Inhibition of protein synthesis in M1 of monkeys disrupts performance of

sequential movements guided by memory

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Abstract

The production of action sequences is a fundamental aspect of human motor skills. To

examine whether primary motor cortex (M1) is involved in maintenance of sequential

movements, we trained two monkeys (*Cebus apella*) to perform two sequential reaching tasks.

In one task, sequential movements were instructed by visual cues, whereas in the other task,

movements were generated from memory after extended practice. After the monkey became

proficient with performing the tasks, we injected an inhibitor of protein synthesis, anisomycin,

into M1 to disrupt information storage in this area. Injection of anisomycin in M1 had a marked

effect on the performance of sequential movements that were guided by memory. In contrast,

the anisomycin injection did not have a significant effect on the performance of movements

guided by vision. These results suggest that M1 of non-human primates is involved in the

maintenance of skilled sequential movements.

(144/150)

Introduction

The ability to perform a sequence of movements is a key component of motor skills, such as

typing and playing a musical instrument. How the brain binds elementary movements together

into meaningful actions has been a topic of much interest. The preparation for and generation

of sequential movements is classically thought to depend on the supplementary motor area

(SMA) and the pre-SMA (Roland et al., 1980; Tanji and Shima, 1994; Gerloff et al., 1997;

Shima and Tanji, 1998; Nakamura et al., 1998; Hikosaka et al., 2002; Picard and Strick, 2001;

Dayan and Cohen, 2011). According to this view, the primary motor cortex (M1) is thought to

produce the patterns of muscle activity that are necessary to implement the motor plans

generated by the premotor areas. There is growing evidence, however, that M1 of humans and

non-human primates is involved in the acquisition and retention of sequential movements. For

example, the activity of some M1 neurons appears to reflect the serial order of potential target

stimuli (Pellizzer et al., 1995; Carpenter et al., 1999). Similarly, a large number of M1 neurons

(~40%) and metabolic activity reflected aspects of learned movement sequences (Lu and Ashe,

2005; Matsuzaka et al., 2007; Picard et al., 2013). In addition, the functional connectivity of M1

neurons was suggested to be associated with sequential planning of movements (Hatsopoulos

et al., 2003). These observations and others imply a more active role for M1 in the acquisition

and retention of skilled movement sequences than previously thought (Karni et al., 1995; Dayan

and Cohen, 2011). In fact, the results of imaging studies in humans underscore the importance

of practice history and skill level for the activation of the motor areas (Krings et al, 2000; Meister

et al., 2005).

However, causal experiments to test M1's involvement in the acquisition or maintenance of

motor sequences have been challenging. M1 is critical for implementing motor output. Lesion

or inactivation of M1 will abolish the motor commands to the spinal cord that generate muscle

activity. Instead, we manipulated protein synthesis in M1 to selectively disrupt information

storage in this cortical area. This approach has been successful in the experimental dissection

and analysis of other memory systems in rodents (Davis and Squire, 1984; Nader et al., 2000;

Kleim et al., 2003). In these studies, local injection of an inhibitor for protein synthesis,

anisomycin, was shown to interfere with learning and maintenance of memory (Davis and

Squire 1984; Nader et al., 2000 a,b; Kandel, 2001; Dudai, 2004, 2012; Kelleher et al. 2004;

Rudy 2013; Kleim et al., 2003) .

Results

We trained two monkeys (*Cebus apella*) on two tasks. In one task, the monkeys performed

sequential movements guided by memory (Repeating task) (Fig. 1 a, b). As a control task, the

monkeys also were trained to perform reaching movements guided by visual cues (Random

task, for details see Methods). After the monkeys became proficient with performing the two

tasks (> 100 training sessions), we made injections of the protein synthesis inhibitor,

anisomycin, in M1. The injections of aniomycin solution (100 mg/ml) were placed at sites in

which intracortical stimulation evoked shoulder or elbow movements (Fig. 1c-f, for details see

Methods). We analyzed the monkey's behavior before and after the injection separately for

each movement in each task. A movement from one target to the next is defined as a trial.

