

1 **Integrating genome-wide association mapping of additive and**
2 **dominance genetic effects to improve genomic prediction**
3 **accuracy in *Eucalyptus***

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18 Running head: GWAS and GS in Eucalyptus

19 **Summary**

20 Genome-wide association studies (GWAS) is a powerful and widely used
21 approach to decipher the genetic control of complex traits. A major challenge for
22 dissecting quantitative traits in forest trees is statistical power. In this study, we use
23 a population consisting of 1123 samples from two successive generations that have
24 been phenotyped for growth and wood property traits and genotyped using the
25 EuChip60K chip, yielding 37,832 informative SNPs. We use multi-locus GWAS
26 models to assess both additive and dominance effects to identify markers
27 associated with growth and wood property traits in the eucalypt hybrids. Additive
28 and dominance association models identified 78 and 82 significant SNPs across all
29 traits, respectively, which captured between 39 and 86% of the genomic-based
30 heritability. We also used SNPs identified from the GWAS and SNPs using less
31 stringent significance thresholds to evaluate predictive abilities in a genomic
32 selection framework. Genomic selection models based on the top 1% SNPs
33 captured a substantially greater proportion of the genetic variance of traits
34 compared to when all SNPs were used for model training. The prediction ability of
35 estimated breeding values was significantly improved for all traits using either the
36 top 1% SNPs or SNPs identified using a relaxed p -value threshold ($p < 10^{-3}$). This
37 study highlights the added value of also considering dominance effects for
38 identifying genomic regions controlling growth traits in trees. Moreover,
39 integrating GWAS results into genomic selection method provides enhanced power

40 relative to discrete associations for identifying genomic variation potentially useful
41 in tree breeding.

42 **Keywords**

43 *Eucalyptus*, dominance, FarmCPU, genome-wide association study, genomic
44 selection

45

46 **Introduction**

47 Deciphering the genetic basis of complex phenotypic traits is of fundamental
48 importance for understanding biological processes and may ultimately provide
49 information that can help enhance selection in plant breeding programs. Genome-
50 wide association studies (GWAS) is a powerful way to identify putative causal
51 genes or genomic segments that underlie phenotypic variation in plants,
52 particularly for traits with complex genetic architectures (Ingvarsson and Street,
53 2011; Kruglyak, 2008). Dissection of complex traits have been undertaken in forest
54 genetics to understand the genetic basis of adaptive phenotypes (Ingvarsson et al.,
55 2008; Olson et al., 2013; Wang et al., 2018) or physiological or morphological
56 traits, such as growth or wood properties. For example, Porth *et. al.* (Porth et al.,
57 2013) and later Chhetri *et. al.* (Chhetri et al., 2019a) performed GWAS for wood
58 traits, biomass, eco-physiological and phenology traits in *Populus trichocarpa* with
59 genotyping based on 6.78 million single nucleotide polymorphisms (SNPs).
60 Similarly, a study of *Salix viminalis* identified 29 SNPs that were associated with
61 bud burst, leaf senescence, number of shoots or shoot diameter (Hallingback et al.,
62 2016). In *Eucalyptus*, the earliest GWAS identified 16 markers that were associated
63 with growth and two markers that were associated with lignin traits (Cappa et al.,
64 2013). Recently, 26 quantitative trait loci (QTLs) were identified for productivity
65 and disease resistance using a regional heritability mapping method that helps
66 increase the genomic heritability to 5-15% from 4-6% when using SNPs
67 individually (Resende et al., 2017a; Resende et al., 2017b).

68 GWAS studies can also provide tools for accelerating the long breeding cycles
69 in tree breeding (reviewed in (Neale and Kremer, 2011)). For example, although
70 many species of *Eucalyptus* display unusually fast growth, breeding cycles aimed
71 at developing elite commercial genotypes still take between 12 to 16 years to
72 complete, since identification of elite genotypes require progeny trials followed by
73 two or more sequential clonal trials (Rezende et al., 2014). However, genomic
74 selection based on genome-wide molecular makers is expected to reduce the time
75 required for completing a cycle of developing elite clones to only 9 years mainly
76 due to the shorter time needed for progeny tests when phenotypes can be predicted
77 from the genomic selection models (Grattapaglia, 2017).

78 The rapid development in genomics has opened up opportunities to identify
79 molecular markers that are associated with traits of interest and use these marker-
80 trait associations to complement and extend traditional breeding programs. Despite
81 the efforts to discover polymorphisms associated with economically relevant traits,
82 much of the genetic contribution to complex traits in forest trees remains
83 unexplained. One of the main reasons is that GWAS methods normally conduct
84 tests on one marker at a time, for instance using a generalized linear model (GLM)
85 or a mixed linear model (MLM). When dealing with complex traits such as growth
86 and wood qualities, where the effect size of individual loci is likely small to
87 moderate, these methods suffer from limited statistical power to detect loci of small
88 effects (Muller et al., 2017). One potential approach to increase the power and to
89 accurately identify more causal variants is so called ‘multi-locus mixed models’

90 (MLMM), which simultaneously test multiple markers by including them as
91 covariates in a stepwise MLM to partially remove confounding between tested
92 markers and kinship (Segura et al., 2012). One such method is the ‘fixed and
93 random model circulating probability unification’ (FarmCPU) that performs
94 marker tests using other associated markers as covariates in a fixed effect model
95 (Liu et al., 2016). Optimization across the associated covariate markers using a
96 random effect model is then performed separately. This approach has been reported
97 to simultaneously reduce computational complexity, remove confounding between
98 population structure, kinship and quantitative trait loci, prevent model over-fitting
99 and control the number of false positives (Liu et al., 2016).

00 Most GWAS analyses to date have been undertaken by implicitly assuming a
01 genetic architecture consisting of additive effects. However, non-additive effects,
02 including dominance (Bruce, 1910), over-dominance (Crow, 1948) and epistasis
03 (Hill, 1982) are known to also play important roles in controlling some traits. One
04 trait where non-additive effects are likely to be pronounced is heterosis, or hybrid
05 vigor, which is the near universally observed phenomenon of phenotypic
06 superiority of hybrid progeny relative to their parents (Charlesworth and Willis,
07 2009). Not surprisingly, heterosis has been and continues to be of great importance
08 in most plant breeding schemes (Duvick, 2001). To date, a limited number of
09 studies have utilized GWAS methods to dissect the genetic basis of heterotic traits
10 in *Arabidopsis thaliana* and rice. In the model plant *A.thaliana*, dominance and
11 over-dominance of flowering time is a well-studied trait and significant loci from a

12 GWAS were shown to explain as much as 20% of the phenotypic variation in a
13 hybrid population consisting of 435 individuals derived from inter-crossing 30
14 parents (Seymour et al., 2016). In rice, genome-wide dissection uncovered multiple
15 non-additive effect loci for yield increase (Li et al., 2016; Zhen et al., 2017). For
16 instance, a major QTL, rice heterosis 8 (*RH8*) was found to regulate grain-yield
17 component traits (Li et al., 2016). In *Eucalyptus* hybrids dominance appears to be
18 an important and widespread contributor to many growth-related traits (Bison et al.,
19 2006; Bouvet and Vigneron, 1995; Volker et al., 2008) and ratios of dominance to
20 additive variances exceeding 1.2 have been estimated for growth in *E. grandis* x *E.*
21 *urophylla* hybrids (Bouvet et al., 2009; Makouanzi et al., 2014; Tan et al., 2017).
22 Such results suggest that there should be ample opportunities to identify SNPs
23 accounting for dominance and/or over-dominance effects in *Eucalyptus* hybrids.

