Molecular Evolution of the Sex Peptide Network in *Drosophila*

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Abstract

Successful reproduction depends on interactions between numerous proteins beyond those involved directly in gamete fusion. While such reproductive proteins evolve in response to sexual selection pressures, how networks of interacting proteins arise and evolve as reproductive phenotypes change remains an open question. Here, we investigated the molecular evolution of the "sex peptide network" of *Drosophila melanogaster,* a functionally well-characterized reproductive protein network. In this species, the peptide hormone sex peptide (SP) and its interacting proteins cause major changes in female physiology and behavior after mating. In contrast, females of more distantly related *Drosophila* species do not respond to SP. In spite of these phenotypic differences, we detected orthologs of all network proteins across 22 diverse *Drosophila* species and found evidence that most orthologs likely function in reproduction throughout the genus. In *D. melanogaster* and closely related species that show similar responses to SP, we detected the recurrent, adaptive evolution of several network proteins, consistent with sexual selection acting to continually refine network function. We also found some evidence for adaptive evolution of several proteins along two key branches of the *Drosophila* phylogeny on which major changes in SP-related phenotypes likely occurred. Finally, we used gene expression profiling to examine the likely degree of functional conservation of the paralogs of an SP network protein that arose via gene duplication. Our results suggest a dynamic history for the SP network in which network members arose before the onset of robust SP-mediated responses and then were shaped by both purifying and positive selection.

Keywords: sex peptide, *Drosophila*, seminal fluid, sexual selection, molecular evolution

25 **Introduction**

26 Successful reproduction requires the fusion of egg and sperm cells, yet this fusion is often
27 facilitated by proteins that are not part of the gametes. For example, non-gametic reproductive
28 proteins provided in male seminal fluid or produced in the female reproductive tract can facilitate
29 sperm motility, induce or manage sperm storage, or cause changes to female reproductive
30 physiology (Wilburn & Swanson, 2016, Schnakenberg et al., 2011). While proteomic and
31 comparative genomic methods have enabled the identification of hundreds of gametic and non-
32 gametic reproductive proteins across diverse taxa (reviewed in McDonough et al., 2016),
33 understanding how these proteins interact, and how such interactions evolve, remain areas of
34 active research.

35 Some of the best-characterized reproductive protein interactions occur in the "sex peptide
36 network" of *Drosophila melanogaster* that regulates female post-mating behavior and
37 physiology. The network centers on the sex peptide (SP), a short peptide hormone transferred
38 from males to females as a non-gametic component of seminal fluid (Chen et al., 1988). The
39 presence of SP in the female reproductive tract stimulates egg production (Soller et al., 1999),
40 reduces receptivity to remating (Liu & Kubli, 2003, Chapman et al., 2003), facilitates the release
41 of sperm from storage prior to fertilization (Avila et al., 2010), and affects numerous other
42 female behaviors, including feeding, defecation and sleep (Carvalho et al., 2006, Apger-
43 McGlaughon & Wolfner, 2013, Isaac et al., 2010). SP-mediated effects on females persist for
44 several days after mating because SP binds to sperm, which become stored in specialized sperm
45 storage organs in the female tract (Peng et al., 2005). SP is then gradually cleaved from sperm
46 and released from the storage organs into the female tract, where it interacts with the sex peptide
47 receptor (SPR), a G-protein coupled receptor that is expressed in a subset of neurons innervating

48 the uterus (Yapici et al., 2008, Hasemeyer et al., 2009, Yang et al., 2009). This gradual "dosing"
49 of SP causes the persistence of the hormone's effects on female behavior and physiology. SPR
50 signaling is also required for the efficient release of sperm from the storage organs (Avila et al.,
51 2015).

52 While the molecule(s) on sperm to which SP binds remain unknown, RNAi screens have
53 identified several additional male seminal fluid proteins and female reproductive tract proteins
54 required for robust SP responses (Ravi Ram & Wolfner, 2007, Ravi Ram & Wolfner, 2009,
55 LaFlamme et al., 2012, Findlay et al., 2014, Singh et al., 2018). Together with SP and SPR,
56 these proteins comprise the SP network. The male-derived proteins include: predicted C-type
57 lectins CG1652 and CG1656; predicted proteases/protease homologs CG9997, seminase,
58 aquarius and intrepid; and, predicted cysteine-rich secretory proteins CG17575 and antares. The
59 female-derived proteins include fra mauro (a predicted metallopeptidase), Esp (a predicted anion
60 transporter) and hadley (which lacks identifiable protein domains). The male-derived proteins
61 act interdependently to facilitate SP binding to sperm (Ravi Ram & Wolfner, 2009, Findlay et
62 al., 2014, Singh et al., 2018), while the female-derived proteins act downstream of SP binding to
63 sperm, potentially by facilitating SP-SPR signaling (Findlay et al., 2014). Other genes expressed
64 in the secondary cells of the male accessory gland are also required for SP-mediated responses,
65 though it remains unclear whether these genes encode proteins that interact directly with the
66 network proteins described above (Sitnik et al., 2016).

67 SP's functions and interactions have been well characterized in *D. melanogaster*, but
68 comparative genomic and functional studies have shown that the SP response is not conserved
69 throughout the *Drosophila* genus. Tsuda et al. (2015) found that only species of the
70 *melanogaster* group of *Drosophila* (Fig. 1) show changes in female remating receptivity and egg

71 production upon injection with synthetic SP, even though SP and SPR orthologs can be detected
72 outside of this group. Furthermore, by incubating GFP-labeled SP with female reproductive
73 tracts from progressively more divergent species, Tsuda et al. (2015) discovered that SP could
74 bind to the female tract only in *melanogaster* group species. This observation suggested that
75 robust expression of SPR in the female tract evolved on the phylogenetic lineage leading to the
76 *melanogaster* group, which the authors tested by comparing *SPR* gene expression between in-
77 group and out-group species (Tsuda et al., 2015). Consistent with *D. melanogaster* expression
78 patterns (Yapici et al., 2008), they found that *SPR* was expressed in non-reproductive areas in
79 both sexes of all species examined. However, its expression in the female reproductive tract was
80 largely limited to the *melanogaster* group. (The only outgroup species that showed expression in
81 this location was *D. virilis*, but conspecific GFP-labeled SP did not bind to female reproductive
82 tracts in this species). Intriguingly, the SP ortholog from *D. pseudoobscura* (a non-*melanogaster*
83 group species) is expressed in *D. pseudoobscura* male reproductive tracts (Yang et al., 2018) and
84 can trigger SP-mediated responses when injected into *D. melanogaster* females, but not when
85 injected into conspecifics (Tsuda et al., 2015). This result suggests that the SP protein might have
86 evolved the potential to affect female post-mating behavior before the emergence of the
87 *melanogaster* group, but this function was not fully realized until the subsequent evolution of
88 SPR expression in the female reproductive tract (and, perhaps, within specific neurons in the
89 tract) (Hasemeyer et al., 2009, Yang et al., 2009, Yapici et al., 2008, Rezaval et al., 2012). It is
90 also possible that the transition to high levels of SPR expression in the female reproductive tract
91 created or increased an evolutionary selective pressure to bind higher levels of SP to stored
92 sperm.