The anisomycin injections had a significant effect on the performance of the movements during

the Repeating task (Figs. 2, 3a,b). The injections resulted in a significant increase in the

number of errors (Fig. 2, 3; χ^2 test, $p < 0.001$) and a significant decrease in the number of

predictive responses, an indication of sequence learning, (Fig. 3c; χ^2 test, $p < 0.001$) during the

Repeating task. In contrast, anisomycin injections did not have a significant effect on

performance of visually guided movements during the Random task (χ^2 test, $p > 0.05$).

The incorrect responses during the Repeating task can be categorized as two types: errors of

accuracy and errors in direction. An accuracy error is a reach performed in the correct direction

(e.g., to the right in move 1-5), but to an endpoint outside of the correct target (e.g., short of

target 5). As shown in Fig. 2a, before the injection, the monkey made correct contact to target

5 on 96% of the trials in the Repeating task (sequence 5-3-1). After the anisomycin injection,

the number of incorrect responses increased dramatically. The monkey then made accuracy

errors on 45% of the trials (Fig. 2a).

A direction error was considered to be a reach performed in the direction opposite to the correct

target (Fig. 2b, E_D). For example, movement 2-4 requires a rightward movement from target 2

to target 4. As shown in Fig. 2b, the monkey made a correct movement to target 4 on 93% of

the trials in the Repeating task (sequence 1-2-4) before the injection. After the anisomycin

injection, the animal moved his arm to the left from target 2 and made direction errors on 18%

of the trials in the Repeating task (Fig. 2b). This type of error suggests a deficit in selecting the

movement component in the sequence. For the injection session illustrated in Figs. 2 and 3,

the error rate increased significantly for five of the six movements performed during the

Repeating task.

Overall, we observed a significant increase in error rate for all of six injection sessions (χ^2 test,

$p < 0.05$; monkey N, n = 4; monkey R, n = 2). The effect of anisomycin was more pronounced

for certain moves of the learned sequences (Figs. 3, 4). The affected moves varied between

sessions. The error rate increased by an average of 42% for the most affected movements

during the Repeating task (Fig. 4b; paired t-test; $p = 0.002$).

As noted above, the anisomycin injections also caused a change in the number of predictive

movements during the Repeating task. We defined predictive responses as movements

completed before the presentation of the next target cue or in less than 150 ms after the

presentation of the next target cue (see Methods). Before the anisomycin injection in monkey

N, session No. 2, the animal made predictive responses on > 90% of the trials (Fig. 3c). After

the anisomycin injection, the number of predictive movements decreased dramatically. For

movements from target 5 to target 3 in sequence 5-3-1, the number of predictive responses

decreased by 55% of the total trials (Fig. 3c, χ^2 test, $p < 0.001$).

Overall, we observed a significant decrease in the number of predictive movements for all of six

injection sessions (χ^2 test, $p < 0.05$). The number of predictive responses decreased an

average of 56 % for the most affected movements during the Repeating task (Fig. 4c; paired t-

test, $p < 0.001$). A decrease in the number of predictive responses suggests an increase in the

time for movement selection.

As control experiments, we inactivated M1 by injecting muscimol, a GABA agonist, into the

shoulder representation of M1 in two sessions. We examined the performance of the animal on

the Random and Repeating tasks as we did for anisomycin injections into M1. We found that

M1 inactivations impaired performance on both the Random and Repeating tasks (χ^2 test, $p <$

0.05). The results indicate that M1 inactivation caused an indiscriminate deficit of motor

production. Injection of saline into M1 did not have any significant effects on performance of

both the Random and Repeating tasks (χ^2 test, $p < 0.05$).

Discussion

In the present study, localized inhibition of protein synthesis in M1 resulted in a selective deficit

in the performance of internally generated sequential movements during the Repeating task.

This observation emphasizes the importance of M1 for the generation of sequential movements

that are memory guided. Our results suggest that, although M1 is critical for movement

production, it also is involved in the maintenance of skilled sequential movements.

The acquisition and maintenance of long-term memory requires a series of cellular and

molecular changes that involve translation (i.e. protein synthesis) (Davis and Squire, 1984).

