1 ORIGINAL RESEARCH PAPER

2 Micronutrient composition and microbial community analysis

across diverse landraces of the Ethiopian orphan crop enset

- 4 Solomon Tamrat^{a,b*}, James S. Borrell^{c*†}, Manosh K. Biswas^d, Dawd Gashu^e, Tigist Wondimu^a, Carlos A.
- 5 Vásquez-Londoño^f, Pat J.S. Heslop-Harrison^d, Sebsebe Demissew^{c,b,g}, Paul Wilkin^c and Melanie-Jayne
- 6 R. Howes^c.
- 7 ^{*}Joint first authorship
- 8 [†]Corresponding author: J.Borrell@kew.org
- 9 ^aDepartment of Plant Biology and Biodiversity Management, Addis Ababa University, Addis Ababa, Ethiopia
- 10 ^bDepartment of Biology, Dilla University, SNNPR, Ethiopia
- 11 ^cRoyal Botanic Gardens, Kew, Richmond, Surrey, TW9 3AE, UK
- ^dDepartment of Genetics and Genome Biology, University of Leicester, LE1 7RH, UK
- 13 ^eCenter for Food Science and Nutrition, Addis Ababa University, Addis Ababa, Ethiopia
- 14 ^fFaculty of Sciences, Universidad Nacional de Colombia, Bogotá, Zip Code 111321, Colombia
- 15 ^gGullele Botanic Garden, P. O. Box 153/1029, Addis Ababa Ethiopia.
- 16
- 17
- 18
- 19
- 1

20 Abstract

Enset (*Ensete ventricosum*) is a major starch staple and food security crop for 20 million people. 21 22 Despite substantial diversity in morphology, genetics, agronomy and utilization across its range, 23 nutritional characteristics have only been reported in relatively few landraces. Here, we survey nutritional composition in 22 landraces from three enset growing regions. We present mineral 24 25 characterization of enset corm tissue, free amino acid characterization of raw and processed 26 (fermented) tissues and genomic analysis of the microbial community associated with fermentation. We show that compared to regionally important tubers and cereals, enset is high in calcium, iron, 27 28 potassium and zinc and low in sodium. We report changes in free amino acid composition due to 29 processing, and establish that the bacteria genera Acetobacter, Lactobacillus and Bifidobacterium, predominate during fermentation. Nutritional and microbial variation presents opportunities to select 30 31 for improved composition, quality or safety with potentially significant impacts in food security and 32 public health.

- 33
- 34
- 35

36 Keywords

37 *Ensete ventricosum*; Ethiopia; fermentation; free amino acids; food security; kocho; micronutrients;
 38 orphan crops.

39

40 **1. Introduction**

Humans currently satisfy most dietary requirements with surprisingly few species (Borrell et al., 41 42 2019), yet a much greater diversity of nutritionally suitable plants have been identified, often with narrow regions of utilization (Mayes et al., 2012). The food products of the large perennial herb 43 44 Ensete ventricosum (Welw.) Cheesman (Musaceae) are an important dietary starch source in Ethiopia (Borrell et al., 2018; Fanta & Satheesh, 2019; Negash & Niehof, 2004). Commonly known as enset (or 45 46 alternatively as the false banana or Abyssinian banana), this major food crop is principally cultivated as a highly resilient staple that withstands a wide range of environmental conditions and can buffer 47 48 seasonal variation in food availability. Enset contributes to the food security of over 20 million people, but is virtually unknown outside of its narrow zone of cultivation in South West Ethiopia, despite 49 growing undomesticated and unutilized across much of East and Southern Africa (Borrell et al., 2018). 50 51 In addition to being processed for multiple food products, enset is also used for livestock fodder, 52 packaging materials, fiber and traditional medicine (Borrell et al., in press; Mohammed, Martin, & 53 Laila, 2013; Olango, Tesfaye, Catellani, & Pè, 2014).

54 Despite the importance of enset for food security and the existence of hundreds of diverse landraces 55 (Borrell et al., 2018), the nutritional composition of the raw plant tissues and processed food products 56 (e.g. kocho) has only been reported in a small number of landraces (Bosha et al., 2016; Daba & 57 Shigeta, 2016; Mohammed et al., 2013; Nurfeta, Tolera, Eik, & Sundstøl, 2008). Reported nutritional composition and the relative concentrations of certain micronutrients show considerable variation. 58 59 Whilst this may be attributable in part to differing analysis methods, the extensive diversity of genetically differentiated enset landraces (Borrell et al., 2018; Tobiaw & Bekele, 2011), heterogeneity 60 61 of farm management practices (Garedew, Ayiza, Haile, & Kasaye, 2017; Olango et al., 2014) and

environmental conditions, particularly soil (Amede & Diro, 2005; Borrell et al., 2018), may also be a contributing factors. Therefore, we highlight the need to profile micronutrient composition across a representative subset of enset landrace diversity, whilst also characterizing the ubiquitous effects of fermentation on the composition, relevant to the quality and safety of enset foods, and the associated microorganism diversity responsible for mediating tissue processing.

67 The edible parts of enset comprise the starch rich pseudopetioles forming the pseudostem (overlapping leaf sheaths) which are decorticated, and the corm (the underground base of the stem 68 69 that serves as a storage organ) which is pulverized and pressed (Borrell et al., 2018). These two main tissues are collectively processed, using fermentation pits, into starch staples including bulla and 70 71 kocho (see Birmeta, Bakeeva, & Passoth, 2018, for a detailed description). Kocho is the bulk of the 72 fermented product and is baked into a thin fibrous bread considered to have a good shelf life. Bulla is a small amount of water-insoluble starchy product separated from the *kocho* during processing by 73 74 squeezing and sometimes consumed separately. The corm of enset is also occasionally consumed boiled, much like potato, and this is called *amicho*. 75

The precise fermentation practice is variable among regions and cultural groups (Garedew et al., 2017; Hunduma & Ashenafi, 2011). Karssa et al., (2014) reports preparation of a starter culture (known as *gamancho* or *gamma*) from selected corms of mature plants, followed by a two-phase process, with surface and then pit fermentation. Bosha et al. (2016) reports a ground mix of several other plant species being added to the mashed corm to initiate fermentation. Whereas many other authors (Birmeta et al., 2018; Gashe, 1987), indicate that fermentation is initiated simply from mashed tissue left for several days at ambient temperature.

