A consensus phylogenomic approach highlights the ancient rapid radiation of Ericales

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Running Head: A consensus approach to resolving the Ericales phylogeny

ABSTRACT

Premise of study: Large genomic datasets offer the promise of resolving historically recalcitrant

phylogenetic problems. However, different methodologies can yield conflicting results,

especially when diversification occurs rapidly. Here, we used an array of dataset filtering

strategies and species tree methods to infer a consensus topology of Ericales and explored

sources of uncertainty associated with an ancient radiation.

Methods: We used a hierarchical clustering approach, along with tree-based homology and

orthology detection, to generate multiple phylogenomic datasets. Support for species

relationships was inferred from multiple lines of evidence including shared gene duplications,

gene tree conflict, gene-wise edge-based analyses, concatenation, and coalescent-based species

tree methods and summarize these results in a consensus framework.

Key Results: Our consensus approach supported a topology largely concordant with the current

theorized relationships, but suggests that the data are not capable of resolving several early

relationships due to lack of informative characters, sensitivity to methodology, and extensive

gene tree conflict. We find evidence of one or more paleopolyploidy events before the radiation

of ericalean families that likely contributes to the high levels of gene tree conflict observed.

*Conclusions***:** Our approach provides a novel hypothesis regarding the history of Ericales and

confidently resolves several nodes. However, we also demonstrate that some ancient divergences

are unresolvable with our current data. Whether this is because of rapid and unresolvable ancient

speciation or lack of data needs to be addressed with additional data collection efforts. The role

that paleopolyploidy plays in generating gene tree conflict warrants further investigation.

Keywords: consensus topology; Ericales; gene duplication; gene tree conflict; multifurcation,

phylogenetic uncertainty; phylogenomics; polytomy; rapid radiation; whole genome duplication

INTRODUCTION

The flowering plant clade Ericales contains several ecologically important lineages that shape the structure and function of ecosystems including tropical rainforests (e.g. Lecythidaceae, Sapotaceae, Ebenaceae), heathlands (e.g. Ericaceae), and open habitats (e.g. Primulaceae) around the globe (ter Steege et al., 2006; Hedwall et al., 2013; He et al., 2014; Memiaghe et al., 2016; Moquet et al., 2017). With 22 families comprising ca. 12,000 species (Chase et al., 2016; Stevens, 2001 onward), Ericales are a diverse and disparate clade with an array of economically and culturally important plants. These include agricultural crops such as blueberries (Ericaceae), kiwifruits (Actinidiaceae), sapotas (Sapotaceae), Brazil nuts (Lecythidaceae), and tea (Theaceae) as well as ornamental plants such as cyclamens and primroses (Primulaceae), rhododendrons (Ericaceae), and phloxes (Polemoniaceae). The group has also given rise to multiple parasitic lineages and the carnivorous American pitcher plants (Sarraceniaceae). Although Ericales has been a well-recognized clade throughout the literature (Chase et al., 1993; Anderberg et al., 2002; Schönenberger et al., 2005; Rose et al., 2018), the evolutionary relationships among major clades within Ericales remain contentious.

One of the first molecular studies investigating these deep relationships used three plastid and two mitochondrial loci; the authors concluded that the dataset was unable to resolve major familial relationships (Anderber et al., 2002). These relationships were revisited with 11 loci (two nuclear, two mitochondrial, and seven chloroplast), where maximum parsimony and Bayesian analyses provided support for the resolution of several early diverging lineages (Schönenberger et al., 2005). Recent work employing three nuclear, nine mitochondrial, and 13 chloroplast loci in a concatenated supermatrix consisting of 49,435 aligned sites and including 4,531 ericalean species but with 87.6% missing data, provided the most statistically robust

hypothesis to date (Rose et al., 2018). Despite the broad sampling utilized in Rose et al. (2018),

many relationships remain poorly supported, including several deep divergences that seem to be

the result of an ancient, rapid radiation. Moreover, these and other studies have recovered several

conflicting interfamilial relationships, highlighting the need to investigate possible biological and

methodological explanations for this topological incongruence (Anderber et al., 2002; Bremer et

al., 2002; Schönenberger et al., 2005).

Large genomic datasets can be used to help understand conflicting species tree results

and as these data have become more affordable, it is possible to conduct thorough investigations

of elusive relationships across the Tree of Life. Yet, despite the increasing availability of

genome-scale datasets, many relationships remain controversial, as research groups recover

different answers for the same evolutionary questions, often with seemingly strong support (Shen

et al., 2017). One benefit of genome-scale data for phylogenetics (i.e. phylogenomics) is the

ability to examine conflicting signal within and among datasets and a key finding in the

phylogenomics literature has been the high prevalence of gene tree conflict at contentious nodes

(Brown et al. 2017b, Reddy et al., 2017; Vargas et al., 2017; Walker et al., 2018a). The

differences seen across previous analyses may in fact be the result of real biological conflict

among the molecular data.. However, biological sources of conflict (e.g. hybridization,

incomplete lineage sorting, horizontal gene transfer) can also provide valuable information

regarding the evolutionary history of lineages. By identifying this conflict, it becomes possible to

locate where focused analyses are warranted and future sampling efforts might prove useful. One

prominent method for examining genomic conflict is analyzing the relationships among gene

trees, which may be obtained through a variety of methods including from transcriptome data

(Yang and Smith, 2014). Transcriptomes provide data on hundreds to thousands of coding

sequences per sample, and have elucidated many of the most contentious relationships in the

green plant phylogeny (e.g. Simon et al., 2012; Wickett et al., 2014; Walker et al., 2017).

Transcriptomes also provide the necessary information to identify gene and genome duplications.

Gene duplications may be associated with important molecular evolutionary events but also

represent a potentially informative event in a lineage's history and shared duplications can

therefore provide additional evidence for how species are related. These multiple lines of

evidence regarding evolutionary history can be applied as metrics of support to generate robust

phylogenetic hypotheses.

In this study, we sought to understand the evolutionary history of Ericales and asked

whether the available genome-scale data robustly supported the resolution of deep relationships

in the clade. We leveraged coding sequences from hundreds/thousands of genes to infer major

evolutionary events, such as ancient gene/genome duplication, rates of evolution, and conflicting

signal among genes. In applying a consensus approach that considers several methodological

alternatives in an effort to resolve the phylogeny, we allow the possibility that methods might

disagree and that there may not be the data available to support a single, bifurcating species tree

may. While more data may be available, in cases where whole genomes and transcriptomes have

been sequenced, a polytomy may represent a more accurate representation of the evolutionary

history of a clade than a fully bifurcating species tree. Our investigation of the evolution of the

Ericales with genomic data is particularly well-suited to instantiate a conversation about when

polytomies may be considered and to what extent biological polytomies occur across the Tree of

Life.

MATERIALS AND METHODS

Taxon sampling and de novo assembly procedures—We assembled a dataset consisting of

coding sequence data from 97 genomes and transcriptomes from the 1KP project and NCBI

databases (Appendix S1; see Supplemental Data with this article). Samples assembled from raw

reads were done so with Trinity version 2.5.1 using default parameters and the "--trimmomatic"

option (Kodama et al. 2011; Grabherr et al., 2011; Matasci et al., 2014). Assembled reads were

translated using TransDecoder v5.0.2 (Haas et al., 2013) with the option to retain blast hits to a

custom protein database consisting of *Beta vulgaris*, *Arabidopsis thaliana*, and *Daucus carota*

obtained from Ensembl plants (https://plants.ensembl.org/). The translated amino acid sequences

from each assembly were reduced by clustering with CD-HIT version 4.6 with settings -c 0.995 -

n 5 (Fu et al., 2012). As a quality control step, the chloroplast genes *rbcL* and *matK* were

extracted from each assembly using a custom script (available on Dryad [TBD]) and then queried

using BLAST against the NCBI online database (Altschul et al., 1990). In cases where the top

hits were to a species not closely related to that of the query, additional sequences were

investigated and if contamination, misidentification, or other problems seemed likely, the

transcriptome was not included in further analyses or homolog clustering. The final

transcriptome sampling included 86 ingroup taxa spanning 17 of the 22 families within Ericales

as recognized in APG VI (Appendix S1; Chase et al., 2016).

Homology inference—We used the hierarchical clustering procedure from Walker et al. (2018b)

for homology identification. In short, the method involves performing all-by-all blast procedure

on user-defined clades; homolog clusters identified within each clade are then combined

recursively with clusters of a sister clade based on sequence similarity until clusters from all

clades have been combined. To distribute taxa among groups for clustering we estimated a

species tree with RAxML v8.2.12 (Stamatakis, 2014) using a concatenated matrix of *rpoC2*,

rbcL, *nhdF*, and *matK*, identified in the quality control step. The resulting tree was used to

manually assign taxa to one of eight clades for initial homolog clustering (Appendix S2).

Homolog clustering within each group was performed following the methods of Yang and Smith

(2014).

Nucleotide sequences for the inferred homologs within each group were aligned with

MAFFT v7.271 (Katoh et al., 2002; Katoh and Standley, 2013) and trimmed with the phyx

command pxclsq (Brown et al., 2017a) such that columns with less than 10% occupancy were

removed. Homolog trees were estimated with RAxML, unless the homolog had >500 tips, in

which case FastTree v2.1.8 was used (Price et al., 2010). Tips with branch lengths longer than

1.5 substitutions/site were trimmed because of their potential to reflect misidentified homology

(Yang and Smith, 2014). When a clade is formed of sequences from a single taxon, it likely

represents either in-paralogs or alternative splice sites and since neither of these provide

phylogenetic information, we retained only the tip with the most character rich sequence data.