The role of protein synthesis in learning and memory has been intensively studied in rodents in

the context of fear conditioning (Davis and Squire 1984; Nader et al., 2003; Kandel, 2001;

Dudai, 2004, 2012; Kelleher et al. 2004; Rudy 2008; Ryan et al., 2015). Similarly, an injection

of a protein synthesis inhibitor, anisomycin, into rat M1 severely disrupted the maintenance

(Kleim et al., 2003) or the learning (Luft et al., 2004) of a skilled forelimb reaching task. The

injection of anisomycin also caused a reduction in synapse density and in the thickness of the

post synaptic density in M1 *in vivo* (Kleim et al., 2003). Studies using brain slices showed that

protein synthesis is required for long-lasting synaptic plasticity (late long-term potentiation)

(reviewed in Kelleher et al. 2004) and spine-head enlargement and growth during learning

(Steward and Levy, 1982; Steward and Fass, 1983; Steward and Schuman, 2003; Tanaka et

al., 2008). Several observations indicate that the effects of protein synthesis inhibitors on long-

lasting synaptic plasticity are likely to be a specific consequence of their translational blockade

(Linden, 1996; Huber et al. 2000; Beaumont et al. 2001; Kelleher III et al., 2004; Sadowski et

al., 2011). These studies in rodents support our finding that inhibition of protein synthesis in M1

of monkeys interfered with performance of sequential movements guided by memory.

M1 is densely interconnected with the dorsal premotor cortex proper (PMd) (Dum and Strick,

2005). We recently demonstrated that inactivation of the PMd resulted in a selective deficit in

the performance of internally generated sequences (Ohbayashi et al., 2016). We proposed that

the PMd contributes to the internal generation of sequential movements through maintaining

motor-motor associations. Our current results are consistent with the PMd functioning as a

major source of input to M1 to guide the performance of internally generated sequences.

Previous studies reported that extended practice on a sequence of movements resulted in

dramatic alterations in the functional activation and neural responses of M1 (Matsuzaka et al.,

2007; Picard et al., 2013). In addition, human imaging studies found a functional and structural

change in M1 of professional musicians (Elbert et al., 1995; Amunts, K et at., 1997; Gaser and

Dchlaug, 2003; Schwenkreis et al., 2007; Draganski and May, 2008; Herholz and Zattore, 2012;

Zatorre et al., 2012; Sampaio-Baptista and Johansen-Berg, 2017; Wenger et al., 2017). These

studies, along with our current results, suggest that M1 is involved in the maintenance of

sequential movements after extensive practice. M1's specific contribution to the acquisition and

retention of sequential movements needs to be further explored in future experiments.

Materials and Methods

The care of the monkeys and the experimental protocols adhered to the National Institutes of

Health *Guide for the Care and Use of Laboratory Animals* and the U.S. Public Health Service

Policy on Humane Care and Use of Laboratory Animals. All procedures used followed

institutional guidelines and were approved by the Institutional Animal Care and Use Committee.

Behavioral task. Two monkeys (*Cebus apella*, two males weighing 3.2 kg and 3.7 kg) were

trained to perform two tasks that required sequential reaching movements to targets on a touch

sensitive monitor with their right arm (Fig. 1a, b) (Matsuzaka et al., 2007; Picard et al., 2013;

Ohbayashi et al., 2016). In the Random task, the reaching movements were guided by visual

targets displayed on a touch screen monitor. Each new target was presented according to a

pseudorandom sequence. Contact of the correct target triggered the display of the next target

after 100 ms delay. After the monkeys became proficient in the performance of the Random

task (~ 50 days of practice), they were introduced to the Repeating task. In the Repeating task,

new targets were presented according to a three-element repeating sequence. We trained

animals on repeating sequences: "5-3-1-5-3-1 . . ." and "1-2-4-1-2-4 . . ." or "2-3-4-2-3-4 ..."

(Fig. 1b, two sequences for monkey N, one sequence for monkey R). New targets during the

Repeating task were presented 400 ms after contact of the preceding target. This delay

promoted the performance of predictive responses during the Repeating task in which the

animal moved to the next target in a sequence before the target was presented. Each task was

performed continuously in blocks of 200–500 trials. A liquid reward was given after every four to

five correct responses. Both monkeys became proficient with performing the two tasks after >

50 training sessions. The injection experiments were performed after a monkey had more than

100 training sessions on each sequence.

Surgery. We implanted a head restraint device, along with an MR compatible chamber for

micro-injection, on an animal's skull using small screws and dental acrylic. All surgical

procedures were performed under general anesthesia (1-3% isofluorane) using aseptic

techniques. The animal received antibiotics and analgesics after surgical procedures. The

chamber's placement over M1 was verified using structural MR images taken prior to and after

the surgery (Fig 1d). When task performance returned to the pre-surgical level, we performed

a craniotomy to expose the cortex in the chamber.

