

## **BowSaw: inferring higher-order trait interactions associated with complex biological phenotypes**

Demetrius DiMucci<sup>a,b</sup>, Mark Kon<sup>a,c</sup>, Daniel Segrè<sup>a,b,d, #</sup>

<sup>a</sup> Bioinformatics Graduate Program, Boston University, Boston, Massachusetts, USA

<sup>b</sup> Biological Design Center, Boston University, Boston, Massachusetts, USA

<sup>c</sup> Department of Mathematics and Statistics, Boston University, Boston, Massachusetts, USA

<sup>d</sup> Department of Biology, Department of Biomedical Engineering, Department of Physics, Boston University, Boston, Massachusetts, USA

# Correspondence to Daniel Segrè, [dsegre@bu.edu](mailto:dsegre@bu.edu)

1 **Abstract**

2 Machine learning is helping the interpretation of biological complexity by enabling the  
3 inference and classification of cellular, organismal and ecological phenotypes based on  
4 large datasets, e.g. from genomic, transcriptomic and metagenomic analyses. A number  
5 of available algorithms can help search these datasets to uncover patterns associated with  
6 specific traits, including disease-related attributes. While, in many instances, treating an  
7 algorithm as a black box is sufficient, it is interesting to pursue an enhanced  
8 understanding of how system variables end up contributing to a specific output, as an  
9 avenue towards new mechanistic insight. Here we address this challenge through a suite  
10 of algorithms, named BowSaw, which takes advantage of the structure of a trained  
11 random forest algorithm to identify combinations of variables (“rules”) frequently used  
12 for classification. We first apply BowSaw to a simulated dataset, and show that the  
13 algorithm can accurately recover the sets of variables used to generate the phenotypes  
14 through complex Boolean rules, even under challenging noise levels. We next apply our  
15 method to data from the integrative Human Microbiome Project and find previously  
16 unreported high-order combinations of microbial taxa putatively associated with Crohn’s  
17 disease. By leveraging the structure of trees within a random forest, BowSaw provides a  
18 new way of using decision trees to generate testable biological hypotheses.

19

20

21

22

## 23 **Introduction**

24           The production of large biological data sets with high-throughput techniques has  
25 increased the utilization of supervised machine learning algorithms to produce  
26 predictions of complex phenotypes (e.g. healthy vs. disease) from measurable traits.  
27 These algorithms use measurements of relevant traits such as gene variants, the  
28 presence/absence of microbial taxa, or metabolic consumption variables as predictors.  
29 Categorical prediction of phenotypes is typically the end goal of these applications.  
30 However, an additional benefit of these algorithms is the potential to extract explanatory  
31 classification rules. In this context, a rule is defined as a Boolean function of a set of  
32 traits, such that the value of the function is 1 (true) when the traits are associated with a  
33 given phenotype. Identifying the relationships between the traits involved in  
34 classification rules may yield key insights into the biological processes associated with  
35 important phenotypes [1, 2]. This realization is creating demand for methods that assist in  
36 the interpretation of supervised machine learning methods [3–5], especially when the  
37 measured traits may be causal agents of disease states, such as genetic variants or  
38 microbial taxa [6]. Identifying classification rules associated with a phenotype of interest  
39 is valuable because these rules are likely to carry information about the causal  
40 mechanisms that generate the phenotype.

41           Algorithms that are particularly valuable in this respect are those involving  
42 decision trees, such as random forests, since decision trees are easily interpretable [7].  
43 Decision trees are rule-based classifiers, where rules arise from a series of “yes-no”  
44 questions that can efficiently divide the data into categorical groups. In a biological

45 context, such rules may arise from sets of genes whose simultaneous modulation could  
46 affect a phenotype, or sets of microbial species whose co-occurrence may be associated  
47 with a disease state. While in several cases it seems like disease phenotypes are uniquely  
48 associated with a single specific pattern (e.g. retinoblastoma [8]), there is increasing  
49 evidence for cases in which multiple distinct patterns can be associated with (and  
50 potentially causing) the same high-level phenotype [9, 10]. A particular example we will  
51 explore in this work is the multiplicity of distinct microbial presence/absence patterns  
52 which may be associated with Crohn's disease [11]. Crohn's disease has five clinically  
53 defined sub-types [12] but studies of the associated microbiome do not usually indicate  
54 which form of Crohn's disease a donor has been diagnosed with. Each sub-type of the  
55 disease may be associated with different microbes, each requiring different treatment  
56 regimes. Thus, identifying rules associated with sub-populations within a given  
57 phenotype label are of great interest due to potential therapeutic implications.

58         The fact that there may be multiple etiologies that generate the same or similar  
59 phenotypes complicates the straightforward interpretation of parameter coefficients or  
60 variable importance scores [13, 14]. Uncovering the multiple interactions between  
61 predictive variables as they relate to phenotypic labels remains a challenging statistical  
62 endeavor, but one that is of paramount importance. Identifying the associated rules that a  
63 random forest uses to classify a given sample as having a particular disease enables the  
64 development of mechanistic hypotheses for follow up-studies. This challenge, and an  
65 overview of the key strategy we propose, are illustrated in Figure 1. In figure 1A we  
66 depict a toy model where measured variables (traits) have only two possible values (e.g.:

67 present/absent), the high-level phenotype (category) is binary (e.g.: no disease/disease),  
68 and two distinct Boolean rules can both generate the phenotype. The goal in this case is  
69 to identify each of the rules that are associated with the phenotype. The multiple Boolean  
70 rules obtained in this manner can be thought of as a consensus decision tree that  
71 possesses the most informative branches of the forest with respect to a given class label.  
72 In this work, we will show how this can be achieved by in-depth analyses of any given  
73 random forest (RF) (Fig. 1B).

74         The random forest algorithm intrinsically takes advantage of non-linear  
75 relationships between variables and is widely used in the life sciences [15–17]. RFs,  
76 when used to distinguish between disease states known to have multiple causes, often  
77 result in excellent classifiers [18, 19]. It has also been reported that RFs capture subtle  
78 statistical interactions between variables [13]. Unfortunately, an RF is not  
79 straightforwardly interpretable despite its hierarchical structure, and recovering those  
80 interactions is notoriously difficult [14] due in large part to the method’s reliance on  
81 ensembles of trees [20]. The difficulties in interpretation created by these properties has  
82 led many to refer to RF as a ‘black box’ model [21].

