Species richness increases fitness differences,

but does not affect niche differences

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Abstract

A key question in ecology is what limits species richness. Modern coexistence theory presents the persistence of species as a balance between niche differences and fitness differences that favor and hamper coexistence, respectively. With most applications focusing on species pairs, however, we know little about if and how this balance changes with species richness. Here, we present the first mathematical proof that the average fitness difference among species increases with richness, while the average niche difference stays constant. Extensive simulations with more complex models and analyses of empirical data confirmed these mathematical results. Taken together, our work suggests that, as species accumulate in ecosystems, ever-increasing fitness differences will at some point exceed constant niche differences, limiting species richness. Our results contribute to the expansion of modern coexistence theory towards multi-species communities.

Introduction

Explaining nature's biodiversity is a key challenge for science (Hutchinson, 1957). One type of approach consists of focusing on the capacity of individual species to persist through time despite occasional pruning to low density or species interactions (Turelli, 1978). Modern coexistence theory is such an approach, and predicts species persistence in a community when niche differences overcome fitness differences. Niche differences measure the strength of negative frequency dependence, i.e. whether a focal species i can recover when reduced to low abundance (Adler *et al.*, 2007; Chesson, 2000; Spaak & De Laender, 2020). Fitness differences measure the competitive ability of that species (Adler *et al.*, 2007; Hart *et al.*, 2018; Spaak & De Laender, 2020).

However, few applications of coexistence theory have explicitly focused on explaining biodiversity, i.e. the persistence of species in species-rich communities. Instead, most applications have considered two-species communities (Chesson, 2000; Letten *et al.*, 2017), using a variety of approaches and case studies, including annual and perennial plants (Adler *et al.*, 2018; Godoy & Levine, 2014), phytoplankton (Gallego *et al.*, 2019; Narwani *et al.*, 2013) and bacteria (Zhao *et al.*, 2016), and under different environmental conditions (Bimler *et al.*, 2018; Cardinaux *et al.*, 2018; Grainger *et al.*, 2019; Lanuza *et al.*, 2018; Matías *et al.*, 2018; Napier *et al.*, 2016; Wainwright *et al.*, 2019). To our knowledge only three empirical studies that report niche and fitness differences in communities composed of more than two species (hereafter multi-species communities) (Chu & Adler, 2015; Petry *et al.*, 2018; Veresoglou *et al.*, 2018). However, none of these three studies explain how niche and fitness differences change with species richness. In order to understand how niche and fitness differences co-determine species persistence in multi-species communities, we need to understand how both variables change when adding species to a community.

Multi-species communities possess at least four complexities that are absent from two-species communities, which may affect niche and fitness differences, and therefore how we interpret coexistence in multispecies communities. (1) First, a multi-species community can host more interaction types than species pairs, e.g. competitive, mutualistic or trophic interactions. Species richness increases the number of possible interactions and the number of combinations of these interaction types. Several metrics exist to summarize

this diversity of interaction types and study their implications for community dynamics (Fontaine *et al.*, 2011). (2) Second, two-species communities are always fully connected and correlations between interspecific interactions (Barabás *et al.*, 2016b) become irrelevant since there is only a single pair of interspecific interactions. In contrast, in an *n*-species community there may be anywhere from $n-1$ to $\frac{n}{2}(n-1)$ connections, and the interspecific effects of species j on species i can be positively or negatively correlated with the interspecific effects i has on j (Barabás *et al.*, 2016b). May (1972) and Allesina & Tang (2012, 2015) and have shown that connectance and correlation can have large effects on the local stability of multi-species communities. We therefore expect these factors to influence coexistence as well. (3) Third, higher-order interactions, through which a third species changes the interaction between a species pair, are by definition absent from two-species communities (but see Letten & Stouffer (2019); Levine *et al.* (2017)). Such higher-order interactions have been found empirically, for example, in communities composed of phytoplankton, bacteria, and ciliates (Mickalide & Kuehn, 2019). In that study, bacteria coexisted with phytoplankton and ciliates, but all three functional groups did not coexist. In the three-species community, phytoplankton inhibited bacterial aggregation, leaving the latter more vulnerable to predation by ciliates. (4) Fourth, indirect effects, whereby a third species changes the dynamics of a species pair by directly interacting with both partners at the same time, are by definition absent from two-species communities (Walsh, 2013). We will refer to these four complexities throughout the text with (1) interaction types, (2) interaction matrix structure, (3) higher-order interactions and (4) indirect interactions.

Studying multi-species coexistence is challenging both theoretically and experimentally. Theoretically speaking, the methods to analyse coexistence via niche and fitness differences in a multi-species community were not available until recently (Carmel *et al.*, 2017; Carroll *et al.*, 2011; Saavedra *et al.*, 2017; Spaak & De Laender, 2020). Experimentally speaking, studying coexistence of multiple species is resource-demanding. For instance, in the simple case of linear direct interactions among species (i.e. as in Lotka-Volterra models) the number of experiments needed to parametrize the community is quadratic in species richness (but see Maynard *et al.* (2019)). Considering higher-order interactions will consequently result in an even higher experimental load. For example, measuring higher-order interactions, sensu Letten & Stouffer (2019), would require 39 experiments in a three species community.

In this paper we investigate the balance between niche and fitness differences along a gradient of species richness. More specifically, we ask how niche and fitness differences change as the number of species increases in randomly assembled communities, and how the additional complexities (1)-(4) influence these changes. We do so using four independent methods that rely on a novel definition for niche and fitness differences that is able to analyse multi-species coexistence through niche and fitness differences (Spaak & De Laender, 2020), and a new compilation of species interaction data. First, we derive equations that quantify how niche and fitness differences respond to species richness in a community with linear interactions and simple cases of higher-order interactions. Second, we give an intuitive explanation of these responses based on the Mac-Arthur consumer-resource model. Third, we perform simulations with more complex models. We run these simulations as a full-factorial virtual experiment, varying direct interactions (type, correlation, connectance), higher-order interactions and indirect interactions. Fourth, we compile data from the literature on empirically measured species interaction matrices and compute niche and fitness differences as a response to species richness. We then compare our results obtained via random species assembly to a community with non-random assembly. All methods support the same general conclusion: species richness does not affect niche differences, but increases fitness differences. Importantly, these conclusions are independent of the four complexities (1)-(4).

Methods

Model assumptions

To include the various additional complexities of multispecies communities we use a generalized Lotka-Voltera model with n species containing second-order interactions to model the per-capita growth rates $f_i(N_i, \mathbf{N}^{-i})$:

$$\frac{1}{N_i}\frac{dN_i}{dt} = \underbrace{r_i\left(1 + \sum_j \alpha_{ij}N_j\left(1 + \sum_k \beta_{ijk}N_k\right)\right)}_{f_i(N_i, \mathbf{N}^{-i})}, \tag{1}$$

where N_i is the density of the focal species i, $\mathbf{N}^{-i}$ is the density of the resident community (vector of length $n-1$), r_i is its intrinsic growth rate, and α_{ij} and β_{ijk} are first (or linear) and second-order species interactions, respectively. A positive α_{ij} indicates a positive interaction between species i and j (facilitation). Negative α_{ij} on the other hand indicate negative interactions (competition). When β_{ijk} is positive or negative species k will intensify or weaken the relationship between species i and j, respectively (second-order interaction). The inclusion of third order interactions did not affect any of our results. Throughout the manuscript we assume $\alpha_{ii} = -1$ with no loss of generality as it can be achieved by rescaling of empirical data. Furthermore, for most of the manuscript we assume a random community assembly. That is α_{ij} and α_{ik} are sampled from any independent distributions for $k \neq j$ and similarly for β_{ijk}.