24 Another genomic-based approach that has become widely used in plant and
25 animal breeding in recent years is genomic selection (GS) or alternatively known
26 as genomic prediction. Unlike GWAS, GS refers to marker-based selection where
27 total genetic variance is captured using genome-wide markers without a prior step
28 of identifying trait-associated markers. GS aims to predict the genetic potential (e.g.
29 genome-estimated breeding values) of breeding individuals without locating genes
30 or QTLs important for the trait(s) of interest. One of most important questions for
31 GS is how to improve the prediction accuracy and methods for accuracy has long
32 been a central research aim in genomic selection. Thus far progress on increasing
33 prediction accuracies have been achieved through the development of new

34 statistical models, more efficient design of training populations, improved quality
35 of phenotypic measurements, a greater number of makers used for model building
36 and by also considering non-additive effects (Grattapaglia, 2017). In this paper we
37 assess methods for improving genomic prediction accuracy by integrating results
38 from GWAS studies into GS to predict the genetic potential of breeding targets. It
39 is well known that using only associated SNPs identified from a GWAS is usually
40 not sufficient for explaining a large fraction of the genetic variation in a trait of
41 interest (the so called “missing heritability” problem, (Makowsky et al., 2011)).
42 However, utilizing GWAS information in the form of associated SNPs, in
43 combination with other types of data has the potential to enhance prediction ability
44 in GS studies (Gowda et al., 2015).

45 In this study, we present results from a GWAS on growth and wood quality
46 traits, in a breeding population comprising two species of *Eucalyptus* and their
47 hybrids. We also integrate the GWAS results in a GS model with the goal of
48 assessing whether this can help increase prediction accuracies for the traits in
49 question. Specifically, our study has two objectives: first, we implement a state of
50 the art GWAS method that consider both additive and dominance effects for
51 dissecting the genetic architecture of growth and wood quality traits. We also
52 evaluate the proportion of phenotypic variation that explained by significant loci
53 for these two genetic effects. Second, we evaluate how different categories of
54 informative SNPs, selected based on the results from the GWAS, can be
55 implemented in a widely used model for genomic prediction, GBLUP, to estimate

56 variance components and to evaluate prediction accuracies of estimated breeding
57 values.

58 **Results**

59 *Characters of growth and wood traits*

60 All growth traits were moderately variable at the different assessment ages
61 (Table 1). We observed a lower phenotypic variation for height at 3 years of age, as
62 judged by the coefficient of variation (Table 1). The F1 population underwent
63 selection based on height in order to identify trees to use for genotyping and this
64 selection process likely contributed to the lower phenotypic variation we see in
65 height at 3 years of age. We also observed low phenotypic variation for basic
66 density and pulp yield, which is commonly observed in many wood quality traits.
67 Generally, variation in CBH was greater than in height but both mean and variance
68 for both traits increased as the trees aged. Growth traits generally had low
69 heritabilities ($h^2 < 0.2$) whereas wood quality traits showed moderate heritabilities
70 (Table 1). Phenotypic correlations between growth traits were generally positive
71 (0.24~0.74) whereas basic density was weakly negatively correlated with pulp
72 yield (-0.28). The wood quality traits were generally independent from growth
73 traits (correlations in the range -0.1 - 0.1) (Figure S1). The greatest positive
74 phenotypic correlations were observed between CBH and height assessed at the
75 same age (0.63 and 0.74 for 3 and 6 years, respectively).

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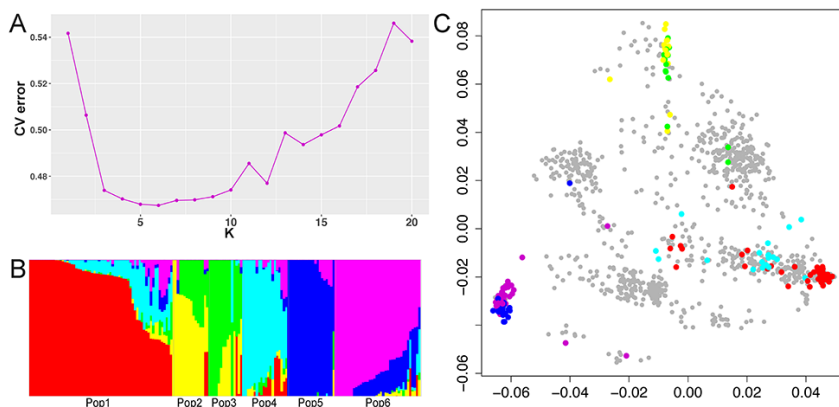
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Table 1. Statistical summary of phenotypes

Trait	Abbr.	No. obs.	Unit	Mean	CV(%) [†]	h ²
Circumference at breast height, age 3 years	CBH3	1123	cm	61.82	13.22	0.143
Height, age 3 years	Ht3	1094	m	22.43	9.81	0.162
Circumference at breast height, age 6 years	CBH6	1104	cm	83.80	18.67	0.186
Height, age 6 years	Ht6	985	m	28.40	13.09	0.182
Basic density	BD	1061	kg/m ³	532.78	6.83	0.381
Pulp yield	PY	1039	%	49.64	8.05	0.42

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[†] CV: coefficient of variance.



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81 **Figure 1.** Population structure of parents and F1 progenies. (A) Cross-validation
82 error in the admixture analysis for K varying from 1-20 for the 174 parents. (B)
83 Population structure of parents inferred using admixture for K=6. (C) PCA plot
84 based on genetic covariance among all individuals. Only the first two principle
85 components are shown. The colours used for the parents are in line with the
86 clustering shown in (B), with grey colour denoting all F1 progeny.

