of the hand microbiota. More importantly, MDR bacteria have been detected in medical equipment and contact surfaces, especially in critical care units [10,15,16]. Studies carried out in Brazil and North America have reported contamination of hospital surfaces by bacteria resistant to antibiotics, especially methicillin-resistant

S. aureus (MRSA) [4,17,18] and vancomycin-resistant enterococci [19,20]. These findings indicate that patients and staff are at risk of contamination by pathogens associated with high mortality rates against which treatment options are restricted.

Given the importance of in-hospital transmission, HAI studies have looked into the epidemiology of hospital-related bacterial contamination to propose preventive measures to reduce the contamination and dissemination of resistant pathogens. This study aimed to analyze the epidemiology of bacterial contamination (contaminated sites, pathogen species and their antimicrobial susceptibility, and tracking of multidrug-resistant microorganisms - MDR) of inert hospital surfaces and medical equipment in two public hospitals in Northern Brazil.

Materials and methods

This was a multicenter, cross-sectional study carried out in the *Hospital Geral Público de Palmas* (HGPP) [Palmas Public General Hospital] (A) located in the state of Tocantins, Brazil, and in the *Hospital Municipal Eurico Dutra* [Eurico Dutra Municipal Hospital] (B) located in the city of Barreiras, Western Bahia, Brazil.

Hospital A is a large teaching hospital which provides tertiary care and is a reference center for medium and high complexity healthcare assistance. The hospital contains approximately 400 beds and is a major health center for the state of Tocantins and neighboring states in Brazil. It includes two intensive care units (ICUs) – a pediatric unit (PICU) with 3 rooms and 8 beds, and an adult unit (AICU) with 26 beds distributed among 18 rooms; pharmacy; laboratory; operating room (OR); elective and emergency surgery services; emergency room (ER) with three on-call specialties – orthopedics, internal medicine, and surgery; conventional hospital ward, and home care and outpatient

facilities, with an average 3,500 appointments per month.

Hospital B is a medium-sized and medium complexity general hospital which provides services such as hospital admission, diagnostic and therapeutic support services, emergency and outpatient care. Patients are admitted through spontaneous and referred demand from the city of Barreiras and surrounding region. The hospital has a total of 10 beds in the Internal Medicine Ward (IMW), 29 beds in the Surgical Ward (SW) and 4 OR with 5 post-anesthesia recovery beds in the SW.

Sample collection

99

100

101

102

103

104

105

106

107

108 A total of 243 samples (n = 208, Hospital A; n = 35, Hospital B) were collected from the following surfaces and medical equipment: Hospital A - ER (door knobs, n =109 56); SW (door knobs, n = 20); AICU (heart monitors, n = 18; infusion pumps, n = 18; 110 medication tables, n = 18; side bed frames, n = 18; with a total n = 72); and PICU (side 111 bed frames, n = 16; bed headboards, n = 8; bed frame feet, n = 8; mattresses, n = 8; bedside 112 tables, n = 8; stethoscopes, n = 8; samples from the sinks, n = 4; with a total n = 60); 113 Hospital B - IMW (bed headboard, n = 3, mattresses, n = 2; side bed frame, n = 2; saline 114 stand, n = 1; door knobs, n = 3, with a total n = 11); SW (bed headboard, n = 5, mattresses, 115 116 n=3; side bed frame, n=1; saline stand, n=3; countertops, n=1; door knobs, n=4; with a total n = 17); OR (surgical light, n = 4; stretcher, n = 3; with a total n = 7). The 117 selected hospital surfaces included those highly touched or nearby patients. 118 119 The samples were collected in the morning and afternoon from July to October 2018 (Hospital A) and April 2018 (Hospital B) by friction with sterile swabs moistened 120 in broth Brain Heart Infusion (BHI, Oxoid, Basingstoke, Hampshire, England) from 121 selected surfaces. The swabs were placed immediately into sterile tubes and transported 122

