Plant budding speciation predominant by ecological and

geographical differentiation: an 'evolutionary snapshot' in

***Iberodes* (Boraginaceae).**

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ABSTRACT

Traditional classification of speciation modes has been focused on physical

barriers to gene flow. Allopatry—geographic separation of populations so that

gene flow between them is more or less severed—has as a consequence been

viewed as the most common mechanism of speciation (allopatric speciation). By

contrast, parapatry and sympatry—no physical barrier is found separating

populations (adjacent or overlapped) so that speciation takes place in the face of

ongoing gene flow— is more difficult to demonstrate. *Iberodes* (Boraginaceae) is a

primarily Iberian genus that comprises a relatively small number of recently

derived species (five) with contrasting diaspore dispersal traits, habitats and

distribution patterns. In particular, species distributions of *Iberodes* offer an ideal system to study drivers of lineage divergence and differentiation in the course of evolution. To reconstruct the evolutionary history of the genus, we addressed an integrative study combining a suitable set of tools: (i) phylogenetics based on restriction-site associated DNA sequencing (RAD-seq), (ii) morphometrics, and (iii) climatic niche modelling. Our robust phylogenetic reconstruction of *Iberodes* reveals a predominantly budding speciation pattern. Climatic niche analyses, together with the morphometric data and species distributions, suggests that ecological and geographical differentiation interact in concert, in such a way that both allopatric and parapatric have been operating in *Iberodes*.

Keywords: Restriction-site associated DNA sequencing (RAD-seq), molecular dating, species distribution modelling, niche overlap, paraphyly, allopatry, parapatry.

INTRODUCTION

The classification of modes of speciation has been one of the more contentious topics in evolutionary biology for several decades (e.g. Templeton, 1981; Rieseberg and Brouillet, 1994; Butlin *et al.*, 2008; Horandl and Stuessy, 2010). Reproductive barriers are the most frequently used criteria for classifying three main speciation modes (Futuyma, 2009). Speciation of adjacent populations in absence of clear physical or reproductive barriers is frequently explained under the process of parapatric speciation, which entails the evolution of reproductive isolation in the face of limited but ongoing gene flow (Coyne and Orr, 2004, p. 112). Parapatric speciation can be considered intermediate between allopatric and sympatric

speciation. In allopatry, gene flow is prevented by geographic barriers between

populations. In sympatry, partial or complete reproductive isolation arises

between subsets of a single population without spatial segregation (keeping

overlapping ranges). Sympatric speciation often entails significant levels of gene

flow at the early stages of differentiation (Gavrilets, 2003; Futuyma, 2009).

Whereas allopatry is widely viewed to be the most common mode of speciation

(Coyne and Orr, 2004; Vargas *et al.*, 2018), the relative importance of parapatry

and sympatry has been the subject of considerable debate (Fitzpatrick *et al.*, 2008,

2009; Arnold, 2015) in large part due to the difficulty of (1) resolving fine-scale

phylogenetic relationships among very closely related populations that may still be

exchanging genes at a low level, and (2) quantifying the amount of gene flow

between populations during population divergence (Barluenga *et al.*, 2006;

Fontdevilla, 2014). In the last decade, new genomic, phylogenetic, and ecological

tools have made non-allopatric modes of speciation increasingly amenable to

study, allowing us to explore with statistical rigor a wider range of speciation

scenarios (e.g. Savolainen *et al.*, 2006; Chozas *et al.*, 2017; Zheng *et al.*, 2017). In

particular, there are some critical points of evidence that would support a

parapatric scenario: (1) phylogenetic relationships usually reflect a progenitor-

derivative (budding) pattern in which the most widely distributed species is the

living progenitor of the most restricted one; (2) the absence of any physical barrier

to gene-flow in both present and past times, i.e. distribution ranges are adjacent;

(3) clear interspecific genetic clustering that putatively discards processes of

secondary contact when testing for ancient and contemporary gene flow; (4) the

need of alternative factors (differential climatic niches, reproductive exclusion,

ploidy variation, among others) of isolation other than physical barriers are

claimed to be in play.

Ecological differentiation is often attributed to non-allopatric speciation (Sobel

et al. 2010; Nosil 2012; Stankowsky *et al.* 2015). Indeed, ecological speciation is

claimed to be more important in plants than in animals because a sessile habit

makes plants more sensitive to fine-scale environmental heterogeneity (Anacker

and Strauss, 2014). Both the influence of niche conservatism (i.e. retention of

ancestral ecological characteristics of species over time, Peterson *et al.* 1999) and

niche evolution (i.e. adaptation of lineages to changes in the environment,

Donoghue and Edwards 2014) have been argued to play a role in speciation and

lineage diversification (Wiens and Graham, 2005; Cavender-Bares, 2019).

Nevertheless, the contribution of ecological factors as primary drivers of

speciation remains more elusive (Mayr, 1954; Pyron and Burbrink, 2010; Yin et al.,

2016).

The advent of inexpensive phylogenomic approaches in the last decade have

made the genetic side of testing recent speciation scenarios tractable (Mc Cormack

et al., 2013; Mc Vay *et al.*, 2017a). High Throughput Sequencing (HTS) approaches

help to analyze loci sampled from the entire genome in reconstructing species

trees (Fernández-Mazuecos *et al.*, 2018) and speciation in the face of gene flow

(Leroy *et al.*, 2017; Folk *et al.*, 2018; Crowl *et al.*, 2019). In particular, restriction-

site associated DNA (RAD-seq) is considerably contributing to our understanding

of recent evolutionary processes, being especially helpful for non-model organisms

since a reference genome is not needed for accurate phylogenetic inference (e.g.

Fitz-Gibbson *et al.*, 2017). RAD-seq has as a consequence been useful in inferring

complex speciation histories: repeated cycles of island connectivity and isolation

(the pump hypothesis; Papadopoulou and Knowles, 2015), incipient sympatric

speciation (Kautt et al., 2016), allopatric speciation despite historical gene flow

(Maguilla *et al.*, 2017), ancient introgression among now-extinct species (Mc Vay *et*

al. 2017b).

The Mediterranean subendemic genus *Iberodes* M.Serrano, R.Carbajal &

S.Ortiz (Boraginaceae, Cynoglossoideae; Serrano *et al.*, 2016) provides an excellent

system for studying recent speciation, morphological and ecological differentiation

because it is clearly demonstrated to be a recently-derived genus (Chacón *et al.*

2017, Otero *et al.* 2019a). *Iberodes* comprises five species and one subspecies that

differ in geographic ranges (including allopatric and parapatric species) and

habitat (ranging from forest understories to scrublands and coastal dunes).