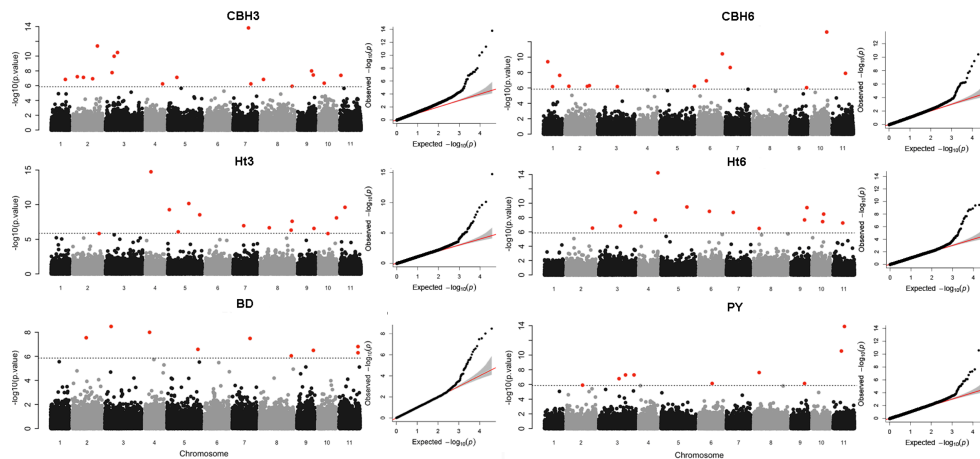
87 *Population structure and model optimization*

88 To examine population structure in the breeding population including both
89 parents and their F1 progenies, we conducted both model-based admixture and
90 fastStructure analyses and principle component analysis (PCA) based on a set of
91 independent SNPs. The admixture analysis could not identify an optimal genetic
92 clustering on account of the minimization of the CV error even for K-values up to

!93 K=100 (Figure S2). In contrast, fastStructure suggested an optimal genetic
!94 clustering of K=1. Due to the inconsistencies between the methods, we repeated the
!95 population structure analyses using only the parents, given that the F1 individuals
!96 were all obtained through crossings between these parents. Admixture analyses
!97 based on the parents alone suggested K=6 minimized the CV error (Fig

!98

!99 ure 1A) and K=6 was also the optimal genetic clustering obtained from
!00 fastStructure. The parents were assigned to the six subpopulations according to
!01 individual ancestry proportions (Figure 1B). We also performed a PCA to
!02 summarize genetic variation among parents and the first six components explained
!03 21.53% of the total genetic variation. Notably, the eigenvalues beyond the first six
!04 PCs were relatively small (Figure S3), consistent with the minimum K identified in
!05 the admixture analyses. Based on first two principle components, parents can be
!06 clearly separated into three clusters and two further sub-clusters can be identified
!07 within in each major cluster. Progenies are inferred to be derived from crossing
!08 parents either with the different major clusters or between them (Figure 1C) and
!09 therefore we used the first six PCs in all subsequent analyses to correct for
!10 population stratification.



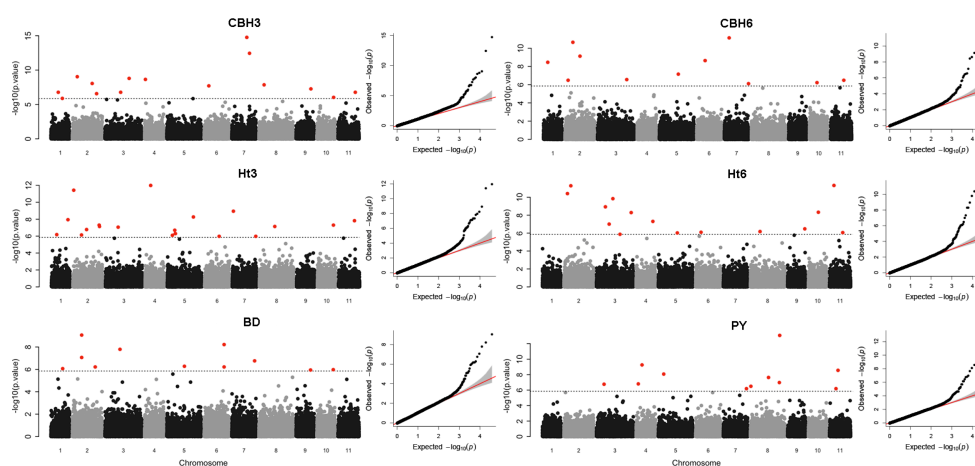
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:12 **Figure 2.** Manhattan plots and quantile-quantile (QQ) plots of the FarmCPU
:13 results using an additive effects model. The traits used are CBH and height at age
:14 3 and 6 (Ht3 and Ht6, respectively) as well as basic density (BD) and pulp yield
:15 (PY). The Manhattan plots show $-\log_{10} p$ -values plotted against SNP positions on
:16 the 11 *Eucalyptus* chromosomes. Associations reaching genome-wide
:17 significance are displayed in red and the horizontal dotted line indicates a
:18 Bonferroni-corrected significant threshold of $p < 1.7E-06$. The QQ plots for each
:19 of the six traits demonstrate the observed versus expected distribution of p -
:20 values. The solid red line represents the expected null distribution assuming no
:21 associations.

:22 ***Genome-wide association study for additive effects***

:23 We first ran FarmCPU with an additive effect encoding to identify loci with
:24 significant additive effects on the different phenotypes. Quantile-quantile (QQ)
:25 plots suggest that population structure and kinship relationships were well
:26 controlled in the GWAS for the different traits (Figure 2). SNPs with p -values $<$
:27 $1.7E-06$ threshold were declared statistically significant. Overall, we identified 78
:28 significant SNPs across the six traits and these significant SNPs were distributed

!29 across all 11 chromosomes (Figure 2). No significant SNPs were identified for
!30 more than one trait, even though both CBH and height show strong genetic
!31 correlations across ages. Comparing the number of significant SNPs found for the
!32 different traits, growth traits had more significant SNPs than wood traits, with
!33 height and CBH at the two different ages having between 14 and 18 significant
!34 SNPs whereas we only identified 9 significant SNPs for the two wood quality traits.
!35 We generally observe lower phenotypic variances explained by individual SNPs
!36 for CBH and height compared to pulp yield and basic density (Table S1). The
!37 maximum percentage of phenotypic variance explained by single associated SNP
!38 was 2.3% (for pulp yield) and the minimum percentage of phenotypic variance
!39 explained by a significantly associated SNP was 0.33% (CBH age 3 years).



!40

!41 **Figure 3.** Manhattan plots and quantile-quantile (QQ) plots of the FarmCPU
!42 results for the dominance effects model. The traits used are CBH and height at
!43 age 3 and 6 (Ht3 and Ht6, respectively) as well as basic density (BD) and pulp
!44 yield (PY). The Manhattan plots show $-\log_{10} p$ -values plotted against SNP

!45 positions on the 11 *Eucalyptus* chromosomes. Associations reaching genome-
!46 wide significance are displayed in red and the horizontal dotted line indicates a
!47 Bonferroni-corrected significant threshold of $p < 1.7E-06$. The QQ plots for each
!48 of the six traits indicate the observed versus expected distribution of p-values.
!49 The solid red line represents the expected null distribution assuming no
!50 associations.

