

1 **Main title: The short-term high fat diet-induced increase in %5-methylcytosine**
2 **expression in peripheral blood T lymphocytes, is attenuated by low-dose aspirin.**

3

4

5 Tinashe Mutize¹, Phiwayinkosi V. Dlodla^{2,3}, Zibusiso Mkandla¹, Bongani B. Nkambule¹

6

7 ¹School of Laboratory Medicine and Medical Sciences (SLMMS), College of Health

8 Sciences, University of KwaZulu-Natal, Durban, South Africa.

9 ²Biomedical Research and Innovation Platform, South African Medical Research Council,

10 Tygerberg, South Africa.

11 ³Department of Life and Environmental Sciences, Polytechnic University of Marche, Ancona,

12 Italy.

13

14 Tinashe Mutize: E-mail address: 217063119@stu.ukzn.ac.za. Tel.: +27 84 053 2069.

15 Zibusiso Mkandla: Email address: 217063126@stu.ukzn.ac.za, Tel.: +27 79 273 1696

16 Bongani B. Nkambule: Email address: nkambuleb@ukzn.ac.za, Tel.: +27 31 260 8964

17 Phiwayinkosi V. Dlodla: pdlodla@mrc.ac.za. Tel.: +27 219380333; fax: +27 219380456.

18

19 **Corresponding author:** Bongani B. Nkambule: Email address: nkambuleb@ukzn.ac.za,

20 Tel.: +27 31 260 8964

21

22

23

24

25

26 Abstract

27 **Objective:**

28 To assess peripheral lymphocyte DNA methylation profiles in prediabetes using a high fat-
29 diet-fed C57BL/6 animal model. We further evaluated whether low dose-aspirin, or low-dose
30 aspirin in combination with metformin, could modulate global DNA methylation levels in
31 peripheral blood lymphocytes.

32

33 **Methods**

34 Twenty-eight (28) male C57BL/6 mice were used in two experimental phases. The first
35 experiment involved animals (n=16) which were randomised to receive a low-fat diet (LFD) or
36 high-fat diet (HFD) (n = 8/group) for 10 weeks. Whereas in the second experiment, HFD-fed
37 mice (n=15) were randomised into 3 treatment groups, a low-dose aspirin (LDA), LDA and
38 metformin group, and a clopidogrel group. DNA methylation profiles of were determined using
39 flow cytometry.

40 **Results**

41 The HFD group showed moderate weight gain and elevated postprandial blood glucose levels
42 when compared to the LFD group after 2 weeks of HFD-feeding ($p < 0.05$). Interestingly, the
43 HFD group had elevated levels of T cells expressing high levels %5-methylcytosine ($p < 0, 05$).
44 Notably, these elevated levels were lowered by short-term low-dose aspirin treatment.

45 **Discussion**

46 T cells are involved in the propagation of the inflammatory response. Persistent T cell
47 activation promotes chronic inflammation and insulin resistance. Low-dose aspirin may be
48 effective in modulating T cell-specific global methylation.

49

50 **Keywords:** Global DNA methylation, type 2 diabetes mellitus, prediabetes, anti-
51 hyperglycaemic, anti-coagulant, anti-inflammatory.

52

53 Introduction

54

55 The global prevalence of type 2 diabetes (T2DM) continues to increase with several studies
56 linking aberrant DNA methylation with the development of these metabolic disease (Van
57 Otterdijk et al., 2017; Zhang et al., 2017; Shah et al., 2019; Wittenbecher et al., 2019). Aberrant
58 DNA methylation profiles are associated with an increased risk of developing T2DM (Zhou et
59 al., 2018; Zhang et al., 2018). Differential DNA methylation has been reported in patients with
60 T2DM (Kuroda et al., 2009; Nilsson et al., 2014; Dayeh et al., 2014; Zhang et al., 2018). DNA
61 hypermethylation (Ban et al., 2002; Yang et al., 2011; Gu et al., 2013; Zou et al., 2013; Seman
62 et al., 2015; Bacos et al., 2016) and DNA hypomethylation (Zhang et al., 2014; Nilsson et al.,
63 2015) levels have been reported in T2DM. Moreover, increased levels of global DNA
64 methylation were associated with insulin resistance (Zhao et al., 2012) and in another study,
65 aberrant DNA methylation was reported in women with obesity-related systemic insulin
66 resistance (Zhang et al., 2018). Hypermethylation on the insulin promoter in type 2 diabetic
67 patients is also associated with the downregulation of the insulin gene in human pancreatic
68 islets (Yang et al., 2011). Furthermore, Kuroda *et al.*, also reported that the expression of the
69 insulin gene was regulated by differential DNA methylation (Kuroda et al., 2009).

70 Several studies have associated dietary patterns with DNA methylation profiles studies and
71 obesity-related conditions such as chronic inflammation, insulin resistance, T2DM and
72 subsequently cardiovascular disease (Mckay & Mathers, 2011; Park et al., 2017; Milagro et
73 al., 2013).

74 Chronic inflammation promotes insulin resistance and the development of type 2 diabetes
75 (Goldfine et al., 2011). Chronic inflammation also provides a link between type 2 diabetes and
76 cardiovascular disease (Goldfine et al., 2011). Previous studies have shown that differential
77 methylation in adipose tissues, muscles and pancreatic islets are associated with localised
78 inflammation in patients with T2DM (Kulkarni et al., 2012; Yang et al., 2012; Ribel-Madsen et
79 al., 2012; Nilsson et al., 2014; Dayeh et al., 2014).