This procedure was repeated with refined clusters twice using the same settings. Homologs

between each of the eight groups were then combined (https://github.com/jfwalker/Clustering)

and homolog trees were again estimated and refined with the same settings except that internal

branches longer than 1.5 subs/site were also cut—again to reduce the possibility of misidentified

homology. Trees were re-estimated and refined by cutting terminal branches longer than 0.8

substitutions/site and internal branches longer than 1.0 substitutions/site. The final homolog set

contained 9469 clusters.

Initial ortholog identification and species tree estimation—Putative orthologs were extracted

from homolog trees using the rooted tree (RT) method which allows for robust orthology

detection even after genome duplications (Yang and Smith, 2014). Orthologs were required to

have at least 50 taxa and seven cornalean taxa as well as *Arabidopsis thaliana* and *Beta vulgaris*

were used as outgroups (Appendix S1). Orthologs were not extracted from homolog groups with

more than 5000 tips because of the uncertainty in reconstructing very large homolog trees

(Walker et al., 2018b). Here, we use the term ortholog to describe clusters of sequences that have

been inferred to be monophyletic based on their position within inferred homolog trees after

accounting for gene duplication. This usage reflects our interest in addressing phylogenetic

questions, rather than those of gene functionality. The tree-aware ortholog identification

employed here provides the best defense available against misidentified orthology that could

mislead phylogenetic analyses (Eisen, 1998; Gabaldón, 2008; Brown and Thomson, 2016).

The resulting ortholog trees were then filtered to require at least one euasterid taxon,

Helianthus annuus or *Solanum lycopersicum*, for use as outgroups, and if both were present the

two were required to be monophyletic. This was done to ensure that orthologs could be rooted in

subsequent analyses. Terminal branches longer than 0.8 were again trimmed, resulting in a

refined dataset containing 387 orthologs. Final nucleotide alignments were estimated with

PRANK v.150803 (Löytynoja, 2014) and cleaned for a minimum occupancy of 30% occupancy

using the pxclsq function in phyx. Alignments for the 387 orthologs were concatenated using

pxcat in phyx and a maximum likelihood (ML) species tree and rapid bootstrap support (200

replicates) with was inferred using RAxML v8.2.4 with the command raxmlHPC-PTHREADS

and a separate GTRCAT model of evolution estimated for each partition; this resulted in a

topology we refer to as the maximum likelihood topology (MLT). In separate runs, 200 regular

bootstraps with and without the MLT as a starting tree were conducted using the -b option. Trees

for each ortholog were also estimated individually using RAxML, with the GTRCAT model of

evolution and 200 rapid bootstraps. A coalescent-based maximum quartet support species tree

(MQSST) was estimated using ASTRAL v5.6.2 (Zhang et al., 2018) with the resulting 387

ortholog trees. In order to investigate the possible effect of ML tree search algorithm on

phylogenetic inference, an ML tree for the 387 ortholog supermatrix was estimated using

RAxML v8.2.11 and the command raxmlHPC-PTHREADS-AVX and with IQ-TREE v1.6.1

with -m GTR+Γ -n 0 and model partitions for each ortholog specified with the -q option.

Gene-wise log-likelihood comparisons—Gene-wise likelihood support for the MLT and an

alternative backbone topology recovered by the consensus all 200 rapid bootstraps (i.e. the rapid

bootstrap topology, RBT) were investigated using a two-topology analysis (Shen et al., 2017;

Walker et al., 2018a). A three-topology comparison was also conducted by calculating the gene-

wise log-likelihoods of a third topology where the MLT was modified such that the inter-familial

backbone relationships were those recovered in the ASTRAL topology (AT). This constructed

tree was used in lieu of the actual ASTRAL topology to ensure that the within-family topological

differences did not affect the likelihood calculations. In both tests, the ML score for each gene is

calculated while constraining the topology under multiple alternatives. Branch lengths are

optimized for each input topology and GTR+Γ is optimized for each supermatrix partition for

each topology (Shen et al., 2017; Walker et al., 2018a). Results from both comparisons were

visualized using a custom R script.

An edge-based analysis was performed to compare the likelihoods of competing

topologies while allowing gene tree conflict to exist outside the relationship of interest (Walker

et al., 2018a). Our protocol was similar to that of Walker et al. (2018a), except that instead of a

defined "TREE SET", we used constraint trees with clades defining key bipartitions

corresponding to each of three competing topologies using the program EdgeTest.py (Walker et

al., 2019). The likelihood for each gene was calculated using RAxML-ng (Kozlov et al., 2019)

with the GTR+Γ model of evolution, using the brlopt nr_safe option and an epsilon value of

0.000001 while a given relationship was constrained. The log-likelihood of each gene was then

summed to give a likelihood score for that relationship. An additional edge-based analysis was

conducted to investigate the placement of Ebenaceae using the program Phyckle (Smith et al.,

2018), which uses a supermatrix and a set of constraint trees specifying conflicting relationships

and reports, for each gene, the ML as well as the difference between the best and second-best

topology. This allows quantification of gene-wise support at a single edge as well as a metric of

how strongly the relationships is supported in terms of likelihood.

Gene duplication comparative analysis—Homolog clusters were filtered such that sequences

shorter than half the median length of their cluster and clusters with more than 4000 sequences

were removed. Homolog trees were then re-estimated with IQ-TREE with the GTR+Γ model and

SH-aLRT support followed by cutting of internal branches longer than 1.0 and terminal branches

longer than 0.8 inferred substitutions per base pair. Rooted ingroup clades were extracted with

the procedure from Yang et al. (2015) with all non-ericalean taxa as outgroups and gene

duplications were inferred with the program phyparts, requiring at least 50% SH-aLRT support

in order to avoid the inclusion of very poorly supported would-be duplications (Smith et al.,

2015). By mapping gene duplications in this way to competing topological hypotheses (i.e. the

MLT, RBT, and AT), as well as several hypothetical topologies not recovered, we determined

the number of duplications uniquely shared among several key ericalean clades.

Owing to how duplications are inferred using tree-based methods, the location to which

duplications are mapped depends on the topology of the phylogeny; a gene duplication can

appear along a branch other than that which the homolog trees actually contains evidence

231 because duplications are mapped to the bipartition in the species tree that contains all the taxa

232 that share a given duplication in a homolog tree. For example, if in a homolog tree, taxon A and

233 taxon B share a gene duplication, but in the species tree taxon C is sister to taxon A and taxon B

234 sister to those, then the duplication will be mapped to the branch that includes all three taxa,

235 because the duplication is mapped to the smallest clade that includes both taxon A and taxon B.

236 A differently specified topology can allow the relevant duplication to instead be mapped to the

237 correct, more exclusive branch that specifies the clade for which there is evidence of that

238 duplication in the homolog tree (i.e. only taxa A and B in the previous example). Once it has

239 been determined how many duplications are actually present in the homolog trees, comparisons

240 between competing relationships can be made. It is important to note that incomplete taxon

241 sampling and other biases should be considered when applying such a comparative test of gene

242 duplication number. Assuming there are duplicated genes present in the taxa of interest, clades

243 with a greater number of sampled taxa or more complete transcriptomes will likely share more

244 duplications simply due to the fact that more will have been sampled in the dataset. Furthermore,

245 the location to which a duplication is mapped will depend on which assemblies contain that

246 transcript, whether or not that gene is present in the genome of any particular organism.

247 Therefore, we used gene duplications shared amongst clades as an additional, relative metric of

248 topological support capable of corroborating other results (i.e. that if clades share many gene

249 duplications unique to them, they are more likely to be closely related), while recognizing that

250 the absolute number of duplications shared by various clades are impacted by imperfect

251 sampling.

252 ***Synthesizing support for competing topologies***—Support from each of the main tree hypotheses

253 (MLT, RBT and AT) was reviewed and the most commonly supported backbone of familial

relationships was determined. Because the comparative gene duplication analysis and constraint

tree analyses both supported the RBT over other candidates, this corroborated topology was the

focus of additional measures of support. Quartet support was assessed on the RBT using the

program Quartet Sampling with 1000 replicates (Pease et al., 2018). We used this procedure to

measure quartet concordance (QC), quartet differential (QD), quartet informativeness (QI), and

quartet fidelity (QF). Briefly, QC measures how often the concordant relationship is recovered

with respect to other possible relationships, QD helps identify if a relationship has a dominant

alternative, and QI correspond to the ability of the data to resolve a relationship of interest, where

a quartet that is at least 2.0 log-likelihood (LL) better than the alternatives is considered

informative. Finally, QF aids in identifying rogue taxa (Pease et al., 2018). Gene conflict with

the corroborated topology was assessed using the bipartition method as implemented in phyparts

using gene trees for the 387 orthologs, which were first rooted on the outgroups *Helianthus*

annuus and *Solanum lycopersicum* using a ranked approach with the phyx command pxrr. Gene

conflict was assessed both requiring node support (BS≥70) and without any support

requirement; the support requirement should help reduce noise in the analysis, but we also ran

the analysis without a support requirement to ensure that potentially credible (but only partially-

supported) bipartitions were not overlooked. Results for both were visualized using the program

phypartspiecharts (https://github.com/mossmatters/phyloscripts/).

Expanded ortholog datasets—In order to explore the impact of dataset construction, orthologs

were inferred from the homolog trees as for the 387 ortholog set but with modifications to taxon

requirements and refinement procedures described below. For each dataset, sequences were

aligned separately with both MAFFT and PRANK and cleaned for 30% occupancy. A

supermatrix was constructed with each and an ML tree was estimated with IQ-TREE with and

without a separate GTR+Γ model partition for each ortholog. Individual ortholog trees were

estimated with RAxML and used to construct an MQSST with ASTRAL.

