Neural representation of bat predation risk and evasive flight in moths: a modelling

approach

Holger R. Goerlitz[a,c,d,*], Hannah M. ter Hofstede[b,c,d], Marc W. Holderied[c]

a. Max Planck Institute for Ornithology, Acoustic and Functional Ecology Group, 82319

Seewiesen, Germany

b. Dartmouth College, Department of Biological Sciences, Hanover, NH, 03755, USA

c. University of Bristol, School of Biological Sciences, Bristol, BS8 1UG, UK

d. Contributed equally

* corresponding author: hgoerlitz@orn.mpg.de

Short title: Neural representation of predator threat

Abstract

Most animals are at risk from multiple predators and can vary anti-predator behaviour based

on the level of threat posed by each predator. Animals use sensory systems to detect predator

cues, but the relationship between the tuning of sensory systems and the sensory cues related

to predator threat are not well-studied at the community level. Noctuid moths have

ultrasound-sensitive ears to detect the echolocation calls of predatory bats. Here, combining

empirical data and mathematical modelling, we show that moth hearing is adapted to provide

information about the threat posed by different sympatric bat species. First, we found that

multiple characteristics related to the threat posed by bats to moths correlate with bat

echolocation call frequency. Second, the frequency tuning of the most sensitive auditory

receptor in noctuid moth ears provides information allowing moths to escape detection by all

sympatric bats with similar safety margin distances. Third, the least sensitive auditory

receptor usually responds to bat echolocation calls at a similar distance across all moth

species for a given bat species. If this neuron triggers last-ditch evasive flight, it suggests that

there is an ideal reaction distance for each bat species, regardless of moth size. This study

shows that even a very simple sensory system can adapt to deliver information suitable for

triggering appropriate defensive reactions to each predator in a multiple predator community.

Keywords: A1, A2, anti-predator behaviour, Chiroptera, Lepidoptera, predator-prey-

interaction

1. Introduction

Sensory system adaptations for detecting and responding to predator cues are well-known in animals, and the specific neural activity required to generate anti-predator behaviour has been documented for many species (e.g. arctiid moths: Ratcliffe et al. 2009; crickets: Nolen & Hoy 1984; locusts: Fotowat & Gabbiani 2007; cockroaches: Ritzmann 1980; crayfish: Edwards et al. 1999; fish: Eaton et al. 2001). Most animals co-occur with multiple predator species, each of which provides different cues and poses a different level of threat (Smolka et al. 2001; Falk et al. 2015). Animals often demonstrate anti-predator behaviours that are proportional to the risk posed by the detected predator (e.g. moths: Roeder 1974, Ratcliffe et al. 2011; crabs: Smolka et al. 2001; frogs: Fraker 2008; fish: Helfman 1989; birds: Templeton et al. 2005), but many aspects of predator threat cannot be directly encoded by sensory systems, such as the speed and manoeuvrability of a predator that is stationary at the time it is detected. Behavioural studies suggest that some animals might overcome this limitation by assessing a single detectable trait that is comparable across predators, such as size, to estimate predator threat (Templeton et al. 2005). Few studies, however, have assessed whether sensory systems are specifically tuned to cues that correlate with the predator threat posed by different predators in the community.

Eared moths and echolocating bats are ideal study animals for addressing this question. Echolocating bats are significant nocturnal predators of moths (reviewed in Fullard 1998). Bats produce ultrasonic echolocation calls for orientation and prey detection, and ultrasound-sensitive ears evolved in many moth families with the primary function of detecting bat echolocation calls (Fullard 1998; ter Hofstede & Ratcliffe 2016). With only 1-4 auditory receptor neurons depending on the moth family, these ears are the simplest ears in nature (Yack 2004). Moths in the family Noctuidae have two auditory receptor cells, called A1 and A2. The A2 cell is approximately 20 dB less sensitive than the A1 cell (**Fig. S1**).

Noctuid moths also have a two-staged anti-bat response; they show directional flight away

from quiet ultrasonic pulses, typical of a distant bat, and last-ditch flight manoeuvres in

response to loud ultrasonic pulses, typical of a close bat (Roeder 1962, 1964; Agee 1969).

Directional flight is likely triggered at intensities just above A1 threshold (Roeder 1964,

1967) and at much greater distances than those at which the bat can detect the moth's echo

(Roeder 1998; Surlykke et al. 1999; Goerlitz et al. 2010), meaning that moths can initiate

directional flight to avoid being detected by the bat. Directional flight, however, should no

longer be effective once the bat has detected the moth (Corcoran & Conner 2016) due to

differences in flight speeds between bats (4-9 m/s: **Table S1**) and moths (0.5-6 m/s; Riley et

al. 1992; Vickers & Baker 1997; Luo et al. 2002; Chapman et al. 2008; Corcoran and Conner

2016). Instead, moths require more drastic erratic flight or other secondary defence strategies

(e.g., Ratcliffe & Fullard 2005, Corcoran & Conner 2012) to avoid being captured by the

attacking bat. Due to the A2 cell's higher thresholds, meaning it is only activated by high

amplitude sounds, Roeder (1974) hypothesized that A2 cell activity might trigger these last-

ditch flight maneuvers. This hypothesis, however, has never been empirically tested in

noctuid moths.

The frequency tuning curves of moth A-cells vary depending on the sympatric bat

community (reviewed in Fullard 1998; ter Hofstede et al. 2013), suggesting a strong

functional link between frequency tuning of moth ears and the echolocation call frequency of

sympatric bats. Although the A1 cell is more sensitive than the A2 cell, both cells have the

same shaped tuning curve (**Fig. S1**), meaning that moths cannot discriminate between

different bat species based on frequency (ter Hofstede et al. 2013). Larger moths have lower

A1 cell thresholds (i.e. are more sensitive) than smaller moths, presumably to compensate for

the greater distances at which bats can detect the louder echo reflected from their larger

surface area (Surlykke et al. 1999; ter Hofstede et al. 2013). Despite differences in tuning and

sensitivity between moth species, the general shape of the auditory tuning curve is similar

across most moth species, with greatest sensitivity to lower ultrasonic frequencies and

decreasing sensitivity at higher ultrasonic frequencies (**Fig. S1**; Fullard 1998; ter Hofstede et

al. 2013; ter Hofstede & Ratcliffe 2016). The relatively consistent shape of the moth auditory

tuning curve across species suggests that it is either constrained by morphological or

physiological factors or that it is a functional adaptation, i.e. a matched filter (Wehner 1987;

Römer 2016; von der Emde & Warrant 2016).

Here we use both empirical data and mathematical modelling to assess whether the

shape of the moth auditory tuning curve is adapted such that moths detect different bat

species at times when they pose a similar level of threat. First, we tested for relationships

between bat call frequency, which varies across bat species in a community, and four

variables that influence the detectability of calls by the moth and echoes by the bat (call

intensity, duration and repetition rate) or the time required for the bat to intercept the moth

(bat flight speed). Strong relationships between these variables would allow for the evolution

of moth ear frequency tuning that reflects the threat posed by different bat species. Second,

we incorporate data on bat echolocation call parameters, bat and moth hearing thresholds and

estimated detection distances between bats and moths into multiple models of moth escape

behaviour to develop testable hypotheses about the adaptive value of moth ear tuning for

evading bat predators at a distance or at close range.

2. Relationships between bat call frequency and bat characteristics relevant to predator

threat

We compiled data from the literature and our own measurements on echolocation call peak

frequency, call duration, apparent call source level, call interval and flight speed for 14

European bat species (**Table S1**) that hunt flying moths (Barlow 1997; Vaughan 1997;

Andersson et al. 1998; Bogdanowicz et al. 1999; Dietz et al. 2009). For our own

measurements, we used acoustic flight path tracking to obtain these data for five European

bat species (**Table S1**, for methods see Goerlitz et al. 2010). Thirteen of the bat species are in

the family Vespertilionidae and produce relatively short duration calls at long intervals to

separate call and echo in time, sometimes referred to as low duty-cycle echolocation (Jones &

Teeling 2006; Fenton et al. 2012). One of the bat species (*Rhinolophus ferrumequinum*) is in

the family Rhinolophidae, which uses a specialized form of echolocation, sometimes referred

to as high duty cycle echolocation, in which they produce long-duration calls and separate

call and echo in frequency, allowing them to detect glints in the echoes caused by insects

wing movements (Jones & Teeling 2006; Fenton et al. 2012). Species in this family,

including *R. ferrumequinum*, often hunt insects close to vegetation (Jones & Rayner 1989;

Dietz et al. 2009; Lazure & Fenton 2011).