80 Low-dose aspirin (LDA) is widely used for the primary prevention of cardiovascular events in
81 patients with type 2 diabetes who are at an increased risk of developing thrombotic
82 complications (Goldfine et al., 2011). Furthermore, the use of low-dose aspirin as a primary
83 preventive measure for patients with type 2 diabetes and heart failure (HF) has been reported
84 to be associated with lower all-cause mortality, though in the absence of other
85 contraindications including a history of myocardial infarction, stroke, coronary artery disease,
86 peripheral artery disease (Abi Khalil et al., 2018) (Chang et al., 2013). Furthermore, it has
87 been suggested that an increased dose of aspirin or twice-daily low-dose aspirin therapy could
88 be possible therapeutic options for cardiovascular prevention in diabetes mellitus patients
89 (Capodanno & Angiolillo, 2016). The combination of aspirin with a potent anti-platelet drug
90 such as clopidogrel is an antiplatelet therapy option of choice in the prevention of
91 macrovascular conditions (Silber et al., 2015; Smith et al., 2006; Anderson et al., 2007; King
92 et al., 2008; Werf et al., 2008; Kushner et al., 2009). Notably, the occurrence of cardiovascular
93 events in coronary artery disease patients receiving dual clopidogrel and low-dose aspirin
94 therapy is associated with a poor response to clopidogrel (Kuliczkowski et al., 2009). The poor
95 response and variability in clopidogrel action, as well as the recurrence of ischemic events in
96 patients with stroke, has been attributed to differential DNA methylation (Gallego-Fabrega et
97 al., 2016). In fact, hypomethylation of the ATP Binding Cassette Subfamily B Member 1
98 (ABCB1) gene promoter has been reported to be associated with a decreased clopidogrel
99 response in ischemic stroke patients via increased ABCB1 mRNA expression (Yang et al.,
100 2015).

101 In this study, we hypothesized that low dose aspirin (LDA) as a monotherapy and in
102 combination with metformin (LDA + Metformin) may induce differential global DNA methylation
103 in B and T lymphocytes (Nishimura et al., 2009; DeFuria et al., 2013). The study aimed to
104 assess the global DNA methylation profiles of the lymphoid cell subsets using a HFD-fed
105 animal model of prediabetes. We further assessed whether low-dose aspirin and clopidogrel
106 modulate the global DNA methylation profiles of peripheral blood lymphocytes.

107 **Methods and Materials**

108 **Animals and animal handling**

109 Male C57BL/6 mice (n = 28) at 6 weeks of age were purchased from the Biomedical Resource
110 Unit (BRU) at the University of KwaZulu-Natal (UKZN). The C57BL/6 mice strain is well
111 characterized and has been shown to become glucose intolerant when kept on a high-fat diet
112 (Pinchuk & Filipov, 2008). The mice were housed in cages at the BRU in a controlled 12-hour
113 light/dark cycle and a temperature range of 23 - 25 °C (relative humidity: approximately 50 %).
114 The animal well-being was monitored in accordance with the principles of laboratory animal
115 care (National Institute of Health publication 80-23, revised 1978). The mice were allowed
116 free access to water throughout the experimental period. The animal study followed the Animal
117 Research: Reporting In Vivo Experiments (ARRIVE) (Kilkenny et al., 2013) (Supplementary
118 File 1). The ARRIVE guideline was used to improve the standard of reporting in animal
119 research. The study received ethical approval from the University of KwaZulu-Natal Animal
120 Research Ethics Committee (AREC), under the ethics registration number AREC/086/016.

121 **Study design and experimental procedures**

122 The study comprised of two major experiments (Figure 1). The first experiment comprised of
123 16 male mice which were randomized into two diet groups, the low-fat diet (LFD) and the high-
124 fat diet (HFD) group (table 1). The LFD group received a low-fat diet (D12450J) (Research
125 Diets, New Brunswick, NJ, USA) containing 10 % kcal fat, 20 % kcal Protein, 70 % kcal
126 carbohydrates and 3.82 kcal/g energy density, whereas the HFD group (D12492) (Research
127 Diets, New Brunswick, NJ, USA) received a high-fat diet containing 60 % kcal fat, 20 % kcal
128 protein, 20 % kcal carbohydrates and 5.21 kcal/g energy density).

129 Table 1. Diet composition (g/kg)

Ingredients	LFD^a	HFD^b
Casein, 30 mesh	200.00	200.00
L-Cystine	3.00	3.00
Corn starch	506.20	-

Lodex 10	125.00	125.00
Sucrose	72.80	72.80
Solka Floc, FCC200 (Fiber)	50.00	50.00
Soybean Oil	25.00	25.00
Lard	20.00	245.00
Mineral mix S10026B	50.00	50.00
Choline Bitartrate	2.00	2.00
Vitamin mix V10001C	1.00	1.00
Dye, Yellow FD&C #5, Alum. Lake 35-42%	0.04	-
Dye, Blue FD&C #1, Alum. Lake 35-42%	0.01	0.05

130 ^a The LFD obtained from Research Diets Inc (#D12450J, rodent diet with 10% kcal% fat) provided 3.82
131 kcal/g from 20%, 70%, and 10% of protein, carbohydrate, and fat, respectively.

132 ^b The HFD obtained from Research Diets Inc (#D12492, rodent diet with 60% kcal% fat) provided 5.21
133 kcal/g from 26.2%, 26.3%, and 34.9% of protein, carbohydrate, and fat, respectively.

134 ^c Typical analysis of cholesterol in lard = 0.72 g/kg.

135

136 **Experiment one:**

137 The first experiment aimed at measuring the baseline DNA methylation profiles of isolated B
138 and T lymphocytes, following 8-weeks of high fat-diet feeding. The study comprised of 5-week
139 old male C57BL/6 mice (n = 16). The mice were allowed a week for acclimatisation while
140 receiving normal mice chow and had free access to water *ad libitum*. The mice were then
141 randomised into two groups, the LFD and HFD group (n = 8/group). The animals were then
142 housed in separate cages (n = 8/cage) based on their respective diets (figure 1). Two-hundred
143 microliters of venous blood was drawn from each animal and baseline haematological
144 measurements, glucose, insulin and DNA methylation measurements (%5-methylcytosine)
145 were determined (Supplementary file 2). Monocyte and granulocytes were depleted from the
146 whole blood samples and B and T lymphocyte were then isolated using the BD™ IMag Cell
147 Separation system (BD Bioscience,USA) (Supplementary file 2).