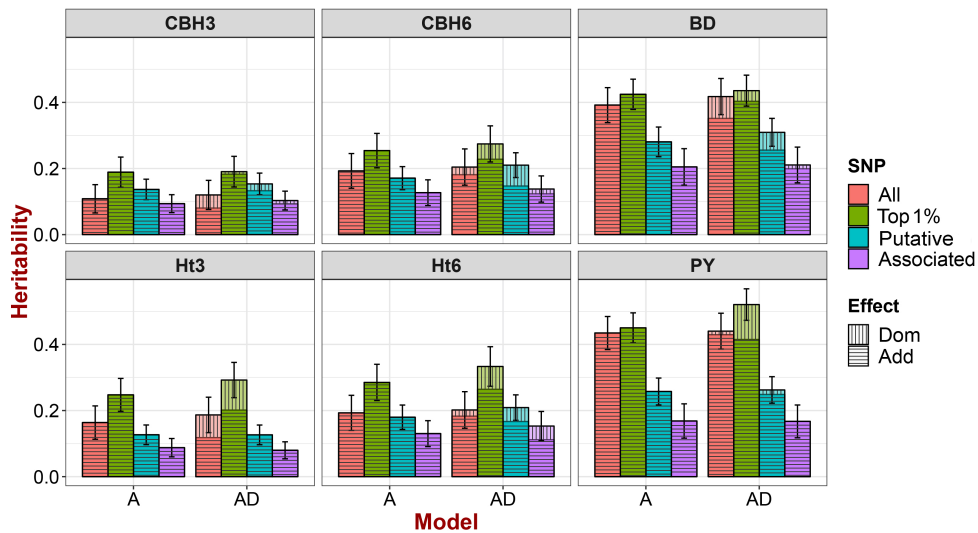
!51 ***Genome-wide association study for dominance effects***

!52 FarmCPU efficiently controlled the false positive rates due to population
!53 structure and sample relationships also when identifying significant loci using
!54 dominance encoding (Figure 3, QQ plots). Under a dominance model we detected a
!55 total of 82 significant SNPs for the six traits. Height at 3 years old (Ht3) had the
!56 greatest number of associations with 19 SNPs displaying significant effects. Fewer
!57 associations were observed for the other traits, with between 11 and 15 significant
!58 SNPs identified (Figure 3, Manhattan plots). Two significant SNPs,
!59 Chr5.40663824 and Chr11.28479550, were found to overlap between CBH and
!60 height at age of 6 years. The maximum percentage of phenotypic variance
!61 explained by an associated SNP was 4%, a much higher value than found in the
!62 additive effect estimations. The smallest percentage of phenotypic variance
!63 explained by an associated SNP for the dominance model was of similar magnitude
!64 to that observed for additive effects model (Table S2). Comparing significant SNPs
!65 identified from the additive and dominance effects models, a total of 10 SNPs
!66 overlap between two models for different traits. This result suggest that the two
!67 genetic effects are not completely independent. Nine out of ten SNPs that overlap

:68 between additive and dominance effects were identified for growth traits and with
:69 the remaining SNP observed for pulp yield.

:70 ***Genome selection by using GWAS results***

:71 To confirm the utility of the SNPs identified from the GWAS and to further
:72 understand the performance of selecting SNPs for each trait based on the GWAS
:73 results, we conducted genomic prediction using four categories of SNPs by using
:74 both an additive genetic model (A) and an additive + dominance genetic model
:75 (AD). The four categories of SNPs used for the GBLUP analyses were selected
:76 from the GWAS results for each trait based on the following criteria: 1) '*associated*
:77 *SNPs*' were identified as significant from the GWAS using the threshold $p < 1.7E-6$;
:78 2) '*putative SNPs*' were identified as significant from the GWAS using a more
:79 relaxed significance threshold $p < 1E-3$ of each trait; 3) the '*top 1% SNPs*' included
:80 the top 1% SNPs for each trait, ranked according to GWAS significance. The
:81 rationale of this category was to ensure that models utilised the same number of
:82 SNPs across all traits. Finally, 4) '*all SNPs*' used all 37,832 available SNPs when
:83 building the genomic selection models (Table S3).



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Figure 4. Genomic-based narrow- and broad-sense heritabilities based on an additive (A) or an additive + dominance (AD) model for the four different categories of SNPs used. The coloured bins represent the different categories of SNPs used, with red indicating ‘all’ SNPs (37,832), green indicates the ‘top 1%’ SNPs ranked according to GWAS p-value, cyan denotes ‘putative’ SNPs selected based on GWAS results with $p < 1E-3$ and purple denoted ‘associated’ SNPs selected based on GWAS results using $p < 1.7E-6$. The fill patterns represent different genetic effects. Vertical lines denote additive effects and horizontal lines denote dominance effects. Error bars indicate the standard error of total genetic variance.

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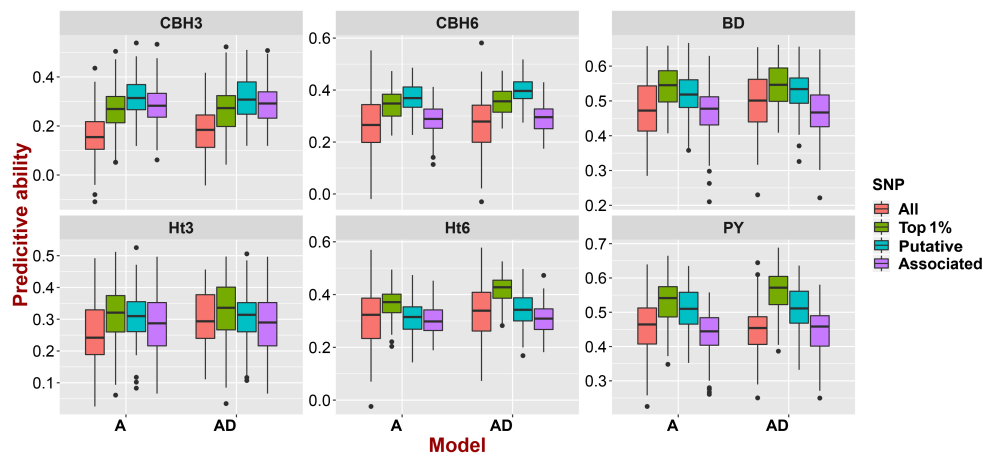
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The narrow-sense and broad-sense heritabilities were estimated using a modified GBLUP model based on different maker-based relationship matrices calculated using the four SNP categories. As expected, basic density and pulp yield had higher realised heritabilities than growth traits, independent of what category of SNPs that were used for the calculations. Broad-sense heritabilities were higher than narrow-sense heritabilities for most traits, demonstrating that dominance plays an important role in the expression of most traits (Figure 4) and in line with earlier

002 observations in this population (Tan et al., 2018). Comparing heritabilities (h^2 and
003 H^2) for the different SNP categories suggest, perhaps surprisingly, that the ‘top 1%
004 SNPs’ category explain more of the genetic variation than any of the other
005 categories, including when all SNPs were used (Table S4). Furthermore, using the
006 ‘top 1% SNP’ set yielded the largest estimates of dominance effects. As expected,
007 using only SNPs that were significantly associated with a trait in the GWAS
008 resulted in lower heritability estimates compared to using all SNPs. Comparing
009 heritability estimates between the ‘putative’ and ‘all’ SNP categories showed that
010 these yielded similar estimates for CBH and height, the ‘putative’ category of SNPs
011 yielding significantly lower heritability estimates than the ‘all’ SNP category for the
012 two wood quality traits (Figure 4).