93 While *Drosophila* species differ in reproductive traits for many reasons (Markow &
94 O'Grady, 2005), some of these differences relate directly to the SP network and could thus be
95 causes or consequences of SP network evolution. For example, a change in sperm length may
96 affect the amount of SP that can bind, and other structural changes to sperm could affect the
97 binding of SP and other network proteins that interact with sperm, such as CG1652 and CG1656
98 (Ravi Ram & Wolfner, 2009, Singh et al., 2018). The amount of SP bound to sperm, the rate and
99 efficacy of its release (Peng et al., 2005), and its ability to bind SPR (Yapici et al., 2008) could
100 affect re-mating rates, while the structure of female sperm storage organs could affect the ability
101 of the network proteins to bind SP to sperm or modulate SP's interaction with SPR. Changes in
102 these traits – sperm length, female remating rate, and female sperm storage structures – have
103 been well documented in the literature (Alpern et al., 2019, Joly & Bressac, 1994, Markow,
104 1996, Markow & O'Grady, 2005, Pitnick et al., 1999, Snook et al., 1994, Snook, 1995, Singh et
105 al., 2002), and we summarize them and infer their timing in Figure 1. Of particular relevance to
106 this study, the phenotypic and phylogenetic data are consistent with SPR evolving to be
107 expressed in female reproductive tracts along the lineage leading to the *melanogaster* group of
108 flies (Tsuda et al., 2015; branch 11), and an increase in sperm length on the lineage leading to *D.*
109 *ananassae* and *D. bipectinata* (Joly & Bressac, 1994, Markow, 1996, Pitnick et al., 1999; branch
110 15).

111 In light of the differences between species in reproductive phenotypes, we used
112 comparative genomics and molecular evolutionary analysis to gain insights into the evolution of
113 the SP network. While robust, long-lasting changes in female behavior and physiology due to
114 SP are found only in the *melanogaster* group of *Drosophila*, we identified orthologs of each SP
115 network protein in numerous outgroup species and verified their expression in the male

116 reproductive system in two such species using published data (Yang et al., 2018, Kelleher et al.,
117 2009). Using PAML (Yang, 2007) we determined that recurrent positive selection has acted on
118 specific sites in several of these proteins. We also detected marginal evidence that positive
119 selection has acted on certain network proteins on key phylogenetic lineages corresponding with
120 major changes in SP-related phenotypes. Finally, we used gene expression analysis to
121 investigate the functional consequences of a gene duplication event that gave rise to one of the
122 SP network proteins, seminase. Taken together, our results suggest that the members of the SP
123 network had the potential to influence reproductive success before the onset of SP/SPR-mediated
124 responses in the reproductive tract of mated females of the *melanogaster* group of species.
125 However, additional adaptive changes in these proteins occurred concurrent with, and
126 subsequent to, these critical changes in the fly reproductive system. These results underscore the
127 strength of sexual selection acting in *Drosophila* and illustrate potential molecular changes that
128 occur in the face of such selection.

129 **Methods**

130 *Identification of SP network proteins across* Drosophila *species*

131 We obtained the protein sequence for each SP network protein in *D. melanogaster* from
132 FlyBase. For species for which protein annotations were available on FlyBase (Drosophila 12
133 Genomes et al., 2007), we obtained orthologous protein-coding DNA sequences using the
134 FlyBase Orthologs feature. These species included *Drosophila simulans, sechellia, yakuba,*
135 *erecta, ananassae, pseudoobscura, persimillis, willistoni, mojavensis, virilis* and *grimshawi.* For
136 species with sequenced genomes that lacked FlyBase protein annotations (Chen et al., 2014), we
137 manually searched for gene orthologs using tBLASTn and the *D. melanogaster* protein sequence
138 as the query. These species include *Drosophila ficusphila, eugracilis, takahashii, elegans,*
139 *rhopoloa, kikkawai, bipectinata, miranda* and *albomicans.* For genes expected to have introns
140 based on the *D. melanogaster* gene structure, we looked in the unannotated species for the
141 approximate location of the *D. melanogaster* intron, and used known intron border consensus
142 sequences and six-frame translation, implemented in EMBOSS SixPack (Madeira et al., 2019),
143 to identify predicted intron borders and remove intronic sequences prior to the analyses below.

144 To study the gene duplication events that gave rise to *seminase* and its tandem gene
145 duplicates (*CG10587* and *CG11037* in *D. melanogaster*), we identified the genes flanking these
146 three genes and used them to identify the syntenic region of the other *Drosophila* genomes. We
147 assumed conservation of gene order within this syntenic region in assigning orthologs for this
148 gene family (Figure S1).

149 For all putative orthologs identified by bioinformatic methods, we verified that the
150 ortholog was the reciprocal best BLAST hit to the expected SP network member of *D.*
151 *melanogaster.* Inferred orthologs with a high degree of similarity, successful reciprocal best hits,

152 and a sequence that could be translated conceptually to produce a polypeptide without premature
153 stops, were retained for study. In cases of duplicate genes (*seminase*, *CG1652* and *CG1656*), we
154 also used gene order and synteny to confirm correct ortholog identification.
155
156 *Sequence alignment*
157 For each SP network protein, we used MUSCLE as implemented in MEGA 6.06 (Tamura
158 et al., 2013) to align amino acid sequences, then visually checked and edited each alignment for
159 accuracy. Amino acid alignments were then back-translated in MEGA to obtain the cDNA
160 alignment.
161
162 *Phylogenetic analysis*
163 To infer a *Drosophila* consensus phylogeny based on all SP network proteins, we
164 concatenated the amino acid alignments of all SP network proteins within each of the 22 species.
165 We used PROML in Phylip (Felsenstein, 2005) to infer an unrooted maximum-likelihood
166 phylogeny (with random input order, slow analysis, and all other default parameters). Gaps in the
167 alignment were used in cases in which a protein was not present in a particular species. The
168 resulting phylogeny matched published *Drosophila* phylogenies, expect for *D. virilis* and *D.*
169 *mojavensis* (Drosophila 12 Genomes et al., 2007, Markow & O'Grady, 2005, Seetharam &
170 Stuart, 2013). We then used this consensus tree for the PAML analyses, with species removed
171 on a gene-by-gene basis as described below.
172
173 *Detection of recombination*

174 Because recombination within a gene sequence can impact the results of analyses to
175 detect selection, we first used GARD with default parameters as implemented in DataMonkey
176 2.0 to check for evidence of recombination within each gene (Kosakovsky Pond et al., 2006,
177 Weaver et al., 2018). Genes were partitioned at breakpoints evaluated as significant by the
178 Kishino-Hasegawa test (p-value < 0.05 for both LH and RH; Table S2), and PAML was run on
179 each gene segment separately. We performed PAML analyses on sequence alignments spanning
180 two different ranges in the *Drosophila* phylogeny: the branch and branch-sites tests (see below)
181 were run on species from the entire genus, while the sites test (see below) was run on species
182 from only the *melanogaster* group. Thus, we generated a set of recombination breakpoints for
183 each set of species (Table S2). For each species set, six SP network genes showed evidence of
184 recombination, but the sets of genes that showed recombination differed between the two sets of
185 species.
186
187 *PAML analyses*
188 For each protein, we used codeml of the PAML package to perform evolutionary
189 analyses on protein-coding DNA sequence (Yang, 2007). To test for heterogeneity in the rate of
190 each protein's evolution across the phylogeny, we utilized the PAML branch test, which uses a
191 likelihood ratio test (LRT) to compare the "free ratio" model, allowing for different ω values for
192 each branch, with model M0, which estimates a single ω for the whole phylogeny (Yang, 1998).
193 For these tests, and for the branch-sites tests below, we used the consensus tree described above
194 that covered the entire *Drosophila* phylogeny, but manually removed from it any species for
195 which: a) an ortholog could not be identified, or b) an ortholog was identified, but it could not be
196 confidently aligned due to ambiguity over an intron position or the end of the protein-coding

197 region. Table S1 shows the set of species used for the molecular evolutionary analyses for each
198 gene.