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

Bacterial strains were considered as multidrug-resistant (MDR) if showing

148

149

150

151

152

153

154

155

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

resistance to at least three classes of antimicrobials [23,24] and associated phenotypes. Some examples included MRSA, Enterobacteria resistant to 4th generation cephalosporins and quinolone-resistant P. aeruginosa, according to definitions of the European Center for Disease Prevention and Control [23]. **Results** Bacterial contamination was detected in 94.7% of the 208 sampled surfaces and equipment in Hospital A. A total of 233 bacterial isolates were obtained therefrom, with no microbial growth in only 11 surfaces and equipment in the PICU. Moreover, 100% of the 35 sampled surfaces and equipment in Hospital B showed bacterial contamination. The microbiological analysis of the surfaces in Hospital A showed a predominance of common bacteria of the human flora, such as S. aureus (62.0%) and nosocomial bacteria, such as Enterobacteria (24.0%) and *P. aeruginosa* (7%). In Hospital B, there was a predominance of environmental microorganisms, such as Gram-positive Bacilli (41%) and *Micrococcus* spp. (9.0%) (Figure 1). Figure 1. Frequency of microorganisms recovered from inert surfaces and medical equipment in the Hospital Geral Público de Palmas-TO, Brazil (A) and Hospital Municipal Eurico Dutra in the city of Barreiras, BA, Brazil (B). In Hospital A, the ER door knobs were mostly contaminated with S. aureus (53.3%), but also with enteric bacteria (30.4%), with a high frequency of samples

contaminated with P. aeruginosa (64.7%) (Table 1). The AICU showed the highest

frequency of S. aureus colonization, particularly on heart rate monitors (83.3%). Infusion

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

(2.78%). The OR in both Hospital A and B had the lowest frequency of MDR bacteria, as shown in Table 3.

Table 3. Multi-drug resistant microorganisms recovered from inert surfaces and equipment of hospitals located in Northern Brazil. Legend - *Multidrug-resistant pathogen; ER: Emergency Room; OR: Operating Room; AICU: Adult Intensive Care Unit; PICU: Pediatric Intensive Care Unit; IMW: Internal Medicine Ward; SW: Surgical Ward; MRSA: Methicillin-Resistant *Staphylococcus aureus*; MR-CoNS: Methicillin-Resistant Coagulase-Negative Staphylococci.

Discussion

Inert hospital surfaces and medical equipment can be a reservoir of MDR pathogens. Understanding the epidemiology of bacterial contamination in this setting is essential to prevent in-patient contamination and HAI development.

Bacterial pathogens can survive and remain viable on inert surfaces and equipment due to their ability to form biofilms and to environmental factors such as surface porosity and humidity [7,25]. This ability to adapt to environmental stress works as a major factor driving pathogen thriving and dissemination. Our results are consistent with these premises as 94.7% (Hospital A) and 100.0% (Hospital B) of the inanimate surfaces analyzed (n = 243) in general hospitals located in Northern Brazil were contaminated with bacterial pathogens. Other factors should also be considered, which include proximity of colonized and/or infected patients to the surfaces analyzed; the physical structure of the hospital; the adoption of antibiotic administration programs; as well as issues with the cleaning and disinfection protocols of surfaces and medical

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

10

justified [21]. The bacterial contamination present therein may result from the patient's

immunocompromised individuals [21].

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

S. aureus was the main microorganism recovered from the surfaces and equipment of both Hospital A (61.8%) and B (68.6%). In Hospital A, there was a predominance of S. aureus in door knobs in the ER (53.3%), heart monitors (83.3%) and medication tables (77.8%) in the AICU, and stethoscopes (100.0%) and mattresses (75.0%) in the PICU. In Hospital B, S. aureus was the main pathogen recovered from door knobs and bed headboards (66.7%), saline stands (100.0%) and mattresses (66.7%). These percentages are similar to those reported in the national literature [4] for mattresses (72.0%) and higher than international percentages [34] for beds (34%) and door knobs (26.0%). Our data show that highly touched surfaces are prone to be contaminated with S. aureus, a microorganism commonly found in the humans' hand microbiota. This suggests that the hands of health professionals were the main vector of contamination of surfaces in the units analyzed, although this was not investigated. Surfaces and equipment can be contaminated and, if not are properly disinfected, re-contaminate sanitized hands during the interruption of patient care to touch the surfaces and equipment such as keyboards, tables, door knobs, stethoscopes etc. The surfaces become permanently contaminated by pathogens [5,7,19] and the hospital environment turns into a reservoir of highly transmissible microorganisms. In our study, bedside tables of the PITU were found to be contaminated with CoNS (42.8%), although at a frequency lower than that of PICU clinicians' mobile phones in Kuwait (62.9%) [35]. CoNS are main cause of late-onset sepsis in developed countries³⁶ and early neonatal infections in Canadá [36], USA [37] and Brazil [38]. Despite the fact that CoNS are not considered pathogenic under normal circumstances, their presence on