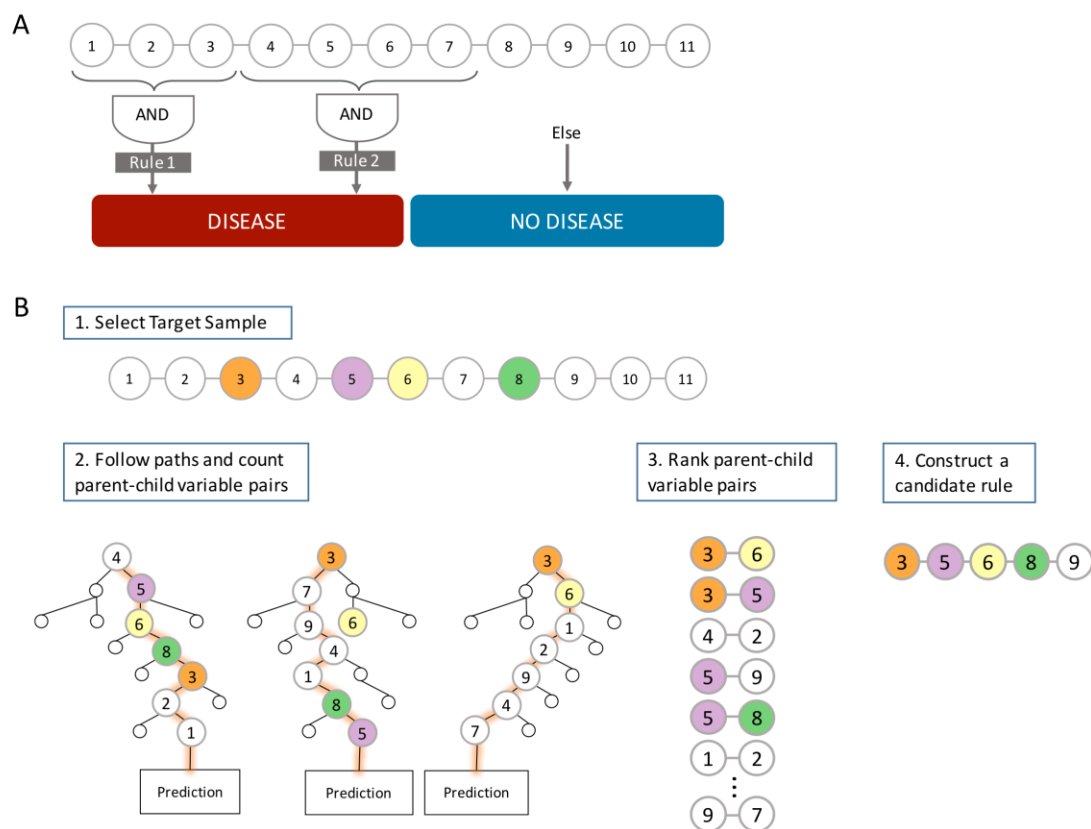
83         Identifying the rules that a RF utilizes in classification tasks is an active area of  
84 research, and many strategies have been developed to address this problem. Effective  
85 strategies have focused on evaluating how individual variables influence the  
86 classification probabilities of specific samples [22, 23], pruning existing decision rules  
87 found in the tree ensemble to produce compact models [24], computing conditional  
88 importance scores [25], or iteratively enriching the most prevalent variable co-

89 occurrences through regularization [26]. These approaches offer valuable methods for the  
90 identification of statistical interactions between variables. However, we and others have  
91 observed that while these methods are capable of recovering a true causal rule in  
92 simulated data when exactly one such rule is present, the existence of multiple rules  
93 associated with one phenotype can confound interpretation efforts [26].

94         Here we describe BowSaw, a new set of algorithms that utilizes variable  
95 interactions in a trained RF model in order to extract multiple candidate explanatory  
96 rules. With BowSaw, we set out to develop a *post hoc* method intended to aid in the  
97 discovery of these rules when the input variables are categorical in nature. The primary  
98 approach of BowSaw is to start by approximating a best combination of variables (i.e. a  
99 rule) that explain the forest's predictions for individual instances of a given class in the  
100 data set and then to curate the collection of best combinations to obtain a concise set of  
101 combinations that collectively segregate a class of interest with high precision. For  
102 individual instances a rule is identified by systematically quantifying the co-occurrence  
103 of specific variable pairs across trees in the forest that attempt to predict the class of the  
104 instance (out-of-bag trees) and then using the frequency of co-occurring variable pairs to  
105 guide the construction of a rule that precisely identifies the instance as its observed class.  
106 For the entire set of instances, we then curate the collection of all rules identified this way  
107 in order to produce a small set of rules that are broadly and precisely applicable to  
108 instances of the given class label.

109         We first demonstrate that BowSaw can recover true rules by applying the  
110 algorithms to simulated data sets of varying complexity. We then apply BowSaw to a

111 study on the role of the gut microbiome on Crohn’s disease [11], and show that it can find  
 112 a previously unreported combination of microbial taxa that is broadly and precisely  
 113 associated with Crohn’s disease instances in the data set. In its current implementation  
 114 BowSaw can be applied to any dataset with categorical or discrete predictors with any  
 115 number of class labels.



116 **A** In a hypothetical dataset there may be two phenotype labels – “Disease” and “No  
 117 Disease”, that we wish to discriminate based on input predictor variables. In this  
 118 example, there are two distinct high-order patterns that both confer the “Disease”  
 119 phenotype. Our goal is to identify a potentially diverse set of patterns (or, in this  
 120 simplified case, all patterns) that are associated with the “Disease” label. **B** Conceptual  
 121 pipeline of BowSaw. In (1) we begin by identifying the vector of a target instance that  
 122 has the target observed label. In this example, the colored nodes indicate a true associated  
 123 pattern, which is unknown to us. In (2) we follow the path of the instance through each of  
 124 its out-of-bag trees and record how often the sample encounters sequential pairs of  
 125 variables. (3) Each ordered pair sequence is sorted in descending order by its observed  
 126

127 frequency. (4) Starting from the top of the list, pair sequences are iteratively evaluated  
128 and added to an undirected network of variables (i.e. a candidate rule) until this network  
129 is maximally associated with the observed phenotype of the target vector or the list of  
130 ordered pairs is exhausted. Each sample with the label of interest yields one such  
131 candidate rule. These rules are then aggregated and curated to obtain a concise set of  
132 rules that explain class-specific classification decisions that occur in the forest.  
133

## 134 **Methods**

### 135 Overview of the pipeline

136        Provided with a trained random forest and a training set, BowSaw goes through  
137 three steps in order to generate a candidate rule (variable-value combination) for each  
138 observation associated with the phenotype of interest. First, for a specific observation, the  
139 *Count* algorithm counts the frequency of unique ordered pairs of variables encountered  
140 along each of its out-of-bag trees in the forest (Figure 1B – step 2). Second, for that  
141 observation, the *Construct* algorithm takes the counts from the first step and generates a  
142 list of ordered pairs, ranked by their frequencies, then uses this list as a guide to construct  
143 a candidate decision rule (which could consist of two or more variables) that is  
144 maximally associated with the observed phenotype (Figure 1B – steps 3 - 4). Finally, the  
145 *Curate* algorithm pools the candidate decision rules from each observation together in  
146 order to select a subset of rules that collectively account for all of the samples with the  
147 desired phenotype (Figure 1B – step 5). Optionally, the *Sub-rule* algorithm can be used to  
148 generate pruned versions of candidate rules prior to applying the Curate algorithm in  
149 order to obtain a more concise, albeit less specific, set of candidate rules. The Count and  
150 Curate algorithms generate the candidate rules for individual observations while the



151 Curate and Sub-rule algorithms produce a combined set of rules that account for all  
152 observations with the chosen phenotype.

153 In the following section, we provide a description of the inputs BowSaw takes and  
154 the algorithms that implement these steps along with pseudocode.

155

### 156 Inputs

157 BowSaw takes as inputs a dataset,  $\mathbf{D}$ , composed of  $N$  observed vectors  $\mathbf{x}_i$   
158 (together with their respective classes  $k_i$ ) each of  $p$  categorical variables. There are  
159 assumed to be  $K$  possible class labels for each vector in  $\mathbf{D}$  which for the purposes of this  
160 discussion denote different phenotypes. A random forest is assumed to be trained on  $\mathbf{D}$  to  
161 distinguish the classes  $k = 1, \dots, K$ . Additionally, BowSaw takes as input the feature  
162 vector  $\mathbf{x}_i$  of a specific observation for which the goal is to identify a set of simplified  
163 rules associated with the phenotype  $k_i$ .