199 To test whether a subset of sites in a protein had evolved under recurrent positive
200 selection, we used LRTs to compare an evolutionary model (M8) that allows a class of sites to
201 have $\omega > 1$ to models M7 and M8a, which do not (Swanson et al., 2003, Yang et al., 2000). For
202 proteins for which model M8 was significantly preferred to models M7 and M8a, we used the
203 Bayes Empirical Bayes (BEB) approach to identify at the 0.9 confidence level the specific
204 residues that have evolved adaptively. These comparisons were done only for species within the
205 *melanogaster* group due to the possibility of synonymous site saturation if more divergent
206 species were included. To check for convergence in the 'free-ratio' and the sites models, we ran
207 codeml twice with the initial omega set at 0.4 and 2, respectively.

208 Finally, we performed the branch-sites test for positive selection (Zhang et al., 2005) to
209 identify classes of sites that had evolved adaptively along either of two specific lineages in the
210 phylogeny that we identified *a priori* because they represent likely evolutionary transitions in
211 key SP-related traits. First, we tested for sites under selection on the branch leading to the
212 *melanogaster* group of species (Fig. 1, branch 11), since this branch corresponds with the
213 inferred timing of when the SP receptor became expressed in the female reproductive tract and,
214 consequently, when females became sensitive to the non-receptivity effect caused by SP (Tsuda
215 et al., 2015). Second, we tested for sites under selection in the lineage that leads to and separates
216 *D. ananassae* and *D. bipectinata* from the rest of the *melanogaster* group species (Fig. 1, branch
217 15), because these species are known to have somewhat longer sperm (Joly & Bressac, 1994,
218 Markow, 1996, Pitnick et al., 1999). Although we inferred other important evolutionary
219 transitions in reproductive traits on the broader *Drosophila* phylogeny (Fig. 1), we limited our

220 branch-sites analyses to these two lineages because of the greater number of available sequenced
221 species in the *melanogaster* group.
222 In the branch-sites test, we used a LRT to compare a null model allowing for only
223 purifying and neutral selection on the focal branch with an alternative model allowing for a class
224 of sites to evolve under positive selection (Yang, 2007, Yang & Dos Reis, 2011, Zhang et al.,
225 2005). Recently, Venkat et al. (2018) found that this branch-sites test can have a high rate of
226 false positives driven by multinucleotide mutations within codons (i.e., mutations at adjacent
227 sites). To control for this issue, we implemented the tests in the Venkat model, a version of
228 PAML developed by these authors that runs the analysis after masking these sites. PAML
229 analyses were implemented using custom batch scripts for GNU parallel (Tange, 2018) and
230 PAML version 4.8a, or HyPhy version 2.5.1 (in the case of the Venkat model).
231
232 *Identification of* seminase *orthologs and paralogs*
233 We identified the predicted amino acid sequences for orthologs of seminase, CG11037
234 and CG10587 in *Drosophila* species using the methods described above. To confirm that calls of
235 orthology for seminase and its paralogs were accurate, we used Phylip's PROML program
236 (Felsenstein, 2005) to infer a maximum-likelihood rooted tree (using the single copies in *D.*
237 *pseudoobscura* and *D. persimillis* as the outgroup, and default PROML parameters). This was
238 consistent with the orthology assignments made using conserved gene order, except for *D.*
239 *ananassae* and *D. bipectinata*, which are likely confounded by their long branch length.
240
241 *Evaluation of gene expression*

242 *D. melanogaster, D. yakuba, D. ficusphila, D. bipectinata, D. annanassae, D.*
243 *pseudoobscura* and *D. willistoni* were raised in the lab as in Tsuda et al. (2015). We CO_2-
244 anesthetized 9-day-old flies of each species, separated them by sex, homogenized male or female
245 whole flies in TRIzol reagent, and purified RNA from samples and synthesized cDNA as
246 previously described (Gubala et al., 2017). We then used species-specific primers to amplify
247 *seminase*, *CG11037*, *CG10587* and *RpL32* as a housekeeping gene control. Genomic DNA was
248 used as a positive control for PCR reactions, and water was used in place of template in negative
249 control reactions.

250 **Results and Discussion**

251 *SP network proteins are present in species outside of the* melanogaster *group*

252 While SP orthologs have been found in species outside of the *melanogaster* group, only
253 females of species within this group appear to show large-scale SP-mediated reproductive
254 responses (Tsuda et al., 2015). One likely factor for this change is the evolution of *SPR*
255 expression in the female reproductive tract in the last common ancestor of the *melanogaster*
256 group (Tsuda et al., 2015). This evolutionary history raises the question of whether the
257 remaining members of the SP network – all of which are critical for SP responses in *D.*
258 *melanogaster* – are present outside of the group. We addressed this question bioinformatically
259 by searching for intact orthologs across 22 *Drosophila* species with sequenced genomes.

260 Figure 2 shows that all SP network protein-coding gene orthologs are detectable in a
261 large majority of the species surveyed, including those outside of the *melanogaster* group. For
262 example, we found all currently known network proteins in *D. pseudoobscura* and *D. willistoni*,
263 and all but one ortholog in *D. virilis*. To assess whether these orthologs were likely to function
264 in reproduction outside of the *melanogaster* group, we examined publicly availably RNAseq data
265 from male reproductive tracts in *D. pseudoobscura* (Yang et al., 2018) and proteomic data from
266 male accessory glands in *D. mojavensis* (Kelleher et al., 2009). Transcripts of orthologs of male-
267 derived network proteins were consistently enriched in (or entirely specific to) samples from
268 whole males, male testes, and male carcasses in *D. pseudoobscura*, while showing either no or
269 low expression in females or in male heads (Fig. S2). This pattern is consistent with
270 expectations for reproductive proteins produced in the male accessory gland, with the "testis"
271 expression likely reflecting contamination of testis dissections with accessory gland tissue. The
272 genes encoding female-derived proteins showed broader expression patterns (Fig. S2), including

273 in whole females and whole males, but this pattern is consistent with their *D. melanogaster*
274 orthologs, the expression of which is not limited to the female reproductive system (Brown et al.,
275 2014). Predicted orthologs of the male-derived network proteins CG1652, CG1656, CG9997,
276 CG17575, seminase, aquarius and antares were also identified in a proteomic analysis of the *D.*
277 *mojavensis* accessory gland (Kelleher et al., 2009). Subsequent work showed that males of this
278 species transfer transcripts of the *antares* ortholog to females during mating (Bono et al., 2011).
279 Thus, RNAseq and proteomic data from two outgroup species are consistent with many SP
280 network proteins functioning in reproduction outside of the *melanogaster* group.
281 It is likely that some SP network proteins function in other processes in certain species
282 that impact their evolutionary trajectories. For example, SPR is expressed in both sexes outside
283 of the reproductive tract (Tsuda et al., 2015, Yapici et al., 2008), and myoinhibitory peptides
284 (MIPs) are known ligands in addition to SP (Kim et al., 2010, Poels et al., 2010, Yamanaka et al.,
285 2010). SPR-MIP interactions outside of the reproductive tract affect sleep patterns in both sexes
286 and remating propensity in females (Jang et al., 2017, Oh et al., 2014). Such interactions, in
287 addition to the sexual selective pressures exerted by SP network-mediated interactions and
288 reproductive phenotypes, have likely contributed to the evolution of SP network proteins in
289 various *Drosophila* lineages. While most male-derived network proteins appear to have male-
290 specific or heavily male-biased expression (in species for which expression data are available),
291 the female-derived proteins show broader expression patterns. Understanding these proteins'
292 non-reproductive functions will shed additional light on evolutionary forces that may have
293 shaped them.
294