Niche and fitness differences

There exist a total of eleven definitions for niche and fitness differences with various assumptions about the community model (Spaak & De Laender, 2020). Five of these definitions can be applied to multi-species communities (Carmel *et al.*, 2017; Carroll *et al.*, 2011; Chesson, 2003; Saavedra *et al.*, 2017; Spaak & De Laender, 2020). Optimally, we would compute niche and fitness differences according to all five methods to understand how species richness affects coexistence.

However, this is not possible, as four of the five community models do not readily apply to the communities investigated here. The definitions of Carmel *et al.* (2017) assume that the species have a minimal growth rate, the mortality rate, it therefore best applies to resource competition models, but it does not apply to Lotka-Volterra models. Chesson (2003) was developed for environmental or spatial fluctuations, which we do not consider here. Furthermore, Chesson (2003) works best for communities driven by only one limiting factor (Barabás *et al.*, 2018), the community models investigated here, however, have as many limiting factors as they have species. For such communities the scaling factors are not uniquely defined and the results will therefore depend on the non-unique choice of the scaling factors (Barabás *et al.*, 2018; Song *et al.*, 2019). Saavedra *et al.* (2017) can only be applied to communities without higher order interactions. Cenci & Saavedra (2018) propose and extension to include non-linear per-capita growth rates, however, the interaction matrix must still be linear, which is not the case for the communities investigated here. Carroll

et al. (2011) can only be applied to communities without strong inter-specific facilitation, that is the intrinsic growth rate must be higher than the invasion growth rate. This excludes about two-thirds of our theoretical and empirical data. One could ask how species richness affects niche and fitness differences according to these last two definitions if the community model were restricted to competitive Lotka-Volterra communities, yet, there is an additional problem. Niche and fitness differences as computed by these two definitions allow inference of coexistence only in two-species communities, not in multi-species communities (Spaak & De Laender, 2020). That is, two different multi-species communities may have identical niche and fitness differences but different outcomes of coexistence (e.g. all species persist in one community, but not in the other). Since we ask how niche and fitness differences jointly determine coexistence in multi-species communities, these methods are not suited to gain new insight. Consequently, of the eleven available definitions of niche and fitness differences only the definition of Spaak & De Laender (2020) applies to the community models and the research question investigated here.

Spaak & De Laender (2020) base their definition of niche and fitness differences ($\mathcal{N}_i$ and $\mathcal{F}_i$) on the comparison of species growth rates in various conditions. If the two species (i and j) of a two-species community have completely separated niches ($\mathcal{N}_i = \mathcal{N}_j = 1$), the species i will grow in presence of j as it would in its absence, which implies $f_i(0, N_j^*) = f_i(0, 0)$ where $f_i(0, N_j^*)$ is the per capita growth rate of species i when it invades a community with j at equilibrium density N_j^*, known as the invasion growth rate (Carroll *et al.*, 2011; Chesson, 1994; Ellner *et al.*, 2019) and $f_i(0, 0)$ is the intrinsic growth rate of species i. Conversely, if the two species have exactly the same niche ($\mathcal{N}_i = \mathcal{N}_j = 0$) they have equivalent effects on each other. It then holds that $f_i(0, N_j^*) = f_i(c_{ij}N_j^*, 0)$ where c_{ij} is the conversion factor allowing to express individuals of species j as individuals of species i.

Unfortunately, the c_{ij} are defined implicitly as the solution of an equation, which often does not have a closed form solution, but must be computed numerically. However, intuitively the c_{ij} are simple. The conversion factors c_{ij} change the frequency of species i and j, but they do not affect the total density. That is, two communities with densities $(0, N_j^*)$ and $(c_{ij}N_j^*, 0)$ have the same total density, but different frequencies of species i. In the first community species i has 0 frequency, while in the second it has 100% frequency, as $c_{ij}N_j^*$ is a density of species i. We refer to Spaak & De Laender (2020) and the appendix S1

138 for a mathematical definition of c_{ij}.

Further intuitive insight can be gained from a Lotka-Volterra model in which the c_{ij} can be computed explicitly as $c_{ij} = \sqrt{\left|\frac{\alpha_{ij}\alpha_{jj}}{\alpha_{ji}\alpha_{ii}}\right|}$ (Appendix S2). Note that the arrangement of the interaction coefficients α_{ij}

141 differs from both, the niche and the fitness differences. The numerator $(\alpha_{ij}\alpha_{jj})$ is the competitive effect of species j, while the denominator $(\alpha_{ji}\alpha_{ii})$ is the competitive effect os species i. The c_{ij} therefore keep total density constant by scaling the effects of the two species.

144 Niche differences are then defined via interpolation between these two extreme cases:

$$\mathcal{N}_i = \frac{f_i(0, N_j^*) - f_i(c_{ij}N_j^*, 0)}{f_i(0, 0) - f_i(c_{ij}N_j^*, 0)} \tag{2}$$

147 This equation of niche differences can be interpreted as the relative frequency dependence of species i. The numerator assesses frequency dependence, as it compares the growth rate of species i when it has 0% frequency to when it has 100% frequency, notably at the same converted density. The denominator assesses

150 density dependence, as it compares the growth rate of species i at 0 density to when it has non-zero density. Importantly, in both growth rates species i has 100% frequency. This definition maps positive frequency dependence, negative frequency dependence and facilitation to $\mathcal{N}_i < 0, 0 < \mathcal{N}_i < 1$, and $1 < \mathcal{N}_i$ respectively.

153 Similarly, they define fitness differences as the scaled growth rate in the absence of niche differences

$$\mathcal{F}_i = \frac{f_i(c_{ij}N_j^*, 0)}{f_i(0, 0)} \tag{3}$$

156 Zero fitness differences imply that the species have equal competitive strength, as $N_i^* = c_{ij}N_j^*$ that is species will have the same equilibrium density after conversion with c_{ij}. Competitive subordinate species then have negative fitness differences, as $N_i^* < c_{ij}N_j^*$, conversely competitive dominant species have positive fitness

159 differences. Note that $\mathcal{F}_i = 0$ means no fitness differences and more negative $\mathcal{F}_i$ means stronger fitness differences. Importantly, these definitions of niche and fitness differences agree with the original definitions on the two species Lotka-Volterra models (Chesson, 2013; Chesson & Kuang, 2008), they are therefore not

162 newly defined niche and fitness differences, but rather a generalization thereof (Appendix S1, Spaak & De Laender (2020)).