164

### 165 Counting stubs

166 Given an RF machine  $\mathbf{M}$  trained on dataset  $\mathbf{D}$  and a feature vector  $\mathbf{x} = (x_1, x_2, \dots, x_p) \in$   
167  $\mathbf{D}$ , the first sub-routine of our method (the *count algorithm*) proceeds as follows. It starts  
168 by identifying among the set of trees in  $\mathbf{M}$ , those sub-paths (sequences of successive  
169 variable indices) encountered by sample  $\mathbf{x}$  as it travels through  $\mathbf{M}_x$ , its set of out-of-bag  
170 trees. An out-of-bag tree is a tree for which  $\mathbf{x}$  was not included in the training set. For a  
171 specific path  $\mathbf{P}$  in  $\mathbf{M}_x$  the sequence of successive variable indices forms a vector  $\mathbf{v} =$   
172  $(v_1, \dots, v_r)$  (note that each  $v_j$  is one of the variables  $x_j$ ). Each stub (ordered pair of

173 sequentially encountered variables  $v_i v_{i+1}$ ) in all out-of-bag along  $\mathbf{P}$  for  $i = 1, \dots, r-1$  is  
174 accounted for in a  $p \times p$  matrix  $\mathbf{C}^x$ , where the element  $C_{ij}^x$  records the number of stubs  
175 containing the ordered pair of variables  $x_i$  and  $x_j$  among all paths of  $\mathbf{M}_x$ .

176

177 **Algorithm 1: Count Algorithm Pseudocode**

178 Initialize  $\mathbf{C}^x$  as a  $p \times p$  matrix of zeros.

179 For each path  $\mathbf{P}$  with feature indices  $\mathbf{v}$  in  $\mathbf{M}_x$  do:

180     For  $i = 1, \dots, r - 1$ ,

181         
$$C_{v_i, v_{i+1}}^x = C_{v_i, v_{i+1}}^x + 1$$

182     End loop

183 End loop

184 Return  $\mathbf{C}^x$ .

185 For simplicity, henceforth we will denote  $\mathbf{C} = \mathbf{C}^x$ , remembering that  $\mathbf{C}$  continues to  
186 depend on the fixed sample  $\mathbf{x}$ .

187

188 **Constructing a candidate rule**

189 A rule for classifying to a test point  $\mathbf{x}$  will have the form “ $\mathbf{x}_I = \mathbf{a}_I$  implies  $\mathbf{x}$  is in class

190  $k$ ”. Here  $\mathbf{I}$  is a designated subcollection of the variable indices  $i = 1, \dots, p$ , and  $\mathbf{x}_I =$

191  $(x_{i_1}, \dots, x_{i_{|\mathbf{I}|}})$  is the sub-vector of current vector  $\mathbf{x} = (x_1, \dots, x_p)$  corresponding just to the

192 indices  $i_j \in \mathbf{I}$ . The vector  $\mathbf{a}_I = (a_{i_1}, \dots, a_{i_{|\mathbf{I}|}})$  will denote an assigned set of values to the

193  $x_i$ , i.e., so that  $x_i = a_i$  for  $i \in \mathbf{I}$ . Thus the condition  $\mathbf{x}_I = \mathbf{a}_I$  means assignment of values

194 to  $x_i$  for  $i \in I$ . The rule is that if training vector  $\mathbf{x}$  satisfies  $\mathbf{x}_I = \mathbf{a}_I$ , we classify  $\mathbf{x}$  into  
195 category  $k$ .

196

197 The second sub-routine (the *construct algorithm*) builds a candidate rule  $\mathbf{R}$ , based  
198 (initially) on a fixed training point, say  $\mathbf{a} \in \mathbf{D}$ , in class  $k$ . This is done by first placing all  
199 of the stubs  $(i, j)$  with non-zero counts  $\mathbf{C}_{ij}$  into a list  $\mathbf{L}$  sorted in descending order by their  
200 values in  $\mathbf{C}$ .

201

202 We define the candidate rule  $\mathbf{R}$  (based on  $\mathbf{a}$ ) through the following steps. We initialize  
203 using the first stub  $L_1 = (i_1, j_1)$  in the list  $\mathbf{L}$ , together with the two fixed values  $x_{i_1} =$   
204  $a_{i_1}$ ,  $x_{j_1} = a_{j_1}$ . This is the initialized form of the rule  $\mathbf{R}$ , which requires that for any test  
205 vector, its values at the above indices  $i_1$  and  $j_1$  match the values

206 of the above fixed training vector  $\mathbf{a} \in \mathbf{D}$ , so that  $x_{i_1} = a_{i_1}$ , and  $x_{j_1} = a_{j_1}$ . For brevity,  
207 denote the pair  $(i_1, j_1) = I_1$  and the corresponding assigned values as  $(a_{i_1}, a_{j_1}) = \mathbf{a}_{I_1}$ .

208 Then the content of rule  $\mathbf{R}$  will be denoted succinctly as  $\mathbf{R}: \mathbf{x}_I = \mathbf{a}_I \Rightarrow \text{class } k$ . Since  
209 ordering of the indices  $i_1, j_1$  does not matter, (as long as the indices are identified), we  
210 will henceforth write  $(i_1, i_2) \rightarrow \{i_1, i_2\}$ .

211 We then update rule  $\mathbf{R}$  as follows. We find all  $\mathbf{x} \in \mathbf{D}$  that satisfy the initial part of rule  $\mathbf{R}$ ,  
212 i.e.,  $\mathbf{x}_I = \mathbf{a}_I$  i.e., all training points matching the two indices  $\{i_1, j_1\}$  of training sample  $\mathbf{a}$ ,  
213 and store them as a subcollection  $\mathbf{D}_1 \subset \mathbf{D}$  of the training set. We call  $F$  the fraction of  
214 data points in  $\mathbf{D}_1$  that have phenotype  $k$ , i.e., match the phenotype of the initial sample  
215  $\mathbf{a} \in \mathbf{D}$ . If  $F = 1$ , we stop and return the current above rule  $\mathbf{R}$ . If  $F < 1$ , we continue by

216 choosing the second stub  $L_2 = \{i_2, j_2\}$  in the above list  $L$ , and augment the current rule  $R$   
217 by adding the condition  $x_{i_2} = a_{i_2}, x_{j_2} = a_{j_2}$  (again written  $x_{I_2} = \mathbf{a}_{I_2}$ ) and maintaining the  
218 assignment of class  $k$  (i.e., the same class as the currently fixed sample  $\mathbf{a} \in \mathbf{D}$ ). If the  
219 second stub  $L_2$  happens to overlap with the initial stub  $L_1$ , this added condition in the rule  
220  $R$  will clearly be consistent, being still based on the fixed sample  $\mathbf{a}$ . We augment the  
221 current index list  $I_1$  to a list  $I_2$ , adding to it the two new indices  $i_2$  and  $j_2$ , so that now  
222  $I_2 = \{i_1, j_1, i_2, j_2\}$  writing the augmented rule as  $R: \mathbf{x}_{I_2} = \mathbf{a}_{I_2} \Rightarrow \text{class } k$ . Again  
223 defining  $F$  to be the fraction of the data subset  $\mathbf{D}_2$  (matching the more restrictive new  
224 rule  $R$ ) with phenotype  $k$ , we stop the algorithm and use the current rule  $R$  if  $F = 1$ , and  
225 otherwise augment rule  $R$  by adding the indices  $L_3 = (i_3, j_3)$  to it, as above, yielding a  
226 larger set  $I_3$  of indices and the augmented rule  $R: \mathbf{x}_{I_3} = \mathbf{a}_{I_3} \Rightarrow \text{class } k$ , with a more  
227 restricted subset  $\mathbf{D}_3 \subset \mathbf{D}$ , and a new value for  $F$ , now the fraction of  $\mathbf{D}_3$  in the class  $k$  of  
228 the fixed  $\mathbf{a} \in \mathbf{D}$ .  
229 This process continues until the fraction  $F = 1$ , i.e., 100% of the samples in  $\mathbf{D}$  match the  
230 current set of indices, and also match the class  $k$  of the current sample  $\mathbf{a}$ . Alternatively,  
231 the algorithm stops when all stubs in  $L$  have been exhausted.

