

A/T/N polygenic risk score for cognitive decline in old age

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Research in Context

Systematic Review

Authors reviewed relevant literature using PubMed and Google Scholar. Key studies that generated and validated polygenic risk scores (PRS) for clinical and pathologic AD were cited. PRS scores have been increasingly used in the literature but clinical utility continues to be questioned.

Interpretation

In the current research landscape concerning PRS clinical utility in the AD space, there is room for model improvement and our hypothesis was that a PRS with integrated risk for AD biomarkers could yield a better model for cognitive decline.

Future Directions

This study serves as proof-of-concept that encourages future study of integrated PRS across disease markers and utility in taking an A/T/N (amyloidosis, tauopathy and neurodegeneration) focused approach to genetic risk for cognitive decline and AD.

Abstract

INTRODUCTION: We developed a novel polygenic risk score (PRS) based on the A/T/N (amyloid plaques (A), phosphorylated tau tangles (T), and neurodegeneration (N)) framework and compared a PRS based on clinical AD diagnosis to assess which was a better predictor of cognitive decline. **METHODS:** We used summary statistics from genome wide association studies of cerebrospinal fluid amyloid- β ($A\beta_{42}$) and phosphorylated-tau ($p\tau_{181}$), left hippocampal volume (LHIPV), and late-onset AD dementia to calculate PRS for 1181 participants in the Alzheimer's Disease Neuroimaging Initiative (ADNI). Individual PRS were

averaged to generate a composite A/T/N PRS. We assessed the association of PRS with baseline and longitudinal cognitive composites of executive function and memory. **RESULTS:** The A/T/N PRS showed superior predictive performance on AD biomarkers and executive function decline compared to the clinical AD PRS. **DISCUSSION:** Results suggest that integration of genetic risk across AD biomarkers may improve prediction of disease progression.

1. Introduction

Alzheimer's disease (AD) currently affects roughly 5.7 million people in the United States[1]. By 2025, it is expected that 7.1 million people will be affected [1], highlighting the urgency for progress in AD-focused research. AD is highly heritable, with as much as 79% shared heritable risk reported in twin studies [2], although the genetic architecture is complex including notable polygenicity. Phenotypically, AD includes a long prodrome in which neuropathology begins to accumulate decades before cognitive symptoms can be detected [3, 4]. For that reason, early detection is critical to prevent progression of the disease [5, 6]. A combined screening approach that integrates biomarker, genomic, and clinical information will likely be required to optimize early identification and prevention of AD progression [7, 8].

Polygenic risk scores (PRS) represent a tool for early AD risk detection; however, studies that used PRS to predict AD case/control status reported predictive accuracy (AUC_{max}) of less than 83% [9-12], and the clinical utility of PRS beyond the *APOE* locus have been questioned [12, 13]. A wealth of genetic data from genome-wide association studies (GWAS) have become available in recent years [14], and incorporation of additional genetic data may represent a strategy to improve PRS predictive ability. In 2010, the largest GWAS of AD involved ~16,000 participants [15], compared to the recently completed AD GWAS that included ~600,000 participants [16]. This increase in GWAS data have enabled an increase in the number of single nucleotide polymorphisms (SNPs) used to calculate AD genetic risk scores.

Some of the first PRS studies used as few as 5 SNPs to calculate AD risk, compared to 205,068 SNPs in a more recent study [17]. Despite the progress made in genomic studies of AD, PRS continue to show limited power, and innovative analytical strategies from novel perspectives are required if PRS are going to attain clinical utility.

Improvement for PRS may come from GWAS of AD endophenotypes which focus on neuropathological features of the disease. In fact, *in vivo* biomarkers of AD neuropathology have become a central feature used to identify cognitively normal individuals who are at the greatest risk of cognitive decline [4, 18]. Additionally, the National Institute on Aging and Alzheimer's Association released a research framework for AD in which the pathological accumulation of amyloid plaques (A), neurofibrillary tangles composed of tau (T), and neurodegeneration (N) are recommended to be included in diagnostic categories of AD used for research [19]. While this A/T/N framework has emerged in studies of preclinical disease, it has not been integrated into PRS for AD despite the availability of GWAS summary statistics for autopsy and *in vivo* measures of A/T/N [20, 21]. Indeed, a previous study suggested that shared genetic drivers for hippocampal volume (a marker of neurodegeneration) and AD may exist [22]. Thus, integrating the common and independent genetic drivers of clinical AD and A/T/N could produce models with higher predictive capacity for cognitive decline in late life.

We set out to determine how PRS for CSF biomarkers of AD (amyloid and ptau₁₈₁), hippocampal volume, and clinical AD relate to biomarkers of AD and longitudinal cognitive performance. We then developed a novel composite PRS using the A/T/N framework to integrate genetic risk for AD biomarkers, hippocampal volume, and clinical AD diagnosis into a single score (A/T/N PRS). We compared the predictive capabilities of the A/T/N score to a PRS score for clinical AD and hypothesized that the A/T/N PRS would serve as a more predictive genetic risk profile compared to a PRS for clinical AD alone.