objects with frequent manual contact (e.g., bedside tables and areas close to patients such as bed headboards and bed frame feet) may pose a risk of contamination and development of infection in hospitalized children. This fact requires attention by local health teams and indicates a need for revision of surface cleaning/disinfection protocols at the PICU in Hospital A.

Our findings showed a high percentage of methicillin-resistant isolates of CoNS and mostly *S. aureus*. The frequency of MRSA recovered from the ER (28.3%) was similar to that reported in the literature [12] (20.0%), while the frequency of MRSA recovered from the AICU (52.7%) was higher than that found in Brazil [39] (41.8%) and lower than international rates [40] (67.3%). The presence of MRSA in both hospitals analyzed reflects the persistence of this pathogen in the environment [34], as observed in another tertiary hospital in the Middle East in five rooms and two nursing stations in the AICU and PICU. MRSA strains often exhibit multidrug resistance, including resistance to beta-lactam antibiotics, and exhibit strong virulence as a result of a multiplicity of factors acting simultaneously to evade the host's defenses. Some of which include the production of enzymes and toxins, intracellular invasion and proliferation, dissemination into tissues and organs, and ability to form biofilms [4,27,41-43]. These virulence factors mediate bacterial adhesion to inert hospital surfaces and medical equipment, which may explain the high prevalence rates of MRSA found in this study, although biofilm production was not investigated herein.

The contamination of surfaces and equipment with Gram-negative bacilli is not as studied as the contamination with MRSA, VRE and *C. difficile* [3]. A study [44] reported the presence of MDR Enterobacteria in 22.2% of 18 sampled surfaces, which is consistent with our findings (19.44%) for the AICU of Hospital A. European data [45] showed a

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

high level of contamination by P. aeruginosa in sink isolates and tap biofilms. In line with this, our study recovered P. aeruginosa from 25% of sink samples, which attests that this pathogen is commonly associated with humid sites in the hospital environment and is the main microorganism isolated in ICUs [46]. In contrast, from a total of seventeen P. aeruginosa isolates found in this study, eleven were non-MDR isolates from door knobs of the ER, which is not a humid environment. This may be associated with the presence of *P. aeruginosa*-infected patients in beds near contaminated door knobs. Once more, this reinforces the understanding that health professionals' hands work as a vector of pathogen transmission. In our study, MDR P. aeruginosa isolates were recovered at a low frequency from AICU (2.78%) and PICU (2.04%) facilities, which differs from the literature showing 40.8% [47] prevalence of MDR *P. aeruginosa* in two hospitals in the West region of Santa Catarina state in Brazil. Even so, our results confirm the imminent risk of contamination of critically ill patients. In our study, we observed that the AICU of Hospital A was highly contaminated with MDR pathogens. This is in line with observational studies that recovered MDR microorganisms from critical care units [5,27,40]. The environmental contamination of ICUs has been a worrisome issue, since severely ill patients are prone to develop infections. It is fundamental to adopt standard measures to prevent and control hospitalacquired infection, considering the patients' risk of death, the proximity of the beds and the presence of monitoring and support equipment, which are highly touched and susceptible to contamination. It is also worth noting the widespread use of antibiotics in critical units, which select MDR clones both in the patient and in the environment, in addition to the fact that viable MDR bacteria have been isolated from biofilm surfaces