232

233 **Algorithm 2: Construct Algorithm Pseudocode**

234 Make ranked list  $L$  of stubs from  $C$

235 Initialize fixed  $\mathbf{a} \in \mathbf{D}$ ,  $R = \phi$   $I = \phi$ ,  $F = 0$ ,

236 For  $i = 1: |L|$ , select stub  $L_i$

237       If  $F = 1$ :

238                   Exit loop

239           Else:

240                    $I' = \{I \cup L_i\}$

241                    $D_{I'} = \{x \in D: x_{I'} = a_{I'}\}$

242                    $F' = \frac{|\{x \in D_{I'}: \text{class } x=k\}|}{|D_{I'}|}$

243                   If  $F' > F$ :

244                                  $I = I'$

245                                  $F = F'$

246   End loop

247   Return  $I, F, D_I$  [all corresponding to the fixed  $a \in D$ ].

248   Return rule  $R: x_I = a_I \Rightarrow \text{class } k$

250   **Curating candidate rules:**

251           The *count* and *construct* algorithms are the heart of BowSaw. In our workflow,

252   we apply these algorithms to each observation  $a \in D$  that has the desired observed

253   phenotype  $k$ . We call the set of these vectors  $D^k \subset D$ . By default, we produce a single

254   candidate rule for each vector in  $a \in D^k$ . We store each candidate rule in list  $Q$  and rank

255   them by their respective values of  $|I|$ , i.e., the number of indices in the respective rules.

256   Since  $Q$  may include many redundant rules, we developed another sub-routine (the *curate*

257   *algorithm*) to generate a concise set of candidate rules that collectively account for all

258   data vectors  $D^k$  in class  $k$ . Briefly, we initialize an empty list  $E$ , to which we add the top

259 ranked rule from  $Q$  (by default this is the rule with the greatest value of  $|I|$ ), and record  
260 the index of samples in  $D$  that match any rule in  $E$  and also have the desired observed  
261 phenotype class  $k$ , into a set  $A$ . Next, we determine how many samples remain  
262 unaccounted for, i.e. are in  $U = D^k \sim A$ , Then we determine which of the remaining rules  
263 in  $Q$  minimizes  $|U|$ , add it to  $E$ , and repeat these steps until  $U$  is an empty set.

264

265 **Algorithm 3: Curate algorithm pseudocode**

266  $Q$  = ranked list of all candidate rules for  $\Phi_t$

267  $E = Q_{best}$  (user defined, default is maximum  $M$ )

268  $I^*$  = which  $D$  match any rule in  $E$  and  $k = K_d$

269  $A = D^k \cap M^*$

270  $U = D^k - A$

271 While  $U$  is not empty:

272      $B = \{ \}$

273     For rule  $i$  in  $Q$ :

274          $E^* = E + Q_i$

275          $I^*$  = which  $D$  match any rule in  $E^*$  and  $k = K_d$

276          $A^* = D^k \cap I^*$

277          $B_i = |U - A^*|$

278     End loop

279      $best = \text{which min } B_i$

280      $E = E + Q_{best}$

281  $M^* =$  which  $D$  match any rule in  $E$  and  $k = K_d$

282  $A = D^k \cap M^*$

283  $U = U - A$

284 End while loop

285 Return  $E$

286

287 **Constructing sub-rules**

288 Since rules are rarely 100% associated with any given phenotype, we devised a  
289 strategy for selecting a set of candidate sub-rules that account for all samples with desired  
290 observed phenotype class  $k$ . Candidate sub-rules are shorter candidate rules derived from  
291 larger candidate rules by omitting one or more variables. For each candidate rule in  $E$ , we  
292 identify sub-rules that meet a user-defined complexity criteria, e.g. only produce sub-  
293 rules that are composed of three or four variables and their corresponding values. We  
294 place each of the unique sub-rules into a new list  $E_{sub}$ . Then the corresponding number of  
295 identical matches,  $I$ , and proportion of  $I$  that have the phenotype  $K_d$ ,  $F$ , are determined.  
296 At this stage, we can apply our third sub-routine (the *Curate* algorithm) to  $E_{sub}$  to obtain a  
297 parsimonious list of sub-rules that accounts for  $\mathbf{x}_{all}$ . In our pipeline, we also choose  
298 thresholds based on desired levels of  $I$  and/or  $F$  in order to eliminate poor candidate sub-  
299 rules from consideration. In this study, we decided on the thresholds after visually  
300 inspecting a plot of  $F$  against  $I$ .

301

302 **Algorithm 4: Sub-rule algorithm pseudocode**

303  $E_{sub} = \{ \}$

304  $Complexity = \{ \text{user defined numeric values} \}$

305 For  $rule$  in  $E$

306     For  $i$  in  $Complexity$

307          $E_{sub} = E_{sub} \cup \left( \frac{rule}{i} \right)$

308     End loop

309 End loop

310

311     The algorithms described above are generalizable to multi-classification tasks but  
312 are currently limited to discretized or categorical representations of the feature space.

313 Pseudocode for implementing each of the algorithms described above along with an  
314 implementation of the algorithms in R [27] can be found in the supplemental files and on  
315 github: <https://github.com/ddimucci/BowSaw>.

316

317

## 318 **Results**

### 319 **Application to simulated Data**

320     To test the capacity of BowSaw to recover multiple decision rules, we applied it  
321 to increasingly challenging simulated data sets. These data set consists of binary vectors  
322 representing different observations. The phenotype associated with each observation is a  
323 function of the corresponding vector. The function consists of a set of multiple mutually  
324 distinct Boolean rules, such that if a rule is satisfied, it will cause the observation to have



325 the phenotype with a certain probability (which we call here “penetrance” because of its  
326 resemblance to the genetics concept). The first dataset (IDEALIZED) we use is relatively  
327 simple, and includes multiple equally prevalent rules. It is also generated under the  
328 assumption that there are no unmeasured confounders, i.e. that if an observation does  
329 have a phenotype, then it must be satisfying at least one of the above rules. We then  
330 apply BowSaw to a more challenging scenario (INTERMEDIATE) in which the  
331 phenotype-generating rules differ in their relative prevalence and the assumption of  
332 unmeasured confounders is violated. Finally, is a set of data sets with complex co-  
333 varying parameters (COMPLEX), we systematically varied the underlying parameters of  
334 the simulation and examined the relationship between summary statistics of the RF  
335 performance and the ability of BowSaw to generate candidate rules containing the true  
336 phenotype-generating rules.