83 The microorganisms responsible for fermentation alter the chemical composition of the raw substrate, which in some cases enriches the nutritional value of fermented products (Tamang, 84 85 Watanabe, & Holzapfel, 2016) by removing anti-nutritionals and breaking down complex components. 86 Furthermore, microbial communities introduced during processing are often critical to food safety 87 and preservation by preventing growth of spoilage and toxic organisms. These microorganisms, often 88 occurring as communities in food products are poorly known in orphan and minor tropical crops 89 cultivated by subsistence farmers (Tamang et al., 2016). However, improvement of these cultures 90 represents a relatively accessible opportunity to enhance the nutritional consistency, bioavailability and quality of neglected food products (Chelule, Mokoena, & Ggaleni, 2010), while urbanization and 91 92 penetration of shops leads to loss of cultures traditionally maintained by smallholders.

93 In a study on kocho production from enset, Gashe (1987) reported that Leuconostoc mensenteroides 94 and Streptococcus faecalis are responsible for initiating fermentation and reducing the pH. These are 95 then superseded by the homofermentative bacteria Lactobacillus coryniformis subsp. coryniformis 96 and L. plantarum which reduced the pH further. More recently, in an analysis of kocho samples from the Wolkite area, Weldemichael, Shimelis, Emire, & Alemu (2019) identified kocho-associated 97 bacteria as predominantly from the genera Lactobacillus and Acetobacteria, whilst a survey of kocho 98 99 samples from the Gamo highlands identified high relative abundance of Enterobacteriaceae and L. 100 mesenteroides subsp. cremoris in the early stages of fermentation (Andeta, Vandeweyer, Woldesenbet, Eshetu, & Hailemicael, 2018). More generally, the process of fermenting enset has 101 102 been reported to reduce total protein and carbohydrates, whilst increasing free amino acids (FAAs) 103 1.6-fold (Urga et al. 1997). However, the specific free amino acids in fermented products have not 104 been characterized. Therefore whether enset is a source of essential amino acids (those the body 105 cannot synthesize in sufficient quantities) in this form is unknown.

106 We hypothesize that significant genomic, phenotypic, environmental and agronomic variation in enset should also result in variation in nutritional composition across landraces. In this study we 107 108 investigate selected micronutrients with a focus on inorganic heavy metal and trace elements and free amino acids in domesticated enset in Ethiopia. We profile both raw tissues and processed 109 (fermented) products in samples from 22 landraces across three major enset growing regions, and 110 present a quantitative genomic survey of the microbial community associated with enset 111 112 fermentation. We place our results in the context of other regionally available staples and the opportunities that nutritional and microbial variation presents for applied biotechnology to improve 113 114 the nutritional quality, consistency and safety of enset derived foods.

115 **2. Material and Methods**

2.1 Sample collection and preparation

117 Fresh samples of six enset tissues (corm, pseudostem, leaf, fruit flesh, fruit peel (exocarp) and seed) 118 were harvested from a selection mature enset plants (n=28, landraces=22) occurring in three regions; 119 Sidama, Wolaita and Gurage zones in the Southern Nations Nationalities and Peoples Region (SNNPR), Ethiopia (Table 1). In selecting samples we sought to capture the full range of phenotypic and cultural 120 121 variation. Selected plants were individually prepared, processed and fermented following traditional local practices and associated qualitative indigenous knowledge related to processing was recorded. 122 Subsequently, samples of three enset products kocho, bulla and mixed kocho and bulla (Ko-Bu; 123 124 whereby the bulla liquid is not isolated during preparation, a common practice in parts of Sidama) 125 from selected plants were collected post-fermentation, air dried and powdered using an electric grinder. 126

- 127 **2.2 Micronutrient and mineral composition**
 - 6

Micronutrient content used only raw corm tissue, as relative concentration of micronutrients should not be affected by fermentation. Total ash content was estimated using 5 g of desiccated corm tissue. Samples were charred in a hot plate (Wagtech Model ST15, Sweden) and ignited at 550°C for 5 hours a in a Muffle furnace (Carbolite Model S302RR, Sweden). Residual ash was calculated as a percentage of the original desiccated sample weight.

133 Mineral content was assessed using 0.5 g corm samples digested in a closed high-performance 134 microwave digestion system (ETHOS) with the addition of trace metal grade 65% HNO₃ (Fisher 135 Scientific, UK) and H_2O_2 according to the manufacturer's recommendations under high pressure and temperature. The digest was washed with distilled water (Milli-Q). Multi-elemental analysis was 136 137 conducted using an inductively coupled ICP-MS (Thermo-Fisher Scientific iCAP-Q; Thermo Fisher 138 Scientific, Bremen, Germany). Samples were introduced from an auto-sampler (Cetac ASX-520) incorporating an ASXpress™ rapid uptake module through a perfluoroalkoxy (PFA) Microflow PFA-ST 139 nebuliser (Thermo Fisher Scientific, Bremen, Germany). Internal standards were introduced to the 140 141 sample stream on a separate line via the ASX press unit and included Ge (10 µg L-1), Rh (10 µg L-1) and Ir (5 μg L-1) in 2% trace analysis grade (Fisher Scientific, UK) HNO₃. External multi-element calibration 142 143 standards (Claritas-PPT grade CLMS-2 from SPEX Certiprep Inc., Metuchen, NJ, USA) including Al, As, Cd, Ca, Co, Cr, Cs, Cu, Fe, K, Mg, Mn, Mo, Na, Ni, P, Pb, S, Se, and Zn, in the range 0 – 100 µg L-1 (0, 20, 144 40, 100 µg L-1) were used. A bespoke external multi-element calibration solution (PlasmaCAL, SCP 145 Science, France) was used to create Ca, Mg, Na and K standards in the range 0-30 mg L-1. In addition, 146 147 KH_2PO_4 and K_2SO4 solutions were used to calibrate the machine during determination of P and S, 148 respectively. Sample processing was undertaken using Qtegra[™] software (Thermo-Fisher Scientific) 149 utilizing external cross-calibration between pulse-counting and analogue detector modes when 150 required. Blank digests containing all except enset samples were used for quality control. In addition, 7

a certified reference material (SRM 1567b – wheat flour) from the National Institute of Standards and
Technology (NIST), USA was used for standardization. Comparative plant nutritional composition data
was sourced from the USDA Food Composition Databases (USDA, 2012) and Feedipedia (Heuze,
Thiollet H., Tran G., Hassoun P., & Lebas F., 2017), and plotted together with data from this study.

155 **2.3 Crude protein and free amino acid composition**

156 Crude protein was estimated using 0.5 g of desiccated corm tissue. Briefly, 6ml of conc H_2SO_4 and 3.5ml of H₂O₂ were added followed by 3 g of a catalytic mixture of CuSO₄ and K₂SO₄ (1:15) for 15 157 158 minutes. The solution was heated at 370°C using a kjeldahl digester until a clear solution was observed and distilled using the Kjeldahl distillation apparatus (Auto K9840 Analyzer, Kjeltec, BCR-159 Technology Co, Ltd., China). The auto distiller was adjusted to add 30 ml distilled water (to avoid 160 161 precipitation of sulfate in the solution), and 40 ml NaOH (35%) in order to neutralize excess acid, 162 break down ammonium sulfate and release ammonia gas. The distillate was collected for 8 minutes in a 250 ml Erlenmeyer flask containing 2% boric acid and five drops of methyl red indicator. The 163 164 distillate was titrated with standardized 0.1M HCl and sample N content determined by subtracting 165 the N content of blank control samples.