295 *SP network proteins demonstrate evolutionary rate heterogeneity across the* Drosophila
296 *phylogeny*
297 Because recombination within a gene can cause false positive results in the PAML
298 analyses, we first analyzed each set of orthologs using GARD (Kosakovsky Pond et al., 2006) to
299 identify high-confidence recombination sites, which were detected for six of the proteins (Table
300 S2). These six proteins were thus split into segments corresponding to the regions between
301 recombination breakpoints, which we analyzed independently.
302 To begin investigating these proteins' molecular evolution across the genus, we used
303 PAML model M0 to estimate a single d_N/d_S ratio (ω) for each full-length protein-coding
304 sequence across all species. For the six genes for which we detected evidence of recombination
305 (Table S2), we also estimated ω across the full length of each segment. We then performed the
306 branch test (Yang, 1998) to assess whether ω varied significantly across different branches of the
307 phylogeny. Most network proteins (and segments of proteins) had full-length ω estimates
308 between 0.2 and 0.3 across the full-genus tree (Table S3). Three proteins showed notably slower
309 evolutionary rates: CG17575 (ω = 0.07), a male-expressed cysteine-rich secretory protein
310 required for binding of SP to sperm (Ravi Ram & Wolfner, 2009); Esp (ω = 0.03), a female-
311 expressed, predicted sulfate membrane transporter also required for long-term fertility (Findlay
312 et al., 2014); and SPR (ω = 0.06), the female-expressed G-protein coupled receptor for SP
313 required for female post-mating changes including egg-laying, resistance to remating and release
314 of sperm from sperm-storage organs (Avila et al., 2015, Yapici et al., 2008). While these
315 proteins' slow rates of evolution could indicate that they play highly conserved roles in
316 reproduction, it is also possible that they have evolved adaptively at only a few sites or on a few
317 lineages (see below).

318 We next ran the "free ratio" model in which PAML estimates an ω value for each branch
319 of the phylogeny. We found significant evidence of evolutionary rate heterogeneity for all but
320 one network protein, intrepid (Table S3). Additionally, all proteins but intrepid had at least one
321 phylogenetic branch for which ω was estimated to be > 1. While the branch test is not a rigorous
322 test of positive selection acting on specific branches, the results indicate that the evolutionary
323 rates of most SP network proteins have varied significantly across their evolutionary histories.
324 In contrast, the constant, slow rate of evolution for intrepid implies that this protein has likely
325 played a conserved and important role since the origin of the genus. Intrepid has undergone less
326 functional characterization than other male-expressed male network proteins, so we cannot
327 speculate further about its specific role(s) in reproduction.
328
329 *Several SP network proteins have undergone recurrent positive selection at specific sites since*
330 *the evolution of SPR expression in female reproductive tracts*
331 To determine the extent to which positive selection has shaped the evolution of the SP
332 network proteins, we used the PAML sites test to ask whether any protein had a particular subset
333 of sites that had undergone recurrent positive selection. Because of the likelihood of
334 synonymous site saturation over longer phylogenetic distances, we limited the sequences used in
335 this analysis to those from the *melanogaster* group. This set of species also represents the likely
336 extent of major SP/SPR-mediated post-mating responses, as only these species express SPR at
337 high levels in the female reproductive tract and respond to injection of synthetic SP (Tsuda et al.,
338 2015). Thus, our analyses identify proteins that might have evolved adaptively to further
339 improve/refine network function in the past ~15 million years (Seetharam & Stuart, 2013).

340 The results of the sites analyses are shown in Table 1. Four proteins – CG9997, fra
341 mauro, CG1652 and hadley – show significant evidence for having a class of amino acid sites
342 that have evolved under recurrent positive selection across the *melanogaster* group of species.
343 Three other proteins (antares, intrepid and CG17575) each have a class of sites found to be under
344 positive selection in the Model M7/M8 comparison, but these results are no longer significant
345 when comparing Models M8 and M8a, suggesting that the class of more quickly evolving sites
346 identified for each protein in Model M8 may be evolving neutrally rather than under positive
347 selection.

348 The male-expressed network proteins that have evolved adaptively are functionally co-
349 dependent. CG9997, a serine protease homolog predicted to be catalytically inactive, must be
350 produced in the male accessory glands for CG1652, a C-type lectin, to be transferred to mated
351 females (Ravi Ram & Wolfner, 2009). Likewise, in the absence of CG1652, CG9997 is not
352 efficiently "processed" from its 45-kDa form to its 36-kDa form in mated females (Ravi Ram &
353 Wolfner, 2009, Singh et al., 2018). The loss of either protein prevents SP from accumulating on
354 stored sperm in females. Recent work has shown that both CG9997 and CG1652 also bind to
355 sperm, though their sperm-binding is detectable only in the hours after mating, while SP binding
356 lasts for several days (Peng et al., 2005, Singh et al., 2018). CG9997 and CG1652 also show
357 significant evidence of evolutionary rate covariation (Findlay et al., 2014). These results suggest
358 that pressure to maintain their functional interactions may be a factor driving the adaptive
359 evolution of CG9997 and CG1652, as has been observed for pairs of interacting reproductive
360 proteins in other systems (Clark et al., 2009, Grayson, 2015).

361 Other work on CG9997 is consistent with its adaptive evolution. Wong et al. (2008)
362 found evidence for recent positive selection acting on this gene by examining patterns of

363 polymorphism and divergence between populations of *D. melanogaster* and *D. simulans*. They
364 hypothesized that non-catalytically active serine protease homologs like CG9997 function as
365 agonists or antagonists for active proteases, while others have speculated that protease homologs
366 bind to other proteins or molecules in the female tract to slow their rate of digestion by active,
367 female-derived proteases (Laflamme & Wolfner, 2013). Under either scenario, protease
368 homologs like CG9997 may need to continually coevolve with their interacting partners,
369 providing the impetus for the recurrent, adaptive evolution detected here. Additionally,
370 knockdown of *CG9997* diminishes male sperm competitive ability (Castillo & Moyle, 2014),
371 suggesting another potential factor in its adaptive evolution.

372 Less functional information exists for the adaptively evolving, female-expressed proteins.
373 Both fra mauro and hadley were identified in a screen for female-expressed proteins that
374 coevolved with a male-expressed SP network protein; in each case, the coevolutionary signal
375 was with CG17575 (Findlay et al., 2014). RNAi knockdown of either gene reduced female
376 fertility, though knockdown females could receive SP and store it properly on sperm (Findlay et
377 al., 2014). These data suggested that the proteins could be involved in maintaining the female
378 long-term response to SP, though *fra mauro* knockdown females also showed a significant
379 fertility defect in the 24 hrs after mating (Findlay et al., 2014). The fra mauro protein encodes a
380 predicted neprilysin protease, which may coevolve with its as yet unknown molecular targets or
381 antagonists (Laflamme & Wolfner, 2013). As noted above, functional domains have not been
382 identified for the hadley protein, so it is difficult to speculate on potential forces driving its
383 adaptive evolution.