There was a high frequency of contaminated surfaces and equipment in our study,

Conclusions

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

with isolation of difficult-to-treat MDR phenotypes, e.g. MRSA, MR-CoNS, Enterobacteria resistant to 4th generation cephalosporins and MDR *P. aeruginosa*. These findings raise concern and point to the need to review the protocols of prevention and control of infections in the hospitals analyzed, mainly Hospital A. The cleaning/sanitizing of inert surfaces and equipment and antibiotic dispensing procedures should also be looked at in critical units, since antibiotics can act as a selective force of resistant bacterial strains. Routine and terminal disinfection of environmental surfaces and medical equipment with germicides is recommended by the literature [32] to decrease the frequency and level of contamination, especially highly touched surfaces near the patient. There is a need for greater attention into hand hygiene before and after contact with patients as much as with the areas close to them. These actions together could reduce contamination of hospital surfaces and equipment and the possibility of crosstransmission of pathogens while minimizing the risk of contamination of patients and the development of HAI. Taken altogether, the association between environmental contamination and the epidemiology of HAI is complex. This study is the pioneer in Western Bahia that describes the epidemiology of contamination of hospital surfaces, opportuning futures studies in this field. Further research should better understand the correlation of bacterial contamination with the underlying pathology of hospitalized patients close to

Acknowledgements

- We would like to thank to board directors from *Hospital Geral Público de Palmas* (HGPP)
- 350 [Palmas Public General Hospital], the board directors from *Hospital Municipal Eurico Dutra*
- 351 [Eurico Dutra Municipal Hospital] for their institutional consent.

References

337

338

339

340

341

342

343

344

345

346

347

348

352

- 1.Bonnet V, Dupont H, Glorion S, Aupée M, Kipnis E, Gérard JL, et al. Influence of
- bacterial resistance on mortality in intensive care units: a registry study from 2000 to
- 355 2013 (IICU Study). J Hosp Infect. 2019; 102 (3): 317-24.
- 2. Weber DJ, Rutala WA, Miller MB, Huslage K, Sickbert-Bennett E. Role of hospital
- surfaces in the transmission of emerging health care-associated pathogens: Norovirus,
- 358 Clostridium difficile, and Acinetobacter species. Am J Infect Control. 2010; 38 (5 Suppl
- 359 1): S25-33.

- 360 3. Weber DJ, Rutala WA. Understanding and Preventing Transmission of Healthcare-
- 361 Associated Pathogens Due to the Contaminated Hospital Environment.
- 362 Infect Control Hosp Epidemiol. 2013; 34 (5): 449-52.
- 4. Ferreira AM, Andrade D, Almeida MTG, Cunha KC, Rigotti MA. Colchões do tipo
- caixa de ovo: um reservatório de *Staphylococcus aureus* resistente à meticilina? Rev Esc
- 365 Enferm. 2011; 45: 161-6.
- 366 5. Russotto V, Cortegiani A, Raineri SM, Giarratano A. Bacterial contamination of
- inanimate surfaces and equipment in the intensive care unit. J Intensive Care. 2015; 3:54-
- 368 61.
- 6. Trindade RC, Bonfim ACR, Resende MA. Conjuntivais flora microbiana de pessoas
- 370 clinicamente normais que trabalham em um ambiente hospitalar. Braz J Microbiol. 2000;
- 371 31 (1): 12-6.
- 372 7. Russotto V, Cortegiani A, Fasciana T, Iozzo P, Raineri SM, Gregoretti C, et al. What
- 373 Healthcare Workers Should Know about Environmental Bacterial Contamination in the
- 374 Intensive Care Unit. Biomed Res Int. 2017; 2017: 6905450.
- 8. Boyce JM. Environmental contamination makes an important contribution to hospital
- infection. J Hosp Infect. 2007; 65 (Suppl 2): 50-4.
- 9. Rossi D, Devienne KF, Raddi MSG. Influência de fluídos biológicos na sobrevivência
- de *Staphylococcus aureus* sobre diferentes superfícies secas. Rev Ciênc Farm Básica Apl.
- 379 2008; 29: 211-4.
- 380 10. Adams CE, Smith J, Watson V, Robertson C, Dancer SJ. Examining the association
- between surface bioburden and frequently touched sites in intensive care. J Hosp Infect.
- 382 2017; 95 (1):76–80.

