THE SPATIAL REGULATION OF CONDENSIN ACTIVITY IN CHROMOSOME CONDENSATION

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Abstract

Condensin mediates chromosome condensation, which is essential for proper chromosome segregation during mitosis. We show that in budding yeast, polo kinase Cdc5p stimulates condensin to promote condensation of the ribosomal DNA (RDN). We show that cohesin, a complex related to condensin, recruits Cdc5p to the RDN. This spatial control of condensin activity prevents misfolding of the RDN into an irreversibly decondensed state. This regulation is also necessary to promote condensation of the RDN into a thin loop, distinct from the rest of the chromosomes. This thin condensed structure is reminiscent of constrictions associated with the pericentric and ribosomal loci of mitotic chromosomes in animals and plants. Therefore, we propose that the localized stimulation of condensin by cohesin and polo kinase described in the yeast RDN could be an evolutionarily conserved mechanism to generate a distinct type of chromosome folding at specific loci.

Introduction

Mitotic chromosomes have two structural features, sister chromatid cohesion and chromosome condensation, which are essential for their proper inheritance during cell division. Sister chromatid cohesion is established concurrently with DNA replication and

is maintained until the onset of anaphase. In contrast, condensation is established in mitotic prophase and persists through anaphase until the end of mitotic division. As a result, cohesion and condensation coexist from prophase until the onset of anaphase, a period henceforth referred to as mid-M. Perturbing proper condensation or cohesion leads to aneuploidy and chromosome rearrangements, hallmarks of cancer, birth defects and several genetic disorders (Hassler et al., 2018). Over the last two decades, scientists have made significant progress in identifying the protein complexes that mediate cohesion and condensation and understanding their cell cycle regulation. However, the potential interplay between these complexes has remained largely uninvestigated.

Cohesion and condensation are mediated respectively by cohesin and condensin, two related protein complexes in the SMC (structural maintenance of chromosomes) family (Figure 1A) (Hassler et al., 2018). Cohesin intermolecularly tethers DNA of the sister chromatids, resulting in cohesion, while condensin promotes condensation by tethering DNA intramolecularly. Condensin is also thought to promote condensation by inducing supercoils and by actively extruding DNA to form loops (Ganji et al., 2018; Hagstrom et al., 2002; Kimura et al., 2001; Kimura and Hirano, 1997; St-Pierre et al., 2009). In principle, the existence of distinct SMC complexes dedicated to cohesion and condensation would allow these processes to occur independently. Consistent with this conclusion, mammalian chromosomes can condense in the absence of cohesin (Losada et al., 1998; Sonoda et al., 2001).

The independence of cohesion and condensation also seems to be supported by studies of their regulation. The establishment of cohesion in S phase is regulated by Eco1p-dependent acetylation of cohesin (Rolef Ben-Shahar et al., 2008; Unal et al., 2008), while its maintenance is mediated by interactions with Pds5p, a cohesin auxiliary factor (Noble et al., 2006). In contrast, the phosphorylation of condensin by different kinases mediates the establishment and maintenance of condensation. Cyclin-dependent kinases (CDKs) are required to establish condensation in prophase (Kimura et al., 1998). Polo kinase and Aurora kinase are important for the maintenance of condensation after the onset of anaphase when CDK activity diminishes (Lavoie et al., 2002, 2004; Robellet et al., 2015; St-Pierre et al., 2009). These observations suggest cohesion and condensation are mediated by distinct protein complexes with distinct regulatory circuits. However, cohesin inactivation abrogates condensation as well as cohesion in mitosis of budding yeast, meiosis of fission yeast, and mouse oocytes and sperm (Ding et al., 2006; Guacci and Koshland, 2012; Guacci et al., 1997b; Novak et al., 2008). A subsequent study in budding yeast showed that cohesin is not required for condensation per se, but rather prevents condensin from permanently misfolding chromosomes into an irreversibly decondensed state (Lavoie et al., 2002). How cohesin regulated condensin in mid-M remained a mystery.

A candidate for mediating this regulation was polo kinase. In budding yeast, polo kinase (Cdc5p) binds to cohesin associated regions (CARs) on chromosomes, and the binding is dependent upon cohesin (Pakchuen et al., 2016). Cdc5p also phosphorylates condensin in mid-M (Archambault et al., 2015; Pakchuen et al., 2016; St-Pierre et al.,

2009; Walters et al., 2014). While these observations connect Cdc5p with cohesin and condensin, inactivation of Cdc5p with temperature-sensitive allele did not lead to a mid-M condensation defect (St-Pierre et al., 2009). In contrast, here we show that Cdc5p is required for both the establishment and maintenance of condensation in mid-M, likely by promoting condensin activity to make chromosome folds. We provide additional evidence that suggests cohesin recruits Cdc5p to specific genomic regions facilitating the activation of adjacently bound condensin. This spatial and temporal activation of condensin promotes condensation by preventing condensin-mediated chromosome misfolding.

Results

The polo kinase Cdc5p is essential for mid-M chromosome condensation.

The absence of cohesion and condensation defects in mid-M for *cdc5* temperature-sensitive alleles suggested that Cdc5p was not required for mid-M chromosome structure. However, we postulated that these temperature-sensitive mutations may not have completely abrogated Cdc5p function (St-Pierre et al., 2009). With this in mind, we generated a more stringent loss-of-function *cdc5* allele to study the role of Cdc5p in cohesion and condensation. To assess how essential is Cdc5p function in these processes, we compared Cdc5p depletion with depletion of cohesin and condensin.

We took advantage of the auxin-induced degron (AID) system to generate a *cdc5* allele (Cdc5p-AID) (Nishimura et al., 2009). Briefly, in this system, the fusion of Cdc5p with an AID tag allows for near wild-type function in the absence of auxin. However, in the presence of auxin, tagged proteins are targeted by the E3 ligase TIR1 for rapid ubiquitin-

dependent degradation (Figure 1B, top). We used the AID system to also deplete cohesin (Mcd1p-AID) and condensin (Brn1-AID). After depletion, the AID proteins were undetectable by western blot and all strains were inviable (Figure 1—figure supplement 1). Indicated yeast strains were first synchronized in G1, then released into media with auxin to deplete the AID-tagged protein and nocodazole to arrest cells in mid-M (Figure 1B, bottom). Using this protocol (referred to as staged mid-M) cells were depleted of the AID-tagged protein from G1 to mid-M. We proceeded in assessing sister chromatid cohesion and condensation.

We measured sister chromatid cohesion using a well-characterized GFP dot assay (Straight et al., 1996). In this system, a lacO array is integrated at a chromosomal locus and bound by lacI-GFP. If proper sister chromatid cohesion is present, the two sister chromatids are held together, and the two arrays appear as one GFP spot. Conversely, if cohesion is absent, two distinct GFP spots are observed (Guacci et al., 1997a; Michaelis et al., 1997). We tested wild type, Cdc5p-AID depleted and Mcd1p-AID depleted cells. Mcd1p depletion served as positive control for the abrogation of cohesion. Cells lacking Cdc5p-AID had a three-fold decrease in sister chromatid cohesion compared to wild-type cells (Figure 1C). This defect was significantly less than the nine-fold decrease in cohesion in cells depleted for Mcd1p-AID. Thus, Cdc5p is important, but not essential for sister chromatid cohesion.

We performed a time-course experiment for cohesion to assess whether Cdc5p functions in the establishment or maintenance of cohesion. Depletion of factors that are important

for the establishment of cohesion resulted in a high level of separated sister chromatids from the end of S-phase till mid-M. While depletion of factors that are involved in the maintenance of cohesion resulted in a gradual loss of cohesion that starts at the end of S-phase and peaks at mid-M (Eng et al., 2014; Jin et al., 2009). We assessed cohesion on wild-type, Cdc5p depleted, and Pds5p depleted cells and assayed cohesion at 15-minute intervals from G1 to mid-M. Pds5p is a factor required for the maintenance of sister chromatid cohesion after S phase. Under this regimen, Cdc5p-AID depleted cells presented a weaker defect in cohesion maintenance compared to cells depleted of Pds5p-AID (Figure 1D). We concluded that Cdc5p is not involved in establishing cohesion but is partially required for cohesion maintenance. Furthermore, cells that expressed a kinase-dead version of Cdc5p in combination with depletion of the endogenous copy of Cdc5p did not present the maintenance defect and were indistinguishable from wild type (Figure 1D). Therefore, the partial requirement of Cdc5p in cohesion maintenance is independent of its kinase activity.

Using the same regimen described above, we also interrogated the role of Cdc5p in mid-M chromosome condensation by assessing the morphology of the ~1.4 megabases ribosomal DNA model locus (RDN). The RDN contains 75-100 tandem repeats of the 9.1 kb rDNA and undergoes a series of distinct, cell cycle-dependent morphological changes (Figure 1E, top) (Guacci et al., 1994). In interphase, the RDN is a puff that is segregated to the periphery of the bulk chromosomal mass. In contrast, in mid-M, the RDN forms a condensed loop. This dramatic cytological RDN change is a tractable assay for condensation. In contrast, the rest of the genome only undergoes two-fold compaction

that is only measurable by laborious FISH or GFP assays (Guacci et al., 1994; Vas et al., 2007). In mid-M arrested wild type, around 25% of cells fail to condense the RDN. In contrast, approximately 75% of Cdc5p-AID depleted cells failed to condense the RDN, similarly to depletion of a core condensin subunit, Brn1p-AID (Figure 1E, bottom and Figure 1—figure supplement 2A). Therefore, Cdc5p-AID depletion caused a severe condensation defect. Ectopic expression of a kinase-dead Ccd5p did not rescue the condensation defect of cells depleted for Cdc5p-AID. Thus, in contrast to cohesion, Cdc5p and its kinase activity play a crucial role in RDN condensation in mid-M.

To assess the role of Cdc5p in the establishment of condensation, we performed a time course following RDN condensation in Cdc5p-AID depleted cells. 90 minutes is the earliest time point after release from G1 where we could observe condensed RDN loops. Cells depleted for Cdc5p did not establish condensation at 90 minutes or later (Figure 1—figure supplement 2B). Instead, control cells depleted for the maintenance factor Pds5p showed condensed loops at 90 minutes that were lost over time. Therefore, Cdc5p is required for the establishment of condensation. To assess whether Cdc5p also functions in condensation maintenance, we modified the auxin regiment of *CDC5-AID* cells. We first arrested them in mid-M by adding nocodazole, allowing RDN condensation to occur (Figure 1—figure supplement 3) and then we added auxin to deplete Cdc5p-AID (Figure 1F, top). We observed a dramatic loss of RDN condensation in cells depleted of Cdc5p-AID in mid-M like that observed in cells depleted for Brn1p-AID (Figure 1F, bottom). Therefore, Cdc5p is an essential regulator for the establishment and maintenance of condensation in mid-M.

Cdc5p stimulates condensin-dependent RDN condensation.

How does Cdc5p regulate condensation? Insights come from further examining the RDN condensation defects. Puffed RDN in mid-M can result from either an unfolded state such as is normally seen in G1 or from a misfolded state (Lavoie et al., 2002). The unfolded state is still competent to be folded, while the misfolded state is irreversibly trapped in a decondensed manner (Figure 2A). These two states were previously distinguished using a simple genetic test referred to here as add later test (Figure 2B, top and (Lavoie et al., 2002)). Briefly, temperature-sensitive condensation factors were inactivated from G1 to mid-M preventing condensation, then re-activated in mid-M and the reversibility of the condensation defect was assessed. With this test, we showed delayed activation of condensin restores RDN condensation, while delayed activation of cohesin did not (Figure 2B, bottom and (Lavoie et al., 2002)). Therefore, condensin inactivation results in an unfolded state, while cohesin inactivation results in an irreversibly misfolded state.

Cdc5p depletion can result in an unfolded state like condensin, or a misfolded state like cohesin. We performed the add later experiments using our AID system to deplete Mcd1p-AID, Brn1p-AID, and Cdc5p-AID (Figure 2—figure supplement 1). As expected, depletion of any of these three proteins between G1 and mid-M prevented the appearance of folded RDN (Figure 2C, gray bars). When Cdc5p-AID was expressed in Cdc5p-AID depleted cells, the RDN condensed (Figure 2C, blue bars). The same result was observed when Brn1p-AID was expressed in Brn1p-AID depleted cells. Conversely, when Mcd1p-AID was expressed in Mcd1p-AID depleted cells, the RDN remained

decondensed. These results show that depletion of Cdc5p-AID, like depletion of condensin, results in a reversible unfolded RDN state, rather than misfolded like depletion of cohesin.

We hypothesized that the RDN misfolding that occurred after cohesin depletion was condensin dependent. If this model was correct misfolding of the RDN due to cohesin depletion could be prevented by co-inactivation of condensin or any factor that stimulates condensin activity. To test this prediction, we depleted and added later two components together (Figure 2—figure supplement 1). When we co-depleted and added later cohesin and condensin we restored condensation, thus preventing RDN misfolding caused by the absence of cohesin alone (Figure 2D and (Lavoie et al., 2002)). When we depleted and added later cohesin and Cdc5p-AID together, we also prevented RDN misfolding caused by the absence of cohesin (Figure 2D). To explain these genetic results, we propose that Cdc5p is a factor that stimulates condensin activity. When cohesin is present, Cdc5p stimulates condensin to promote proper RDN folding generating condensation. When cohesin is missing, Cdc5p inappropriately stimulates condensin resulting in a misfolded decondensed RDN.

Cohesin-dependent recruitment of Cdc5p to the RDN promotes its binding to condensin in the RDN.

To dissect the molecular mechanism for how Cdc5p, cohesin, and condensin regulate condensation, we examined their genomic localization by ChIP-seq. We analyzed the binding of Cdc5p, cohesin, and condensin in wild-type and Cdc5p-, cohesin-, and

condensin-depleted cells. We also performed an important ChIP-seq control in yeast by immunoprecipitating GFP from GFP-expressing cells. This ChIP-seq provided us with the background signal of genomic regions prone to entrapment by ChIP-seq in the absence of a real signal (Teytelman et al., 2013).

Using the staged mid-M regimen, we first assessed the binding of condensin (Ycg1p-Myc), cohesin (Mcd1p), and Cdc5p (Cdc5p-Flag) in wild-type cells (Figure 3). We observed Ycg1p enrichment in a large pericentric region peaking at centromeres and within the end of the non-transcribed region of the RDN (NTS1), corroborating previous results (Figure 3 and (Leonard et al., 2015)). Additional peaks of enrichment for Ycg1-Myc that were previously reported (D'Ambrosio et al., 2008) were also observed along the chromosome arms, but they were likely false positives because they coincided with peaks in GFP-control (Figure 3A). Mcd1p was bound to centromeres, pericentric regions, cohesin associated regions (CARs) along chromosome arms, and a region between the two non-transcribed spacers (NTS1 and NTS2) in the rDNA repeat (Figure 3 and (Glynn et al., 2004; Laloraya et al., 2000; Pakchuen et al., 2016)). Finally, the binding pattern of Cdc5p-Flag mirrored cohesin binding genome-wide (Figure 3 and (Pakchuen et al., 2016; Rossio et al., 2010)). Importantly, we showed that Cdc5p co-localizes with cohesin on the RDN.