Additionally, this definition very naturally generalizes to multi-species communities. The three growth

165 rates remain conceptually the same. The intrinsic growth rate $f_i(0, \mathbf{0})$, where $\mathbf{0}$ denotes the absence of all

other species. The invasion growth rate $f_i(0, \mathbf{N}^{(-i,*)})$, where $\mathbf{N}^{(-i,*)}$ denotes the equilibrium density of the community in absence of species i. And the no-niche growth rate $f_i(\sum_j c_{ij} N_j^{(-i,*)})$, where all densities of the resident species have been converted to densities of the focal species. Doing so makes clear that $\mathscr{N}_i$ and $\mathscr{F}_i$ are species-specific properties, i.e. in general we have $\mathscr{N}_i \neq \mathscr{N}_j$ and $\mathscr{F}_i \neq \mathscr{F}_j$ in multi-species communities. However, $\mathscr{N}_i$ and $\mathscr{F}_i$ compare the effect of the focal species on itself to the effect of the rest of the community on the focal species. They therefore do depend on the traits of the other species in the community.

Analyses and Simulations

We first examined analytically how $\mathscr{N}_i$ and $\mathscr{F}_i$ change with species richness. We found a generic solution for first-order interactions and for a simplified case of higher-order interactions.

Second, we designed a full-factorial virtual experiment in which we numerically computed $\mathscr{N}_i$ and $\mathscr{F}_i$ for a wider range of different simulated communities (see table 1). For these we solve numerically for equilibrium densities and invasion growth rates using the 'fsolve' function from the scipy package in Python. Communities with higher order interactions can have multiple equilibria (Aladwani & Saavedra, 2019). To compare communities with and without higher order interactions we computed $\mathscr{N}_i$ and $\mathscr{F}_i$ only for one equilibrium in communities with higher order interactions. Specifically, we always chose the equilibrium that is closest to the equilibrium of the community without higher order interactions. The factors were (i) first-order interaction type (competitive, facilitative or both, i.e. $\alpha_{ij} < 0, > 0$ or unrestricted); (ii) first-order interaction strength (strong or weak); (iii) connectance of the interspecific interaction ($c \in \left\{ 1, \frac{4}{5}, \frac{2}{3} \right\}$); (iv) correlation between the interspecific interaction ($\rho(\alpha_{ij}, \alpha_{ji}) = \rho_{ij}(\beta_{ijk}, \beta_{jik}) \in \{-1, 0, 1\}$); (v) inclusion of indirect effects (present or absent); (vi) second-order interaction type ($\beta_{ijk} < 0, > 0$, unrestricted, or absent); To exclude indirect effects we set equilibrium densities of resident species to their monoculture equilibrium density. In this way, we cancel out interactions among residents that will change the residents' densities. The intrinsic growth rate r_i does not affect $\mathscr{N}_i$ and $\mathscr{F}_i$, therefore we set it to 1 in all simulations.

This design leads to a total of $3 \cdot 2 \cdot 3 \cdot 3 \cdot 2 \cdot 4 = 432$ parameter settings. We ran 1000 repetitions for each of the five species richness levels ($2 \le n \le 6$), leading to a total of $432 \cdot 5 \cdot 1000 = 2'160'000$ simu-

Factor	Parameter	Levels	Interpretation	Complexity investigated
Interaction type first-order	α_{ij}	< 0 > 0 no restriction	competition facilitation mixed	(1)
Strength of interaction first-order	α_{ij}	strong weak		(1)
Connectance	$P(\alpha_{ij} \neq 0)$	$1, \frac{4}{5}, \frac{2}{3}$		(2)
Interaction correlation	$\mathrm{cor}(\alpha_{ij}, \alpha_{ji})$ $\mathrm{cor}_{ij}(\beta_{ijk}\beta_{jik})$	1 0 -1	equal unrelated opposite	(2)
Presence of indirect effects		Yes No	absent present	(4)
Interaction type second-order	β_{ijk}	> 0 < 0 no restriction $= 0$	intensify weaken mixed no second-order	(1) and (3)

Table 1: Design of full factorial virtual experiment.

lations. We parametrized the first-order interactions using distributions of empirically obtained first-order interactions (see supplementary informations S2). We sampled "strong" first-order interactions between the $Q_1 - 1.5(Q_3 - Q_1)$ and $Q_3 + 1.5(Q_3 - Q_1)$ of those distributions, where Q_1 and Q_3 are the first and third quartile, to remove outliers (defined as species interactions outside this range, Appendix S3). Similarly, we sampled "weak" first-order interactions between the Q_1 and Q_3 of the empirical distributions of interaction strength. We fitted linear regressions to measure the effect of species richness on $\mathscr{N}_i$ and $\mathscr{F}_i$. With this approach we were able to investigate the effects of all complexities (1)-(4).

Literature data

We found three review papers that collected multi-species Lotka-Volterra interaction matrices (Adler *et al.*,
2018; Fort, 2018; Keddy & Shipley, 1989), representing a total of 33 interaction matrices, ranging from 3 to
9 species, and containing 29 plant, 2 phytoplankton, 1 zooplankton and 1 ciliate communities. We normal-
ized all these data such that $\alpha_{ii} = -1$. The interaction matrices were obtained through pairwise experiments,
measuring the interspecific effect of one species on the other. For each multi-species community we con-
structed all possible sub-communities with at least two species, leading to a total of 2544 communities that
varied in species richness from 2 to 9. We excluded all communities in which not all interaction strengths
were available, e.g. because of a "NA" entry in the sampled sub-community, leading to 2296 communi-
ties. For 1376 communities we could not compute $\mathcal{N}_i$ and $\mathcal{F}_i$. That is because, like any method seeking to
quantify frequency dependence, our approach is based on invasion analysis: the capacity of an invader to
grow with the other species at their non-zero equilibrium. Thus, one must be able to compute the invasion
growth rate of each species, which is the per capita growth rate $f_i(0, N_j^{-i,*})$ when the focal species i is nearly
absent (mathematically equal to 0) and the other species are at their equilibrium density N^{-i*}. $\mathcal{N}_i$ and $\mathcal{F}_i$
are thus only computable for communities where all species in each subcommunity (the community without
the invading species) coexist stably. We therefore computed $\mathcal{N}_i$ and $\mathcal{F}_i$ for the remaining 920 communities
(see Appendix S4).

For each interaction matrix obtained from the literature we computed $\mathcal{N}_i$ and $\mathcal{F}_i$ using equation 2 and 3.
For each of the 33 interaction matrices, we regressed $\mathcal{N}_i$ against species richness of the sub-communities.
These data contained many outliers, which skewed the results of our linear regressions. We therefore used
a Theil-Sen estimator for the slope, which is more robust to outliers than linear regression based on least
squares (Sen, 1968). We fitted (using least squares) a saturating function $\mathcal{F}_i = \frac{n-2}{(n-2)+H}$ for the fitness
differences. This saturating response was chosen for $\mathcal{F}_i$, because our analytical results suggested such a
response.