Moreover, all species are endangered and narrowly endemic, except for the

widely-distributed *I. linifolia* of the Iberian Peninsula and southeastern France

(Talavera *et al.*, 2012). Sanger sequencing has failed to resolve phylogenetic

relationships among *Iberodes* species because of scarcely information of traditional

markers (Otero *et al.*, 2014; Holstein *et al.*, 2016ab, Otero *et al.*, 2019a). The

question remains as to what is the relative importance of extrinsic geographical

and ecological barriers in the complex speciation pattern observed for *Iberodes*.

Given the narrow endemicity in most *Iberodes* species, we hypothesize that

geographic barriers rather than ecological factors have been predominant in the

speciation history of *Iberodes*. To test this hypothesis, we aimed to: (1) reconstruct

phylogenetic relationships of the five species; (2) infer speciation patterns and

lineage diversification during the last geological epochs comparing diverse

morphologies and ecologies; and (3) evaluate the relative contribution of

geographical and ecological processes in speciation.

MATERIAL AND METHODS

Taxon sampling

Based on species distributions, between one and four individuals from one to nine populations were sampled for the five species of *Iberodes* including ssp. *gallaecica* and ssp. *littoralis* of *I. littoralis*) (Fig. 1, Table S1, in Supporting Information). Samples of *I. littoralis*, *I. kuzinskyana* and *I. brassicifolia* were collected in the field during 2015 and stored in silica gel until extraction. Since these species are considered threatened according to IUCN criteria (Lopes and Carvalho, 1990; Serrano and Carbajal, 2003; ICNB, 2007; Moreno, 2008; IUCN, 2019; INPN, 2019), permits were obtained prior to collecting. Mature seeds of *I. commutata* were collected in 2015 and germinated in a glasshouse to obtain green leaves for DNA extraction. Both field and herbarium materials were used to represent the large distribution of *I. linifolia* (Table S1). Three species (17 individuals) of the tribe Omphalodeae were included as the outgroup (Otero *et al.*, 2019b): *Omphalodes nitida*, *Myosotidium hortensia*, and *Gyrocaryum oppositifolium* (Table S1).

DNA extraction and RAD library preparation

DNA was extracted from leaf tissue using a modified CTAB protocol from Doyle & Doyle (1987) and Shepherd & McLay (2011), including a precipitation step in isopropanol and ammonium acetate 7.5M at -20ºC overnight to improve the yield. DNA extractions were visualized and quantified with NanoDrop 2000/2000c (Thermo-Fisher, Waltham, Massachusetts) and Qubit Fluorometric Quantification (Thermo-Fisher, Waltham, Massachusetts) in the *Instituto de Investigaciones Biomédicas* (IIBm, CSIC-UAM). Final DNA sample concentrations were standardized to 10 ng/ul. Preparation of single-end RAD-seq libraries using restriction enzyme PstI from genomic DNA was conducted at Floragenex Inc.

149   (Eugene, Oregon) following Baird *et al.* (2008) and sequenced as single-end, 100-

150   bp reactions on an Illumina HiSeq 2500 at the University of Orgeon Genomics &

151   Cells Characterization Core Facility (library C606). Processed data were returned

152   in the Illumina 1.3 variant of the FASTQ format, with Phred quality scores for all

153   bases.

Data clustering

Sequences from Illumina were analyzed through *PyRAD* v. 3.0.6, a pipeline with

seven main steps that filter and cluster putatively orthologous loci from RAD

sequencing reads (Eaton, 2014). This pipeline is able to cluster highly divergent

sequences, taking into account indel variation and nucleotide (SNP) variation. It is

thus well-suited to the inter- and intraspecific scales addressed in our study. As no

evidence of polyploidy was found from flow cytometry, we did not assess

sensitivity of paralog detection to sequencing depth or number of heterozygotic

positions allowed per loci or shared among individuals. Base calls with a Phred

Quality score <20 were replaced by the ambiguous base code (N). Three similarity

percentages were set for the clustering (c) threshold of reads: 85%, 90% and 95%.

Four levels of minimum coverage (m) of a sample in a final locus (4, 12, 20, 28)

were tested for each of the three percentages of clustering similarity, resulting in a

fully factorial set of 12 matrices from the combination of filtering parameters, so

we could assess sensitivity of our phylogenetic inferences to clustering

assumptions. Consensus base calls were made for clusters with a minimum depth

of coverage >5. From each parameter combination we obtained three kinds of

output: (1) a matrix with all loci retrieved for each individual (concatenated

matrix), (2) a matrix with single nucleotide polymorphisms (SNP matrix), and (3) a

matrix with one SNP per locus (unlinked SNP matrix). The loci matrix was

explored with the RADami (Hipp, 2014) and vegan (Oksanen *et al.*, 2008) packages

of R version 3.3.2 (R Core Team, 2013).

Phylogenetic analysis

Maximum likelihood analyses of concatenated matrices obtained in *PyRAD* were

performed in RAxML v8.2.10 (Stamatakis, 2014) through the Cipres Portal (Miller

et al., 2010). We used rapid bootstrapping, GTRGAMMA model, and automatic-stop

bootstrapping with the majority rule criterion. We conducted a maximum

likelihood analysis for each of the 12 *PyRAD* concatenated matrices. Finally, we

compared the topologies based on average bootstrap support (BS). Between the

two topologies with average BS>98%, we chose the one maximizing the number of

loci (HBS tree) for the remaining phylogenetic-based analyses. Moreover, we used

the unlinked SNP matrix from the HBS tree (HBS matrix) to perform a coalescent-

based phylogeny using SVDquartets (Chifman and Kubatko, 2014) in PAUP

(Swofford, 2001) through the Cipres portal (Miller *et al.*, 2010). For SVDquartets

we grouped individuals based on main clades obtained from concatenated analysis

in RAxML and evaluated all possible quartets with 100 bootstrap replicates.