384 Notably, several proteins in the SP network showed no evidence of recurrent adaptive
385 evolution within the *melanogaster* group, while others had subsets of sites with evolutionary

386 rates that were elevated, but approximated neutrality. These data suggest that while some
387 network proteins may contain regions that are under relaxed constraint, much of the functionality
388 and interdependence of the network might have already existed at the origin of the *melanogaster*
389 group.
390
391 *Several network proteins underwent adaptive evolution on specific lineages correlating with*
392 *changes in reproductive phenotypes*
393 While the PAML sites test described above detects recurrent adaptive evolution, protein
394 networks can also be shaped by bursts of episodic positive selection acting on specific
395 phylogenetic lineages. One important evolutionary transition for the SP network occurred at the
396 base of the *melanogaster* group, when *SPR* evolved expression in the lower female reproductive
397 tract (Tsuda et al., 2015). This change likely created (or exacerbated) a selective pressure for
398 higher SP levels in this location, as prolonged SP-SPR signaling could promote continued egg
399 production and prolong female non-receptivity to re-mating. Because a primary purpose of the
400 male-expressed SP network proteins in *D. melanogaster* is to bind SP to sperm to prolong the
401 post-mating response, we hypothesized that some of these proteins might have experienced a
402 burst of adaptive evolution on the same phylogenetic branch on which female reproductive *SPR*
403 expression is inferred to have evolved. Likewise, the increase in *SPR* expression in females
404 could have created a selective pressure for other female-expressed members of the network to
405 evolve. To test these ideas, we used the Venkat model, a modified PAML branch-sites test
406 (Venkat et al., 2018, Zhang et al., 2005), to ask whether any network protein had a subset of sites
407 under selection on the branch leading to the *melanogaster* group (i.e., branch 11 in Fig. 1).

408 Table 2 shows the results of these tests. Two proteins show marginal evidence for
409 adaptive evolution on branch 11: CG1656 and SPR. As originally formulated (Zhang et al.,
410 2005), the LRT for the branch-sites test follows a null distribution described as an equal mixture
411 of point mass 0 and a chi-square distribution with 1 degree of freedom (df). Under this null
412 distribution, the test statistic corresponding with a *p*-value of 0.05 is 2.71, a value exceeded by
413 each gene. However, the test is typically conducted conservatively (Venkat et al., 2018, Zhang
414 et al., 2005), following only a chi-square distribution with 1 df. The p-values listed in Table 2
415 are calculated based on this latter distribution, and they are marginally significant ($0.05 < p <$
416 0.1) for CG1656 and SPR.
417 The potential adaptive evolution of sites in the SPR protein along branch 11 is
418 interesting, because this lineage also represents the time during which the protein became
419 expressed in the female reproductive tract (Tsuda et al., 2015). Thus, it is possible that the SPR
420 gene underwent both regulatory and protein-coding adaptations that altered how the female post-
421 mating response is controlled. The other protein that potentially underwent adaptive evolution
422 along this lineage is the predicted C-type lectin CG1656, which functions similarly to its
423 recurrently rapidly evolving paralog described above, CG1652. Both lectins are required for
424 SP's long-term binding to stored sperm, and both proteins themselves bind sperm temporarily in
425 the hours after mating (Singh et al., 2018). Given the potential selective pressure to bind more
426 SP to stored sperm in female tracts expressing *SPR*, it is possible that the adaptive evolution of
427 CG1656 on this key phylogenetic branch could have helped to improve the efficiency of SP's
428 binding to sperm. This idea could be tested in future experiments by either identifying and
429 mutating the residues likely to have changed along branch 11 and/or by substituting an outgroup

430 ortholog of CG1656 (and potentially its duplicate, CG1652) into *D. melanogaster* and examining
431 the effects on SP's sperm binding and on the female long-term post-mating response.
432 Prior work demonstrated that SP binds to the full length of *D. melanogaster* sperm (Peng
433 et al., 2005, Ravi Ram & Wolfner, 2009, Singh et al., 2018). Indeed, the ability of SP (and
434 potentially other molecules) to bind sperm and then influence post-mating responses is one
435 hypothesis for why sperm tails have evolved to be so long in many *Drosophila* species. Within
436 the *melanogaster* group species that experience SP-mediated post-mating responses, one notable
437 change in reproductive physiology is that the sperm of *D. ananassae* and its closely related
438 species are considerably longer than those of *D. melanogaster* (*D. ananassae* sperm length: 3.3
439 mm; *D. melanogaster* and other *melanogaster* group species sperm length: just under 2 mm
440 (Pitnick et al., 1999, Joly & Bressac, 1994, Markow, 1996)). We thus infer that a major (>50%)
441 increase in sperm length occurred on the branch of the phylogeny leading to *D. ananassae* and
442 its close sister species *D. bipectinata* (branch 15 in Fig. 1).
443 To test for whether any SP network proteins experienced adaptive evolution concurrent
444 with this change in sperm length, we again used the modified branch-sites test. Two network
445 proteins, antares and CG17575, show evidence of positive selection acting on specific sites on
446 the lineage leading to *D. ananassae* and *D. bipectinata* (Table 3). Antares' signal of selection is
447 significant under both null distributions described above, while CG17575's signal is significant
448 under the mixed null distribution and marginally significant ($p = 0.0504$) under the conservative
449 test. In addition to facilitating SP's long-term binding to sperm, antares also binds to sperm itself
450 for a shorter period (Findlay et al., 2014, Singh et al., 2018). Thus, antares might have evolved
451 adaptively to facilitate greater or more efficient binding of either itself or SP to sperm as sperm
452 tails lengthened. Interestingly, the antares ortholog in outgroup species *D. mojavensis* and *D.*

453 *arizonae* was also found to evolve under diversifying selection (Bono et al., 2015), even though
454 *D. mojavensis* does not have a currently detectable SP ortholog (Tsuda et al., 2015) (Fig. 2).
455 Heterospecific matings between these species fail due to post-mating, pre-zygotic isolating
456 barriers, which include problems with sperm storage in the female reproductive tract (Kelleher &
457 Markow, 2007). It is thus possible that antares plays an essential role in binding molecules to
458 sperm and/or facilitating sperm storage, and that the male reproductive activity of antares has
459 been refined by different selective pressures in different lineages.

460 CG17575 is a male-expressed, cysteine-rich secretory protein required for SP and other
461 sperm-binding network proteins to localize from the female uterus, where seminal proteins and
462 sperm are first deposited, into the seminal receptacle (SR), the primary site of sperm storage in
463 *D. melanogaster* (Ravi Ram & Wolfner, 2009, Singh et al., 2018). Since CG17575 does not
464 itself bind sperm (Singh et al., 2018), further details of how CG17575 provides for proper
465 localization of other seminal proteins to the seminal receptacle are needed before we can
466 speculate on the selective forces that might have contributed to its evolution in this lineage.

467 The branch-sites tests for branches 11 and 15 reported above were conducted using full-
468 length gene sequences, since the test has limited power. However, we repeated this analysis on
469 all segments of the six genes for which recombination was detected. These results (Table S4)
470 found marginal evidence for selection for antares on branch 11 and for a segment of CG1652 on
471 branch 15. CG1656 was not among the genes for which recombination was detected (Table S2),
472 so its results above are unaltered.

473

474 *Seminase gene duplicates retain male-specific expression patterns across* melanogaster *group*
475 *species*

476 In addition to CG17575, the male-expressed serine protease seminase is required for the
477 localization of SP and other male-expressed proteins to the SR after mating (LaFlamme et al.,
478 2012, Singh et al., 2018). Seminase arose through gene duplication in the lineage leading to the
479 *melanogaster* group of flies. The genomes of *D. pseudoobscura* and other outgroup species
480 contain only one detectable copy of the gene, but in *D. melanogaster* and its fellow *melanogaster*
481 group members, there are three tandemly arrayed, intron-containing copies, suggesting two
482 distinct DNA-based duplication events (Figure S1). The other genes are *CG10587* and
483 *CG11037*. Like *seminase*, both are expressed specifically in the male accessory gland in *D.*
484 *melanogaster* (Brown et al., 2014, Leader et al., 2018). While we detected no recurrent or
485 episodic positive selection acting on seminase after these duplications (Tables 1-2), we were
486 curious whether it or its paralogs might have evolved different expression patterns (and, thus,
487 potential functions) after duplication. We thus performed RT-PCR to amplify each paralog from
488 cDNA isolated from males or females of a variety of species from the *melanogaster* group. We
489 also assessed the expression of the single-copy parent gene from *D. pseudoobscura* and *D.*
490 *willistoni*. Our results (Figure 3) show that both the single-copy genes from the outgroup
491 species, as well as all of paralogs from all *melanogaster* group species tested, are expressed
492 specifically in adult males. This result is consistent with the ancestral single copy of seminase
493 also functioning in male reproduction (and potentially with other SP network proteins).
494 Given that seminase itself has additional reproductive functions beyond its role in the SP
495 network (LaFlamme et al., 2012), it is possible that the paralogs have sub- or neo-functionalized
496 to have unique roles, in spite of their conserved expression patterns. Future studies should
497 evaluate how the paralogs contribute to reproduction, which may suggest possible evolutionary
498 forces that affected their evolution after the gene duplication events.