Intracortical microstimulation. We used intracortical microstimulation to identify the arm

representation in M1 and to physiologically define the border between M1 and the PMd (Dum

and Strick, 2005 and Ohbayashi et al., 2016). We used glass-coated Elgiloy microelectrodes

(0.6–1.5 MΩ at 1 kHz) to deliver intracortical microstimuli. A constant-current stimulator was

used to deliver cathodal pulses (1- 40 μA intensity, 10-20 cathodal pulses, 333 Hz, 0.2 ms

duration) at a depth of 1500 μm below the cortical surface (Dum and Strick, 2005; Ohbayashi et

al, 2016). Stimulus intensity was measured with a current monitor (Ion Physics). The motor

response evoked by stimulation was determined by visual observation and muscle palpation.

The response threshold was defined as the lowest stimulus intensity necessary to evoke a

response on ~80% of the trials.

Injections of pharmacological agents. We injected anisomycin (an inhibitor for protein

synthesis), muscimol solution or sterile saline at 1.5 mm below the cortical surface using a 30

gauge cannula connected to a 10 μl Hamilton syringe (0.2 μl every 30–60 s). We prepared

solutions of anisomycin (100 μg/μl in ACSF, pH 7.2-7.4) and muscimol (5 mg/ml in saline) from

commercially available powders (Sigma-Aldrich, MO). The cortical sites of anisomycin injection

in monkey N are displayed in Figure 1e (injection session No. 2). For each injection session,

we injected total of 5 or 10 μl anisomycin solution into M1 (5μl, n=5; 10μl, n=1). To cover a

large portion of the arm representation in M1, the anisomycin solution was injected at two to

four sites in the arm representation area of M1 (Fig. 1f, 1-3 μl at each site). The cannula was

left in place for ~5 min to allow diffusion of the solution and prevent its reflux and then removed.

The effect on performance of the tasks was tested 20-24 hours after the injection of an inhibitor

of protein synthesis. We did experiments with anisomycin injections 4 times in monkey N and 2

times in monkey R. Injections were spaced at least 7 days apart. In separate experiments, we

also injected 1-3 μl of muscimol to inactivate M1 function to compare the results with

anisomycin injections. The effect of muscimol on task performance was tested 20 min after the

injection to allow for some diffusion of the chemical in the brain tissue before resuming the task.

Saline (5 µl) was injected as a control in separate experiments. During a post-injection test

session, blocks of Random and Repeating trials were alternated at frequent intervals to sample

the animal's performance evenly as the effect of the test substance emerged and intensified.

Measurement of performance on days prior to an injection and/or on trial blocks preceding an

injection was used as the baseline for comparison with post-injection performance.

Analysis of performance.   For every trial of each task, we recorded various task parameters

and measures of performance. The effect of an injection was assessed by examining the

following: changes in the percentage of correct responses, types of incorrect responses,

changes in Movement Time (MT) and Response Time (RT), percentage of predictive responses

(defined as RT < 150ms) and contact points on the touch screen (Ohbayashi et al., 2016). We

defined MT as the interval between the release of contact from one target to touch of the next

target. We defined RT during the Random task as the time between the presentation of a new

target and contact of that target minus 100 ms delay time. We defined RT during the

Repeating task as the time between contact of two targets in a sequence minus 400 ms delay

time. A negative RT indicates that the animal is internally generating the response before the

visual cue is presented. After ~50 days of training on the first Repeating sequence, the monkey

made predictive responses (RT < 150 ms) without using visual cues in more than 80% of the

trials. When RT was less than 150 ms the trial was considered to be a predictive response.

We excluded the following trials from analysis: 1) trials after errors because in this case the trial

was repeated; 2) trials during the Random task with an RT less than 150 ms because the

monkey may not have been paying attention to the task, tried to guess the next target or have

attempted to perform a move of a Repeating sequence. For each movement, we used χ^2 tests

with Holm–Bonferroni's correction to examine the significance of changes in error rate and

predictive responses. We used t-tests with Holm–Bonferroni's correction to examine changes

of MT and RT. The effect of anisomycin was more pronounced for certain moves of the

learned sequences. For each injection session, a movement of the Repeating task that showed

the largest and weakest change of error rate were identified. The data of these movements

during the Repeating task and the data of corresponding movements during the Random task

were used for population analysis, average and SE of error rates and percentage of predictive

responses, in Figure 4.