Estimates of divergence times

We used penalized likelihood as implemented in TreePL (Smith and O'Meara,

2012) to estimate a time-calibrated tree for all bootstrap trees (with branch

lengths) obtained in RAxML from the HBS tree. TreePL is suitable for divergence

time estimation when dealing with large amounts of data, as those yielded by high

throughput sequencing such as RAD-sequencing (Zheng and Wiens, 2015). Two

calibration points were used: (1) tribe Omphalodeae (root) (minimum age

(minAge) = 9.062; maximum age (maxAge) = 24.4097) and (2) clade *Iberodes*

(ingroup) (minAge = 0.6591; maxAge = 3.7132) based on the averaged ages and

standard deviation inferred in Otero *et al.* (2019a). Average node ages over all

posterior trees from the latter study were calculated using *treeStat* from the

BEAST v 1.8.2 package (Drummond *et al.*, 2012). We first conducted an analysis

under the "prime" option to select the optimal set of parameter values and we set

random subsample and replicate cross-validation (RSRCV) to identify the best

value for the smoothing parameter lambda. For this first analysis we used the

thorough analysis option and set 200,000 iterations for penalized likelihood and

5,000 iterations for cross validation. Consequently, we repeated the same

thorough analysis and iterations but setting the smoothing value of lambda to 10,

gradient based (opt) and auto-differentiation based optimizers (optad) to five, and

auto-differentiation cross-validation-based optimizers to two (optcvad). Although

TreePL does not draw a prior distribution for the phylogenetic tree, we took into

account the bootstrapped variance in topology and branch length by running the

same TreePL analysis for each bootstrap tree and then a maximum clade

credibility tree using the mean heights was reconstructed in *TreeAnnotator* from

the BEAST v. 1.8.2 package (Drummond *et al.*, 2012).

Phylogenetic inferences showed two cases of paraphyly within the lineage

of *I. linifolia* that involved three other taxa (*I. kuzinskyana, I. littoralis* ssp. *littoralis*

and ssp. *gallaecica*; see results). To evaluate these cases of paraphyly, a set of

analyses were done to analyze the degree of genetic, morphological and ecological

differentiation among the four taxa involved. Therefore, the following analyses

were performed on a subset of data only including these four taxa.

Genetic structure

The HBS matrix was used to perform a Discriminant Analysis of Principal

Components (DAPC) to identify and describe clusters of genetically related

individuals (Jombart *et al.*, 2010). The format of the HBS matrix was converted

from *vcf* to *genlight* with vcfR (Knaus and Grünwald, 2017). DAPC analysis was

performed in adegenet (Jombart, 2008). We fixed the number of clusters to four,

based on the natural groups of four taxonomic species and subspecies (the

Linifolia clade, see results). We used the function 'optim.a.score' to estimate the

optimal number of principal components for discriminant analysis. We

represented genetic differentiation using scatterplots of the principal components

and a barplot representing assignment probabilities of individuals to groups. In

addition, analysis of molecular variance (AMOVA) was performed in Arlequin v.

3.5.2.1 (Excoffier and Lischer, 2010). AMOVA was run to evaluate the level of

genetic differentiation of the paraphyletic *I. linifolia* as a whole with respect to the

other two species (*I. kunzinskyana* and *I. littoralis*) of the Linifolia clade. Therefore,

we considered only two groups, *I. linifolia* on one hand, and the two coastal species

together (*I. kusinskyana* and *I. littoralis*) on the other. Finally, to evaluate a

potential role of historical introgression in determining paraphyly, we conducted

D-statistic tests in PyRAD (see Methods S1 for details).

Morphometric evaluation

We performed a morphometric exploration using principal component analysis

(PCA). We consulted the taxonomic literature to identify diagnostic characters in

Iberodes (Brand, 1921; Ferguson, 1972; Fernández and Talavera, 2012; Neto *et al.*,

2015). We measured 12 diagnostic macromorphological vegetative and

reproductive characters for 111 individuals from 58 vouchers (two different

individuals per voucher except for 5 vouchers with only one individual). We

averaged measures from each two individuals of the same voucher. As a result, we

obtained a final matrix of 58 accessions from 47 locations (74 individuals from 35

locations of *I. linifolia*, 14 from five locations of *I. kuzinskyana*, and 23 from seven

locations of *I. littoralis* (including the two subspecies) (Fig. 1, Table S2). Ten

morphological characters were quantitatively continuous: four vegetative

characters (length of stem, leaves and pedicels, and width of leaves) and six

reproductive characters (length of fruiting calix, width and length of nutlet, length

of nutlet margin, length of the nutlet margin teeth, and hair length). In addition,

one binomial character (presence or absence of bracts), and one quantitative,

discrete character (hair number per mm^2 on the abaxial surface of the nutlet) were

also considered.

PCA was run using the function 'prcomp' from Stats R package (R Core Team,

2013). Results were visualized with the function 'ggbiplot' in the ggbiplot R

package (Vu, 2011).

Climatic niche differentiation and distribution modelling

Occurrences of the four taxa were collected from GBIF (https://www.gbif.org/).

We filtered the resulting dataset by removing points suspected to be incorrect,

such as those placed clearly outside of the known species distribution range (based

on taxonomic literature and databases; Fernández and Talavera, 2012; Muséum

national d'Histoire Naturelle, 2003-2019). Additional geographic coordinates were

obtained from herbarium material and our own fieldwork data. We also estimated

the geographic coordinates of non-georeferenced herbarium specimens, whenever

the locality provided could be pinpointed with an accuracy of ± 5 km (Table S3, Fig.

1). To reduce sampling bias, we filtered the dataset by randomly removing points

that were 0.2° of each other for *I. linifolia* and 0.1° for *I. littoralis* ssp. *littoralis* and

ssp. *gallaecica*. Filtering was not applied for *I. kuzinskyana* because of the low

number of occurrences. The final dataset after filtering included 115 occurrences

of *I. linifolia*, 11 of *I. kuzinskyana*, 16 of *I. littoralis* ssp. *littoralis* and 11 of ssp.

gallaecica.

We obtained 19 bioclimatic variables (resolution of 30") from WorldClim 1.4

(http://www.worldclim.org/; Hijmans et al, 2006) for the study region (34° to 50°

N; 11° W to 7° E). To avoid collinearity among bioclimatic variables, first we

excluded variables displaying a high correlation coefficient with other variables

($|r| > 0.7$) in the study area (Dormann *et al.*, 2013). Then, we calculated the

variance inflation factor (VIF) for each remaining variable using the HH package in

R (Heiberger, 2017) and selected variables with VIF <5 following Benitez-Benitez

et al. (2018). As a result, the following six variables were selected: bio1, bio3, bio4,

bio8, bio12 and bio14. We assessed climatic niche overlap among the four taxa by

pairwise comparisons. The E-space (i.e. environmental space resulting from the six

climatic variables) of each taxon was analyzed in a Principal Component Analysis

(PCA) as implemented in the R package ecospat (Di Cola *et al.*, 2017). To visualize

niche differences along each of the first two principal components, the result was

plotted using the niceOverPlot function (Fernández-López and Villa-Machío, 2017),

which relies on the ggplot2 package (Wickham, 2016). We calculated values of

Schoener's index (D) as a measure of niche overlap (Schoener, 1968; Warren *et al.*,

2008). Then, we conducted tests of niche equivalency and niche similarity using

ecospat (Di Cola *et al.*, 2017). The niche equivalency test evaluates whether the

observed niche overlap is significantly different from a null simulated by randomly

reallocating the occurrences of both entities between their ranges (Broennimann

et al., 2012; Warren *et al.*, 2008). The niche similarity test checks whether the

overlap between two niches is significantly different from the overlap obtained if

random shifts within each environmental space are allowed (Schoener, 1968;

Warren *et al.*, 2008). In both cases, we tested for niche divergence by using the

alternative="lower" option. All tests were based on 100 iterations. In similarity

tests, both ranges were randomly shifted (rand.type=1).