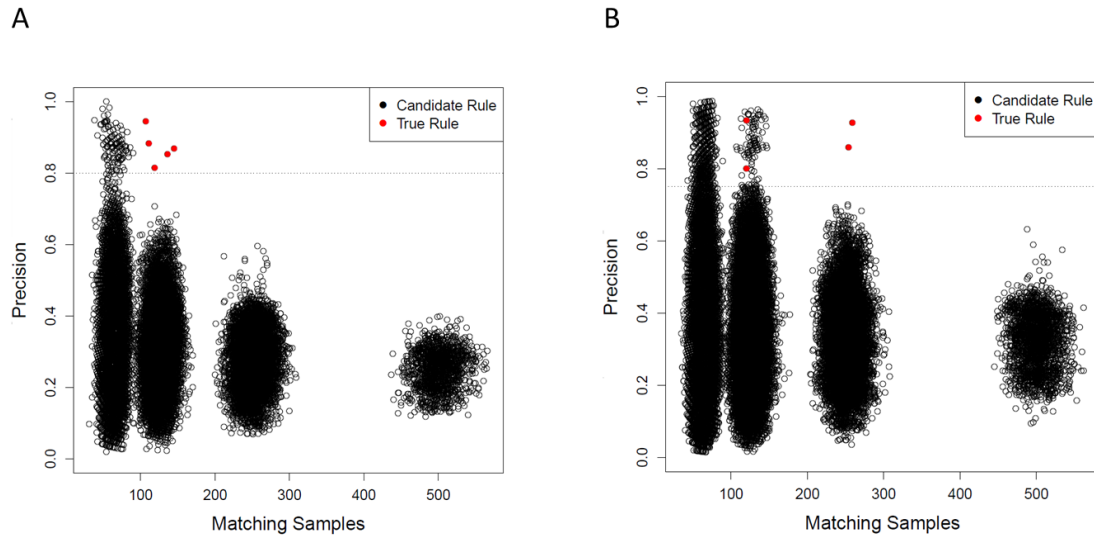
337 For the IDEALIZED scenario, we simulated data set of 100 independent and  
338 identically distributed random binary variables and 2,000 observations. We randomly  
339 defined five rules that each required four randomly selected variables each to have  
340 specific values (e.g. all variables equal to 1) in order to assign a hypothetical phenotype  
341 with likelihood between .8 and .9. Here we present the results of this scenario with a  
342 specified random seed, but other seeds and parameters can be explored using the scripts  
343 provided in the supplemental files. Using these parameters 479 samples were assigned  
344 the phenotype and BowSaw produced a set of 135 unique candidate rules ranging in  
345 complexity from six to fourteen variables. From these rules, we produced all sub-rules  
346 ranging involving anywhere from two to five variables, which resulted in unique 50,034

347 sub-rules. We calculated the number of matches  $|I|$ , the proportion of samples with the  
348 phenotype,  $F$ , for each sub-rule, and visualized these values in order to select an  
349 association threshold (Figure 2A). To reduce the number of sub-rules that the curate  
350 algorithm would need to examine, we eliminated from consideration any rules that had an  
351  $F$  below 80%. We selected an 80% threshold because in the cluster centered around 125  
352 matching samples there is a small cloud of rules that are clearly segregating the  
353 phenotype more efficiently than the others are. We selected the sub-rule with largest  $|I|$   
354 among these as the top candidate rule. This produced a final list consisting of five  
355 candidate rules that accounted for all of the samples with the phenotype and were each  
356 one of the true phenotype generating rules (Figure 3A red points). These results  
357 demonstrate that in an ideal scenario with no phenotype diagnosis errors, BowSaw is  
358 indeed capable of recovering multiple true rules.

359 For the more challenging scenario (INTERMEDIATE), we generated the data set  
360 the same as before except this time we allowed the five underlying rules to vary in  
361 complexity from three to five variables. Varying the complexities of rules resulted in  
362 different prevalence among them, as rules that are more complicated are less likely to  
363 appear in the data. In this case, we had one rule of complexity five, two that required four  
364 variables, and two that used three variables. We also added background noise by  
365 randomly assigning the phenotype to 2% of samples that did not possess any of the rules.  
366 BowSaw produced 176 unique candidate rules involving between six to thirteen  
367 variables. From this list we generated 68,938 sub-rules and chose an association threshold  
368 of 75% because there are two clusters at  $\sim|I| = 125$  that begin to clearly separate in that

369 range and the two outlier points at  $\sim|I| = 250$  do not combine to account for all of the  
370 phenotype (Figure 3B). Applying the curate algorithm to the rules meeting this threshold  
371 produced 20 candidate sub-rules the top four (when ranked by  $|I|$ ) of which were true  
372 rules. The rule of five variables was not recovered. These results show that BowSaw is  
373 able to recover strongly associated patterns (and in this case, causal patterns) even in the  
374 presence of noise, but low prevalence rules can be masked by high prevalence rules.

375         We used the same data generation method to investigate BowSaw's ability to  
376 produce candidate rules containing true rules when the underlying parameters change.  
377 We applied BowSaw to 20,000 simulated data sets where we randomly altered the  
378 number of features, sample size (200 or 2,000 samples), complexity of the rules, number  
379 of rules, the likelihood of each rule assigning the phenotype, and the background noise.  
380 We identified scenarios where rule recovery with BowSaw performs very well and  
381 situations in which it fails to recover any rules at all. Additionally, we found a strong  
382 linear relationship between BowSaw's performance measured as the average fraction of  
383 rules recovered and the of number of samples, number of features, and two evaluation  
384 metrics for RF model – the area under the curve for both the receiver operator  
385 characteristic and precision recall curves (Figure S1).



386

387 **Figure 2**

388 **A** Precision of candidate sub-rules against the number of exactly matching samples for  
389 the ideal scenario. Each point represents a unique sub-rule. X-axis is the number of  
390 samples that exactly match the pattern defined by the rule. Y-axis is the fraction of  
391 matching samples with the observed phenotype (i.e. precision of the rule). Each cluster of  
392 points corresponds to decreasing rule complexity from 5 variables per rule to 2 on the  
393 right most cluster. These clusters appear because the values of each variable is produced  
394 by an identical binomial distribution. Dashed line is the precision threshold we set. Only  
395 candidate rules with precision above this threshold were considered for the curate  
396 algorithm. Red points are the causative sub-rules we defined. BowSaw correctly  
397 identified all five red points in this scenario. **B** Candidate sub-rules generated for the  
398 more challenging scenario. We defined 5 causative rules of varying lengths in this  
399 scenario and allowed 2% of samples without a causative rule to be assigned the label.  
400 BowSaw completely 4 of the causative rules (red points). The longest rule which  
401 involved 5 variables was not recovered.

402

403 **Application to Human Microbiome Data**

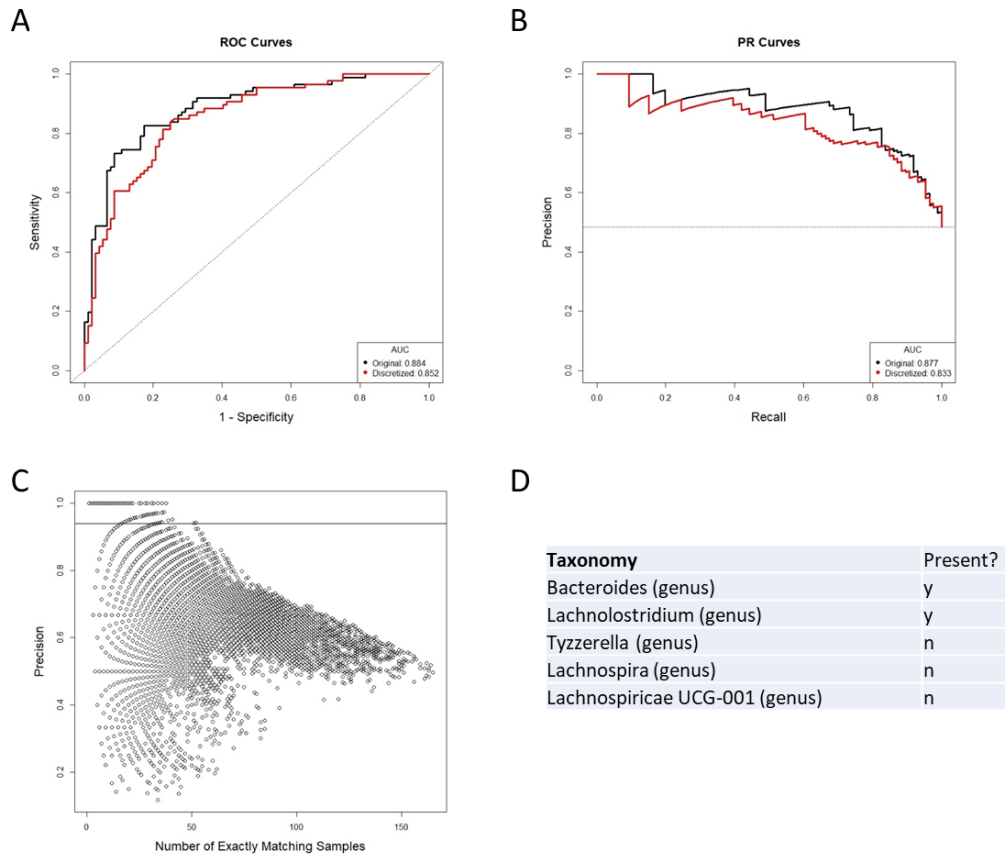
404 Irregular distributions of microbial taxa within the gut are often associated with  
405 serious illnesses such as Crohn's disease or ulcerative colitis [28, 29]. Human  
406 microbiome studies regularly use 16s sequencing methods and extensive reference  
407 databases to report on microbial taxa found in samples as operational taxon units (OTUs).  
408 RF classifiers are frequently built using counts of OTUs to accurately discriminate