013 We further estimated the prediction ability of breeding values for the *A* model
014 and the prediction ability of genetic values for the *AD* model using a ten-fold cross-
015 validation approach. The distribution of predictive abilities for each of the models
016 and SNP, obtained using 100 replications, are displayed in Figure (5). Generally,
017 we observe higher prediction abilities for wood quality traits, in line with the
018 higher heritability values we observe for these traits. The *AD* model yielded
019 slightly higher prediction abilities than the *A* model for most of traits. When
020 comparing the different SNP categories, both the ‘top 1%’ and ‘putative’ SNP
021 categories yielded substantial improvements in the predictive ability compared to
022 both the ‘all’ and the ‘significant’ SNP categories. The ‘associated’ SNPs yielded
023 more or less similar results to that obtained using ‘all’ SNPs. Moreover, the ‘top

124 1% SNP category yielded the highest average prediction ability for height, basic
125 density and pulp yield with values being much higher than those calculated from
126 the ‘all’ SNP category (Table S5).



127

128 **Figure 5.** Predictive abilities of the additive (A) and the additive + dominance
129 (AD) genomic prediction models for the four different categories of SNPs. The
130 coloured boxplots display the distribution of predictive ability across 100
131 replicates of ten-fold cross-validation for the different categories of SNPs.
132 Colours used as in Figure 4.

133

134 Discussion

135 We used GWAS with both additive and dominance effects to dissect the
136 genetic architecture of growth and wood quality traits in hybrid *Eucalyptus*. The
137 method we employed for GWAS, FarmCPU, was able to control the false positive
138 rate induced by both the complex population structure and kinship that characterise
139 our mapping population and efficiently identified significantly associated SNPs for
140 both additive and dominance effects. Using top-ranking SNPs based on the GWAS

141 yielded higher genomic heritability estimates compared to using all available SNPs.
142 We were also able to achieve more accurate genomic prediction results by filtering
143 SNPs based on their associations in the GWAS and this help shed light on future
144 directions in the application of genomic selection in *Eucalyptus* breeding.

145 ***FarmCPU perform superior in GWAS analysis***

146 Most economic traits targeted for breeding in forestry, such as growth and
147 wood properties, are quantitative traits and usually have a complex genetic
148 architecture controlled by many loci of small effect. Here we utilised a recently
149 developed method for the dissection of complex traits, FarmCPU, that has been
150 proposed to efficiently address problems confounding between testing markers and
151 covariates that often arise in GWAS (Liu et al., 2016). Several empirical studies
152 have verified that FarmCPU offers enhanced power for GWAS of complex traits
153 (Vanous et al., 2019; Ward et al., 2019; Zhu et al., 2018). In this study, we
154 identified 78 and 82 significant associations having additive and dominant effects,
155 respectively, with average of 13 SNPs identified per trait studied. These results are
156 more efficient compared to another commonly used GWAS method, Genome-wide
157 Efficient Mixed Model Association (GEMMA). In preliminary analyses we found
158 that GEMMA was also able to control the false positive rate very well, but this
159 came at the price of a relatively low statistical power. GEMMA consequently only
160 identified two significant SNPs for additive effects (Figure S4) and a total of 13
161 significant SNPs for the dominance effects across traits (Figure S5). The FarmCPU
162 methods therefore appears to be an attractive method that strike a good balance

163 between the identification of false positives and false negatives and thus have good
164 power for the dissection complex traits.

165 ***Dominance effects play important roles in hybrid population***

166 GWAS have traditionally assumed only additive effects of individual SNPs
167 (Bush and Moore, 2012; Marjoram et al., 2014) but here we show the added value
168 of also considering dominance effects for identifying genomic regions controlling
169 growth and wood quality traits. By assessing also dominance effects, we identify
170 an additional 72 associated SNPs across the traits, in addition to the 78 SNPs we
171 identified using additive effects. Furthermore, a considerable proportion of the
172 genetic variation in our hybrid population is attributable to non-additive effects and
173 our results show that the alleles underlying this variation can be identified when
174 dominance effects are explicitly considered in a GWAS setting. Several previous
175 studies have used controlled crosses in crop species, particularly in maize and rice,
176 to identified loci that exhibit dominance effects. For heterosis-related traits, data
177 from maize is frequently cited as supporting the dominance model (Cui et al., 2017;
178 Wallace et al., 2014), while rice has been proposed as a system that supports the
179 over-dominance hypothesis (Li et al., 2016; Zhen et al., 2017). Our approach
180 quantifies the contribution of dominance to the “missing heritability” in a
181 *Eucalyptus* hybrid breeding population and we collectively show that up to 10% of
182 the genomic-based heritability can be explained by associated SNPs that were
183 identified using a dominance model (Table S4).

184 ***The benefit of integrating GWAS results on genomic prediction***

185 Even if we capture a substantially larger number of associated SNPs by
186 considering both additive and dominance effects, a large fraction of the genomic-
187 based heritabilities (14%-62%) cannot be explained by only considering
188 significantly associated SNPs (Table S4) and these observations are in line with
189 several earlier reports (Chhetri et al., 2019b; Tang et al., 2019; Zhao et al., 2019).
190 Also, when using significantly associated SNPs from the GWAS to investigate the
191 accuracy of genomic prediction, we find that this yields no improvement in
192 accuracy, and sometimes even reduced accuracy, compared to predictions based on
193 all available SNPs which mirrors results seen in other similar studies (Gowda et al.,
194 2015; Wallace et al., 2016). Regions identified in a GWAS are consequently not
195 able to explain all of the genetic variation in the traits of interest and this problem
196 is greater for quantitative traits that are controlled by many genes of small effect.
197 These are the traits where current GWAS methods often suffers from insufficient
198 power to detect loci of small effect, unless sample sizes are substantially larger
199 than what is commonly used in most studies of plants.

