Topology-driven analysis of protein-protein interaction networks detects functional genetic modules regulating reproductive capacity

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Abstract

Understanding the genetic regulation of organ structure is a fundamental problem in developmental biology. Here, we use egg-producing structures of insect ovaries, called ovarioles, to deduce systems-level gene regulatory relationships from quantitative functional genetic analysis. We previously showed that Hippo signalling, a conserved regulator of animal organ size, regulates ovariole number in *Drosophila melanogaster*. To comprehensively determine how Hippo signalling interacts with other pathways in this regulation, we screened all known signalling pathway genes, and identified Hpo-dependent and Hpo-independent signalling requirements. Network analysis of known protein-protein interactions among screen results identified independent gene regulatory modules regulating one or both of ovariole number and egg laying. These modules predict involvement of previously uncharacterised genes with higher accuracy than the original candidate screen. This shows that network analysis combining functional genetic and large-scale interaction data can predict function of novel genes regulating development.

Keywords : *Drosophila melanogaster*, Reproduction, Ovariole, Ovary, Egg laying, Topology, Network analysis, Interactome, Hippo signalling.

Introduction

The final shape and size of an organ is critical to organismal function and viability. Defects in human organ morphology cause a multitude of pathologies, including cancers, organ hypertrophies and atrophies (e.g. Yang and Xu, 2011). It is thus critical to understand the regulatory mechanisms underlying the stereotypic shape and size of organs. To this end, assessing the genetic regulation of size is significantly facilitated by using quantifiable changes in organ size and shape.

The *Drosophila melanogaster* female reproductive system is a useful paradigm to study quantitative anatomical traits. In these organs, the effects of multiple genes and the environment combine to produce a quantitative phenotype: a species-specific average number of egg-producing ovarian tubes called ovarioles. Fruit fly ovaries can contain as few as one and as many as 50 ovarioles per ovary, depending on the species (Kambysellis and Heed, 1971; King, 1970; Markow et al., 2009; Sarikaya et al., 2019), with each ovariole capable of producing eggs. Ovariole number, therefore, may affect the reproductive fitness of *Drosophila* species by determining the potential of an adult female to produce eggs (Klepsatel et al., 2013b; R´kha et al., 1997). While ovariole number within a species can vary across temperatures (Azevedo et al., 1996), altitudinal and latitudinal clines (Capy et al., 1994; David and Bocquet, 1975), under constant environmental conditions ovariole number is highly stereotypic (Capy et al., 1993; Klepsatel et al., 2013a; R'Kha et al., 1991; R´kha et al., 1997). The reproducibility of ovariole number thus indicates a strong genetic component (Sarikaya et al., 2019). Genome wide association studies and quantitative trait locus mapping have demonstrated that the ovariole number is a highly polygenic trait (Bergland et al., 2008; Lobell et al., 2017; Orgogozo et al., 2006; Wayne et al., 2001; Wayne et al., 1997; Wayne and McIntyre, 2002). In contrast, functional genetic studies have identified only a small number of genes whose activity regulates ovariole number (discussed below). Thus, the complexity of the genetic regulation of this important trait remains largely unknown.

The determination of ovariole number in *D. melanogaster* occurs during late larval and pupal development (King et al., 1968). Each ovariole in the adult fly arises from a single primordial structure called a terminal filament (TF), which forms in the late third instar larval ovary (Godt and Laski, 1995) by convergent extension (Keller, 2006) of the terminal filament cells (TFCs) (Godt and Laski, 1995; Sahut-Barnola et al., 1996). TFCs are first specified from an anterior

population of somatic cells in the larval ovary by the expression of transcription factors including

Bric-à-brac 1/2 (*bric-à-brac 1/2; bab1/2*) and Engrailed (*engrailed; en*) (Godt and Laski, 1995)

(Sahut-Barnola et al., 1995). Initially a loosely arranged group in the anterior of the larval ovary,

TFCs undergo morphogenetic movements to give rise to the ordered columns of cells that are

TFs. Cell intercalation during convergent extension is dependent on the actin regulators Cofilin

(*twinstar*) and the large Maf factor Traffic Jam (*traffic jam; tj*), and on E-cadherin dependent

adhesion (Chen et al., 2001; Godt and Laski, 1995). Regulation of ovariole number is thus

largely dependent on the specification of the TFCs and their rearrangement into TFs (Sarikaya

and Extavour, 2015).

We previously showed that the regulation of both TFC and TF number is dependent on the

Hippo signalling pathway (Sarikaya and Extavour, 2015) , a pan-metazoan regulator of organ

and tissue size (Hilman and Gat, 2011; Sebe-Pedros et al., 2012). At the core of the Hippo

kinase cascade are two protein kinases, Hippo (*hippo; hpo*) and Warts (*warts*), which prevent

the nuclear localisation of the transcriptional co-activator Yorkie (*yorkie; yki*). Yki and the

transcription factor Scalloped (*scalloped*) together initiate the transcription of multiple gene

targets, including those that promote cell proliferation and survival. In the *D. melanogaster* larval

ovary, loss of Hpo in the somatic cells causes an increase in nuclear Yki, leading to an increase

in TFCs, TFs, ovariole number and egg laying in adults (Sarikaya and Extavour, 2015).

Production of fertile eggs from a stereotypic number of ovarioles requires a spatially and

temporally coordinated interplay of signalling between the somatic and germ line cells of the

ovary. Thus, signalling amongst somatic and germ line cells in the larval ovary is crucial to all

stages of ovarian development (Ables and Drummond-Barbosa, 2017; Gilboa, 2015; Green II et

al., 2011; LaFever and Drummond-Barbosa, 2005; LaFever et al., 2010; Sarikaya and Extavour,

2015). For instance, disruptions in insulin or Tor signalling affect both somatic and germ line cell

proliferation (Gancz and Gilboa, 2013; Green II and Extavour, 2012; Hsu and Drummond-

Barbosa, 2009; LaFever and Drummond-Barbosa, 2005; LaFever et al., 2010; Sarikaya et al.,

2012). Similarly, ecdysone pulses from the prothoracic gland regulate the timely differentiation

of the primordial germ cells (PGCs) and the somatic TFCs (Gancz et al., 2011; Hodin and

Riddiford, 1998, 2000b). Both Hpo and ecdysone signalling also control the proportion of germ

line to somatic cells by differentially regulating proliferation of both cell types (Gancz et al.,

2011; Sarikaya and Extavour, 2015).