For all bat species, we compiled data on call peak frequency (frequency with

maximum energy), call duration (time from the start to the end of the call), apparent call

source level (apparent emitted sound level at 10 cm from the bat's mouth; i.e., the lower

boundary of the source level because measurements also include off-axis calls), call interval

(time from start of one call to start of the next call), and flight speed (the mean speed of

multiple flight paths). We included these variables in our analyses due to their relevance to

predator threat. Bats calling at lower frequency can detect prey, and can be detected by their

prey, over greater distances because lower sound frequencies experience lower atmospheric

attenuation than higher sound frequencies (Griffin 1971; Goerlitz 2018). Call duration was

tested because moths are known to have lower hearing thresholds for longer duration sounds

(Tougaard 1998), meaning that bats with longer calls would be detected by moths at greater

distances than those with shorter calls. Likewise, bats that produce greater amplitude calls,

measured as apparent call source level, can be detected by moths at greater distances than

those with low amplitude calls. Long call intervals provide less information to the moth about

predator positions than faster call rates. Finally, bats with faster flight speeds will close the

distance between themselves and prey more quickly than bats with slower flight speeds. We

used reduced major axis regression (Smith 2009; Trujillo-Ortiz & Hernandez-Walls 2010) to

test for a linear relationships between log10(call peak frequency) and the log10 of each of the

Figure 1. Bat echolocation call peak frequency predicts multiple characteristics of 14 European bat species: a) apparent source level (aSL), b) call interval, c) call duration, and d) flight speed.
Solid and dashed lines show the relationship between bat characteristics and bat call frequency, including and excluding *R. ferrumequinum*, respectively. Note that we performed linear reduced major axis regressions of the logarithmized values, which we backtransformed to the here presented potential function of the non-logarithmized data. P-values report the results for testing the hypothesis that there is no relationship between the log-transformed data with sequential Bonferroni-correction for seven tests..
We excluded the apparent source level of *B. barbastella* due to this species' unusual low-amplitude stealth echolocation strategy (Goerlitz et al. 2010; Lewanzik & Goerlitz 2018) and the duration of *R. ferrumequinum* due to this species' use of high duty-cycle echolocation (Schnitzler 1968) as outliers from all regression analyses (open circles in panels a and c).
For data sources, see Table S1.

four other bat characteristics. We used reduced major axis regression (also called geometric

mean regression) because both variables in the regression equation are random and subject to

error.

Call duration, apparent call source level, call interval, and flight speed all had

significant negative relationships with call peak frequency (**Fig. 1**, major axis regression, all

$p<0.05$ after sequential Bonferroni correction). Bats calling at lower frequencies emit calls of

higher amplitude and longer duration, call less often and fly faster than those calling at higher

frequencies. These strong relationships between bat call frequency and bat characteristics

relevant to predator threat suggests that the frequency tuning of moth auditory receptor cells

might provide an adaptive filter for encoding predation threat across multiple sympatric bat

species.

3. Calculating detection distances between bats and moths

To test whether the frequency tuning of the moth ear might function as a filter to match

detection thresholds with the threat posed by different sympatric bat species, we calculated

the maximum distances over which bats can detect moth echoes (bat detection distance, or

bat-DD) and over which the A1 and A2 cells can be triggered by bat calls (A1-DD and A2-

DD, **Fig. S2**). We calculated these values for all combinations of 14 European bat species

(**Table S1**) and 12 European moth species (**Table S2**). Detection distances were calculated

using the sonar equation (Møhl 1988), sound attenuation values based on mean weather

conditions during our recordings (16°C, 75% relative humidity), estimated bat and measured

moth hearing thresholds (see below), bat call source levels (**Table S1**) and moth target

strength measurements (TS: echo level relative to the impinging sound level, **Table S2**; for

details, see Goerlitz et al. 2010 supplementary methods).

Bat hearing thresholds have been estimated to be approximately between 0-20 dB SPL

(Kick 1982, Neuweiler et al 1984). Therefore, we set hearing threshold to 20 dB SPL for all

bat species to account for intrinsic noise of the auditory system and (often even higher)

behavioural reaction thresholds (Troest & Møhl 1986, Møhl 1988, Surlykke et al. 1999;

Lewanzik & Goerlitz 2018). Moth hearing thresholds for bat calls were derived from

published A1 and A2 thresholds for 20 ms pure tones (ter Hofstede et al. 2013; **Fig. 1**). First,

we calculated the threshold at the bat species' peak frequency by linear interpolation between

the measured pure-tone thresholds at the frequencies below and above the bats' peak

frequency (for *R. ferrumequinum* calling at 82 kHz, we conservatively used the highest

measured threshold at 80 kHz). Second, to account for differences in duration between pure

tones and bat calls, we increased or reduced this threshold by 1.85 dB per halving or doubling

of duration, respectively, to match the bat species' call duration. This correction value is

based on the relationship between sound duration and auditory threshold for two noctuid

moth species (Tougaard 1998). These audiogram-derived A1 and A2 detection distances were

additionally divided by 1.5 and 1.2, respectively. This is an accurate correction to obtain

realistic estimates of maximum detection distances for bat calls in the field based on lab-

measured audiograms (own unpublished data). Target strength was obtained from each

moth's surface-area as TS @ 10 cm = 13.5 dB * lg(SA) – 48.6 dB (Surlykke et al. 1999: Fig.

2a, mean of 30 and 100 kHz). Moth surface area was measured (ImageJ, National Institute of

Health, U.S.A.) from digital photographs of moth specimens with wings completely spread

(7±3 specimens per species (mean ± std), **Table S2**).

4. Evading distant bats: modelling moth behaviour in response to A1 cell activity

4.1 The Constant Buffer Hypothesis

Noctuid moths show directional flight away from quiet pulses of ultrasound (Roeder 1962,

1964; Agee 1969). The A1 cell of the moth ear is more sensitive than the A2 cell (**Fig. S1**;

Fullard 1998; ter Hofstede et al. 2013), and directional flight is likely triggered at intensities

just above A1 threshold and below A2 threshold (Roeder 1964, 1967). Previous studies have

shown that A1-DD is much greater than bat-DD (Roeder 1998; Surlykke et al. 1999; Goerlitz

et al. 2010), meaning that moths can initiate directional flight before being detected by the

bat. Bats with lower frequency echolocation calls can detects moths at greater distances than

those with higher frequency echolocation calls, and most moths can detect bats with lower

frequency echolocation calls at greater distances than bats with higher frequency

echolocation calls (Surlykke et al. 1999). It is not known, however, how this correlation

between detection distances relates to moth evasive behaviour.

The *Constant Buffer Hypothesis* (**Fig. 2A**) suggests that the shape of the A1 tuning

curve allows moths to detect bats with different echolocation call frequencies such that moths

initiate directional flight in time to avoid detection by each bat species. Specifically, it

predicts that if the shape of the moth auditory tuning curve is adaptive and the moth initiates

directional flight away from the bat close to A1 threshold, then the moth will fly out of the

path of an approaching bat with a similar buffer distance for each bat species.

Figure 2. Predictions of the constant buffer hypothesis (a) and model results (b).

(a) Conceptual figure showing the predicted results if the constant buffer hypothesis is correct and moths initiate directional evasive flight at the onset of A1 cell activity. The constant buffer hypothesis postulates that directional evasive flight allows moths to escape detection by a bat at a constant buffer distance before detection by the bat would have occurred, across all sympatric bat species for any given moth species. The shape of the curves in the bat-DD and A1-DD plots reflect calculated values (see **Fig. S2**).