2045 ortholog set—Orthologs were filtered such that there was no minimum number of taxa and

at least two tips from each of the following five groups: 1) Primulaceae; 2) Polemoniaceae and

Fouquieriaceae; 3) Lecythidaceae; 4) outgroups including *Solanum* and *Helianthus* as well as

Marcgraviaceae and Balsaminaceae (i.e. the earliest diverging ericalean clade in all previous

analyses); and 5) all other taxa. This resulted in a dataset with 2045 orthologs. Ortholog tree

support for conflicting placements of Ebenaceae was assessed for the PRANK-aligned orthologs

using Phyckle.

4682 ortholog set—Orthologs were not filtered for any taxon requirements. Sequences were

aligned with MAFFT, ortholog trees were estimated with RAxML and terminal branches longer

than 0.8 were trimmed. Sequences were then realigned separately with MAFFT and PRANK and

cleaned as before.

1899, 661, and 449 ortholog sets—To assess the effect of requiring *Helianthus* and *Solanum*

outgroups in the 387 ortholog set and to further explore the effect of taxon requirements on the

inferred topology, each homolog tree that produced an ortholog in that dataset was re-estimated

in IQ-TREE with SH-aLRT support. Calculating this support allowed visual assessment of

whether uncertain homolog tree construction was affecting the ortholog identification process.

This did not appear to be a major issue as orthologs (i.e. clades of ingroup tips subtended by

outgroup tips) within homolog trees typically received strong support as monophyletic.

Following homolog tree re-estimation, orthologs were identified as described above with no

minimum taxa requirement. Sequences were aligned with MAFFT and any taxon with >75%

missing data was removed. Filtered alignments were then realigned separately with both MAFFT

and PRANK, and cleaned for 30% column occupancy. This resulted in a dataset with 1899

orthologs. Two subsets of this 1899 ortholog set were generated by requiring a minimum of 30

and 50 taxa, resulting in 661 and 449 orthologs respectively. Gene tree conflict was assessed for

the 449 MAFFT-aligned dataset by rooting all ortholog trees on all taxa in Balsaminaceae and

Marcgraviaceae (i.e. the balsaminoid Ericales) since all previous analyses showed this clade to

be sister to the rest of Ericales; phyparts was then used to map ortholog tree bipartitions to the

ML tree for this dataset and the results were visualized with phypartspiecharts.

Estimating substitutions supporting contentious clades—In order to gain additional insight into

the amount of signal present in ortholog alignments informing various relationships, we

developed a custom script that identifies clades of interest within an ortholog tree and uses the

estimated branch length leading to that clade, multiplied by the length of the corresponding

sequence alignment, to estimate the number of substitutions implied by that branch. Applying

this approach to the trees from the 449 ortholog MAFFT-aligned dataset with appropriate taxon

sampling to allow rooting on a member of the balsaminoid Ericales, we calculated the

approximate number of substitutions that informed the branch leading to the most recent

common ancestor (MRCA) of two or more of the following: Primulaceae,

Polemoniaceae+Fouquieriaceae, Lecythidaceae, Ebenaceae, and Sapotaceae. In addition, we

assessed substitution support for several non-controversial relationships, namely that each of the

following was monophyletic: Polemoniaceae+Fouquieriaceae, Lecythidaceae, and the non-

balsaminoid Ericales. Mean and median values for substitution support were calculated and a

distribution of these values was plotted using custom R scripts.

Synthesis of uncertainty, consensus topology, and genome duplication inference—We took

into consideration all previous results, including those of the expanded datasets, gene tree

conflict, and substitution support, to determine which relationships were generally well-

supported by the results of this study, and which were not. In cases where an interfamilial or

intrafamilial relationship remained irresolvable when considering the preponderance of the

evidence, that relationship was not included in the consensus topology. Gene duplications were

mapped to the consensus topology using the methods of the comparative duplication analysis

described above. Internodes on inferred species trees with notably high numbers of gene

duplications were used as one line of evidence for assessing putative whole genome duplications.

Further investigation into possible genome duplications was conducted by plotting the number of

synonymous substitutions (K_s) between paralogs in K_s plots according to the methods of Yang et

al. (2015) after removing sequences shorter than half the median length of their cluster.

Multispecies K_s values for selected combinations of taxa were generated according to the

methods of Wang et al. (2018). The effect of evolutionary rate-heterogeneity among species was

investigated by conducting a multispecies K_s analysis of each non-balsaminoid ericalean taxon

against *Impatiens balsamifera*, since all evidence suggests each of these pairs have the same

MCRA. Because of this, the ortholog peak resulting from each of these pairs represents the same

speciation event, and therefore differences in the location of this peak in the K_s plots should be

the result of differences in evolutionary rate among species. Plots where the location of the

ortholog peak was ambiguous were not considered for this rate-heterogeneity analysis. All single

and multispecies K_s plots were generated using custom R scripts.

RESULTS

Initial 387 ortholog dataset—The concatenated supermatrix for the 387 ortholog dataset

contained 441,819 aligned sites with 76.1% ortholog occupancy and 57.8% matrix occupancy.

The MLT recovered by RAxML v8.2.4 is shown in Figure 1. A consensus of the rapid

bootstrapping trees unanimously supported a backbone topology (i.e. the RBT) that differed from

that of the MLT by one relationship. The MLT contained the clade

Polemoniaceae+Fouquieriaceae as sister to a clade consisting of Ebenaceae, Sapotaceae, and

what is referred to here as the "Core" Ericales: the clade that includes the Actinidiaceae,

Roridulaceae, Sarraceniaceae, Theaceae, Diapensiaceae, Styracaceae, Symplocaceae,

Pentaphylacaceae, and Ericaceae. The RBT instead placed Lecythidaceae sister to Ebenaceae,

Sapotaceae, and the Core (a hypothetical clade herein referred to as ESC). A gene-wise log-

likelihood analyses comparing these two topologies is shown in Appendix S3. The cumulative

log-likelihood difference between the MLT and RBT was approximately 3.57; there were 27

orthologs that supported the MLT over the RBT by a score larger than this and the exclusion of

any of these from the supermatrix could cause the RBT to become the topology with the best

likelihood. Both of these topologies were considered as candidates in a search for a corroborated

interfamilial backbone. Full bootstrapping resulted in 6.5% support for

Polemoniaceae+Fouquieriaceae sister to ESC unless the ML tree was given as a starting tree,

under which conditions the that topology received 100% bootstrap support. Unguided, full

bootstrapping instead provided strong support for a clade consisting of Lecythidaceae and ESC

(90%, Appendix S4). The ASTRAL topology (AT) for the 387 ortholog set is available in

Appendix S5. Interfamilial relationships in the AT were largely congruent with those of the MLT

except that Primulaceae was recovered as sister to Lecythidaceae+ESC but with only 54% local

posterior probability. A total of seven intrafamilial relationships differed between the AT and

MLT. The AT was considered as a third candidate for an interfamilial backbone.

There was an effect of the ML tree search algorithm that may be important to note for

future phylogenomic studies. Re-conducting an ML tree search for the 387 ortholog supermatrix

with RAxML v8.2.11 using PTHREADS-AVX architecture and -m GTRCAT or with IQ-TREE

and GTR+Γ resulted in a species tree with a different topology than under any other conditions

for the 387 ortholog dataset. The final log-likelihood of the original ML tree was -5948732.774.

Using the PTHREADS-AVX architecture returned a final likelihood of -5948752.263, while the

best log-likelihood reported by IQ-TREE was -5949077.382 (19.489 and 344.607 log-likelihood

points worse than the original results, respectively).

Gene-wise log-likelihood comparative analysis across three candidate topologies—The results

from the gene-wise comparisons of likelihood contributions showed that there were 155

orthologs in the dataset that most strongly supported the MLT, 90 supported the RBT, and 142

supported the AT (Appendix S6). The AT had a cumulative log-likelihood score that was >300

points worse, even though more individual genes support the AT over the RBT (Appendix S6).

Edge-based comparative analyses across three candidate topologies—Of the three candidate

topologies investigated by constraining key edges, Lecythidaceae sister to ESC received the best

score, while Primulaceae sister to ESC received the worst (Appendix S7). Similarly, the clade

Lecythidaceae+Polemoniaceae+Fouquieriaceae+ESC received a better likelihood score than the

clade Lecythidaceae+Primulaceae+ESC. Regarding the placement of Ebenaceae for this dataset,

Ebenaceae+Sapotaceae was supported by the highest number of orthologs whether or not two

log-likelihood support difference was required (Appendix S8). However, a number of orthologs

support each of the investigated placements for Ebenaceae and less than half of genes supported

any placement over the next best alternative by at least two log-likelihood points.

Gene duplication comparative analysis—Gene duplications mapped to each candidate backbone

topology and the five additional hypothetical topologies revealed differing numbers of shared

duplications that can be used as a metric of support among candidate topologies (Fig. 2).

Regarding which clade is better supported as sister to ESC, Lecythidaceae uniquely shares 433

duplications with ESC, more than twice as many as either alternative.

Polemoniaceae+Fouquieriaceae shares 1300 unique duplications with Lecythidaceae+ESC, more

than Primulaceae which shares 626 unique duplications (Fig. 2).

A corroborated topology for the 387 ortholog set—The above comparative analyses supported

one topology among the three candidates identified for the 387 ortholog dataset, namely the

RBT. In this tree, all taxonomic families are recovered as monophyletic. Marcgraviaceae and

Balsaminaceae (i.e. the balsaminoid Ericales) are found to be sister, and form a clade that is

sister to the rest of Ericales (i.e. the non-balsaminoid Ericales). Ebenaceae and Sapotaceae form

a clade that is sister to the Core. The monogeneric Fouquieriaceae is sister to Polemoniaceae. A

clade containing Symplocaceae, Diapensiaceae, and Styracaceae is sister to Theaceae.

Roridulaceae is sister to Actinidiaceae and these together form a clade that is sister to Ericaceae.