Free amino acid composition was assessed in a subset of samples using an EZ:faast[™] kit (Phenomenex, Macclesfield, UK). Powdered samples (100 mg) were extracted in 100% water and sonicated for 20 min prior to centrifugation. Supernatants were derivatized using the method described in the manufacturer's instructions, and as described previously (Dziągwa-becker, Weber, Zajączkowska, & Oleszek, 2018). Analyses of derivatized amino acids were performed with 2µl sample injections on a Thermo Scientific system, consisting of an 'Ultimate 3000' UHPLC unit and an 'LTQ Velos Pro' mass spectrometer (Thermo Scientific, Waltham, MA, USA). Chromatography (qualitative

and quantitative analyses) was performed as described in the EZ:faast[™] kit's instructions, with amino
acids assigned by comparison with the calibration amino acid standards provided with the kit. For
each enset sample, all analyses were performed in triplicate, and the base peak areas for detected
amino acid derivatives analyzed using Thermo Xcalibur (Thermo Fisher Scientific, USA) and R software
(R Core Team, 2017), and averaged per sample.

178 **2.4 Genomic characterization of enset fermentation**

DNA was extracted from kocho using a standard CTAB protocol. High quality DNA was successfully isolated from the kocho of three landraces (Ganticho, Agade, and Ado), one sample failed (Hala). Library preparation followed the double digest restriction site associated sequencing protocol (tGBS) of Data2Bio (lowa, USA). Sequencing was performed using the lon Proton platform (Thermo Fisher Scientific, USA). Raw reads were subsequently trimmed and quality controlled following the Data2Bio pipeline.

185 To identify microorganisms present in kocho samples, retained sequences were mapped to 24,610 186 reference bacteria 285 reference fungi genomes, obtained from NCBI and (https://www.ncbi.nlm.nih.gov/, accessed 20th March 2019). A local blast database was built and 187 blast searches performed with e-value threshold of e^{-20} and scoring (match-mismatch) =1-2. The 188 percentage of retained reads that aligned to a bacterial or fungal genome were recorded. As a 189 190 control, DNA was also extracted from the leaf tissue of multiple farmer-grown enset accessions and a 191 similar pipeline used to characterize bacteria or fungi present on the surface or within plant tissue. 192 This approach provides a comparative line of evidence that where specific microbes identified in 193 kocho samples are present at high concentrations, this is indicative that they are associated with 194 fermentation (rather than simply present). It also provides an indication on whether the microbial

community responsible for fermentation originates from the plant surface/tissue or elsewhere, such
as the wider environment (e.g. soil) or a starter culture.

197 **3. Results and discussion**

3.1 Sample collection, processing and associated knowledge

199 Fermentation was initiated from mashed pseudostem tissue left for several days at ambient 200 temperature without contrived additives of cultures or previous products. Kocho and Bulla were then 201 fermented for three months, and Ko-Bu was fermented for six months. We recorded the ubiquitous 202 use of fresh enset leaves as a work surface for processing and the prevailing views that fermentation 203 will not be successful if the fermentation pit is not positioned within the enset growing area. 204 Positioning of the enset fermentation pit within the enset growing area may indicate that temperature (or shade) is important, or alternatively may be related to the presence of specific 205 206 microorganisms in the soil or elsewhere in the environment associated with enset plants. The 207 resulting fermentation products can be stored for long periods (>6 months). As expected, farmers reported that different enset landraces produce kocho (or other products) of different qualities or 208 209 preferred uses (including as medicines), however multiple landraces are normally processed together. 210 In times of extreme famine, wild enset may be blended with harvested domestic plants, despite generally being considered unpalatable, though the resulting product is regarded as having lower 211 212 quality, perhaps due to taste.

3.2 Inorganic composition of enset and comparison with other regional crops

214 Mean ash content across 19 corm samples was 5.0% (sd=1.4%), which is similar to previously 215 published ash content (4.5%) for enset corms (Nurfeta et al., 2008). Concentrations of inorganic

micronutrients and trace elements in raw enset corm tissue were averaged across 19 corm samples, each with two replicates, and plotted in Figure 1A. Inorganic micronutrient composition varied both by individual sample, and region (Figure 1B), although these data are unable to distinguish the relative importance of these two factors. Samples originating from Gurage zone were most variable on the first axis (27.1% of the variation), whilst samples from Sidama vary on the second axis (22.7% of the variation).

222 These data illustrate important variation in micronutrient composition across enset landraces and 223 growing regions, which mirrors the high vernacular and genetic diversity reported elsewhere for this crop (Borrell et al., 2018; Yemataw, Mohamed, Diro, Addis, & Blomme, 2014). This indicates the 224 225 influence of genetic, environmental and/or management factors, and as a result, there are likely to be 226 appropriate genetic targets for breeding to enhance nutritional composition. Similar performance 227 gains have been achieved in a range of other species for which there is high variability in 228 micronutrient composition (Welch & Graham, 2004). These data also suggest that climate or soil 229 conditions, together with enset management practices may influence the uptake and accumulation of 230 micronutrients in enset corm tissue, in particular because micronutrient composition appears to 231 cluster weakly by region (Figure 2) - though many landraces are region specific making further 232 investigation of this pattern challenging. We note that consumption of some micronutrients in high 233 amounts is not recommended (e.g. Ni, Cd) and high levels of these metals in samples from Sidama (Figure 1B, 1C) may be associated with groundwater contamination. 234

In a comparison with regionally important tuber and cereal crops, enset reports particularly high values for calcium, iron, potassium and zinc and relatively low values for sodium (Figure 2). Both iron and zinc deficiencies are widespread in the region (Grebmer et al., 2018). Anemia (iron deficiency) is 238 reported in 56% of children and 24% of adult women in Ethiopia (Gebru et al., 2018), with anemia also 239 included in the World Health Organization's Global Nutrition Monitoring Framework for Ethiopia 240 (WHO, 2019). This emphasizes the importance of enset as a potential dietary source of iron. Iron 241 concentrations have previously been reported as higher in enset pseudostem compared to corm 242 (Heuze et al., 2017) and we find further support for this pattern across a larger range of landraces 243 (Figure 2), though we show almost 8-fold variation between samples. Zinc deficiency is also reported 244 in diets in Ethiopia (Gebru et al., 2018; Kebede & Modes, 2013). In this study we provide further 245 evidence that enset is an important source of Zinc, and that enset corm contains higher levels of zinc 246 than pseudostem tissue. Overall, most micronutrients occur at higher concentrations in the 247 pseudostem than the corm (Figure 2).