Although it is clear that genes function together in regulatory networks (Gonzalez and Kann,

2012), determining how the few genes functionally verified as required for ovariole development

and function, work together to coordinate ovariole number and ovarian function more generally,

is a challenge because most genes or pathways have been considered individually. An

alternative approach that is less often applied to animal developmental genetics, is a systems

biology representation of complex biological systems as networks (Barabási and Albert, 1999;

Watts and Strogatz, 1998). Protein-protein interaction (PPI) networks are such an example

(Albert and Barabási, 2002). The availability of high throughput molecular biology datasets from,

for example, yeast two-hybrid, protein CHiP and microarrays has allowed for the emergence of

large scale interaction networks representing both functional and physical molecular interactions

(Barabási and Oltvai, 2004; Berger et al., 2007; Giot et al., 2003; Gonzalez and Kann, 2012).

With ample evidence that signalling in the ovary can affect ovarian development, but few genes

functionally verified to date, we aimed to identify novel regulators of ovariole development by

functionally testing all known members of all characterized *D. melanogaster* signalling

pathways. We used tissue-specific RNAi to systematically knock down 463 genes in the larval

ovary, and looked for modifiers of the *hpo* loss of function egg laying and ovariole number

phenotypes. To analyse the results of this phenotypic analysis, we used topology-driven

network analysis to identify genetic modules regulating these phenotypes, thus generating

hypotheses about the relationships between these modules. With this systems biology

approach, we identify not only signalling pathway genes, but also previously untested genes

that affect these reproductive traits. Functional testing showed that these novel genes affect

ovariole number and/or egg laying, providing us with a novel *in silico* method to identify target

genes that affect ovarian development and function. We use these findings to propose putative

developmental regulatory modules underlying one or both of ovariole formation and egg laying.

122 Results

123 An RNAi modifier screen for signalling pathway involvement in ovariole
124 number

To systematically ascertain the function of signalling pathway genes and their interactions with

Hippo signalling in the development of the *D. melanogaster* ovary, we first curated a list of all

known and predicted signalling genes (Gramates et al., 2016; Kanehisa et al., 2010; Mbodj et

al., 2013). We identified 475 genes belonging to the 14 developmental signalling pathways

characterised in *D. melanogaster* (Table 1; Table S1), and obtained UAS:RNAi lines for 463 of

these genes from the Vienna *Drosophila* RNAi centre (VDRC) or the TRiP collections at the

Bloomington *Drosophila* Stock centre (BDSC) (all *D. melanogaster* genetic lines used are listed

in Methods).

We previously showed that reducing the levels of *hpo* in the somatic cells of the larval ovary

using *traffic jam* Gal4 (*tj*:*Gal4*) driving *hpo[RNAi]* increased both ovariole number and egg laying

of adult female flies (Sarikaya and Extavour, 2015). To identify genes that modify these

phenotypes, we used *tj*:*Gal4* to drive simultaneous *hpo[RNAi]* and *RNAi* against a signalling

candidate gene, and quantified the phenotypic change (Figure 1a-d). We observed that on

driving two copies of *hpo[RNAi]* using *tj*:*Gal4*, we obtained a further increase in both egg laying

and ovariole number (Figure 1e). This indicates that ovaries have further potential to increase

ovariole number and egg laying beyond the increase induced by *tj:Gal4* driving one copy of

hpo[RNAi], and that *tj:Gal4* can drive the expression of two RNAi constructs, indicating that our

screen could identify both enhancers and suppressors of the *tj:Gal4>hpo[RNAi]* phenotype.

We proceeded to identify modifiers of the *tj:Gal4>hpo[RNAi]* phenotype by crossing males of

each of the 463 candidate genes RNAis individually with *tj:Gal4>hpo[RNAi]* females, and

performing three phenotypic screens on the offspring. In the first screen (Figure 1a), we

measured egg laying of three F1 female offspring (*tj:Gal4>hpo[RNAi], signalling*

candidate[RNAi]) over 5 days. To address batch variation (Figure S1), we standardized egg

laying measurements by calculating the Z scores (Z_{gene} = number of standard deviations from

the mean) for each candidate line relative to its batch controls. 190 genes had an egg laying

|Z_{gene}| below 1. Previous studies have shown that the egg laying of newly eclosed adult mated

females correlates with ovariole number during the first five days (Klepsatel et al., 2013b). We

therefore eliminated these 190 genes from subsequent screening, because the change in egg

laying was so modest that we considered these candidates were unlikely to show changes in

ovariole number when compared to controls.

In the second screen (Figure 1b), we measured egg laying in a wild-type background

(*tj>signalling candidate[RNAi]*) for the 273 remaining candidate genes. For the third screen

(Figure 1c), we quantified the ovariole number of *tj:Gal4>hpo[RNAi], signalling candidate[RNAi]*

F1 adult females for the same 273 candidate genes. To choose candidates from the second and

third screens for further study, we wished to account for the fact that the two screens had

different effective numbers of data points. This was because egg laying data were obtained from

individual vials of three females over five days, while ovariole numbers were obtained from 20

ovaries from ten females (see methods). We therefore selected the 67 genes with a $|Z_{gene}|$ above

two for ovariole number (Figure 1c, 1d; Table 2), and the 49 genes with a more conservative

$|Z_{gene}|$ above five for egg laying (Figure 1a, 1b, 1d; Table 2), for a total of 116 positive candidates

for subsequent analyses.

170   Ovariole number is weakly correlated with egg laying

Standardization of the results from the three screens using Z scores allowed us to compare the

effects of individual genes on one or both of egg laying and ovariole number. We performed a

pairwise comparison of the Z_{gene} values for all combinations of screens, and considered genes

with $|Z_{gene}|$ values that were above the thresholds set for the phenotype in each screen (above

two for ovariole number, above five for egg laying; green dots in Figure 2a-c. Across all three

screens, loss of function of our positive candidates yielded reductions in ovariole number and

egg laying more commonly than increases (Figure 2a-c). Comparing the $|Z_{gene}|$ values of egg

laying and ovariole number of *tj:Gal4>hpo[RNAi], signalling candidate[RNAi]* adult females

revealed that genes that caused a change in egg laying did not always similarly affect ovariole

number, and vice versa (Figure 2a). We therefore hypothesise that egg laying and ovariole

number may be regulated by genetically separable mechanisms. This hypothesis

notwithstanding, we observed a weak but statistically significant correlation between egg laying

and ovariole number (p=$1e10^{-5}$; Figure 2d), and this correlation was most significant in adult

females that had a drastic reduction in both phenotypes (Figure 2a).