In addition, we modeled the potential range of the four taxa of the Linifolia

clade under present conditions, which was then projected to last interglacial (LIG,

c. 120–140 kya) and last glacial maximum (LGM, c. 21 kya) conditions. To this end,

we used the same six WorldClim bioclimatic variables employed for niche

differentiation analyses, including present layers, LIG layers from Otto-Bliesner *et*

al. (2006) and LGM layers based on the CMIP5 project (three global climate models

(GCMs): CCSM4, MIROC-ESM and MPI-ESM-P). Resolution was 30″ for current and

LIG, and 2.5′ for LGM layers. We performed species distribution modeling (SDM)

using the maximum entropy algorithm, as implemented in Maxent v3.4 (Phillips *et*

al., 2006). We used the same occurrences for each taxon employed for niche

differentiation analyses (see above). 80% of occurrences for each taxon were used

for model training and 20% for model evaluation. For each taxon, ten subsample

replicates were run by randomly partitioning the data into a training set and

evaluation set, and a mean model was calculated. For the LGM, an average of the

three   GCMs   was   calculated.   Logistic   outputs   were   converted   into

presence/absence using the maximum training sensitivity plus specificity logistic

threshold.

Ploidy level estimation

Flow cytometry was used to estimate genome size, explore the possible role of

polyploidization in speciation, and assess the risk of clustering paralogs in RAD-

seq *de novo* clustering. Earlier cytogenetic studies for three *Iberodes* species

showed them as diploids (2n=28 for *I. linifolia*, *I. commutata* and *I. kuzinskyana*

(Franco, 1984; Saly, 1997; Talavera *et al.*, 2012), and aneuploidy was detected for *I.*

littoralis with 2n=24 (Fernández-Casas, 1975). Nevertheless, some of these

references do not cite the information sources. Individuals of the five *Iberodes*

species were cultivated at the glasshouse and five individuals per species were

sampled. In addition, samples of the diploid *Solanum lycopersicum* of known

genome size (2n=24, Rick, 1979; 2C=1.88-2.07 pg, Grandillo *et al.*, 2011) were also

cultivated to compare standardized results with our study species and estimate

genome sizes (Doležel and Bartoš, 2005). We performed a two-step nuclear DNA

Content Analysis using Partec Buffer (de Laat *et al.*, 1987) and DAPI fluorochrome

through a Cell Lab Quanta SC flow cytometer (Beckman Coulter, Fullerton, CA,

USA) equipped with a mercury lamp following the protocol of Doležel et al. (2007).

Samples were visualized through the Cell Lab Quanta SC Software package. FL1

detector (530 nm / 28 nm; DAPI emission maximum=461) and FL1-Area was used

as directly correlated to DNA content. A minimum number of 1000 nuclei for G1

peak were analyzed. Only histograms with a coefficient of variation (CV) lower

than 10% for the G1 peak were accepted.

RESULTS

Phylogenetic analysis

After filtering and processing all samples under the different parameter

combinations in *PyRAD*, 12 matrices of 54 taxa with different numbers of loci and

SNPs were analysed (see Table S4). The same topology was obtained in RAxML

from each of the 12 matrices, differing only in bootstrap support (BS) values for

the nodes (Fig. S1). The highest BS supports and highest number of loci were

obtained for the matrix c95m20 (95% similarity within clusters and 20 minimum

coverage of samples for a final locus). Consequently, this matrix was preferred for

the remaining phylogenetic analyses (HBS matrix). *Iberodes brassicifolia* is the

earliest-diverging lineage, sister to a clade that contains *I. commutata* sister to

another subclade with the remaining species (Fig. 2). Paraphyly was inferred for *I.*

linifolia since a clade including *I. kuzinskyana* and *I. littoralis* is nested within.

Likewise, paraphyly was inferred for *I. littoralis* ssp. *gallaecica* since ssp. *littoralis*

is nested within populations of subsp. *gallaecica* (Fig. 2).

Taxon groups for coalescent-based phylogeny (SVDquartets) were based on

the eight main lineages of RAxML (*Iberodes brassicifolia*, *I. commutata*, populations

of *I. linifolia* from Seville, core clade of *I. linifolia*, populations of *I. linifolia* from SW

Spain, populations of *I. linifolia* from central Portugal, *I. kusinskyana*, and *I.*

littoralis, Fig. S2). The species tree topology was mostly congruent with the one

obtained from the concatenated data-based phylogeny. The only difference was the

placement of *I. linifolia* populations from Seville and SW Spain, which were nested

within the core of *I. linifolia* (Fig. S2). In any case, paraphyly of *I. linifolia* is still

inferred because of the nested position of *I. kuzinskyana* and *I. littoralis* sister to

populations of *I. linifolia* from Portugal (Fig. S2).

Estimate of divergence times

Node ages for main lineages of *Iberodes* are represented in Fig. 2. The stem age

estimated for *Iberodes* was 17.56 myr (17.26-17.86 Bootstrapped Variance, BV)

with a crown age of 5.32 myr (5.32-5.32 BV). Crown ages for *I. brassicifolia* and *I.*

commutata were 1.29 (1.21-1.33 BV) and 2.22 myr (2.17-2.28 BV), respectively.

The crown age of the Linifolia clade (*I. linifolia* s.l., *I. kuzinskyana* and *I. littoralis*)

was inferred in 4.33 myr (4.25-4.59 BV). Divergence time between Portuguese

populations of *I. linifolia* and ancestor of the coastal species (*I. kuzinskyana* and *I.*

littoralis) was 3.67 myr (3.60-3.85 HPD). The origin of *I. littoralis* ssp. *littoralis*,

possibly from an *I. littoralis* ssp. *gallaecica* ancestor is inferred 10,000 years ago

(Fig. 2).

Genetic structure

We set four as the *a priori* number of clusters (k=4) based on the four taxa

circumscribed within the Linifolia clade. DAPC scatterplot revealed two well-

differentiated clusters: *I. linifolia* (LIN) on the one side, and the coastal *I.*

kuzinskiana (KUZ) and *I. littoralis* (LIT_LIT, LIT_GAL) on the other (Fig. 3c,). Within

the coastal species cluster, *I. kuzinskyana* occupied a position closer to *I. linifolia*,

while the two subspecies of *I. littoralis* were intermingled (Fig. 3c). Compoplot

assigned all individuals of *I. linifolia* to a separate cluster, without admixture (Fig.