Author contributions: M.O. designed research, performed research, analyzed data and wrote

the paper.

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Competing interests

The authors declare no competing financial interests.

References

Amunts, K., G. Schlaug, L. Jancke, H. Steinmetz, A. Schleicher, A. Dabringhaus and K. Zilles (1997). "Motor cortex and hand motor skills: structural compliance in the human brain." Hum Brain Mapp 5(3): 206-215.

Beaumont, V., N. Zhong, R. Fletcher, R. C. Froemke and R. S. Zucker (2001). "Phosphorylation and local presynaptic protein synthesis in calcium- and calcineurin-dependent induction of crayfish long-term facilitation." Neuron 32(3): 489-501.

Carpenter, A. F., A. P. Georgopoulos and G. Pellizzer (1999). "Motor cortical encoding of serial order in a context-recall task." Science 283(5408): 1752-1757.

Davis, H. P. and L. R. Squire (1984). "Protein synthesis and memory: a review." Psychol Bull 96(3): 518-559.

Dayan, E. and L. G. Cohen (2011). "Neuroplasticity subserving motor skill learning." Neuron 72(3): 443-454.

Draganski, B. and A. May (2008). "Training-induced structural changes in the adult human brain." Behav Brain Res 192(1): 137-142.

Dudai, Y. (2004). "The neurobiology of consolidations, or, how stable is the engram?" Annu Rev Psychol 55: 51-86.

Dudai, Y. (2012). "The restless engram: consolidations never end." Annu Rev Neurosci 35: 227-247.

Dum, R. P. and P. L. Strick (2005). "Frontal lobe inputs to the digit representations of the motor areas on the lateral surface of the hemisphere." J Neurosci 25(6): 1375-1386.

Elbert, T., C. Pantev, C. Wienbruch, B. Rockstroh and E. Taub (1995). "Increased cortical representation of the fingers of the left hand in string players." Science 270(5234): 305-307.

Gaser, C. and G. Schlaug (2003). "Gray matter differences between musicians and nonmusicians." Ann N Y Acad Sci 999: 514-517.

Gerloff, C., B. Corwell, R. Chen, M. Hallett and L. G. Cohen (1997). "Stimulation over the human supplementary motor area interferes with the organization of future elements in complex motor sequences." Brain 120 (Pt 9): 1587-1602.

Hatsopoulos, N. G., L. Paninski and J. P. Donoghue (2003). "Sequential movement representations based on correlated neuronal activity." Exp Brain Res 149(4): 478-486.

Herholz, S. C. and R. J. Zatorre (2012). "Musical training as a framework for brain plasticity: behavior, function, and structure." Neuron 76(3): 486-502.

Hikosaka, O., K. Nakamura, K. Sakai and H. Nakahara (2002). "Central mechanisms of motor

skill learning." Curr Opin Neurobiol 12(2): 217-222.

Huber, K. M., M. S. Kayser and M. F. Bear (2000). "Role for rapid dendritic protein synthesis in hippocampal mGluR-dependent long-term depression." Science 288(5469): 1254-1257.

Kandel, E. R. (2001). "The molecular biology of memory storage: a dialogue between genes and synapses." Science 294(5544): 1030-1038.

Karni, A., G. Meyer, P. Jezzard, M. M. Adams, R. Turner and L. G. Ungerleider (1995). "Functional MRI evidence for adult motor cortex plasticity during motor skill learning." Nature 377(6545): 155-158.

Kelleher, R. J., 3rd, A. Govindarajan and S. Tonegawa (2004). "Translational regulatory mechanisms in persistent forms of synaptic plasticity." Neuron 44(1): 59-73.

Kleim, J. A., R. Bruneau, K. Calder, D. Pocock, P. M. VandenBerg, E. MacDonald, M. H. Monfils, R. J. Sutherland and K. Nader (2003). "Functional organization of adult motor cortex is dependent upon continued protein synthesis." Neuron 40(1): 167-176.