148

149 **Experiment two:**

150 In experiment two we assessed whether the short-term treatment with low-dose aspirin (LDA),
151 LDA with metformin (LDA+MET), and clopidogrel modulates DNA methylation profiles in HFD-
152 fed (HFF) mice. Fifteen (n=15) HFF mice were randomized into three treatment groups (n =
153 5/group). These included; (1) a low-dose aspirin (LDA) (3 mg/kg) (Kroesen et al., 2018) group;
154 (2) LDA and metformin (150 mg/kg) (Saisho, 2015) group and clopidogrel (0.25 mg/kg)
155 (Iannacone et al., 2007). The mice received their respective treatment for 6 weeks via daily
156 oral gavage (figure 1). Blood was then drawn and the serum insulin levels, haematological
157 indices, DNA methylation profiles were determined (Supplementary file 2).

158

159

160 **Statistical analysis**

161

162 All statistical analysis was performed using GraphPad Prism 5 (GraphPad Software Inc.; La
163 Jolla, CA, USA). The comparisons of baseline measurements between the two diets groups
164 the LFD and HFD were performed using an unpaired t-test for parametric data and reported
165 as mean and standard deviation. In addition, the one-way analysis of variance (ANOVA) was
166 used for comparisons of DNA methylation levels across the three treatment groups, followed
167 by the Bonferroni post-hoc test. While non-parametric data was analysed using the Mann
168 Whitney U test and reported as median and interquartile range. For comparisons across the
169 three treatment groups the Kruskal-Wallis test followed by the Dunn's multiple comparison test
170 were used. The dependent variable was the global DNA methylation level (% 5-
171 Methylcytosine) while the independent variables were the lymphocyte subsets, treatment
172 drug, and diet group. A p-value of < 0.05 was considered statistically significant.

173

174 **Results**

175

176 **Baseline characteristics and haematological parameters following 8-weeks of HFD-**
 177 **feeding.**

178

179 The HFD-fed (HFF) group showed increased body weight gain after two weeks of HFD-feeding
 180 ($p < 0.05$). The weight gain were noticeable in weeks 2, 4, 6 and 8 weeks of HFD-feeding; with
 181 mean percentage weight gain of 7.9%, 22.12%, 27.18% and 31.33% ($p < 0.0001$) respectively.

182 The HFF group had an elevated 2-hour postprandial blood glucose and insulin levels when
 183 compared to the LFD group ($p < 0.05$), indicating impaired glucose metabolism following 8-
 184 week HFD-feeding (Table 2). The haematological indices were comparable between the HFD
 185 and LFD groups ($p > 0.05$).

186

187 **Table 2.** Baseline metabolic and haematological characteristics

Parameter	LFD	HFD	P value
Weight (g)	25.0 ± 2.5	26.0 ± 1.9	0.43
Metabolic profile			
2hr postprandial glucose			
Glucose Levels (mg/dL)	6.1 (5.4 - 6.9)	8.7 (8.5 - 9.2)	0.008
AUC mmol/L*120 min	636.0 (559.9 - 702.0)	765.0 (715.5 - 784.5)	0.032
Insulin conc (µIU/ml)	4.5 (4.4 - 4.6)	4.8 (4.6 - 8.1)	0.026
Haematological parameters			
RBC ($10^3\mu\text{l}$)	7.2 ± 0.2	6.3 ± 1.0	0.14
WBC ($10^3\mu\text{l}$)	4.9 ± 1.0	5.5 ± 3.0	0.62
MO (%)	2.0 (2.0 - 2.2)	2.6 (1.9 - 3.3)	0.32
LY (%)	89.2 ± 1.4	87.4 ± 3.1	0.31
LY # ($10^3\mu\text{l}$)	4.4 ± 0.9	4.8 ± 2.6	0.72
PLT ($10^3\mu\text{l}$)	701.0 ± 156.3	548.8 ± 276.6	0.43

188 **LFD:** low fat diet, **HFD:** high fat diet, **AUC:** area under curve, **RBC:** red blood cell, **WBC:** white blood
 189 cells, **MO (%):** monocyte percentage, **PLT:** platelet count, **LY (%):** lymphocyte percentage, **LY #:**
 190 absolute lymphocyte count.

191 **Global DNA methylation levels comparison among different cell subsets in**
 192 **prediabetic mice (HFD).**

193

194 The HFF group had increased %5mC expression on circulating peripheral blood lymphocytes
 195 when compared to the LFF group ($p = 0.049$). Notably, T lymphocytes isolated from the HFD
 196 group also showed elevated levels of %5mC ($p = 0.038$) when compared to the LFD group.

197 However, in the isolated B lymphocytes the levels of %5mC were comparable between the
198 two diet groups ($p = 0.43$) (Table 3).

199 Table 3. %5-methyl cytosine levels in HFD-fed compared to LFD-fed mice

Parameter	LFD	HFD	P value
CD45 ⁺ Lymphocytes	5.5 (2.8 – 12.0)	13.25 (10.3 – 15.9)	0.049
CD3 ⁺ T cells	30.9 (14.6 – 43.2)	68.5 (28.9 – 74.5)	0.038
CD19 ⁺ B cells	99.8 ± 0.1	99.93 ± 0.03	0.432

200 **LFD:** low fat diet, **HFD:** high fat diet.
201 Significant values ($p < 0.05$) shown in boldface.

202

203

204 **Changes in haematological indices in HFD-fed mice treated mice.**

205

206 There were significant differences in the levels of circulating lymphocyte ($F_{(3,16)} = 5.44$, $p =$
207 0.010), monocytes ($F_{(3,16)} = 3.69$, $p = 0.033$) and platelets ($\chi^2 = 9.08$, $p = 0.028$). The post-hoc
208 test showed that the lymphocytes were significantly elevated in the HFD group as compared
209 to the LDA+MET group ($p < 0.05$). In addition, the levels of circulating platelets in the LDA group
210 as compared to the HFD group (Table 4). While no significant differences in the levels of
211 circulating monocytes were observed between the treatment groups ($p > 0.05$) (Table 4).