2. Methods

2.1 Participants

Participants were drawn from the Alzheimer's Disease Neuroimaging Initiative database (ADNI; adni.loni.usc.edu) launched in 2003 as a public-private partnership. The ADNI, ADNI-GO and ADNI-2 studies enrolled more than 1,500 participants, aged 55–90 years, excluding serious neurological disease, other than AD, and history of brain lesion, head trauma, or psychoactive medication use (for full inclusion/exclusion criteria see <http://www.adni-info.org>). Written informed consent was obtained from all participants at each site, and analysis of ADNI's publicly available database was approved by our local Institutional Review Board prior to data analysis.

We accessed publicly available participant data from ADNI on July 12, 2018. ADNI enrollment criteria are outlined in the ADNI protocol (<http://www.adniinfo.org/Scientists/AboutADNI.aspx>). For the cognitive analyses, we included all participants who had genomic data and longitudinal cognitive (memory and executive function) data, yielding a sample size of 1,181 participants. From this sample, 1,086 subjects also had brain MRI data and were included in neuroimaging analyses. CSF biomarker data was available for 826 participants of those whom had genomic and cognitive data. Demographics of participants with genetic and cognitive data are shown in **Table 1**.

2.2 CSF collection and assays for A β ₄₂ and ptau₁₈₁

The ADNI protocol for CSF collection and quantification of A β ₄₂ and ptau₁₈₁ biomarkers has been detailed previously, and used the multiplex xMAP Luminex platform [23, 24]. For this study, the UPenn master data set that was available on the ADNI website was downloaded. The first biomarker measurement for each participant was used as a continuous variable in statistical models.

2.3 Diagnostic criteria

Full details of these diagnostic criteria have been previously published [25]. Briefly, Normal Cognition (NC) participants had a Mini-Mental State Exam [26] (MMSE) score between 24-30 (inclusive), a Clinical Dementia Rating (CDR) of 0, and were non-depressed. Mild Cognitive Impairment (MCI) participants scored between 24-30 (inclusive) on the MMSE, had a memory complaint or objective memory loss as measured by the Wechsler Memory Scale-Revised (WMS-R) Logical Memory II, a CDR of 0.5, and absence of impairments significant enough to fit criteria for dementia. An AD diagnosis required MMSE scores between 20-26 (inclusive), a CDR score of 0.5-1.0 and meeting probable AD criteria [27].

2.4 Brain Imaging

The ADNI neuroimaging protocol has been reported in detail elsewhere [28]. Images for the current study included original uncorrected 1.5T (ADNI-1) and 3.0T (ADNI-2, ADNI-GO) T1-weighted high-resolution three-dimensional structural data. Cortical reconstruction and volumetric segmentation were performed with the FreeSurfer image analysis suite version 4.3 in ADNI-1 and 5.1 in ADNI-2 [29-31] (<http://surfer.nmr.mgh.harvard.edu/>). FreeSurfer processing in ADNI has been described in detail elsewhere [32]. FreeSurfer scans were assigned a quality control (QC) value of pass, fail or partial [33]. We excluded all scans that did not have a pass QC value. We used left hippocampal volume as our primary neuroimaging outcome measurement and included a measurement of intracranial volume (ICV) and scanner strength as covariates in all volumetric analyses, with volumetric measurements defined in FreeSurfer [34].

2.5 Neuropsychological Testing

The ADNI neuropsychological protocol, including calculation of episodic memory and executive function composite measures, has been reported previously [35, 36]. Memory (ADNI-

MEM) and executive function (ADNI-EF) composite scores were used for this study. ADNI-MEM included a composite z-score based on item-level data from Rey Auditory Verbal Learning Test, Mini-Mental State Examination (MMSE), AD Assessment Scale-Cognitive Test, and Logical Memory I and II. ADNI-EF included item-level data from Trail Making Test Parts A and B, Digit Symbol, Digit Span Backward, Animal Fluency, Vegetable Fluency, and Clock Drawing Test.

Genotyping and Genetic Quality Control Procedures

ADNI genotyping was performed using the Illumina Human610-Quad BeadChip (ADNI-1), the HumanOmniExpress BeadChip (ADNI-GO/2), or the Illumina Omni 2.5M WGS platform (Illumina, Inc., San Diego, CA). Quality control was performed using PLINK [37] (<http://pngu.mgh.harvard.edu/purcell/plink>) with a 95% threshold for genotyping efficiency applied and a minimum minor allele frequency of 0.01. SNPs outside of Hardy-Weinberg equilibrium ($p < 1 \times 10^{-6}$) were removed. Participants were excluded if they had a call rate $< 99\%$, if there was an inconsistency between reported and genetic sex, or if relatedness to another sample was established ($P_{\text{ihat}} > 0.4$), in which case one participant was selected to remain in the dataset at random (9 samples removed). Imputation was performed on the Michigan Imputation Server (<https://imputationserver.sph.umich.edu/index.html>) using the HRC r1.1.2016 reference panel. Population structure was analyzed using the fastStructure software package [38].

2.6 Polygenic Risk Score Calculation

We utilized data from a GWAS for CSF biomarkers published by Deming and colleagues [20]. The original data for this study was collected from 3,146 individuals across nine cohort studies, including ADNI. After removing ADNI participants [$N=390$ (ADNI-1), 397 (ADNI-2)], the

GWAS was re-run using data from 2,359 participants in the remaining seven cohorts. Summary results of the re-analyses excluding ADNI participants are available in Supplementary Materials.