We then examined Brn1p, Mcd1p and Cdc5p binding patterns after depletion of Cdc5p-AID, Brn1p-AID or Mcd1p-AID. We depleted the relevant factor from G1 to mid-M using the staged mid-M regimen. The depleted factor could not be detected by western blot. The chromosomal binding pattern of Ycg1p-Myc did not change upon the depletion

of either cohesin or Cdc5p (Figure 3—figure supplement 1, Figure 3—figure supplement 2, Figure 3—figure supplement 3). Similarly, the binding pattern of Mcd1p remained unchanged upon depletion of either condensin or Cdc5p (Figure 3—figure supplement 1, Figure 3—figure supplement 2, Figure 3—figure supplement 3). Thus, cohesin and condensin bind to specific sites on chromosomes independently of each other and Cdc5p. Therefore, cohesin and Cdc5p do not regulate RDN condensation by changing condensin localization to the RDN.

We then analyzed whether changes in the localization of Cdc5p might provide insights into RDN condensation. We examined cells in which Mcd1p-AID or Brn1p-AID were depleted from G1 to mid-M using the staged mid-M regimen. Upon Mcd1p-AID depletion, the binding of Cdc5p was abolished genome-wide (Figure 3 and Figure 3—figure supplement 1, Figure 3—figure supplement 2, Figure 3—figure supplement 3), despite similar levels of total Cdc5p (Figure 3—figure supplement 4). This result recapitulated previous work showing the cohesin-dependent binding of Cdc5p at CARs (Mishra et al., 2016; Pakchuen et al., 2016). A few small peaks of binding were observed at centromere and RDN regions, but they coincided with false positives in the GFP-control. In contrast, upon Brn1p-AID depletion, the pattern of Cdc5p was largely unaffected genome-wide (Figure 3 and Figure 3—figure supplement 1, Figure 3—figure supplement 2, Figure 3—figure supplement 3). These results showed that the recruitment of Cdc5p to chromosomes (including the RDN) required the presence of cohesin over some period between G1 to mid-M.

We reasoned the purpose of cohesin recruitment of Cdc5p was to promote localized Cdc5p binding and stimulation of condensin. Therefore, we predicted that a transient association of cohesin with the RDN would be sufficient to bring some Cdc5p to condensin. To test this possibility, we synchronized *MCD1-AID* cells in mid-M in the absence of auxin to allow cohesin, condensin, and Cdc5p to normally localize to the RDN. We then added auxin to deplete Mcd1p-AID and remove cohesin. We observed that Cdc5p was eliminated throughout the genome except at a portion of the condensin-bound regions of the RDN and centromeres (Figure 4 and Figure 4—figure supplement 1). In these regions, we observed peaks of Cdc5p binding that were 3-5 fold greater than those observed when Mcd1p-AID was removed from G1 to mid-M. In contrast, the levels of the non-specific peaks, as defined by the GFP-control, did not change. The remaining portion of the Cdc5p binding when Mcd1p-AID was depleted in mid-M, was not sufficient to promote proper condensation (Supplemental Figure). Therefore, the co-localization of Cdc5 and condensin was revealed upon transient recruitment of cohesin. We propose that in wild type, high levels of cohesin at the RDN bring high levels of Cdc5p to neighboring condensin, promoting condensin activation and proper condensation.

Localization of Cdc5p with CRISPR-Cas9 to the RDN partially suppresses the condensation defect of cohesin-depleted cells.

If cohesin localization of Cdc5p to condensin at the RDN did indeed promote condensation, this function might be mimicked by ectopic localization of Cdc5p. To test this possibility, we engineered a strain harboring a fusion gene of Cdc5p and catalytically dead dCas9 under the control of the galactose promoter (Figure 5A). The fusion protein

supported the growth of yeast as the sole source of Cdc5p (Figure 5—figure supplement 1A), indicating that this fusion protein retained activity. We introduced the fusion gene into *MCD1-AID* cells with a guide RNA targeting the region of condensin binding at the RDN (NTS1 gRNA), or a distal guide RNA targeting 500 bp away (RDN5 gRNA) (Figure 5A). These strains were subjected to the staged mid-M regimen to deplete Mcd1p-AID. Expression of dCas9-Cdc5p was induced from G1 by the addition of galactose to the media (Figure 5—figure supplement 1B).

In Mcd1p depleted cells, only 6% of the cells show normal condensed RDN loops (Figure 5B). In contrast, in the presence of guide RNA proximal to condensin, we observed 17% of cells with normally condensed loops, in addition to 6% partially condensed (Figure 5B and Figure 5—figure supplement 2). The partial restoration of condensation was absent from cells with the distal guide RNA. These results showed that ectopic targeting of Cdc5p to condensin in the RDN can partially promote RDN condensation.

Discussion

In this study, we have identified an interactome of early condensation factors (comprised of cohesin, condensin, and the polo kinase Cdc5p) that control the establishment of chromosome condensation for the 1.4 megabases RDN locus of budding yeast. Our results suggest that cohesin recruits Cdc5p to the RDN and that Cdc5p actively promotes proper condensation, likely by stimulating condensin activity. When cohesin is absent, Cdc5p stimulates condensin to misfold chromosomes into an irreversibly decondensed state. Our data suggest that productive condensin activity is reliant on cohesin's spatial

regulation of Cdc5p. Altogether, our study describes a novel mechanism by which cohesin regulates condensin activity and begins to provide mechanistic insight into how SMC complexes can be spatially and temporally coordinated to achieve their functions.

Cdc5p promotes the establishment and maintenance of condensation prior to anaphase

A key finding of our study is that Cdc5p is required for both the establishment and maintenance of condensation in mid-M of budding yeast. Our results suggest that Cdc5p activates condensin and that the kinase activity is needed for this activation. We favor a model where Cdc5p-dependent phosphorylation of condensin or a condensin regulator is the key step for the establishment of condensation, and that continued phosphorylation is necessary to maintain condensation. Condensin will likely be the main Cdc5p substrate since a previous study showed that Cdc5p phosphorylates condensin prior to anaphase (St-Pierre et al., 2009). Identifying the putative Cdc5p-dependent phosphorylation sites in condensin necessary for pre-anaphase condensation will be an important, but challenging future direction given the complexity of condensin phosphorylation pattern in mitosis (Bazile et al., 2010).

The requirement for both Cdc5p and CDK kinases for the establishment of condensation prior to anaphase might reflect that Cdc5p binding to its targets requires their prior phosphorylation by CDK. Alternatively, CDKs and Cdc5p might have distinct roles in condensation. They may regulate different condensin activities like tethering DNA, supercoiling DNA and extruding DNA loops (Ganji et al., 2018; Hagstrom et al., 2002; Kimura et al., 2001; Kimura and Hirano, 1997; Lavoie et al., 2002; St-Pierre et al., 2009).

Consistent with this idea, CDK phosphorylation is limited to the Smc4p subunit of condensin (Robellet et al., 2015) while Cdc5p phosphorylates Ycg1p, Ycs4p, and Brn1p subunits.

Cdc5p and cohesin impact condensin-dependent condensation by distinct mechanisms. Here we describe three genetic pieces of evidence that differentiate the decondensed states of the RDN after depletion of cohesin, condensin, and Cdc5p. Depletion of Cdc5p or condensin results in a decondensed RDN that can be condensed upon subsequent addition of the depleted factor. These results imply that the depletion of these two factors leaves the decondensed RDN in a condensation-competent state. However, the depletion of cohesin leads to a decondensed RDN state in mid-M that cannot be condensed upon subsequent cohesin addition (this study and (Lavoie et al., 2004)). This result implies that in the absence of cohesin, the decondensed RDN is trapped in condensation-incompetent state. Finally, co-inactivation of cohesin with either condensin or Cdc5p results in decondensed RDN that can be condensed upon later addition of these factors (this study). This third observation is critical as it shows that the condensation-incompetent state in the absence of cohesin is dependent upon condensin and Cdc5p.

A simple explanation for this dependence is that cohesin normally directs Cdc5p-stimulated condensin to properly fold chromosomes. In the absence of cohesin, the RDN is misfolded into a condensation-incompetent state by Cdc5p and condensin. In the absence of cohesin and either condensin or Cdc5p, the chromosomes remain in an unfolded condensation-competent state, because misfolding is blocked. This model

introduces a critical nuance to condensin regulation. While previous studies have focused on kinase-dependent activation of condensin during the cell cycle (Bazile et al., 2010), our study reveals that polo kinase also plays an important role in controlling the quality of condensin function.

Cdc5 recruitment to the condensin-bound region of the RDN stimulates condensin promoting proper RDN condensation.

In this study, we provide important clues on how the regulation of condensin by cohesin and Cdc5p is important for the folding of the RDN. First, cohesin is required for the initial recruitment of Cdc5p to chromosomes in mid-M (this study, and (Mishra et al., 2016; Pakchuen et al., 2016)). We showed that cohesin recruitment of Cdc5p to chromosomes promotes Cdc5p binding to neighboring condensin. We suggest that cohesin recruitment either directly hands off Cdc5p to neighboring condensin or generates a high local concentration of Cdc5p that promotes its efficient interaction with condensin. By constraining the interaction of Cdc5p with condensin at the RDN, condensin activity is spatially controlled to promote proper RDN folding. In support of this model, we show that cohesin's recruitment function can be replaced by ectopic localization of Cdc5p to the RDN by its fusion with Cas9.

The spatial activation of condensin through a localized kinase adds a critical second feature to the regulation of condensation. Indeed, modulating the kinase recruitment to the target protein is a very common theme in kinase regulation (Ferrell and Cimprich, 2003; Langeberg and Scott, 2015). It will be interesting to determine whether

the regulation of condensin by CDKs and aurora kinase are also controlled spatially. However, the fact that condensation was not fully restored by the Cas9-Cdc5p fusion suggests that simply localizing Cdc5p to the RDN is not enough for efficient condensation. The localization of single Cas9-Cdc5p to the RDN might not allow sufficient phosphorylation of condensin to fully promote condensation. Alternatively, cohesin may have a Cdc5p-independent role in condensation. Discovering these features of condensin regulation will require additional studies.

In summary, the co-localization of cohesin, polo kinase and condensin underlies a regulatory pathway that spatially modulates condensin function to generate a thin loop of condensed RDN. The pericentric regions are the only other genomic loci showing similar co-localization of cohesin, condensin, and polo kinase. We predict that these factors regulate condensation of the pericentric regions to generate thinly condensed structures as they do at the RDN. In yeast, these structures would be difficult to detect cytologically because of the small size of the pericentric regions (~10 kilobases). Intriguingly, the pericentric regions are much larger in animals and plants. These regions are visible as thinner areas of the mitotic chromosomes also known as primary constrictions. Thin regions of constrictions are also observed in the nucleolus organizer regions, the rDNA loci of multicellular organisms. We propose that these constrictions are the result of the same regulatory pathway we described here for the condensation of yeast RDN. Indeed, in vitro condensed chromosomes without cohesin and polo kinase lack constrictions (Shintomi et al., 2015). In vivo, cohesin and polo kinase depleted mammalian cells also fail to form primary constrictions (Gandhi et al., 2006). More broadly this regulatory

pathway may serve as a general paradigm for how cohesin, condensin, and other SMC complexes communicate to achieve proper chromosome structure in other biological processes.

Figure 1

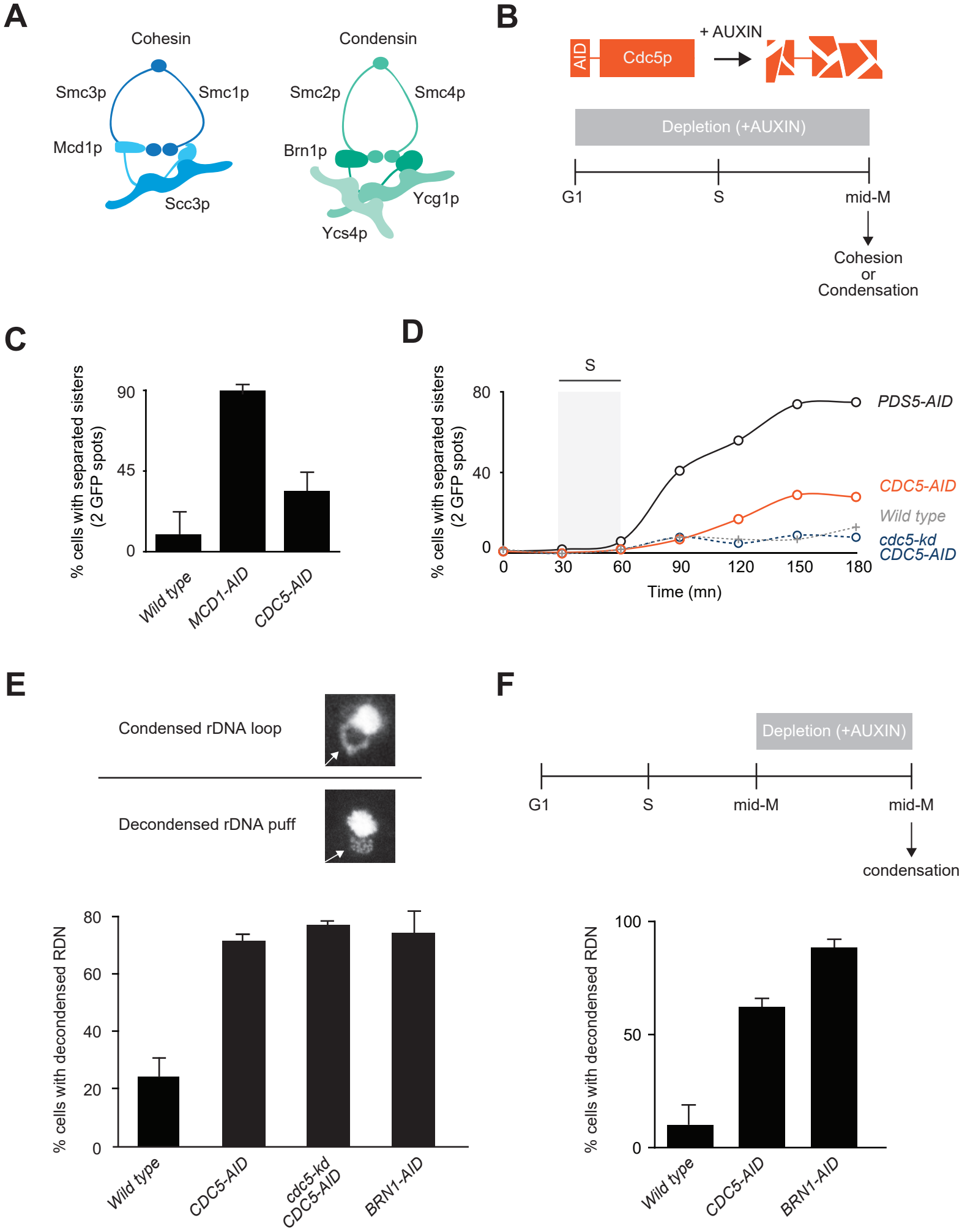

A
Cohesin
Smc3p
Smc1p
Mcd1p
Scc3p
Condensin
Smc2p
Smc4p
Brn1p
Ycg1p
Ycs4p
B
AID
Cdc5p
+ AUXIN
Depletion (+AUXIN)
G1
S
mid-M
Cohesion
or
Condensation
C
% cells with separated sisters
(2 GFP spots)
90
45
0
Wild type
MCD1-AID
CDC5-AID
D
S
80
40
0
% cells with separated sisters
(2 GFP spots)
PDS5-AID
CDC5-AID
Wild type
cdc5-kd
CDC5-AID
0
30
60
90
120
150
180
Time (mn)
E
Condensed rDNA loop
Decondensed rDNA puff
% cells with decondensed RDN
80
60
40
20
0
Wild type
CDC5-AID
cdc5-kd
CDC5-AID
BRN1-AID
F
Depletion (+AUXIN)
G1
S
mid-M
mid-M
condensation
% cells with decondensed RDN
100
50
0
Wild type
CDC5-AID
BRN1-AID

Figure 1: The polo kinase Cdc5p is essential for mid-M chromosome condensation

A. Schematic of cohesin and condensin. Cohesin is comprised of a Smc1p/Smc3p heterodimer joined at their head domain through Mcd1p. Mcd1p is likewise bound to Scc3p resulting in the cohesin holocomplex. Condensin is comprised of a Smc2p/Smc4p heterodimer linked by Brn1p. Brn1p is bound to Ycg1p and Ycs4p. Together, these proteins form the pentameric condensin complex.