212

213

214

215 **Global DNA methylation levels in HFD-fed compared to treated mice**

216

217 In order to assess whether DNA methylation profiles are modulated following short-term
218 treatment of 6 weeks, we compared the % 5mC levels across three treatment groups. There
219 were significant changes in the levels of T cells expressing 5mC ($F_{(4, 20)} = 6.34$, $p = 0.0016$) but
220 not in the total lymphocyte ($F_{(3, 16)} = 1.59$, $p = 0.2311$) and B cell population ($F_{(3, 16)} = 1.472$,

221 p=0.2597). The post-hoc test comparing the levels of %5mC expression on T cells showed
222 that LDA significantly lowered levels of %5mC on T cells (p=0.0045). In addition, clopidogrel
223 treatment also lowered the levels of %5mC expressed on T cells when compared to the
224 LDA+MET group (p=0.0051) (Table 4).

225

226 Table 4. %5-methyl cytosine levels in untreated HFD-fed compared to treated HDF-fed mice

Parameter	HFD	LDA	LDA + MET	CLO	P Value
Lymphocytes	12.5 ± 12.4	6.7 ± 6.4	14.1 ± 9.6	7.000 ± 4.7	0.453
CD19⁺ B cells	100.0 (99.9 - 100.0)	99.7 (99.5 - 100.0)	99.7 (99.1 - 99.9)	100.0 (99.7 - 100.0)	0.087
CD3⁺ T cells	41.2 ± 21.9	18.8 ± 19.3 ^a	49.88 ± 26.2	35.4 ± 28.3 ^b	0.001

227 ^a compared to the HFD group,
228 ^b compared to the LDA+MET group.
229 Statistical significance (p < 0.05) is shown in boldface.

230
231

232 Discussion

233 This study aimed at measuring the global DNA methylation levels of the major lymphoid cell
234 subsets following short-term high-fat diet feeding. We further assessed whether the levels of
235 DNA methylation are modulated by clopidogrel, low dose aspirin, or low-dose aspirin
236 combined with metformin. The increase in T lymphocyte-specific global DNA methylation in
237 the high-fat diet (HFD) group persisted even during short-term clopidogrel treatment. In fact,
238 our findings showed that elevated DNA methylation levels in T cells following high-fat diet
239 feeding. This may suggest that T cell-specific DNA methylation changes could affect diverse
240 biological signalling that modulate the inflammatory response. Previous studies have reported
241 on an association between obesity and global DNA hypermethylation in lymphocyte
242 subpopulations, including T cells, T cytotoxic cells, and B cells (Jacobsen et al., 2016). Taken
243 together this may suggest that patients with metabolic diseases like T2DM may present with
244 a perturbed epigenetic profile in the subpopulations of circulating lymphocytes. A strong link

245 between inflammation, insulin resistance, and epigenetic modifications has been reported
246 (Zhao et al., 2012; Van Otterdijk et al., 2017). T cells play a crucial role in the initiation as well
247 as maintenance of inflammation in adipose tissue, through persistent macrophage recruitment
248 which may as a consequence promote the development of insulin resistance (Winer et al.,
249 2011; Winer et al., 2012; Nishimura et al., 2009; DeFuria, A. C. Belkina, et al., 2013).

250 Epigenetic modification, precisely DNA methylation, may affect clopidogrel response (Zhang
251 et al., 2017). Age and body mass index (BMI), which are some of the risk factors for type 2
252 diabetes and metabolic syndrome, have also been reported to be significantly associated with
253 clopidogrel response (Cuisset et al., 2009; Khalil et al., 2016). It has also been reported that
254 diabetic patients exhibit a poor antiplatelet effect upon treatment with clopidogrel when
255 compared to non-diabetics (Angiolillo et al., 2005; Serebruany et al., 2008), a phenomenon
256 that is yet to be elucidated.

257 Although low-dose aspirin and clopidogrel have been used as dual therapy in the primary
258 prevention of cardiovascular events in patients with T2DM. A substantial incidence of major
259 adverse cardiac events (MACE) has been reported. Exploration of the role and association of
260 T cell-specific DNA methylation with clopidogrel action could help in the elucidation of the
261 possible factors associated with the varied patient responses (Khalil et al., 2016). In fact,
262 Garcia-Calzon *et al.*, reported that DNA methylation on metformin transporter genes in the
263 human liver, which differed according to anti-diabetic drug that was administered (Garcia-
264 Calzon et al., 2017). In our study, LDA significantly decreased the levels of T cell specific
265 global DNA methylation in a prediabetic state. Surprisingly, no synergetic modulation of T cell
266 global DNA methylation was observed in our study. These novel findings could suggest
267 potential insight in the variable responses to anti-inflammatory drugs amongst patients living
268 with T2DM. Persistent T cell activation that is initiated at the prediabetic phase may persist
269 during treatment and lead to increased thrombotic risk.

270 The current study was limited to the major cell lymphocyte lineages and no T cell subtyping
271 was performed to delineate whether differences in the T cell subsets exist. Activated T cells
272 have been implicated in coronary artery disease remain one of the macrovascular
273 complications associated with type 2 diabetes mellitus. In addition, an epigenome-wide study
274 revealed that hypomethylation within the tumour necrosis factor receptor-associated factor 3
275 (TRAF3) gene was associated with increased platelet aggregation and vascular recurrence in
276 ischemic stroke patients who were under clopidogrel treatment (Gallego-Fabrega et al., 2016).
277 Moreover higher TRAF3 expression due to decreased methylation may lead to an increase in
278 the CD40 signal pathway (Song et al., 2011; Kuijpers et al., 2015). CD40 is involved in the co-
279 stimulation and activation of T cells (Song et al., 2011). It remains unclear whether
280 hypermythylated T cells retain the functional capacity and whether in this may affect
281 immunological responses in patients living with T2DM.

282

283 **Conclusion**

284 T cells are involved in the initial perturbation of DNA methylation profiles in the pathogenesis
285 of inflammation, insulin resistance and subsequently type 2 diabetes. Low-dose aspirin is
286 effective in modulating T cell-specific global methylation, whereas clopidogrel showed no
287 modulatory effect on the DNA methylation profile following a short term high fat diet feeding.
288 This may suggest that the early changes in T cell DNA methylation profiles are mediated by
289 inflammation and may be reversed by using low-dose aspirin.