186   No single signaling pathway dominates regulation of ovariole number or
187   egg laying

We found that at least some genes from all tested signalling pathways could affect both egg

laying and ovariole number (Figure 3). To determine if some pathway(s) appeared to play a

more important role than others in these processes, we asked whether any of our screens were

enriched for genes from a specific signalling pathway. To measure enrichment, we compared

the distribution of individual pathway genes among the positive candidates in each screen to a

bootstrapped null distribution of pathway genes among a group of the same number of genes

randomly selected from our curated list of 463 signalling genes (Figure 3a). Involvement of a

pathway in the regulation of a phenotype would be reflected in a difference between the

representation of pathway genes in an experimentally derived list and a randomly selected

group of signalling genes. We found that rather than only one or a few pathways showing

functional evidence of regulating ovariole number or egg laying, nearly all pathways affected

both phenotypes (Figure 3a). We further tested this result by calculating the hypergeometric p-

value for the enrichment of each signaling pathway, in each of the three groups of genes.

Consistent with the results of the bootstrapping approach (Figure 3a), we found that most

pathway members were not significantly enriched for egg laying or ovariole number phenotypes

(Figure 3b). The absence of significant enrichment of any specific pathway is not simply

attributable to the pool of genes that were screened, because our experimental manipulations of

ovariole number and egg laying did cause a change in the distribution of signalling pathway

members (Figure S2a). Instead, both phenotypes appeared to be regulated by members of

most or all signalling pathways (Figure 3; Figure S2). The only two exceptions to this trend were

a greater than twofold enrichment of (1) genes from the Notch signalling pathway in the

regulation of ovariole number (p-value < 0.05, pink bar in Figure 3a, b), and (2) members of the

Hedgehog (Hh) signaling pathway in the regulation of Hippo-dependent egg laying (p-value <

0.05, brown bar in Figure 3a, 3b; Figure S3).In summation, our analyses of the enrichment of

signalling pathways within the different screens indicated that both ovariole number and egg

laying are regulated by genes from nearly all described animal signalling pathways (Figure 3a),

rather than being dominated by any single pathway.

Comparing the results of the Egg Laying screens performed in a wild type background (Figure

1b) or in a *hpo[RNAi]* background (Figure 1a), revealed that most of the genes that met a

threshold of $|Z_{gene}|$ >5 in one screen, did not meet that threshold in the other screen (Figure 2c).

This result suggests the existence of both Hippo-dependent and Hippo-independent

mechanisms of regulation of egg laying. The interpretations of separable Hippo-dependent and -

independent regulation of egg laying, and of the separable regulation of ovariole number and

egg laying, was supported by the results of the network analysis described in the following

section.

Centrality of genes in the ovarian protein-protein interaction networks can predict the likelihood of loss of function phenotypic effects

The finding that these reproductive traits were regulated by the genes of all signalling pathways led us to consider the broader topology of putative gene regulatory networks in the analysis of our data. Previously characterized genes in the ovary are often pleiotropic and can regulate both ovariole number and egg laying (Gilboa, 2015; Sarikaya and Extavour, 2015). As with proteins in a linear pathway, proteins in a protein-protein interaction (PPI) network are more likely to function in conjunction with genes that are connected to them within the network (e.g. Ideker and Sharan, 2008; Jeong et al., 2001). Centrality is one measure of the connectedness of a gene in the PPI and can be used to identify the most important functional centres within a protein network (Hahn and Kern, 2005; Ma'ayan, 2011). Most centrality measures use path length, which is a measure of the number of other proteins required to link any two proteins in the network. Here we used four commonly used metrics to quantify gene centrality, each measuring slightly different properties (Jalili et al., 2016; Koschutzki and Schreiber, 2008). (1) *Degree centrality* is proportional to the number of proteins that a given protein directly interacts with. (2) *Betweenness centrality* measures the number of shortest paths amongst all the shortest paths between all pairs of proteins that require passing through a particular protein. (3) *Closeness centrality* measures the average shortest path that connects a given protein to all other proteins in the network. (4) *Eigenvector centrality* is a measure of the closeness of a given protein to other highly connected proteins within the network.

We hypothesised that if the candidate genes we identified in our screen as playing roles in ovarian function worked together as a PPI network, then the degree of centrality of a gene might be an indicator of function. To test this hypothesis, we calculated the four centrality measures, described above, for all genes within the *D. melanogaster* PPI (Figure S4). We then rank ordered only the genes tested in each screen by their score for each centrality measure, and asked whether their rank order correlated with the results of the screen, plotting these results as a receiver operating characteristic (ROC) curve. Positive correlations between centrality (a continuous variable) and phenotype (a binary variable: above or below the $|Z_{gene}|$ threshold) are reflected in an area under the curve (AUC) of more than 0.5. We found that the higher the centrality score, the greater the likelihood that a gene had $|Z_{gene}|$ values above our threshold for effects on ovariole number and egg laying (Figure 4a; Table S3). This supports the premise that the positive candidates identified in our screen function together as a network in the regulation

of either ovariole number or egg laying. Interestingly, while the centrality of genes did predict

whether a gene would affect our phenotypes of interest, it could only weakly predict the strength

of that effect (Figure S5).

Genes regulating egg laying and ovariole number regulation form non-

random interaction modules

The centrality analyses above suggested that the genes implicated in ovariole number and egg-

laying displayed characteristics of a functional network. PPI networks can often be further sorted

into a collection of sub-networks. A sub-network is a smaller selection of proteins from the PPI.

Examples of such sub-networks could be proteins within the same subcellular organelle (Foster

et al., 2006) or genes that are expressed at the same time (Spellman et al., 1998), thus making

them likely to function together (Srinivasan et al., 2007). A module is a sub-network that can

perform regulatory functions independent of other sub-networks, and has key measurable

features (Barabási and Oltvai, 2004; Hartwell et al., 1999; Ravasz et al., 2002; Yook et al.,

2004). We therefore asked if our sub-networks consisting of genes that showed similar mutant

phenotypes might constitute such functional modules. To determine whether genes that were

implicated in regulation of ovariole number and egg laying interacted with each other in specific

groups more than would be expected by chance, we created four lists of genes, called "seed"

lists, based on their individual phenotypic effects based on our screen results: (1) the core seed

list, including genes positive in all three screens (Figure 4b); (2) the egg laying seed list,

including genes positive in the wild type background egg-laying screen (Figure 1b; Figure 4c);

(3) the *hpo[RNAi]* egg laying seed list, including genes positive in the *hpo[RNAi]* background

egg laying screen (Figure 1a; Figure 4c); and (4) the *hpo[RNAi]* ovariole seed list, including

genes positive in the *hpo[RNAi]* background ovariole number screen (Figure 1c; Figure 4c).

Interestingly, the core seed list, comprising genes that affected all three measured phenotypes,

only consisted of genes that caused a reduction in both ovariole number and egg laying (Figure

4b).