A GWAS for clinical AD using data from the International Genomics of Alzheimer's Project (IGAP)[39, 40] was re-run excluding 441 ADNI participants (55,931 participants remained across the Alzheimer's Disease Genetic Consortium (ADGC), the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) Consortium, the European Alzheimer's Disease Initiative (EADI) and the Genetic and Environmental Risk in Alzheimer's Disease (GERAD) Consortium). Summary results of the re-analysis are presented in Supplementary Materials.

Publicly available summary statistics from a UK Biobank GWAS of brain imaging phenotypes that included 8,428 participants [21] was utilized to generate ADNI participant PRS for left hippocampal volume.

PRS were calculated for each outcome in PLINK 1.9 using the scoring function (<https://www.cog-genomics.org/plink/1.9/>) based on summary statistics from overlapping SNPs in the ADNI cohort and the GWAS studies detailed above. The number of overlapping SNPs to calculate PRS are shown in **Supplemental Table 1**. We calculated PRS scores using a linkage disequilibrium threshold of 0.25 and physical distance of 200kb for clumping to select independent SNPs. A significance threshold of 0.01 was used for index SNPs, with a secondary significance threshold of 0.5 for clumped SNPs. Scores were generated including and excluding the *APOE* region, defined as 1MB up and downstream of the gene (chromosome 19, position 44,409,039 to 46,412,650). The PRS for CSF A β_{42} was multiplied by -1 to align a higher risk score with decreased CSF A β_{42} (and increased brain A β_{42} concentration). Scores were rank inverse-normal transformed to place them on the same scale [41]. Individual scores were averaged to generate a composite A/T/N score.

All studies were approved by the institutional review board (IRB) of each participating location. Data sharing was carried out within the guidelines of IRB protocols.

2.7 Statistical Analyses

Data were analyzed using R (version 3.5.1, <https://www.r-project.org>). All associations were run on each of the four individual PRS (ie, clinical AD, $A\beta_{42}$, ptau, and LHIPV) and on the combined A/T/N PRS. Linear regression models assessing PRS score associations with CSF biomarkers (ptau₁₈₁, $A\beta_{42}$) covaried for age, sex, years of education, and CSF measurement batch. Linear regression models that evaluated PRS associations with left hippocampal volume (LHIPV) covaried for age, sex, years of education, baseline intracranial volume, and strength of magnet used for brain image acquisition. A binary logistic regression model assessed PRS associations with last available diagnosis (AD compared to NC, MCI diagnosed participants were excluded), and covaried for age, sex, and years of education.

We also assessed associations between PRS and baseline and longitudinal cognitive performance. A linear regression model covaried for age, sex, and years of education assessed associations between PRS and baseline cognition (memory and executive function). A mixed effects linear regression model assessed associations between PRS and longitudinal cognition (memory and executive function). Fixed effects in the model included age, sex, years of education and an interaction term for PRS x interval representing years between visits and baseline. Random effects included the intercept and interval term (slope expressed as years from the baseline visit). The pseudo R^2 reported was the marginal coefficient of determination associated with the fixed effects of the model. A Bonferroni correction was applied to account for all 40 models tested in this study (8 outcomes x 5 PRS scores) and results that remained statistically significant after correction are indicated.

PRS performance was measured using p-value significance and adjusted R^2 values to compare the variance explained across PRS and overall models. For mixed effects models, the marginal coefficient of determination is reported as pseudo R^2 . Sensitivity analyses were performed excluding individuals diagnosed with clinical AD or excluding clinically diagnosed AD

and MCI participants. Additionally, all PRS were generated including and excluding the *APOE* region. Sex interactions on all outcomes were also investigated, and if overall interactions were found, sex-stratified analyses were performed.

3. Results

3.1 PRS Validation

The predictive performance of PRS on CSF $A\beta_{42}$ were compared. Not surprisingly, the CSF $A\beta_{42}$ PRS was negatively associated with measured CSF $A\beta_{42}$ levels, indicating a higher CSF $A\beta_{42}$ PRS is associated with higher brain $A\beta_{42}$ burden. The CSF $A\beta_{42}$ PRS model explained 4.6% of the variance in CSF $A\beta_{42}$ levels (**Supplemental Table 2**, $p=4.1E-06$). The association remained significant after removing the *APOE* region ($p=6.5E-04$, $R^2=0.035$) and when restricting the sample to cognitively normal and MCI individuals ($p=3.7E-05$, $R^2=0.045$). Notably, the A/T/N score was the best predictor of CSF $A\beta_{42}$, with the model explaining 7.6% of the variance (**Supplemental Figure 1**, $p=6.8E-12$) and this score was predictive of CSF $A\beta_{42}$ in normal cognition controls ($p=1.1E-03$, $R^2=0.076$).