200 In order to assess if the GWAS results could be used to enhance genomic
201 prediction in our breeding population, we also tried to identify possible ‘candidate’
202 SNPs that were not detected as significant using the stringent significant threshold
203 we applied in our GWAS. The rationale here is that, as outlined above, most GWAS
204 methods fail to detect loci of small effect but that the GWAS would nevertheless
205 serve as a useful ‘filter’ for ranking SNPs for their possible effects on the traits of
206 interest. We therefore selected two categories of SNPs using two different criteria

‡07 of relaxed significance and used these to estimate genomic heritabilities and
‡08 perform genomic prediction. The first category, ‘putative’ SNPs include all SNPs
‡09 that were found to be associated with the traits of interest based a more relaxed p -
‡10 value ($p < 1E-3$). Using this more relaxed p -value we identify between 70 to 184
‡11 SNPs for the different of trait when combined across the additive and dominance
‡12 effect models. Using the ‘putative’ SNP category we observed large improvements
‡13 in the heritability estimates for the growth traits, to the point where almost all of
‡14 the genetic variation could be explained (Table S4). For wood quality traits,
‡15 however, about 40% of the genetic variation remain unexplained compared to
‡16 when using all SNPs for heritability estimation (Table S4). The second category of
‡17 SNPs we considered consisted of the top 1% of SNPs, ranked by the p -value from
‡18 the GWAS. Using this criterion ensures that the same number of SNPs are used for
‡19 prediction across the different traits. Surprisingly we were able to explain a
‡20 substantially greater proportion, up to 174%, of the genetic variation explained
‡21 when using all SNPs (Table S4). When we performed genomic prediction using
‡22 these two categories of SNPs we also observe a substantial increase in the
‡23 prediction ability for all traits compared to predictions based on all available SNPs.
‡24 This suggests that using all available SNPs introduce noise in the prediction models
‡25 that negatively affects our prediction ability. Our method for analysing genomic
‡26 selection and increasing prediction accuracy clearly benefited from integrating
‡27 results from the GWAS analyses, but the number of associated SNPs that needs to
‡28 be incorporated depends on the study trait in questions.

‡29 ***Detection of associations for complex traits in forest trees***

‡30 Identifying candidate genes underlying growth and wood traits has long been
‡31 an active area of research in forest trees, such as in *Eucalyptus* (Cappa et al., 2013;
‡32 Müller et al., 2019; Muller et al., 2017; Resende et al., 2017a), *Populus* (Allwright
‡33 et al., 2016; Du et al., 2016; Fahrenkrog et al., 2017; Porth et al., 2013) and *Pinus*
‡34 (Bartholomé et al., 2016; Lu et al., 2017). To ensure good statistical power, both
‡35 common and rare genetic variants needs to be considered to have a comprehensive
‡36 understanding of the genetic regulation of complex traits, since many low-
‡37 frequency variants were identified as associated with growth and wood
‡38 composition traits (Fahrenkrog et al., 2017). For instance, regional heritability
‡39 mapping (RHM), has previously been shown to successfully utilise information
‡40 from both common and rare variants and can therefore capture a larger proportion
‡41 of the genomic heritability in *Eucalyptus* (Müller et al., 2019; Resende et al.,
‡42 2017a).

‡43 Furthermore, both additive and non-additive effects play important roles in
‡44 association studies for many traits. Adding dominance effects to a GWAS analysis
‡45 increase the possibility to identify additional variants that can help capture a greater
‡46 fraction of the genetic variance (Du et al., 2016; Lu et al., 2017). Other methods,
‡47 such increasing the sample size using meta-analysis (Müller et al., 2019) or using
‡48 multi-locus GWAS approaches instead of single marker methods (Fahrenkrog et al.,
‡49 2017) are methods that also can help increase statistical power in GWAS.

‡50 ***Putative genes for plant growth and stress response***

‡51 Among the significantly associated SNPs we observe across additive and
‡52 dominance effects estimations, we identified a total of 49 candidate genes that have
‡53 known functions relevant for the traits in question. The details of these genes,
‡54 including information on the position of associated SNPs and the putative functions
‡55 of the genes, are summarised Table S6. In general, candidate genes can be
‡56 separated into two groups, with one group containing genes that have direct
‡57 functions associated to the morphological formation of different tissues or organs.
‡58 The other group contain genes related to general responses to abiotic and biotic
‡59 stress, which, more indirectly, influence plant growth and biomass.

‡60 Among the significant SNPs associated with morphology, a number of
‡61 associations are linked to genes which are related to cell wall biosynthesis. For
‡62 example, SNP Chr3.46653967 is associated with Ht6 using an additive effects
‡63 model. This SNP is located on chromosome 3 and encodes a missense variant in a
‡64 gene coding for a pectin lyase-like superfamily protein (PME). This gene is
‡65 expressed in stamen and is involved in cell wall loosening and have previously
‡66 been implicated in floral development (Francis et al., 2006). We also identified a
‡67 significant SNP on chromosome 11 (Chr11.20479646) which is associated with
‡68 Ht6 (Table S6). This SNP is located in the gene *Eucgr.K01691* which encodes a
‡69 homolog to the *Arabidopsis* alpha-L-arabinofuranosidase 1 (*ARAF1*) gene.
‡70 Expression of the *ARAF1* gene is localized to several cell types in the vascular
‡71 system of roots and stems and the protein is known to be involved in cell type-
‡72 specific alterations of cell wall structure (Chávez Montes et al., 2008). Many other

‡73 studies have also identified cell wall biosynthesis related genes from GWAS
‡74 performed using growth traits in forest trees. Du *et al.* (2016) identified four
‡75 significant SNPs that were located in genes involved in secondary cell wall
‡76 biosynthesis when analysing growth traits in the *Populus* (Du et al., 2016). A SNP
‡77 associated with volume in *E. pellita* is located in a gene whose function is known
‡78 to be involved in cell wall cellulose biosynthesis (Muller et al., 2017). Similarly,
‡79 Muller *et al.* (Müller et al., 2019) used a joint-GWAS approach in four *Eucalyptus*
‡80 breeding populations and identified eight SNPs associated with growth traits that
‡81 were all linked to genes which were related to cell wall biosynthesis.

‡82 Many of the candidate genes putatively related to abiotic and biotic stress
‡83 show response to adverse conditions. It is perhaps not surprising that these genes
‡84 show up in our GWAS, as the planting area of the study population alternates
‡85 between extremely dry (from July to August) and wet (from August to October)
‡86 conditions in most years, which often leads to stress-induced damage and high
‡87 incidence of diseases. In line with this, we identified several candidate genes
‡88 involved in stress response to adverse climate conditions. The SNP Chr2.1760161
‡89 is highly associated (p -value=3.72E-12) with height at 3 years age in the
‡90 dominance model. This SNP is located upstream of the gene *Eucgr.B00092*, which
‡91 encodes a putative *HVA22* homologue E (*HVA22E*). *HVA22E* is upregulated to
‡92 varying degrees in response to cold and salt stress, ABA treatment or dehydration
‡93 (Chen et al., 2002; Shen et al., 2001). Another variant (Chr3.41941452), associated
‡94 with CBH at age 6, is located in the upstream region of *High-affinity K+*

i95 *transporter 1* (HKT1) gene. *HKT1* is expressed in root stelar cells and leaf cells
i96 (Hamamoto et al., 2015) and provides a key mechanism for protecting leaves from
i97 Na⁺ over-accumulation and salt stress (Berthomieu et al., 2003; Maser et al., 2002).
i98 The SNP Chr6.23066996 is associated with CBH at 6 years of age in the additive
i99 effects model, is located on chromosome 6 inside the gene *Eucgr.F01775* that
i00 encodes catalase 2 (*CAT2*). *CAT2* controls levels and sensitivity to H₂O₂ (Bueso et
i01 al., 2007), photo-oxidative stress (Konert et al., 2015) and auxin levels (Gao et al.,
i02 2014).