(b) Modelled A1-buffer distances as a function of bat call frequency. Each coloured line represents one moth species. The initial off-axis position of the moth in relation to the bat flight and sonar beam direction is given at the top left of each panel. Open/closed symbols: moths that escaped detection and moths that were detected by the bat, respectively. Model parameters: 20° sonar beam width, 5 m/s moth flight speed.
N: number of moth-bat-combinations (of 168) where the moth (i) escaped / (ii) was detected (incl. those where the moth's initial position was within the bat's search cone) / (iii) was outside the bat's detection tunnel at the initial position and thus never detectable for the bat.
Sign.: number of linear regressions (of 12 moth species) whose slope is significantly different from Zero, separately for regressions including / excluding *B. barbastellus*. Arrowhead: *B. barbastellus* (36 kHz).

215
216

4.2 Model design

To test the Constant Buffer Hypothesis, we designed a geometric model that simulated linear

bat search flight and directional evasive flight of moths upon detecting a bat call (cf.,

Corcoran & Conner 2016; Domenici 2002; Domenici et al. 2011; **Fig. 3**). Modelled bats flew

along a straight path and emitted search calls of average duration, average call interval,

average source level (**Table S1**), and half-amplitude beam widths of 15, 20, 25, 30 and 35° in

the direction of flight (Ghose & Moss 2006) to cover documented variation (Jakobsen et al.

2012; Surlykke et al. 2009). We used the piston model (Surlykke et al. 2009) to calculate

angle-dependent source levels (**Fig. 3c**). We used multiple angles of the moth's initial

position relative to the bat's flight and sonar beam direction to determine how this off-axis

angle would influence the moth's escape probability (15 off-axis angles in total: 0.5, 1, 1.5,

2.5, 5, 7.5, 10, 15, 20, 25, 30, 40, 50, 70 and 90°). The initial distance between the modelled

bat and moth was determined by the A1 detection distance (**Fig. 3a**), meaning that the

received sound pressure level of the bat calls were just at A1 threshold (Roeder 1964, 1967).

The model simulated the flight trajectory of the bat and moth with time steps

equalling the call interval of the bat. After the emission of each call, bat and moth positions

were altered by the distances they fly within one call interval. Bats flew with their average

flight speed (**Table S1**) along their linear search path, starting with the first time step. Moths

in the model initiated directional flight after the second call because moths only demonstrate

sustained directional flight in response to multiple calls at repetition rates typical for bat

echolocation (Roeder 1967). Moths moved directly away from the bat (negative phonotaxis:

Roeder 1962) at six different flight speeds (1, 2, 3, 4, 5 and 6 m/s), encompassing the range

of measured noctuid moth flight speeds (Riley et al. 1992; Vickers & Baker 1997; Luo et al.

2002; Chapman et al. 2008; Corcoran and Conner 2016). After each call, we calculated the

new bat-moth-distance, moth off-axis angle and respective detection distances for the new

positions. The model run was stopped if (i) the moth escaped detection by the bat (i.e., it flew

outside of the bat's detection tunnel formed by the maximum lateral extent of the bat's sonar

beam and its forward movement; **Fig. 3b1**), or (ii) the moth was detected by the bat (bat-

moth-distance was less than or equal to the maximum distance at which the bat can detect the

moth; **Fig. 3b2**). If the moth escaped detection, we calculated the moth's "buffer distance" as

Figure 3. Diagrammatic representation of the model for directional evasive flight.

(a) Model features and terms. Bats searched for prey along a linear trajectory, detecting any prey within their search cone as defined by their maximum detection distance (DD; red line). The maximum width of the bat's search cone determines the bat detection tunnel (dashed grey lines), from which moths need to escape to avoid detection. The moths' starting point was at the maximum detection distance of bats by their A1 cell (blue solid line) and at various off-axis angles relative to the bat's flight trajectory. With each emitted call, bats flew along their linear search trajectory according to their flight speed, while moths flew directly away from the bat (negative phonotaxis) at various potential flight speeds.

(b) End points of the model. The model run ended when the moth escaped by crossing the edge of the bat detection tunnel (b1), or when the moth was detected by crossing the bat's maximum detection distance (b2). For escaped moths, the buffer distance is the shortest distance ever between any moth position and the front of the bat's search cone (b1).

(c) Examples of sonar beam patterns for three different bat species and 20° half-width beam angle, showing the call level emitted into all directions relative to the on-axis call level. While (a) and (b) show simplified sketches of the spatial search cone, (c) shows the realistic sonar beam pattern used in the model to calculate the realistic detection distances for the search cones in (d) and (e). Values below bat names are call peak frequency, duration and apparent source level (at 10 cm distance).

(d) Examples of maximum detection distances of moths by bats (red) and bats by moths (blue, solid: A1, dotted: A2) for three different bat and moth species. Bat species are the same as in (c). Values below moth names are target strength (at 10 cm distance) and surface area.

(e) Example of a model run showing moth positions (blue, each dot is a position per bat call) relative to bat position (red dot) for moth starting positions (large dot) at four different off-axis angles.

the shortest distance ever between the front of the bat's echolocation search beam and the

moth's position during the entire modelled evasive flight (**Fig. 3b,e**). The buffer distance is a

measure for the moth's spatial safety margin, i.e., the shortest distance ever during evasive

directional flight before the moth would have been detected by the bat. If the moth was

detected by the bat, buffer distance was set to zero. If the moth's initial position was already

outside the detection tunnel, as was the case for many of the larger off-axis angles, then

buffer distance was undefined.

We did not attempt to model the behaviour of bats and moths after the bat detected the

moth for two reasons. First, bats have stereotyped echolocation while searching for prey, but

change the structure of their calls and calling pattern once they have detected and pursue prey

(Moss & Surlykke 2010), meaning that we could only effectively model the bats' flight and

calling behaviour prior to moth detection. Second, moths switch from directional evasive

flight to more unpredictable last-ditch evasive flight while being approached by a bat. This

switch might occur around the time when the bat detects the moth. The last-ditch behaviour

varies widely, including loops, zig-zags and dives (Roeder 1962; Agee 1969), and varies

between and sometimes within species (Hügel & Goerlitz 2019), making it currently

impossible to model moth behaviour after the initiation of last-ditch behaviour. Therefore, we

only modelled the interval during which we would expect to see moth directional flight away

from the bat, i.e. after the moth detects the bat but before the bat detects and starts actively

pursuing the moth.

For each moth species, we ran linear regressions between log10(buffer distance) and

log10(bat echolocation call frequency). The log-transformation was used to linearize the data

for statistical analyses. If the Constant Buffer Hypothesis is correct, we predict that buffer

distances within each moth species would be the same for all bat species regardless of

echolocation call frequency, resulting in horizontal lines when buffer distance is plotted

against frequency and a lack of significant regressions (**Fig. 2a**). We evaluated model

performance by counting the number of linear regressions with slopes that are significantly

different from zero (after sequential Bonferroni correction for twelve tests). Twelve

insignificant regressions with slopes of zero would indicate strong support for the hypothesis,

and 12 significant regressions would indicate rejection of the hypothesis.

4.3 Model results

We initially present results (**Fig. 2b**) for a typical sonar beam width of 20° (Surlykke et al.

2009; Jakobsen et al. 2012) and for a moth flight speed of 5 m/s, assuming that moths use a

high flight speed during escape. Moths likely increase their flight speed when they hear bats,

with evidence coming from observations of moths responding to bat attacks in free flight

(Agee 1967, 1969; Corcoran & Conner 2016) and increased wingbeat frequency by tethered

moths during their turning response to low amplitude ultrasound (Treat 1955; Roeder 1967).