248 **3.3 Organic composition and the nutritional implications of enset fermentation**

249 Mean total protein content across 19 corm samples in this study was 4.6% (sd=1.5%), broadly 250 consistent with values published elsewhere (Feedipedia: 3.5%, sd=1.1%). The mean concentration of 251 FAAs across raw enset tissues and processed food products are reported in Figure 3A with overall 252 concentrations for each FAA shown in Figure 3B. Of the edible components of enset, corm tissue contained the highest concentrations of FAAs, but was also most variable. FAA variation across 253 254 samples is presented in Figure 3C with variable loadings for the first and second principal components 255 reported in Figure 3D. Based on principal component analysis, concentrations for pseudostem were 256 mostly clustered but showed higher variation for corm tissue. The most variable FAAs on the second 257 axis associated with corm variability included arginine, histidine and aspartic acid. The relative 258 concentrations of individual FAAs before and after fermentation is reported in Figure 4.

259 Processing of crops is normally performed to improve the digestibility of the product, removing anti-260 nutritionals and allowing storage without microbial or animal (including insect) contamination. 261 However, there is no evidence that unprocessed enset is toxic, contrasting with cassava (cyanide) or 262 many legumes (with both highly toxic lectin glycoproteins and frequent association with fungi 263 producing mycotoxins). In this instance, enset processing appears to have a significant impact on the 264 composition of derived food products, with FAAs present at higher concentrations in raw tissues 265 (particularly leaf and corm tissue; Figure 4A), suggesting that microorganisms are net consumers of 266 FAAs during enset fermentation. These data also show that certain FAAs may be localized to specific 267 tissues, for example arginine and glutamine are both detected in corm, but at low concentrations or absent in other tissues both pre- and post-fermentation (Figure 4). Similarly, phenylalanine and 268 269 glycine occur at higher concentrations in fermented products than raw tissues, whilst many other 270 FAAs decline. Overall, several essential amino acids (isoleucine, leucine, phenylalanine and valine) are 271 increased in fermented products, particularly the main staple food, *Kocho*, suggesting that traditional 272 fermentation practices may contribute to nutrition.

273 When protein or amino acids are ingested, the vast majority of digestion products that reach the 274 blood stream are single amino acids, with the completeness of protein digestion dependent on metabolic factors (Bhutta & Sadig, 2012). Thus in some cases ingested FAAs may be more bioavailable 275 276 whilst in others a protein-rich diet may be poorly digested and of reduced nutritional value (Bhutta & Sadig, 2012). Furthermore, certain amino acids contribute to the flavour of foods (Kato, Rhue, & 277 278 Nishimura, 1989), thus influencing the selection of processing or fermentation methods and dietary 279 choices of consumers, with potential consequences for human nutrition. We note wild enset is not 280 consumed because it is considered bitter and unpalatable. It is interesting, therefore, that 281 fermentation appears to markedly reduce aspartic acid and glutamic acid, both of which produce a 13

potent sour taste at relatively low concentrations, as well as reducing arginine and histidine concentrations, which are characteristic of bitter tastes. Whether development of processing techniques has been concomitant with domestication to produce food with improved palatability is an interesting area for further research.

286 This study also reveals detection of varying concentrations of the essential amino acids isoleucine, 287 leucine, lysine, phenylalanine, threonine, tryptophan, and valine in different enset tissues (Figure 4), 288 in the form of FAAs. Consequently, these data provide the first evidence that enset, as part of a 289 broader diet, may contribute to intake of these essential amino acids. The principal FAA detected 290 across all enset samples analysed in this study was arginine (Figure 4), which occurred at the highest 291 levels in corm tissue. Although arginine is not an essential amino acid, some evidence suggests that 292 increased dietary arginine can improve outcomes in critically ill individuals (Emery, 2012) and it is 293 considered essential for infant growth, with histidine also being important for the latter (Brayfield, 294 2019), and also detected in enset tissue, especially the corm (Figure 5).

295 A traditional use of enset in Ethiopia is its intake in the form of amicho (boiled corm), which is reputed 296 to heal bone fractures (Borrell et al., 2018). Arginine is involved in collagen formation, tissue repair 297 and wound healing via proline, which is hydroxylated to form hydroxyproline, and it may also 298 stimulate collagen synthesis as a precursor of nitric oxide (Van de Poll, Luiking, Dejong, & Soeters, 299 2012). The analysis of free amino acids in this study has revealed high levels of arginine, compared to 300 other amino acids, detected in different edible parts of enset. This novel finding provides a rational 301 scientific basis for the first time that may explain the reputed traditional use of enset to aid bone 302 healing after breakage. Intriguingly, three landraces often reported as having medicinal properties (Koshkowashiye (Gurage), Astara (Sidama) and Lochingiya (Wolaita)) report three of the four highest 303

304 arginine values, whilst Lochingiya and Astara also report the highest calcium concentrations, a mineral

305 critical for bone development if not healing.

306 3.4 Microbial community characterization and signatures of contamination

307 Sequencing of kocho samples resulted in 9.6M raw reads. After quality control 8.6M reads were 308 retained with a length of 50-209bp. Of these, 1.9M and 24,300 reads returned BLAST hits above a threshold of e⁻²⁰ for bacteria and fungi genomes, respectively. The bacterial species *Acetobacter* 309 pasteurianus was the most frequent in the kocho samples Ganticho and Ado, whereas Raoultella 310 311 ornithinolytica was found to be most frequent in Agade (Figure 5A). The fungal species Penicillium 312 chrysogenum, Pichia kudriavzevii and Aspergillus fischeri were found most frequent in the Ganticho, Ado and Agade samples respectively (Figure 5B). A total of 797 bacterial species were found in 313 314 common across the three kocho samples, though comparatively only eight fungi species were found 315 across all samples (Figure 5C). As a control, BLAST results from leaf tissue is provided in Figure 5D and 316 largely identifies distinct species from the main kocho analysis. The same analysis is presented at 317 genus level in Figure S1.