B. *Top:* Schematic of the auxin-induced degron (AID) system to study functions of essential proteins. Proteins of interest, like Cdc5p, were fused to a 3V5-AID tag that allows for wild-type functions in the absence of auxin. Upon addition of auxin, 3V5-AID-tagged proteins were rapidly degraded in a proteasome-mediated manner.
Bottom: 'Staged mid-M' regimen used to assess strains function in mid-M. Cultures were synchronized in G1 and released in media containing auxin and nocodazole, to depleted AID-tagged protein and arrest cells in mid-M.

C. Depletion of Cdc5p results in a moderate cohesion defect. Cells were treated as in 1B and cohesion assayed in mid-M through a GFP spot assay. In these experiments, LacO arrays are integrated at LYS4, a centromere-distal arm site. Wild type, Mcd1p depleted, and Cdc5p depleted cells were scored for sister chromatid cohesion. The average percentage of separated sister chromatids from three independent experiments (100 cells) are reported, and error bars represent standard deviation.

D. Depletion of Cdc5p results in a moderate cohesion maintenance defect. Wild type, Pds5p depleted, Cdc5p depleted, and Cdc5 kinase-dead (kd) expressing in combination with depletion of Cdc5p-AID cells were treated as in B and cohesion assayed at 15mn intervals after release from G1 to mid-M. The average percentage of separated sister chromatids from two independent experiments (100 cells) are reported.

E. Depletion of Cdc5p results in condensation loss. *Top:* Example of rDNA locus (RDN) morphologies assayed. RDN is either assayed as a condensed loop or decondensed puff. The percentage of cells displaying decondensed RDN puffs is quantified.
Bottom: Wild type, Cdc5p depleted, Cdc5 kinase-dead (kd) expressing in combination with depletion of Cdc5p-AID, and Brn1p depleted cells were treated as in B and processed to make chromosome spreads to score RDN condensation (see Materials and Methods). Average of three experiments scoring 200 cells are shown and error bars represent the standard deviation.

F. Depletion of Cdc5p results in loss of maintenance of condensation . *Top:* Cells were arrested in mid-M using nocodazole. The arrest was confirmed by analysis of bud morphology (>95% large budded cells). Auxin was added to ensure Cdc5p-AID degradation. *Bottom:* Wild type, Cdc5p depleted, and Brn1p depleted cells were processed to assess condensation as described in E.

Figure 2

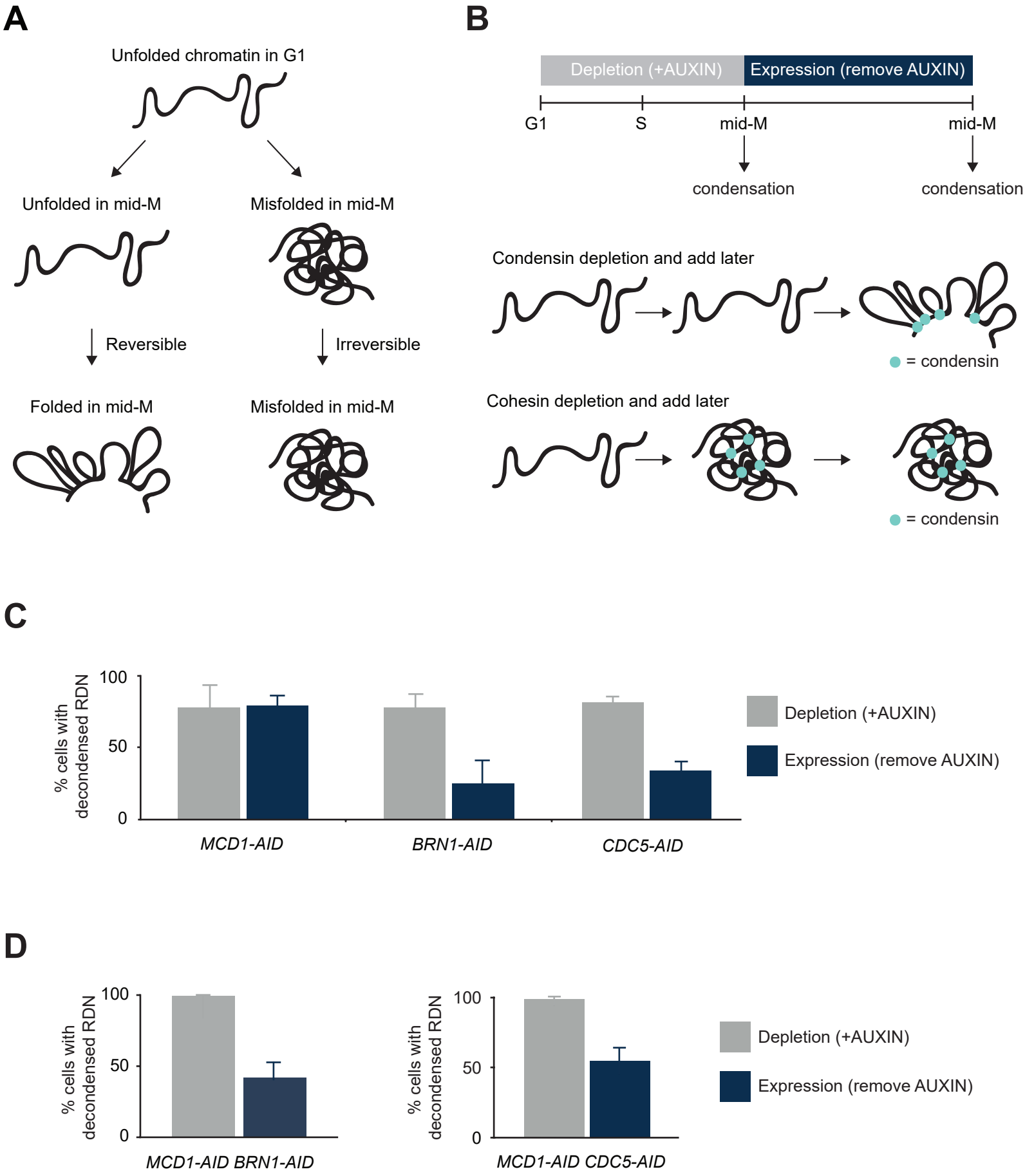

A
Unfolded chromatin in G1
Unfolded in mid-M
Misfolded in mid-M
Reversible
Irreversible
Folded in mid-M
Misfolded in mid-M
B
Depletion (+AUXIN)
Expression (remove AUXIN)
G1
S
mid-M
mid-M
condensation
condensation
Condensin depletion and add later
= condensin
Cohesin depletion and add later
= condensin
C
% cells with decondensed RDN
100
50
0
MCD1-AID
BRN1-AID
CDC5-AID
Depletion (+AUXIN)
Expression (remove AUXIN)
D
% cells with decondensed RDN
100
50
0
MCD1-AID BRN1-AID
% cells with decondensed RDN
100
50
0
MCD1-AID CDC5-AID
Depletion (+AUXIN)
Expression (remove AUXIN)

Figure 2: Cdc5p stimulates condensin-dependent RDN condensation

A. Schematic of defects in condensation. RDN is in a puffed unfolded state in G1. Depletion of condensation factors results in a puffed RDN that can either be in an unfolded state or in a misfolded state. The unfolded state is reversible, once the factor is expressed again the RDN can be folded. Misfolded state is irreversible, even after expressing again the condensation factor the RDN remains misfolded.

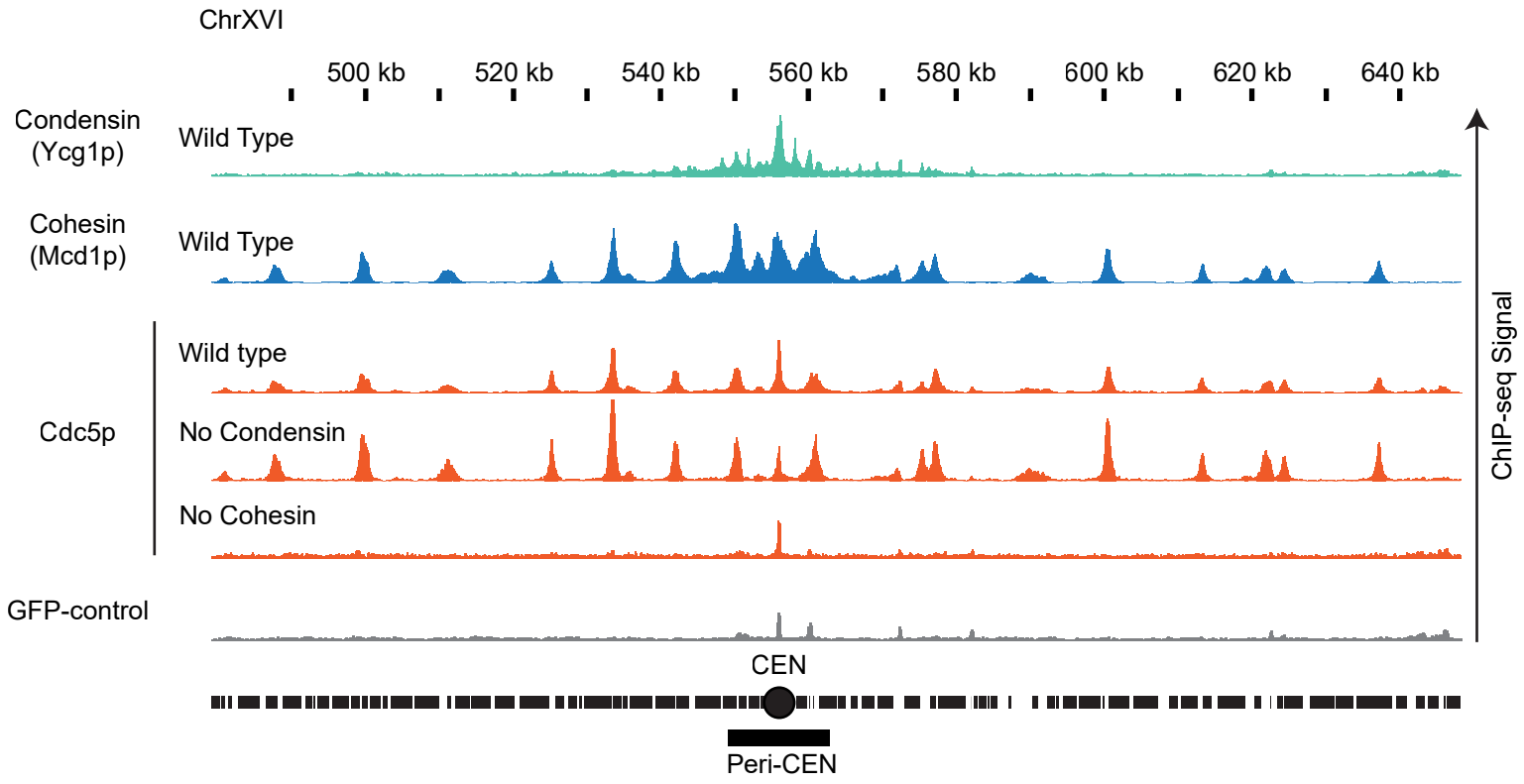
B. Schematic of 'add later' genetic test setup. *Top:* Indicated cultures were arrested in G1 (>95% schmooed morphology), and AID-tagged proteins (Mcd1p-AID, Brn1p-AID, Cdc5p-AID) were depleted by the addition of auxin. G1 arrest was relieved by resuspending in media with auxin and nocodazole to arrest cells in mid-M. Cells were then resuspended in media containing only nocodazole for two hours to allow the expression of the previously depleted AID-tagged proteins.
Bottom: Depletion of condensin resulted in unfolded RDN that was folded upon later expression of condensin. Depletion of cohesin resulted in misfolded RDN that stayed misfolded upon later expression of cohesin.

C. Cdc5p depletion results in a reversible unfolded RDN state. **'**Add later' genetic test results from *MCD1-AID, BRN1-AID*, and *CDC5-AID* strains were treated as described in B and scored for RDN condensation. The percentage of cells displaying puffed RDN (decondensed) is quantified. Average of two experiments scoring 300 cells are shown and error bars represent standard deviation.

D. Condensin and Cdc5p promote misfolding of the RDN in the absence of cohesin. 'Add-later' genetic test results from MCD1-AID/BRN1-AID and MCD1-AID/CDC5-AID strains were treated as described in B and scored for RDN condensation. The percentage of cells displaying decondensed puffed RDN (decondensed) is quantified. Average of three experiments scoring 300 cells are shown and error bars represent standard deviation.