3c). The other three clusters were shared among coastal species with different

membership probability (Fig. 3c). One of the two individuals of *I. kuzinskyana* was

uniquely assigned to one of the clusters, whereas the other individual showed a

mixed ascription between the three genetic groups (Fig. 3c). Most individuals

(nine) of both subspecies of *I. littoralis* were placed in the same two clusters, while

two of them showed also admixture with the same cluster shared by the two

samples of *I. kuzinskyana* (Fig. 3c). Given the interdigitated pattern of the two

subspecies of *I. littoralis*, we repeated the analysis by considering the two

subspecies as part of a single group (K=3) to evaluate the genetic clustering of *I.*

littoralis as a genetic group. Compoplot for K=3 assigned most individuals (nine) of

I. littoralis to the same cluster, while two samples had mixture assignment

probability to the cluster of *I. kuzinskyana* (Fig. S3). Likewise, AMOVA of the two

groups (*I. linifolia* vs coastal species) showed significant among-group variance

(58.88%) higher than within-population variance (38.09%) or variance within

populations among groups (3.03%), with a fixation index of F_{ST}= 0.619 (Table 1).

D-statistic tests did not conclusively support a role of historical introgression in

determining paraphyletic pattern since ancient introgression was inferred

indiscriminately among lineages of *I. linifolia* and both coastland species (see

Methods S1).

Morphometric analysis

Principal Component Analysis (PCA) revealed morphological differentiation

between the three taxa. *I. linifolia* held the greatest morphological variation

ranging from the two extremes of both PC1 and PC2 compared to the more

restricted variation of coastal species that is concentrated in higher scores of PC1

and lower of PC2. Three species of the Linifolia clade were almost completely

differentiated, which supports the previous taxonomic observations. *I. linifolia* and

I. littoralis occupied the two extremes of variation ranging from higher length of

vegetative characters, higher nutlet sizes and nutlet teeth for *I. linifolia* to higher

length and density of nutlet trichomes and presence of bracts for *I. littoralis* (Fig.

3a). *Iberodes kuzinskyana* occupied an intermediate position between *I. littoralis*

and *I. linifolia*, overlapping very slightly with the latter. The two subspecies of *I.*

littoralis were intermingled at the highest scores of PC1 and the lowest of PC2 (Fig.

3a).

Climatic niche differentiation and distribution modelling

The two main principal components of the 6 climatic variables explained 73.59%

(PC1=48.86% and PC2=24.73%) of the climatic variability in the study region. The

variables that contributed most to PC1 were: bio1 (annual mean temperature),

bio8 (mean temperature of wettest quarter), bio12 (annual precipitation), and

bio14 (precipitation of the driest month). Bio3 (isothermality) and bio4 (temperature seasonality) were the two variables most related to PC2. The visualization of the e-space showed little overlap among taxa (Fig. 3b). The geographically widespread *Iberodes linifolia* showed the widest niche, whereas the more narrowly distributed coastal species are more restricted in climatic niche space (Fig. 3b). Among coastal species, *I. kuzinskyana* differs from *I. littoralis* in PC1 (less precipitation and higher temperatures for *I. kuzinskyana*) and overlaps with *I. littoralis* ssp. *gallaecica* but not with *I. littoralis* ssp. *littoralis* in PC2 (higher isothermality in *I. kuzinskyana* and *I. littoralis* ssp. *gallaecica*). The two subspecies of *I. littoralis* have well-differentiated niches particularly along PC2. *I. kuzinskyana* occupies an intermediate position between *I. linifolia* and *I. littoralis* subsp. *gallaecica*. Thus, *I. kuzinskyana* marginally overlapped with *I. linifolia* for those parts of the niche of *I. linifolia* with higher isothermality, lower temperature and higher precipitation. Indeed, minimum niche overlap was inferred between both subspecies of *I. littoralis* and *I. linifolia* due to lower temperature and higher precipitation for coastal taxa. Consequently, although tests of similarity indicated that observed values of niche overlap are not significantly lower than random overlap, all pairwise equivalency tests significantly rejected the equivalency of niches and this is supported by the low values of D obtained (Table 2).

The potential ranges for the species of the Linifolia clade in the present are broadly congruent with known geographic distributions, with overlapping distributions of *I. linifolia* and *I. kuzinskyana* (Fig. 4). In contrast, a contraction of distribution ranges was inferred for the LIG, including the disappearance of the potential range of *I. littoralis* ssp. *gallaecica* (Fig. 4), but maintaining the overlap and absence of physical barriers between *I. linifolia* and *I. kuzinskyana*. Projections

to the LGM showed a range similar to the present one for *I. linifolia* and expanded

ranges for both *I. kuzinskyana* and *I. littoralis* subsp. *gallaecica*. The range of *I.*

littoralis subsp. *gallaecica* is inferred to have occupied most of the current range of

both subspecies in the LGM, before subsp. *littoralis* differentiated from subsp.

gallaecica.

Ploidy level estimation

Mean peaks of fluorescence, CV values, and genome sizes obtained for each species

are summarized in Fig. S4 and Table S5. Fluorescence peaks were similar among all

Iberodes species, ranging from 45.2 to 62.6 nm for G1. No evidence of duplicated

DNA content was observed, suggesting no ploidy differences among taxa or

populations. Likewise, similar genome sizes were obtained for the five species

ranging from 1.788 to 1.980 pg. *I. kuzinskyana* and *I. littoralis* ssp. *littoralis* were

the *Iberodes* species with the largest and the smallest genome sizes, respectively.

The five species are similar in genome size to the *S. lycopersicum* standard (1.88-

2.07 pg, Grandillo *et al.*, 2011).

DISCUSSION

Phylogenetic relationships among *Iberodes* species are robustly inferred by

RAD-sequencing, which unveiled the monophyly of populations into species except

for *I. linifolia*. Two early-diverging monophyletic species (*I. brassicifolia* and *I.*

commutata) contrast with paraphyly in the Linifolia clade of three species (Fig. 1).