Genomic analysis of the microbial community associated with enset fermentation showed that the 318 319 most abundant genus of bacteria was Acetobacter, which is generally aerobic and known for 320 producing acetic acid, reducing the pH of the fermenting enset pulp. It both gives a desirable flavor to 321 the product and lowers the pH which inhibits growth of other organisms, thus allowing safe storage of 322 the product. Whilst these species may occur as endophytes, the most common species identified in 323 the kocho samples (Acetobacter pasteurianus) was not identified in raw leaf tissue (Figure 5D). 324 The second most abundant genus was Lactobacillus, a group of anaerobic bacteria often associated 325 with controlled fermentation in foods. In a comparison with the bacteria identified by Gashe (1987) 326 we also found *L. mensenteroides*, *L. coryniformis* and *L. plantarum* but not *S. faecalis*. In the genome 327 sequencing of leaf-derived DNA, Harrison et al., (2014) reported extensive hits (>8%) against 328 Pseudomonas fluorescens and Methylobacterium radiotolerans, which they propose are endophytes 329 associated with enset; we also find a small number of *P. fluorescens* hits in our kocho samples, but 330 only one hit to *M. radiotolerans*. Overall, our analysis identified many more bacterial than fungal 331 sequences. We also find a much higher proportion of bacteria species in common between samples 332 (39.2%), compared to fungi (1.6%) including the yeasts. This suggests that bacteria are principally responsible for enset fermentation. 333

334 These analyses provide an opportunity to identify potentially harmful microorganisms such as those associated with spoilage or food poisoning (Bhunia, 2018). The number of hits associated with 335 336 Escherichia, Campylobacter, Salmonella, Clostridium (except in the Ganticho kocho sample) and 337 Listeria was generally very low in all samples. However, whilst the microbial community composition of Ganticho and Ado were very similar, Agade was dominated by Raoultella ornithinolytica, a species 338 339 associated with human infections. Stenotrophomonas maltophilia, Acinetobacter johnsonii, Rahnella 340 aquatilis are other species potentially harmful to health were also among the most frequently 341 identified bacteria in this sample. Similarly, the highest fungal hit for Agade was Aspergillus (Neosartorya) fischeri, a close relative of the major pathogen Aspergillus fumigatus also associated 342 with hypersensitivity pneumonitis (farmer's lung, immunologically mediated inflammation). This 343 suggests that in comparison to Ganticho and Ado, Agade could be characterized as a contaminated 344 345 food product. The prevalence of enset food product contamination in Ethiopia is unknown.

346 **4. Conclusions**

347 Ethiopia has historically been the world's largest recipient of targeted food aid (World Food Programme, 2013) and is 93rd of 119 qualifying countries in the 2018 Global Hunger Index (Grebmer 348 et al., 2018). Nationally, undernourishment affects 21.4% of the population and 38.4% of children 349 under the age of five are affected by stunting (Grebmer et al., 2018). Therefore effective utilization of 350 351 agricultural diversity is a priority to achieve food security and address public health needs, particularly 352 in the context of climate change and population growth (Pironon et al., 2019). However despite being 353 a center of diversity for plant domestication, dietary diversity over much of Ethiopia is extremely low 354 due to overdependence on starchy staples (Gebru et al., 2018) with major dietary deficiencies in iron 355 and zinc. This is indicative of a However, whilst child stunting prevalence in the enset growing region (reflecting *chronic* undernutrition) is largely consistent with the national average (CSA and ICF, 2016). 356 357 these areas have some of the lowest national levels of child wasting (an indicator of acute 358 undernutrition). This provides compelling indirect evidence of the food security potential of enset in mitigating acute food insecurity events. 359

The results presented here show that there is significant potential for enhanced nutritional benefits 360 361 (e.g. iron, zinc, FAAs) from enset that could impact the chronic health and welfare challenges 362 experienced by millions of Ethiopian farmers whom rely on enset as a resilient starch staple. We show that there is significant variation in enset nutritional diversity, partitioned across multiple stages of 363 364 enset cultivation and processing from the selection of landraces, environmental conditions and 365 management practices, to the timing and selection of tissues for harvest and the microbial 366 community associated with enset processing. Many of these sources of variation are not currently 367 understood, controlled or investigated and represent significant opportunities for optimization or 368 improvement. We also highlight that in addition to there being few enset germplasm collections 369 (Borrell et al., 2018), there are no collections of the microbial communities associated with kocho 17

370	processing. We therefore suggest that it is important to further document, collect and preserve their
371	diversity as they would be lost should farm structures, agronomy or management change for social or
372	economic reasons. In summary, more than 20 million Ethiopians rely on enset-derived products as a
373	starch staple or co-staple, and this population is projected to grow significantly in the coming decades
374	(Borrell et al., 2018). Therefore, selection of enset landraces with improved raw nutritional content or
375	enhanced processing techniques that improve the composition, quality or safety of enset based foods
376	has the potential for significant public health impacts.
377	
378	
379	
380	
381	
382	
383	
384	
385	
386	
387	
388	

389 Acknowledgements

390 We thank farmers for processing enset tissues for study, and field assistants for data collection.

391 Ethics Statement

- 392 All indigenous knowledge associated with enset was collected with prior informed consent and in
- 393 accordance relevant Access and Benefit Sharing Agreements.

394 **Funding Sources**

- 395 This work was supported by the GCRF Foundation Awards for Global Agricultural and Food Systems
- 396 Research, entitled, 'Modelling and genomics resources to enhance exploitation of the sustainable and
- diverse Ethiopian starch crop enset and support livelihoods' [Grant No. BB/P02307X/1].

398 **Conflict of Interest Statement**

	399	The authors declare n	o conflict of interests.
--	-----	-----------------------	--------------------------

400

401

402

403

404

405

406

407 **References**

- 408 Amede, T., & Diro, M. (2005). Optimizing soil fertility gradients in the Enset systems of the Ethiopian
 409 Highlands.
- 410 Andeta, A. F., Vandeweyer, D., Woldesenbet, F., Eshetu, F., & Hailemicael, A. (2018). Fermentation of
- 411 enset (Ensete ventricosum) in the Gamo highlands of Ethiopia^D: Physicochemical and microbial
 412 community dynamics. *Food Microbiology*, *73*, 342–350.
 413 https://doi.org/10.1016/j.fm.2018.02.011
- 414 Bhunia, A. K. (2018). *Foodborne microbial pathogens: mechanisms and pathogenesis*. Springer.
- 415 Bhutta, Z. A., & Sadiq, K. (2012). Protein Digestion and Bioavailability. Encyclopedia of Human
 416 Nutrition. (A. Prentice, L. H. Allen, & B. Caballero, Eds.) (3rd ed.).
- 417 Birmeta, G., Bakeeva, A., & Passoth, V. (2018). Yeasts and bacteria associated with kocho, an 418 Ethiopian fermented food produced from enset (Ensete ventricosum). Antonie van 419 Leeuwenhoek, International Journal of General and Molecular Microbiology, 8. https://doi.org/10.1007/s10482-018-1192-8 420
- 421 Borrell, J. S., Biswas, M. K., Goodwin, M., Blomme, G., Schwarzacher, T., Heslop-Harrison, P. J. S., ...
- 422 Wilkin, P. (2018). Enset in Ethiopia: a poorly characterised but resilient starch staple. *Annals of*
- 423 *Botany, xx–xxx,* 1–20. https://doi.org/10.1093/aob/mcy214
- 424 Borrell, J. S., Dodsworth, S., Forest, F., Perez-Escobar, O. A., Lee, M. A., Mattana, E., ... Pironon, S.
- 425 (2019). The climatic challenge: Which plants will people use in the next century? (ACCEPTED).
- 426 Environmental and Experimental Botany.
- Borrell, J. S., Goodwin, M., Blomme, G., Jacobsen, K., Wendawek, A. M., Gashu, D., ... Wilkin, P. (n.d.).
 20

428	Enset based agri-systems in Ethiopia: A systematic review of production trends, agronomy,
429	processing and the wider food security applications of a neglected banana relative. Plants,
430	People, Planet.