A grade containing Primulaceae, Polemoniaceae and Fouquieriaceae, and Lecythidaceae leading

to ESC is supported by the rapid bootstrapping in RAxML, duplication, and constraint-tree

analyses.

Quartet sampling—In our results and discussion we consider a QC score of (≥0.5) to be strong

support as this signifies strong concordance among quartets (Pease et al., 2018). We found

varying levels of support for several key relationships in the RBT (Appendix S9). The

monophyly for all families received strong support (QC ≥ 0.90). There was strong support

(QC=0.54) for the node placing Lecythidaceae sister to ESC, while equivocal support

(QC=0.035) for Polemoniaceae+Fouquieriaceae sister to Lecythidaceae and ESC. Within ESC,

there was moderate support (QC=0.28) for Ebenaceae sister to Sapotaceae, but poor (QC=-0.13)

support for this clade as sister to the Core. There was strong support (QC=0.85) for the clade

including Symplocaceae, Diapensiaceae, and Styracaceae and moderate support (QC=0.26) for

this clade as sister to Theaceae. Roridulaceae was very strongly supported (QC=0.99) as sister to

Actinidiaceae but there was no support for this clade as sister to Ericaceae (QC=-0.23).

However, the monophyly of the clade that includes Roridulaceae, Actinidiaceae, Ericaceae, and

Sarraceniaceae received very strong support (QC=0.90).

The QD scores for several contentious relationships indicate that discordant quartets

tended to be highly skewed towards one conflicting topology as indicated by scores below 0.3

(Pease et al., 2018). However, the QD score for the relationship placing Lecythidaceae sister to

ESC in the RBT was 0.53, indicating relative equality in occurrence frequency of discordant

topologies. Similarly, the relationship placing Polemoniaceae+Fouquieriaceae sister to

Lecythidaceae+ESC received a QD score of 0.83, indicating that among alternative topologies

(e.g. Primulaceae sister to the clade Lecythidaceae+ESC), there was no clear alternative to the

RBT recovered through quartet sampling for this dataset. The QI scores for all nodes defining

interfamilial relationships were above 0.9, indicating that in the vast majority of sampled quartets

there was a tree that was at least two log-likelihoods better than the alternatives. The QF scores

for all but one taxon were above 0.70 and the majority were above 0.85, which shows that rogue

individual taxa were not a major issue (Pease et al., 2018).

Conflict analyses—Assessing ortholog tree concordance and conflict for the 387 ortholog set

mapped to the RBT showed that backbone nodes that differed between candidate topologies

were poorly supported with the majority of orthologs failing to achieve ≥70% bootstrap support

(Appendix S10). Among informative orthologs, the majority of trees conflict with any candidate

topology at these nodes and there is no dominant alternative to the RBT. There were 77 ortholog

trees with appropriate taxon sampling that placed Primulaceae sister to the rest of the non-

balsaminoid Ericales and 37 did so with at least 70% bootstrap support. The clade

Lecythidaceae+Primulaceae+ESC was recovered in 47 ortholog trees, and in 17 with at least

70% bootstrap support. Of the 33 ortholog trees that contained the clade

Primulaceae+Ebenaceae, nine did so with at least 70% bootstrap support.

Expanded ortholog sets—Seven combinations of relationships along the backbone were

recovered in analyses of the expanded ortholog sets (Fig. 3). Full output for each of these

analyses are available on Dryad [TBD]. Among these, the ML tree estimated from the 2045-

ortholog PRANK-aligned, partitioned supermatrix placed Lecythidaceae and Ebenaceae in a

clade sister to Sapotaceae and the Core, with the other interfamilial relationships recapitulating

those of the RBT; the topology recovered by ASTRAL for these orthologs placed

Polemoniaceae+Fouquieriaceae sister to Sapotaceae and the Core, with the clade

Primulaceae+Ebenaceae sister to that and Lecythidaceae sister to a clade including all of those.

In regard to the placement of Ebenaceae for the 2045 PRANK-aligned set, the edge-based

Phyckle analysis showed that 432 of orthologs in this dataset with appropriate taxon sampling

supported Ebenaceae sister to Primulaceae, while 202 did so by at least two log-likelihood over

any alternative (Appendix S11). However, 416 orthologs supported Ebenaceae sister to

Sapotaceae (158 with ≥2LL), and 316 supported Ebenaceae sister to Lecythidaceae (140 with

≥2LL). When the 2045 ortholog set was aligned with MAFFT to produce a supermatrix, the

resulting ML topology was such that the clade Primulaceae+Ebenaceae were sister to

Sapotaceae, with that clade sister to the Core and Polemoniaceae+Fouquieriaceae sister to all of

those. When the same orthologs were aligned with MAFFT, the ASTRAL topology recovered

placed Polemoniaceae+Fouqieriaceae sister to the Core, with the clade Ebenaceae+Primulaceae

sister to that, and Lecythidaceae sister to all of those.

The backbone topology resulting from the 4682 ortholog PRANK-aligned, partitioned

supermatix was the same as that recovered from the 2045 ortholog, partitioned, PRANK-aligned

ML tree. The ASTRAL topology for this dataset placed Lecythidaceae sister to the Core, the

clade Primulaceae+Ebenaceae sister to that, and Polemoniaceae+Fouquieriaceae sister to all of

those. When the 449, 661, or 1899 ortholog set were aligned with MAFFT to produce a

supermatrix, the resulting ML backbone topology was the same as that of the MAFFT-aligned

2045 and 4682 ortholog sets, except when the 449 ortholog supermatrix was used without

partitioning. When ortholog alignments were instead produced with PRANK, the ML backbone

topology recovered from the 449 and ortholog set was the same as that of the 2045 PRANK-

aligned ML tree (i.e Ebenaceae sister to Lecythidaceae and that clade sister to the Core). The

backbone of the ML trees produced from the 1899 and 661 ortholog sets recapitulated that of the

2045 ortholog ASTRAL tree, namely that Polemoniaceae+Fouquieriaceae was sister to

Sapotaceae and the Core, with the clade of Primulaceae+Ebenaceae sister to that and

Lecythidaceae sister to all of those.

Synthesis of uncertainty and determination of an overall consensus—In the species trees

generated with the 387, 449, 661, 1899, 2045, and 4682 ortholog datasets, the placement of

several taxonomic families were resolved in conflict with one another. Many of these

relationships also conflict with the thoroughly investigated RBT, which was shown to be very

well-corroborated by the 387 ortholog set. Nine out of ten ML trees generated with MAFFT-

aligned supermatrices recovered Primulaceae and Ebenaceae sister to one another and forming a

clade with Sapotaceae, however this pattern was not recovered under any other circumstances

(Fig. 3). In all, Primulaceae was recovered as sister to Ebenaceae in 17 of the 33 species trees

generated in this study (51.5%), but these families were sister in only eight of the 24 trees

(33.3%) where they did not form a clade with Sapotaceae. Sapotaceae was recovered as sister to

the Core in 21 of the 33 species trees (63.6%). Thus, there is no majority consensus that

reconciles these conflicting relationships; Primulaceae is only sister to Ebenaceae in a majority

of trees if those that contain the clade Sapotaceae+Ebenaceae+Primulaceae are considered and

that topology is in direct conflict with Sapotaceae+Core, a relationship that is recovered in a

majority of trees. In addition, edge-wise support for the position of Ebenaceae was dataset

dependent and showed equivocal gene-wise support. The positions of Lecythidaceae or the clade

containing Polemoniaceae and Fouquieriaceae that is not agreed by a majority of species trees.

Given this, the relationships among Primulaceae, Polemoniaceae and Fouqieriaceae,

Lecythidaceae, Ebenaceae, and Sapotaceae are not resolved in the consensus topology. All other

interfamilial relationships were unanimously supported by all species trees except for the

relationships between the clade Symplocaceae, Diapensiaceae, and Styracaceae sister to

Theaceae, which was recovered in 29 of the 33 trees (87.9%).

Genome duplication inference—Gene duplications mapped to the consensus topology shows

that the single largest number of duplications occurs on the branch leading the non-Balsaminoid

Ericales (Fig. 4). The other largest numbers of duplications appear along branches leading to

Actinidia (Actinidiaceae), *Camellia* (Theaceae), several members of Primulaceae, *Rhododendron*

(Ericaceae), *Impatiens* (Balsaminaceae), and Polemoniaceae (i.e. *Phlox* and *Saltugilia*). If gene

duplications are instead mapped to the species tree resulting from the 449 ortholog, MAFFT-

aligned supermatrix with partitioning, 3485 duplications occur along the branch leading to the

non-balsaminoid Ericales (Appendix S12). The K_s plots suggest support for several of the recent

lineage-specific duplications, indicated by K_s peaks near zero (Appendix S13). The K_s plots for

most, but not all, taxa also contain a peak that appears between 0.4 and 0.8, which is consistent

with a genome duplication shared by all Ericales. The multispecies K_s plots for some ericalean

taxa paired with a member of Cornales show an ortholog peak with a higher K_s value (i.e. farther

to the right) than the paralog peaks near 1.0, which would be indicative of two separate

duplication events occurring after the divergence of these species. However, other combinations

of ingroup and outgroup taxa result in an ortholog peak to the left of the paralogs peaks, which

would suggest that the implied duplication event occurred in a shared ancestor of the two orders

(Appendix S14). When comparing the locations of unambiguous ortholog peaks of all non-

balsaminoid ericalean taxa to *Impatiens balsamifera*, the location of the peak varied between

values of 1.01 to 1.57 in *Schima superba* and *Primula poissonii* respectively, indicating the

presence of rate heterogeneity in the accumulation of synonymous substitutions in Ericales

(Appendix S15).