CSF Ptau PRS showed a significant positive association with CSF $ptau_{181}$ concentrations (**Supplemental Table 3**, $R^2=0.038$, $p=0.003$), however this association did not meet the Bonferroni threshold for all 40 models tested. This association remained significant after excluding the *APOE* region ($p=0.01$, $R^2=0.046$) and after exclusion of AD participants ($p=1.1E-03$, $R^2=0.059$). Again, the A/T/N PRS showed the strongest association with CSF $ptau_{181}$ concentrations compared to other scores, where a higher A/T/N PRS was associated with higher CSF ptau levels ($p=8.3E-06$, $R^2=0.051$).

We also validated the hippocampal volume PRS, whereby a higher PRS for left hippocampal volume was associated with greater left hippocampal volume at baseline and the model explained 22.2% of variance in the LHIPV data (**Supplemental Table 4**, $p=8.9E-04$).

Notably, this association was significant after excluding the *APOE* region. The PRS for clinical AD (calculated using a subset of the original IGAP data), and CSF A β_{42} PRS were also significantly associated with baseline left hippocampal volume. The clinical AD PRS score showed the strongest association and accounted for the highest variance in this outcome ($R^2=0.223$, $p=3.0E-04$), although all PRS had similar effect sizes (R^2 ranging from 0.215 to 0.223).

As expected, the clinical AD PRS showed the strongest association with AD diagnosis (**Supplemental Table 5**, $p=7.4E-08$, $R^2=0.111$) and remained significant after exclusion of the *APOE* region ($p=1.3E-03$, $R^2=0.079$). The ATN and CSF A β_{42} PRS were also significantly associated with AD diagnosis and remained significant after *APOE* region exclusion, however the CSF A β_{42} PRS association did not remain significant after Bonferroni correction.

3.2 PRS Association with Baseline Cognition

Higher A/T/N PRS and clinical AD PRS were significantly associated with lower baseline executive function in ADNI (**Table 2, Supplemental Figure 2**; A/T/N $p=0.004$, $R^2=0.115$; Clinical AD $p=0.02$, $R^2=0.112$), and these PRS models accounted for 11.2-11.4% of variability in baseline executive function but did not meet the Bonferroni correction threshold. These associations were not significant after exclusion of the *APOE* region but remained significant after exclusion of AD participants. It is noteworthy that the CSF A β_{42} PRS model also showed a nominal association with baseline executive function ($p=0.04$).

The clinical AD and A/T/N PRS were associated with baseline memory performance (**Table 3, Supplemental Figure 3**; A/T/N $p=0.048$, $R^2=0.093$; Clinical AD $p=6.8E-05$, $R^2=0.102$), and remained significant after excluding AD participants from the analysis. However, the associations were not statistically significant when excluding the *APOE* region and the A/T/N PRS associations did not meet the Bonferroni p-value threshold. LHIPV PRS was also nominally associated with baseline memory ($p=0.049$, $R^2=0.093$).

3.3 PRS Performance on Longitudinal Cognition

The A/T/N PRS showed the strongest association with longitudinal executive function, where a higher score was associated with a faster decline in executive function, and this association remained statistically significant when removing the *APOE* region or excluding AD participants (**Table 4, Figure 1**; A/T/N $p=4.6E-07$, $R^2=0.125$; A/T/N excluding *APOE* $p=2.4E-04$, A/T/N excluding AD $p=2.3E-07$). Clinical AD, CSF $A\beta_{42}$, and CSF ptau PRS models were also significantly associated with longitudinal executive function (Clinical AD $p=1.9E-04$, $R^2=0.116$; CSF $A\beta_{42}$ $p=0.005$, $R^2=0.118$; CSF ptau $p=3.6E-04$, $R^2=0.117$). However, no PRS was associated with change in executive function when restricting the sample to participants with normal cognition at baseline.

The A/T/N PRS also showed a significant association with longitudinal memory, where a higher score was associated with a greater rate of memory decline and the association remained statistically significant when removing the *APOE* region or removing AD participants from the analyses (**Table 5, Figure 2**; A/T/N $p=1.6E-09$, $R^2=0.118$; A/T/N excluding *APOE* $p=1.2E-05$, A/T/N excluding AD $p=1.5E-09$). Higher clinical AD, CSF $A\beta_{42}$, and CSF ptau PRS were significantly associated with greater decline in memory performance, none of which remained statistically significant when the sample was restricted to individuals with normal cognition at baseline.

In additional sensitivity analyses, we did not observe any sex interactions with PRS on the baseline or longitudinal cognition outcomes detailed above (**Supplemental Table 6**).

4. Discussion

We found that the A/T/N PRS with combined genetic risk for CSF ptau₁₈₁, CSF A β ₄₂, LHIPV, and clinical AD was a strong predictor of biomarker levels during the preclinical stages of AD, and a better predictor of longitudinal executive function compared to a PRS for late-onset clinical AD. The A/T/N PRS also performed comparably to the clinical AD PRS on baseline cognitive measures (memory, executive function) and longitudinal memory performance. Together, our findings suggest that utilizing the A/T/N framework by combining genetic risk for AD with genetic risk for AD biomarkers yields a better fitting prediction model, particularly in the earliest stages of disease.