Results

Analytical solutions

For the linear Lotka-Volterra model without higher-order interactions (i.e. $\beta_{ijk} = 0$), we can explicitly compute $\mathscr{F}_i$ and $\mathscr{N}_i$ the fitness and niche differences of species i in the multispecies community (see supplementary informations S1):

$$1 - \mathscr{F}_i \quad = \sum_{j,\alpha_{ij} \neq 0} \left(1 - \mathscr{F}_{ij}^{(2)}\right) \frac{N_j^{-i,*}}{N_j^*} \tag{4}$$

$$\mathscr{N}_i \quad = \frac{\sum_{j,\alpha_{ij} \neq 0} \left(c_{ij} N_j^{-i,*}\right) \mathscr{N}_{ij}^{(2)}}{\sum_{j,\alpha_{ij} \neq 0} c_{ij} N_j^{-i,*}} \tag{5}$$

with $\mathscr{F}_{ij}^{(2)}$ and $\mathscr{N}_{ij}^{(2)}$ the fitness and niche difference of species i in a two-species community consisting of species i and j. $N_j^{-i,*}$ is the equilibrium density of species j in the absence of species i, N_j^* is the equilibrium density of species j in monoculture and c_{ij} is the conversion factor from species j to species i (see methods). The sums are taken across all species j with which species i interacts directly, i.e. $\alpha_{ij} \neq 0$.

Eq. 4 and 5 illustrate our two main results. First, $1 - \mathscr{F}_i$ is the weighted *sum* of the two-species fitness differences $1 - \mathscr{F}_{ij}^{(2)}$, across all species pairs with which i interacts. The weights $\frac{N_j^{-i,*}}{N_j^*}$ are the relative yields, as known from biodiversity ecosystem functioning research (Fox, 2005; Hector *et al.*, 2001). The effect of species richness on fitness differences will therefore be similar to the effect of species richness on the sum of the relative yield, known as the relative yield total ($\sum_{j \neq i} \frac{N_j^{i-,*}}{N_j^*}$), which is known to increase with species richness in many communities (Carroll *et al.*, 2011; Grace *et al.*, 2016; Loreau, 2004). Hence, $1 - \mathscr{F}_i$ (and therefore of $\mathscr{F}_i$ decreases) will on average increase with species richness. Second, $\mathscr{N}_i$ is the weighted *average* of the two-species niche overlaps $\mathscr{N}_{ij}^{(2)}$. Hence, species richness will, on average, not affect $\mathscr{N}_i$. Since we did not make assumptions about the α_{ij}, these results are independent of the details of interspecific interactions, i.e. the results apply regardless of complexities (1) and (2).

Equation 5 and 4 can be approximated by assuming constant interspecific interaction strength $\overline{\alpha}$ (see supplementary informations 1). This yields $\mathscr{N}_i \approx 1 - \overline{\alpha}$ and $\mathscr{F}_i \approx 1 - \frac{n-1}{1-(n-2)\overline{\alpha}}$, from which it is clear that $\mathscr{N}_i$ is independent of species richness n and $\mathscr{F}_i$ is an increasing but saturating function of species richness.

The saturation occurs because the sum of the weights $\frac{N_j^{-i,*}}{N_j^*}$, the relative yield total, will saturate as well in the Lotka-Volterra model (Loreau, 2004; Spaak *et al.*, 2017). Including other complexities does not affect these two main results (Appendix S2).

Link to resource competition

The fact that $1 - \mathscr{F}_i$ is a weighted sum, while $\mathscr{N}_i$ is a weighted average makes intuitive sense when realising that the interaction coefficients α_{ij} can, under certain conditions, be related to resource utilisation (Chesson, 1990; MacArthur, 1970). We want to compute the $\mathscr{N}_1$ and $\mathscr{F}_1$ of the yellow focal species (yellow, Fig. 1) in presence of one up to five competitors (Fig 1 A-E). We assume that the species only differ in resource utilisation rates, not in other parameters such as mortality.

In a two species community, the species with the higher total utilisation rate, denoted $\|A_i\|$ the area under the curve A_i, will have a competitive advantage and consequently the higher fitness difference. One could therefore intuit that the fitness difference is linked to the ratio of total resource utilisation rates, i.e. $\mathscr{F}_1 \approx 1 - \frac{\|A_2\|}{\|A_1\|}$. Fitness differences therefore increase with species richness, as each competitor increases the total resource utilisation rates of all competitors combined (Fig. 1 F), i.e. $\mathscr{F}_1 \approx 1 - \frac{\sum \|A_j\|}{\|A_1\|}$. It turns out that this intuition is almost correct; we only have to add weights to the sum according to the densities of the species at equilibrium (compare this equation to eq. 4). $\mathscr{F}_1$ thus increases (becomes more negative), as species richness increases (note that $\mathscr{F}_i = 0$ means no fitness differences and more negative $\mathscr{F}_i$ means stronger fitness differences).

Intuitively, $1 - \mathscr{N}_i$ is the proportion of shared resources between the focal species and its competitors, that is the amount of shared resources scaled by the total consumption of the species. In a two species community we therefore intuit $\mathscr{N}_1 \approx 1 - \frac{\|A_1 \cap A_2\|}{\|A_1\| \cdot \|A_2\|}$. Increasing species richness will increase the sum of shared resources, as the focal species will share resources with each competitor (Fig. 1 G), but also the sum of the total consumption of the species (Fig. 1 F). We therefore expect $\mathscr{N}_1 \approx 1 - \frac{\|A_1 \cap \sum_j A_j\|}{\|A_1\| \cdot \sum_j \|A_j\|}$ to be independent of species richness (Fig 1 H). Again, this intuition is correct up to the inclusion of the weights according to the species equilibrium densities.

Full-factorial simulations

The simulations with strong first-order interactions only partially seem to confirm the predictions made by theory (Fig. 2). That is, $\mathcal{N}_i$ is not invariant but approaches 1 as species richness increases (Fig. 2 A), which seems to contradict the theoretical results. Yet, species richness does not directly affect $\mathcal{N}_i$, but rather affects the average interaction strength $\bar{\alpha}$, which in turn affects $\mathcal{N}_i$, which can be approximated by $\mathcal{N}_i \approx 1 - \bar{\alpha}$ (see Appendix S2). That is because, by design, any method based on invasion growth rates (such as those to compute $\mathcal{N}_i$ and $\mathcal{F}_i$) can only be applied to communities in which invasion analysis is possible. Hence, too strong negative interactions prevent the invasion into highly-diverse communities, and will often impede feasible $n - 1$ subcommunities to begin with (Kokkoris *et al.*, 2002). Hence, species richness selects against communities with overly strong negative interactions, which leads to on average weaker interactions at higher species richness (see supplementary Fig. S3). Similarly, species densities in communities with strong positive interactions will tend to grow to infinity, and more so in species-rich communities, because interspecific facilitation is stronger than intraspecific limitation (self-regulation). Again, species richness selects against strong positive interactions, weakening the average interaction strength (see supplementary Fig. S3). This selection of weak (negative and positive) interspecific interactions causes $\mathcal{N}_i$ to approach 1. $\mathcal{F}_i$ increases with species richness for all parameter settings, as predicted by the theory (see Fig 2 B).