290

291 **Funding**

292 This study was funded by the University of KwaZulu-Natal, College of Health Sciences under
293 the Research cost funding. Furthermore, the work is based on the research supported in part
294 by the National Research Foundation of South Africa [Grant Numbers: 107519]; and by the
295 South African Medical Research Council under a Self-Initiated Research Grant (number

296 9894). The views and opinions expressed are those of the author(s) and do not necessarily
297 represent the official views of the SA MRC. The funders had no role in study design, data
298 collection and analysis, decision to publish, or preparation of the manuscript.

299 **Authors' contributions**

300 TM and BBN conceived the idea and design of the study as well as results analysis. ZM and
301 PVD helped draft the article. All authors wrote and approved the final manuscript.

302 **Competing interests**

303 The authors have no competing interests to declare.

304 **Ethics approval**

305 The study was approved by the University of KwaZulu Natal's Animal Research Ethics
306 Committee (AREC) with the ethics number: AREC/086/016.

307 **Acknowledgements**

308 The University of KwaZulu-Natal (UKZN) Biomedical Resource Unit (BRU) is acknowledged
309 for the assistance with mice laboratory procedures and animal housing facilities. Furthermore,
310 the UKZN Department of Human Physiology, College of Health Sciences (CHS) is
311 acknowledged for providing access to the flow cytometry analysis facility.

312 **References**

313 Abi Khalil, C., Omar, O.M., Al Suwaidi, J. & Taheri, S. 2018. Aspirin use and cardiovascular
314 outcome in patients with type 2 diabetes mellitus and heart failure: A population-based
315 cohort study. *Journal of the American Heart Association*, 7(21): 1–11.

316 Anderson, J.L., Adams, C.D., Antman, E.M., Bridges, C.R., Califf, R.M., Casey, D.E., Li,
317 W.E.C., Fesmire, F.M., Hochman, J.S., Levin, T.N., Lincoff, A.M., Peterson, E.D.,
318 Theroux, P., Wenger, N.K., Wright, R.S., Smith, S.C., Jacobs, A.K., Adams, C.D.,
319 Anderson, J.L., Antman, E.M., Halperin, J.L., Hunt, S.A., Krumholz, H.M., Frederick, G.,
320 Lytle, B.W., Nishimura, R., Joseph, P., Page, R.L. & Riegel, B. 2007. ACC / AHA
321 Guideline Revision ACC / AHA 2007 Guidelines for the Management of Patients With
322 Unstable Angina / Non – ST-Elevation Myocardial Infarction A Report of the American

- 323 College of Cardiology / American Heart Association Task Force on Practice Guidel.
- 324 Bacos, K., Gillberg, L., Volkov, P., Olsson, A.H., Hansen, T., Pedersen, O., Gjesing, A.P.,
325 Eiberg, H., Tuomi, T., Almgren, P., Groop, L., Eliasson, L., Vaag, A., Dayeh, T. & Ling,
326 C. 2016. Blood-based biomarkers of age-associated epigenetic changes in human
327 islets associate with insulin secretion and diabetes. *Nature communications*, 7: 11089.
- 328 Ban, N., Yamada, Y., Someya, Y., Miyawaki, K., Ihara, Y., Hosokawa, M., Toyokuni, S.,
329 Tsuda, K. & Seino, Y. 2002. Hepatocyte nuclear factor-1alpha recruits the
330 transcriptional co-activator p300 on the GLUT2 gene promoter. *Diabetes*, 51(5): 1409–
331 1418.
- 332 Capodanno, D. & Angiolillo, D.J. 2016. Aspirin for Primary Cardiovascular Risk Prevention
333 and beyond in Diabetes Mellitus. *Circulation*, 134(20): 1579–1594.
- 334 Chang, P.Y., Chen, Y.J., Chang, F.H., Lu, J., Huang, W.H., Yang, T.C., Lee, Y.T., Chang,
335 S.F., Lu, S.C. & Chen, C.H. 2013. Aspirin protects human coronary artery endothelial
336 cells against atherogenic electronegative LDL via an epigenetic mechanism: A novel
337 cytoprotective role of aspirin in acute myocardial infarction. *Cardiovascular Research*,
338 99(1): 137–145.
- 339 Dayeh, T., Volkov, P., Salö, S., Hall, E., Nilsson, E., Olsson, A.H., Kirkpatrick, C.L.,
340 Wollheim, C.B., Eliasson, L., Rönn, T., Bacos, K. & Ling, C. 2014. Genome-Wide DNA
341 Methylation Analysis of Human Pancreatic Islets from Type 2 Diabetic and Non-
342 Diabetic Donors Identifies Candidate Genes That Influence Insulin Secretion. *PLoS*
343 *Genetics*, 10(3): e1004160.
- 344 DeFuria, J., Belkina, A.C., Jagannathan-Bogdan, M., Snyder-Cappione, J., Carr, J.D.,
345 Nersesova, Y.R., Markham, D., Strissel, K.J., Watkins, A.A., Zhu, M., Allen, J.,
346 Bouchard, J., Toraldo, G., Jasuja, R., Obin, M.S., McDonnell, M.E., Apovian, C., Denis,
347 G. V. & Nikolajczyk, B.S. 2013. B cells promote inflammation in obesity and type 2