At small moth off-axis angles (up to 2.5°), buffer distance was constant across all bat species

for most moth species (0 significant regression; the bat *B. barbastellus* was excluded because

of its unusually low call source levels, **Fig. 2b**). With increasing moth off-axis angle, buffer

distance increased for low-frequency bat species, resulting in fewer relationships that had

constant buffer distances across bat species (only four and three moth species at 5° and 10°,

respectively, excluding *B. barbastellus*). When the moth off-axis angle increased further, the

initial position of a greater percentage of moths (10°: 17%, 15°: 48%, 20°: 74%) was already

outside the detection tunnel, preventing the bat from ever detecting the moth.

Moth flight speed strongly influences the success of directional evasive flight (**Fig.**

4a, top to bottom). At slow flight speeds of 1-2 m/s, all moths flying in the centre of the bat

detection tunnel (0-5° off-axis) are detected by all searching bats (blue line**, Fig. 4a**, top). At

10° off-axis, the proportion of bat-moth-combinations for which the moth escapes detection

298 (green line) reaches almost 50% at 1 m/s and about 90% at 2 m/s. Increasing flight speed

299 further allows more and more moths to escape from the centre of the bat detection tunnel

300 without being detected by the bat. In contrast to moth flight speed, the bat's sonar beam width

301 has a very minor effect on the moths' escape success (**Fig. S4**).

302    The behavioural reaction thresholds of bats and moths are generally not well-known,

303 and might differ from neural detection thresholds. For moths, we used the neural detection

304 thresholds of A1- and A2-cells as estimates for the onset of directional and erratic evasive

305 flight, respectively. However, the exact mechanism linking neuronal activity to evasive flight

306 is debated (Ratcliffe et al. 2009; Surlykke 1984; ter Hofstede & Ratcliffe 2016) and multiple

307 acoustic call properties determine A-cell spiking and evasive flight (Roeder 1964; Waters

308 1996; Gordon & ter Hofstede 2018), which might only start at received levels of 10 dB and

309 more above neuronal thresholds (summarized in Lewanzik & Goerlitz 2018). Likewise, bats'

Figure 4. Effects of variation in behavioural threshold (a-d), moth flight speed (top to bottom) and moth off-axis angle (x-axes) on the success of directional evasive flight. Numbers above each column (+0 or +10 dB) are relative levels of reaction thresholds relative to moth A1 threshold (measured for each species) / bat hearing threshold (20 dB). Blue lines: proportion of moths that were detected by the bat; green lines: moths that escaped detection by the bat, either by flying out of the detection tunnel (black line, "fly") or whose initial position was already outside of the detection tunnel (grey line, "out"). Dotted vertical lines are visual reference lines at 10° and 45° off-axis angle. Model parameters: 20° sonar beam width.

reaction threshold can be 7 dB and more above our assumed threshold of 20 dB SPL

(Lewanzik & Goerlitz 2018). We thus calculated all models again after increasing reaction

thresholds by 10 dB for bats only, moths only and both bats and moths at the same time.

Increasing the reaction thresholds relative to the moth's A1 detection thresholds and

to 20 dB SPL bat threshold also influences the success of directional evasive flight.

Increasing moth reaction thresholds by 10 dB generally lowers the proportion of moths that

escape detection (**Fig. 4a,b)** from about 80% (135 of 168) to 45% (75 of 168, **Fig. S5a**, top

two panels) at a moth flight of 5 m/s. Buffer distance of the escaped moths is somewhat

lower, yet again constant across bat species (**Fig. S5a**), and the proportion of escaped moths

increases with increasing moth flight speed as before with default thresholds (**Fig. 4b**).

Likewise, a similar opposite trend exists when only increasing bat reaction threshold by 10

dB, allowing more moths to escape at larger buffer distances (**Fig. S5a**, third panel), lower

speeds and lower off-axis angles (**Fig. 4c**). When increasing hearing thresholds of both moths

and bats by 10 dB, the proportion of escaped moths, the buffer distance and the overall

pattern is very similar to the default non-increased reaction thresholds (**Fig. 4d, S5a**). This

suggests that the proportion of escaped and detected moths depends more on the relative

difference in reaction threshold between bats and moths, and less on their absolute value,

making our model results robust to variation in the exact reaction thresholds.

329
5. Evading close bats: modelling moth behaviour in response to A2 cell activity

5.1 The Matched Onset and Fixed Onset Hypotheses

To explain how the shape of the A2 cell tuning curve might be adaptive, we propose two

mutually exclusive hypotheses. The *Matched Onset Hypothesis* (**Fig. 5a**) postulates that last-

ditch flight is initiated by the moth at the same distance at which the moth is detected by the

bat (or possibly a constant spatial offset prior to that). In other words, using A2-DD as proxy

for the initiation of last-ditch flight, this hypothesis postulates that the distance at which the

moth A2 cell responds to bat calls (A2-DD) matches the maximum distance at which the bat

can detect the moth (bat-DD). Thus, this hypothesis predicts 1) a slope of 1 for the

relationship between A2-DD and bat-DD (overall, and also within each moth species and

within each bat species); and 2) because bats detect larger moths over larger distances, i.e.

bat-DD is positively correlated with moth size, A2-DD will also be positively correlated with

moth size (**Fig. 5a**).

Alternatively, the *Fixed Onset Hypothesis* (**Fig. 5b**) postulates that last-ditch flight

might be most effective when initiated at a distance to impact that is specific to each bat

species, not necessarily when the bat first detects the moth. This constant distance would be

dependent on the flight speed and manoeuvrability of the attacking bat, but independent of

moth characteristics. In other words, using A2-DD as proxy for the initiation of last-ditch

flight, this hypothesis postulates that the distance at which the moth A2 cell responds to bat

calls (A2-DD) is constant per bat species. Thus, this hypothesis predicts that 1) *within each*

bat species, the slope of the relationship between A2-DD and bat-DD will be 0; and 2) *within*

each bat species, A2-DD will be constant across moth species regardless of moth size (slope

= 0), but this distance might vary between bat species (**Fig. 5b**). Both the Matched Onset and

Fixed Onset hypotheses are based on the assumption that last-ditch flight is correlated with

activity of the A2 cell, and this hypothesis has not been empirically tested in noctuid moths

(ter Hofstede & Ratcliffe 2016). Our purpose is to determine how well the model results fit

this assumption and to refine future hypotheses about the relationship between neural input

and behavioural output in moths.

Figure 5. Predictions for the Matched Onset and Fixed Onset hypotheses (a,b) and model results (c).

(a,b) Conceptual figure showing the predicted results if the matched onset hypothesis (a) or fixed onset hypothesis (b) is correct and moths initiate last-ditch evasive flight at the onset of A2 cell activity.
(a) The Matched Onset hypothesis postulates that onset of erratic flight (approximated by A2-DD) is matched to the detection of the moth by the bat (bat-DD), resulting in a slope of 1 for the relationship between A2-DD and bat-DD, and a positive relationship between A2-DD and moth size.
(b) The Fixed Onset hypothesis postulates that onset of erratic flight (approximated by A2-DD) is a fixed value for each bat species across all moth species, predicting no relationship between A2-DD and bat-DD (slope = 0), and no relationship between A2-DD and moth size (slope = 0).

(c) Model results: relationship between maximum detection distances of bat calls by the A2-cells (A2-DD) and of moth echoes by bats (bat-DD; c1), and between A2-DD and moth size (c2). Each colour represents one moth (blue to green) or bat (red to yellow) species.
Sign.: number of linear regressions whose slope is significantly different from Zero, separately for regressions including / excluding *B. barbastellus*, for results before and after sequential Bonferroni correction. Arrowhead: *B. barbastellus* (36 kHz).