Figure 3

A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540 kb
560 kb
580 kb
600 kb
620 kb
640 kb
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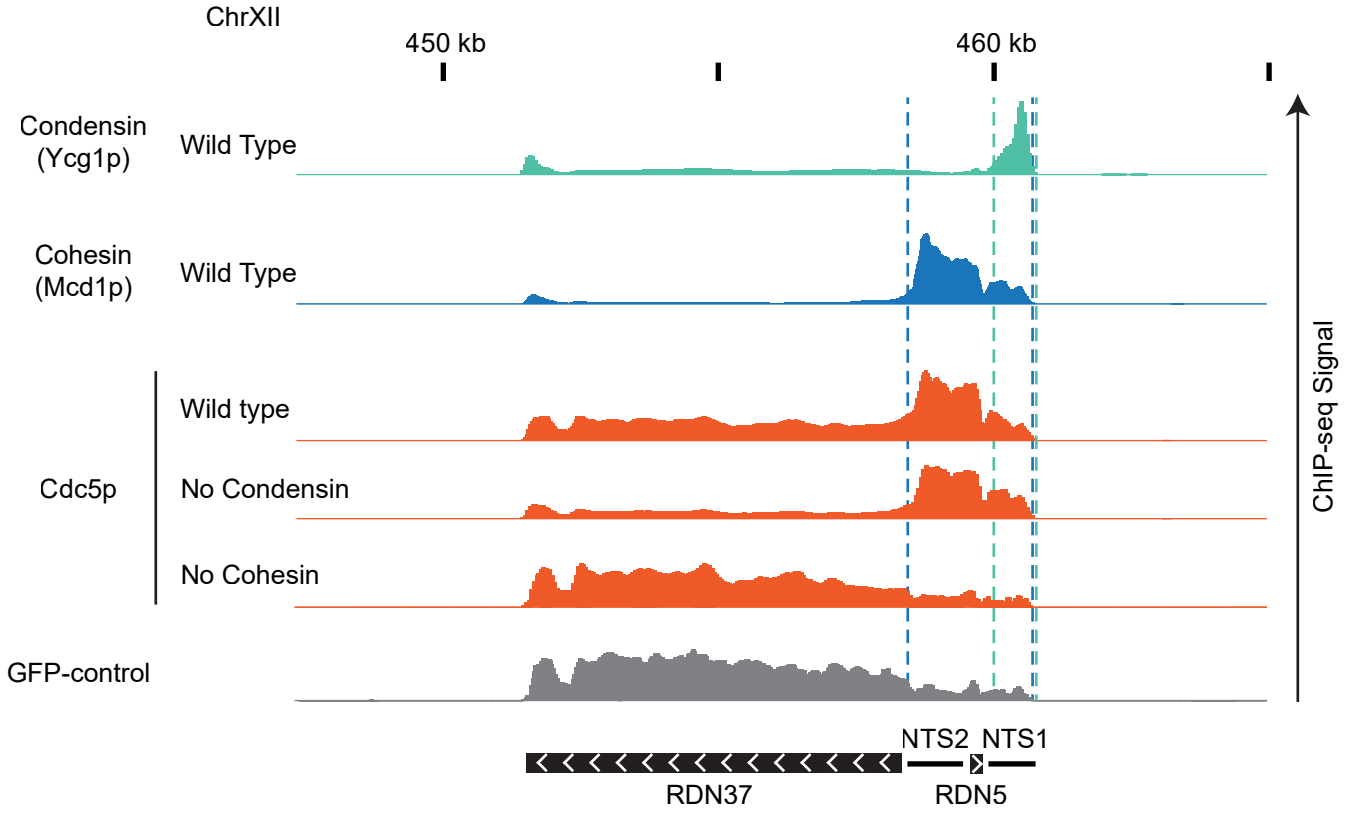
Wild Type
Cohesin (Mcd1p)
Wild Type
Cdc5p
Wild type
No Condensin
No Cohesin
GFP-control
ChIP-seq Signal
CEN
Peri-CEN

B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Wild Type
Cohesin (Mcd1p)
Wild Type
Cdc5p
Wild type
No Condensin
No Cohesin
GFP-control
ChIP-seq Signal
NTS2
NTS1
RDN37
RDN5

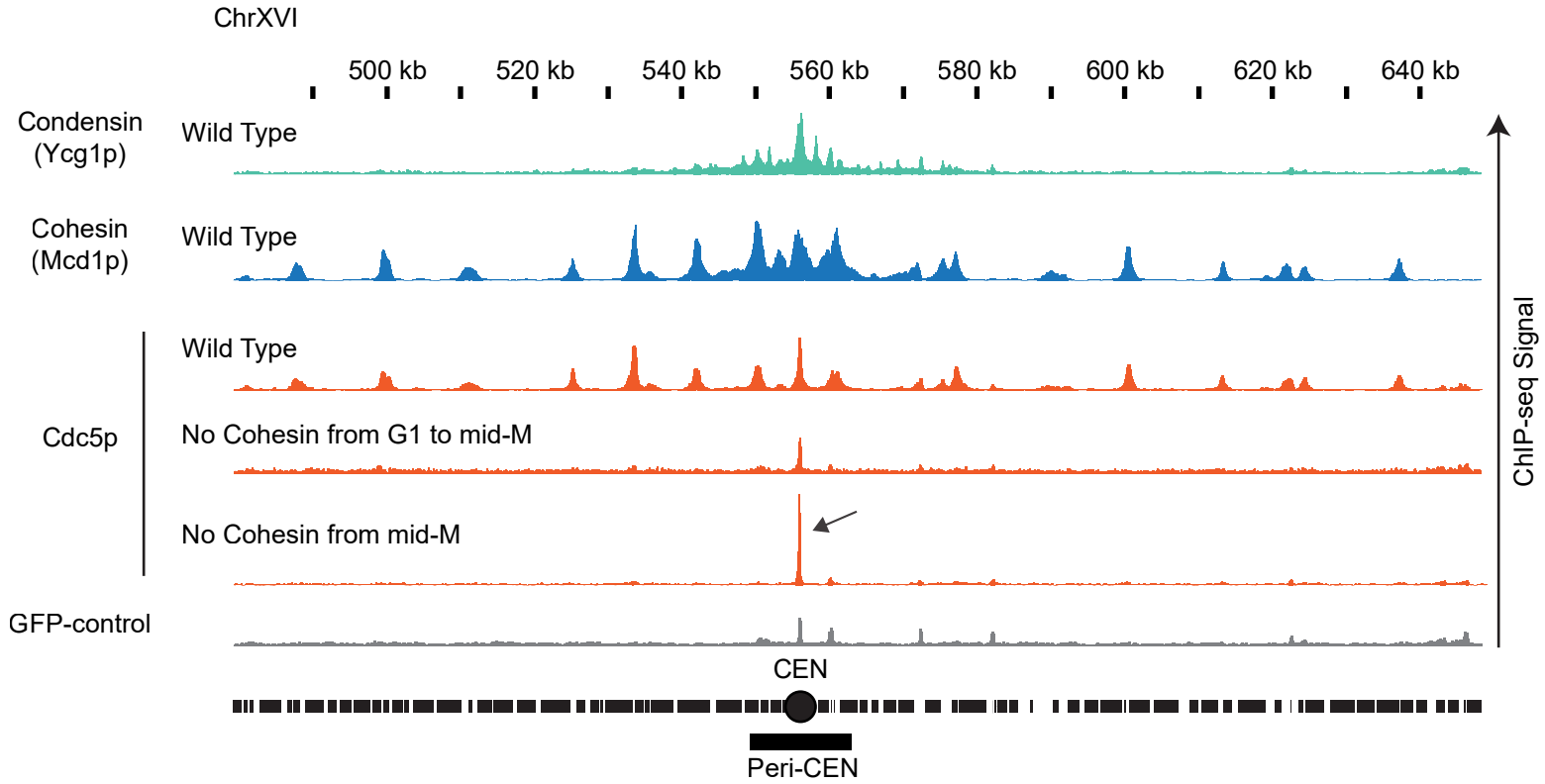
Figure 3: Genome-wide localization of condensin, cohesin and Cdc5p.

A. Cohesin recruits Cdc5p at unique sequences genome-wide. ChIP-seq signals from a representative portion of chromosome XVI are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-Flag is in orange, and signal coming from GFP-control is in grey. Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of condensin subunit Brn1p-AID is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID is in the third orange lane. All strains were depleted of the indicated AID-protein from G1 to mid-M and arrested in mid-M (staged mid-M).

B. Cohesin recruits Cdc5p at the repetitive RDN locus. ChIP-seq signals from a portion of chromosome XII are presented with bar charts. ChIP-seq signal from strains as in (A). The region presented includes one copy of the rDNA repeat (9.1kb) made of the transcribed regions RND37 and RND5 (black box with white arrows indicating the direction of transcription) and the non-transcribed regions NTS1 and NTS2 (black line). Region of enrichment of cohesin signal is marked by two blue dashed lines (from NTS2 to NTS1). Region of enrichment of condensin signal is marked by two green dashed lines (end of NTS1).

Figure 4

A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Condensin (Ycg1p)
Wild Type
Cohesin (Mcd1p)
Wild Type
Cdc5p
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No Cohesin from mid-M
GFP-control
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CEN
Peri-CEN

B

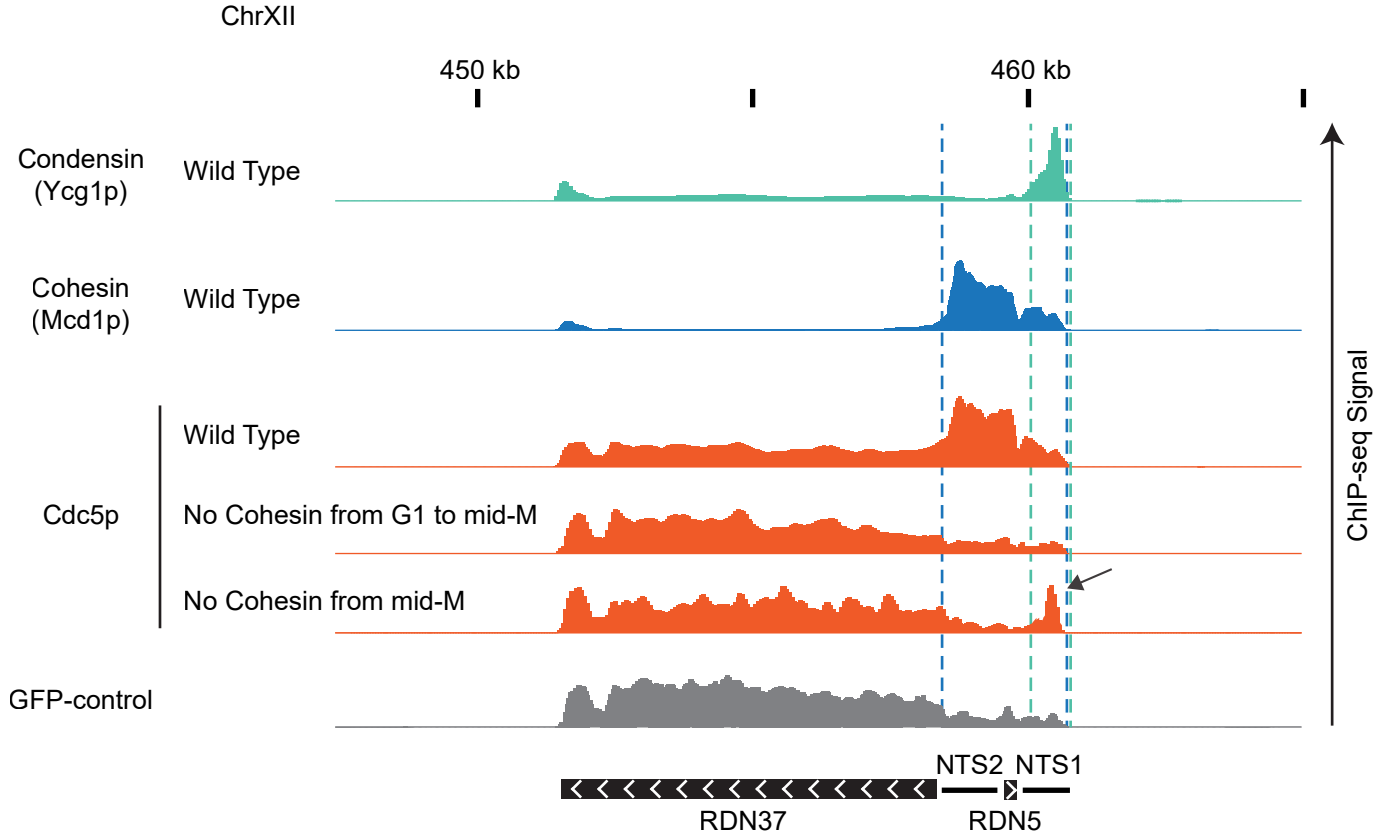
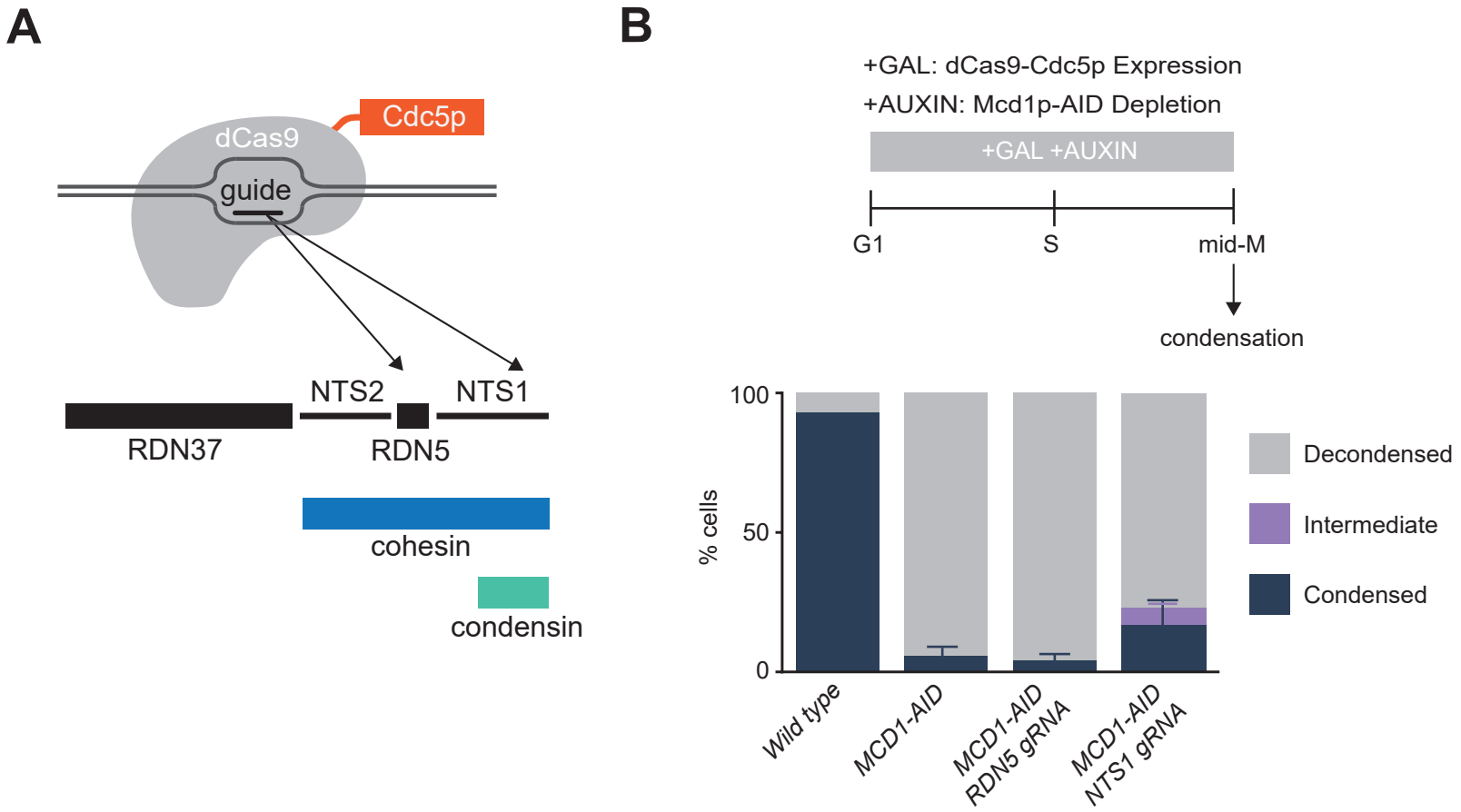

ChrXII
450 kb
460 kb
Condensin (Ycg1p)
Wild Type
Cohesin (Mcd1p)
Wild Type
Cdc5p
Wild Type
No Cohesin from G1 to mid-M
No Cohesin from mid-M
GFP-control
ChIP-seq Signal
NTS2
NTS1
RDN37
RDN5

Figure 4: Cdc5p is recruited by cohesin and brought to condensin at centromeres and rDNA.

A. Cdc5p is recruited by cohesin and brought to condensin at centromeres. ChIP-seq signals from a representative portion of chromosome XVI are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-V5 is in orange, and signal coming from GFP-control is in grey. Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of cohesin subunit Mcd1p-AID from G1 to mid-M is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID in mid-M is in the third orange lane. Black arrow indicates enrichment of Cdc5p at the centromere when cohesin is transiently associated with chromosomes.