The simulation results based on weak interaction strengths allow assessing the direct effect of species richness on $\mathcal{N}_i$ and $\mathcal{F}_i$ without the confounding effect of species richness on interspecific interaction strength α_{ij}. In these simulations, the effect of species richness on interspecific interactions was much weaker (see supplementary Fig. S3). These simulations confirmed our theoretical results; $\mathcal{N}_i$ was on average unaffected by species richness (see Fig. 3 A) and $\mathcal{F}_i$ increased with species richness (Fig. 3B). We illustrate how $\mathcal{N}_i$ and $\mathcal{F}_i$ values jointly varied with species richness, using weak interaction strength: no higher-order interactions ($\beta_{ijk} = 0$), no correlation between the α_{ij}, and maximum connectance (Fig. 3 C). Again, these results hold independently of the complexities (Appendix S3).

Literature Data

The results for the empirical communities reflect those obtained for the simulated communities. The absolute values of the slopes of the linear regressions of $\mathcal{N}_i$ were small (< 0.05) for all but 6 datasets. The slope for the overall regression of $\mathcal{N}_i$ against species richness (Fig. 4A, black line) was small (-0.028). $\mathcal{F}_i$ increased with richness in all but one dataset. Overall, we conclude that the response of $\mathcal{N}_i$ and $\mathcal{F}_i$ to richness for empirical communities did not qualitatively differ from that of randomly generated communities.

The empirical data also revealed cases in which coexistence is possible even though some of the species have negative $\mathcal{N}_i$. This is possible as long as $\mathcal{F}_i$ is sufficiently positive such that $\mathcal{F}_i \geq \frac{-\mathcal{N}_i}{1-\mathcal{N}_i}$. A total of 95 (4.1%) communities were found with species persisting despite having negative $\mathcal{N}_i$.

Non-random community assembly

So far we have focused on a random community assembly, where the location and width of the resource utilisation A_i of species i were chosen randomly (Fig. 1 A-E). We here focus on two different possibilities of non-random community assembly. First, given a species pool, we can rearrange the order of species arrival. By choosing a non-random community assembly, species richness can increase or decrease niche differences, but will always increase fitness differences (Appendix S5). Additionally, averaged over all possible community assemblages species richness does not affect niche differences.

Alternatively, one might ask how species richness affects niche and fitness differences, if species optimize their resource utilisation according to the prevailing species richness (Barabás *et al.* (2016a), Fig. 5 A-E, Appendix S6). Importantly, in this case the two-species community is *not* a subcommunity of the three-species community. This links to the traditional question of species packing, which is how close can species be packed in a given environment and still coexist (Barabás *et al.*, 2014; MacArthur, 1970). In this scenario, species richness still increases fitness differences (Fig. 5 F). However, because of the limited niche space, species in species rich communities are located closer to each other and therefore have a stronger interaction strength on average (Fig. 5 G). Niche differences therefore slightly decrease with species richness (Fig. 5 H).

Discussion

It is well-established that the likelihood for stable coexistence drops with species richness (Godoy *et al.*, 2017; Goh & Jennings, 1977; May, 1972; Serván *et al.*, 2018). Here, we explain this well known result in the context of modern coexistence theory by examining how niche and fitness differences ($\mathscr{N}_i$ and $\mathscr{F}_i$) change with species richness. We found that species richness, *per se*, does not affect $\mathscr{N}_i$ but does increase $\mathscr{F}_i$. This conclusion is based on four independent approaches: mathematical computation, biological intuition, numerical simulations, and metanalysis of experimental data. Overall, the influence of species richness on $\mathscr{N}_i$ and $\mathscr{F}_i$ is robust to inclusion or omission of the complexities (1)-(4), and all their combinations. The fitness differences of a species increases with species richness, as fitness differences measure the fitness of a species compared to the combined fitness of all other species. In multi-species communities, most species will therefore have negative fitness differences, as rarely one species will have higher fitness than all other species combined.

The niche differences of a species measure the proportion of limiting factors, e.g. resources, that are limiting to other species as well. Increasing species richness increases the amount of limiting factors shared with other species, but also the amount of limiting factors that are not shared with other species. The proportion of shared limiting factors is therefore unaffected, on average. Species-rich communities are therefore less likely to coexist (all else being equal), as fitness differences become to strong to be overcome by niche differences.

Limitations

The available experimental data only represented fully connected communities, with no correlation among interactions (complexity (2)) and, most notably, did not contain cases of higher-order interactions (complexity (3)). We do therefore not know whether the parameter values used to describe these higher-order interactions in our simulations (and therefore the simulation results) are realistic. The available experimental data were biased towards competitive communities of terrestrial plants with relatively low species richness. Our simulations suggest that our conclusions hold for other networks as well, but we were not able to sup-

port this claim with empirical data. Computing $\mathcal{N}_i$ and $\mathcal{F}_i$ on a larger collection of natural communities would help to refine our understanding of this process. However, obtaining the full interaction matrix for species-rich communities is challenging. To obtain interaction matrices, various approaches exist. For example, one uses the frequency of interaction between species (e.g. number of visits of a pollinator on a plant) as a proxy for interaction strength. The robustness of this approach, however, still needs to be tested (García-Callejas *et al.*, 2018). Other methods consist of estimating interaction strength based on, for example, biomass (Moore *et al.*, 1996; Zhao *et al.*, 2019), mass ratio (Emmerson & Raffaelli, 2004) or production and consumption rates of species (Christensen V. & D., 1992; Jacquet *et al.*, 2016). These different methods rely on different assumptions and may therefore influence the resulting matrix estimate (Carrara *et al.*, 2015).

Given these limitations, one can ask to what extent addressing them would change our conclusions. In communities where species richness increases total abundance, which is the case for various communities (Grace *et al.*, 2016; Loreau, 2004; Turnbull *et al.*, 2013), we expect the no-niche growth rate $f_i(\sum_j c_{ij} N_j^{-i,*}, 0)$ to become more negative, as $\sum_j c_{ij} N_j^{-i,*}$ increases (eq. 2, 3). Consequently, we expect species richness to increase fitness differences, i.e. make $\mathcal{F}_i$ more negative, as it is linear in the no-niche growth rate. Conversely, in communities where species richness decreases total abundance we expect the opposite, that is: fitness differences might decrease with species richness. It is less clear how species richness will affect niche differences in models not explored in the current paper , e.g. with different per-capita growth rates functions f_i, or with a community with age structure (Chu & Adler, 2015). Niche differences depend on the invasion growth rate and the no-niche growth rates, which both depend on the species richness and total abundance. When species richness has a stronger negative effect on the no-niche growth rate than on the invasion growth rate, then niche differences will increase with species richness. If the invasion growth rate is decreases more, niche differences will decrease.