The paraphyly of *I. linifolia* with respect to *I. kuzinskyana* and *I. littoralis* suggests a

progenitor-derivative relationship between the species, with the widespread

species giving rise to the coastal endemics in the course of a northward

colonization from the coast of Portugal to the west coast of France. Our results

provide a multiple lines of evidence that support parapatric divergence of the

coastland lineage (*I. kuzinskyana-I. littoralis* clade): (1) lack of any physical barrier

to gene flow between inland and coastland in the present or the past; (2) low level

of current gene flow (despite historical introgression) that support budding

differentiation rather secondary contact; (3) recent niche differentiation from

inland habitats with higher temperature seasonality to coastland ones with higher

precipitation; (4) paraphyly of *I. linifolia*, which can be interpreted as the extant

inland ancestor of the coastland lineage; and (5) incipient morphological

differentiation between *I. linifolia* and the coastal lineage. Subsequently,

differentiation in allopatry of the two coastal species during the mid-Pleistocene

led to the modern-day species *I. kuzinskyana* and *I. littoralis*, most likely mediated

by geographic isolation and niche differentiation. Thus ecological and geographic

differentiation contributed to both parapatric and allopatric speciation in the

clade.

A predominant paraphyletic pattern during *Iberodes* evolution

Our RAD-seq analyses revealed a pattern of predominant paraphyly within the

recently diverged Linifolia clade (Fig. 2). While the populations of *I. brassicifolia*

and those of *I. commutata* form monophyletic groups, paraphyly is inferred for *I.*

linifolia populations with respect to the two coastal species. While apparent

paraphyly of species may be a consequence of introgression dragging populations

toward the species to which they have introgressed (e.g., Eaton *et al.*, 2015), our

analyses of historical introgression do not conclusively support a role of this

process in determining the paraphyletic topology (Methods S1) and genetic

structure shows no evidence of recent admixture involving *I. linifolia* (Fig. 3c).

Despite the tendency in taxonomy to only consider monophyletic species,

paraphyletic ones have been reported to be particularly common in groups

showing a recent local speciation pattern, i.e. groups composed of a 'mother'

species that coexists in time and space with more restricted derivative species

(Rieseberg and Brouillet, 1994).

Recent ecological speciation in parapatric *Iberodes*

Although parapatry has been argued to be common in plants (Rieseberg and

Brouillet, 1994), few empirical examples are found on the literature (Table 3). Our

study points to demonstrate parapatry through a process of budding (or

progenitor-derivative) speciation between *I. linifolia* and the ancestor of *I.*

kuzinskyana and *I. littoralis*. Indeed, the projection of distribution models to past

climatic periods during the Quaternary revealed a similar or greater degree of

geographic overlap between the potential ranges of *I. linifolia* and *I. kuzinskyana*

(Fig. 4), which makes allopatry unlikely (Zheng *et al.*, 2017). Peripheral

populations of the widespread *I. linifolia* close to the coast of Portugal are the most

closely related and possible living progenitor of coastland species (*I. kuzinskyana*

and *I. littoralis*; Fig. 2), which seem to have differentiated in an adjacent area with

no apparent physical barrier. Congruently, paraphyly underlies such budding

speciation event (Stuessy and Hörandl, 2013). In addition, despite the fact that

historical introgression was inferred among *I. linifolia* and coastland species

(Methods S1), it does not seem to underlie the phylogenetic gradation since

introgression is found indiscriminately with all lineages of *I. linifolia*. Moreover,

our results of AMOVA and DAPC pointed to lack of recent gene flow between the

nearby species (*I. kuzinskyana* and *I. linifolia*). This pattern supports strong

isolation rather than secondary contacts as suggested in other cases of ecological

differentiation (e.g. *Mimulus*, Stankowski *et al.*, 2015; *Stauracanthus*, Chozas *et al.*,

2017; Table 3). Nevertheless, a deeper population genetics study focused on the

adjacent populations of Portugal is necessary to obtain more detailed estimates of

gene flow.

In the earliest stages of parapatric speciation mediated by ecological

differentiation, reproductive barriers are expected to evolve in response to

selection towards locally adapted, divergent ecotypes (Sakaguchi *et al.*, 2019). The

niche identity tests points to a role of ecologically-based selection in the speciation

of the coastal lineage from *I. linifolia.* The coastal lineage is distributed in niches

with less temperature seasonality and higher precipitation means than *I. linifolia*

(Fig. 3b). Interestingly, although niches are not equivalent and overlap is close to

zero, similarity tests indicated a certain degree of similarity within the E-space,

which points to recent divergence and suggests ongoing differentiation or some

degree of niche conservatism. Besides, edaphic factors such as the specialization to

coastal dunes substrates can have also contributed to ecological differentiation,

although this is a fine-scale soil variable that is difficult to include in niche models.

The differentiation of coastal lineages (*I. kuzinskyana* and *I. littoralis*) from inland

populations of *I. linifolia* seems to have started in western Iberian Peninsula during

the Mid Pliocene (3.85-3.60 myr, Fig. 2), when increasing seasonality led to the

origin of Mediterranean climate with its summer droughts (Suc, 1984). The onset

of Mediterranean climate favored the differentiation and configuration of modern

Mediterranean flora (Vargas *et al.*, 2018), which agrees with recent *Iberodes*

speciation (Fig. 2). Posterior glacial/interglacial periods during the Late Pliocene

promoted the expansion and contraction of geographic ranges and the formation

of sources and sinks of genetic diversity for most Mediterranean species (Médail

and Diadema, 2009). Interestingly, the overlapping area inferred for *I. linifolia* and

I. kuzinskyana in the past is congruent with the location of glacial refugia

(Estremadura and Beira litoral) proposed by Médail and Diadema (2009) and plant

species previously studied as glacial refugial species (e.g. *Drosophyllum*

lusitanicum, Müller and Deil, 2001; *Cistus monspeliensis*, Fernández-Mazuecos and

Vargas, 2010; *Linaria elegans*, Fernández-Mazuecos and Vargas, 2013).

In addition, ecological differentiation in parapatric speciation involves

divergent natural selection towards contrasting environments that promotes

adaptation to different environments and subsequent barriers to gene flow

(Rundle and Nosil, 2005; Schluter, 2001). The strength of divergent selection can

modulate the degree of completeness of a speciation process (Nosil *et al.*, 2009). In

our case, both the high interspecific genetic differentiation and clear

morphological and ecological divergence (Fig. 3) between inland and coastal

species may reflect strong divergent selection promoting rapid speciation

(Pliocene-Pleistocene, Fig. 2). Indeed, our study reinforces the common trend for

Mediterranean plants in which narrow endemics arise from ecological

differentiation of peripheral populations of more widespread progenitor species

(Papuga *et al.*, 2018). Moreover, this trend highlights that this peripheral

populations may set the scene for diversification, thus revealing their conservation

value (Debussche and Thompson, 2003).