Estimating substitutions supporting contentious clades—The estimated number of substitutions

informing clades containing at least two backbone families whose placement is contentious

tended to be much less than for relationships that garnered widespread support. These backbone

clades had a median value of 8.65 estimated substitutions informing them, compared to 30.08,

75.09, and 144.00 informing the monophyly of Polemoniaceae and Fouqieriaceae,

Lecythidaceae, and the non-Balsaminoid Ericales, respectively, in the MAFFT-aligned ortholog

trees (Fig. 4). In trees for the same orthologs with alignments generated instead with PRANK,

the corresponding median values were 8.96, 30.73, 74.16 and 145.98 respectively.

DISCUSSION

Our focal dataset, consisting of 387 orthologs, supports an evolutionary history of

Ericales that is largely consistent with previous work on the clade for many, but not all,

relationships (Appendix S9). The balsaminoid clade, which includes the families Balsaminaceae,

Tetrameristaceae (not sampled in this study), and Marcgraviaceae was confidently recovered as

sister to the rest of the order as has been shown previously (e.g. Geuten et al., 2004; Rose et al.,

2018; Gitzendanner et al., 2018). Similarly, the monogeneric family Fouquieriaceae is sister to

Polemoniaceae, the para-carnivorous Roridulaceae are sister to Actinidiaceae and the

circumscription of Primulaceae sensu lato is monophyletic (Rose et al., 2018). The majority of

analyses for this dataset recovered a sister relationship between Sapotaceae and Ebenaceae, with

that clade sister to the Core Ericales and Lecythidaceae sister to those. Notably, the topology

supported suggests that Primulaceae diverged earlier than has been recovered in most previous

phylogenetic studies and does not form a clade with Sapotaceae and Ebenaceae as has been

suggested by others, including Rose et al. (2018). The topology recovered by Gitzendanner et al.

(2018) using coding sequences from the chloroplast placed Primulaceae sister to Ebenaceae, with

those sister to the rest of the non-basaminoid Ericales, though that study did not include sampling

from Lecythidaceae. In addition to the traditional phylogenetic reconstruction methods, by

applying the available data on gene duplications as a metric of support and leveraging methods

that make use of additional phylogenetic information present in the supermatrix, we were able to

more holistically summarize the evidence present in a 387 ortholog dataset in an effort to resolve

the Ericales phylogeny (Fig. 1; Appendix S3-S10).

It has been shown repeatedly that large phylogenetic datasets have a tendency to resolve

relationships with strong support, even if the inferred topology is incorrect (Seo, 2008).

However, our results suggest extreme sensitivity to tree-building methods. For example, the

initial ML analysis resulted in an ML topology (MLT) with zero rapid bootstrap support for the

placement of Lecythidaceae (Fig. 1). The rapid bootstrap consensus instead unanimously

supported Lecythidaceae sister to a clade including Ebenaceae, Sapotaceae and the Core Ericales

553 (ESC), a hypothesis we refer to as the rapid bootstrap topology (RBT). Gene-wise investigation

554 of likelihood contribution confirmed that these two topologies have very similar likelihoods but

555 did not identify outlier genes that seemed to have an outsized effect on ML calculation

556 (Appendix S3). Instead, the cumulative likelihood influence of the 387 genes in the supermatrix

557 provides nearly equal support for the two topologies, while ASTRAL resulted in a third, and full

558 bootstrapping recovered an even more diverse set of topologies (Appendix S4). These results

559 suggest that there are several topologies with relatively close likelihood scores for this dataset

560 and despite the fact that the additional comparative analyses applied to the 387 ortholog dataset

561 supported a single alternative among those investigated (i.e. the RBT), the recovery of multiple

562 topologies by various tree-building and bootstrapping methods suggested that the criteria used to

563 generate and filter orthologs could have marked potential to influence the outcome of our efforts

564 to resolve relationships amongst the families of Ericales, possibly due to the evolutionary history

565 of the group and the amount of data available to inform these relationships.

566   In addition to sensitivities associated with tree building methods, variation in dataset

567 assembly parameters and methods can result in different topologies. We investigated additional

568 datasets constructed with a variety of methods and filtering parameters to shed further light on

569 the nature of the problem of resolving the ericalean phylogeny. While it is clear from

570 investigation of gene tree topologies in the 387 ortholog dataset that phylogenetic conflict is the

571 rule rather than the exception for those data, the expanded datasets show that this is true for a

572 variety of approaches to dataset construction and not simply an artifact of one approach. While

573 the monophyly of most major clades and the relationships discussed above were recovered

574 across these datasets, we also demonstrate that many combinations of contentious relationships

can be recovered depending on the methods used in dataset construction, alignment, and analysis

(Fig 2).

An unresolved consensus topology—Based on the data available, we suggest that while the

relationships recovered in the Core Ericales and within most families are robust to

methodological alternatives, there is insufficient evidence to resolve several early-diverging

relationships along the ericalean backbone. We, therefore, suggest that the appropriate

representation is as a polytomy (Fig. 4). Whether this is biological or the result of data

limitations remains to be determined. A biological polytomy (i.e. hard polytomy) can be the

result of three or more populations diverging rapidly without significant time for the

accumulation of substitutions to distinguish lineage divergence. The major clades of Ericales

may or may not have diverged simultaneously; however, if divergence occurred rapidly enough

as to preclude the evolution of genomic synapomorphies, then a polytomy may be a reasonable

representation of such historical events rather than signifying a failure of methods or lack of data.

Most of the inferred orthologs contain little information useful for inferring relationships along

the backbone of the phylogeny; we investigated this explicitly by estimating the number of

nucleotide substitutions that inform these backbone relationships and find that branches that

would resolve the polytomy were based on a fraction of the number of substitutions that

informed better-supported clades (Fig. 5). The information that is present results in extensive

gene tree conflict, albeit mostly with low support (Appendix S11). Due to the timing of these

divergence events (80–100 million years ago; Magallón et al., 2015; Rose et al., 2018), coding

DNA sequences may be particularly well suited for inference of these deep relationships due to

their slow rate of evolution relative to noncoding regions, while allowing the inclusion of

potentially informative silent substitutions invisible in amino acid alignments. Our use of a tree-

based orthology inference method provides the best protection available against misidentified

orthology that would introduce systematic error into the analysis. Additionally, the stringent

filtering criteria applied during the initial 387 ortholog supermatix construction, requirement of

an outgroup taxon, and repeated trimming of long branches meant that any sequences that were

included in this dataset were required to be relatively slowly evolving sequences that were

robustly inferred to be orthologous (Yang et al., 2014).

The high levels of gene tree conflict and lack of a clear consensus among datasets for a

resolved topology is likely to have multiple causes. Among these is the fact that this series of

divergence events seems to have happened relatively rapidly over the course of around 10

million years (Rose et al., 2018). We also find evidence that a whole genome duplication is

likely to have occurred before or during this ancient radiation. If this is the case, differential gene

loss and retention during the process of diploidization is likely to complicate our ability to

resolve the order of lineage divergences. In addition, we cannot exclude the possibility of

hybridization and introgression between these ancient lineages at some point in the past, as

hybridization has been documented between plants that have been diverged for tens of millions

of years (Arias et al., 2014; Rothfels et al., 2015). It is even possible that some of the lineages

involved in such introgression have gone extinct in the intervening 100 million years.

Hybridization with such now-extinct "ghost lineages" could therefore represent a potentially

insurmountable obstacle to fully understanding the events that lead to the diversity of forms we

now find in Ericales. We chose not to test explicitly for evidence of hybridization here, due to

the seeming equivocal phylogenetic signal present in most gene trees for these contentious

relationships. We feel that to interpret the generally weak signal present in most conflicting gene

trees as anything other than a lack of reliable information, runs a high risk of overinterpreting

these data since network analyses generally treat tree topology as a state known without error.

However, future studies with more complete taxon sampling could find such an approach to be

appropriate for explaining the high levels of conflict among orthologs, but should carefully

consider alternative explanations for discordance.

Genome duplications—Gene duplication analyses showed evidence for several whole genome

duplications in Ericales, including at least two, those in *Camellia* and *Actinidia*, that have been

verified using sequenced genomes (Huang et al., 2013; Xia et al., 2017; Shi et al., 2010).

Incomplete taxon sampling precludes resolving the precise location of these duplication events

within their respective families, but the 379 duplications shared by *Schima* and *Camellia*

suggests the genome duplication within Theaceae occurred before the divergence of these two

genera, though additional taxa from the family should be investigated in the future. Similarly,

our results show that all species of *Actinidia* included in our study share at least one whole

genome duplication. The corroboration of our results with those of previous work provides

support that the transcriptomic methods we employ here provide a viable means of assessing the

location of recent genome duplications in Ericales (Huang et al., 2013; Xia et al., 2017; Shi et al.,

2010).

The pattern of gene duplication observed along the backbone (i.e. hundreds to thousands

of inferred gene duplications at several contentious nodes) is difficult to interpret in any of our

resolved phylogenies for several reasons (Fig. 2; Appendix S12). The observed pattern could be

the result of biological processes, incomplete transcript sampling or assembly, systematic error in

phylogenetic reconstruction, or a combination of these factors (Sankoff and Zheng, 2018). If

these results were strictly due to biological processes, then the data would suggest that there were

repeated, nested, large-scale gene or whole genome duplications between successive divergence

events. It is also possible that there was a single genome duplication event followed by

cladogenesis and subsequent differential gene loss in descendent lineages. This would result in

the appearance of additional nested duplications because of the differential gene loss. It is

important to note that either of these biological processes produces evidence of shared ancestry

that is potentially useful in inferring evolutionary relationships, as we investigate here. In

addition to biological processes, failure to sequence or assemble transcripts in a given paralog, as

well as phylogenetic uncertainty in homolog trees, can result in a failure to correctly identify the

existence and/or location of duplications.

Our results strongly suggest that uncertainty should be considered when inferring

duplications with tree-based methods and/or from K_s plots (Zwaenepoel and Van de Peer, 2019).