- 348 diabetes through regulation of T-cell function and an inflammatory cytokine profile.
349 *Proceedings of the National Academy of Sciences*, 110(13): 5133–5138.
- 350 Gallego-Fabrega, C., Carrera, C., Reny, J.L., Fontana, P., Slowik, A., Pera, J., Pezzini, A.,
351 Serrano-Heras, G., Segura, T., Martí-Fàbregas, J., Muiño, E., Cullell, N., Montaner, J.,
352 Krupinski, J. & Fernandez-Cadenas, I. 2016. TRAF3 Epigenetic Regulation Is
353 Associated With Vascular Recurrence in Patients With Ischemic Stroke. *Stroke*, 47(5):
354 1180–1186.
- 355 Goldfine, A.B., Fonseca, V. & Shoelson, S.E. 2011. Therapeutic approaches to target
356 inflammation in type 2 diabetes. *Clinical Chemistry*, 57(2): 162–167.
- 357 Gu, T., Gu, H.F., Hilding, A., Sjöholm, L.K., Östenson, C., Ekström, T.J., Brismar, K.,
358 Östenson, C.-G., Ekström, T.J. & Brismar, K. 2013. Increased DNA methylation levels
359 of the insulin-like growth factor binding protein 1 gene are associated with type 2
360 diabetes in Swedish men. *Clinical epigenetics*, 5(1): 21.
- 361 Iannacone, M., Sitia, G., Narvaiza, I., Ruggeri, Z.M. & Guidotti, L.G. 2007. Antiplatelet drug
362 therapy moderates immune-mediated liver disease and inhibits viral clearance in mice
363 infected with a replication-deficient adenovirus. *Clinical and Vaccine Immunology*,
364 14(11): 1532–1535.
- 365 Kilkenny, C., Browne, W.J., Cuthill, I.C., Emerson, M. & Altman, D.G. 2013. Improving
366 bioscience research reporting: The arrive guidelines for reporting animal research.
367 *Animals*, 4(1): 35–44.
- 368 King, S.B., Smith, S.C., Hirshfeld, J.W., Jacobs, A.K., Morrison, D.A., Williams, D.O., Smith,
369 S.C., Feldman, T.E., Hirshfeld, J.W., Jacobs, A.K., Kern, M.J., Iii, S.B.K., Morrison,
370 D.A., Neill, W.W.O., Schaff, H. V, Whitlow, P.L., Williams, F.D.O., Smith, S.C., Jacobs,
371 A.K., Adams, C.D., Anderson, J.L., Buller, C.E., Creager, M.A., Ettinger, S.M., Halperin,
372 J.L., Hunt, S.A., Krumholz, H.M., Kushner, F.G., Lytle, B.W., Nishimura, R., Page, R.L.,

- 373 Riegel, B., Dns, C., Tarkington, L.G. & Yancy, C.W. 2008. Focused Update of the ACC
374 / AHA / SCAI 2005 Guideline Update for Percutaneous Coronary Intervention A Report
375 of the American College of Cardiology / American Heart Association Task Force on
376 Practice Guidelines 2007 Writing Group to Review New Evidence and. *Circulation*,
377 117(2): 261–295.
- 378 Kroesen, V.M., Rodríguez-Martínez, P., García, E., Rosales, Y., Díaz, J., Martín-Céspedes,
379 M., Tapia, G., Sarrias, M.R., Cardona, P.J. & Vilaplana, C. 2018. A beneficial effect of
380 low-dose aspirin in a murine model of active tuberculosis. *Frontiers in Immunology*,
381 9(4): 1–12.
- 382 Kuliczowski, W., Witkowski, A., Polonski, L., Watala, C., Filipiak, K., Budaj, A., Golanski, J.,
383 Sitkiewicz, D., Pregowski, J., Gorski, J., Zembala, M., Opolski, G., Huber, K., Arnesen,
384 H., Kristensen, S.D. & De Caterina, R. 2009. Interindividual variability in the response to
385 oral antiplatelet drugs: A position paper of the Working Group on antiplatelet drugs
386 resistance appointed by the Section of Cardiovascular Interventions of the Polish
387 Cardiac Society, endorsed by the Working . *European Heart Journal*, 30(4): 426–435.
- 388 Kulkarni, S.S., Salehzadeh, F., Fritz, T., Zierath, J.R., Krook, A. & Osler, M.E. 2012.
389 Mitochondrial regulators of fatty acid metabolism reflect metabolic dysfunction in type 2
390 diabetes mellitus. *Metabolism: Clinical and Experimental*, 61(2): 175–85.
- 391 Kuroda, A., Rauch, T. a., Todorov, I., Ku, H.T., Al-Abdullah, I.H., Kandeel, F., Mullen, Y.,
392 Pfeifer, G.P. & Ferreri, K. 2009. Insulin gene expression is regulated by DNA
393 methylation. *PLoS ONE*, 4(9): e6953.
- 394 Kushner, F.G., Hand, M., Smith, S.C., King, S.B., Anderson, J.L., Antman, E.M., Bailey,
395 S.R., Bates, E.R., Blankenship, J.C., Casey, D.E., Green, L.A., Hochman, J.S., Jacobs,
396 A.K., Krumholz, H.M., Morrison, D.A., Ornato, J.P., Pearle, D.L., Peterson, E.D., Sloan,
397 M.A., Whitlow, P.L. & Williams, D.O. 2009. 2009 focused updates: ACC/AHA guidelines
398 for the management of patients with st-elevation myocardial infarction (Updating the