- 431 Bosha, A., Dalbato, A. L., Tana, T., Mohammed, W., Tesfaye, B., & Karlsson, L. M. (2016). Nutritional
- 432 and chemical properties of fermented food of wild and cultivated genotypes of enset (Ensete
- 433
 ventricosum).
 Food
 Research
 International,
 89,
 806–811.

 434
 https://doi.org/10.1016/j.foodres.2016.10.016
 https://doi.org/10.1016/j.foodres.2016.10.016
 https://doi.org/10.1016/j.foodres.2016.10.016
- 435 Brayfield, A. (2019). Martindale: The Complete Drug Reference. Retrieved from
 436 https://www.medicinescomplete.com
- 437 Chelule, P. K., Mokoena, M. P., & Gqaleni, N. (2010). Advantages of traditional lactic acid bacteria
 438 fermentation of food in Africa, 1160–1167.
- CSA (Central Statistical Agency) [Ethiopia] and ICF. (2016). *Ethiopia Demographic and Health Survey*.
 Addis Ababa, Ethiopia, and Rockville, MD, USA.
- 441 Daba, T., & Shigeta, M. (2016). Enset (Ensete Ventricosum) Production in Ethiopia: Its Nutritional and
- 442 Socio-Cultural Values. Agriculture and Food Sciences Research, 3(2), 66–74.
 443 https://doi.org/10.20448/journal.512/2016.3.2/512.2.66.74
- Dziągwa-becker, M., Weber, R., Zajączkowska, O., & Oleszek, W. (2018). Free amino acids in Viola
 tricolor in relation to different habitat conditions. *Open Chemistry*, *16*(2), 833–841.
- Emery, P. W. (2012). Amino Acids: Chemistry and Classification. In *Encyclopedia of Human Nutrition*(3RD ed., pp. 64–71). Academic Press.
- Fanta, S. W., & Satheesh, N. (2019). Nutrition & Food Science A review on nutritional profile of the 21

449	food from enset ² : A staple diet for more than 25 per cent population in Ethiopia Article
450	information [®] : For Authors A r Article information [®] :, (March). https://doi.org/10.1108/NFS-11-
451	2018-0306

- 452 Garedew, B., Ayiza, A., Haile, B., & Kasaye, H. (2017). Indigenous Knowledge of Enset (Ensete
- 453 ventricosum (Welw.) Cheesman) Cultivation and Management Practice by Shekicho People,
- 454 Southwest Ethiopia. Journal of Plant Sciences, 5(1), 6–18.
 455 https://doi.org/10.11648/j.jps.20170501.12
- 456 Gashe, B. A. (1987). Kocho fermentation. *Journal of Applied Bacteriology*, 62(6), 473–477.
 457 https://doi.org/10.1111/j.1365-2672.1987.tb02679.x
- Gebru, M., Remans, R., Brouwer, I., Baye, K., Melesse, M. B., Covic, N., ... Vandenberg, M. (2018). Food
 Systems for Healthier Diets in Ethiopia. *IFPRI Discussion Paper*, (April). Retrieved from
 https://a4nh.cgiar.org/files/2018/04/DP1050 Formatted.pdf
- Grebmer, K. von, Bernstein, J., Patterson, F., Sonntag, A., Maria Klaus, L., Fahlbusch, J., ... Fritschel, H.
 (2018). Global Hunger Index: Forced migration and hunger. *International Food Policy Institute*,
 68.
- 464 Harrison, J., Moore, K., Paszkiewicz, K., Jones, T., R.Grant, M., Ambacheew, D., ... J.Studholme, D.
- 465 (2014). A Draft Genome Sequence for Ensete ventricosum, the Drought-Tolerant "Tree Against
- 466 Hunger." *Agronomy*, *4*(1), 13–33. https://doi.org/10.3390/agronomy4010013
- 467 Heuze, V., Thiollet H., Tran G., Hassoun P., & Lebas F. (2017). *Enset (Ensete ventricosum) corms and*468 *pseudostems. Feedipedia, a programme by INRA, CIRAD, AFZ and FAO.*
- 469 Hunduma, T., & Ashenafi, M. (2011). Traditional Enset (Ensete ventricosum) Processing Techniques in

- 470 Some Parts of West Shewa Zone, Ethiopia. *Journal of Agriculture and Development*, 2(1), 37–57.
- 471 Retrieved from http://opendocs.ids.ac.uk/opendocs/handle/123456789/8730
- 472 Karssa, T. H., Ali, K. A., & Gobena, E. N. (2014). The microbiology of Kocho: An Ethiopian Traditionally
- 473 Fermented Food from Enset (Ensete ventricosum). International Journal of Life Sciences, 8(1), 7–
- 474 13. https://doi.org/10.3126/ijls.v8i1.8716
- 475 Kato, H., Rhue, M. R., & Nishimura, T. (1989). Role of Free Amino Acids and Peptides in Food Taste. In
- 476 *Flavor Chemistry* (Vol. 388, pp. 13–158). American Chemical Society.
- 477 https://doi.org/doi:10.1021/bk-1989-0388.ch013
- Kebede, A., & Modes, T. (2013). Ethiopia National Food Consumption Survey. *Ethiopian Public Health Institute, 3*, 54–67.
- 480 Mayes, S., Massawe, F. J., Alderson, P. G., Roberts, J. A., Azam-Ali, S. N., & Hermann, M. (2012). The
- 481 potential for underutilized crops to improve security of food production. *Journal of Experimental*

482 *Botany*, *63*(3), 1075–1079. https://doi.org/10.1093/jxb/err396

- 483 Mohammed, B., Martin, G., & Laila, M. K. (2013). Nutritive values of the drought tolerant food and 484 fodder crop enset. *African Journal of Agricultural Research, 8*(20), 2326–2333. 485 https://doi.org/10.5897/AJAR12.1296
- 486 Negash, A., & Niehof, A. (2004). The significance of enset culture and biodiversity for rural household
- 487 food and livelihood security in southwestern Ethiopia. Agriculture and Human Values, 21(1), 61–
- 488 71. https://doi.org/10.1023/B:AHUM.0000014023.30611.ad
- 489 Nurfeta, A., Tolera, A., Eik, L. O., & Sundstøl, F. (2008). Yield and mineral content of ten enset (Ensete
 490 ventricosum) varieties. *Tropical Animal Health and Production*, 40(4), 299–309.