As more GWAS data for AD endophenotypes have become available, it was important that we validate whether PRS for A/T/N are truly predictive of biomarkers in this independent dataset. All PRS were validated for their respective outcomes, and importantly, we found that combining PRS across A/T/N provided a more robust predictor of the individual AD biomarkers. It could be the case that genetic variants with small effects have functional impact across biomarkers, positioning a composite A/T/N score to capture more genetic overlap between biomarkers and amplify relevant signals for associations. As sample sizes continue to increase in genetic studies of amyloid, tau, and hippocampal atrophy in late-life, it is quite likely that the sensitivity of these PRS will continue to improve. Currently, the scores appear to provide a small boost in sensitivity to early deposition of pathology.

Similarly, the A/T/N PRS appeared to provide more sensitivity to risk of executive function decline, particularly when including participants across the clinical spectrum of AD. In contrast, the clinical AD PRS appeared to perform comparably to the A/T/N score in predicting memory performance, suggesting that the current polygenic predictors of clinical AD may be particularly sensitive to memory dysfunction. Given that many of the cohorts included in the AD GWAS studies come from memory clinics across the country, it is not surprising to see such robust associations with memory performance. However, when the *APOE* region is excluded

the A/T/N score remains significantly associated with memory decline while the clinical AD score does not, suggesting that the A/T/N score may provide genetic prediction above and beyond the *APOE* region in the context of memory decline. However, the clinical utility of the additional sensitivity provided by the A/T/N PRS in non-memory domains remains to be determined.

It is notable that the associations we observed with biomarkers of neuropathology and longitudinal cognitive decline explained variance above and beyond *APOE*, which has been a limitation of previous PRS analyses [42]. Certainly, *APOE* is a critical genetic component in AD, but AD remains a complex polygenic trait including many genetic effects with small effect sizes. Pooling across these small effects does appear to provide a score that shows more robust associations than reliance on the top genetic signal alone.

Previous work has shown that the clinical AD PRS derived from IGAP summary statistics was predictive of bilateral hippocampal volume in a Brazilian youth cohort [43], aligning with the clinical AD PRS association with left hippocampal volume demonstrated in this study. Together, these results may suggest that genetic drivers of hippocampal volume are influential throughout life. This association also fits with previous literature which reported a PRS for late-onset AD was associated with hippocampal function, as measured by functional magnetic resonance imaging (fMRI) [44]. Few studies to date have tested PRS associations with late life cognitive change, and results have been mixed [45, 46].

A strength of our chosen validation set was the wealth of clinical and genetic data including more than 1,100 participants. However, the GWAS summary statistics and the present analyses were sampled from highly-educated, Non-Hispanic White populations and generalizability of findings to more diverse groups is limited. PRS analyses can capture the common genetic risk for disease, however other disease contributors such as rare variants, gene-gene and gene-environment interactions may not be as accurately modeled. Larger sample sizes for GWAS of endophenotypes in particular would improve power of PRS models.

In their current form, PRS models of CSF biomarkers explain a small portion of the variance, suggesting substantial gains could be reached as sample sizes grow.

Overall, this study suggests an integration of genetic components in the A/T/N framework can be used to explain a greater percentage of variability in longitudinal cognition data compared to previously published genetic risk scores for late-onset clinical AD. Importantly, the calculated A/T/N PRS was significantly associated with longitudinal cognition in the absence of the *APOE* region, suggesting a greater degree of genetic risk for cognitive decline can be captured above and beyond *APOE*. Future PRS development should employ endophenotype-specific approaches to predict cognitive trajectories.

Table 1. Cohort demographics and summary statistics

Participants with genetic and cognitive data	Clinical Diagnosis			Total (N=1182)	P
	Normal Cognition (n=336)	Mild Cognitive Impairment (n=640)	Alzheimer's Disease (n=205)		
Age at baseline, years	75.3 ± 5.4	73.6 ± 7.5	75.6 ± 7.8	74.4 ± 7.1	0.72
Female, no. (%)	161 (48)	250 (39)	88 (43)	499 (42)	0.03
Education, years	16.3 ± 2.7	15.9 ± 2.9	15.0 ± 3.0	15.9 ± 2.9	7.4E-7
<i>APOE</i> -ε4 carrier	90 (27%)	313 (49%)	138 (67%)	541 (46%)	6.1E-24
Baseline Memory	1.0 ± 0.5	0.1 ± 0.7	-0.9 ± 0.6	0.2 ± 0.9	7.9E-178
Baseline Executive Function	0.8 ± 0.5	0.2 ± 0.8	-0.9 ± 0.6	0.2 ± 1.0	1.2E-97

Values are presented as mean±standard deviation, unless otherwise indicated. **Boldface** indicates $P < 0.05$.

Table 2. PRS associations with baseline executive function performance.

PRS Score	Beta	P	Adj R ²	No <i>APOE</i>	NC/MCI	NC
				P	P	P
A/T/N	-0.14	0.004	0.115	0.100	1.56E-04*	0.187
Clinical AD	-0.06	0.019	0.112	0.602	7.81E-04*	0.472
CSF AB₄₂	-0.05	0.044	0.111	0.187	0.066	0.524
CSF Ptau	-0.05	0.088	0.110	0.168	0.064	0.220
LHIPV	-0.01	0.763	0.108	0.769	0.210	0.767

Boldface indicates $P < 0.05$. * $P < 1.25E-03$ Bonferroni threshold. N=1,182 unless noted otherwise. No *APOE* denotes PRS excluding *APOE* region results. NC/MCI and NC indicate sensitivity analysis results with the denoted diagnostic groups (as assessed at baseline).