5.2 Model design

To test the Matched Onset and Fixed Onset hypotheses, we tested for the existence of two relationships (i.e., having a slope significantly different from 0) within each bat species: 1) A2-DD vs bat-DD, and 2) A2-DD vs. moth surface area. We log10-transformed data before statistical analyses to achieve linearity. We assessed the support for each hypothesis based on the slopes and proportion of statistically significant regressions. First, the Fixed Onset hypothesis would be supported by significant positive relationships with a slope of 1 between A2-DD and bat-DD, and by significant positive relationships between A2-DD and moth size within bat species (**Fig. 5a**). In contrast, the Fixed Onset hypothesis would be supported by a lack of significant relationships and horizontal lines (slope of 0) between A2-DD and bat-DD and between A2-DD and moth size within bat species (**Fig. 5b**). We evaluated model performance by counting the number of linear regressions with slopes that are significantly different from zero (after sequential Bonferroni correction for 28 tests: 14 tests for A2-DD vs. bat-DD (one per bat species) and 14 tests for A2-DD vs. moth surface area (one per bat species)).

5.3 Model results

The mutually exclusive Matched Detection and Fixed Onset Hypotheses for the function of last-ditch evasive flight postulate that the A2-DDs are either matched to the bat's detection distance of the moth, or alternatively are constant (fixed) for each bat species. For the test of prediction 1 (**Fig. 5c1**), approximately one third of the bat species (5 of 14 and 13) had significant relationships between A2 detection distance and bat detection distance. The five bat species with statistically significant slopes were the two bat species with the lowest call frequencies (*Nyctalus lasiopterus* and *N. noctula*) and three of the four bat species with

the highest call frequencies (*Myotis* sp.*, Pipistrellus pygmaeus* and *Rhinolophus*

ferrumequinum). Only one of these relationships (*N. lasiopterus*) remained significant after

sequential Bonferroni correction. Therefore, the A2 cells of all moth species start to fire at

similar distances within almost all bat species, regardless of the bat's detection distances for

the moth, providing support for the Fixed Onset hypothesis and refuting the Matched Onset

hypothesis.

For the test of prediction 2 (**Fig. 5c2**), approximately one third of the bat species (5 of

14 and 13) had significant relationships between A2 detection distance and moth size. The

five bat species with statistically significant slopes were the two bat species with the lowest

call frequencies (*Nyctalus lasiopterus* and *N. noctula*) and the three bat species with the

highest call frequencies (*Miniopterus schreibersii, Pipistrellus pygmaeus* and *Rhinolophus*

ferrumequinum). None of these relationships remained significant after sequential Bonferroni

correction. Therefore, the A2 cells of all moth species start to fire at similar distances within

those bat species, regardless of moth size, providing support for the Fixed Onset hypothesis

and refuting the Matched Onset hypothesis.

Increasing the moths' reaction threshold relative to the A2 cell detection threshold

reduces the A2 detection distance for bat calls (**Fig. S5b2, second row**). Despite this, the

number of non-significant relationships of A2-DD with moth size did not change, providing

continued support for the Fixed Onset hypothesis. The number of significant relationships

between A2-DD and bat-DD did not change when the moths' reaction threshold was

increased, and increased from 1 to 2 significant relationships when the bat reaction threshold

increased by 10 dB (**Fig. S5b1**). In summary, the Fixed Onset hypothesis is almost always

supported for all tested moth and bat reaction thresholds.

6. Discussion

Our model results support the hypotheses that 1) echolocation call frequency of bats is

correlated with variables that correspond with their predator threat, 2) the shape of the A1 cell

tuning curve is adapted to allow moths to avoid detection by bats with a similar spatial safety

margin across bat species in their community, and 3) bats of the same species are detected by

the moth A2 cell at a similar distance, regardless of moth species or size. In addition, the

model suggests that the A1 cell is adapted for the worst-case scenario, allowing moths to

avoid detection by bats when they are directly in the path of an oncoming bat, resulting in

unnecessary defensive behaviour at greater angles away from a bat flying in a straight line.

Together, these results suggest that the tuning of the moth ear is adapted to allow moths to

respond to each bat species at an appropriate distance based on the frequency of its

echolocation calls and suggest avenues for future empirical research and hypothesis testing.

6.1 Bat call frequency predicts threat-related bat characteristics

Larger bat species generally emit calls of lower frequency, greater call source level, longer

duration and lower repetition rate than smaller bats (Barclay & Brigham 1991; Vaughan et al.

1997; Jones 1999; Holderied & von Helversen 2003). Our results demonstrate for the first

time that many of these bat traits (call source level, call duration, call interval) as well as

flight speed are tightly correlated with bat call peak frequency (cf. Vaughan et al. 1997;

Waters et al. 1995 for call duration). These traits determine the maximum prey detection

distance and the minimum time to prey interception, thus strongly influencing the risk that

any given bat species poses for its prey. This correlation turns the bats' (perceptible) call

frequency into a proxy for its (imperceptible) predation threat. We suggest that the frequency-

dependent tuning of moth auditory cells is a functional adaptation exploiting this correlation,

allowing moths to respond to different sympatric bat species at appropriate distances.

6.2 Constant Buffer Hypothesis (onset of directional evasive flight)

Moths can detect bats at much greater distances than bats can detect moths (e.g.

Holderied & von Helversen 2003; Goerlitz et al. 2010), giving moths the opportunity to

initiate evasive flight before being detected. Our model of evasive directional flight based on

measured moth auditory thresholds suggests that moth ears are adapted for the worst-case

scenario, allowing moths to avoid detection by a bat even when they are in the centre of the

bat's flight path and echolocation beam. Remarkably, under these worst-case conditions, the

buffer distance at which the modelled moths escape detection is relatively constant across bat

species (0-10 m; **Fig. 2b**). This result suggests that the shape of the moth auditory tuning

curve is adapted to allow moths to escape sympatric bats with a similar safety margin

regardless of the frequency of a bat's echolocation calls.

Since larger moths are detected by bats over longer ranges then small moths, Surlykke

et al. (1999) already suggested that larger moths might need to detect bats at greater distances

and require more time to escape the echolocation beam of bats than smaller moths. Our

model supports this hypothesis, demonstrating that differences in detection range between

large and small moths result in similar buffer distances of escape from the echolocation beam

of an oncoming bat. Likewise, the different detection ranges for lower and higher frequency

calls results in similar buffer distances of escape for low- and high-frequency bats.

Our model suggests that the buffer distances for escaping detection by a bat become

more variable for larger off-axis angles of the moth relative to the bat's flight and sonar beam

direction. This large variation could be an accurate representation of buffer distances or it

could be due to limitations of our model. If these values are accurate, moths detect lower

frequency bats at much greater distances than necessary when they are off-axis from the

oncoming bat. Unnecessary evasive flight reduces feeding and mating opportunities, but this

might be an acceptable cost given the importance of avoiding predation. The simplicity of the

moth ear likely constrains optimal adaptation to the full range of potential bat-moth

orientations. On the other hand, our model limited the bat to fly in a straight line and to direct

its echolocation calls straight forward, whereas foraging bats turn frequently and move their

sonar beam to scan their surroundings (Surlykke & Moss 2000). The detection tunnel of a

naturally scanning bat will be wider than modelled here, and the bat's turns will increase the

chances that a moth suddenly finds itself in the echolocation beam of a bat that is too close

for effective escape by directional flight. The seemingly larger than necessary buffer

distances might help moths to escape from turning and scanning bats. Lastly, the largest

variation in buffer distances occurred at the lowest frequency (17 kHz), corresponding with

the bat *Nyctalus lasiopterus*. We included *N. lasiopterus* to cover the full frequency range of

European aerial hawking bat species, but it does not occur in the UK. Therefore, this bat

species is not sympatric with the tested moth specimens in this study, which might result in

relaxed selection at this frequency range for these moths (c.f. ter Hofstede et al. 2013).

Another unknown is the directionality of hearing of bats, which is additive to the sonar beam

directionality and will narrow the width of the bat's detection tunnel. Hearing directionality is

mostly unknown for bats, although for *Rhinolophus ferrumequinum* it is between 13-25°

(Schnitzler & Grinnell 1977, Matsuta et al 2013), i.e., in the range of the sonar beam

directionality, or even more narrow. More data on hearing directionality, hearing direction,

their variation due to ear movements (e.g., Griffin et al 1962; Kugler et al 2017, Lattenkamp

et al 2018, Yin & Mülller 2019), and on natural scanning behaviour will be required to

estimate the average prey detection cone of free-flying bats.