491 https://doi.org/10.1007/s11250-007-9095-0

- Olango, T., Tesfaye, B., Catellani, M., & Pè, M. (2014). Indigenous knowledge, use and on-farm
 management of enset (Ensete ventricosum (Welw.) Cheesman) diversity in Wolaita, Southern
 Ethiopia. Journal of Ethnobiology and Ethnomedicine, 10(1), 41. https://doi.org/10.1186/17464269-10-41
- 496 Pironon, S., Etherington, T. R., Borrell, J. S., Kuhn, N., Macias-Fauria, M., Ondo, I., ... Willis, K. J. (2019).
- 497 Potential adaptive strategies for 29 Sub-Saharan crops under future climate change. *Nature*498 *Climate Change*, *9*, 758–763.
- R Core Team. (2017). R: A language and environment for statistical computing. R Foundation for
 Statistical Computing, Vienna, Austria.
- 501 Tamang, J. P., Watanabe, K., & Holzapfel, W. H. (2016). Review: Diversity of microorganisms in global
- 502 fermented foods and beverages. Frontiers in Microbiology, 7(MAR).
 503 https://doi.org/10.3389/fmicb.2016.00377
- 504 Tobiaw, D. C., & Bekele, E. (2011). Analysis of genetic diversity among cultivated enset (Ensete
- 505 ventricosum) populations from Essera and Kefficho, southwestern part of Ethiopia using inter
- simple sequence repeats (ISSRs) marker. *African Journal of Biotechnology*, *10*(70), 15697–15709.
- 507 https://doi.org/10.5897/AJB11.885
- 508 Urga, K., Fite, A., & Biratu, E. (1997). Natural fermentation of Enset (Ensete ventricosum) for the
 509 production of Kocho. *Ethiopian Journal of Health Development*, *11*(1), 75–81.
- 510 USDA. (2012). Composition of Foods Raw, Processed, Prepared USDA National Nutrient Database for
 511 Standard Reference, Release 25. USDA National Nutrient Database for Standard Reference,

512 *Release 25*, (September). https://doi.org/10.13140/RG.2.1.2550.5523

513	Van de Poll, M. C. G., Luiking, Y. C., Dejong, C. H. C., & Soeters, P. (2012). Amino Acids: Specific
514	Functions. In Encyclopedia of Human Nutrition (3rd ed.).

515 Welch, R. M., & Graham, R. D. (2004). Breeding for micronutrients in staple food crops from a human

516 nutrition perspective, 55(396), 353–364. https://doi.org/10.1093/jxb/erh064

517 Weldemichael, H., Shimelis, W., Emire, A., & Alemu, M. (2019). Selection and characterisation of the

518 predominant Lactobacillus species as a starter culture in the preparation of kocho, fermented

519 food from enset. *Food Science and Biotechnology*. https://doi.org/10.1007/s10068-019-00555-2

520 WHO. (2019). Nutrition Landscape Information System. Retrieved from http://www.ncdrisc.org/data-

521 downloads.html

522 World Food Programme. (2013). *2012 Food Aid Flows*. Retrieved from 523 http://documents.wfp.org/stellent/groups/public/documents/newsroom/wfp262299.pdf

Yemataw, Z., Mohamed, H., Diro, M., Addis, T., & Blomme, G. (2014). Enset (Ensete ventricosum)
clone selection by farmers and their cultural practices in southern Ethiopia. *Genetic Resources and Crop Evolution*, *61*(6), 1091–1104. https://doi.org/10.1007/s10722-014-0093-6

527

528

529

530

531

Tables

Table 1. Enset tissue and food product types evaluated in this study.

Туре	Tissue	Samples	Uses and food products
Tissue	Corm	19	A raw, underground storage organ used to make both kocho and bulla. Also boiled and eaten as
			'Amicho'.
	Pseudostem	9	A raw aboveground tissue comprising overlapping leaf sheaths. The pseudostem is the main tissue
			type making up the bulk of both kocho and bulla. Petioles and leaf midribs are fed to cattle as
			fodder.
	Leaf	1	Used as a protective layer in baking bread and to line the kocho processing area and fermentation
			pits
	Fruit flesh	1	Not currently utilized
	Fruit peel	1	Not currently utilized
	Seed	2	Not currently utilized
Product	Kocho	11	A traditional fermented flatbread
	Kocho-Bulla	8	A traditional fermented flatbread, but made without extracting the bulla liquid. In this study this
	(KoBu)		practice occurs in Sidama region
	Bulla	8	The water-insoluable starchy liquid extracted from kocho by squeezing. Bulla can be dried and
			used as a flour, or made into a gelatinous porridge. It is produced in small amounts and often
			considered the most valuable enset food product.
			considered the most valuable enset food product.

558 Figures

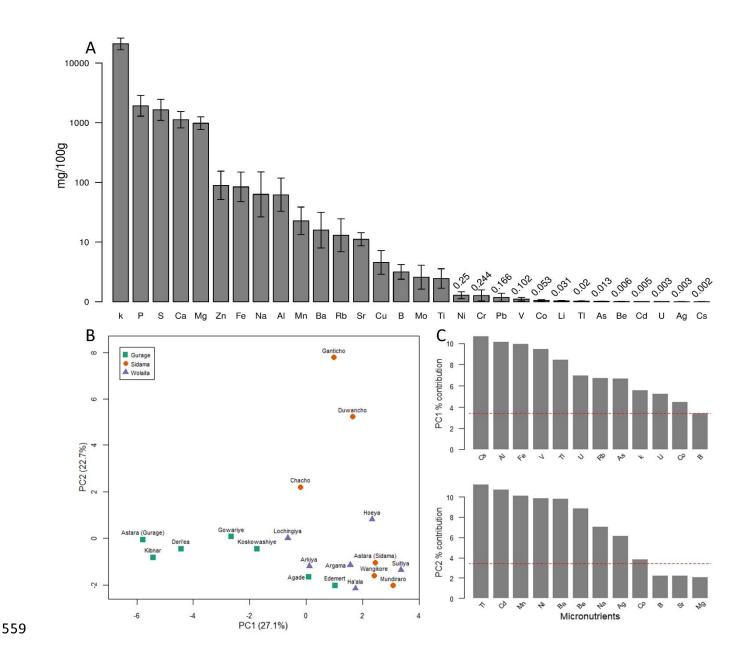


Figure 1. Concentration of inorganic minerals in enset corm tissue. A) Mineral values averaged across 19 enset corm samples. Bars denote standard deviation. B) Principal component analysis of enset corm inorganic composition across three geographical regions. C) Axis loading plots for the first and second principal components. The horizontal red line shows the level at which axis contributions would be equal.