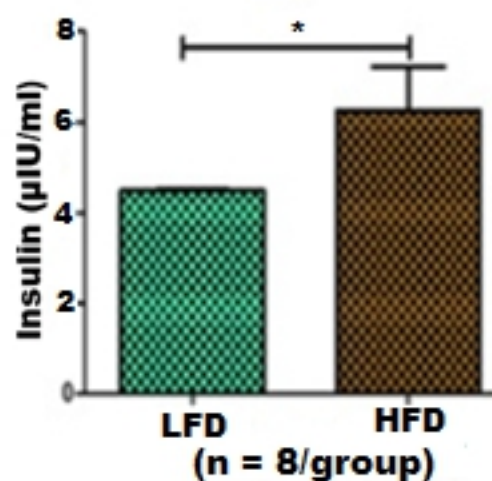
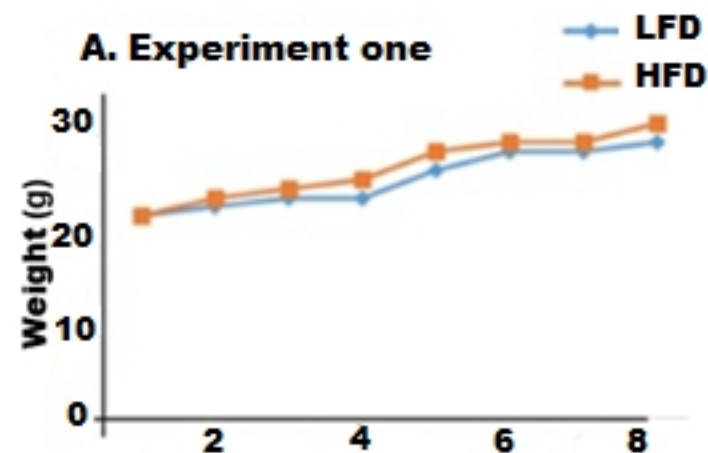
- 399 2004 guideline and 2007 focused update) and ACC/AHA/SCAI guidelines on
400 percutaneous coronary intervention (Updating the 2005 Guid. *Circulation*, 120(22):
401 2271–2306.
- 402 Mckay, J.A. & Mathers, J.C. 2011. Diet induced epigenetic changes and their implications for
403 health. *Acta Physiologica*, 202(2): 103–118.
- 404 Milagro, F.I., Mansego, M.L., De Miguel, C. & Martínez, J. a. 2013. Dietary factors,
405 epigenetic modifications and obesity outcomes: Progresses and perspectives.
406 *Molecular Aspects of Medicine*, 34(4): 782–812.
- 407 Nilsson, E., Jansson, P., Perfilyev, A., Volkov, P., Pedersen, M., Svensson, M.K., Poulsen,
408 P., Ribel-Madsen, R., Pedersen, N.L., Almgren, P., Fadista, J., R??nn, T., Pedersen,
409 B.K., Scheele, C., Vaag, A. & Ling, C. 2014. Altered DNA methylation and differential
410 expression of genes influencing metabolism and inflammation in adipose tissue from
411 subjects with type 2 diabetes. *Diabetes*, 63(9): 2962–2976.
- 412 Nilsson, E., Matte, A., Perfilyev, A., De Mello, V.D., Käkelä, P., Pihlajamäki, J. & Ling, C.
413 2015. Epigenetic alterations in human liver from subjects with type 2 diabetes in parallel
414 with reduced folate levels. *Journal of Clinical Endocrinology and Metabolism*, 100(11):
415 E1491–E1501.
- 416 Nishimura, S., Manabe, I., Nagasaki, M., Eto, K., Yamashita, H., Ohsugi, M., Otsu, M., Hara,
417 K., Ueki, K., Sugiura, S., Yoshimura, K., Kadowaki, T. & Nagai, R. 2009. CD8+ effector
418 T cells contribute to macrophage recruitment and adipose tissue inflammation in
419 obesity. *Nature Medicine*, 15(2009): 914–920.
- 420 Van Otterdijk, S.D., Binder, A.M., Szarc Vel Szic, K., Schwald, J. & Michels, K.B. 2017. DNA
421 methylation of candidate genes in peripheral blood from patients with type 2 diabetes or
422 the metabolic syndrome. *PLoS ONE*, 12(7): 1–13.
- 423 Park, J., Kim, S., Soo, M. & Kim, M. 2017. Molecular Aspects of Medicine Epigenetic

- 424 modification by dietary factors: Implications in metabolic syndrome. *Molecular Aspects*
425 *of Medicine*, 54(4): 58–70.
- 426 Pinchuk, L.M. & Filipov, N.M. 2008. Differential effects of age on circulating and splenic
427 leukocyte populations in C57BL/6 and BALB/c male mice. *Immunity & Ageing*, 5(1): 1.
- 428 Ribel-Madsen, R., Fraga, M.F., Jacobsen, S., Bork-Jensen, J., Lara, E., Calvanese, V.,
429 Fernandez, A.F., Friedrichsen, M., Vind, B.F., Hojlund, K., Beck-Nielsen, H., Esteller,
430 M., Vaag, A. & Poulsen, P. 2012. Genome-wide analysis of DNA methylation
431 differences in muscle and fat from monozygotic twins discordant for type 2 diabetes.
432 *PloS one*, 7(12): e51302.
- 433 Saisho, Y. 2015. Metformin and inflammation: its potential beyond glucose-lowering effect.
434 *Endocrine, Metabolic & Immune Disorders-Drug Targets (Formerly Current Drug*
435 *Targets-Immune, Endocrine & Metabolic Disorders)*, 15(3): 196–205.
- 436 Seman, N.A., Mohamud, W.N.W., Östenson, C.-G., Brismar, K. & Gu, H.F. 2015. Increased
437 DNA methylation of the SLC30A8 gene promoter is associated with type 2 diabetes in a
438 Malay population. *Clinical Epigenetics*, 7(1): 1–7.
- 439 Shah, U.J., Xie, W., Flyvbjerg, A., Nolan, J.J., Hojlund, K., Walker, M., Relton, C.L. & Elliott,
440 H.R. 2019. Differential methylation of the type 2 diabetes susceptibility locus KCNQ1 is
441 associated with insulin sensitivity and is predicted by CpG site specific genetic
442 variation. *Diabetes Research and Clinical Practice*, 148(2): 189–199.
- 443 Silber, S., Albertsson, P., Avilés, F.F., Camici, P.G., Colombo, A., Hamm, C., Jørgensen, E.,
444 Marco, J., Nordrehaug, J.E., Ruzyllo, W., Urban, P., Stone, G.W., Wijns, W., Deckers,
445 J., Bassand, J.P., Battler, A., Bertrand, M., Betriu, A.G., Cokkinos, D., Danchin, N., Di
446 Mario, C., de Feyter, P., Fox, K., Indolfi, C., Karsch, K., Steg, P.G., Tendera, M., Van
447 de Werf, F., Verheugt, F.W.A. & Widimski, P. 2015. Guidelines for percutaneous
448 coronary interventions. *European Heart Journal*, 26(8): 804–847.