Our model necessarily simplifies bat and moth behaviours and contains only a subset

of the variables that could influence buffer distance. Future studies that collect data on the

following variables would contribute to more accurate and precise models of bat-moth

behaviour and help answer questions about the evolution of moth hearing for defence against

bats. First, more detailed information on behavioural thresholds and how they relate to neural

thresholds are needed. While correlative studies show that moths initiate directional flight at

sound levels close to A1 cell threshold (Roeder 1962, 1967), direct measurements of A1

activity in behaving moths would help to confirm the exact relationship between neural

activity and behaviour. Likewise, bat reaction thresholds are unknown and were set to 20 dB

SPL, an estimate of hearing threshold for many bat species, although behavioural reaction

thresholds can be higher (Lewanzik & Goerlitz 2018). Our analysis, however, shows that

model results are rather robust against some variation in threshold and mostly depend on their

relative values. Although our model did not include the moths' reaction time (ranging

between 45-250 ms from sound arrival to response onset; Roeder 1998), the model moths

started their directional flight after the second bat call, reducing the influence of this variable

on the results. Second, we assumed that moths would fly directly away from the sound source

based on observational descriptions of moth behaviour (Roeder 1962), but three-dimensional

flight path data are needed to confirm this. Third, incorporating species-specific data on moth

flight speeds would improve the accuracy of the model. Currently, we do not have data for

the species in our sample to incorporate this variable into the model. Most studies that report

flight speeds for moths focus on a few species of economic importance and measure this

variable in the context of flight in a pheromone plume. Studies have shown, however, that

multiple moth species significantly reduce their flight speeds once they enter a pheromone

plume and flight speed decreases with increasing pheromone concentration (*Lymantria*

dispar: Carde & Hagaman 1979, Charlton et al. 1993; *Heliothis virescens*: Vickers & Baker

1997; *Spodoptera littoralis*: Murlis & Bettany 1977). In addition, moths increase flight speed

with height above the ground (Keunen & Baker 1982), but the majority of studies investigate

moth flight in wind tunnels and encounters between bats and moths would typically occur at

greater distances from the ground. Therefore, we only considered flight speeds that were

measured in noctuid moths in the absence of pheromone and in free flight outside of a wind

tunnel for the model. Moth size also likely influences flight speed. Kuenen & Carde (1993)

show that larger individuals of the moth *Lymantria dispar* fly faster than smaller individuals,

and Dudley & Srygley (1994) found that larger butterflies flew faster than smaller butterflies.

In both cases, this relationship is driven by weight and wing loading, not surface area. To

overcome these limitations, we modelled all moth species at the same flight speed, but varied

flight speed to determine its effect on the model results.

Despite these simplifications, our model predicts remarkably similar buffer distances

across bat and moth species, suggesting that the tuning of the moth ear is a functional

adaptation to the threat posed by bats with different call frequencies. Moths only escaped bats

at higher flight speeds, providing specific predicted values for moth flight speeds while

evading bats that can be tested in empirical studies. Future studies should consider collecting

comparative data across a range of moth species of different sizes on 1) neural thresholds for

turning responses, 2) 3D flight path tracking of moths evading distant moths (as measured by

Corcoran and Conner 2016 for close range attacks), and 3) the relationship between moth size

and flight speed in different contexts (both searching flight and directional evasive flight).

6.3 Matched Onset and Fixed Onset Hypotheses (onset of last-ditch evasive flight)

Our model results provide support for the hypothesis that the A2 cell is adapted to fire

at a fixed distance between a particular bat species and all sympatric moths regardless of

moth species and size. If the A2 cell activity triggers last-ditch behaviour, this suggests that

last-ditch flight might be most effective when initiated at a distance to impact that is specific

to each bat species, not necessarily when the bat first detects the moth (as predicted by the

matched onset hypothesis). In particular, it suggests that the ideal distance for moths to react

to bats might be more influenced by aspects of bat flight speed and manoeuvrability than

moth-specific parameters. Variation in the difference between A1 and A2 best thresholds

across moth species of different sizes might contribute to this pattern. Although larger moths

are more sensitive to ultrasound than smaller moths, the relationships between A1 threshold

and moth surface area has a much steeper slope than that of A2 and moth surface area (ter

Hofstede et al. 2013). Therefore, larger and smaller moths are more similar in A2 thresholds

than A1 thresholds, contributing to this pattern of similar A2 detection distances seen across

moths of different sizes in our model.

Despite almost complete support for the fixed onset over the matched onset

hypothesis, there was sometimes still marked variation in A2-DD across moth species within

a bat-species, particularly for low-frequency bats. Two factors contributed to this: bat-DD

varied more with moth size for low- than high-frequency bats (**Fig. S6**), and A2-DD varied

little with moth size for most bats, except for those with the lowest and highest call

frequencies. Moth ears might be constrained to provide the most reliable representation of

predator threat at intermediate frequencies, with bat species calling at extremely low or high

frequencies having an advantage over moths, as proposed by the allotonic frequency

hypothesis (Fullard 1998). Indeed, bats that call at extremely low or high frequencies are

known to be moth specialists (*Euderma maculatum* 12 kHz: Fullard & Dawson 1997, Painter

2009; *Hipposideros ater* 160 kHz: Pavey & Burwell 1998).

Our model suggests two hypotheses for future empirical studies on the relationship

between moth hearing and last-ditch anti-bat behaviour. Future studies should test whether

A2 cell activity triggers last-ditch flight behaviour and whether this is true across all noctuid

moth species or perhaps varies between species. Likewise, distances between bats and moths

of known species and auditory sensitivity should also be measured with 3D tracking to

confirm or refute that moths initiate last-ditch flight at a constant specific distance for each

bat species.

6.4 Conclusions

To avoid escaping a predator too late or too early, prey animals must correctly estimate the

threat of multiple predators. Threat, however, depends on many predator-specific parameters

and cannot always be directly perceived or assessed. In contrast, biophysical constraints in

predators can result in correlations between particular predator cues that are detectable by

prey, such as size or sound frequency, and the level of threat posed by a predator. Prey

animals can benefit from sensory systems tuned to detectible cues in predators that provide

information about potential predation risk across different predators. In echolocating bats, the

sound frequency of their echolocation calls is a reliable and perceptible predictor of predation

threat for eared insects. The frequency tuning of auditory receptor cells of eared moths

appears to exploit this relationship, showing that even a very simple sensory system can adapt

to deliver information suitable for triggering appropriate defensive reactions to each predator

in a multiple predator community.

Acknowledgements: Data collection was funded by the BBSRC (BB/f002386/1) to MWH.

Model development and analysis was funded by the German Research Foundation (Emmy

Noether, GO 2091/2-1) to HRG. HtH was supported by funding from Dartmouth College. We

thank J. Memmott and G. Jones for assistance with collecting moths, Jim and Di McPetrie

(Middlegrounds Farm Slapton), Slapton Ley Field Center, Mottisfont and Bristol City

Council for site access and logistic support, D. Schröder and O. Nommensen for help with

data analysis, L. Jakobsen for providing the Matlab code to estimate call beam shape, T.

Hügel for discussion, and B. Hedwig, K. Kostarakos, T. Bayley, J. Memmott and G. Jones for

constructive feedback on earlier versions of this manuscript.

Author contributions:

Devised the general research idea: HRG, HMtH, MWH

Collected and analysed moth neurophysiological data: HMtH

Collected and analysed bat data: HRG

Devised and developed the model: HRG, HMtH

Run and analysed the models: HRG

Wrote the manuscript: HRG, HMtH

Reviewed the final manuscript: HRG, HMtH, MWH

References

Agee H.R. 1967 Response of acoustic sense cell of bollworm and tobacco budworm to ultrasound. *J*
Econ Entomol **60**(2), 366-369.