In the case of the former, gene duplications are mapped based on a species tree and therefore the

location to which a given duplication appears can depend on which topologies are considered

(Fig. 2 A-H; Fig. 3; Appendix S12), while the latter may be affected by heterogeneity in

evolutionary rate with faster-evolving taxa accumulating synonymous substitutions at faster rates

than more slowly evolving lineages (Appendix S15; Smith and Donoghue, 2008). Many of our

transcriptomes appear to share two peaks, one around 2.0 that is well-known to be the result of a

genome duplication shared by all angiosperms (Jiao et al., 2011) and another that in most taxa

occurs at K_s values between 0.4 and 0.8 (Appendix S13-S14), though the location of the latter

varies quite substantially, possibly due to the rate heterogeneity in K_s accumulation we find

across ericalean taxa (Appendix S15). Several transcriptomes of outgroup taxa in Cornales also

contain a peak near these values, suggesting that this group also experienced a whole genome

duplication around the same time as Ericales.

Recent nuclear analyses (e.g. Zeng et al., 2017) have provided evidence that Ericales is

sister to Cornales, rather than the two as successive sister-groups to the other asterids, as several

earlier studies had suggested (Bremer et al., 2002, Wikström et al., 2015). Our use of Cornales as

outgroups prohibits us from addressing this topological question directly, but if the two clades

are sister, evidence of similar K_s peaks in both clades would be compatible with a shared

duplication event predating their divergence (Jiao et al., 2012; Wang et al., 2016, Landis et al.,

2018). The multispecies K_s plots do not appear to definitively resolve how many duplication

events took place, since some results suggest a whole genome duplication occurring before the

MRCA of Cornales and Ericales, while results from combinations of taxa directly conflict with

such an interpretation (Appendix S14). We show that the location of ortholog peaks can differ

quite dramatically among species pairs that share the same MRCA (Appendix S14-S15), which

has important implications for the interpretation of multispecies K_s plots. Given this and the

topological uncertainty that remains, we interpret our results to mean that at least one genome

duplication occurred around the time (i.e. before, during, or after) the divergence of Ericales

from other Asterids. Our sampling prevents us from explicitly testing the number of shared

duplications between Ericales and Cornales, but the tree-based methods we employ here detect

duplications in an ingroup (i.e. Ericales) relative to an outgroup (i.e. Cornales, *Helianthus*,

Solanum, *Arabadopsis*, and *Beta*). The fact that we observed thousands of gene duplications

along the backbone of Ericales not present in any outgroup taxon suggests that a whole genome

duplication occurred after Ericales diverged from other Asterids, since these duplicated genes are

not present in the seven cornalean or other outgroup transcriptomes based on reference genomes.

Additional data and expanded taxon sampling will likely shed light on whether a whole genome

duplication occurred before or after the MCRA of Ericales, but the evidence presented here

suggests that choice of taxa and methodology could have important effects on the inferred

locations of such events given that there are several lineage-specific whole genome duplications

towards the tips of the ericalean tree (Fig. 4; Appendix S13-S15), and that the backbone of the

phylogeny is highly sensitive to methodology (Fig. 3).

CONCLUSIONS

The first transcriptomic dataset broadly spanning Ericales and constructed from publicly

available data resolves many of the relationships within the clade and supports several

relationships that have been proposed previously. Our results confirm genome duplications in

Actinidiaceae and Theaceae and support at least one whole genome duplication close to the

origin of Ericales. We find evidence to suggest additional duplication events in Balsaminaceae,

Ericaceae, Polemoniaceae, and Primulaceae. As genomic sampling improves, we may discover

additional duplications and refine placement of others. While trees were largely concordant

within taxonomic families, our results concerning the topological resolution of the deep

divergences in Ericales are less decisive. We demonstrate that with the available data, there is

not enough information to strongly support any resolution. While these relationships have been

resolved in previous studies, our analyses strongly suggest that uncertainty should be thoroughly

investigated in phylotranscriptomic datasets, as strong support can be given by different methods

for conflicting topologies. Even in a dataset containing hundreds of genes and hundreds of

thousands of characters, the criteria used in dataset construction altered the inferred topology,

even without the presence of outlier genes. The high prevalence of conflict among datasets and

the lack of clear consensus in regards to the relationships among several major ericalean clades

led us to conclude that a single, fully resolved tree is not supported by these transcriptome data,

though we acknowledge that future improvements in sampling might justify their resolution. Our

results suggest that future work should employ a range of methodologies and support metrics so

that topological uncertainty can be fully explored and reported.

Data Accessibility Statement

Data and custom software used in this study are available at Dryad [TBD].

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Author Contributions

The study was conceived by D.A.L. and S.A.S. Dataset construction was led by D.A.L. with

contributions by O.M.V. and input from J.F.W. Analyses were led by D.A.L. and J.F.W with

contributions and input by S.A.S. D.A.L. wrote the manuscript with contributions and input from

O.M.V, J.F.W., and S.A.S. All authors read and approved the final draft of the manuscript.

LITERATURE CITED

Altschul, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment

search tool. *Journal of molecular biology* 215: 403-410.

Anderberg, A. A., C. Rydin, and M. Källersjö. 2002. Phylogenetic relationships in the order

Ericales sl: analyses of molecular data from five genes from the plastid and mitochondrial

genomes. *American Journal of Botany* 89: 677-687.

Arias, T., M. A. Beilstein, M. Tang, M. R. McKain, and J. C. Pires. 2014. Diversification times

among Brassica (Brassicaceae) crops suggest hybrid formation after 20 million years of

divergence. *American Journal of Botany* 101: 86-91.

Bremer, B., K. Bremer, N. Heidari, P. Erixon, R. G. Olmstead, A. A. Anderberg, M. Källersjö,

and E. Barkhordarian. 2002. Phylogenetics of asterids based on 3 coding and 3 noncoding

chloroplast DNA markers and the utility of noncoding DNA at higher taxonomic levels.

Molecular Phylogenetics and Evolution 24: 274–301.

Brown, J. M., and R. C. Thomson. 2016. Bayes factors unmask highly variable information

content, bias, and extreme influence in phylogenomic analyses. Systematic Biology 66:

517-530.

Brown, J. W., J. F. Walker, and S. A. Smith. 2017. Phyx: phylogenetic tools for unix.

Bioinformatics 33: 1886-1888.

Brown, J. W., N. Wang, and S. A. Smith. 2017. The development of scientific consensus:

Analyzing conflict and concordance among avian phylogenies. Molecular Phylogenetics

and Evolution 116: 69-77.

Chase, M. W., M. Christenhusz, M. Fay, J. Byng, W. S. Judd, D. Soltis, D. Mabberley, et al.

2016. An update of the Angiosperm Phylogeny Group classification for the orders and

families of flowering plants: APG IV. *Botanical Journal of the Linnean Society* 181: 1-

20.

Chase, M. W., D. E. Soltis, R. G. Olmstead, D. Morgan, D. H. Les, B. D. Mishler, M. R. Duvall,

et al. 1993. Phylogenetics of seed plants: an analysis of nucleotide sequences from the

plastid gene rbcL. *Annals of the Missouri Botanical Garden*: 528-580.

Eisen, J. A. 1998. Phylogenomics: improving functional predictions for uncharacterized genes by

evolutionary analysis. *Genome research* 8: 163-167.

Fu, L., B. Niu, Z. Zhu, S. Wu, and W. Li. 2012. CD-HIT: accelerated for clustering the next

generation sequencing data. *Bioinformatics* 28: 3150–3152.

Gabaldón, T. 2008. Large-scale assignment of orthology: back to phylogenetics? *Genome*

biology 9: 235.

Geuten, K., E. Smets, P. Schols, Y.-M. Yuan, S. Janssens, P. Küpfer, and N. Pyck. 2004.

Conflicting phylogenies of balsaminoid families and the polytomy in Ericales: combining

data in a Bayesian framework. *Molecular Phylogenetics and Evolution* 31: 711-729.

Gitzendanner, M. A., P. S. Soltis, G. K. S. Wong, B. R. Ruhfel, and D. E. Soltis. 2018. Plastid

phylogenomic analysis of green plants: a billion years of evolutionary history. *American*

Journal of Botany 105: 291-301.

Grabherr, M. G., B. J. Haas, M. Yassour, J. Z. Levin, D. A. Thompson, I. Amit, X. Adiconis, et

al. 2011. Full-length transcriptome assembly from RNA-Seq data without a reference

genome. *Nature biotechnology* 29: 644.

Haas, B. J., A. Papanicolaou, M. Yassour, M. Grabherr, P. D. Blood, J. Bowden, M. B. Couger,

et al. 2013. De novo transcript sequence reconstruction from RNA-seq using the Trinity

platform for reference generation and analysis. *Nature Protocols* 8: 1494–1512.

He, Y., C. Kueffer, P. Shi, X. Zhang, M. Du, W. Yan, and W. Sun. (2014) Variation of biomass

and morphology of the cushion plant Androsace tapete along an elevational gradient in

the Tibetan Plateau. *Plant species biology* 29: e64-e71.

Hedwall, P. O., J. Brunet, A. Nordin, and J. Bergh. 2013. Changes in the abundance of keystone

forest floor species in response to changes of forest structure. *Journal of vegetation*

science 24: 296-306.

Jiao, Y., J. Leebens-Mack, S. Ayyampalayam, J. E. Bowers, M. R. McKain, J. McNeal, M. Rolf,

et al. 2012. A genome triplication associated with early diversification of the core

eudicots. *Genome biology* 13: R3.

Jiao, Y., N. J. Wickett, S. Ayyampalayam, A. S. Chanderbali, L. Landherr, P. E. Ralph, L. P.

Tomsho, et al. 2011. Ancestral polyploidy in seed plants and angiosperms. *Nature* 473:

97.