499 **Conclusions**

500 Sex peptide is directly responsible for major changes in female post-mating behavior and
501 physiology and is therefore one of the best characterized reproductive proteins to date. SP-
502 mediated responses appear to have arisen specifically in the *melanogaster* group of *Drosophila*,
503 and they manifest in full only with the help of a suite of male- and female-derived proteins, the
504 SP network. We have shown that these proteins are present and expressed in species outside of
505 the *melanogaster* group, suggesting they likely function in reproduction in these species and that
506 they did so in a common ancestor. Within the *melanogaster* group, several network proteins
507 (CG9997, CG1652, fra mauro, and hadley) have experienced recurrent positive selection,
508 suggesting that continued, adaptive evolution refined SP network function. A non-overlapping
509 set of proteins, including CG1656, SPR, antares, and CG17575, showed some evidence of bursts
510 of adaptive evolution on specific phylogenetic lineages corresponding with major changes in SP
511 network reproductive phenotypes. Taken together, these data suggest that SP network proteins
512 may have interacted to affect reproduction before the evolution of major SP-mediated changes in
513 the *melanogaster* group. However, once SPR became expressed at high levels in the female
514 reproductive tract in the common ancestor of this group (Tsuda et al., 2015), a combination of
515 both quick bursts of adaptation on specific lineages and recurrent changes at specific protein sites
516 helped the network evolve into the present form observed in *D. melanogaster*. This study
517 demonstrates how changes in both regulatory and protein-coding regions can affect the evolution
518 of protein networks and motivates future functional studies of the SP network proteins in
519 *Drosophila* species both within and outside of the *melanogaster* group.

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696
697

698 **Figure Legends**
699
700 **Figure 1. Phylogeny of *Drosophila* species examined in this study.** The gray box indicates the
701 *melanogaster* group. Each branch is numbered for reference in the main text. Key changes in
702 reproductive tracts are indicated by letters a-d and are based on an examination of the literature
703 (references cited in main text). The PAML branch-sites tests (see Results) were conducted on
704 branches 11 and 15. Branch lengths are not proportional to evolutionary distances.
705
706 **Figure 2. Bioinformatic identification of SP network proteins across 22 *Drosophila* species.**
707 Identified orthologs that were also reciprocal best BLAST hits are noted with a + sign, while a –
708 sign indicates no ortholog could be identified.
709
710 **Figure 3. RT-PCR on *seminase* and paralogs shows conserved, male-biased expression after**
711 **duplication.** Orthologs of *seminase*, *CG10586* and *CG11037* show male-specific expression in
712 various *melanogaster* group species, though the level of expression between paralogs and species
713 is somewhat variable. The single-copy parent gene in *D. pseudoobscura* and *D. willistoni* is also
714 expressed in a male-specific manner.
715

716 **Table 1. PAML sites tests for positive selection acting on SP network genes.** Asterisks
717 indicate genes for which recombination was detected, which were split into numbered segments
718 as indicated. Specific codons that were inferred to be under selection by PAML's Bayes
719 Empirical Bayes (BEB) analysis with Pr > 0.9 are shown for genes or segments for which
720 positive selection was detected (i.e., in which model M8 was a significantly better fit to the data
721 than models M7 and M8a). Amino acid site positions and identities refer to the *D. melanogaster*
722 protein sequence.
723

Gene	Segment Sites	M0 ω estimate	M7 vs. M8		M8 vs. M8a		% sites in ω > 1	Sites with BEB Pr > 0.90
			2*ΔlnL	*p*-value	2*ΔlnL	*p*-value		
aqrs		0.21	3.950	0.1388	0.773	0.3792		
antr		0.22	10.194	0.0061	1.935	0.1643		
intr		0.21	6.719	0.0348	0.547	0.4597		
CG9997		**0.25**	**21.001**	**2.75E-05**	**5.974**	**0.0145**	**10%**	**152S**
CG1652*		0.19	0.000	1.0000	0	1.0000		
CG1652_1	1-70	0.08	5.877	0.0529	0.797	0.3721		
CG1652_2	71-106	0.01	0	1.0000	0	1.0000		
CG1652_3	107-144	0.01	4.822	0.0897	0	1.0000		
CG1652_4	145-187	0.03	0	1.0000	0	1.0000		
CG1652_5	**188-322**	**0.67**	**6.953**	**0.0309**	**4.398**	**0.0360**	**38%**	**233P, 234G, 250V**
CG1656*		0.26	3.829	0.1474	3.221	0.0727		
CG1656_1	1-69	0.19	0.816	0.6651	0	1.0000		
CG1656_2	70-328	0.08	2.981	0.2252	0	1.0000		
CG17575*		0.06	24.088	5.88E-06	2.154	0.1422		
CG17575_1	1-139	0.03	0	1.0000	0	1.0000		
CG17575_2	140-298	0.08	16.612	0.0002	1.373	0.2413		
SP		0.22	1.463	0.4813	0	1.0000		
SPR		0.04	1.817	0.4031	8.828	0.0030		
Esp		0.03	0	1.0000	2.096	0.1477		
frma*		**0.24**	**15.986**	**0.0003**	**4.812**	**0.0283**	**7%**	**392A**
frma_1	**1-347**	**0.26**	**13.421**	**0.0012**	**4.515**	**0.0336**	**11%**	**none**
frma_2	**348-611**	**0.21**	**8.188**	**0.0167**	**6.106**	**0.0135**	**2%**	**392A**
hdly*		**0.29**	**36.156**	**1.41E-08**	**31.463**	**2.03E-08**	**6%**	**173V, 201I, 229S, 239I, 304A**
hdly_1	**1-364**	**0.30**	**42.194**	**0.0000**	**37.383**	**0.0000**	**7%**	**173V, 201I, 229S, 239I, 304A**
hdly_2	365-445	0.12	0.494	0.7813	0.463	0.4964		
sems		0.20	0	1.0000	0	1.0000		
sems_1	1-93	0.28	0.640	0.7262	0.409	0.5227		
sems_2	94-275	0.18	0.899	0.6379	0.320	0.5715		

724
725

726 **Table 2. Venkat model branch-sites tests for positive selection acting on specific sites of SP**
727 **network proteins on the lineage (Branch 11) leading to the *melanogaster* group of**
728 ***Drosophila.*** *P*-values are calculated based on a χ_1^2 distribution. Asterisks indicate likelihood
729 ratio test statistics that reach the $p < 0.05$ significance threshold for a null distribution derived
730 from a 50:50 ratio of point mass 0 and the χ_1^2 distribution.
731