409 between disease and healthy patient samples [30, 31]. Despite their demonstrated  
410 effectiveness as good classifiers of Crohn's disease, studies that look to discover  
411 associations with disease status typically focus on individual OTUs while specific  
412 microbial association rules found by RF are not discussed, as a result it is uncertain how  
413 heterogeneous study cohorts are. To investigate potential rule heterogeneity in a human  
414 microbiome cohort we downloaded processed files from the Human Microbiome Project  
415 for inflammatory bowel disease (IBD) [11] which contain information on the taxonomic  
416 profiles of 982 OTUs in 178 patients – 86 of which have been diagnosed with Crohn's  
417 disease, 46 diagnosed with ulcerative colitis, and 46 diagnosed as non-IBD. We were  
418 specifically interested in finding rules that separate the Crohn's disease samples from  
419 ulcerative colitis and non-IBD, so we framed the problem as a binary classification task  
420 with Crohn's disease as the target phenotype.

421         Since the current implementation of BowSaw is limited to finding rules when the  
422 variables have categorical values, we first converted the OTU counts of each taxon to a  
423 simple presence/absence scheme. This resulted in nearly equivalent RF performance  
424 relative to training RF with the original continuous OTU inputs: ROC AUC of 0.862  
425 (binary) vs 0.882 (continuous) and PR AUC of 0.846 (binary) vs 0.886 (continuous)  
426 (Figure 3A-B). This is an important result because it allows us to think about associations  
427 just in terms of presence or absence of an OTU without sacrificing much in model  
428 performance. We applied BowSaw to the Crohn's disease samples and visualized 56,902  
429 resultant sub-rules ranging in complexity from 2 to 7 variables (Figure 3C). There were  
430 1,941 sub-rules with  $F = 1$ . We selected the most general of these rules ( $\max|I|$ ) to be the

431 top candidate for the curate algorithm and found that it considers the status of 5 OTUs  
432 and accounts for 38 of 86 Crohn's disease samples (Figure 3C). We set an association  
433 threshold of 90% and ended up with 10 sub-rules that together account for all 86 Crohn's  
434 disease samples and an additional 11 non-Crohn's disease samples (4 non-IBD, 7  
435 ulcerative colitis). The top five rules combine to account for 78 of 86 Crohn's disease  
436 samples and include 10 non-Crohn's disease samples (Table 1).

437 The top candidate rule is comprised of the presence of *Bacteroides* and  
438 *Lachnoclostridium* and the absence of three genera from the family *Lachnospiraceae*:  
439 *Lachnospira*, *Tyzerrella*, and *Lachnospiraceae* UCG 001 (Figure 3D). Detection of  
440 *Bacteroides* was nearly ubiquitous within the cohort, it was found in 170 of 178 total  
441 samples, but only 3 of the samples in which it was missing are diagnosed as Crohn's  
442 disease. For the remaining taxa we performed a t-test comparing the distribution of the  
443 taxa in Crohn's disease versus ulcerative colitis and versus healthy samples.  
444 *Lachnoclostridium* was frequently found in Crohn's disease (67/86) but not in ulcerative  
445 colitis (27/46,  $p = .02$ ) and was detected at roughly the same rate in non-IBD samples  
446 (34/46,  $p = .616$ ). Detection of *Lachnospira* was depleted in Crohn's disease samples  
447 (20/86) relative to ulcerative colitis (20/46,  $p = .022$ ) and to non-IBD samples (31/46,  $p =$   
448  $9.9^{-7}$ ). *Tyzerrella* was also detected at a lower rate in Crohn's disease (63/86) relative to  
449 ulcerative colitis (24/46,  $p = .019$ ) and non-IBD (24/46,  $p = .019$ ). *Lachnospiraceae* UCG  
450 001 was rarely detected in Crohn's disease (4/86) which is a lower rate than it was  
451 detected in ulcerative colitis (9/46,  $p = .022$ ) and in non-IBD samples (19/46,  $p = 1.45^{-5}$ ).



452

453 **Figure 3**

454 **A** Performance of the random forest classifier as measured by area under the receiver  
455 operator curve (ROC-AUC) is not strongly perturbed by simplifying OTU representation  
456 to a presence/absence scheme versus the original continuous count. Dashed line indicates  
457 the performance of a perfectly random classifier. **B** The area under the curve of the  
458 precision recall curve is similarly not strongly affected by the new representation scheme.  
459 Dashed horizontal line is the random performance line. **C** Each point represents a unique  
460 candidate sub-rule. On the x-axis is the number of samples in the data matrix that are  
461 subject to that rule. The y-axis represents what fraction of matching samples were  
462 diagnosed as Crohn's disease. **D** The taxon identities of the OTUs that make up the most  
463 generally applicable of the sub-rules where all matching samples have the Crohn's  
464 disease label.  
465

Rule	CD Samples	Non CD Samples	New Samples Covered	Taxonomy	Presence
1	38	0	38	<i>Bacteroides (genus)</i>	y
				<i>Lachnolostridium (genus)</i>	y
				<i>Tyzzerella (genus)</i>	n
				<i>Lachnospira (genus)</i>	n
				<i>Lachnospiraceae UCG-001 (genus)</i>	n
2	41	4	20	<i>Dialister (genus)</i>	y
				<i>Christensenellaceae R7 group (genus)</i>	n
				<i>Christensenellaceae R7 group (genus)</i>	n
				<i>Collinsella (genus)</i>	n
				<i>Ruminococcaceae (family)</i>	n
				<i>Finegoldia (genus)</i>	n
				<i>Ruminococcus 1 (genus)</i>	n
3	9	1	9	<i>Ruminococcus 1 (genus)</i>	y
				<i>Ruminococcaceae UCG-002 (genus)</i>	n
				<i>Lachnospiraceae (family)</i>	n
4	24	2	6	<i>Streptococcus (genus)</i>	y
				<i>Tyzzerella (genus)</i>	n
				<i>Lachnospiraceae (family)</i>	n
				<i>Hafnia Obesumbacterium</i>	n
5	27	3	5	<i>Lachnospiraceae UCG-008 (family)</i>	y
				<i>Ruminococcus 1 (genus)</i>	n
				<i>Eubacterium eligens group</i>	n
6	5	0	2	<i>Ruminococcus 1 (genus)</i>	y
				<i>Dorea (genus)</i>	n
7	7	0	2	<i>Bacteroides (genus)</i>	y
				<i>Dialister (genus)</i>	n
				<i>Eubacterium rectale group</i>	n
8	15	0	2	<i>Lachnospiraceae NK4A136 group</i>	y
				<i>Eubacterium eligens group</i>	y
				<i>Tyzzerella (genus)</i>	n
				<i>Christensenellaceae R7 group (genus)</i>	n
				<i>Lachnospira (genus)</i>	n
9	3	0	1	<i>Ruminococcus gnavus group</i>	y
				<i>Veillonella (genus)</i>	n
				<i>Bacteroides (genus)</i>	n
				<i>Finegoldia (genus)</i>	n
10	10	1	1	<i>Parabacteroides (genus)</i>	y
				<i>Eubacterium eligens group</i>	y
				<i>Ruminococcaceae UCG-003 (genus)</i>	n
				<i>Lachnospiraceae ND3007 group</i>	n

466

467

468 **Table 1** Association rules identified by BowSaw that account for all Crohn's disease

469 samples.