Katoh, K., K. Misawa, K. i. Kuma, and T. Miyata. 2002. MAFFT: a novel method for rapid

multiple sequence alignment based on fast Fourier transform. *Nucleic acids research* 30:

3059-3066.

Katoh, K., and D. M. Standley. 2013. MAFFT multiple sequence alignment software version 7:

improvements in performance and usability. *Molecular biology and evolution* 30: 772-

780.

Kozlov, A., D. Darriba, T. Flouri, B. Morel, and A. Stamatakis. 2019. RAxML-NG: A fast,

scalable, and user-friendly tool for maximum likelihood phylogenetic inference. *bioRxiv*:

447110.

Landis, J. B., D. E. Soltis, Z. Li, H. E. Marx, M. S. Barker, D. C. Tank, and P. S. Soltis. 2018.

Impact of whole-genome duplication events on diversification rates in angiosperms.

American Journal of Botany 105: 348-363.

Löytynoja, A. 2014. Phylogeny-aware alignment with PRANK, Multiple sequence alignment

methods, 155-170. Springer.

Magallón, S., S. Gómez-Acevedo, L. L. Sánchez-Reyes, and T. Hernández-Hernández. 2015. A

metacalibrated time-tree documents the early rise of flowering plant phylogenetic

diversity. *New Phytologist* 207: 437-453.

Matasci, N., L.-H. Hung, Z. Yan, E. J. Carpenter, N. J. Wickett, S. Mirarab, N. Nguyen, et al.

2014. Data access for the 1,000 Plants (1KP) project. *GigaScience* 3: 17.

Memiaghe, H. R., J. A. Lutz, L. Korte, A. Alonso, and D. Kenfack (2016). Ecological

importance of small-diameter trees to the structure, diversity, and biomass of a tropical

evergreen forest at Rabi, Gabon. *PLoS One* 11: e0154988.

Kodama, Y., M. Shumway, and R. Leinonen. 2011. The sequence read archive. *Nucleic Acids*

Research 39: D19–D21.

Moquet, L., M. Vanderplanck, R. Moerman, M. Quinet, N. Roger, D. Michez, and A. L.

Jacquemart. 2017. Bumblebees depend on ericaceous species to survive in temperate

heathlands. *Insect Conservation and Diversity* 10: 78-93.

Pease, J. B., J. W. Brown, J. F. Walker, C. E. Hinchliff, and S. A. Smith. 2018. Quartet Sampling

distinguishes lack of support from conflicting support in the green plant tree of life.

American Journal of Botany 105: 385-403.

Price, M. N., P. S. Dehal, and A. P. Arkin. 2010. FastTree 2 -Approximately maximum-

likelihood trees for large alignments. *PLoS One* 5: e9490.

Reddy, S., R. T. Kimball, A. Pandey, P. A. Hosner, M. J. Braun, S. J. Hackett, K.-L. Han, et al.

2017. Why do phylogenomic data sets yield conflicting trees? Data type influences the

avian tree of life more than taxon sampling. *Systematic Biology* 66: 857-879.

Rose, J. P., T. J. Kleist, S. D. Löfstrand, B. T. Drew, J. Schönenberger, and K. J. Sytsma. 2018.

Phylogeny, historical biogeography, and diversification of angiosperm order Ericales

suggest ancient Neotropical and East Asian connections. *Molecular Phylogenetics and*

Evolution 122: 59-79.

Rothfels, C. J., A. K. Johnson, P. H. Hovenkamp, D. L. Swofford, H. C. Roskam, C. R. Fraser-

Jenkins, M. D. Windham, and K. M. Pryer. 2015. Natural hybridization between genera

that diverged from each other approximately 60 million years ago. *The American*

Naturalist 185: 433-442.

Sankoff, D., and C. Zheng. 2018. Whole Genome Duplication in Plants: Implications for

Evolutionary Analysis, Comparative Genomics, 291-315. Springer.

Shen, X.-X., C. T. Hittinger, and A. Rokas. 2017. Contentious relationships in phylogenomic

studies can be driven by a handful of genes. *Nature ecology & evolution* 1: 0126.

Schönenberger, J., A. A. Anderberg, and K. J. Sytsma. 2005. Molecular phylogenetics and

patterns of floral evolution in the Ericales. *International Journal of Plant Sciences* 166.2:

265-288.

Shi, T., H. Huang, and M. S. Barker. 2010. Ancient genome duplications during the evolution of

kiwifruit (Actinidia) and related Ericales. *Annals of Botany* 106: 497-504.

Simon, S., A. Narechania, R. DeSalle, and H. Hadrys. 2012. Insect phylogenomics: exploring the

source of incongruence using new transcriptomic data. *Genome biology and evolution* 4:

1295-1309.

Smith, S.A., J. F. Walker, J. W. Brown, and N. Walker-Hale, 2018. Nested phylogenetic

conflicts, combinability, and deep phylogenomics in plants. *bioRxiv*, 371930.

Smith, S. A., and M. J. Donoghue. 2008. Rates of molecular evolution are linked to life history

in flowering plants. *Science* 322: 86-89.

Smith, S. A., M. J. Moore, J. W. Brown, and Y. Yang. 2015. Analysis of phylogenomic datasets

reveals conflict, concordance, and gene duplications with examples from animals and

plants. *BMC Evolutionary Biology* 15: 150.

Stamatakis, A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of

large phylogenies. Bioinformatics 30: 1312-1313.

Stevens, P. F. 2001 onward. Angiosperm phylogeny website, version 14, July 2017 [more or less

continuously updated]. Website http://www.mobot.org/MOBOT/research/APweb/

[accessed 11 July 2019]

Ter Steege, H., N. C. Pitman, O. L. Phillips, J. Chave, D. Sabatier, A. Duque, J.-F. Molino, et al.

2006. Continental-scale patterns of canopy tree composition and function across

Amazonia. *Nature* 443: 444.

Vargas, O. M., E. M. Ortiz, and B. B. Simpson. 2017. Conflicting phylogenomic signals reveal a

pattern of reticulate evolution in a recent high-Andean diversification (Asteraceae:

Astereae: Diplostephium). *New Phytologist* 214: 1736-1750.

Walker, J. F., J. W. Brown, and S. A. Smith. 2018. Analyzing contentious relationships and

outlier genes in phylogenomics. *Systematic Biology* 67: 916-924.

Walker, J. F., N. Walker-Hale, O. M. Vargas, and D. A. Larson, G. W. Stull. 2019.

Characterizing gene tree conflict in plastome-inferred phylogenies. *bioRxiv*: 512079.

Walker, J. F., Y. Yang, T. Feng, A. Timoneda, J. Mikenas, V. Hutchison, C. Edwards, et al.

2018. From cacti to carnivores: Improved phylotranscriptomic sampling and hierarchical

homology inference provide further insight into the evolution of Caryophyllales.

American Journal of Botany 105: 446-462.

Walker, J. F., Y. Yang, M. J. Moore, J. Mikenas, A. Timoneda, S. F. Brockington, and S. A.

Smith. 2017. Widespread paleopolyploidy, gene tree conflict, and recalcitrant

relationships among the carnivorous Caryophyllales. *American Journal of Botany* 104:

858-867.

Wang, N., Y. Yang, M. J. Moore, S. F. Brockington, J. F. Walker, J. W. Brown, B. Liang, et al.

2018. Evolution of Portulacineae marked by gene tree conflict and gene family expansion

associated with adaptation to harsh environments. *Molecular biology and evolution* 36:

112-126.

Wang, Y., S. P. Ficklin, X. Wang, F. A. Feltus, and A. H. Paterson. 2016. Large-scale gene

relocations following an ancient genome triplication associated with the diversification of

core eudicots. *PloS one* 11: e0155637.

Wickett, N. J., S. Mirarab, N. Nguyen, T. Warnow, E. Carpenter, N. Matasci, S. Ayyampalayam,

et al. 2014. Phylotranscriptomic analysis of the origin and early diversification of land

plants. *Proceedings of the National Academy of Sciences* 111: E4859-E4868.

Wikström, N., K. Kainulainen, S. G. Razafimandimbison, J. E. E. Smedmark, and B. Bremer.

2015. A revised time tree of the asterids: Establishing a temporal framework for

evolutionary studies of the coffee family (Rubiaceae). *PLoS One* 10: e0126690.

Xia, E.H., H.B. Zhang, J. Sheng, K. Li, Q.J. Zhang, C. Kim, Y. Zhang, et al. 2017. The tea tree

genome provides insights into tea flavor and independent evolution of caffeine

biosynthesis. *Molecular plant*, *10*: 866-877.

Yang, Y., M. J. Moore, S. F. Brockington, J. Mikenas, J. Olivieri, J. F. Walker, and S. A. Smith.

2018. Improved transcriptome sampling pinpoints 26 ancient and more recent polyploidy

events in Caryophyllales, including two allopolyploidy events. New Phytologist 217:

855-870.

Yang, Y., M. J. Moore, S. F. Brockington, D. E. Soltis, G. K.-S. Wong, E. J. Carpenter, Y.

Zhang, et al. 2015. Dissecting molecular evolution in the highly diverse plant clade

Caryophyllales using transcriptome sequencing. Molecular biology and evolution 32:

2001-2014.

Yang, Y., and S. A. Smith. 2014. Orthology inference in nonmodel organisms using

transcriptomes and low-coverage genomes: improving accuracy and matrix occupancy

for phylogenomics. *Molecular biology and evolution* 31: 3081-3092.

Zeng, L., N. Zhang, Q. Zhang, P. K. Endress, J. Huang, and H. Ma (2017). Resolution of deep

eudicot phylogeny and their temporal diversification using nuclear genes from

transcriptomic and genomic datasets. *New Phytol.* 214, 1338–1354.