- 11. Shams AM, Rose LJ, Edwards JR, Cali S, Harris AD, Jacob JT, et al. Assessment of
- the Overall and Multidrug-Resistant Organism Bioburden on Environmental Surfaces in
- Healthcare Facilities. Infect Control Hosp Epidemiol. 2016; 37 (12):1426-32.
- 12. Silva AS, Deuschle RAN, Garlet CCM. Pesquisa de Staphylococcus aureus nas
- maçanetas das portas dos quartos de um hospital na região Noroeste, Rio Grande do Sul
- 388 Rev Saúde (Santa Maria). 2012; 38 (1):115-24.
- 389 13. Oliveira AC, Damasceno QS. Superfícies do ambiente hospitalar como possíveis
- reservatórios de bactérias resistentes: uma revisão. Rev Esc Enferm. 2010; 44 (4): 1118-
- 391 23.
- 392 14. Caetano JA, Lima MA, Miranda MC, Serufo JC, Ponte PRL. Identificação de
- 393 contaminação bacteriana no sabão líquido de uso hospitalar. Rev Esc Enferm. 2011; 45
- 394 (1):153-60.
- 15. Costa DM, Johani K, Melo DS, Lopes LK, Lima LKOL, Tipple AFV, et al. Biofilm
- 396 contamination of high-touched surfaces in intensive care units: epidemiology and
- potential impacts. Lett Appl Microbiol. 2019; 68 (4): 267-8.
- 398 16. Galvin S, Dolan A, Cahill O, Daniels S, Humphreys H. Microbial monitoring of
- the hospital environment: why and how? J Hosp Infect. 2012; 82 (3):143-51.
- 400 17. Sexton T, Clark P, O'Neill E, Dillane T, Humphreys H. Environmental reservoirs of
- 401 methicillin-resistant *Staphylococcus aureus* in isolation rooms: correlation with patient
- isolates and implications for hospital hygiene. J Hosp Infect. 2006; 62 (2):187-94.
- 403 18. Dancer CJ. Importance of the environment in methicillin resistant *Staphylococcus*
- 404 aureus acquisition: the case for hospital cleaning. Lancet Infect Dis. 2008; 8 (2):101-13.

- 405 19. Hayden MK, Blom DW, Lyle EA, Moore CG, Weistein RA. Risk of hand or glove
- 406 contamination after contact with vancomycin resistant Enterococcus or the colonized
- patients environment. Infect Control Hosp Epidemiol. 2008; 29 (2):149-54.
- 408 20. Drees M, Snydman DR, Schmid CH, Barefoot L, Hansjosten K, Vue PM, et al. Prior
- 409 environmental contamination increases the risk of acquisition of vancomycin-resistant
- 410 enterococci. Clin Infect Dis. 2008; 46 (5):678-85.
- 411 21. Koneman EW, Allen SD, Janda WM, Schreckenberger PC, Winn WC. Diagnóstico
- 412 microbiológico: texto e atlas colorido. 8. ed. Rio de Janeiro: Guanabara Koogan; 2010,
- 413 1565 p.
- 414 22. Clinical and Laboratory Standards Institute (CLSI). M100-S24 Performance
- 415 Standards for Antimicrobial Susceptibility Testing; Twenty-Fourth Informational
- 416 Supplement. January 2014.
- 417 23. Siegel JD, Rhinehart E, Jackson M, Chiarello L. Management of multidrug-resistant
- organisms in health care settings, 2006. Am J Infect Control. 2007; 35(10 Suppl 2): S165-
- 419 93.
- 420 24. Hidron AI, Edwards JR, Patel J, Horan TC, Sievert DM, Pollock DA.
- NHSN annual update: Antimicrobial-resistant pathogens associated with healthcare-
- associated infections: annual summary of data reported to the National Healthcare Safety
- Network at the Centers for Disease Control and Prevention, 2006–2007. Infect Control
- 424 Hosp Epidemiol. 2008; 29 (11): 996-1011.