Additionally, we have mostly took the simplifying assumption that species richness is independent of other factors such as the average interaction strength, connectance or correlation (Cohen & Briand, 1984; Kokkoris *et al.*, 2002).

The main determinant of niche differences is the average interaction strength, which might decrease

with species richness due to coexistence requirements (Fig. 2A) or might increase due to increasing overlap of resource requirements (Fig. 5 G).

New insights

Our results yield two new insights, other than the main result on how $\mathcal{N}_i$ and $\mathcal{F}_i$ varies with species richness.

A first insight is that negative niche differences do not necessarily preclude coexistence. Negative niche differences have been attributed to priority effects and therefore viewed as precluding coexistence (Fukami *et al.*, 2016; Ke & Letten, 2018). Our framework confirms this finding for the case of competitive two-species communities (Spaak & De Laender, 2020). However, in contrast to species in two-species communities, species in multi-species communities will not all have the same niche differences (example community in Fig 4). This implies that species *a*, with negative niche differences and low fitness differences, can coexist with species *b* and *c* that have positive niche differences and negative fitness differences. Consequently, multiple species can have negative niche differences in a multi-species communities and still persist. In our empirical data set, we found six three-species communities in which all but one species had negative niche differences. In general, we argue that a community in which all species have negative niche differences and coexist is theoretically possible. However, the kind of model and how it should be parametrized remains to be examined.

A second insight is that one can infer $\mathcal{N}_i$ and $\mathcal{F}_i$ in multi-species communities from $\mathcal{N}_i$ and $\mathcal{F}_i$ measured in pairwise interaction experiments. If one measures $\mathcal{N}_i$ and $\mathcal{F}_i$ for each two-species sub-community of an *n* species community, which is typically done (Gallego *et al.*, 2019; Godoy & Levine, 2014; Narwani *et al.*, 2013; Petry *et al.*, 2018), one can estimate $\mathcal{N}_i \approx \frac{\sum_j \mathcal{N}_{ij}^{(2)}}{(n-1)}$. With one additional multi-species experiment to estimate the relative yield RY_i we obtain an estimation of $\mathcal{F}_i \approx 1 - \sum_j (1 - \mathcal{F}_{ij}^{(2)}) \cdot \mathrm{RY}_j$ as well. This indicates that two-species experiments are sufficient to predict $\mathcal{N}_i$ and $\mathcal{F}_i$ in multi-species communities.

Finally, one of the key questions in community ecology is whether niche differences are strong enough to overcome fitness differences and allow coexistence. Often, niche differences are found to be not only sufficiently strong, but much stronger than strictly needed (Chu & Adler, 2015; Levine & HilleRisLambers, 2009). The present results offer a potential explanation for this observation. That is, niche differences not

only need to be sufficiently strong to overcome fitness differences of one or few competitors, as typically considered in empirical studies, but sufficiently strong to overcome fitness differences of the entire resident community, as niche differences is independent of species richness. Our results therefore allow asking the more general question of how many species one can pack in a community, given its niche difference.

References

Adler, P.B., HilleRislambers, J. & Levine, J.M. (2007). A niche for neutrality. *Ecology Letters*, 10, 95–104.

Adler, P.B., Smull, D., Beard, K.H., Choi, R.T., Furniss, T., Kulmatiski, A., Meiners, J.M., Tredennick, A.T. & Veblen, K.E. (2018). Competition and coexistence in plant communities: intraspecific competition is stronger than interspecific competition. *Ecology Letters*, 21, 1319–1329.

Aladwani, M. & Saavedra, S. (2019). Mathematical Biosciences Is the addition of higher-order interactions in ecological models increasing the understanding of ecological dynamics ? *Mathematical Biosciences*, 315, 108222.

Allesina, S. & Tang, S. (2012). Stability criteria for complex ecosystems. *Nature*, 483, 205–208.

Allesina, S. & Tang, S. (2015). The stability–complexity relationship at age 40: a random matrix perspective. *Population Ecology*, 57, 63–75.

Barabás, G., D'Andrea, R., D'Andrea, R. & D'Andrea, R. (2016a). The effect of intraspecific variation and heritability on community pattern and robustness. *Ecology Letters*, 19, 977–986.

Barabás, G., D'Andrea, R. & Stump, S.M. (2018). Chesson's coexistence theory. *Ecological Monographs*, 88, 277–303.

Barabás, G., Michalska-Smith, M.J. & Allesina, S. (2016b). The effect Of intra- and interspecific competition on coexistence in multispecies communities. *American Naturalist*, 188, E1–E12.

Barabás, G., Pásztor, L., Meszéna, G. & Ostling, A. (2014). Sensitivity Analysis of Coexistence in Ecological Communities : Theory and Application Supporting Information. pp. 1–22.

Bimler, M.D., Stouffer, D.B., Lai, H.R. & Mayfield, M.M. (2018). Accurate predictions of coexistence in natural systems require the inclusion of facilitative interactions and environmental dependency. *Journal of Ecology*, 106, 1839–1852.

Cardinaux, A., Hart, S.P. & Alexander, J.M. (2018). Do soil biota influence the outcome of novel interactions between plant competitors? *Journal of Ecology*, 106, 1853–1863.

Carmel, Y., Cornell, S.J., Belmaker, J., Suprunenko, Y.F., Kent, R., Kunin, W.E. & Bar-Massada, A. (2017). Using exclusion rate to unify niche and neutral perspectives on coexistence. *Oikos*, 126, 1451–1458.

Carrara, F., Giometto, A., Seymour, M., Rinaldo, A. & Altermatt, F. (2015). Inferring species interactions in ecological communities: A comparison of methods at different levels of complexity. *Methods in Ecology and Evolution*, 6, 895–906.

Carroll, I.T., Cardinale, B.J. & Nisbet, R.M. (2011). Niche and fitness differences relate the maintenance of diversity to ecosystem function. *Ecology*, 92, 1157–1165.

Cenci, S. & Saavedra, S. (2018). Structural stability of nonlinear population dynamics. *Physical Review E*, 97.

Chesson, P. (1990). MacArthur's consumer-resource model. *Theoretical Population Biology*, 37, 26–38.

Chesson, P. (1994). Multispecies Competition in Variable Environments. *Theoretical Population Biology*, 45, 227–276.

Chesson, P. (2000). Mechanisms of maintenance of speciesdiversity. *Annual Review of Ecology, Evolution, and Systematics*, 31.

Chesson, P. (2003). Quantifying and testing coexistence mechanisms arising from recruitment fluctuations. *Theoretical Population Biology*, 64, 345–357.

Chesson, P. (2013). Species Competition and Predation. In: *Ecological Systems*. Springer New York, pp. 223–256.

Chesson, P. & Kuang, J.J. (2008). The interaction between predation and competition. *Nature*, 456, 235–238.

Christensen V. & D., P. (1992). ECOPATH II - a software for balancing steady-state ecosystem models and calculating network characteristics. *Ecological Modelling*, 61, 169–185.