Based on published molecular interactions, putative functional modules of genes can be

predicted by algorithms that use either the shortest path method (Bromberg et al., 2008) or the

Steiner Tree approach (Huang and Fraenkel, 2009). Such methods identify and predict

functional connections between the seed proteins, as well as additional nodes (proteins or

genes) that have not been experimentally tested within the given parameters, but are known to

interact with the seed genes in the PPI (Albert and Albert, 2004)((Yu et al., 2006). This process

results in a predicted module, and subsequent functional genetic testing of this module can

confirm its functionality. Given its recent success in predicting gene modules, we used the

previously published Seed Connector Algorithm (SCA), a member of the Steiner Tree algorithm

family (Wang et al., 2017; Wang and Loscalzo, 2018), to identify putative functional modules

formed by genes that had similar phenotypic effects in our screens (Figure 4b, 4c). The SCA

connects seed genes and previously untested novel genes (connectors) to each other using a

known PPI network, producing the largest possible connected putative module given the data.

We compiled a PPI network consisting of all described interactions between *D. melanogaster*

proteins, from the combination of publicly available PPI studies in the DroID database (see

Methods). Using this PPI network and the aforementioned four seed lists, we applied a custom

python implementation of the SCA (Methods: 04_Seed-Connector.ipynb) to build and extract the

largest possible (given our PPI) connected putative modules that regulate egg laying and

ovariole number.

This SCA method yielded four putative modules, one for each seed list, which we refer to as the

core module (Figure 5b), *hpo[RNAi]* Egg Laying module (Figure S6a), Egg Laying module

(Figure S6b), and *hpo[RNAi]* Ovariole Number Module (Figure S6c) respectively. Each of the

modules contained seed genes, which had been functionally evaluated in our screens (green

and red circles in Figure 5), as well as connector genes, which were genes newly predicted as

regulators of these phenotypes (green and red triangles in Figure 5).

We then asked whether these four putative modules were more connected than we would

expect by chance; in other words, we formally tested them for modularity. Meeting our criteria

for modularity would suggest that the genes in these modules operated together as functional

sub-networks within the *Drosophila* PPI. We defined our modularity test using four commonly

measured network metrics: (1) Largest Connected Component (LCC) (the number of proteins

connected together by at least one interaction), (2) network density (the relative number of

edges as compared to the theoretical maximum), (3) total number of edges, and (4) average

shortest path (average of the minimum distances connecting any two proteins). We considered

a group of genes to form a module if they showed higher LCC, higher network density, more

edges, and shorter average shortest path length than a random, bootstrapped selection of the

same number of genes from the PPI.

To determine whether these criteria would correctly identify genes of the same signaling

pathway, which are known to function together, as a module, we measured these four

parameters in the original set of genes used in this study (Table 1). We found that all the genes

of a given signalling pathway were identified as a module based on these parameters (Figure

S7a). We then used this approach to test the modularity of the four phenotypic sub-networks, as

compared to 1000 bootstrapped "control sub-networks" consisting of a group of the same

number of genes as contained in the sub-network, but chosen randomly from among the

candidate genes from our initial screen list (Table 1). We found that the four predicted

phenotypic modules showed a significantly increased Largest Connected Component (LCC)

value, network density, number of edges and decreased average shortest path (Figure S7b),

compared to our "control module". This result indicates that these sub-networks are likely to

function as modules within the PPI, to regulate one or both of ovariole number or egg laying.

Low edge densities between modules indicates genetically separable mechanisms of ovariole number and egg laying

Our network analysis identified four highly connected networks of genes that regulate two

distinct developmental processes, together with or independently of Hippo signalling activity:

ovariole number determination, which occurs primarily during larval development, and egg

laying, which takes place in adult life (Figure 5). We wished to assess the degree to which there

were any shared genetic components between the four modules. To understand potential

interactions between the modules in the regulation of both ovariole number and egg laying, we

constructed a composite network of all genes in each of the four modules (Figure 5b; Figure

S6), which we refer to as the "meta network" (Figure 7a). We then grouped the genes based on

their phenotypic effects as measured in the three screens, resulting in seven sub-networks (I-VII

in Figure 7a). We then asked if these sub-networks were as connected to each other, as were

the genes within each of the sub-networks. To do this, we used an edge density map, which

reflects the number of interactions between the genes within a sub-network and between each

of the sub-networks (Figure 7b).

This analysis yielded three principal findings. First, edge densities between the three sub-
networks corresponding to the three scored phenotypes (I, II and III in Figure 7a) were very low
(Figure 7b). This indicates that genes in each of these sub-networks function as largely
independent networks, rather than interacting substantially with any genes in the other non-core
sub-networks. Second, the core sub-network (IV in Figure 7a) displayed a higher edge density
with the other three sub-networks (I, II and III in Figure 7a) than any of them did with each other
(Figure 7b). Consistent with the definition of core module genes as regulating all three scored
reproductive phenotypes, this result suggests that the core module genes, in contrast to those
from the other three sub-networks, may share substantial functional interactions with genes of
the other sub-networks. Finally, three small additional sub-networks emerged from this analysis
(V, VI and VII in Figure 7a), suggesting small functional networks of genes that work together to
regulate two of the three scored phenotypes. In sum, this meta network analysis supports the
hypothesis of three largely independent genetic networks that regulate Hippo-dependent
ovariole number, Hippo-dependent egg laying, and Hippo-independent egg laying. Moreover,
each of these genetically separable networks included genes in multiple signalling pathways
(Figure 7c).

Network analysis predicts novel genes involved in egg laying and ovariole number

The four predicted phenotypic modules produced by the SCA approach included connector
genes that were not included in our original screen, and thus had not been tested for possible
effects on our phenotypes of interest (triangles in Figure 5b; Figure S6). Given that prior work in
human disease models showed that predicted disease modules can correctly predict gene
involvement in the relevant diseases (Chen et al., 2006; Gonzalez et al., 2007; Wang et al.,
2017; Wang and Loscalzo, 2018), we asked whether our deployment of the SCA had likewise
successfully predicted novel, functionally important genes in each module. To do this, we
measured the effects of knocking down each the connector genes (triangles in Figure 5b and
Figure S6) on ovariole number and egg laying, using *UAS:RNAi* lines for each connector, driven
by *tj:Gal4*.

Of the ten predicted novel connectors in the core module, loss of function of several of these
had significant effects on at least one of the three scored phenotypes. Five affected ovariole

number two affected Hpo-dependent egg laying, and one affected Hpo-independent egg laying.
However, only one of them significantly altered all three scored phenotypes (Figure 6a; Table
S4).