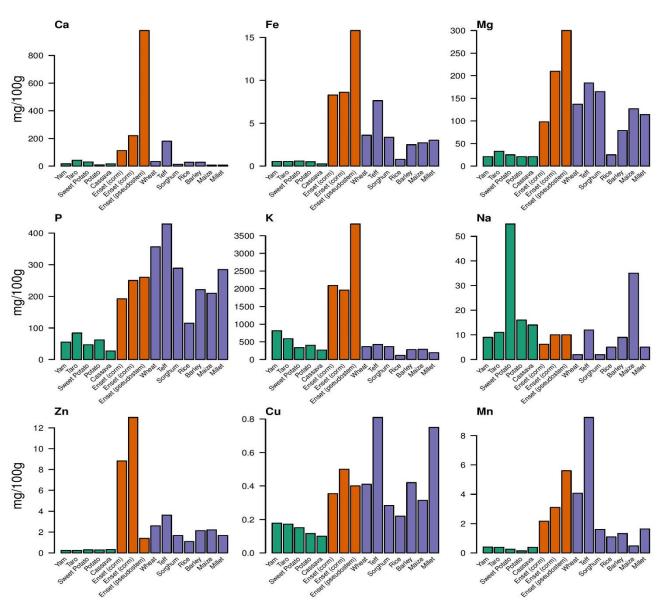


Figure 2. Comparison of enset tissue inorganic micronutrients (corm and pseudostem) with regionally occurring tubers and cereal crops. For enset three values are provided (from left to right); i) Enset (corm) results from this study (note: unavailable for K), ii) Enset (corm) from published sources and iii) Enset (pseudostem) from published sources. Comparative values for tuber and cereal crops are sourced from Feedipedia and the USDA Food Composition Databases.

572

566

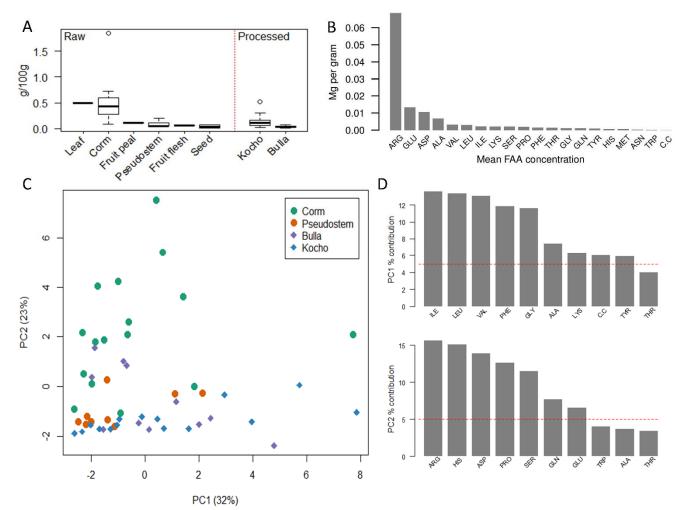


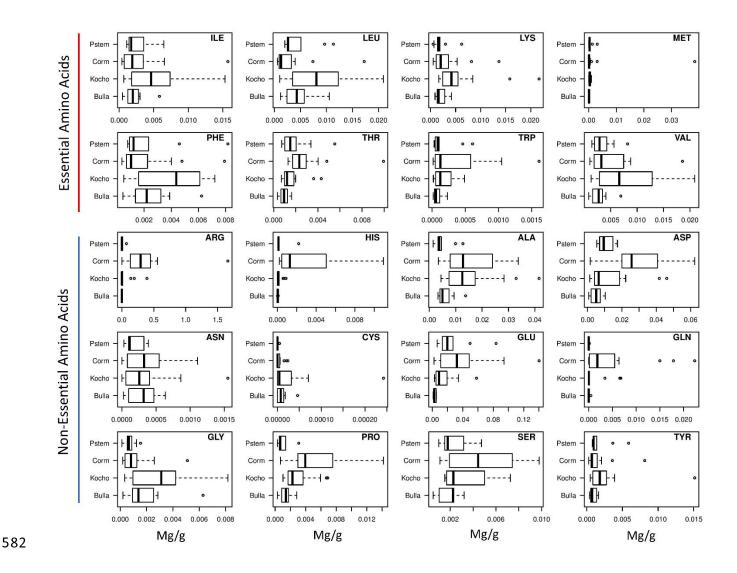
Figure 3. Free Amino Acid composition and variation in enset. A) Quantities of Free Amino Acids
(FAAs) present across raw enset tissues and processed enset food products. B) Mean concentrations
of free amino acids across all samples. C) Principal component analysis of enset FAAs across four
tissue types. D) Axis loading plots for the first and second principal components.

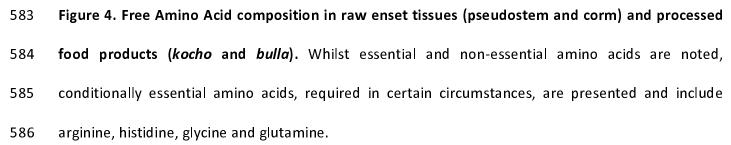
578

579

580

581





587

588

589

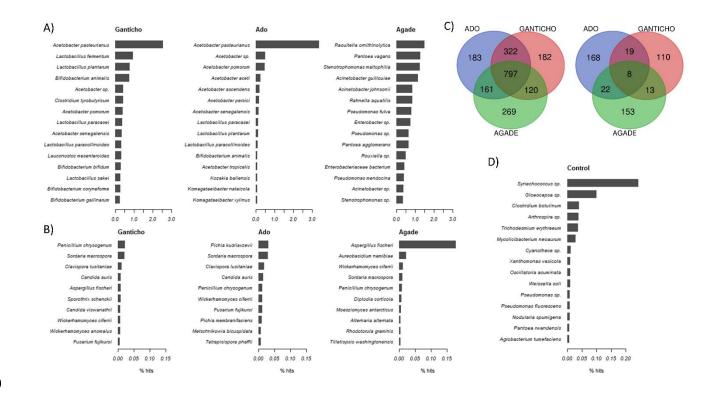




Figure 5. Microbial composition of enset kocho samples. Bacteria (A) and Fungi (B) genomes with
highest percentage hit rates across three kocho samples. C) Venn diagrams illustrating the number of
species in common between samples for Bacteria (left) and Fungi (right). D) Top bacteria and fungi
hits from enset leaf tissue