- 425 25. Esteves DC, Pereira VC, Souza JM, Keller R, Simões RD, Winkelstroter Eller LK,
- et al. Influence of biological fluids in bacterial viability on different hospital surfaces and
- 427 fomites. Am J Infect Control. 2016; 44 (3): 311-4.
- 428 26. Moraes CL, Ribeiro NFG, Costa DM, Furlan VG, Palos MAP, Vasconcelos LSNOL.
- 429 Contaminação de equipamentos e superfícies de unidades de terapia intensiva de uma
- 430 maternidade pública por *Staphylococcus* coagulase negativa. Rev Patol Trop. 2013; 42
- 431 (4): 387-94.
- 432 27. Johani K, Abualsaudc D, Costa DM, Hua H, Whiteleye G, Deva A, et al.
- 433 Characterization of microbial community composition, antimicrobial resistance and
- biofilm on intensive care surfaces. J Infect Public Health. 2018; 11: 418-24.
- 28. Carling PC, Parry MF, von Beheren SM. Healthcare Environ-mental Hygiene Study
- 436 Group. Identifying opportunities to enhance environmental cleaning in 23 acute care
- hospitals. Infect Control Hosp Epidemiol. 2008; 29 (17): 1-7.
- 438 29. Goodman ER, Platt R, Bass R, Onderdonk AB, Yokoe DS, Huang SS. Impact of
- 439 environmental cleaning intervention on the presence of methicillin-resistant
- 440 Staphylococcus aureus and vancomycin-resistant enterococci on surfaces in intensive
- care unit rooms. Infect Control Hosp Epidemiol. 2008; 29: 593-9.
- 30. Rutala WA, Weber DJ. Healthcare Infection Control Practices Advisory Committee
- 443 (HICPAC). Guideline for Disinfection and Sterilization in Healthcare Facilities, 2008.
- Department of health and human services USA. Centers for diseases control; 2017 Feb.
- 445 161p.
- 31. Lessa FC, Gould CV, McDonald LC. Current status of *Clostridium difficile* infection
- epidemiology. Clin Infect Dis. 2012; 55 Suppl 2: S65-70.

- 448 32. Cortegiani A, Russotto V, Maggiore A, Attanasio M, Naro AR, Raineri SM, et al.
- 449 Antifungal agents for preventing fungal infections in non-neutropenic critically ill
- patients. Cochrane Database of Systematic Reviews. 2016; Issue 1. Art. No.: CD004920.
- 451 33. Ledwoch K, Dancer SJ, Otter JA, Kerr K, Roposte D, Rushton L, et al. Beware
- biofilm! Dry biofilms containing bacterial pathogens on multiple healthcare surfaces; a
- 453 multi-centre study. J Hosp Infect. 2018; 100 (3): 47-56.
- 454 34. Nkuwi EJ, Kabanangi F, Joachim A, Rugarabamu S, Majigo M. Methicillin-resistant
- 455 Staphylococcus aureus contamination and distribution in patient's care environment at
- 456 Muhimbili National Hospital, Dar es Salaam-Tanzania. BMC Res Notes. 2018; 11(1):
- 457 484.
- 458 35. Heyba M, Ismaiel M, Alotaibi A, Mahmoud M, Baqer H, Safar A, et al.
- 459 Microbiological contamination of mobile phones of clinicians in intensive care units and
- neonatal care units in public hospitals in Kuwait. BMC Infect Dis. 2015; 15: 434-43.
- 36. Sgro M, Shah PS, Campbell D, Tenuta A, Shivananda S, Lee SK. Early-onset neonatal
- sepsis: rate and organism pattern between 2003 and 2008. J Perinatol. 2011; 31(12):794–
- 463 8.
- 464 37. Edwards RK, Jamie WE, Sterner D, Gentry S, Counts K, Duff P. Intrapartum
- antibiotic prophylaxis and early-onset neonatal sepsis patterns. Infect Dis Obstet Gynecol.
- 466 2003;11(4): 221-6.
- 38. Pereira CA, Marra AR, Camargo LF, Pignatari AC, Sukiennik T, Behar PR, et al.
- 468 Nosocomial Bloodstream Infections in Brazilian Pediatric Patients: Microbiology,
- 469 Epidemiology, and Clinical Features. PLoS One. 2013; 8 (7): e68144