The predicted connector genes from two of the other three phenotypic modules showed high
positive prediction rates. RNAi against seven out of 18 of the *hpo[RNAi]* Egg Laying module
connectors, three out of 11 of the *hpo[RNAi]* Ovariole Number module connectors, and none of
the 11 Egg Laying module connectors, significantly affected the module phenotype (Table S4).
Thus, although the Egg Laying module connectors failed to impact this phenotype in our assay,
41.2% and 27.3% of the connectors from the other two modules were correctly predicted
(Figure 6b; Table S4). These positive hit rates exceed those obtained in our initial candidate
screens, where 59/463 (12.7%) and 67/273 (24.5%) tested genes affected *hpo*-dependent egg
laying and *hpo*-dependent ovariole number respectively (Figure 6d; Table 2). In sum, taken
across all modules (Figure 6c; Table S4, Table S5), this network analysis correctly identified
genes regulating all scored reproductive phenotypes, at rates higher than those obtained in the
original screen of 463 members of all known signalling pathways. By this measure, testing
network-predicted regulatory modules derived from experimentally obtained data was even
more successful than testing signalling pathways as a means of identifying novel genes that
regulate ovariole number and egg laying.

Discussion

Identification of regulatory modules for ovariole development and egg laying

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The *D. melanogaster* ovary is a commonly studied model for organogenesis (Chen et al., 2001;
Godt and Laski, 1995; Lobell et al., 2017; Sarikaya and Extavour, 2015), stem cell maintenance
(Gilboa, 2015) and interactions of development and ecology (Cohet and David, 1978; Hodin and
Riddiford, 2000a; Klepsatel et al., 2013a; Sarikaya et al., 2019). Nevertheless, our
understanding of the genetic mechanisms that regulate these processes remains fragmentary.
In this paper, we have identified four distinct protein interaction modules that regulate ovariole
number and egg laying in the *D. melanogaster* ovary. These modules consist of both novel and

previously characterized genes that regulate either ovariole number or egg laying or both, thus

enhancing our understanding of the genetic underpinnings of this reproductive system.

Of the four modules, the core module affects both ovariole number and egg laying. The core

module contains numerous housekeeping genes, including regulators of transcription,

translation and cell division such as *polo* (Llamazares et al., 1991), *cyclin K* (Edwards et al.,

1998), *nucleosome assembly protein 1* (Ito et al., 1996) and *eukaryotic translation release factor*

1 (Chao et al., 2003). While *polo* and *eukaryotic translation release factor 1* are members of

signalling pathways, *cyclin K* and n*ucleosome assembly protein 1* are genes predicted by the

SCA. Given that the core module largely consists of genes whose loss of function decreases

both of these parameters, we hypothesise that these are essential genes for the basic structure

and function of the ovaries. Essential genes are more interconnected in a PPI with higher

centrality measures (Jeong et al., 2001) and interestingly, we find that the genes in the core

module also have higher connectivity than those in the other three modules (Figure S4).

In addition to genes that regulate basic cellular processes, the core module is enriched for the

core components of the Hh signalling cascade, namely *patched (ptc)*, *smoothened* (*smo*) and

costa (cos) (Lee et al., 2016). However, we find that the loss of Hh ligand, which is expressed in

the TF cells in the developing larval ovary (Lai et al., 2017), does not significantly affect either

ovariole number or egg laying. Though surprising, ligand-independent activation of Hedgehog

signalling has been observed before. For example, in the *Drosophila* eye, loss of either *ptc* or

cos in clones leads to non-cell autonomous proliferation in wild type cells, as well as growth

disadvantages in the mutant tissue (Christiansen et al., 2012). In another example, sufficient

intracellular *smo* levels can also activate downstream transcription of Hh pathway targets,

showing that Hh itself is not always required to activate the cascade (Jiang et al., 2018).

435   Development of the larval ovary

The *hpo[RNAi]* Ovariole Number module is composed of genes that affect the Hippo signalling

activity-dependent determination of ovariole number during development. Establishment of

ovariole number occurs largely during the third instar stage of larval development in *D.*

melanogaster (Godt and Laski, 1995; Hodin and Riddiford, 1998; King et al., 1968; Sahut-

Barnola et al., 1996). During this period, the TFCs are specified in the anterior of the ovary and

undergo rearrangement into stacks of cells called TFs, each of which gives rise to an ovariole

(Godt and Laski, 1995; Sahut-Barnola et al., 1995). TF specification requires the expression of

engrailed (En) (Bolívar et al., 2006) and the transcription factors Bab1 and Bab2, encoded by

the *bric-à-brac* locus (Couderc et al., 2002; Godt et al., 1993). A third transcription factor,

Lmx1a, was recently found to be necessary for the specification of the TFCs (Allbee et al.,

2018). Our *hpo[RNAi]* Ovariole Number module identifies numerous additional novel

transcription factors including *bunched* (*bun*) and *retinoblastoma-family protein* (*rbf*), which we

hypothesize could also be involved in the specification of ovariole number. *bun* and *rbf* have

been implicated in the migration (Dobens et al., 2005) and endoreplication (Cayirlioglu et al.,

2003) of the follicle cells during oogenesis, but have not, to our knowledge, been previously

identified as playing a role in the context of larval ovary development.

The TFCs specified in the larval ovary undergo a process of convergent extension to form TFs.

This process of convergent extension requires cell intercalation, and the actin depolymerizing

factor Cofilin, encoded by the gene *twinstar*, is essential to this process (Chen et al., 2001).

During intercalation, the cells also dynamically modify their actin cytoskeleton and their

expression of E-cadherin (Godt and Laski, 1995). Our *hpo[RNAi]* Ovariole Number module

further identifies *Rho1* (Barrett et al., 1997) and *Rho kinase* (*Rok*) (Mizuno et al., 1999) as

necessary for correct ovariole number. During the extension of the *D. melanogaster* embryonic

germ band, a commonly studied model of convergent extension, the localised activation of the

actin-myosin network facilitated by *Rho1* and *Rok* is necessary for cell intercalation (Kasza et

al., 2014). Given the known roles of *Rho1* and *Rok* as regulators of the actin cytoskeleton

(Ridley, 2006), we propose that TF assembly in the ovary requires both these proteins for

correct cell intercalation. A third actin cytoskeleton regulator, *misshapen* (*msn*), was also

identified by our *hpo[RNAi]* Ovariole Number module. *msn* encodes a MAP kinase previously

shown to regulate the polarisation of the actin cytoskeleton during oogenesis (Lewellyn et al.,

2013), but has not, to our knowledge, been studied to date in the context of larval ovarian

development.

We propose that the polarity of the somatic cells in the ovary is also necessary for correct larval

ovary development, given the presence of the lateral membrane proteins *discs large 1* (*dlg1*)

and *prickle* (*pk*) in the ovariole module. During the maturation of the TFs during larval

development, the TFCs undergo significant cell shape changes, coincident with localised

expression of beta-Catenin and actin to the lateral edges of the TFCs (Godt and Laski, 1995).