Krings, T., R. Topper, H. Foltys, S. Erberich, R. Sparing, K. Willmes and A. Thron (2000). "Cortical activation patterns during complex motor tasks in piano players and control subjects. A functional magnetic resonance imaging study." Neurosci Lett 278(3): 189-193.

Linden, D. J. (1996). "A protein synthesis-dependent late phase of cerebellar long-term depression." Neuron 17(3): 483-490.

Lu, X. and J. Ashe (2005). "Anticipatory activity in primary motor cortex codes memorized movement sequences." Neuron 45(6): 967-973.

Luft, A. R., M. M. Buitrago, T. Ringer, J. Dichgans and J. B. Schulz (2004). "Motor skill learning depends on protein synthesis in motor cortex after training." J Neurosci 24(29): 6515-6520.

Matsuzaka, Y., N. Picard and P. L. Strick (2007). "Skill representation in the primary motor cortex after long-term practice." J Neurophysiol 97(2): 1819-1832.

Meister, I., T. Krings, H. Foltys, B. Boroojerdi, M. Muller, R. Topper and A. Thron (2005). "Effects of long-term practice and task complexity in musicians and nonmusicians performing simple and complex motor tasks: implications for cortical motor organization." Hum Brain Mapp 25(3): 345-352.

Nader, K. (2003). "Memory traces unbound." Trends Neurosci 26(2): 65-72.

Nader, K., G. E. Schafe and J. E. Le Doux (2000a). "Fear memories require protein synthesis in the amygdala for reconsolidation after retrieval." Nature 406(6797): 722-726.

Nader, K., G. E. Schafe and J. E. LeDoux (2000b). "The labile nature of consolidation theory." Nat Rev Neurosci 1(3): 216-219.

Nakamura, K., K. Sakai and O. Hikosaka (1998). "Neuronal activity in medial frontal cortex during learning of sequential procedures." J Neurophysiol 80(5): 2671-2687.

Nakamura, K., K. Sakai and O. Hikosaka (1999). "Effects of local inactivation of monkey medial frontal cortex in learning of sequential procedures." J Neurophysiol 82(2): 1063-1068.

Ohbayashi, M., N. Picard and P. L. Strick (2016). "Inactivation of the Dorsal Premotor Area Disrupts Internally Generated, But Not Visually Guided, Sequential Movements." J Neurosci 36(6): 1971-1976.

Pellizzer, G., P. Sargent and A. P. Georgopoulos (1995). "Motor cortical activity in a context-recall task." Science 269(5224): 702-705.

Picard, N., Y. Matsuzaka and P. L. Strick (2013). "Extended practice of a motor skill is associated with reduced metabolic activity in M1." Nat Neurosci 16(9): 1340-1347.

Picard, N. and P. L. Strick (2001). "Imaging the premotor areas." Curr Opin Neurobiol 11(6): 663-672.

Roland, P. E., B. Larsen, N. A. Lassen and E. Skinhoj (1980). "Supplementary motor area and other cortical areas in organization of voluntary movements in man." J Neurophysiol 43(1): 118-136.

Rudy, J. W. (2008). "Destroying memories to strengthen them." Nat Neurosci 11(11): 1241-1242.

Rudy, J. W. (2013). The neurobiology of learning and memory. Sinauer Associates, Inc.; 2 edition

Ryan, T. J., D. S. Roy, M. Pignatelli, A. Arons and S. Tonegawa (2015). "Memory. Engram cells retain memory under retrograde amnesia." Science 348(6238): 1007-1013.

Sampaio-Baptista, C. and H. Johansen-Berg (2017). "White Matter Plasticity in the Adult Brain." Neuron 96(6): 1239-1251.

Schwenkreis, P., S. El Tom, P. Ragert, B. Pleger, M. Tegenthoff and H. R. Dinse (2007). "Assessment of sensorimotor cortical representation asymmetries and motor skills in violin players." Eur J Neurosci 26(11): 3291-3302.

Shima, K. and J. Tanji (1998). "Both supplementary and presupplementary motor areas are crucial for the temporal organization of multiple movements." J Neurophysiol 80(6): 3247-3260.

Steward, O. and B. Fass (1983). "Polyribosomes associated with dendritic spines in the denervated dentate gyrus: evidence for local regulation of protein synthesis during reinnervation." Prog Brain Res 58: 131-136.