B. Cdc5p is recruited by cohesin and brought to condensin at RDN. ChIP-seq signals from a portion of chromosome XII are presented with bar charts. ChIP-seq signal from strains as in (A). The region presented includes one copy of the rDNA repeat (9.1kb) made of the transcribed regions RND37 and RND5 (black box with white arrows indicating the direction of transcription) and the non-transcribed regions NTS1 and NTS2 (black line). Region of enrichment of cohesin signal is marked by two blue dashed lines (from NTS2 to NTS1). Region of enrichment of condensin signal is marked by two green dashed lines (end of NTS1). Black arrow indicates enrichment of Cdc5p at the condensin-bound region of the RDN when cohesin is transiently associated with chromosomes.

Figure 5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Cdc5p
dCas9
guide
NTS2
NTS1
RDN37
RDN5
cohesin
condensin
B
+GAL: dCas9-Cdc5p Expression
+AUXIN: Mcd1p-AID Depletion
+GAL +AUXIN
G1
S
mid-M
condensation
100
50
0
% cells
Decondensed
Intermediate
Condensed
Wild type
MCD1-AID
MCD1-AID RDN5 gRNA
MCD1-AID NTS1 gRNA

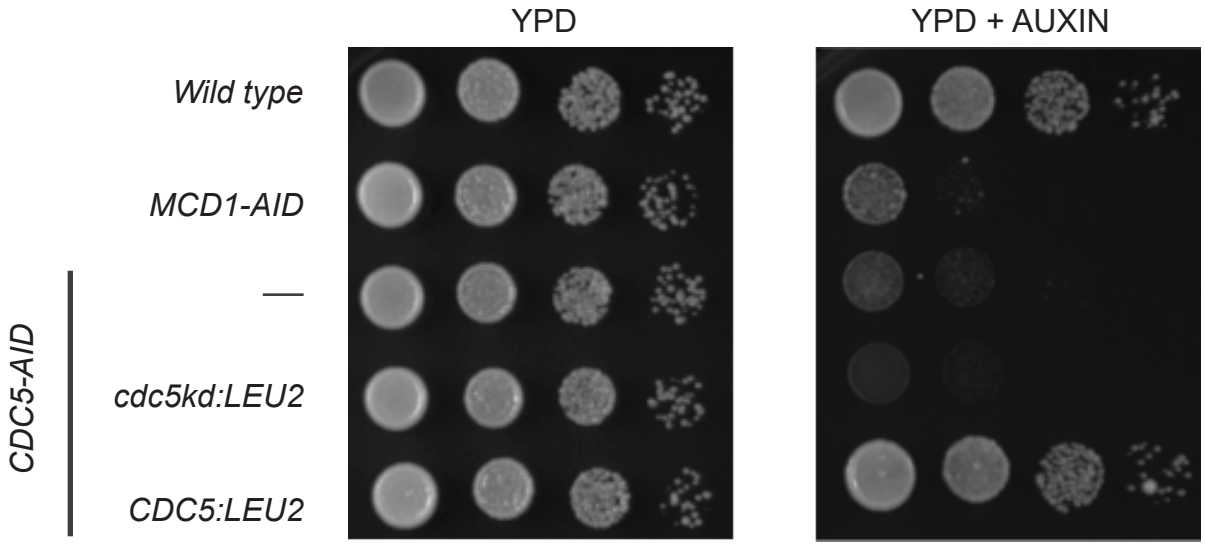
Figure 5: Localization of Cdc5p with CRISPR-Cas9 to the condensin-bound RDN partially restores the RDN condensation defect of cohesin-depleted cells.

A. Galactose addition to the media drives the expression of a catalytically dead dCas9 fused to Cdc5p. Guide RNAs recruit dCas9-Cdc5p fusion protein either to the condensin-bound region NTS1 (RDN5 gRNA) or 500 bp to the left at RDN5 (RDN5 gRNA) where only cohesin is bound.

B. *Top:* 'Staged mid-M' regimen used to assess condensation in mid-M. Cultures were synchronized in G1 and released in media containing galactose and auxin to drive the expression of dCas9-Cdc5p concomitantly to the depletion of Mcd1p-AID. Media also contained nocodazole to arrest cells in mid-M and assess condensation.
Bottom: Wild type, Mcd1p depleted, Mcd1p depleted with dCas9-Cdc5p and RDN5 gRNA expressing, and Mcd1p depleted with dCas9-Cdc5p and NTS1 gRNA expressing cells were treated with 'staged mid-M' regimen and processed to score RDN condensation. Average of three experiments scoring 100 cells are shown and error bars represent the standard deviation.

Figure 1—figure supplement 1

A



B

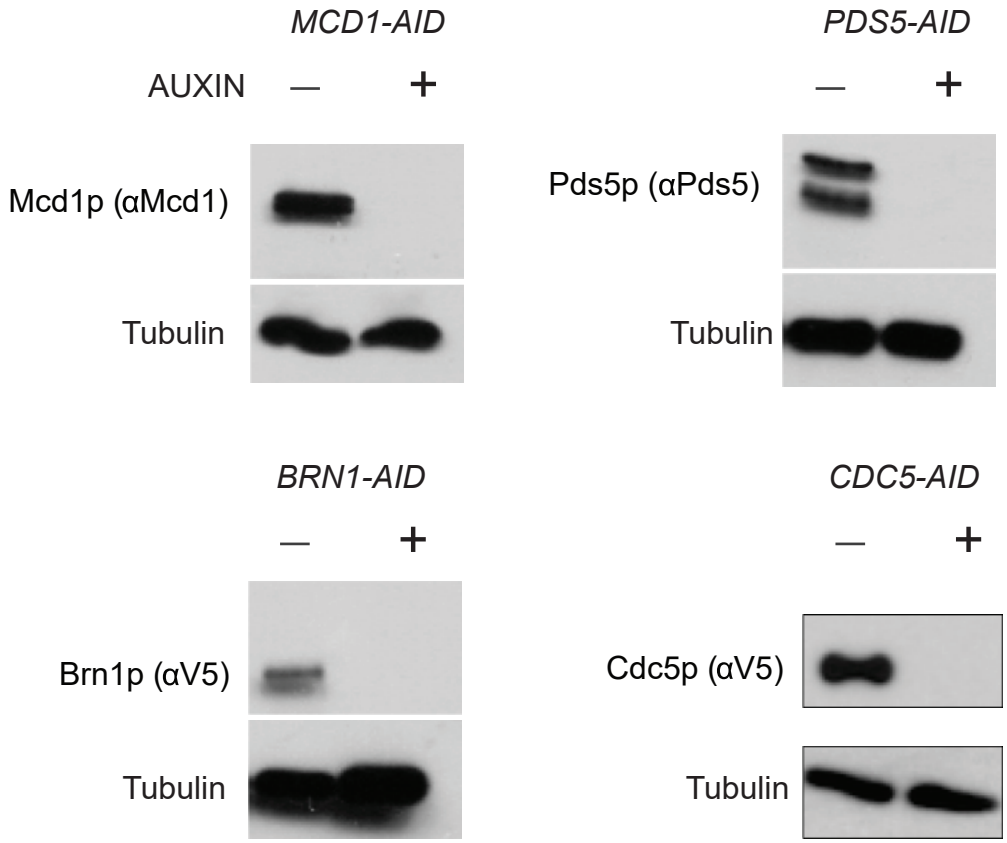


Figure 1—figure supplement 1: Characterization of AID-tagged strains.

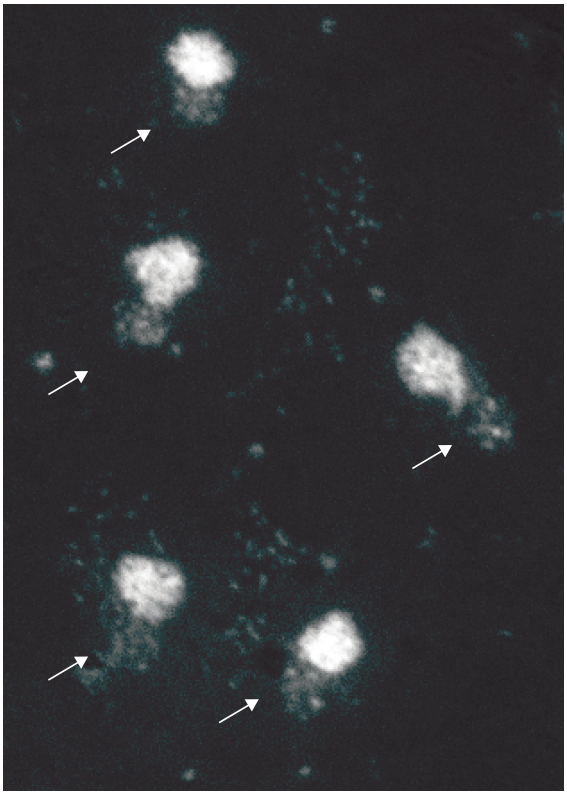
A. Cdc5p depletion using the AID degron system reduces viability. Asynchronous cells were serially diluted and plated on YPD (left) or YPD with 750uM auxin (right). Plates were then incubated at 23C for three days and imaged. Wild Type, Mcd1p-AID, Cdc5p-AID, Cdc5-AID with expression of Cdc5p kinase-dead from the LEU2 locus, and Cdc5p-AID with expression of Cdc5p from the LEU2 locus cells were plated.

B. Depletion of AID-tagged proteins assayed by Western blotting. *CDC5-AID, BRN1-AID, MCD1-AID, PDS5-AID,* strains were grown to mid-log in YPD. Cells were then split and grown either in the presence or absence of auxin for 1h. Cells were then pelleted and protein extracted by TCA as described in the materials and methods. Western blot analysis used antibodies for the endogenous Mcd1p and Pds5p, anti V5 for Brn1p-3V5 and Cdc5p-3V5. Tubulin antibody was used for loading control

Figure 1—figure supplement 2

A

Decondensed RDN puffs
after depletion of Cdc5p



B

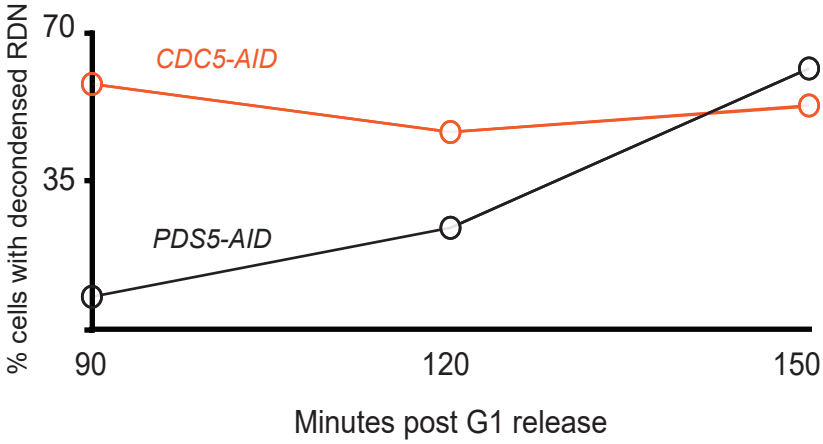


Figure 1—figure supplement 2: Cdc5p depletion results in loss of RDN condensation

A. Loss of rDNA condensation after Cdc5p-AID depletion. Representative image showing chromosome spreads of Cdc5p-AID cells after auxin depletion. The puffed RDN next to the bulk of the chromosomes indicates loss of condensation in the RDN.

B. Cdc5p is required to establish condensation of the RDN. Pds5p depleted and Cdc5p depleted, cells were treated with ‘‘Staged mid-M’ regimen used to assess strains function in mid-M. Cultures were synchronized in G1 and released in media containing auxin and nocodazole, to depleted AID-tagged protein and arrest cells in mid-M. RDN condensation assayed at 30mn intervals after release from G1 to mid-M. 90 minutes is the earliest time point were RDN condensation is clearly measurable. The average percentage of cells with decondensed RDN (100 cells) are reported.

Figure 1—figure supplement 3

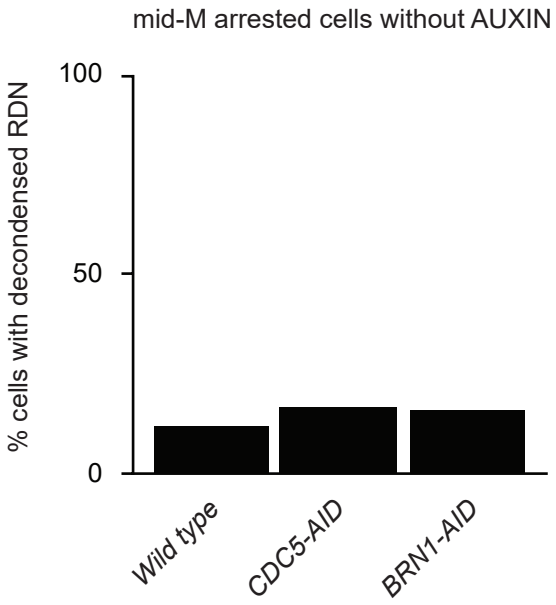


Figure 1—figure supplement 3: Addition of the AID-tag to Cdc5p and Brn1 does not affect RDN condensation in the absence of auxin

rDNA condensation levels of AID-tagged strains are comparable to wild type without auxin. Wild type, Cdc5p depleted, and Brn1p depleted cells were treated as in B omitting auxin addition and processed to make chromosome spreads to score RDN condensation. 200 cells were scored.

Figure 2—figure supplement 1

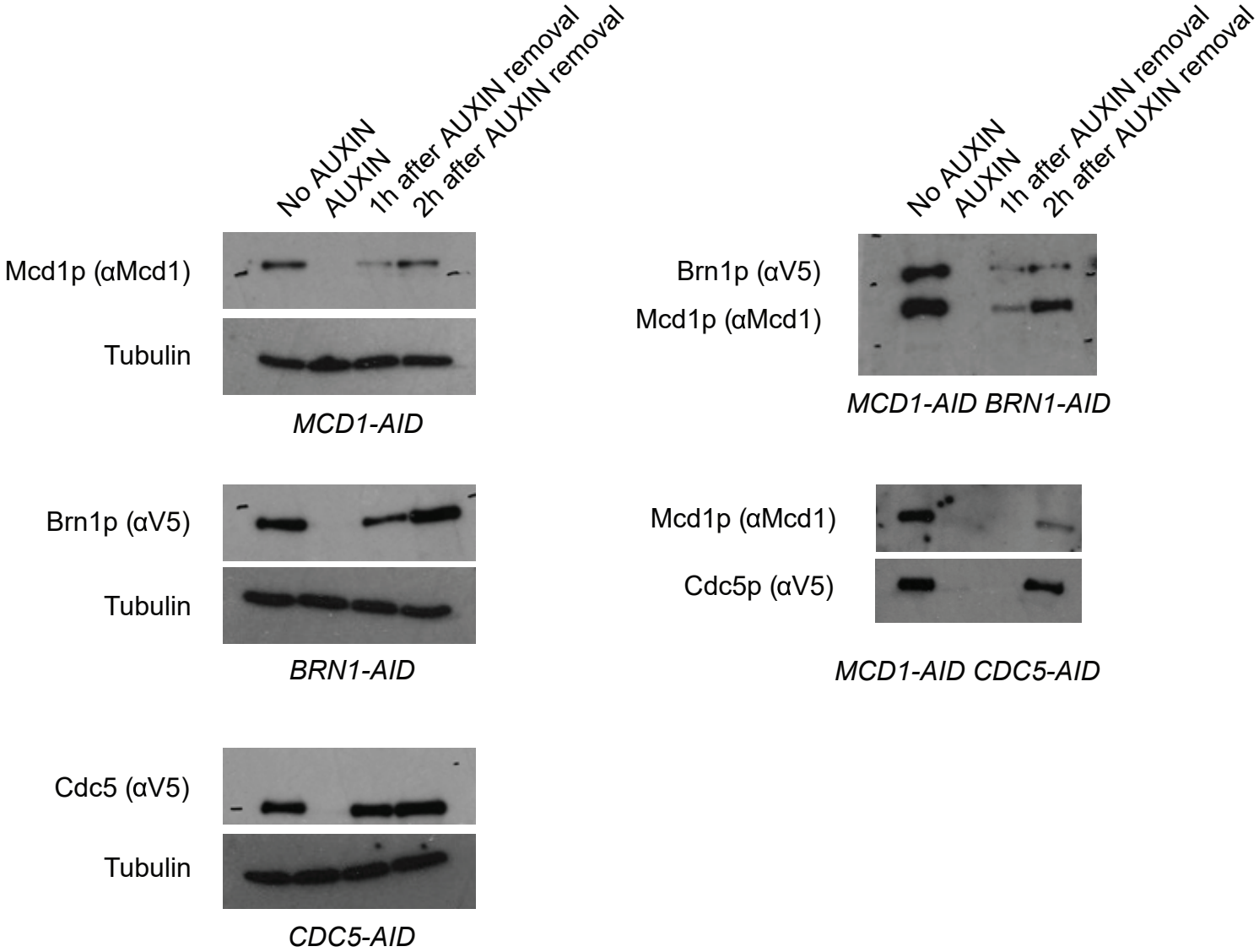


Figure 2—figure supplement 1: AID-tagged protein levels analyzed after depletion and re-expression in the 'add later' experiment.