Table 3. PRS associations with baseline memory performance.

				No APOE	NC/MCI	NC
					N = 845	N = 336
PRS Score	Beta	P	Adj R²	P	P	P
A/T/N	-0.09	0.048	0.093	0.761	0.049	0.072
Clinical AD	-0.10	6.79E-05*	0.102	0.107	1.58E-05*	0.881
CSF AB ₄₂	-0.04	0.072	0.093	0.335	0.148	0.243
CSF Ptau	-0.01	0.593	0.090	0.970	0.990	0.127
LHIPV	0.05	0.049	0.093	0.050	0.163	0.187

Boldface indicates $P < 0.05$. * $P < 1.25E-03$ Bonferroni threshold. N=1,182 unless noted otherwise. No APOE denotes PRS excluding APOE region results. NC/MCI and NC indicate sensitivity analysis results with the denoted diagnostic groups (as assessed at baseline).

Table 4. PRS associations with longitudinal executive function performance.

				No APOE	NC/MCI	NC
					N = 845	N = 336
PRS Score	Beta	P	Pseudo R²	P	P	P
A/T/N	-0.06	4.62E-07*	0.125	2.40E-04*	2.27E-07*	0.455
Clinical AD	-0.02	1.93E-04*	0.116	0.083	6.90E-05*	0.794
CSF AB₄₂	-0.02	0.005	0.118	0.039	0.005	0.081
CSF Ptau	-0.02	3.61E-04*	0.117	0.002	9.00E-04*	0.689
LHIPV	-0.01	0.346	0.112	0.352	0.207	0.908

Boldface indicates $P < 0.05$. * $P < 1.25E-03$ Bonferroni threshold. N=1,182 unless noted otherwise. No APOE denotes PRS excluding APOE region results. NC/MCI and NC indicate sensitivity analysis results with the denoted diagnostic groups (as assessed at baseline).

Table 5. PRS associations with longitudinal memory performance.

				No APOE	NC/MCI	NC
					N = 845	N = 336
PRS Score	Beta	P	Pseudo R ²	P	P	P
A/T/N	-0.05	1.63E-09*	0.118	1.15E-05*	1.53E-09*	0.971
Clinical AD	-0.02	1.69E-05*	0.117	0.065	1.16E-05*	0.474
CSF AB ₄₂	-0.02	1.31E-03	0.110	0.020	1.47E-03	0.895
CSF Ptau	-0.02	2.55E-04*	0.108	1.3E-03	1.31E-03	0.790
LHIPV	-0.01	0.044	0.105	0.044	0.014	0.749

Boldface indicates $P < 0.05$. * $P < 1.25E-03$ Bonferroni threshold. $N=1,182$ unless noted otherwise. No APOE denotes PRS excluding APOE region results. NC/MCI and NC indicate sensitivity analysis results with the denoted diagnostic groups (as assessed at baseline).

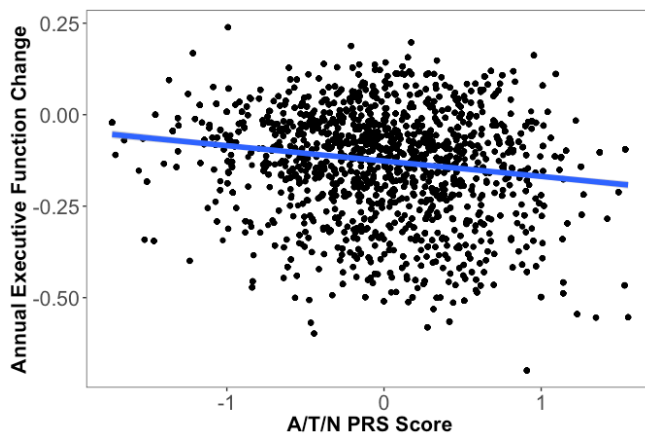


Figure 1. A/T/N PRS score association with annual change in executive function. The A/T/N model accounted for 12.5% of variability in annual executive function change and showed a significant association with longitudinal change in executive function ($p=4.62E-07$).

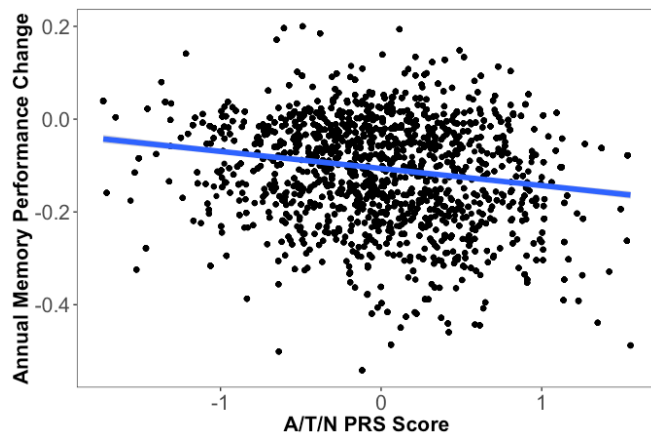


Figure 2. A/T/N PRS score association with annual change in memory performance. The A/T/N model accounted for 11.8% of variability in annual memory performance change, and showed a significant association with longitudinal change in memory performance ($p=1.63E-09$).