Chu, C. & Adler, P.B. (2015). Large niche differences emerge at the recruitment stage to stabilize grassland coexistence. *Ecological Monographs*, 85, 373–392.

Cohen, J. & Briand, F. (1984). Trophic links of community food webs. *Proceedings of the National Academy of Sciences*, 81, 4105–4109.

Ellner, S.P., Snyder, R.E., Adler, P.B. & Hooker, G. (2019). An expanded modern coexistence theory for empirical applications. *Ecology Letters*.

Emmerson, M.C. & Raffaelli, D. (2004). Predator-prey body size, interaction strength and the stability of a real food web. *Journal of Animal Ecology*, 73, 399–409.

Fontaine, C., Guimarães, P.R., Kéfi, S., Loeuille, N., Memmott, J., van der Putten, W.H., van Veen, F.J. & Thébault, E. (2011). The ecological and evolutionary implications of merging different types of networks. *Ecology Letters*, 14, 1170–1181.

Fort, H. (2018). On predicting species yields in multispecies communities: Quantifying the accuracy of the linear Lotka-Volterra generalized model. *Ecological Modelling*, 387, 154–162.

Fox, J.W. (2005). Interpreting the 'selection effect' of biodiversity on ecosystem function. *Ecology Letters*, 8, 846–856.

Fukami, T., Mordecai, E.A. & Ostling, A. (2016). A framework for priority effects. *Journal of Vegetation Science*, 27, 655–657.

Gallego, I., Venail, P. & Ibelings, B.W. (2019). Size differences predict niche and relative fitness differences between phytoplankton species but not their coexistence. *ISME Journal*.

García-Callejas, D., Molowny-Horas, R. & Araújo, M.B. (2018). Multiple interactions networks: towards more realistic descriptions of the web of life. *Oikos*, 127, 5–22.

Godoy, O. & Levine, J.M. (2014). Phenology effects on invasion success: Insights from coupling field experiments to coexistence theory. *Ecology*, 95, 726–736.

Godoy, O., Stouffer, D.B., Kraft, N.J. & Levine, J.M. (2017). Intransitivity is infrequent and fails to promote annual plant coexistence without pairwise niche differences. *Ecology*, 98, 1193–1200.

Goh, B. & Jennings, L. (1977). No Feasibility and stability in randomly assembled Lotka-Volterra models. *Ecological Modelling*, 3.

Grace, J.B., Anderson, T.M., Seabloom, E.W., Borer, E.T., Adler, P.B., Harpole, W.S., Hautier, Y., Hillebrand, H., Lind, E.M., Pärtel, M., Bakker, J.D., Buckley, Y.M., Crawley, M.J., Damschen, E.I., Davies, K.F., Fay, P.A., Firn, J., Gruner, D.S., Hector, A., Knops, J.M., MacDougall, A.S., Melbourne, B.A., Morgan, J.W., Orrock, J.L., Prober, S.M., Smith, M.D. & Rinke_suppl (2016). Integrative modelling reveals mechanisms linking productivity and plant species richness. *Nature*, 529, 390–393.

Grainger, T.N., Letten, A.D., Gilbert, B. & Fukami, T. (2019). Applying modern coexistence theory to priority effects. *Proceedings of the National Academy of Sciences*, 116, 6205–6210.

Hart, S.P., Freckleton, R.P. & Levine, J.M. (2018). How to quantify competitive ability. *Journal of Ecology*, 106, 1902–1909.

Hector, A., Joshi, J., Lawler, S., Spehn, E.M. & Wilby, A. (2001). Conservation implications of the link between biodiversity and ecosystem functioning. *Oecologia*, 129, 624–628.

Hutchinson, G.E. (1957). Concluding Remarks. *Cold Spring Harbor Symposia on Quantitative Biology*, 22, 415–427.

Jacquet, C., Moritz, C., Morissette, L., Legagneux, P., Massol, F., Archambault, P. & Gravel, D. (2016). No complexity-stability relationship in empirical ecosystems. *Nature Communications*, 7, 1–8.

Ke, P.J. & Letten, A.D. (2018). Coexistence theory and the frequency dependence of priority effects. *biorXiv*.

Keddy, P.A. & Shipley, B. (1989). Competitive heirarchies in herb plant communities.

Kokkoris, G.D., Jansen, V.A.A., Loreau, M. & Troumbis, A.Y. (2002). Variability in interaction strength and implications for biodiversity. *Journal of Animal Ecology*, 71, 362–371.

Lanuza, J.B., Bartomeus, I. & Godoy, O. (2018). Opposing effects of floral visitors and soil conditions on the determinants of competitive outcomes maintain species diversity in heterogeneous landscapes. *Ecology Letters*, 21, 865–874.

Letten, A.D., Ke, P.J. & Fukami, T. (2017). Linking modern coexistence theory and contemporary niche theory. *Ecological Monographs*, 87, 161–177.

Letten, A.D. & Stouffer, D.B. (2019). The mechanistic basis for higher-order interactions and non-additivity in competitive communities. *Ecology Letters*, 22, 423–436.

Levine, J.M., Bascompte, J., Adler, P.B. & Allesina, S. (2017). Beyond pairwise mechanisms of species coexistence in complex communities. *Nature*, 546, 56–64.

Levine, J.M. & HilleRisLambers, J. (2009). The importance of niches for the maintenance of species diversity. *Nature*, 461, 254–257.

Loreau, M. (2004). Does functional redundancy exist? *Oikos*, 104, 606–611.

MacArthur, R. (1970). Species packing and competitive equilibrium for many species. *Theoretical Population Biology*, 1, 1–11.

Matías, L., Godoy, O., Gómez-Aparicio, L. & Pérez-Ramos, I.M. (2018). An experimental extreme drought reduces the likelihood of species to coexist despite increasing intransitivity in competitive networks. *Journal of Ecology*, 106, 826–837.

May, R.M. (1972). Will a large complex system be stable? *Nature*, 238, 413–414.

Maynard, D.S., Serván, C.A., Capitán, J.A. & Allesina, S. (2019). Phenotypic variability promotes diversity and stability in competitive communities. *Ecology Letters*, 22, 1776–1786.

513   Mickalide, H. & Kuehn, S. (2019). Higher-order interaction inhibits bacterial invasion of a phototroph-predator microbial community. *bioRxiv*, p. 564260.

Moore, J.C., De Ruiter, P.C., Hunt, H.W., Coleman, D.C. & Freckman, D.W. (1996). Microcosms and soil
516   ecology: Critical linkages between field studies and modelling food webs. *Ecology*, 77, 694–705.

Napier, J.D., Mordecai, E.A. & Heckman, R.W. (2016). The role of drought- and disturbance-mediated competition in shaping community responses to varied environments. *Oecologia*, 181, 621–632.

519   Narwani, A., Alexandrou, M.A., Oakley, T.H., Carroll, I.T. & Cardinale, B.J. (2013). Experimental evidence that evolutionary relatedness does not affect the ecological mechanisms of coexistence in freshwater green algae. *Ecology Letters*, 16, 1373–1381.