i03 Four of the candidate genes we identify in our GWAS have functions in both
i04 morphological formation and stress response. One common SNP (Chr11.
i05 28479550), associated with CBH6 in both the additive and dominance models as
i06 well as with Ht6 for the dominance model is located in the vicinity of the gene
i07 *Eucgr.K02133* which encodes a nucleotide-diphospho-sugar transferase (*QUA1*).
i08 This enzyme is expressed in vascular tissues and affects homogalacturonan, pectin
i09 and hemicellulose cell wall synthesis (Orfila et al., 2005). Recent studies have
i10 shown that *QUA1* also functions in chloroplast-dependent calcium signalling under
i11 salt and drought stresses (Zheng et al., 2016). Finally, the SNP Chr4.11644680 is
i12 associated with Ht3 and is a synonymous variant located in the *SFR6/MED16* gene
i13 which plays important roles in cold- and drought-inducible gene expression
i14 (Knight et al., 2009, defence gene expression {Wathugala, 2012 #739) as well as
i15 modulating iron uptake (Zhang et al., 2014) in response to cell wall defects (Sorek

i16 et al., 2015). These findings suggest that stress resistance also plays an important
i17 role in affecting tree growth traits.

i18 **Conclusions**

i19 In this study, we have used a GWAS approach in a *Eucalyptus* hybrid
i20 population to dissect the genetic basis of growth and wood quality traits by
i21 accounting for both additive and dominance genetic effects. Altogether we identify
i22 78 and 82 significant SNPs using additive and dominance models, respectively,
i23 with 10 SNPs showing an overlap between the two effect models, suggesting that
i24 additive and dominance effects are not completely independent. The associated
i25 genes could be grouped into two broad functional categories relating to how they
i26 influence tree growth and biomass. One group contain genes associated with
i27 morphological formation, such as cell wall biosynthesis, and the other group
i28 contain genes related to abiotic and biotic stress responses, such as oxidative,
i29 hormone-based and disease-induced stress. These results provide novel targets for
i30 possible transgenic or genome editing approaches in the future to directly improve
i31 growth and biomass related traits.

i32 We also applied our results from the GWAS in a genomic selection analysis
i33 by using different categories of SNPs selected based on the GWAS results and used
i34 them to evaluate genomic-based heritabilities and predictive abilities. Our results
i35 show that prediction abilities of the estimated breeding values improved for all
i36 traits when using SNPs selected based on the GWAS results. Integrating GWAS

i37 results into genomic selection thus appear to be a promising avenue to increase the
i38 efficiency of genomic selection in forest breeding.

i39 **Experimental procedures**

i40 *Populations, phenotypic and genotypic data*

i41 A total of 1123 *Eucalyptus* individuals were used in this study, including 90
i42 *E.grandis*, 84 *E.urophylla* parents and 949 F1 progenies derived from a random
i43 mating design that has previously been described (Tan et al., 2017). Briefly, F1
i44 individuals were identified to be comprised of inter- and intra-crossing of the two
i45 parental species. Of the 949 F1 individuals, 57% were interspecific *E.grandis* ×
i46 *E.urophylla* hybrids, 21% were intraspecific *E.grandis* × *E.grandis* progeny and 22%
i47 were intraspecific *E.urophylla* × *E.urophylla* progeny (Tan et al., 2018).

i48 The phenotypic and genotypic data utilized in this study has been previously
i49 described in detail (Tan et al., 2017). The phenotypes include height and
i50 circumference at breast height (CBH), where F1 individuals were evaluated at ages
i51 three and six and the pure species parents were evaluated at age five. In addition,
i52 we obtained data on two wood quality traits, basic density and pulp yield, that were
i53 assessed at age five. Genotyping was performed using an Illumina Infinium
i54 EuCHIP60K SNP chip that contains probes for 60,904 unique SNPs (Silva-Junior
i55 et al., 2015). Across the 1123 individuals, 37,832 SNPs were retained after quality-
i56 control based on call rates (>0.7) for both SNPs and samples and following
i57 filtering based on minor allele frequencies (>0.01) and deviations from Hardy-

i58 Weinberg equilibrium ($>1e-7$). Any missing data remaining in the 37,832 SNPs
i59 were subsequently imputed using BEAGLE 4.1 (Browning and Browning, 2007).

i60 ***Phenotypic data analyses***

i61 Phenotype data for the parental and F1 population were adjusted separately to
i62 minimize environmental variation by fitting a mixed linear model for each trait:

i63
$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}_b\mathbf{r} + \boldsymbol{\varepsilon} \quad (1)$$

i64 where \mathbf{y} is the vector of phenotypic observation, $\boldsymbol{\beta}$ is the vector of overall mean as
i65 fixed effect, \mathbf{r} is the vector of random replication effects following $\mathbf{r} \sim N(0, I\sigma_r^2)$,
i66 where σ_r^2 is the replication variance, $\boldsymbol{\varepsilon}$ is the vector of random residual effects. \mathbf{X}
i67 and \mathbf{Z}_b is design matrix for $\boldsymbol{\beta}$ and \mathbf{r} , respectively. For the F1 population, the
i68 residual variance-covariance matrix is $R = I\sigma_e^2 + AR1(\rho_r) \otimes AR1(\rho_c)\sigma_\eta^2$, where
i69 $AR1(\rho_r)$ and $AR1(\rho_c)$ are autoregressive correlation matrices for the row model
i70 (autocorrelation parameter ρ_r) and column model (autocorrelation parameter ρ_c),
i71 respectively. σ_e^2 is the independent residual variance while σ_η^2 is the spatial variance.
i72 For the pure parental species, we fitted the model in Equation 1 by setting the
i73 residual matrix $R = I\sigma_e^2$ since spatial coordinates and position information were
i74 not available for these individuals. All mixed-linear model analyses were
i75 performed in ASReml 4 (Gilmour et al., 2015). Phenotypes of individuals from the
i76 F1 and parental populations were adjusted for random block effects (r) and spatial
i77 effects (s), respectively.