References:

- [1] Association As. 2018 Alzheimer's Disease Fact and Figures. *Alzheimers Dementia* 2018. p. 367-429.
- [2] Gatz M, Reynolds CA, Fratiglioni L, Johansson B, Mortimer JA, Berg S, et al. Role of Genes and Environments for Explaining Alzheimer Disease. *JAMA Psychiatry*. 2006;63:168-74.
- [3] Logue MW, Panizzon MS, Elman JA, Gillespie NA, Hatton SN, Gustavson DE, et al. Use of an Alzheimer's disease polygenic risk score to identify mild cognitive impairment in adults in their 50s. *Molecular Psychiatry*. 2019;24:421-30.
- [4] Jack CR, Jr., Knopman DS, Jagust WJ, Petersen RC, Weiner MW, Aisen PS, et al. Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol*. 2013;12:207-16.
- [5] Waldemar G, Phung KTT, Burns A, Georges J, Hansen FR, Iliffe S, et al. Access to diagnostic evaluation and treatment for dementia in Europe. *International Journal of Geriatric Psychiatry*. 2007;22:47-54.
- [6] DeCarli C. Mild cognitive impairment: prevalence, prognosis, aetiology, and treatment. *The Lancet Neurology*. 2003;2:15-21.
- [7] Louwersheimer E, Wolfsgruber S, Espinosa A, Lacour A, Heilmann-Heimbach S, Alegret M, et al. Alzheimer's disease risk variants modulate endophenotypes in mild cognitive impairment. *Alzheimer's & Dementia*. 2016;12:872-81.
- [8] Tan CH, Fan CC, Mormino EC, Sugrue LP, Broce IJ, Hess CP, et al. Polygenic hazard score: an enrichment marker for Alzheimer's associated amyloid and tau deposition. *Acta Neuropathologica*. 2018;135:85-93.
- [9] Escott-Price V, Shoai M, Pither R, Williams J, Hardy J. Polygenic score prediction captures nearly all common genetic risk for Alzheimer's disease. *Neurobiology of Aging*. 2017;49:214.e7-.e11.
- [10] Escott-Price V, Myers AJ, Huentelman M, Hardy J. Polygenic risk score analysis of pathologically confirmed Alzheimer disease. *Annals of Neurology*. 2017;82:311-4.
- [11] Escott-Price V, Sims R, Bannister C, Harold D, Vronskaya M, Majounie E, et al. Common polygenic variation enhances risk prediction for Alzheimer's disease. *Brain*. 2015;138:3673-84.

- [12] Chaudhury S, Patel T, Barber IS, Guetta-Baranes T, Brookes KJ, Chappell S, et al. Polygenic risk score in postmortem diagnosed sporadic early-onset Alzheimer's disease. *Neurobiology of Aging*. 2018;62:244.e1-e8.
- [13] Torkamani A, Wineinger NE, Topol EJ. The personal and clinical utility of polygenic risk scores. *Nature Reviews Genetics*. 2018;19:581-90.
- [14] Desikan RS, Fan CC, Wang Y, Schork AJ, Cabral HJ, Cupples LA, et al. Genetic assessment of age-associated Alzheimer disease risk: Development and validation of a polygenic hazard score. *PLOS Medicine*. 2017;14:e1002258.
- [15] Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, et al. Genome-wide association study identifies variants at *CLU* and *PICALM* associated with Alzheimer's disease. *Nat Genet*. 2009;41:1088-93.
- [16] Jansen IE, Savage JE, Watanabe K, Bryois J, Williams DM, Steinberg S, et al. Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk. *Nature Genetics*. 2019;51:404-13.
- [17] Stocker H, Möllers T, Perna L, Brenner H. The genetic risk of Alzheimer's disease beyond *APOE* ϵ 4: systematic review of Alzheimer's genetic risk scores. *Translational Psychiatry*. 2018;8:166.
- [18] Mantzavinos V, Alexiou A. Biomarkers for Alzheimer's Disease Diagnosis. *Current Alzheimer research*. 2017;14:1149-54.
- [19] Jack CR, Jr., Bennett DA, Blennow K, Carrillo MC, Dunn B, Haeberlein SB, et al. NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 2018;14:535-62.
- [20] Deming Y, Li Z, Kapoor M, Harari O, Del-Aguila JL, Black K, et al. Genome-wide association study identifies four novel loci associated with Alzheimer's endophenotypes and disease modifiers. *Acta neuropathologica*. 2017;133:839-56.
- [21] Elliott LT, Sharp K, Alfaro-Almagro F, Shi S, Miller KL, Douaud G, et al. Genome-wide association studies of brain imaging phenotypes in UK Biobank. *Nature*. 2018;562:210-6.
- [22] Foley SF, Tansey KE, Caseras X, Lancaster T, Bracht T, Parker G, et al. Multimodal Brain Imaging Reveals Structural Differences in Alzheimer's Disease Polygenic Risk Carriers: A Study in Healthy Young Adults. *Biological Psychiatry*. 2017;81:154-61.
- [23] Jagust WJ, Landau SM, Shaw LM, Trojanowski JQ, Koeppe RA, Reiman EM, et al. Relationships between biomarkers in aging and dementia. *Neurology*. 2009;73:1193-9.
- [24] Shaw LM, Vanderstichele H, Knapik-Czajka M, Figurski M, Coart E, Blennow K, et al. Qualification of the analytical and clinical performance of CSF biomarker analyses in ADNI. *Acta neuropathologica*. 2011;121:597-609.
- [25] McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR, Kawas CH, et al. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's & Dementia*. 2011;7:263-9.
- [26] Folstein MF, Folstein SE, McHugh PR. "Mini-mental state": A practical method for grading the cognitive state of patients for the clinician. *Journal of Psychiatric Research*. 1975;12:189-98.
- [27] McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease. *Neurology*. 1984;34:939.
- [28] Jack CR, Jr., Bernstein MA, Fox NC, Thompson P, Alexander G, Harvey D, et al. The Alzheimer's Disease Neuroimaging Initiative (ADNI): MRI methods. *J Magn Reson Imaging*. 2008;27:685-91.
- [29] Dale AM, Fischl B, Sereno MI. Cortical Surface-Based Analysis: I. Segmentation and Surface Reconstruction. *NeuroImage*. 1999;9:179-94.
- [30] Fischl B, Sereno MI, Dale AM. Cortical Surface-Based Analysis: II: Inflation, Flattening, and a Surface-Based Coordinate System. *NeuroImage*. 1999;9:195-207.