Allopatric differentiation along the Atlantic coast

An allopatric pattern is also found in the Linifolia clade. Given the current

separation between ranges of the three coastal taxa (Fig. 1), allopatry seems seems

to have triggered speciation process. On the one hand, the widely separated ranges

of *I. kuzinskyana* and *I. littoralis* subsp. *gallaecica* both in current times and in

projections to LIG (despite probable intermittent contact during LGM) suggest

either a colonization mediated by LDD followed by geographic isolation and niche

shift (Fig. 3, 4) or a vicariant process mediated by an early expansion and

contraction of distribution ranges. We find similar examples of recent allopatric

speciation for both hypothesis: by LDD among coastal Mediterranean flora in the

same period (e.g. Jakob *et al.*, 2007; Carnicero *et al.*, 2017; Herrando-Moraira *et al.*,

2017); and expansion/contraction of distribution ranges (e.g. Ortiz *et al.*, 2007; Lo

Presti and Oberprieler, 2011; Fernández-Mazuecos and Vargas, 2011). On the

other hand, the allopatric differentiation of the two subspecies of *I. littoralis* was

likely preceded by an expansion of *I. littoralis* during the LGM, followed by

geographic isolation as its range contracted and niche shifted, leading to the

differentiation of *I. littoralis* subsp. *littoralis* in more recent times (Fig. 3, 4). In this

case, the probable role of long distance dispersal (LDD) is further supported by the

fact that the expanded area during the LGM does not seem to have formed a

continuous range (Fig. 4). Indeed, unlike inland species of *Iberodes*, the three

coastal taxa seem to have LDD specializations in the form of uncinated hooks (Fig.

1) in the nutlet, related to the attachment to feathers and fur in other closely

related Boraginaceae genera (Selvi *et al.*, 2011).

CONCLUSIONS

Our integrative approach has let us overcome previous barriers to understanding

parapatric speciation. The combination RAD-sequencing, morphometry and

climatic niche has provided notable evidences of parapatry revealing this

workflow as a powerful resource for scenarios of non-allopatric processes in other

groups of plants. In particular, RAD-sequencing once again is revealed as a useful

tool to disentangle recent speciation events in non-model organisms.

Consequently, we have empirically shown the recurrence of progenitor-derivative

speciation in plants as previously suggested. Processes such us budding speciation

bring to light the need to extend traditional species concepts and advocate the

consideration of paraphyletic species as natural units of taxonomical classification.

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AUTHOR CONTRIBUTIONS

627   AO, PJM, VV and PV contributed to developing the question and experimental

628   design. AO led the material collection. AO, MFM and ALH conducted the data

629   analysis. AO wrote the manuscript with the contribution of all authors in both

630   interpretation and writing.

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FIGURE LEGENDS

Fig. 1 Locations of the *Iberodes* species sampled for the different analyses

performed. The star symbols indicate locations for RAD-sequencing, squares

represent the locations for the morphometric analysis, and circles indicate

locations for the ecological niche modelling. Color legend for each taxa of *Iberodes*

is shown in the figure. Images of the six taxa are framed following each taxa color

in the map.

Fig. 2 Time-calibrated phylogeny obtained from TreePL by using maximum clade

credibility from 1000 bootstrap trees of the maximum likelihood analysis of

c95m20 matrix. Branch thickness indicates the ML bootstrap supports above 98.

Node bars in blue indicate the node age ranges taking into account the branch

length variance along the 1000 bootstrap trees. No branch length variation was

inferred for nodes without blue bar. The international stratigraphic scale is

included from 26 myr until present. Text codes for each individual location are

detailed in the Table S1. The color for each lineage of the Linifolia clade is

geographycally represented through the colored ellipses of the map. Grey ellipses

group the different lineages of the four taxa within the Linifolia clade.

Fig. 3 Morphometric, niche overlap and genetic clustering analyses of the Linifolia

clade. Codes for each taxa are: LIN for *I. linifolia*, KUZ for *I. kuzinskyana*, LIT-GAL

for *I. littoralis* ssp. *gallaecica*, and LIT-LIT for *I. littoralis* ssp. *littoralis*. (a)

Morphometric analysis. Principal Component Analaysis (PCA) of the 12

morphologic chartacters: LS, length of the stem; LL, length of the leaf; WL, wide of

the leaf; PINFL, pedicel of inflorescence; LCFR, length of fruit calix; DN, diameter of

the nutlet; LN, length of the nutlet; LM, length of the margin nutlet; LT, length of

the margin teeth; DH, density of trichomes; LH, length of trichome; BR: absence=0,

presence=1 of flowering bracts). The contribution of each trait to the two principal

components (PC1, PC2) as well as the positive or negative sense of the relationship

is represented through the length of each arrow and the directionality of the

arrow, respectively. The percentages of the variability explained by the two PCs

are indicated close to the axes. (b) Niche overlap analysis. PCA of the six climatic

variables obtained from Worldclim. BIO1, Annual Mean Temperature; BIO3,

Isothermality; BIO4, Temperature Seasonality; BIO8, Mean Temperature of

Wettest Quarter; BIO12, Annual Precipitation; BIO14, Precipitation of Driest

Month. The strength of contribution of each variable to PCs and the sense of the

relationship is represented, as explained above, through the length and the

directionality of the arrows. The percentages of the variability explained by the

two PCs are indicated close to the axes. Values of the variables are also

represented for each PC separately following Fernández-López and Villa-Machío

(2017). (c) Genetic clustering. Scatterplot from the DAPC analysis showing the

genetic differentiation retained in one PC as the optimal number of PC for the

discriminant function. Compoplot for K=4 is also represented indicating

membership probability to each genetic cluster for each taxa.

Fig. 4 Species distribution modelling obtained through the entropy algorithm, as

implemented in Maxent for the four taxa of the Linifolia clade. Codes for each taxa

are: LIN for *I. linifolia*, KUZ for *I. kuzinskyana*, and LIT for *I. littoralis* (LIT-GAL for *I.*

littoralis ssp. *gallaecica*, LIT-LIT for *I. littoralis* ssp. *littoralis*). Analyses are based

on WorldClim and projected to past conditions: last interglacial (LIG, c. 120–140

kya) and last glacial maximum (LGM, c. 21 kya). The presence/absence was

estimated using the maximum training sensitivity plus specificity logistic

threshold. The projection to the past is only presented according to the inferred

times of divergence of each taxa.

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1075   **TABLES**

1076   **Table 1** Analysis of molecular variance (AMOVA) results for individuals from two

1077   contrasted groups: (1) mainland, *I. linifolia* and (2) coastal, *I. kuzinskyana* and *I.*

1078   *littoralis.*

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Source of Varitation	d.f.	Sum of squares	Variance components	Percentage of variation
Among groups	1	1602.666	106.33509 Va	58.88
Among populations within groups	2	181.372	5.47321 Vb	3.03
Within populations	24	1651.033	68.79306 Vc	38.09
Total	27	3435.071	180.60135	

1080   Note: Variance components are as in Excoffier *et al.* (1992). F_{st} = 0.61909; F_{sc} = 0.07370; F_{ct}
1081   = 0.58878.