Agee H.R. 1969 Response of flying bollworm moths and other tympanate moths to pulsed
ultrasound. *Ann Entomol Soc Am* **62**(4), 801-807.

Andersson S., Rydell J., Svensson M.G.E. 1998 Light, predation and the lekking behaviour of
the ghost swift *Hepialus humuli* (L.) (Lepidoptera: Hepialidae). *Proc R Soc Lond B*
264(1403): 1345-1351.

Barlow, K.E. 1997 The diets of two phonic types of the bat *Pipistrellus pipistrellus* in Britain. *J Zool,*
Lond **243**(3), 597-609.

Barclay R.M.R., Brigham R.M. 1991 Prey detection, dietary niche breadth, and body size in bats -
why are aerial insectivorous bats so small. *Am. Nat.* **137**(5), 693-703.

Bogdanowicz W., Fenton M.B., Daleszcyk K. 1999 The relationship between echolocation calls,
morphology and diet in insectivorous bats. *J Zool, Lond* **247**(3), 381-393.

Chapman J.W., Reynolds D.R., Mouritsen H., Hill J.K., Riley J.R., Sivell D., Smith A.D., Woiwod
I.P. 2008 Wind selection and drift compensation optimize migratory pathways in a high-flying
moth. *Curr. Biol.* **18**(7), 514-518.

Corcoran A.J., Conner W.E. 2012 Sonar jamming in the field: effectiveness and behavior of a unique
prey defense. *J. Exp. Biol.* **215**, 4278-4287.

Corcoran A.J., Conner W.E. 2016 How moths escape bats: predicting outcomes of predator-prey
interactions. *J. Exp. Biol.* **219**(17), 2704-2715.

Dietz C., von Helversen O., Nill D. 2009 Bats of Britain, Europe and Northwest Africa. A & C Black
Publishers Ltd., London.

Domenici P. 2002 The visually mediated escape response in fish: Predicting prey responsivness and
the locomotor behaviour of predators and prey. *Mar Freshw Behav Phy* **35**(1-2), 87-110.

Domenici P., Blagburn J.M., Bacon J.P. 2011 Animal escapology I: theoretical issues and emerging
trends in escape trajectories. *J. Exp. Biol.* **214**(15), 2463-2473.

Eaton R.C., Lee R.K.K., Foreman M.B. 2001 The Mauthner cell and other identified neurons of the
brainstem escape network of fish. *Prog. Neurobiol.* **63**(4), 467-485.

Edwards D.H., Heitler W.J., Krasne F.B. 1999 Fifty years of a command neuron: the neurobiology of
escape behavior in the crayfish. *Trends in Neurosciences* **22**(4), 153-161.

Falk J.J., ter Hofstede H.M., Jones P.L., Dixon M.M., Faure P.A., Kalko E.K.V., Page R.A. 2015
Sensory-based niche partitioning in a multiple predator - multiple prey community. *Proc. R. Soc.*
Lond. B **282**(1808).

Fenton M.B., Faure P.A., Ratcliffe J.M. 2012 Evolution of high duty cycle echolocation in
bats. *J Exp Biol* **215**(17), 2935-2944.

Fotowat H., Gabbiani F. 2007 Relationship between the phases of sensory and motor activity during
a looming-evoked multistage escape behavior. *J. Neurosci.* **27**(37), 10047-10059.

Fraker M.E. 2008 The dynamics of predation risk assessment: responses of anuran larvae to chemical
cues of predators. *J. Anim. Ecol.* **77**(4), 638-645.

Fullard J.H. 1998 The sensory coevolution of moths and bats. In *Comparative Hearing: Insects* (eds.
Hoy R.R., Popper A.N., Fay R.R.), pp. 279-326. New York, Springer-Verlag.

Fullard J., Dawson J. 1997 The echolocation calls of the spotted bat *Euderma maculatum* are
relatively inaudible to moths. *J. Exp. Biol.*, **200**(1), 129-137.

Ghose K., Moss C.F. 2006 Steering by hearing: A bat's acoustic gaze is linked to its flight motor
output by a delayed, adaptive linear law. *J. Neurosci.* **26**(6), 1704-1710.

Goerlitz H.R. 2018 Weather conditions determine attenuation and speed of sound: Environmental
limitations for monitoring and analyzing bat echolocation. *Ecol Evol* **8**(10), 5090-5100.

Goerlitz H.R., ter Hofstede H.M., Zeale M.R.K., Jones G., Holderied M.W. 2010 An aerial-hawking

bat uses stealth echolocation to counter moth hearing. *Curr. Biol.* **20**(17), 1568-1572.

Gordon S.D., ter Hofstede H.M. 2018 The influence of bat echolocation call duration and timing on

auditory encoding of predator distance in noctuoid moths. *J. Exp. Biol.*, **221**(6), jeb171561.

Griffin D.R. 1971 The importance of atmospheric attenuation for the echolocation of bats

(Chiroptera). *Anim. Behav.* **19**(1), 55-61.

Griffin D.R., Dunning D.C., Cahlander D.A., Webster F.A. 1962 Correlated orientation sounds and

ear movements of horseshoe bats. *Nature* **196**, 1185-1186.

Helfman G.S. 1989 Threat-sensitive predator avoidance in damselfish-trumpetfish interactions.

Behav. Ecol. Sociobiol. **24**(1), 47-58.

Holderied M.W., von Helversen O. 2003 Echolocation range and wingbeat period match in aerial-

hawking bats. *Proc. R. Soc. Lond. B* **270**, 2293-2299.

Hügel T., Goerlitz H.R. 2019 Species-specific strategies increase unpredictability of escape flight in

eared moths. *Func. Ecol.* (online). DOI: 10.1111/1365-2435.13383.

Jakobsen L., Ratcliffe J.M., Surlykke A. 2012 Convergent acoustic field of view in echolocating

bats. *Nature* **493**, 93-96.

Jones G. 1999 Scaling of echolocation call parameters in bats. *J. Exp. Biol.* **202**(23), 3359-

3367.

Jones G., Rayner J.M. 1989 Foraging behavior and echolocation of wild horseshoe bats

Rhinolophus ferrumequinum and *R. hipposideros* (Chiroptera, Rhinolophidae). *Behav.*

Ecol. Sociobiol. **25**(3), 183-191.

Jones G., Teeling E.C. 2006. The evolution of echolocation in bats. *TREE* **21**(3), 149-156.

Kick S.A. 1982 Target-detection by the echolocating bat, *Eptesicus fuscus*. *J. Comp. Physiol.*

A **145**, 431-435.

Kugler K., Wiegrebe L. 2017 Echo-acoustic scanning with noseleaf and ears in phyllostomid

bats. *J. Exp. Biol.* **220**, 2816-2824.

Lazure L., Fenton M.B. 2011. High duty cycle echolocation and prey detection by bats. *J.*

Exp. Biol. **214**(7), 1131-1137.

Lattenkamp E.Z., Kaiser S., Kaučič S., Großmann M., Koselj K., Goerlitz H.R. 2018

Environmental acoustic cues guide the biosonar attention of a highly specialised

echolocator. *J. Exp. Biol.* **221**, jeb165696.

Lewanzik D., Goerlitz H.R. 2018 Continued source level reduction during attack in the low-

amplitude bat *Barbastella barbastellus* prevents moth evasive flight. *Func. Ecol.* **32**(5), 1251-

1261.

Luo L.Z., Johnson S.J., Hammond A.M., Lopez J.D., Geaghan J.P., Beerwinkle K.R., Westbrook
J.K. 2002 Determination and consideration of flight potential in a laboratory population of true
armyworm (Lepidoptera : Noctuidae). *Environ Entomol* **31**(1), 1-9.

Matsuta, N., Hiryu, S., Fujioka, E., Yamada, Y., Riquimaroux, H., Watanabe, Y. 2013 Adaptive
beam-width control of echolocation sounds by CF-FM bats, *Rhinolophus ferrumequinum nippon*,
during prey-capture flight. *J. Exp. Biol.* **216**, 1210-1218.