Gene	Whole-Gene ω estimate	Venkat Model	
		2*ΔlnL	*p*-value
antr	298.77	2.464	0.116
aqrs	1.00	0	1.000
CG1652	7.62	1.428	0.232
CG1656	122.47	3.238*	0.072
CG9997	1.04	0	1.000
CG17575	424.52	2.425	0.119
Esp	1.08	0	1.000
frma	1.08	0.034	0.854
hdly	1.08	0	1.000
intr	1.00	0.039	0.843
sems	1.00	0	1.000
SP	1.00	0	1.000
SPR	9999.99	3.064*	0.080

732
733
734

735 **Table 3. Venkat model branch-sites tests for positive selection acting on specific sites of SP**
736 **network proteins on a lineage (Branch 15) corresponding to increased sperm length in *D.***
737 ***ananassae.*** *P*-values are calculated based on a χ_1^2 distribution. Asterisks indicate likelihood
738 ratio test statistics that reach the $p < 0.05$ significance threshold for a null distribution derived
739 from a 50:50 ratio of point mass 0 and the χ_1^2 distribution.
740

Gene	Whole-Gene ω estimate	Venkat Model	
		2*ΔlnL	*p*-value
antr	**6.19**	**3.948**	**0.047**
aqrs	2.53	0.388	0.533
CG1652	1.05	0.006	0.938
CG1656	1.04	0	1.000
CG9997	1.05	0.022	0.882
CG17575	422.98	3.827*	0.050
Esp	1.08	0	1.000
frma	1.04	0.012	0.913
hdly	1.64	0.008	0.929
intr	1.02	0	1.000
sems	403.51	0.962	0.327
SP	1.04	0.002	0.963
SPR	1.08	0	1.000

741
742

743 **Supplemental Materials**
744
745 **Table S1. Orthologs used for each gene in PAML analyses**. Some orthologs that were
746 identified in Table 2 were excluded from PAML analysis due to unresolved intron borders and/or
747 poor alignment quality. Only species above the dotted line (the *melanogaster* group) were
748 analyzed in the sites tests.
749
750 **Table S2. GARD results showing inferred recombination breakpoints.** Breakpoint positions
751 refer to nucleotide positions in the alignment files used. However, since alignments include
752 gaps, these positions do not necessarily have a 3:1 correspondence with the *D. melanogaster*
753 amino acid positions reported in Tables 1-3. The first table shows recombination breakpoints
754 detected for aligned sequences from the entire *Drosophila* genus, which were used for the branch
755 and branch-sites tests. The second table shows recombination breakpoints detected for aligned
756 sequences from only the *melanogaster* group, which were used for the sites tests.
757
758 **Figure S1. RNAseq data from *D. pseudoobscura* support reproductive functions for SP**
759 **network proteins in a species that lacks full-scale SP responses.** A) *D. pseudoobscura*
760 expression patterns for each member of the SP network. Dark shading indicates high expression
761 levels, stripes indicate low (but detectable) expression, and no shading indicates no expression
762 detected in a given sample. Male-derived network proteins show male-biased or male-specific
763 expression, consistent with reproductive functions. B) Examples of *D. pseudoobscura*
764 expression data for several SP network genes; shading in part (A) is based on these data. The
765 RNAseq data were accessed via FlyBase and generated by Yang et al. (Yang et al., 2018).
766
767 **Table S3. Branch tests for rate heterogeneity.** Partitions were implemented in PAML
768 analyses if they were significant in both the LH and RH tests.
769
770 **Table S4. Venkat model branch-sites tests for positive selection acting on specific sites of**
771 **SP network proteins detected by GARD to have multiple recombination segments.** The
772 table shows results for both branch 11 and branch 15 tests. Only the six genes for which
773 recombination was detected in the relevant trees are shown in the table. *P*-values are calculated
774 based on a χ_1^2 distribution. Asterisks indicate likelihood ratio test statistics that reach the $p <$
775 0.05 significance threshold for a null distribution derived from a 50:50 ratio of point mass 0 and
776 the χ_1^2 distribution.