Depletion followed by expression of AID-tagged proteins assayed by Western blotting. Strains were subjected to the 'Add later' regimen. Indicated cultures were arrested in G1 (>95% schmooed morphology), and AID-tagged proteins (Mcd1p-AID, Brn1p-AID, Cdc5p-AID, MCD1-AID/BRN1-AID, and MCD1-AID/CDC5-AID) were depleted by the addition of auxin. G1 arrest was relieved by resuspending in media with auxin and nocodazole to arrest cells in mid-M. Cells were then resuspended in media containing only nocodazole for two hours to allow the expression of the previously depleted AID-tagged proteins. Cells were then pelleted and protein extracted by TCA as described in the materials and methods. Western blot analysis used antibodies for the endogenous Mcd1p, anti V5 for Brn1p-3V5 and Cdc5p-3V5. Tubulin antibody was used for loading control.

Figure 3—figure supplement 1

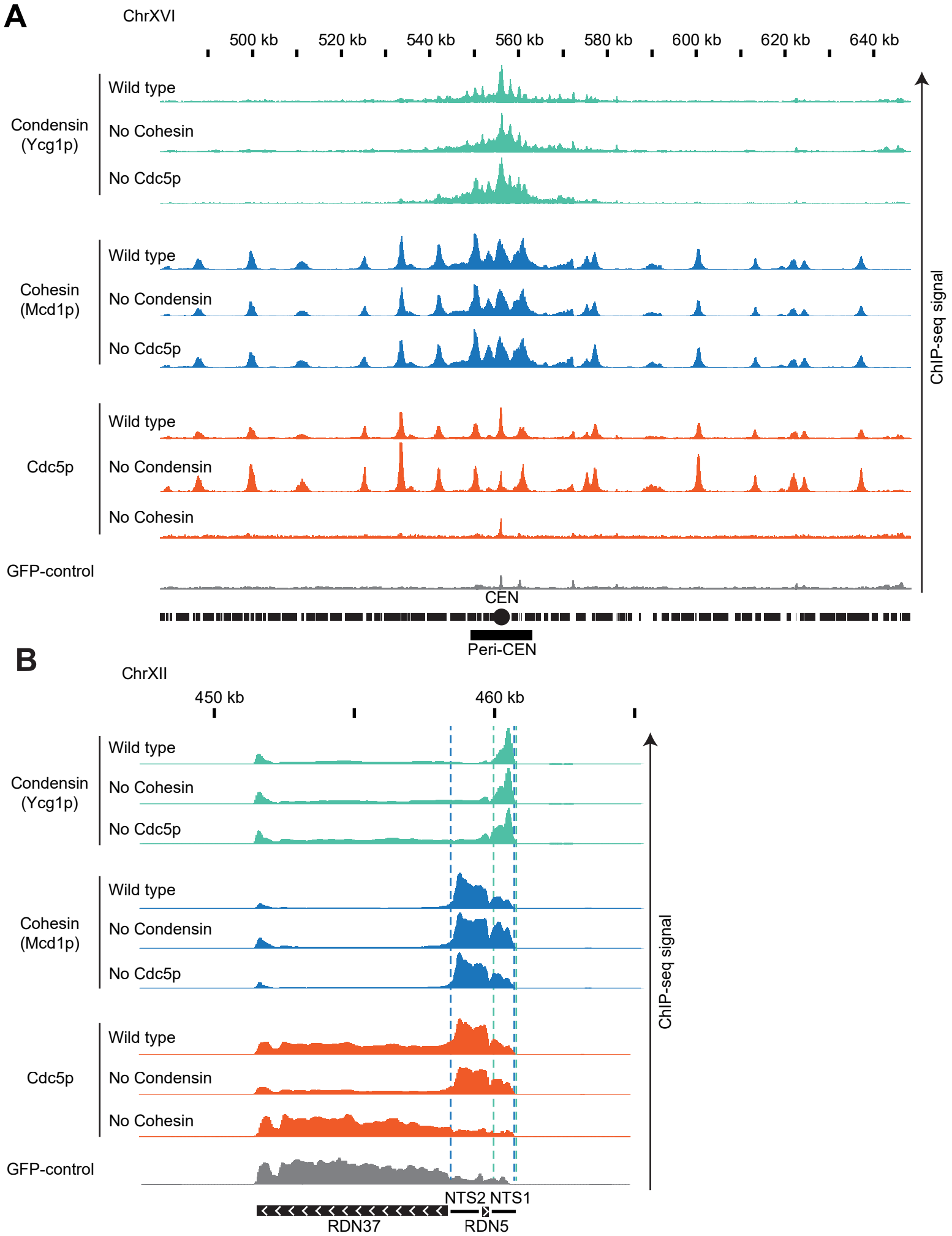


Figure 3—figure supplement 1: Cohesin and condensin bind to specific sites on chromosomes independently of each other and Cdc5p.

A. Cohesin, condensin and Cdc5p localization at unique sequences genome-wide. ChIP-seq signals from a representative portion of chromosome XVI are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-Flag is in orange, and signal coming from GFP-control is in grey. Ycg1p-Myc signal from wild type is in the first green lane, from cells depleted of cohesin subunit Mcd1p-AID is in the second green lane, from cells depleted of Cdc5p-AID is in the third green lane. Mcd1p signal from wild type is in the first blue lane, from cells depleted of condensin subunit Brn1p-AID is in the second blue lane, from cells depleted of Cdc5p-AID is in the third blue lane.Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of condensin subunit Brn1p-AID is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID is in the third orange lane. All strains were depleted of the indicated AID-protein from G1 to mid-M and arrested in mid-M (staged mid-M).

B. Cohesin, condensin and Cdc5p localization at the repetitive RDN locus. ChIP-seq signals from a portion of chromosome XII are presented with bar charts. ChIP-seq signal from strains as in (A). The region presented includes one copy of the rDNA repeat (9.1kb) made of the transcribed regions RND37 and RND5 (black box with white arrows indicating the direction of transcription) and the non-transcribed regions NTS1 and NTS2 (black line). Region of enrichment of cohesin signal is marked by two blue dashed lines (from NTS2 to NTS1). Region of enrichment of condensin signal is marked by two green dashed lines (end of NTS1).

Figure 3—figure supplement 2

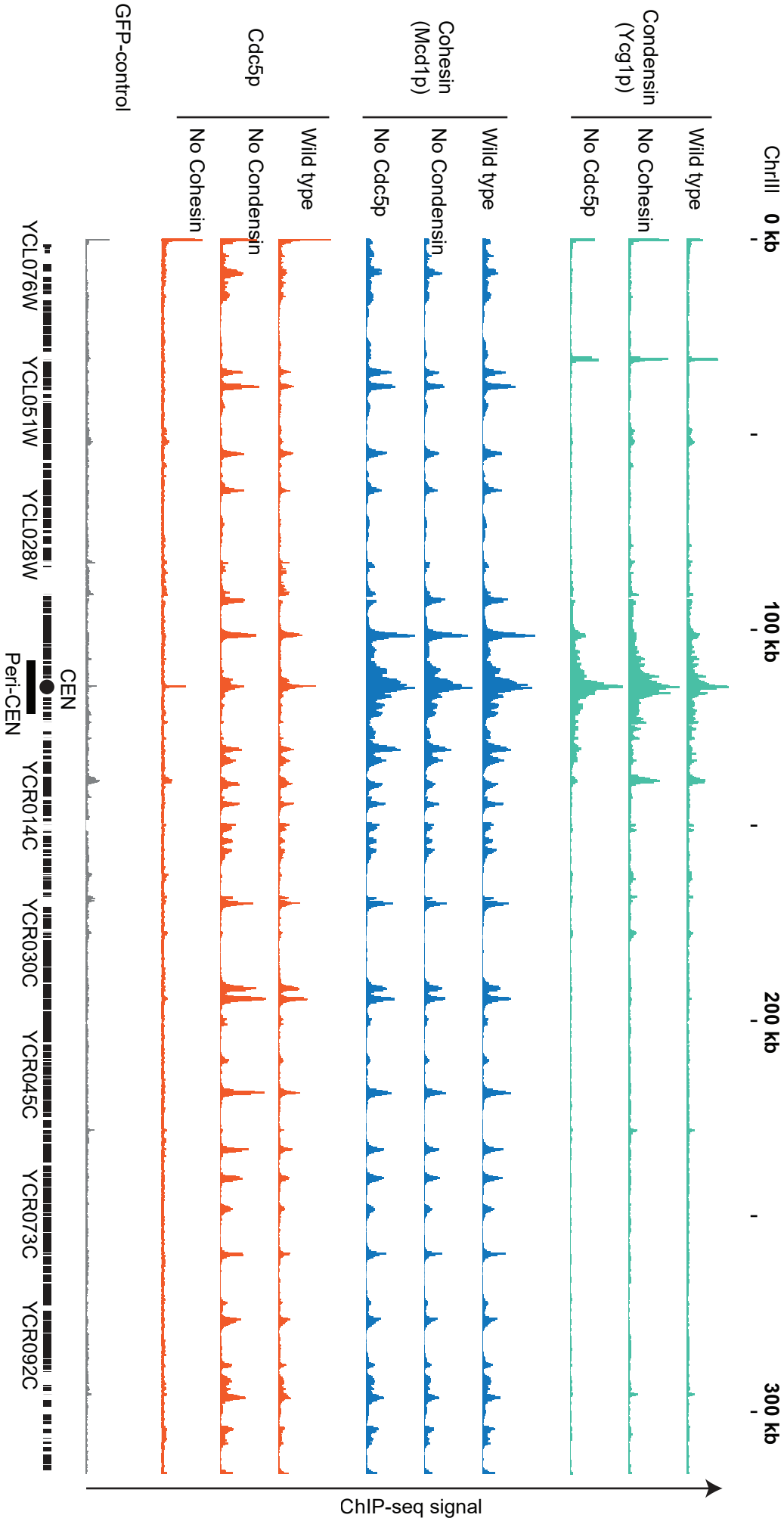


Figure 3—figure supplement 2: Cohesin, condensin, and Cdc5p bind to specific sites on chromosome III.

Cohesin, condensin and Cdc5p localization at unique sequences genome-wide. ChIP-seq signals from a representative portion of chromosome III are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-Flag is in orange, and signal coming from GFP-control is in grey. Ycg1p-Myc signal from wild type is in the first green lane, from cells depleted of cohesin subunit Mcd1p-AID is in the second green lane, from cells depleted of Cdc5p-AID is in the third green lane. Mcd1p signal from wild type is in the first blue lane, from cells depleted of condensin subunit Brn1p-AID is in the second blue lane, from cells depleted of Cdc5p-AID is in the third blue lane.Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of condensin subunit Brn1p-AID is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID is in the third orange lane. All strains were depleted of the indicated AID-protein from G1 to mid-M and arrested in mid-M (staged mid-M).

Figure 3—figure supplement 3

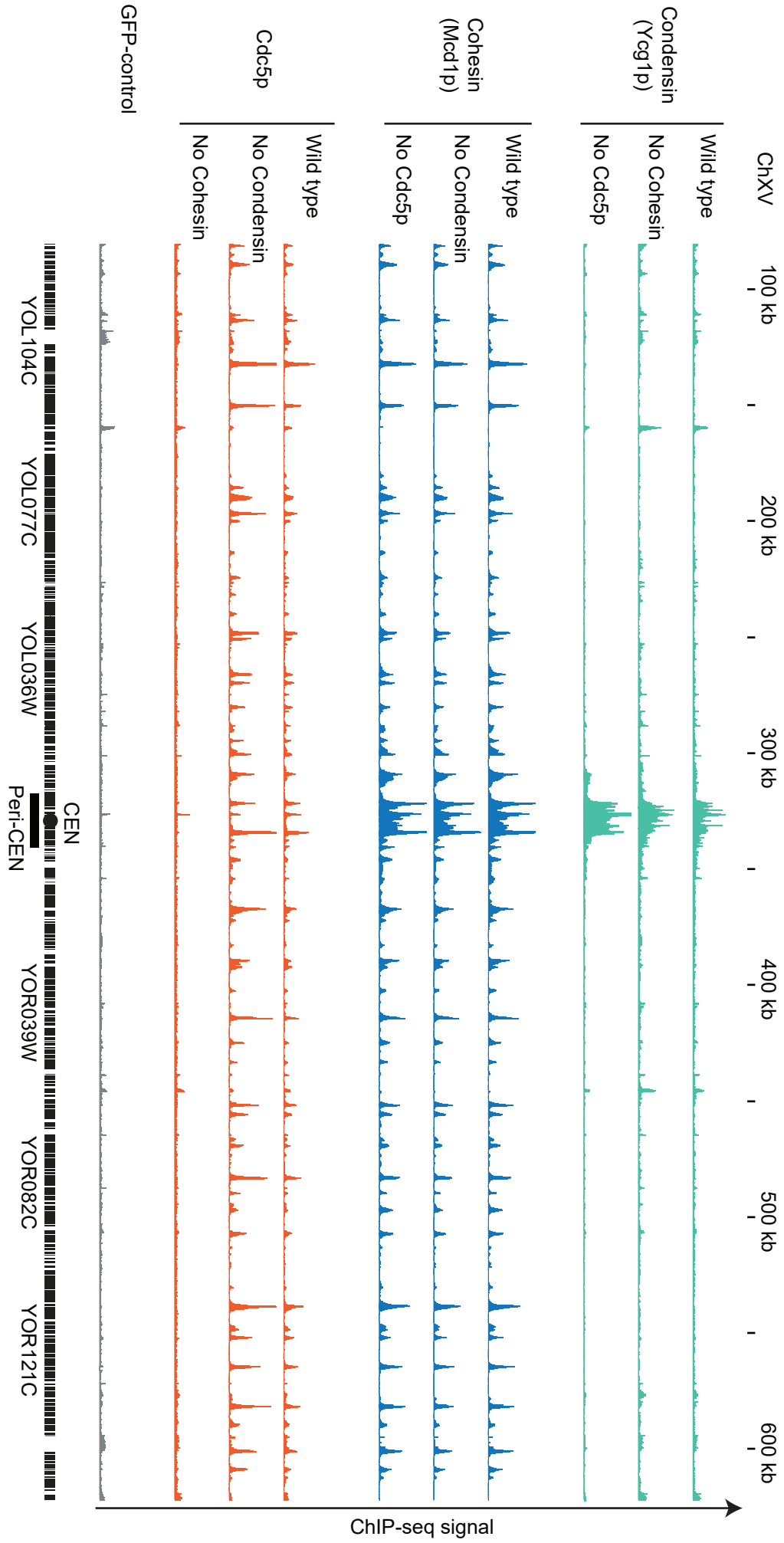


Figure 3—figure supplement 3: Cohesin, condensin, and Cdc5p bind to specific sites on chromosome XV.