470

471

472

473 **Discussion**



474 Interpretation of random forest models for classification may be confounded when  
475 there are multiple rules (combinations of variables and their specific values) associated  
476 with a phenotype of interest. We have developed BowSaw, which is an algorithmic  
477 approach for identifying the rules that a trained random forest model uses to make  
478 classifications when the values are categorical in nature. By taking advantage of the  
479 structure of trees found within a random forest, BowSaw produces a set of multiple  
480 decision rules that combine to account for each sample with a given observed phenotype.  
481 When the variables are the presumed causal agents, these rules represent plausible  
482 mechanistic relationships.

483 Results on simulated data demonstrate that when there are multiple rules  
484 associated with a single phenotype label that BowSaw is capable of faithfully identifying  
485 them. Application to data from the human microbiome project offers further evidence  
486 that BowSaw provides an efficient way of generating plausible hypotheses for high  
487 through put metagenomics studies. In particular we identified a rule that utilizes a  
488 presence/absence pattern of five microbial taxa (present: *bacteroides*, *lachnoclostridium*,  
489 absent: *lachnospira*, *lachnospiracea*, *tyzerrella*) that accounts for nearly half of all  
490 Crohn's disease samples in the cohort (38/86). This specific pattern of microbial  
491 colonization in the guts of Crohn's disease patients is unreported, but each taxon's  
492 respective enrichment or depletion status and association with disease status has been  
493 reported. If the cohort of patients in the human microbiome study are representative of all  
494 people afflicted by Crohn's disease then this rule represents a significantly large sub-set  
495 of those suffering. Inquiries into the relationship of the taxa included in this rule with

496 disease status may yield important insights into the mechanisms of the disease and  
497 potential therapeutic strategies for this sub-population. Of the five associated taxa, we  
498 suspect that the absence of *lachnospira*, *lachnospiracea UCG 001*, and *tyzzerella* are  
499 biologically meaningful. We have reason to believe so because it has been reported that  
500 the *lachnospiraceae* family is generally suppressed in Crohn's disease [32–34].  
501 *Lachnospira* has been reported as depleted with respect to Crohn's disease several times  
502 [35, 36]. The depletion of *tyzzerella* has been associated with chronic intestinal  
503 inflammation and supplementation suggested as a probiotic for Crohn's disease [37, 38].  
504 While the relationship of *lachnospiracea UCG 001* with Crohn's disease is still unclear,  
505 its depletion has been reported in mice displaying symptoms of anhedonia and it was  
506 significantly enriched in anhedonia resilient mice [39]. Partly because IBD is frequently  
507 accompanied by depression, anhedonia has been suggested as an important symptom in  
508 the diagnosis of IBD [40]. The associations of the individual OTUs defined by this rule  
509 are consistent with previously reported findings in the existing literature and describe a  
510 taxonomic profile that exclusively identifies a large sub-population of Crohn's disease  
511 samples within this cohort. The presence of *bacteroides* does not appear to be particularly  
512 useful and in this context is probably preserved because it causes a perfect association,  
513 although high levels of some species are implicated in the pathology of Crohn's disease  
514 [41]. *Lachnoclostridium*, is differentially distributed across the three classes. Notably it is  
515 less frequently detected in ulcerative colitis relative to Crohn's and non-IBD samples,  
516 which roughly resemble one another. Increased levels of this genus was detected in rats

517 that showed relief of colitis symptoms after treatment with a proposed therapeutic agent  
518 [42].

519         The current implementation of the algorithms are restricted to classification tasks  
520 with categorical predictor values, this is a challenge that we will need to address in order  
521 to make the approach more generally applicable. Future work will also focus on  
522 extending these for the interpretation of regression models. Such additions will greatly  
523 increase the number of systems to which we can apply BowSaw.

524

## 525 **Acknowledgments**

526 We are grateful to members of the Segrè lab for helpful discussions and for feedback on  
527 the manuscript. DS and DD acknowledge funding from the Defense Advanced Research  
528 Projects Agency (Purchase Request No. HR0011515303, Contract No. HR0011-15-C-  
529 0091), the U.S. Department of Energy (DE-SC0012627), the NIH (T32GM100842,  
530 5R01DE024468, R01GM121950 and Sub\_P30DK036836\_P&F), the National Science  
531 Foundation (1457695), the Human Frontiers Science Program (RGP0020/2016), and the  
532 Boston University Interdisciplinary Biomedical Research Office. DD is grateful to Dr.  
533 Nisha Rajagopal for her patience in conversations about random forests and her valuable  
534 insight.



## References

1. Visscher, P. M., Wray, N. R., Zhang, Q., Sklar, P., McCarthy, M. I., Brown, M. A., & Yang, J. (2017). 10 Years of GWAS Discovery: Biology, Function, and Translation. *American Journal of Human Genetics*. doi:10.1016/j.ajhg.2017.06.005
2. Furqan, M. S., & Siyal, M. Y. (2016). Inference of biological networks using Bi-directional Random Forest Granger causality. *SpringerPlus*. doi:10.1186/s40064-016-2156-y
3. Le, V., Quinn, T. P., Tran, T., & Venkatesh, S. (2019). Deep in the Bowel: Highly Interpretable Neural Encoder-Decoder Networks Predict Gut Metabolites from Gut Microbiome. *bioRxiv*. doi:10.1101/686394
4. Azmi, M., Runger, G. C., & Berrado, A. (2019). Interpretable regularized class association rules algorithm for classification in a categorical data space. *Information Sciences*. doi:10.1016/j.ins.2019.01.047
5. Nguyen, M., Wesley Long, S., McDermott, P. F., Olsen, R. J., Olson, R., Stevens, R. L., ... Davisa, J. J. (2019). Using machine learning to predict antimicrobial MICs and associated genomic features for nontyphoidal Salmonella. *Journal of Clinical Microbiology*. doi:10.1128/JCM.01260-18
6. LaPierre, N., Ju, C. J. T., Zhou, G., & Wang, W. (2019). MetaPheno: A critical evaluation of deep learning and machine learning in metagenome-based disease prediction. *Methods*. doi:10.1016/j.ymeth.2019.03.003
7. Brodley, C. E., & Friedl, M. A. (1997). Decision tree classification of land cover

- from remotely sensed data. *Remote Sensing of Environment*. doi:10.1016/S0034-4257(97)00049-7
8. Knudson, A. G. (1971). Mutation and cancer: statistical study of retinoblastoma. *Proceedings of the National Academy of Sciences of the United States of America*. doi:10.1073/pnas.68.4.820
  9. Emily, M., Mailund, T., Hein, J., Schausser, L., & Schierup, M. H. (2009). Using biological networks to search for interacting loci in genome-wide association studies. *European Journal of Human Genetics*. doi:10.1038/ejhg.2009.15
  10. Leem, S., Jeong, H. H., Lee, J., Wee, K., & Sohn, K. A. (2014). Fast detection of high-order epistatic interactions in genome-wide association studies using information theoretic measure. *Computational Biology and Chemistry*. doi:10.1016/j.compbiolchem.2014.01.005
  11. Proctor, L. M., Creasy, H. H., Fettweis, J. M., Lloyd-Price, J., Mahurkar, A., Zhou, W., ... Huttenhower, C. (2019). The Integrative Human Microbiome Project. *Nature*. doi:10.1038/s41586-019-1238-8
  12. Reading, D. (2014). Crohn Disease: Pathophysiology, Diagnosis, and Treatment, 85(3), 297–320.
  13. Louppe, G. (2014). *Understanding Random Forests*. Cornell University Library.
  14. Wright, M. N., Ziegler, A., & König, I. R. (2016). Do little interactions get lost in dark random forests? *BMC bioinformatics*. doi:10.1186/s12859-016-0995-8
  15. Boulesteix, A. L., Janitza, S., Kruppa, J., & König, I. R. (2012). Overview of random forest methodology and practical guidance with emphasis on