i78 The heritability (h^2) was estimated using a mixed model $\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}\mathbf{u} + \boldsymbol{\varepsilon}$,
i79 where \mathbf{y} represents the adjusted phenotypes of single trait, $\boldsymbol{\beta}$ is the vector of fixed
i80 effects, including overall mean and age difference. \mathbf{u} is a vector of random additive
i81 or dominance genetic effect of individuals with a normal distribution, $\mathbf{u} \sim N(0, \mathbf{A}\sigma_a^2)$,
i82 \mathbf{A} being the revised, pedigree-based genetic relationships among individuals (Tan
i83 et al., 2018); and $\boldsymbol{\varepsilon}$ is a heterogeneous random residual effects represented different
i84 experimental sites. \mathbf{X} and \mathbf{Z} are incidence matrices for $\boldsymbol{\beta}$ and \mathbf{u} , respectively. We
i85 obtained restricted maximum likelihood (REML) estimates of σ_a^2 and $\boldsymbol{\varepsilon}$, and
i86 estimated $h^2 = \sigma_a^2 / (\sigma_a^2 + \sum \varepsilon/n)$.

i87 ***Population structure, kinship analysis and GWAS***

i88 In the association analyses, confounding effects of population structure and
i89 kinship between individuals need to be accounted for. Population structure (Q) was
i90 estimated using a model-based clustering and through principle component analysis
i91 (PCA) using 13,245 independent SNPs obtained by LD-pruning the original SNP
i92 data set and including only SNPs that have pairwise linkage disequilibrium (LD)
i93 values (r^2) less than 0.2. Model-based clustering was implemented using the
i94 software *admixture* v.1.3.0 which infer population structure by estimating
i95 individual admixture proportions using multi-locus SNP data through a maximum-
i96 likelihood method (Alexander et al., 2009). The number of ancestral populations (P)
i97 was varied from 1 to 100 when using *admixture* and *fastStructure* (Raj et al., 2014).
i98 Five-fold cross-validation (CV) was performed to choose the optimal P value in

599 admixture. A PCA was also performed using the *smartpca* program in *eigensoft*
600 v6.0 to estimate individual ancestry proportions (Price et al., 2006).

601 GWAS was conducted using a recently developed method, FarmCPU, which
602 explicitly takes into account the confounding that exists between covariates and test
603 marker by using both a fixed effect model and a random effect model (Liu et al.,
604 2016). The results from the PCA and the kinship matrix were used as covariates in
605 FarmCPU to account for population structure and relatedness among samples,
606 respectively. We ran the GWAS using the R package *FarmCPU*. False positive
607 errors due to multiple testing were controlled by an adjusted Bonferroni method,
608 *simpleM* (Gao et al., 2008). This method infers the number of independent SNPs by
609 filtering on LD and performs a standard Bonferroni correction to correct for
610 multiple testing based on the number of ‘independent tests’ performed. For the
611 present data a p -value $< 1.7E-06$ was selected as a cut-off to indicate significant
612 associations.

613 ***GWAS for additive and dominance models***

614 We conducted GWA analyses in FarmCPU using either an additive or
615 dominance encoding of genotypes. For the additive encoded data, the homozygous
616 major allele was encoded with 0, the heterozygous genotype with 1 and the
617 homozygous minor allele with 2. For the dominance encoding, both homozygous
618 minor and major alleles were encoded as 0 whereas the heterozygous genotype was
619 encoded as 1 (Seymour et al., 2016).

620 ***Genomic selection (GS) with different informative SNPs***

i21 Genomic selection (GS) models were constructed based on four different
i22 categories of SNPs using the Genomic Best Linear Unbiased Prediction (GBLUP)
i23 method. Details on how the genomic-based additive and dominance relationship
i24 matrices are estimated have been previously described in detail in Tan *et al.* (Tan et
i25 al., 2018). Here we focus on the details of how the four categories of SNPs we have
i26 used in all subsequent analyses were selected. The four categories of SNPs
i27 employed for estimating the additive and dominance relationship matrices were: 1)
i28 ‘*associated SNPs*’ which contain only SNPs that were identified as significantly
i29 associated with the corresponding trait at the Bonferroni-adjusted p -value $< 1.7E-6$;
i30 2) ‘*putative SNPs*’ are all SNPs that were significant in the GWAS for the
i31 corresponding trait using a more relaxed p -value threshold ($p < 1E-3$) in order to
i32 capture also possible causal SNPs that do not reach significance using the more
i33 stringent criteria in the original GWAS; 3) The ‘*top 1% SNPs*’ category use the top
i34 378 SNPs for each trait in the GWAS ranked after p -value in the GWAS in order to
i35 evaluate the same number of SNPs across different traits when building genomic
i36 selection models; and finally 4) ‘*all SNPs*’ which use all of the 37,832 SNPs
i37 available and is therefore identical to the models originally used in Tan et al (2017)
i38 and Tan et al (2018).

i39 Two separate GBLUP models were evaluated that included i) either only
i40 additive (A) or ii) both additive and dominance (AD) genetic effects using the four
i41 SNP categories described above to create marker-based relationship matrices. The
i42 A and AD models have been well described earlier in Tan *et al.* (Tan et al., 2018).

i43 The genomic-based narrow- and broad-sense heritability (h^2 and H^2 respectively)
i44 were calculated after fitting each model across the different traits. Narrow-sense
i45 heritability in the A model was estimated as $h^2 = \sigma_a^2 / \sigma_p^2$ and the broad-sense
i46 heritability of AD model was estimated as $h^2 = (\sigma_a^2 + \sigma_d^2) / \sigma_p^2$, where σ_a^2 , σ_d^2 and
i47 σ_p^2 represented the estimated additive, dominance and phenotypic variance,
i48 respectively. The prediction ability was estimated for all models and relationship
i49 matrices using a ten-fold cross-validation scheme where 100 replications was
i50 implemented to evaluate the prediction accuracy of the different models. For each
i51 replication, the dataset was randomly divided into 10 subsets and nine out of the
i52 ten partitions were used as the training population to fit a model using both
i53 phenotypes and genotypes, while the remaining partition was used as the validation
i54 set where phenotypic data was removed and then used to predict breeding values or
i55 total genetic values for the model in question. The predictive ability of the model
i56 was evaluated by estimating the correlation between phenotypes and
i57 breeding/genetic values, $r(\hat{A}_{vali}, Y_{vali})$ or $r(\hat{G}_{vali}, Y_{vali})$.

i58 ***Assigning significant SNPs to putative candidate genes***

i59 Genes within ± 5 kb away from a SNP that was significantly associated with a
i60 measured phenotypic trait were extracted from the *E.grandis* v2.0 reference
i61 genome (BRASUZ1) in Phytozome (www.phytozome.net) using SnpEff v4.2
i62 (Cingolani et al., 2012). The 5kb window threshold used was based on the distance
i63 over which LD decays in this population (Tan et al., 2017). The putative functions

064 of these candidate genes were determined based on their homology to functionally
065 characterized genes in *A. thaliana* (TAIR).

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