Restriction of the E-cadherin domain in epithelia requires establishment of the basolateral

domain (Harris and Peifer, 2004) and we propose that testing a similar requirement for *dlg1* and

pk in the larval ovary would be a fruitful avenue for future studies.

Network analysis as a tool in developmental biology

Using a systems biology approach to analyse RNAi screening data has proven fruitful, providing

us with new insights into the development and function of the *D. melanogaster* ovary by

identifying novel and previously understudied genes that regulate this process. Systematic

analysis of the function of single genes in development has been a historical convention and

has provided valuable and precise genetic interaction information (Jansen et al., 2002; von

Mering et al., 2002). With the advent of genome-wide analysis, however, we can use data from

a larger number of genes to predict the identity of additional functionally significant genes with

relative ease (Yu et al., 2006). We note that the novel gene prediction rate ranged from as high

as 41.2% from the *hpo[RNAi]* Ovariole Number module to as low as 0% from the Egg Laying

module (Figure 6b; Tables S4, S5). We suggest that this may be due to multiple factors. Firstly,

the possible incompleteness of the PPI is expected to lead to some areas of the network being

sparse or non-existent (von Mering et al., 2002). If the module of interest happens to fall in such

regions of the network, prediction algorithms will fail. Secondly, the initial restriction of tested

genes to signaling pathway members might have provided a seed list too sparse to usefully

predict functional connectors. Finally, it could be the case that "Egg Laying" is such a complex

phenotype that its gene regulation cannot be adequately captured within a highly connected

network of the type suited for identification by the analyses we have used here. Ovariole

number in *D. melanogaster* is the outcome of a discrete developmental process with a clear

beginning and end, comprising a specific series of cellular behaviors that take place in the

confines of one organ (Godt and Laski, 1995; Hodin and Riddiford, 2000a; Sahut-Barnola et al.,

1996). Once established during larval life, ovariole number in *Drosophila* remains unaltered

through to and during adulthood, even if oogenesis within those ovarioles suffers congenital or

age-related defects (King, 1970). Because previous work suggested that ovariole number in

Drosophila could have at least some predictive relation to egg laying (Cohet and David, 1978;

Klepsatel et al., 2013b; Sarikaya and Extavour, 2015), we reasoned that scoring the latter

phenotype in a primary screen (Figure 1a) could be an effective way to uncover ovariole number

regulators (Figure 1c). While our results showed that this was true in many cases, it was also

clear that these two traits can vary independently (Figure 2), highlighting the fact that ovariole

number is not the only determinant of egg laying. Egg-laying dynamics, even during the limited

five day assay used in our study, are likely influenced not just by a single anatomical parameter

such as ovariole number, but rather by many biological, biomechanical, hormonal and

behavioural processes. Consequently, the functional module we were able to extract from the

results of this screen (Figure 1b) might be too coarse to extract novel genes that participate in

potentially complex gene interactions regulating egg laying.

The predictive rates of the approach we have used here, although encouraging, are likely

limited by the degree of noise in the high throughput data used to generate the PPI (Li et al.,

2010), the sparseness of the PPI, and the degree of misidentification of protein interactions

(Zhang et al., 2017). Addressing one or more of these parameters could improve the outcomes

of future network predictions from developmental genetics data. For example, the problem of

sparseness, which is a paucity of high confidence detectable interactions relative to all

biologically relevant interactions, has been addressed in other studies by using an "Interolog

PPI" (Matthews et al., 2001) in place of an organism-specific PPI. The Interolog PPI combines

known interactions from multiple organisms, and has been used successfully to identify, for

example, gene modules relevant in squamous carcinoma, based on a starting dataset of

microarray data on differentially expressed genes between cancer cells and the surrounding

tissue (Wachi et al., 2005). Future studies applying the Interolog PPI to the outcomes of genetic

screens for developmental processes of interest could potentially overcome the problem of

sparseness, as well as the biases towards proteins that are more heavily studied and thus

better represented in organism-specific PPIs.

Tables

Table 1

Signalling pathway	Number of genes in screen
EGF	45
FGF	25
FOXO	67
Hippo	60
JAK/STAT	31
JNK	28
MAPK	29
Notch	48
SHH	54
TGF B	52
Toll	36
VEGF	17
Wnt	125
mTOR	36

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531 **Table 1: Number of candidate genes tested in each signalling pathway.** Candidate genes
532 are grouped by their reported roles in one or more signalling pathways based on published
533 literature. Genes in this list are not necessarily unique to a single pathway, but may function in
534 more than one signalling pathway. The list of specific genes per pathway that were included in
535 the screen for functional analysis (Figure 1) is found in Table S1.

Table 2

Egg Laying Screens	*hpo[RNAi]* Egg Laying (Figure 1a)	Egg Laying (Figure 1b)	Ovariole Number Screen	*hpo[RNAi]* Ovariole Number (Figure 1c)
RNAi stocks unavailable	12	0	RNAi stocks unavailable	0
Primary filter ($|Z_{gene}| < 1$)	190	N/A	Primary filter ($|Z_{gene}| < 1$)	N/A
No effect ($-5 < |Z_{gene}| < 5$)	214	224	No effect ($-2 < |Z_{gene}| < 2$)	206
Negative effect ($Z_{gene} < -5$)	**48**	**44**	Negative effect ($Z_{gene} < -2$)	**54**
Positive effect ($Z_{gene} > 5$)	**11**	**5**	Positive effect ($Z_{gene} > 2$)	**13**
Total	**475**	**273**	**Total**	**273**

Table 2: Results of the three functional genetic screens. Number of genes tested in each screen and cumulative results.
"Negative effect" corresponds to a reduction in eggs laid or number of ovarioles below the Z score (Z_{gene}) threshold for each
phenotype. "Positive effect" indicates an increase above the set Z_{gene} thresholds. Z_{gene} thresholds for each category in each screen
are indicated in brackets. The primary filter of $|Z_{gene}| < 1$ was applied only to the *hpo[RNAi]* Egg Laying screen shown in Figure 1a.
The list of specific genes that exceeded our chosen Z_{gene} thresholds for each scored phenotype (Figure 1), and were therefore
considered to have a positive or negative effect on the phenotype, is found in Table S1. The 12 genes for which RNAis stocks were
unavailable at the time of testing are listed in Table S2.

548 Figure 1

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Figure 1. Screen methodology. a,b,c) Diagrammatic representation of screen workflow. **d**)
Distributions of results of genes in the three screens. n = number of genes tested in each
screen (see also Table 2). **e**) Total eggs laid by three female flies over five days (left panel) and
ovariole number (right panel) of Oregon R (top row), *tj:Gal4* driving one copy of *UAS:hpo[RNAi]*
(middle row), and *tj:Gal4* driving two copies of *UAS:hpo[RNAi]* (bottom row), showing that, the
previously reported *tj:Gal4>hpo[RNAi]* ovariole number and egg laying phenotypes (Sarikaya
and Extavour, 2015) can be modified by further UAS:RNAi-mediated gene knockdown.