522   Petry, W.K., Kandlikar, G.S., Kraft, N.J., Godoy, O. & Levine, J.M. (2018). A competition–defence trade-off both promotes and weakens coexistence in an annual plant community. *Journal of Ecology*, 106, 1806–1818.

525   Saavedra, S., Rohr, R.P., Bascompte, J., Godoy, O., Kraft, N.J. & Levine, J.M. (2017). A structural approach for understanding multispecies coexistence. *Ecological Monographs*, 87, 470–486.

Sen, P.K. (1968). Estimates of the Regression Coefficient Based on Kendall's Tau. *Journal of the American*
528   *Statistical Association*, 63, 1379–1389.

Serván, C.A., Capitán, J.A., Grilli, J., Morrison, K.E. & Allesina, S. (2018). Coexistence of many species in random ecosystems. *Nature Ecology and Evolution*, 2, 1237–1242.

531   Song, C., Barabás, G. & Saavedra, S. (2019). On the consequences of the interdependence of stabilizing and equalizing mechanisms. *The American Naturalist*, pp. 000–000.

Spaak, J.W., Baert, J.M., Baird, D.J., Eisenhauer, N., Maltby, L., Pomati, F., Radchuk, V., Rohr, J.R.,
534   Van den Brink, P.J. & De Laender, F. (2017). Shifts of community composition and population density substantially affect ecosystem function despite invariant richness. *Ecology Letters*, 20, 1315–1324.

Spaak, J.W. & De Laender, F. (2020). Intuitive and broadly applicable definitions of niche and fitness differences. *Ecology Letters*, p. doi: https://doi.org/10.1101/482703.

Turelli, M. (1978). Does environmental variability limit niche overlap? *Proceedings of the National Academy of Sciences of the United States of America*, 75, 5085–9.

Turnbull, L.A., Levine, J.M., Loreau, M. & Hector, A. (2013). Coexistence, niches and biodiversity effects on ecosystem functioning. *Ecology Letters*, 16, 116–127.

Veresoglou, S.D., Rillig, M.C. & Johnson, D. (2018). Responsiveness of plants to mycorrhiza regulates coexistence. *Journal of Ecology*, 106, 1864–1875.

Wainwright, C.E., HilleRisLambers, J., Lai, H.R., Loy, X. & Mayfield, M.M. (2019). Distinct responses of niche and fitness differences to water availability underlie variable coexistence outcomes in semi-arid annual plant communities. *Journal of Ecology*, 107, 293–306.

Walsh, M.R. (2013). The evolutionary consequences of indirect effects. *Trends in Ecology & Evolution*, 28, 23–29.

Zhao, L., Zhang, Q.G. & Zhang, D.Y. (2016). Evolution alters ecological mechanisms of coexistence in experimental microcosms. *Functional Ecology*, 30, 1440–1446.

Zhao, Q., Van den Brink, P.J., Carpentier, C., Wang, Y.X.G., Rodríguez-Sánchez, P., Xu, C., Vollbrecht, S., Gillissen, F., Vollebregt, M., Wang, S. & De Laender, F. (2019). Horizontal and vertical diversity jointly shape food web stability against small and large perturbations. *Ecology Letters*, pp. 1152–1162.

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Figure 1: A-E: Resource utilisation of the yellow focal species and its competitors. F: Increasing the species richness will increase the total utilisation of the resident species $\sum_j \|A_j\|$. Similarly, we expect $\mathscr{F}_i \approx 1 - \frac{\sum_j \|A_j\|}{\|A_1\|}$ to increase with species richness, as it scales with the ratio of total resource consumption. G: The amount of shared resources (hatched region from panels A-E) increases with species richness. H: As both the amount of shared resources increase (panel G) and the total utilisation (panel F) increase, we expect the ratio to be independent of species richness. Similarly, we expect $\mathscr{N}_i \approx 1 - \frac{\|A_i \cap \sum_j A_j\|}{\|A_i\| \cdot \sum_j \|A_j\|}$ to be, on average, independent of species richness. F-H: The colours of the bar correspond to the contribution of each of the resident species to the total of the bar.

Figure 2: $\mathcal{N}_i$ and $\mathcal{F}_i$ as a function of species richness in simulated communities with strong first-order interspecific interactions. A: Contrary to predictions from theory, niche differences change with species richness when first-order interspecific interactions are either positive (green) or negative (red; results for unconstrained interspecific interactions are shown in blue). However, this is because interaction strength decreases with increasing species richness in these cases (see Fig. S2). Each line represents a linear regression of niche differences as a function of species richness for one factorial setting of the full-factorial experiment (see table 1). B: Species richness, however, makes fitness differences more negative (i.e. larger). Note the differences in y-scale between panel A and B. C: Distribution of $\mathcal{N}_i$ and $\mathcal{F}_i$ for simulated theoretical communities that are fully connected, and exhibit first-order interactions without correlations, i.e. similar to the experimental communities (see Fig. 4). Each dot represents $\mathcal{N}_i$ and $\mathcal{F}_i$ of one species in a community composed of 2-6 species (see colour legend). The black line indicates the persistence line, species below this line are assumed to persist in the community. Note the inverted y-axis.

Figure 3: As Fig. 2, but with weak first-order interspecific interactions. A: As predicted by the mathematical results, species richness does not affect niche differences, because communities with different species richness had comparable interaction strengths. B: Species richness, however, makes fitness differences more negative (i.e. larger). C: Distribution of $\mathcal{N}_i$ and $\mathcal{F}_i$ for simulated theoretical communities that are fully connected, and exhibit first-order interactions without correlations, i.e. similar to the experimental communities (see Fig. 4).

Figure 4: Similar to Fig 2 for empirically observed communities. Each grey line corresponds to a fit of a linear ($\mathcal{N}_i$, A) and saturating ($\mathcal{F}_i$, B) regression model to one dataset. The black line represents a fit through all $\mathcal{N}_i$ (A) and $\mathcal{F}_i$ (B) values. Grey dots in panel A and B represent the raw $\mathcal{N}_i$ and $\mathcal{F}_i$ values. Facilitation, i.e. species having a positive net effect on another, and therefore $\mathcal{N}_i > 1$ is common in the datasets we found. We highlight one specific three-species community (grey line) where all species coexist, even though species a has $\mathcal{N}_i < 0$, a property associated with priority effects and therefore exclusion. Axis from C are truncated to show $\sim$95% of all data points.

Figure 5: A-E: Resource utilisation of two to six competing species. Species evolve to optimal foraging strategies according to Barabás *et al.* (2016a). F: Increasing species richness increases fitness differences, as predicted by theory G: As the niche width is limited, species are located closer to each other in species rich communities and therefore have a stronger interaction strength on average. Each dot represents one value of the interaction matrix α_{ij}. H: Consequently, niche differences decrease with species richness, contrary to theory. The example deviates from the theoretical predictions, as one of the key hypothesis of random assembly is not meat.