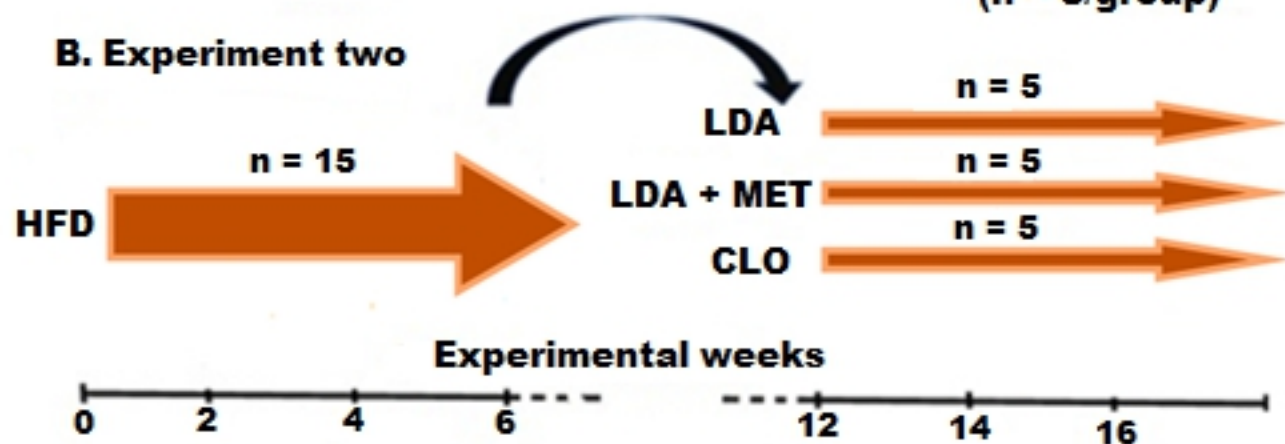
- 449 Smith, S.C., Feldman, T.E., Hirshfeld, J.W., Jacobs, A.K., Kern, M.J., King, S.B., Morrison,
450 D.A., O'Neill, W.W., Schaff, H. V, Whitlow, P.L., Williams, D.O., Antman, E.M., Smith,
451 S.C., Adams, C.D., Anderson, J.L., Faxon, D.P., Fuster, V., Halperin, J.L., Hiratzka,
452 L.F., Hunt, S.A., Jacobs, A.K., Nishimura, R., Ornato, J.P., Page, R.L. & Riegel, B.
453 2006. ACC/AHA/SCAI 2005 Guideline Update for Percutaneous Coronary Intervention.
454 *Journal of the American College of Cardiology*, 47(1): e1–e121.
- 455 Song, Z., Jin, R., Yu, S., Rivet, J.J., Smyth, S.S., Nanda, A., Granger, D.N. & Li, G. 2011.
456 CD40 is essential in the upregulation of traf proteins and NF-KappaB-dependent
457 proinflammatory gene expression after arterial injury. *PLoS ONE*, 6(8): 1–11.
- 458 Werf, F. Van De, Falk, V., Uk, K.F., Germany, A.K., France, P.G.S., Verheugt, F., Weidinger,
459 F., France, C.F., Hellemans, I., Dalby, S., Denmark, K. & France, N.A. 2008.
460 Management of acute myocardial infarction in patients presenting with persistent ST-
461 segment elevation The Task Force on the management of ST-segment elevation acute
462 myocardial infarction of the European Society of Cardiology : *Eur Heart J.*, 29(23):
463 2909–2945.
- 464 Wittenbecher, C., Ouni, M., Kuxhaus, O., Jahnert, M., Gottmann, P., Teichmann, A.,
465 Meidtnr, K., Kriebel, J., Grallert, H., Pischon, T., Boeing, H., Schulze, M.B. &
466 Schurmann, A. 2019. Insulin-Like Growth Factor Binding Protein 2 (IGFBP-2) and the
467 Risk of Developing Type 2 Diabetes. *Diabetes*, 68(1): 188–197.
- 468 Yang, B.T., Dayeh, T.A., Kirkpatrick, C.L., Taneera, J., Kumar, R., Groop, L., Wollheim, C.B.,
469 Nitert, M.D. & Ling, C. 2011. Insulin promoter DNA methylation correlates negatively
470 with insulin gene expression and positively with HbA1c levels in human pancreatic
471 islets. *Diabetologia*, 54(2): 360–367.
- 472 Yang, J., Zhou, J.S., Zhao, Y.X., Yang, Z.H., Zhao, H.D., Zhang, Y.D. & Zou, J.J. 2015.
473 ABCB1 hypomethylation is associated with decreased antiplatelet effects of clopidogrel
474 in Chinese ischemic stroke patients. *Pharmazie*, 70(2): 97–102.

- 475 Yang, M., Sun, J., Sun, Y., You, W., Dai, J. & Li, G. 2012. Association between leptin gene
476 promoter methylation and type 2 diabetes mellitus. *Chinese Journal of Medical
477 Genetics*, 29(4): 474–477.
- 478 Zhang, H., Cai, X., Yi, B., Huang, J., Wang, J. & Sun, J. 2014. Correlation of CTGF gene
479 promoter methylation with CTGF expression in type 2 diabetes mellitus with or without
480 nephropathy. *Molecular medicine reports*, 9(6): 2138–2144.
- 481 Zhang, J., Wang, C., Ha, X., Li, W., Xu, P., Gu, Y., Wang, T., Wang, Y. & Xie, J. 2017. DNA
482 methylation of tumor necrosis factor- α , monocyte chemoattractant protein-1, and
483 adiponectin genes in visceral adipose tissue is related to type 2 diabetes in the Xinjiang
484 Uygur population. *Journal of Diabetes*, 9(7): 699–706.
- 485 Zhang, S.J., Wang, Y., Yang, Y.L. & Zheng, H. 2018. Aberrant DNA methylation involved in
486 obese women with systemic insulin resistance. *Open Life Sciences*, 13(1): 201–207.
- 487 Zhao, J., Goldberg, J., Bremner, J.D. & Vaccarino, V. 2012. Global DNA methylation is
488 associated with insulin resistance: A monozygotic twin study. *Diabetes*, 61(2): 542–546.
- 489 Zhou, Z., Sun, B., Li, X. & Zhu, C. 2018. DNA methylation landscapes in the pathogenesis of
490 type 2 diabetes mellitus. *Nutrition and Metabolism*, 15(1): 1–8.
- 491 Zou, L., Yan, S., Guan, X., Pan, Y. & Qu, X. 2013. Hypermethylation of the PRKCZ gene in
492 type 2 diabetes mellitus. *Journal of Diabetes Research*, 2013(2013): 4.
- 493

A. Experiment one



B. Experiment two



Figure