- 470 39. Campos GB, Souza SG, Lobão TN, Silva DCC, Sousa DS, Oliveira OS, et al.
- 471 Isolation, molecular characteristics and disinfection of methicillin-resistant
- 472 Staphylococcus aureus from ICU units in Brazil. New Microbiol. 2012; (35):183-90.
- 40. Tajeddina E, Rashidana M, Razaghi M, Javadi SSS, Sherafat SJ, Alebouyeha M, et
- al. The role of the intensive care unit environment and health-care workers in the
- 475 transmission of bacteria associated with hospital acquired infections. J Infect Public
- 476 Health. 2016; 9: 13-23.
- 41. Rosenthal VD, Bijje H, Maki DG, Mehta Y, Apisarnthanarak A, Medeiros EA, et al.
- 478 International Nosocomial Infection Control Consortium (INICC) report, data summary
- of 36 countries, for 2004-2009. Am J of Infect Control. 2012; 40 (5): 396-407.
- 42. Laabei M, Recker M, Rudkin JK, Aldeljawi M, Gulay Z, Sloan TJ, et al. Predicting
- the virulence of MRSA from its genome sequence. Genome Res. 2014; 24 (5): 839-49.
- 43. Heilmann C. Adhesion mechanisms of staphylococci. Adv Exp Med Biol. 2011; 715:
- 483 105-23.
- 484 44. Judge C, Galvin S, Burke L, Thomas T, Humphreys H, Fitzgerald- Hughes D. Search
- and you will find: detecting extended-spectrum b-lactamase-producing Klebsiella
- 486 pneumoniae from a patient's im-mediate environment. Infect Control Hosp Epidemiol.
- 487 2013; 34: 534-6.
- 488 45. Abreu PM, Farias PG, Gabriel Silva Paiva GS, Almeida AM, Morais PV. Persistence
- of microbial communities including *Pseudomonas aeruginosa* in a hospital environment:
- a potential health hazard. BMC Microbiol. 2014, 14:118-27.
- 491 46. Ferrareze MVG, Leopoldo VC, Andrade D, Silva MFI, Haas VJ. Pseudomonas
- 492 *aeruginosa* multirresistente em unidade de cuidados intensivos: desafios que procedem?
- 493 Acta Paul Enferm. 2007; 20 (1): 7-11.

501

Figure 1. Frequency of microorganisms recovered from inert surfaces and medical equipment in the Hospital Geral Público de Palmas-TO, Brazil (A) and Hospital Municipal Eurico Dutra in the city of Barreiras, BA, Brazil (B).

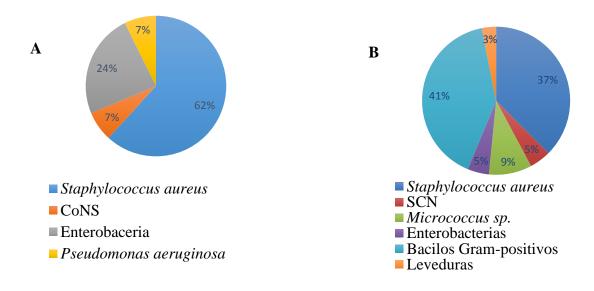


Table 1. Frequency of clinically important microorganisms isolated from inert surfaces and medical equipment in the Hospital Geral Público de Palmas -TO, Brazil (A).

Hospital units	Surfaces (N)	S.aureus N (%)	CoNS N (%)	Enterobacteria N (%)	P. aeruginosa N (%)	Total of bacteria (N)
ER	door knobs (56)	49 (53,3)	4 (4,3)	28 (30,4)	11 (11,9)	92
OR	door knobs (20)	12 (60,0)	2 (10,0)	5 (25,0)	1 (5,0)	20
AICU	heart monitors (18)	15 (83,3)	-	3 (16,7)	-	18
	infusion pumps (18)	13 (72,2)	-	3 (16,7)	2 (11,1)	18
	medication tables	14 (77,8)	-	4 (22,2)	-	18
	(18)					
	side bed frames (18)	11 (61,1)	-	6 (33,3)	1 (5,6)	18
PICU	side bed frames (16)	8 (57,1)	1 (7,4)	5 (35,7)	-	14
	bed headboards (8)	3 (50,0)	2 (33,3)	1(16,7)	-	6
	bed frame feet (8)	3 (50,0)	2 (33,3)	-	1 (16,7)	6
	mattresses (8)	3 (75,0)	1 (25,0)	-	-	4
	bedside tables (8)	3 (42,8)	3(42,8)	1(14,3)	-	7
	stethoscopes (8)	8 (100,0)	-	-	-	8
	samples from the sinks (4)	2(50,0)	1(25,0)		1(25,0)	4
Total (N)	208	144 (61,8)	16 (6,9)	56 (24,0)	17 (7,3)	233

N: total sample size; %: Percentage; ER: Emergency Room; OR: Operating Room; AICU:

Adult Intensive Care Unit; PICU: Pediatric Intensive Care Unit; CoNS: Coagulase-Negative Staphylococci.