Cohesin, condensin and Cdc5p localization at unique sequences genome-wide. ChIP-seq signals from a representative portion of chromosome XV are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-Flag is in orange, and signal coming from GFP-control is in grey. Ycg1p-Myc signal from wild type is in the first green lane, from cells depleted of cohesin subunit Mcd1p-AID is in the second green lane, from cells depleted of Cdc5p-AID is in the third green lane. Mcd1p signal from wild type is in the first blue lane, from cells depleted of condensin subunit Brn1p-AID is in the second blue lane, from cells depleted of Cdc5p-AID is in the third blue lane.Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of condensin subunit Brn1p-AID is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID is in the third orange lane. All strains were depleted of the indicated AID-protein from G1 to mid-M and arrested in mid-M (staged mid-M).

Figure 3—figure supplement 4

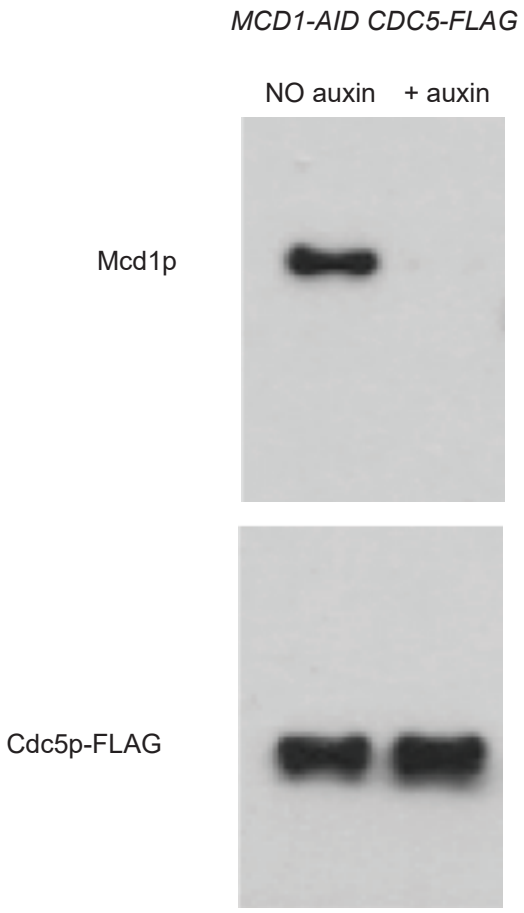


Figure 3—figure supplement 4: Cdc5p levels after Mcd1p depletion analyzed by western blotting.

Depletion of Mcd1p-AID does not affect levels of Cdc5p. Indicated cultures were arrested in G1 (>95% schmooed morphology), and Mcd1p-AID was depleted by the addition of auxin. G1 arrest was relieved by resuspending in media with auxin and nocodazole to arrest cells in mid-M. Cells were then pelleted and protein extracted by TCA as described in the materials and methods. Western blot analysis used antibodies for the endogenous Mcd1p, anti FLAG for Cdc5p-FLAG.

Figure 4—figure supplement 1

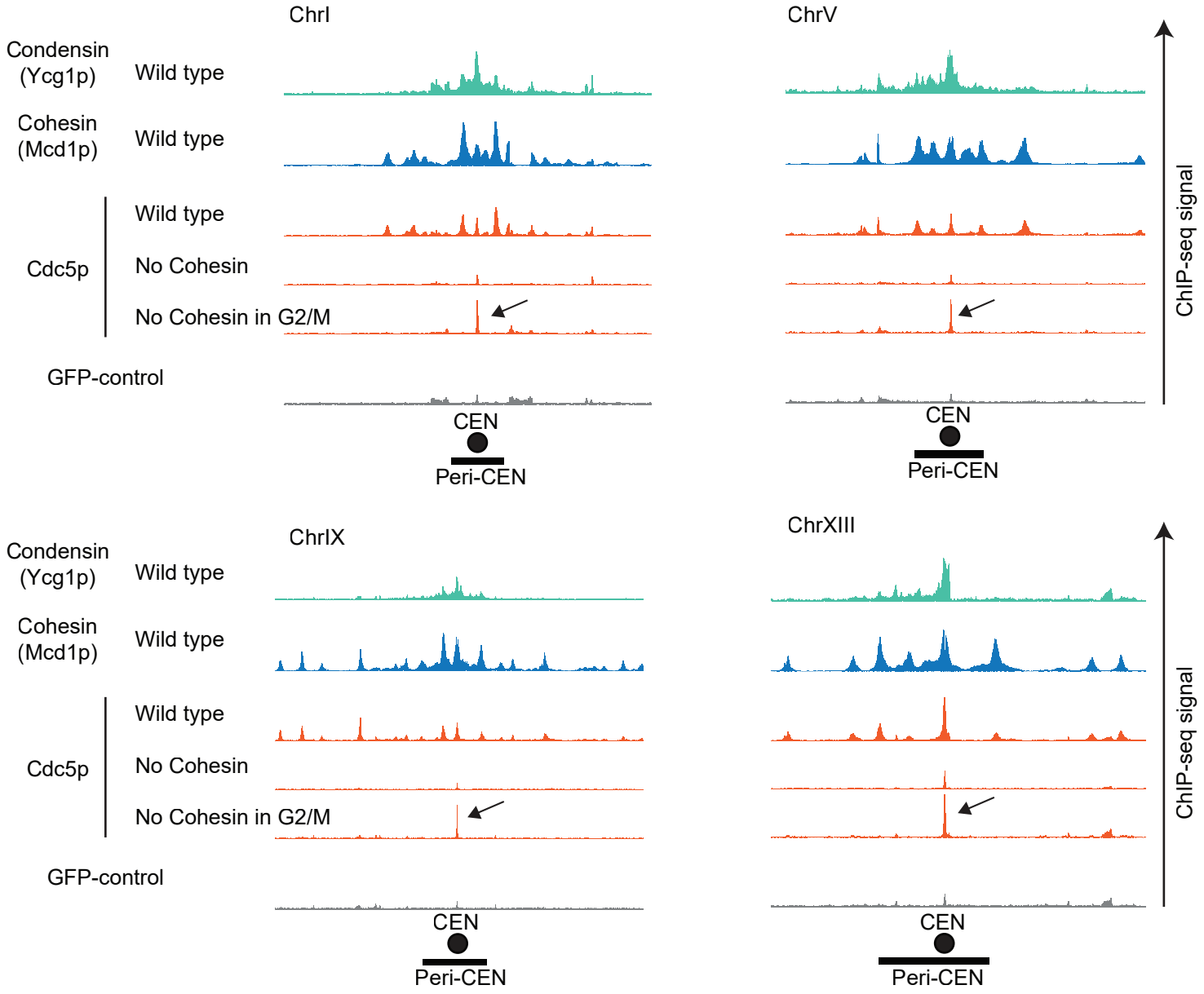


Figure 4—figure supplement 1: Cdc5p binding to centromeres is revealed by a transient association of Mcd1p with chromosomes.

Cdc5p is recruited by cohesin and brought to condensin at centromeres. ChIP-seq signals from a representative portion of chromosomes are presented with bar charts. The region presented includes the centromere (CEN black circle) and the pericentric region (peri-CEN black box). Signal coming from condensin subunit Ycg1p-Myc in wild type strain is in green, signal from cohesin subunit Mcd1p in wild type strain is in blue, signal coming from Cdc5p-V5 is in orange, and signal coming from GFP-control is in grey. Cdc5p ChIP-seq signal from wild type is in the first orange lane, from cells depleted of cohesin subunit Mcd1p-AID from G1 to mid-M is in the second orange lane, from cells depleted of cohesin subunit Mcd1p-AID in mid-M is in the third orange lane. Black arrow indicates enrichment of Cdc5p at the centromere when cohesin is transiently associated with chromosomes.

Figure 4—figure supplement 2

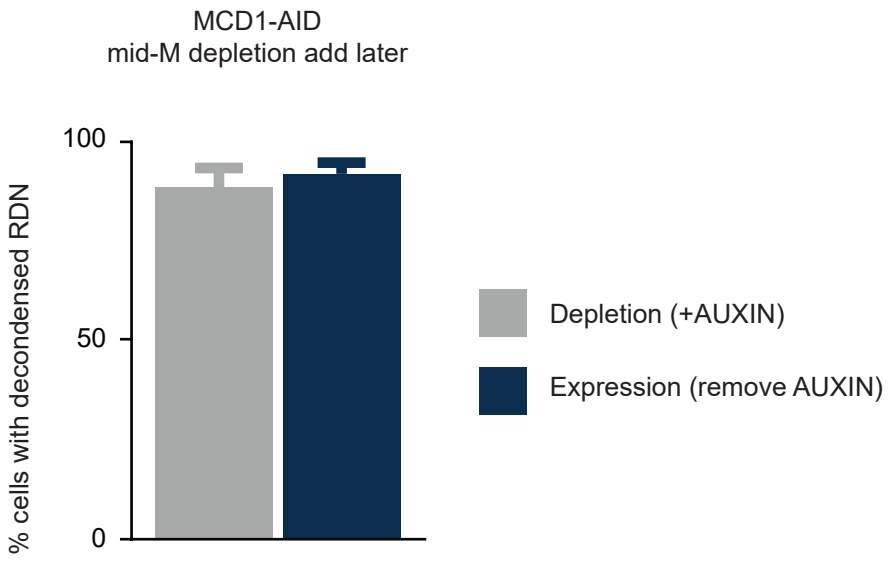
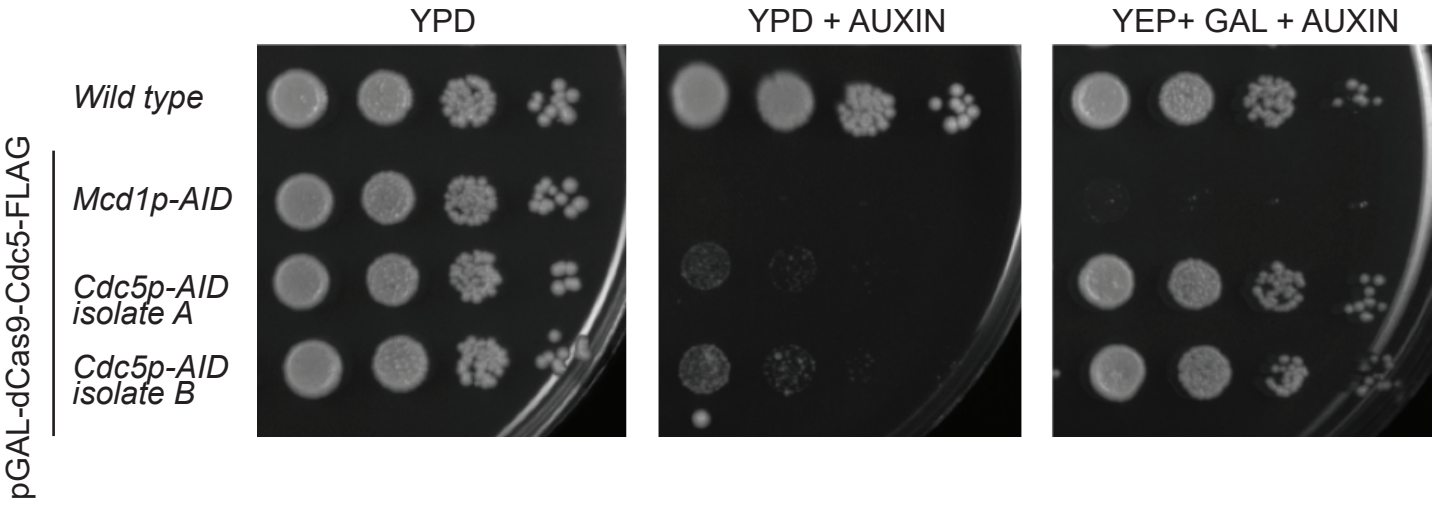


Figure 4—figure supplement 2: Mcd1p transient association with chromosomes does not restore condensation of the RDN in Mcd1p depleted cells.

Depletion of Cdc5p results in loss of maintenance of condensation. Mcd1p-AID cells were arrested in mid-M using nocodazole. The arrest was confirmed by analysis of bud morphology (>95% large budded cells). Auxin was added to ensure Mcd1p-AID degradation for 90min (depletion grey bar). Mcd1p-AID was then expressed by washing out auxin (expression blue bar). Cells were processed to make chromosome spreads to score RDN condensation. Two independent experiments with 200 cells were scored and error bars indicate the standard deviation.

Figure 5—figure supplement 1



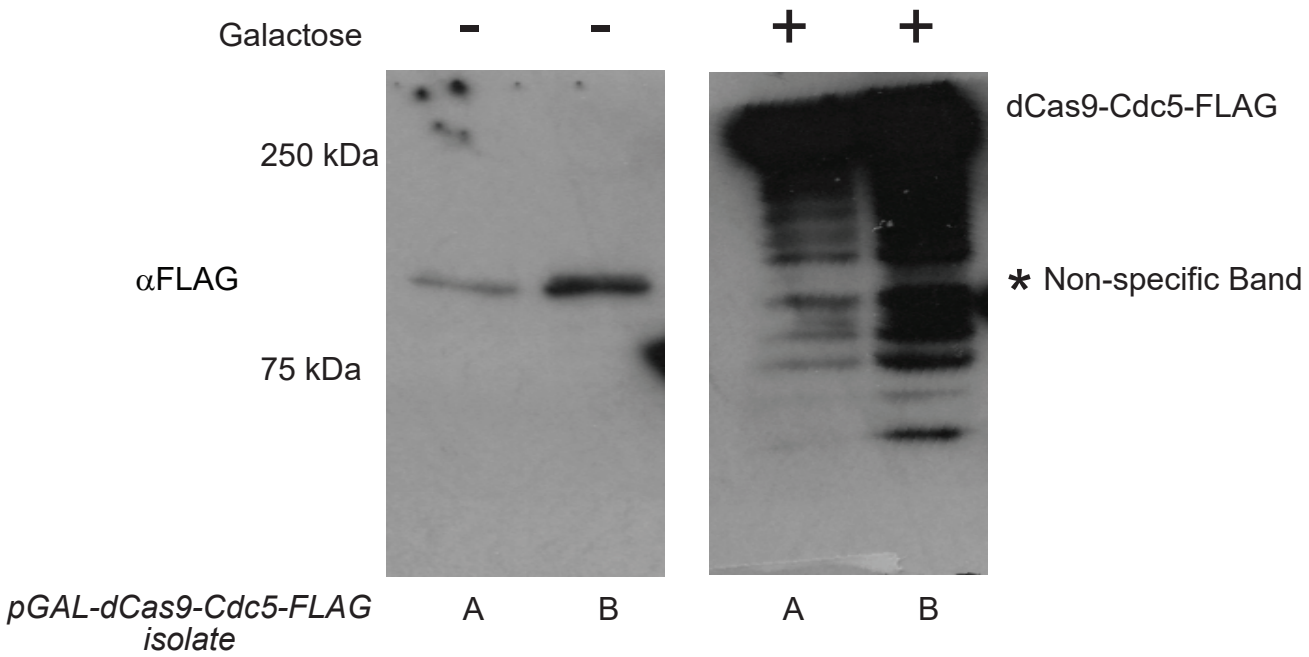


Figure 5—figure supplement 1: Characterization of dCas9-Cdc5p strains.