Fig. 1

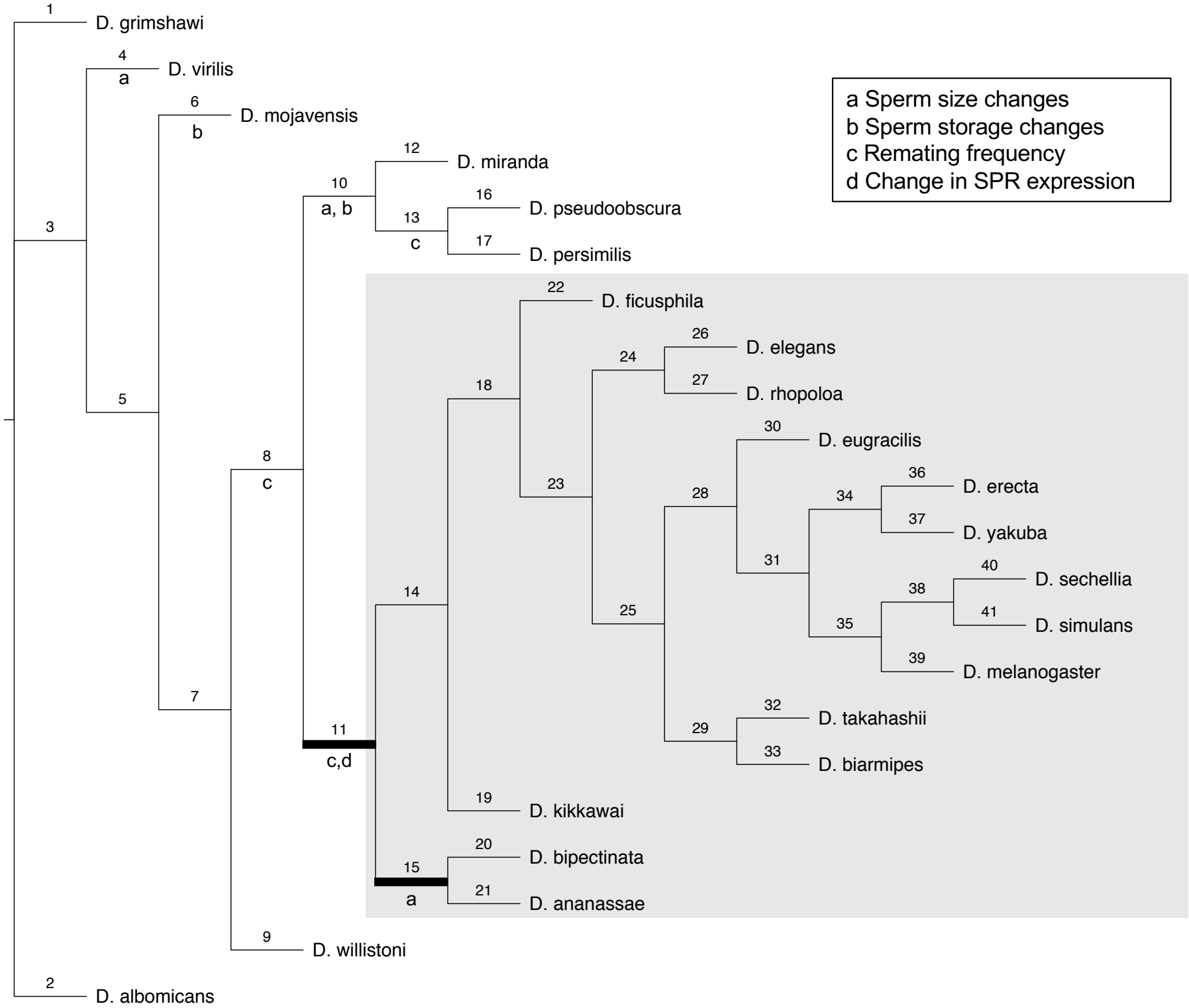
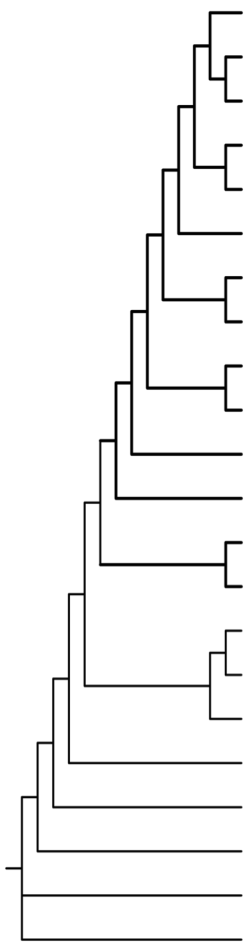
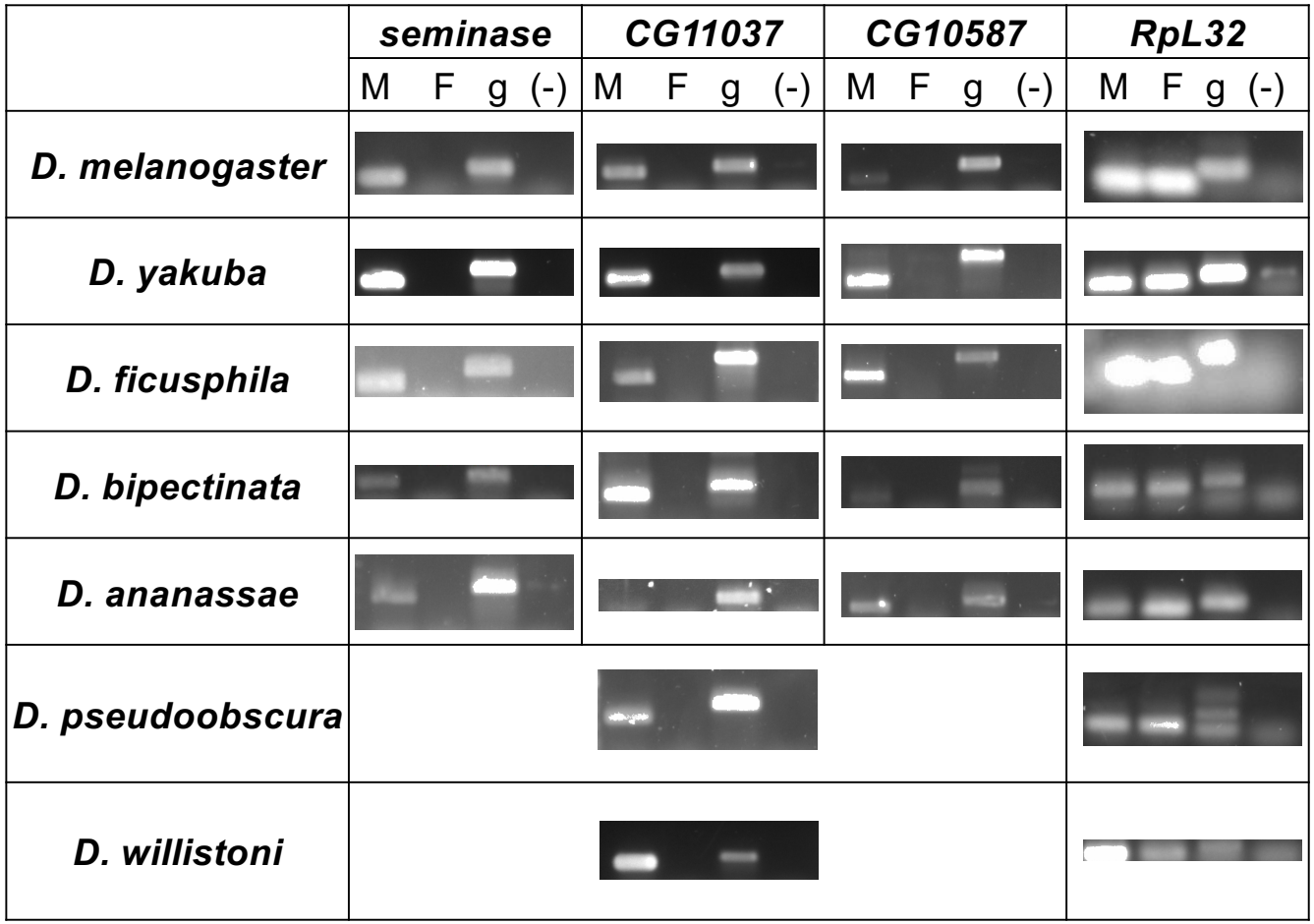

a Sperm size changes
b Sperm storage changes
c Remating frequency
d Change in SPR expression
1
D. grimshawi
4
a
D. virilis
6
b
D. mojavensis
12
D. miranda
10
a, b
16
D. pseudoobscura
13
c
17
D. persimilis
3
22
D. ficusphila
26
D. elegans
24
27
D. rhopoloa
18
5
30
D. eugracilis
8
c
36
D. erecta
23
28
34
37
D. yakuba
31
40
D. sechellia
14
38
41
25
35
D. simulans
39
D. melanogaster
7
32
D. takahashii
11
29
c,d
33
D. biarmipes
19
D. kikkawai
20
D. bipectinata
15
21
a
D. ananassae
9
D. willistoni
2
D. albomicans

Fig. 2



	intrepid	CG1652	CG1656	aquarius	antares	CG9997	CG17575	seminase	sex peptide	fra mauro	hadley	Esp	SPR
D.melanogaster	+	+	+	+	+	+	+	+	+	+	+	+	+
D.sechellia	+	+	+	+	+	+	+	+	+	+	+	+	+
D.simulans	+	+	+	+	+	+	+	+	+	+	+	+	+
D.erecta	+	+	+	+	+	+	+	+	+	+	+	+	+
D.yakuba	+	+	+	+	+	+	+	+	+	+	+	+	+
D.eugracilis	+	+	+	+	+	+	+	+	+	+	+	+	+
D.takahashii	+	+	+	+	+	+	+	+	+	+	+	+	+
D.biarmipes	+	+	+	+	+	+	+	+	+	+	+	+	+
D.elegans	+	+	+	+	+	+	+	+	+	+	+	+	+
D.rhopoloa	+	+	+	+	+	+	+	+	+	+	+	+	+
D.ficusphila	-	+	+	-	+	+	+	+	+	+	+	+	+
D.kikkawai	+	+	+	+	+	+	+	-	+	+	+	+	+
D.bipectinata	+	+	+	+	+	+	+	+	+	+	+	+	+
D.ananassae	+	+	+	+	+	+	+	+	+	+	+	+	+
D.pseudoobscura	+	+	+	+	+	+	+	+	+	+	+	+	+
D.persimilis	+	+	+	+	+	+	+	+	+	+	+	+	+
D.miranda	+	+	+	+	+	+	+	+	+	+	+	+	+
D.willistoni	+	+	+	+	+	+	+	+	+	+	+	+	+
D.mojavensis	-	+	+	+	+	+	+	+	-	+	+	+	+
D.virilis	+	+	+	+	+	+	+	+	+	+	+	-	+
D.albomicans	-	-	-	-	-	-	-	+	-	-	-	-	-
D.grimshawi	-	-	-	-	-	-	-	+	-	+	+	-	+

Fig. 3


seminase
CG11037
CG10587
RpL32
M F g (-)
M F g (-)
M F g (-)
M F g (-)
D. melanogaster
D. yakuba
D. ficusphila
D. bipectinata
D. ananassae
D. pseudoobscura
D. willistoni