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1083   **Table 2** Pairwise statistical test for comparison of ecological niche overlap

1084   between the different taxa within the Linifolia clade. The statistical significance is

1085   represented by p-values (Warren *et al.*, 2008). Asterisk (*) indicate significant

1086   values for *p*-values < 0.01.

Taxa comparison	Schoener's D	Niche equivalency p-value	Niche similarity p-value
I. linifolia vs *I. kuzinskyana*	0.026	0.0099*	0.94059
I. linifolia vs *I. littoralis* ssp. *gallaecica*	0.003	0.003	0.60396
I. linifolia vs *I. littoralis* ssp. *littoralis*	0.004	0.0099*	0.69307
I. kuzinskyana vs *I. littoralis* ssp. *gallaecica*	0.000	0.0099*	0.64356
I. kuzinskyana vs *I. littoralis* ssp. *littoralis*	0.000	0.0099*	0.71287
I. littoralis ssp. *gallaecica* vs *I. littoralis* ssp. *littoralis*	0.000	0.0099*	0.67327

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1088   **Table 3** Examples of parapatric speciation in plants.

Genus	Family	Event suggested	Reference
Ephedra	Ephedraceae	Ecological divergence mediated by bioclimatic niche	Yin *et al.* (2016)
Fosterella	Bromeliaceae	Ecological divergence mediated by polyploidization and	Paule *et al.* (2017)
Helianthus	Asteraceae	Ecological divergence mediate by bioclimatic niche	Andrew & Rieseberg (2013)

Mimulus	Phrymaceae	Ecological divergence mediated by pollinator shift	Sobel & Streisfeld (2015), Stankowski *et al.* (2015)
Miscanthus	Poaceae	Ecological divergence mediated by bioclimatic niche	Huang *et al.* (2014)
Oryza	Poaceae	Ecological divergence mediated by bioclimatic niche	Zheng & Ge (2010)
Populus	Salicaceae	Ecological divergence mediated by bioclimatic niche	Zheng *et al.* (2017)
Roscoea	Zingiberaceae	Ecological divergence mediated by bioclimatic niche	Zhao *et al.* (2016)
Senecio	Asteraceae	Ecological divergence mediated by ecological niche	Richards and Ortiz-Barrientos (2016)
Stauracanthus	Fabaceae	Ecological divergence mediated by bioclimatic niche	Chozas *et al.* (2017)
Columnea	Gesneriaceae	Ecological divergence mediated by bioclimatic niche and pollinator shift	Schulte *et al.* (2015)

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SUPPORTING INFORMATION

Additional Supporting Information may be found online in the Supporting

Information section at the end of the article.

Fig. S1 RAxML trees of *Iberodes* using three different similarity percentages (85%,
90%, 95%) and four levels of minium coverage (m4, m12, m20, m28). Bootstrap
support are indicated when is lower than 100.

Fig. S2. SVDquartets species tree of *Iberodes* using the eight main lineages obtained in the RAxML analyses.

Fig. S3 Compoplot showing individual assignment probability to different species-groups of the Linofolia clade of *Iberodes*, considering the two subspecies of *I. littoralis* as one group (k=3). Codes for each taxa are: LIN for *I. linifolia*, KUZ for *I. kuzinskyana*, LIT-GAL for *I. littoralis* ssp. *gallaecica*, and LIT-LIT for *I. littoralis* ssp. *littoralis*.

Fig. S4 Flow cytometry peaks for each of five samples used for each of the five species of *Iberodes* (including the two subspecies of *I. littoralis*. Peak of the standard *Solanum lycopersicum* is included. FL1 in the X axis indicates the signal intensity (530 nm / 28 nm; DAPI emission maximum=461). Y axis indicates the number of events found. Mean and median values of FL and percentage of variation coefficient is shown for each colored peak.

Table S1 Data information and Genbank numbers of all individuals sampled for RADseq.

Table S2 Morphological characters analyzed. Values represent the mean of two individuals measured per herbarium sheet. LS: length of the stem, LL: length of the leaf, WL: wide of the leaf, PINFL: pedicel of inflorescence, LCFR: length of fruit calix, DN: diameter of the nutlet, LN: length of the nutlet, LM: length of the margin nutlet, LT: length of the margin teeth, DH: density of the hair, LH: length of trichome.

Table S3 Data points for environmental analysis

Table S4 Number of loci, SNPs retrieved for each of the 12 parameter combinations. Parameter abreviations indicate: c, clustering threshold; m, minimum taxon coverage to consider a locus.

Table S5 Genome size estimation for the five species of *Iberodes* (including the two subspecies of *I. littoralis*.

Methods S1 Introgression analysis.

Taxa

- 🟥 *I. brassicifolia*
- 🟨 *I. commutata*
- 🟩 *I. linifolia*
- 🟥 *I. kuzinskyana*
- 🟦 *I. littoralis* ssp. *gallaecica*
- 🟪 *I. littoralis* ssp. *littoralis*

Analyses

- ☆ RAD sequencing
- □ Morphometry
- ○ Ecological niche

LINIFOLIA CLADE

Outgroup

I. brassicifolia

I. commutata

I. littoralis ssp. *littoralis*

I. littoralis ssp. *gallaecica*

I. kuzinskyana

I. linifolia

I.linifolia SEV 2
I.linifolia SEV 1
I.linifolia BUR 2
I.linifolia JAE 1
I.linifolia MAD 15
I.linifolia MAD 16
I.linifolia ZAR 2 2
I.linifolia ZAR 2 3
I.linifolia ZAR 1 3
I.linifolia ZAR 1 1
I.linifolia BAD 1
I.linifolia BAD 2
I.linifolia BAD 3
I.linifolia SET 2
I.linifolia LIS 2
I.kuzinskyana LIS 2 1
I.kuzinskyana LIS 1 1
I.littoralis ssp. gallaecica XUN 1
I.littoralis ssp. gallaecica XUN 2
I.littoralis ssp. gallaecica PC 1
I.littoralis ssp. gallaecica DON 1
I.littoralis ssp. gallaecica DON 2
I.littoralis ssp. gallaecica BAL 1
I.littoralis ssp. littoralis POI 1
I.littoralis ssp. littoralis POI 2
I.littoralis ssp. littoralis LOI 1
I.littoralis ssp. littoralis LOI 2
I.littoralis ssp. littoralis BRE 1

Oligocene	Miocene		Pliocene	Pleistocene	Holocene
Paleogene	Neogene			Quaternary	

20   10   0 (myr)

(a) Morphometric analysis

(b) Niche overlap

(c) Genetic clustering