A. dCas9-Cdc5p supports viability in the absence of endogenous Cdc5p. Asynchronous cells were serially diluted and plated on YPD (left), YPD with 750uM auxin (center), or YPD with 750uM auxin and 2% galactose (right). Plates were then incubated at 23C for three days and imaged. Wild Type, Mcd1p-AID with pGAL-dCas9-Cdc5p-FLAG, Cdc5p-AID with pGAL-dCas9-Cdc5p-FLAG isolate 1 and 2 were plated.

B. Overexpression of dCas9-Cdc5p-FLAG by Western blotting.pGAL-dCas9-Cdc5p-FLAG isolate 1 and 2 strains were grown to mid-log in YPD. Cells were then split and grown either in the presence or absence of galactose for 1h. Cells were then pelleted and protein extracted by TCA as described in the materials and methods. Western blot analysis used FLAG antibody for dCas9-Cdc5p-FLAG.

Figure 5—figure supplement 2

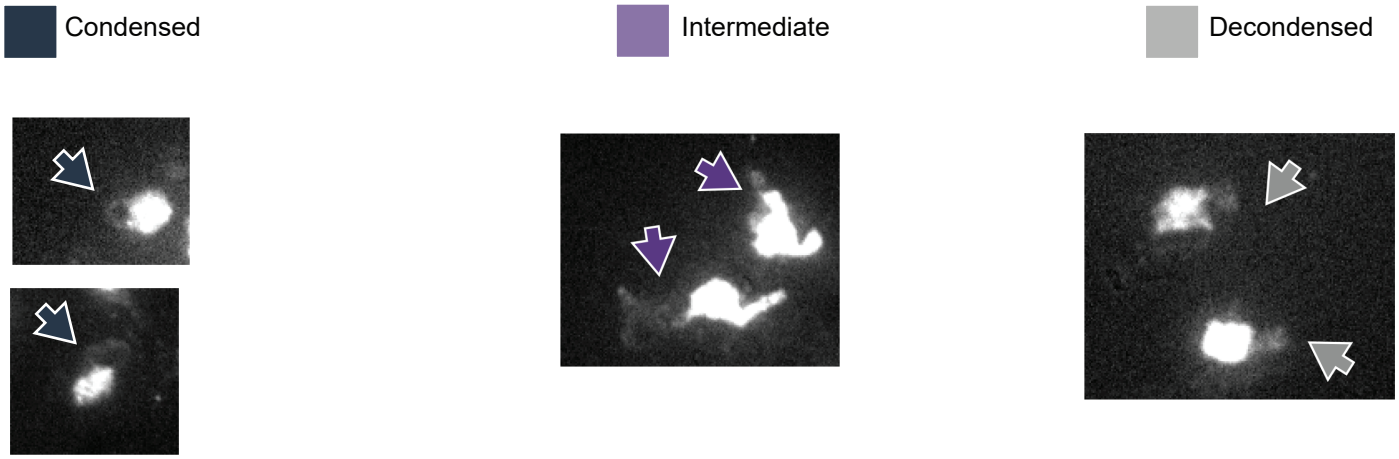


Figure 5—figure supplement 2: Chromosomes spread pictures representing the different RDN morphologies observed upon dCas9-Cdc5p expression.

dCas9-Cdc5p partially condenses RDN in the absence of endogenous Mcd1p. 'Staged mid-M' regimen: cultures were synchronized in G1 and released in media containing galactose and auxin to drive the expression of dCas9-Cdc5p concomitantly to the depletion of Mcd1p-AID. Media also contained nocodazole to arrest cells in mid-M and assess condensation. Mcd1p depleted with dCas9-Cdc5p and NTS1 gRNA expressing cells were subjected to staged mid-M' regimen and processed to score RDN condensation. Representative images of chromosome spreads show a condensed loop of the RDN in the 'condensed' class, a partially condensed line in the 'intermediate' class, and a puffed RDN in the 'decondensed' class.

Materials and Methods

Strain table

Strain	Genotype	Reference
DK5542	MATa MCD1-AID-KANMX6 ADH1-OsTIR1- URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	Eng et al. 2014
TE228	MATa PDS5-3V5-AID2-KANMX6 lys4::LacO(DK)-NAT pHIS3-GFP-LacIHIS3::his3-11,15 trp1-1 ura3-52	Eng et al. 2014
RL06	MATa CDC5-3V5-AID2-HPHMX ADH1-OsTIR1- URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL401	MATa BRN1-D375-3V5-AID2-HPHMX trp1Δ::pGPD1-TIR1-CaTRP1 lys4::LacO(DK)-NAT leu2-3,112 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
VG3620-4C	MATa trp1Δ::pGPD1-TIR1-CaTRP1 lys4::LacO(DK)-NAT leu2-3,112 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	Çamdere et al. 2015
RL404	MATa CDC5-3V5-AID2-HPHMX YCG1-S406-6MYC-KANMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL405	MATa MCD1-AID-KANMX6 YCG1-S406-6MYC-HPHMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL406	MATa MCD1-AID-KANMX6 BRN1-D375-3V5-AID2-HPHMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL407	MATa MCD1-AID-KANMX6 CDC5-3V5-AID2-HPHMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL410	MATa MCD1-AID-KANMX6 CDC5-3FLAG-HPHMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL411	MATa BRN1-D375-3V5-AID2-HPHMX CDC5-3FLAG-KANMX ADH1-OsTIR1-URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study

RL413	MATa CDC5-3V5-AID2-HPHMX cdc5-K110A::LEU2 ADH1-OsTIR1- URA3::ura3-52 lys4::LacO(DK)-NAT trp1-1 GFPLacI-HIS3:his3-11,15 bar1 leu2-3,112	This study
RL414	MATa YCG1-S406-6MYC-KANMX trp1Δ::pGPD1-TIR1-CaTRP1 lys4::LacO(DK)-NAT leu2-3,112 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
RL415	MATa CDC5-3FLAG-HPHMX trp1Δ::pGPD1-TIR1-CaTRP1 lys4::LacO(DK)-NAT leu2-3,112 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
RL416	MATa MCD1-AID-KANMX6 TIR1-LEU2 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
RL417	MATa MCD1-AID-KANMX6 pGAL-dCas9-CDC5-3FLAG-TRP1:trp1Δ (RDN5 2u URA3) TIR1-LEU2 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
RL418	MATa MCD1-AID-KANMX6 pGAL-dCas9-CDC5-3FLAG-TRP1:trp1Δ (NTS1 2u URA3) TIR1-LEU2 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study
RL419	MATa CDC5-AID-HPHMX TIR1-LEU2 pHIS3-GFPLacI-HIS3:his3-11,15 ura3-52 bar1	This study

Reagents table

Reagent type or resource	**Designation**	**Source or reference**	**Identifiers**	**Additional information**
Genetic reagent (*S. cerevisiae*)	NCBI Taxon:4932	This paper	Yeast strains	Supplementary file 1
Primers	DNA primer	IDT		
Antibody	Rabbit polyclonal Anti-Mcd1p	V. Guacci (via Covance)	RbαMcd1p (555)	WB (1:10,000) ChIP-seq (1:1,000)

Antibody	Rabbit polyclonal Anti-Mcd1p	V. Guacci (via Covance)	RbαPds5p (556)	WB (1:20,000) ChIP-seq (1:1,000)
Antibody	Mouse monoclonal Anti-MYC	Roche	MαMYC (9E10) Cat#116666006001	WB (1:10,000) ChIP-seq (1:1,000)
Antibody	Mouse monoclonal Anti-FLAG	Sigma Aldrich	MαFLAG Cat#F3165	WB (1:10,000) ChIP-seq (1:1,000)
Antibody	Mouse monoclonal Anti-V5	Invitrogen	MαV5 Cat# 46–0705	WB (1:10,000) ChIP-seq (1:1,000)
Antibody	Goat polyclonal HRP Anti-rabbit	Biorad	Cat# 170–6515	WB (1:10,000)
Antibody	Goat polyclonal HRP Anti-mouse	Biorad	Cat# 170–6516	WB (1:10,000
Antibody	Rabbit polyclonal Anti-Tub2p	P. Meluh (via Covance)	RbαTUB2	WB (1:40,000)
Dynabeads	Protein A	Invitrogen	Ref# 10002D	IP: Use 100 µl per IP
Chemical compound	Auxin (3-indole acetic acid)	Sigma Aldrich	Cat# C9911	500 µM for liquid (dissolved in Ethanol)
Chemical compound	Alpha factor	Sigma Aldrich	αF (αFactor) Cat# T6901	10-8M
Chemical compound	Nocodazole	Sigma Aldrich	Nz Cat# M1404	Double check
Reagent	Bioruptor beads	Diagenode	Cat# C01020031	200µL per sample

Sequencing and library kits	Accel-NGS 1S Plus DNA Library Kit	Swift Bioscience	Cat #10024	

Yeast strains, growth and media

All strains used in this study are derived from the A364A background and their genotypes can be found in Table 1. YEP (yeast extract, peptone, dextrose) media and YPG (yeast extract, peptone and galactose) were prepared as previously described (Guacci 1997). Conditional AID degron strains were grown in YPD or YPG and auxin (3-indoleacetic acid; Sigma Aldrich Cat. I3750) at a final concentration of 500µM to deplete AID-tagged proteins. YPD auxin agar plates were prepared by cooling molten YPD 2% agar to 55°C and supplemented with auxin at a final concentration of 750µM.

Sister chromatid cohesion

Sister chromatid cohesion was assessed as previously described (Robison and Koshland 2017). In brief, in our strains, a lacO array is integrated at the LYS4 locus on chromosome IV. These arrays can be visualized by binding of GFP-lacI, which is integrated at the HIS3 locus. To assess cohesion, cells were grown to early-log phase (OD600 0.1-0.2) at 23°C overnight and arrested in G1 using the pheromone alpha factor at a concentration of 10-8M (Sigma-Aldrich T6901-5MG). If indicated, auxin was added to a final concentration of 500µM to deplete AID-tagged proteins after a 2:30h arrest. Cells were released from G1 by washing with YPD containing auxin (if indicated) and 0.15mg/mL pronase E (Sigma Aldrich) three times, and once with YPD plus auxin. After the last wash Cells were resuspended in YPD medium containing auxin and 15 µg/mL nocodazole (Sigma Aldrich). Cultures were then once again incubated at 23°C for 3h. 1mL of these cells was then fixed in 70% ethanol and stored at 4C until ready for visualization. Cohesion was assessed by scoring the number of GFP-lacI foci in each cell on an Axioplan2 microscope (Zeiss, Thornwood, NY) using the 100X objective (numerical aperture 1.40) equipped with a Quantix charge-coupled camera (Photometrics). For time course experiments, 1mL of cells was collected every 15mn after release from alpha factor.

Assessing condensation at the RDN locus

Cells were grown as described for the sister chromatid cohesion assay. After 3h in nocodazole, cells were fixed, spheroplasted and prepared for FISH as previously described (Guacci et al 1994). DNA masses were then visualized with DAPI (4’,6-

diamidino-2-phenylindole) and imaged as described above. RDN morphology was scored as condensed “looped” or decondensed “puffy” as previously described (Guacci et al. 1994).

Maintenance Assay

For maintenance assays, cells were grown to early log phase at 23C and arrested in mid-M using 15µg/mL nocodazole. For AID-tagged proteins, auxin was added after synchronous mid-M arrest, 2h after nocodazole addition. Cells were collected after 1h of auxin addition for condensation assay.

Add-later time course

Cells were grown as described in the sister chromatid cohesion assay to get synchronous population of cells arrested in mid-M depleted for the AID-tagged protein(s). To restore the presence of the AID-tagged proteins, these cells were washed 3 times in YPD medium containing nocodazole and grown in media containing nocodazole for 2h. Samples were collected after 2h to assess condensation state and protein levels by TCA extraction followed by Western.

Chromatin immunoprecipitation (ChIP) and sequencing

Cells were grown and arrested at 30oC as described for assessing condensation. 60OD of mid-M cells were collected, fixed for one hour with 1% formaldehyde, quenched with 5mM glycine and processed for ChIP as previously described (Robison and Koshland 2017, with the following modifications). Frozen cell pellets were disrupted 3 times using a FastPrep-24 5G instrument (40 seconds, 6.0m/sec each) (MP Bio, Santa Ana, CA). Chromatin was sheared 10 x 30s ON/30s OFF with a Bioruptor Pico (Diagenode, Denville, NJ). For ChIP-qPCR, 1/20 of sheared and cleared lysate was reserved as input. Chromatin extracts were incubated with 4uL of the following antibodies monoclonal mouse anti-MYC (Roche), monoclonal mouse anti-FLAG (Sigma Aldrich), polyclonal rabbit anti-Pds5p (Covance Biosciences, Princeton, NJ), or polyclonal rabbit anti-Mcd1p (Covance Biosciences, Princeton, NJ) overnight at 4C. Antibody-bound lysates were incubated with 120uL Protein A dynabeads (Invitrogen, Cat#: 10001D) for 1h. Following ChIP, the library was prepared using the Accel-NGS 1S Plus DNA Library Kit (Swift Bioscience) following the manufacturer protocol. Libraries were sequenced using Illumina Hiseq 4000. The sequencing files were aligned to the SacCer3 yeast genome using Bowtie2 tool.

TCA protein extraction and Western blotting

To assess protein level, cells were collected, pelleted, washed with 1X PBS and frozen at -80. Frozen pellets were resuspended in 200uL 20% TCA and broken for 40s using a FastPrep-24 5G instrument (6.0m/sec each) (MP Bio, Santa Ana, CA). Lysates were diluted with 1mL 5%TCA and spun for 10mn at 14,000rpm at 4C. Pellets were resuspended in 2X Laemmli buffer and boiled for 7mn and spun. Cleared lysates were used for Western blotting on 6% SDS-PAGE gels.

Acknowledgements

We thanks Thomas Eng and Vincent Guacci for building some of the strains used. We thank Elcin Unal for sharing the dCas9 plasmid and Fred Winston for critical reading of the manuscript. We thank the Koshland lab and the Unal lab for imputs and discussions. This work was funded by National Science Foundation GRFP grant to RL and National Institutes of Health Grant (1R35 GM-118189-01 to DK).

Competing interests

No competing interests to declare

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