Figure 2

Figure 2. Relationship between Egg Laying and Ovariole Number phenotypes generated in the screens. a) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying versus the ovariole number of adult *tj>hpo[RNAi], candidate[RNAi]* females. **b**) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying of adult *tj>candidate[RNAi]* females versus the ovariole number of adult *tj>hpo[RNAi], candidate[RNAi]* females. **c**) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying of adult *tj>candidate[RNAi]* females versus egg laying of adult *tj>hpo[RNAi], candidate[RNAi]* females. In **a**, **b** and **c**, bar graphs on the top and right sides of each panel show the distribution of genes in each axis of the adjacent scatter plots . Green dots = genes that meet the Z_{gene} threshold for the indicated phenotype. Grey dots = genes that do not meet the Z_{gene} threshold for the indicated phenotype. Dark grey dotted lines = thresholds for each phenotype: $|Z_{gene}| > 5$ for Egg Laying and $|Z_{gene}| > 2$ for Ovariole Number. In **a** and **c**, the white vertical bar removes all genes in the *tj>hpo[RNAi], candidate[RNAi]* with a $|Z_{gene}| < 1$ for egg laying. These genes were not measured in the other two conditions and are therefore not represented in the scatter plots. **d**) Correlation between non-zero Ovariole Number and Egg Laying values.

Figure 3

Figure 3. Enrichment of genes of individual signalling pathways among the
experimentally obtained positive candidates of each screen. a) Depletion/enrichment
analysis to identify over- or under- represented members of individual signalling pathways
among positive candidates of each screen. Positive Z scores represent an enrichment, and
negative Z scores represent depletion, of genes of a pathway among those genes that
experimentally affected the phenotype Enrichment and depletion are defined relative to a null
distribution of the expected number of members of a signalling pathway among a group
containing the same number of randomly selected signalling genes. **b)** Fold enrichment and
hypergeometric p-value calculation to identify over- or under-representation of the genes of a
pathway in each screen. Significantly enriched pathways (colored bars: brown = Hedgehog;
pink = Notch) are defined by having a hypergeometric p-value less than 0.05.

Figure 4

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Figure 4. Screened genes function as a network. a) Receiver operating characteristic (ROC)
curves of genes ordered by rank for each of four network centrality metrics (Betweenness
centrality, Eigenvector centrality, Closeness centrality and Degree centrality) versus a binary
outcome (above or below Z score threshold) for each of the three screens. For each screen and
metric, the Area Under the Curve (AUC) is > 0.5 (Table S3). **b)** Genes whose $|Z_{gene}|$ value was
above the threshold (green dots; Table 2) in all three screens were assigned to the Core seed
list. **c)** Genes whose $|Z_{gene}|$ value was above the threshold (green dots; Table 2) in each screen
were assigned to the corresponding seed list.

Figure 5

Figure 5. Representation of Seed Connector Algorithm and output. a) Schematic
representation of the seed connector algorithm. The algorithm initializes by creating a
subnetwork of seed genes from the PPI, and computes the Largest Connected Component
(LCC) and coverage (number of genes from the seed set in the LCC). At each iteration, genes
in the direct neighborhood of the LCC (distance = 1) are added one at a time to the seed set,
and the coverage and LCC are recomputed. This process is repeated for each gene in the direct
neighborhood, each time restarting from the seed set of the preceding iteration. If any gene
outside the seed set but in the direct neighbourhood is found to maximize coverage while
minimizing the LCC, it is added to the seed set as a connector gene. Black arrows indicate the
path taken by the algorithm for which the criteria of maximal coverage and minimal LCC are
met; such a path would be used to proceed to the subsequent iteration. Grey arrows indicate
paths that fail to meet these criteria; such paths would be disregarded. The iteration repeats
until the coverage cannot be increased; in this schematic example, this state is achieved in
iteration 3. **b)** The Core Module generated by the Seed Connector Algorithm (SCA) based on
the results of the genetic screens (Figure 1a-c).The size of the shapes indicate the relative Z_{gene}
score of the gene. Circles indicate seed genes (functionally tested in the screen; Table 2; Table
S1) while triangles are connector genes (novel predicted genes; Tables S1, S4, S5). Green =
genes with a positive Z_{gene} score above the threshold; red = genes with a negative Z_{gene} score
above the threshold; grey = genes with Z_{gene} values below the threshold.

Figure 6

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Figure 6. Positive prediction rates of the connector genes in each of the four modules. a)
Proportion of core module connector genes with $|Z_{gene}|$ above the threshold in each of the three
screens. The "All phenotypes" category includes the genes with $|Z_{gene}|$ above the threshold in all
three screens. **b)** Proportion of tested connector genes in each of the three modules with $|Z_{gene}|$
above the threshold within their corresponding screen. **c)** Proportion of all unique connector
genes predicted by all four modules with $|Z_{gene}|$ above the respective threshold in any of the
three screens. **d)** Proportion of positive candidate genes emerging from the three original
signalling candidate screens with $|Z_{gene}|$ above the threshold relative to the total number of
genes tested in each screen.

Figure 7

Figure 7. Phenotypically separable sub-networks formed by analysis of the combined genes from all modules. The meta network is generated by the union of the genes in the four phenotypic modules: *hpo[RNAi]* Egg Laying (Figure S6a), Egg Laying (Figure S6b), *hpo[RNAi]* Ovariole Number (Figure S6c) and Core (Figure 5b). **a)** The meta network is represented as a Venn diagram, in which each grey dotted outline represents the screen in which a given gene was identified as affecting the scored phenotype. Within each sub-network, grey circles indicate seed genes, and blue circles indicate connector genes. A single gene, *sloppy paired 1*, was a seed in the Egg Laying module and also a connector in the *hpo[RNAi]* Egg Laying module; it fell within sub-network VII in the meta network, and is marked as a seed (grey) in this figure. Solid grey lines indicate interactions between genes in the meta network from the PPI. **b)** Edge densities between the seven sub-networks of the meta-network. **c)** Relative enrichment of screened members of the 14 tested developmental signalling pathways within the seven sub-networks of the meta-network.

647 # Figure S1

a. *hpo[RNAi]* Egg Laying

b. Egg Laying

c. *hpo[RNAi]* Ovariole Number

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650 **Figure S1. Violin plots of egg laying and ovariole number of controls in each screen**
651 **batch. a)** Distribution of number of eggs laid by five replicates of three *tj:Gal4>hpo[RNAi]*
652 females over five days for each batch. **b)** Distribution of number of eggs laid by five replicates of
653 three *tj:Gal4* females over five days for each batch. **c)** Distribution of number of ovarioles per
654 ovary in 20 ovaries from ten *tj:Gal4>hpo[RNAi]* females in each batch.