Steward, O. and W. B. Levy (1982). "Preferential localization of polyribosomes under the base of dendritic spines in granule cells of the dentate gyrus." J Neurosci 2(3): 284-291.

Steward, O. and E. M. Schuman (2003). "Compartmentalized synthesis and degradation of proteins in neurons." Neuron 40(2): 347-359.

Tanaka, J., Y. Horiike, M. Matsuzaki, T. Miyazaki, G. C. Ellis-Davies and H. Kasai (2008). "Protein synthesis and neurotrophin-dependent structural plasticity of single dendritic spines." Science 319(5870): 1683-1687.

Tanji, J. and K. Shima (1994). "Role for supplementary motor area cells in planning several movements ahead." Nature 371(6496): 413-416.

Wenger, E., C. Brozzoli, U. Lindenberger and M. Lovden (2017). "Expansion and Renormalization of Human Brain Structure During Skill Acquisition." Trends Cogn Sci 21(12): 930-939.

Zatorre, R. J., R. D. Fields and H. Johansen-Berg (2012). "Plasticity in gray and white: neuroimaging changes in brain structure during learning." Nat Neurosci 15(4): 528-536.

Figure 1: Task and cortical maps for monkey N.

a, b. Sequences in the Repeating task. c. Lateral view of cebus brain. Dashed lines indicate
the M1-PMd border and the pre-PMd-PMd border. ArS: arcuate sulcus; CS: central sulcus;
IPS: intra parietal sulcus; LS: lateral sulcus; pre-PMd: pre-dorsal premotor cortex; PMd: dorsal
premotor cortex; M1: primary motor cortex. R: rostral; L: lateral. d. MRI image after the
chamber implantation for monkey N. The dotted circle indicates the chamber outline. e.
Intracortical stimulation map from monkey N. Letter indicate the movements evoked at each
site. S:Shoulder; E: Elbow; W: Wrist; D: Digit. f. Anisomycin solution was injected in the area
indicated by a gray oval.

Figure 2: Reaching end points before and after anisomycin injection.
Left: pre-injection; right: post-injection. E_A: Accuracy errors; E_D: Direction errors; gray dots:
correct response; black dots: error response. The anisomycin injection was placed at sites
shown in Fig. 1f in monkey N. **a**, Reaching end points for movement from target 1 to target 5.
The monkey was performing sequence 5-3-1 during the Repeating task. **b**, Reaching end
points for movement from target 2 to target 4. The monkey was performing sequence 1-2-4
during the Repeating task. Percentage of trials ending in each target are given below the
targets. Touches between targets were counted as touches to the closest target. *$p < 0.05$.

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Figure 3: Effect of anisomycin injection on performance of the task.

a– c, The performance data for an injection session #2 in monkey N (Fig. 1f, Fig. 2). a, Error rate in the Random task. Anisomycin injection did not have any effects on number of errors during the Random task (χ^2 test, p = 0.219 for move 3-1, p = 0.507 for move 5-3, p = 0.467 for move 1-5, df = 1; not significant for move 4-1; p = 0.939 for move 1-2; p = 0.206 for move 2-4). b, Error rate in the Repeating task. After the anisomycin injection, the number of errors increased dramatically in 5 of 6 movements in the Repeating task (χ^2 test, p < 0.001 for moves 5-3, 1-5, 4-1,1-2 and 2-4, p=0.572 for move 3-1, df = 1). c, Predictive responses. The percentage of predictive responses decreased significantly after the anisomycin injection (χ^2 test, p < 0.001 for all movements, df = 1).

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Figure 4: Population data for anisomycin injections.
a, Average error rate in the Random task. Anisomycin injections did not have an effect on the
number of error responses in the Random task (paired t-test, 6 injection experiments, $p = 0.796$
for strongest effect; $p = 0.590$ for weakest effect; df = 5). b, Average error rate in the
Repeating task. The error rate in the Repeating task increased significantly after the
anisomycin injections (paired t-test, 6 injection experiments, $p = 0.002$ for strongest effect; $p =$
0.497 for weakest effect; df = 5). c, Average predictive responses in the Repeating task. The
percentage of predictive responses decreased after anisomycin injections in M1 (paired t-test, p
< 0.001 for strongest effect; $p = 0.419$ for weakest effect; df = 5). *$p < 0.05$.