- [31] Fischl B, Sereno M, Tootell R, Dale A. High-resolution intersubject averaging and a coordinate system for the cortical surface. *Human Brain Mapping*. 1999;8:272-84.
- [32] Mormino EC, Kluth JT, Madison CM, Rabinovici GD, Baker SL, Miller BL, et al. Episodic memory loss is related to hippocampal-mediated beta-amyloid deposition in elderly subjects. *Brain*. 2009;132:1310-23.
- [33] Mormino EC, Kluth JT, Madison CM, Rabinovici GD, Baker SL, Miller BL, et al. Episodic memory loss is related to hippocampal-mediated β -amyloid deposition in elderly subjects. *Brain*. 2008;132:1310-23.
- [34] Desikan RS, Ségonne F, Fischl B, Quinn BT, Dickerson BC, Blacker D, et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. *NeuroImage*. 2006;31:968-80.
- [35] Gibbons LE, Carle AC, Mackin RS, Harvey D, Mukherjee S, Insel P, et al. A composite score for executive functioning, validated in Alzheimer's Disease Neuroimaging Initiative (ADNI) participants with baseline mild cognitive impairment. *Brain imaging and behavior*. 2012;6:517-27.
- [36] Crane PK, Carle A, Gibbons LE, Insel P, Mackin RS, Gross A, et al. Development and assessment of a composite score for memory in the Alzheimer's Disease Neuroimaging Initiative (ADNI). *Brain imaging and behavior*. 2012;6:502-16.
- [37] Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81:559-75.
- [38] Raj A, Stephens M, Pritchard JK. fastSTRUCTURE: Variational Inference of Population Structure in Large SNP Data Sets. *Genetics*. 2014;197:573.
- [39] Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nature genetics*. 2013;45:1452-8.
- [40] Kunkle BW, Grenier-Boley B, Sims R, Bis JC, Damotte V, Naj AC, et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates A β , tau, immunity and lipid processing. *Nature Genetics*. 2019;51:414-30.
- [41] Blom G. Transformations of the Binomial, Negative Binomial, Poisson and χ^2 Distributions. *Biometrika*. 1954;41:302-16.
- [42] Darst BF, Kosciak RL, Racine AM, Oh JM, Krause RA, Carlsson CM, et al. Pathway-Specific Polygenic Risk Scores as Predictors of Amyloid- β Deposition and Cognitive Function in a Sample at Increased Risk for Alzheimer's Disease. *Journal of Alzheimer's disease : JAD*. 2017;55:473-84.
- [43] Axelrud LK, Santoro ML, Pine DS, Talarico F, Gadelha A, Manfro GG, et al. Polygenic Risk Score for Alzheimer's Disease: Implications for Memory Performance and Hippocampal Volumes in Early Life. *Am J Psychiatry*. 2018;175:555-63.
- [44] Xiao E, Chen Q, Goldman AL, Tan HY, Healy K, Zoltick B, et al. Late-Onset Alzheimer's Disease Polygenic Risk Profile Score Predicts Hippocampal Function. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*. 2017;2:673-9.
- [45] Harris SE, Davies G, Luciano M, Payton A, Fox HC, Haggarty P, et al. Polygenic Risk for Alzheimer's Disease is not Associated with Cognitive Ability or Cognitive Aging in Non-Demented Older People. *Journal of Alzheimer's Disease*. 2014;39:565 - 74.
- [46] Marden JR, Mayeda ER, Walter S, Vivot A, Tchetgen Tchetgen EJ, Kawachi I, et al. Using an Alzheimer Disease Polygenic Risk Score to Predict Memory Decline in Black and White Americans Over 14 Years of Follow-up. *Alzheimer Dis Assoc Disord*. 2016;30:195-202.