Møhl B. 1988 Target detection by echolocating bats. In *Animal Sonar. Processes and Performances.*
(eds. Nachtigall P.E., Moore P.W.B.), pp. 435-450. New York, London, Plenum Press.

Moss C.F., Surlykke A. 2010 Probing the natural scene by echolocation in bats. *Front. Behav.*
Neurosci. **4**(33).

Neuweiler G., Singh S., Sripathi K. 1984 Audiograms of a South-Indian bat community. *J. Comp.*
Physiol. A **1984**, 133-142.

Nolen T.G., Hoy R.R. 1984 Initiation of behavior by single neurons: the role of behavioral context.
Science **226**(4677), 992-994.

Painter ML., Chambers C.L., Siders M., Doucett R.R., Whitaker, Jr J.O., Phillips D.L. 2009 Diet of
spotted bats (*Euderma maculatum*) in Arizona as indicated by fecal analysis and stable isotopes.
Can. J. Zool., **87**(10), 865-875.

Pavey C.R., Burwell C.J. 1998 Bat predation on eared moths: a test of the allotonic frequency
hypothesis. *Oikos*, **81**, 143-151.

Ratcliffe J.M., Fullard J.H. 2005 The adaptive function of tiger moth clicks against
echolocating bats: an experimental and synthetic approach. *J. Exp. Biol.*, **208**(24), 4689-
4698.

Ratcliffe J.M., Fullard J.H., Arthur B.J., Hoy R.R. 2009 Tiger moths and the threat of bats: decision-
making based on the activity of a single sensory neuron. *Biol. Lett.* **5**(3), 368-371.

Ratcliffe J.M., Fullard J.H., Arthur B.J., Hoy R.R. 2011 Adaptive auditory risk assessment in the
dogbane tiger moth when pursued by bats. *Proc. R. Soc. Lond. B* **278**(1704), 364-370.

Riley J.R., Armes N.J., Reynolds D.R., Smith A.D. 1992 Nocturnal observations on the emergence
and flight behavior of *Helicoverpa armigera* (Lepidoptera, Noctuidae) in the post-rainy season in
Central India. *B Entomol Res* **82**(2), 243-256.

Ritzmann R.E., Tobias M.L., Fourtner C.R. 1980 Flight activity initiated via giant interneurons of the
cockroach: evidence for bifunctional trigger interneurons. *Science* **210**(4468), 443-445.

Roeder K.D. 1962 The behaviour of free flying moths in the presence of artificial ultrasonic pulses.
Anim. Behav. **10**(3-4), 300-304.

Roeder K.D. 1964 Aspects of the noctuid tympanic nerve response having significance in the
avoidance of bats. *J. Insect Physiol.* **10**(4), 529-546.

Roeder K.D. 1967 Turning tendency of moths exposed to ultrasound while in stationary flight. *J.*
Insect Physiol. **13**(6), 873-888.

Roeder K.D. 1974 Acoustic sensory responses and possible bat-evasion tactics of certain moths. In
Proc. Canadian Society of Zoologists' Annual Meeting (ed. Burt M.D.B.), pp. 71-78. Fredericton,
University of New Brunswick Press.

Roeder K.D. 1998 *Nerve cells and insect behavior. Revised edition of the first edition of 1976.*
Cambridge, Harvard University Press.

Römer H. 2016 Matched filters in insect audition: tuning curves and beyond. In *The Ecology*
of Animal Senses: Matched Filters for Economical Sensing (eds. von der Emde G.,
Warrant E.), pp. 83-109. Cham, Switzerland, Springer.

Schnitzler H.-U. 1968 The ultrasonic sounds of horseshoe bats (Chiroptera, Rhinolophidae) in
different orientation situations. *Zeitschrift für Vergleichende Physiologie* **57**(4), 376-408.

Schnitzler H.-U., Grinnell, A. D. 1977 Directional sensitivity of echolocation in the horseshoe bat,
Rhinolophus ferrumequinum I. Directionality of sound emission. *J. Comp. Physiol. A* **116**, 51-61.

Smith R.J. 2009 Use and Misuse of the Reduced Major Axis for Line-Fitting. *Am J Phys Anthropol*
140(3), 476-486.

Smolka J., Zeil J., Hemmi J.M. 2011 Natural visual cues eliciting predator avoidance in fiddler crabs.
Proc. R. Soc. Lond. B **278**(1724), 3584-3592.

Surlykke A. 1984 Hearing in Notodontid moths: a tympanic organ with a single auditory neuron. *J.*
Exp. Biol. **113**(Nov), 323-335.

Surlykke A., Filskov M., Fullard J.H., Forrest E. 1999 Auditory relationships to size in noctuid
moths: bigger is better. *Naturwissenschaften* **86**(5), 238-241.

Surlykke A., Moss C.F. 2000 Echolocation behavior of big brown bats, *Eptesicus fuscus*, in the field
and the laboratory. *J. Acoust. Soc. Am.* **108**(5), 2419-2429.

Surlykke A., Pedersen S.B., Jakobsen L. 2009 Echolocating bats emit a highly directional sonar
sound beam in the field. *Proc. R. Soc. Lond. B* **276**(1658), 853-860.

Templeton C.N., Greene E., Davis K. 2005 Allometry of alarm calls: Black-capped chickadees
encode information about predator size. *Science* **308**(5730), 1934-1937.

ter Hofstede H.M., Goerlitz H.R., Ratcliffe J.M., Surlykke A., Holderied M. 2013 The simple ears of
noctuid moths are tuned to the calls of their sympatric bat community. *J. Exp. Biol.* **216**, 3954-
3962.

ter Hofstede H.M., Ratcliffe J.M. 2016 Evolutionary escalation: the bat-moth arms race. *J. Exp. Biol.*
219(11), 1589-1602.

Tougaard J. 1998 Detection of short pure-tone stimuli in the noctuid ear: what are temporal
integration and integration time all about? *J. Comp. Physiol. A* **183**(5), 563-572.

Treat A.E. 1955 The response to sound in certain Lepidoptera. *Ann Entomol Soc Am* **48**(4), 272-284.

Troest N., Møhl B. 1986 The detection of phantom targets in noise by serotine bats; negative
evidence for the coherent receiver. *J. Comp. Physiol A* **159**, 559-567.

Trujillo-Ortiz A., Hernandez-Walls R. 2010 gmregress: Geometric Mean Regression (Reduced
Major Axis Regression. A MATLAB file. *URL:*
http://www.mathworks.com/matlabcentral/fileexchange/27918-gmregress.

Vaughan N. 1997 The diets of British bats (Chiroptera). *Mammal Rev* **27**(2): 77-94.

Vaughan N., Jones G., Harris S. 1997 Identification of British bat species by multivariate analysis of
echolocation call parameters. *Bioacoustics* **7**, 189-207.

Vickers N.J., Baker T.C. 1997 Flight of *Heliothis virescens* males in the field in response to sex
pheromone. *Physiol. Entomol.* **22**(3), 277-285.

von der Emde G., Warrant E. (Eds.). 2016 The Ecology of Animal Senses: Matched Filters
for Economical Sensing. Springer, Cham, Switzerland.

Waters D. 1996 The peripheral auditory characteristics of noctuid moths: information encoding and
endogenous noise. *J Exp Biol*, **199**(4), 857-868.

Waters D.A., Rydell J., Jones G. 1995 Echolocation call design and limits on prey size: a case-study
using the aerial hawking bat *Nyctalus leisleri Behav. Ecol. Sociobiol.* **37**(5), 321-328.

Wehner R. 1987 'Matched filters' – neural models of the external world. *J. Comp. Physiol. A* **161**,
511-531.

Yack J.E. 2004 The structure and function of auditory chordotonal organs in insects. *Microsc. Res.*
Techniq. **63**(6), 315-337.

Yin X., Müller R. 2019 Fast-moving bat ears create informative Doppler shifts. *Proc. Nat. Acad.*
Scie. USA **116**(25), 12270-12274.