Zhang, C., M. Rabiee, E. Sayyari, and S. Mirarab. 2018. ASTRAL-III: polynomial time species

tree reconstruction from partially resolved gene trees. *BMC bioinformatics* 19: 153.

Zwaenepoel A., Y. Van de Peer. 2019. Inference of Ancient Whole-Genome Duplications and

the Evolution of Gene Duplication and Loss Rates. *Mol Biol Evol*. 36: 1384–1404.

FIGURES

Figure 1. Maximum likelihood topology recovered for a 387 ortholog dataset using RAxML.

Nodes receiving less than 100% rapid BS support are labeled. Branch lengths are substitutions

per base pair. The node that determined the placement of Lecythidaceae received zero support

(i.e. the ML topology was never recovered by a rapid bootstrap replicate).

Figure 2. (A-H) Gene duplications obtained by mapping duplicated clades with at least 50% SH-aRLT support in homolog trees to several topologies recovered with a 387 ortholog dataset including the maximum likelihood topology (A) the rapid bootstrap topology (B) the ASTRAL topology (C) and several hypothetical topologies that demonstrate evidence for shared duplications in clades not recovered with species tree methods (D-H). Names of clades are abbreviated to four letters, ESC represents the clade Ebenaceae+Sapotaceae+Core Ericales, and "Pole" represents the clade Polemoniaceae+Fouquieriaceae in all cases. I) Bar chart showing the number of uniquely shared gene duplications between clades that can be considered a metric of support for distinguishing among conflicting topological relationships.

Figure 3. Topologies recovered from several combinations of ortholog datasets and species tree methods representing alternative approaches to those used for a 387 ortholog dataset. Each color corresponds to a unique backbone topology recovered in these analyses. Names of clades are abbreviated to four letters and "Pole" represents the clade Polemoniaceae+Fouquieriaceae in all cases.

Figure 4. Gene duplications mapped to a cladogram of the consensus topology. The diameter of circles corresponds to the number of inferred duplications, and cases with at least 250 are labelled along branches. The single largest number of duplications occurs on the branch leading the non-Balsaminoid Ericales. Verified genome duplications in the Theaceae and the Actinidiaceae appear as the second and third largest numbers of gene duplications respectively.

Figure 5. Estimated number of substitutions supporting various clades using rootable ortholog

trees from the 449 ortholog MAFFT-aligned dataset. The number of substitutions supporting a

clade was estimated by multiplying the relevant branch length by the length of the corresponding

alignment for each ortholog tree in which that clade is present. Substitution support for clades

that would resolve the polytomy in the consensus topology had a median value of 8.65 estimated

substitutions informing them, compared to 30.08, 75.09, and 144.00 informing the monophyly of

Polemoniaceae and Fouqieriaceae, Lecythidaceae, and the non-Balsaminoid Ericales

respectively. In trees for the same orthologs aligned with PRANK, the corresponding values

were 8.96, 30.73, 74.16 and 145.98.

SUPPLEMENTAL MATERIALS

Supplemental Appendix 1. The origins of transcriptome assemblies, reference genomes, and

raw reads used in homolog clustering. Citations associated with raw reads available on the

National Center for Biotechnology Information Sequence Read Archive are included where such

information was provided in the accession record or could be confidently identified through a

Google Scholar query of the relevant accession information.

Supplemental Appendix 2. Phylogram inferred using maximum likelihood on a supermatrix

consisting of the genes *rpoC2*, *rbcL*, *nhdF*, and *matK*. The topology of the tree was used to

assign taxa into eight groups for hierarchical homolog clustering, such that each clustering group

was monophyletic and not prohibitively large.

Supplemental Appendix 3. Results of a two topology test comparing gene-wise support in the

387 ortholog dataset for two alternative placements of Lecythidaceae recovered by the maximum

likelihood (ML) search. Delta gene-wise log-likelihood represents the extent to which an

ortholog supports the ML topology over that recovered unanimously in rapid bootstraps. The

dashed line represents the cumulative difference in log-likelihood between the two competing

topologies, such that removing any of the 27 orthologs with a score more positive than that

would cause the ML topology to change, even in the absence of obvious outlier genes.

Supplemental Appendix 4. Results of unguided, full bootstrapping with the 387 ortholog

supermatrix in RAxML. The number of bootstrap replicates in which various contentious

backbone relationships were recovered are reported.

Supplemental Appendix 5. The maximum quartet support species tree topology for the 387

ortholog set generated with ASTRAL. Node support values are ASTRAL local posterior

probabilities and nodes receiving support less than 1.0 are labeled. Branch lengths are in

coalescent units.

Supplemental Appendix 6. Results from the gene-wise comparisons of likelihood contributions

for the three candidate topologies for the 387 ortholog dataset. A) The absolute log-likelihood for

each ortholog, sorted by which topology they best support and organized in descending order of

likelihood. B) The cumulative difference in log-likelihood relative to the ML topology as

orthologs are added across the supermatrix. Delta gene-wise log-likelihood (ΔGWLL) represents

the extent to which the topology has a worse score than the ML topology; values below zero

indicate that a topology has a better likelihood than the ML topology. C) Schematic of the three

candidate topologies in question.

Supplemental Appendix 7. Log-likelihood penalties incurred by constraining key edges

consistent with each of the three candidate topologies for a 387 ortholog dataset using EdgeTest.

Direct comparisons of scores between conflicting clades can be used as a metric of support.

Supplemental Appendix 8. The Phyckle results regarding the placement of Ebenaceae for the

387 ortholog dataset. Of the relationships examined, Ebenaceae+Sapotaceae was supported by

the highest number of orthologs whether or not two log-likelihood support difference was

required.

Supplemental Appendix 9. Cladogram showing results from quartet sampling on the rapid

bootstrap topology (RBT) from a 387 ortholog dataset. Branch labels show quartet concordance

(QC), quartet differential (QD), and quartet informativeness (QI) respectively for each

relationship. Quartet fidelity (QF) for each taxon is shown in parentheses after the relevant taxon

label.

Supplemental Appendix 10. A) Ortholog tree concordance and conflict for the 387 ortholog set

mapped to the rapid bootstrap topology. B) The same analysis except requiring 70% ortholog

tree bootstrap support for an ortholog to be considered informative. Blue indicates the proportion

of informative orthologs that are concordant with the topology, green indicates the proportion of

informative orthologs that support the single most common conflicting topology, red indicates all

other informative ortholog conflict and grey indicates orthologs that are uninformative, either

due to support requirements or lack of appropriate taxon sampling.

1008

Supplemental Appendix 11. The Phyckle results regarding the placement of Ebenaceae for the

2045 ortholog dataset. Ebenaceae+Primulaceae was supported by the highest number of

orthologs whether or not two log-likelihood support difference was required.

1012

Supplemental Appendix 12. Gene duplications mapped to a cladogram of the 449 ortholog

MAFFT-aligned, partitioned, supermatrix ML tree. Number of inferred gene duplications in

shown along branches. The diameter of circles at nodes are proportional to the number of

duplications.

1017

Supplemental Appendix 13. Single species K_s plots for pairs of paralogs within each taxa as a

density plot. Density peaks indicate evidence for a large proportion of genes having been

duplicated at approximately the same time, as would occur during a whole genome duplication.

For each taxa, the x-axis ranges from 0.0 to 3.0 with each tick representing 0.5 synonymous

substitutions between paralogs. The y-axis is scaled to the maximum density value for each

transcriptome and ticks correspond to 0.25, 0.5, 0.75, and 1.0 of the maximum density

respectively. Input transcriptomes have been filtered to remove short sequences and sample-

specific duplications since these can represent transcript splice-site variants or errors in

assembly.

1027

Supplemental Appendix 14. Multispecies K_s plots representing a broad range of taxonomic

pairings. Single species K_s density plots (i.e. pairs of paralogs) within each taxa are plotted on the

same axis as a K_s density plot representing pairs of orthologs between the two taxa. The density

peak for the orthologs corresponds to the time of divergence between the two taxa, since

orthologs in both taxa begin accumulating synonymous substitutions after speciation. If

evolutionary rate in the two taxa are equal and the accumulation of synonymous substitutions is

clock-like, then the timing the duplication events relative to divergence of the two taxa can be

compared, with older events occurring farther to the right. The x-axis ranges from 0.0 to 3.0 with

each tick representing 0.5 synonymous substitutions. The y-axis is scaled to the maximum

density value of any of the three density distributions (therefore the magnitudes of all peaks are

relative) and ticks correspond to 0.25, 0.5, 0.75, and 1.0 of the maximum density respectively.

Input transcriptomes have been filtered to remove short sequences and sample-specific

duplications since these can represent transcript splice-site variants or errors in assembly.

1041

Supplemental Appendix 15. Multispecies K_s plots representing each non-balsaminoid ericalean

taxa paired with *Impatiens balsamifera* for which there is an unambiguous ortholog peak. Single

species K_s density plots (i.e. pairs of paralogs) within each taxa are plotted on the same axis as a

K_s density plot representing pairs of orthologs between the two taxa. The density peak for the

orthologs corresponds to the time of divergence between the two taxa, since orthologs in both

taxa begin accumulating synonymous substitutions after speciation. The dashed line represents

the point along the x-axis where the ortholog peak achieves its maximum value and would be

expected to occur at approximately the same point in all pairings under clock-like accumulation

of synonymous substitutions since all pairs share the same MRCA. The x-axis ranges from 0.0 to

3.0 with each tick representing 0.5 synonymous substitutions. The y-axis is scaled to the

maximum density value of any of the three density distributions (therefore the magnitudes of all

peaks are relative) and ticks correspond to 0.25, 0.5, 0.75, and 1.0 of the maximum density

respectively. Input transcriptomes have been filtered to remove short sequences and sample-

specific duplications since these can represent transcript splice-site variants or errors in

assembly.