- computational biology and bioinformatics. *Wiley Interdisciplinary Reviews: Data Mining and Knowledge Discovery*. doi:10.1002/widm.1072
16. Touw, W. G., Bayjanov, J. R., Overmars, L., Backus, L., Boekhorst, J., Wels, M., & Sacha van Hijum, A. F. T. (2013). Data mining in the life science swith random forest: A walk in the park or lost in the jungle? *Briefings in Bioinformatics*. doi:10.1093/bib/bbs034
  17. Nguyen, C., Wang, Y., & Nguyen, H. N. (2013). Random forest classifier combined with feature selection for breast cancer diagnosis and prognostic. *Journal of Biomedical Science and Engineering*. doi:10.4236/jbise.2013.65070
  18. Franzosa, E. A., Sirota-Madi, A., Avila-Pacheco, J., Fornelos, N., Haiser, H. J., Reinker, S., ... Xavier, R. J. (2019). Gut microbiome structure and metabolic activity in inflammatory bowel disease. *Nature Microbiology*. doi:10.1038/s41564-018-0306-4
  19. Duvallet, C., Gibbons, S. M., Gurry, T., Irizarry, R. A., & Alm, E. J. (2017). Meta-analysis of gut microbiome studies identifies disease-specific and shared responses. *Nature Communications*. doi:10.1038/s41467-017-01973-8
  20. Breiman, L. (2001). Random forests. *Machine Learning*, 45(1), 5–32. doi:10.1023/A:1010933404324
  21. Castelvechi, D. (2016). Can we open the black box of AI? *Nature*. doi:10.1038/538020a
  22. Welling, S. H., Refsgaard, H. H. F., Brockhoff, P. B., & Clemmensen, L. H. (2016). Forest Floor Visualizations of Random Forests. *arXiv*. Retrieved from

<http://arxiv.org/abs/1605.09196>

23. Palczewska, A., Palczewski, J., Robinson, R. M., & Neagu, D. (2013). Interpreting random forest classification models using a feature contribution method (extended). *2013 IEEE 14th International Conference on Information Reuse & Integration (IRI)*, 1–30. doi:10.1109/IRI.2013.6642461
24. Deng, H. (2019). Interpreting tree ensembles with inTrees. *International Journal of Data Science and Analytics*. doi:10.1007/s41060-018-0144-8
25. Strobl, C., Boulesteix, A. L., Kneib, T., Augustin, T., & Zeileis, A. (2008). Conditional variable importance for random forests. *BMC Bioinformatics*. doi:10.1186/1471-2105-9-307
26. Basu, S., Kumbier, K., Brown, J. B., & Yu, B. (2018). Iterative random forests to discover predictive and stable high-order interactions. *Proceedings of the National Academy of Sciences*. doi:10.1073/pnas.1711236115
27. Dessau, R. B., & Pipper, C. B. (2008). [”R”--project for statistical computing]. *Ugeskrift for laeger*.
28. Carding, S., Verbeke, K., Vipond, D. T., Corfe, B. M., & Owen, L. J. (2015). Dysbiosis of the gut microbiota in disease. *Microbial Ecology in Health & Disease*. doi:10.3402/mehd.v26.26191
29. Levy, M., Kolodziejczyk, A. A., Thaiss, C. A., & Elinav, E. (2017). Dysbiosis and the immune system. *Nature Reviews Immunology*. doi:10.1038/nri.2017.7
30. Ai, D., Pan, H., Han, R., Li, X., Liu, G., & Xia, L. C. (2019). Using Decision Tree Aggregation with Random Forest Model to Identify Gut Microbes Associated with



- Colorectal Cancer. *Genes*, 10(2), 112. doi:10.3390/genes10020112
31. Vangay, P., Hillmann, B. M., & Knights, D. (2019). Microbiome learning Repo (ML Repo): A public repository of microbiome regression and classification tasks. *GigaScience*. doi:10.1093/gigascience/giz042
32. Loh, G., & Blaut, M. (2012). Role of commensal gut bacteria in inflammatory bowel diseases. *Gut Microbes*. doi:10.4161/gmic.22156
33. Nagao-Kitamoto, H., & Kamada, N. (2017). Host-microbial Cross-talk in Inflammatory Bowel Disease. *Immune Network*. doi:10.4110/in.2017.17.1.1
34. Geirnaert, A., Calatayud, M., Grootaert, C., Laukens, D., Devriese, S., Smagghe, G., ... Van De Wiele, T. (2017). Butyrate-producing bacteria supplemented in vitro to Crohn's disease patient microbiota increased butyrate production and enhanced intestinal epithelial barrier integrity. *Scientific Reports*. doi:10.1038/s41598-017-11734-8
35. Wang, Y., Gao, X., Ghozlane, A., Hu, H., Li, X., Xiao, Y., ... Zhang, T. (2018). Characteristics of faecal microbiota in paediatric Crohn's disease and their dynamic changes during infliximab therapy. *Journal of Crohn's and Colitis*. doi:10.1093/ecco-jcc/jjx153
36. Wright, E. K., Kamm, M. A., Wagner, J., Teo, S. M., Cruz, P. De, Hamilton, A. L., ... Kirkwood, C. D. (2017). Microbial Factors Associated with Postoperative Crohn's Disease Recurrence. *Journal of Crohn's & colitis*. doi:10.1093/ecco-jcc/jjw136
37. Y.-J., C., H., W., S.-D., W., N., L., Y.-T., W., H.-N., L., ... Shen, X.-Z. (2018).

Parasutterella, in association with irritable bowel syndrome and intestinal chronic inflammation. *Journal of Gastroenterology and Hepatology (Australia)*.

doi:10.1111/jgh.14281

38. Berry, D., Rahman, S., Kaplan, J., & Gordon, N. (2018). Probiotic and prebiotic compositions, and methods of use thereof for treatment and prevention of graft versus host disease. *USPTO*.
39. Yang, C., Fang, X., Zhan, G., Huang, N., Li, S., Bi, J., ... Hashimoto, K. (2019). Key role of gut microbiota in anhedonia-like phenotype in rodents with neuropathic pain. *Translational Psychiatry*. doi:10.1038/s41398-019-0379-8
40. Carpinelli, L., Bucci, C., Santonicola, A., Zingone, F., Ciacci, C., & Iovino, P. (2019). Anhedonia in irritable bowel syndrome and in inflammatory bowel diseases and its relationship with abdominal pain. *Neurogastroenterology and Motility*. doi:10.1111/nmo.13531
41. Rabizadeh, S., Rhee, K. J., Wu, S., Huso, D., Gan, C. M., Golub, J. E., ... Sears, C. L. (2007). Enterotoxigenic *Bacteroides fragilis*: A potential instigator of colitis. *Inflammatory Bowel Diseases*. doi:10.1002/ibd.20265
42. Wang, K., Yang, Q., Ma, Q., Wang, B., Wan, Z., Chen, M., & Wu, L. (2018). Protective effects of salvianolic acid a against dextran sodium sulfate-induced acute colitis in rats. *Nutrients*. doi:10.3390/nu10060791