Table 2. Frequency of clinically important microorganisms isolated from inert surfaces and medical equipment in the Hospital Municipal Eurico Dutra in the city of Barreiras, BA, Brazil (B).

Hospital units	Surfaces (N)	S. aureus N (%)	CoNS N (%)	Enterobacteria N (%)	Micrococcus N (%)	Total of bacteria (N)
IMW	bed headboard (3)	2 (66,7)	-	1 (33,3)	-	3
	mattresses (2)	1 (50,0)	-	-	1 (50,0)	2
	side bed frame (2)	1 (50,0)	-	-	1 (50,0)	2
	saline stand (1)	-	-	-	1 (100,0)	1
	door knobs (3)	2 (66,7)	-	1 (33,3)	-	3
\mathbf{SW}	bed headboard (5)	3 (60,0)	1 (20,0)	1 (20,0)	-	5
	mattresses (3)	2 (66,7)	-		1(33,3)	3
	side bed frame (1)	1 (100,0)	-	-	-	1
	saline stand (3)	3 (100,0)	-	-	-	3
	countertops (1)	1 (100,0)	-	-	-	1
	door knobs (4)	2 (50,0)	1 (25,0)	-	1 (25,0)	4
OR	surgical light - BU (2)	2 (100,0)	-	-	-	2
	surgical light - AU (2)	2 (100,0)	-	-	-	2
	stretcher - BU (1)	1 (100,0)	-	-	-	1
	stretcher - AU (2)	1 (50,0)	1 (50,0)	-	-	2
otal (N)	35	24 (68,6)	3 (8,6)	3 (8,6)	4 (11,4)	35

N: total sample size; %: Percentage; IMW: Internal Medicine Ward; SW: Surgical Ward;

OR: Operating Room; BU: Before Use; AU: After Use; CoNS: coagulase-negative staphylococci.

Table 3. Multi-drug resistant microorganisms recovered from inert surfaces and equipment of hospitals located in Northern Brazil.

Hospital	Hospital units	Number of samples	Culture positive	Microorganisms (N)	MDR* isolated (Frequency)
	ER	56	92	S.aureus (49)	MRSA (28,3%)
				CoNS (4)	MR CoNS (4,3%)
				Enterobacteria (28)	Enterobacteria resistant to 4 th generation
				P.aeruginosa (11)	cephalosporins (8,7%)
-	OR	20	20	S.aureus (12) CoNS (2)	MRSA (25,0%) MR CoNS (5,0%)
				Enterobacteria (5) P.aeruginosa (1)	- -
_	AICU	72	72	S.aureus (53)	MRSA (52,7%)
Hospital A				Enterobacteria (16)	Enterobacteria resistant to 4 th generation
(Palmas-TO)				P.aeruginosa (3)	cephalosporins (19,44%) multi-resistant P.aeruginosa (2,78%)
•	PICU	60	49	S.aureus (30) CoNS (10)	MRSA (4,08%) MR CoNS (20,40%)
				Enterobacteria (7)	Enterobacteria resistant to 4 th generation
				P.aeruginosa (2)	cephalosporins (6,12%) multi-resistant P.aeruginosa (2,04%)
	IMW	11	11	S.aureus (6)	MRSA (27,27%)
				Enterobacteria (2)	-
				Micrococcus. (3)	-
Hospital B	SW	17	17	S.aureus (13)	MRSA (29,4%)
(Barreiras-BA)				CoNS (2) Enterobacteria (1)	MR CoNS (5,88%)
				Micrococcus(1)	-
-	OR	7	7	S.aureus (6) CoNS (1)	MRSA (14,3%)

^{*}Multidrug-resistant pathogen; ER: Emergency Room; OR: Operating Room; AICU: Adult Intensive Care Unit; PICU: Pediatric Intensive Care Unit; IMW: Internal Medicine Ward; SW: Surgical Ward; MRSA: Methicillin-Resistant *Staphylococcus aureus*; MR-CoNS: Methicillin-Resistant Coagulase-Negative Staphylococci.