Figure S2

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Figure S2. Biological aspects of the network modules. a) Enrichment/depletion analysis of
the 273 signalling pathway genes above the threshold $|Z_{gene}| > 1$ (Figure 1a) against all
signalling candidates. We also measured the enrichment/depletion of positive signalling
candidate genes in the *hpo[RNAi]* Ovariole (Figure 1c) and Egg Laying (Figure 1b) screens from
the 273 genes tested in those screens. **b)** Signalling pathway depletion enrichment analysis. For
each module, a null distribution of the expected number of members of a signalling pathway
from a group of the same number of randomly selected signalling pathway genes was
calculated. The Z score from the expected distribution was then calculated. Negative Z scores
represent a depletion, while positive Z scores represent an enrichment. No single pathway is
enriched in any of those modules. **c)** Fold enrichment and hypergeometric p-value calculation
for each pathway in the four modules. Pathway members in color (orange = mTor; brown =
Hedgehog; pink = Notch) have a p-value < 0.05.

Figure S3

Figure S3. Comparison of egg laying candidate genes by pathway. Z_{gene} of egg laying of
adult females of *tj>hpo[RNAi],candidate[RNAi]* plotted against Z_{gene} of egg laying of
tj>candidate[RNAi] adult females displayed by pathway. Contour plots indicate a 2D gaussian
kernel density estimation.

Figure S4

Figure S4. Box plots of the four centrality measures calculated for the genes in each of the four phenotypic modules. See modules in Figures 5 and S6.

Figure S5

Figure S5. Comparisons of the Z_{gene} scores of the positive candidate genes sorted by centrality metrics. In each screen (a, b, c), the $|Z_{gene}|$ values of the first (dark grey) and fifth (light grey) quintiles of positive candidate genes ordered by rank for each of the four chosen centrality metrics, are plotted as a bar plot. Bars indicate standard error.

Figure S6

Figure S6. Three modules generated by the Seed Connector Algorithm (SCA). a)
hpo[RNAi] Egg Laying Module **b)** Egg Laying Module **c)** *hpo[RNAi]* Ovariole Number Module.
The size of the shapes indicate the Z_{gene} score of the gene. Circles = seed genes; triangles =
connector genes. Green = genes with a positive Z_{gene} above the threshold. Red = genes with a
negative Z_{gene} above threshold. Grey = genes with Z_{gene} values below the threshold. All
connectors were phenotypically tested (Table S1) except *eukaryotic translation initiation factor 3*
subunit j (*eIF3J*), in the *hpo[RNAi]* Egg Laying Module (black triangle), for which no RNAi stock
as available at the time of testing.

Figure S7

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**Figure S7. Comparison of network metrics before (Seed Network) and after (Module
Network) application of the seed connector algorithm. a)** Comparison of network metrics of
all screened genes (red line) to a null distribution of network metrics derived by bootstrapping an
equal number of randomly selected genes in the PPI (grey curve). **b)** Comparisons of the
Largest Connected Component (LCC), network density, number of edges and average shortest
path between the seed network (light red line) and the module network (dark red line). The
bootstrapped null distribution (1000 bootstraps) of both the seed network (light grey curve) and
the module network (dark grey curve) are indicated..

Supplementary tables

Table S1

Table S1: Tabulation of raw data and analysis for every gene in the screen.
https://github.com/extavourlab/hpo_ova_eggL_screen/blob/master/Results/MasterTable.csv.

This table contains a summary representation of the data generated by the three screens as well as results from the analysis. Each line corresponds to an independent measurement of a particular RNAi line. Some genes which did not pass the first filter of $|Z_{gene}| > 1$ in the *hpo[RNAi]* Egg Laying screen where then predicted as connectors, therefore they have two entries as they have been independently measured again. The Z scores have been rounded up to 4 significant digits in this table and the Centrality metrics rounded up to 10 significant digits due to their low values, but the full values for both are available in the raw data files provided in the supplementary files in Data/Screens for the Z scores and Results for the centrality values. Moreover this is a summary table and does not contain values for controls as well as batch numbers, all are available in the supplementary files in Data/Screens.

- **FbID**: FlyBase ID of the tested gene.
- **CG number**: CG Number of the tested gene.
- **NAME**: Common name (as per FlyBase nomenclature) of the gene if existing, else it is a -.
- **SYMBOL**: Symbol (as per FlyBase nomenclature) of the gene if existing, else CG number
- **[*ScreenName*]_[*Variable*]_(*Metric*)_Count**: Within the screen [*ScreenName*], the count of the measured variable [*Variable*]. Optional: (*metric*) will indicate if a particular operation was done over the data, such as sum, mean or standard deviation.
e.g. [HippoRNAi_EggL]_[Day_4_Egg]_Count is the count of eggs, on day 4, of the *hpo[RNAi]* Egg Laying screen.
- **[*ScreenName*]_[*Variable*]_(*Metric*)_Zscore**: Within the screen [*ScreenName*], the Z score of the measured variable [Variable] as calculated to batch control. Optional: (*metric*) will indicate if a particular operation was done over the data, such as sum, mean or standard deviation.
e.g. [EggL]_[All_Days_Egg]_(Sum)_Zscore is the Z score of the sum of eggs count, of the Egg Laying screen.
- **PPI_[*Metric*]_centrality**: Within the PPI used in this paper, the calculated centrality value for the metric [*Metric*].
- **[*ModuleName*]_Network**: Presence of absence of a gene in the module [*ModuleName*]. If the gene is in the module, this value is True, if it is absent it is False. (An exception is made for the Meta Network displayed in Figure 7 where instead of True/False, the group assignment I-VII is written)
- **[*ModuleName*]_Connector**: Status of a gene in the module [*ModuleName*] as a connector. If True, the gene is a connector, else if False, the gene is not a connector.
- **[*PathwayName*]_Pathway**: Participation of a gene to the signalling pathway [*PathwayName*]. If the gene participates in the pathway the value is 1, else it is 0.

749 # Table S2

FbID	CG Number	Name	Symbol
FBgn0283468	CG3412	*supernumerary limbs*	*slmb*
FBgn0267821	CG5102	*daughterless*	*da*
FBgn0266724	CG5161	*TRAPP subunit 20*	*Trs20*
FBgn0267378	CG7085	*sauron*	*sau*
FBgn0267487	CG9181	*Protein tyrosine phosphatase 61F*	*Ptp61F*
FBgn0267912	CG9819	*Calcineurin A at 14F*	*CanA-14F*
FBgn0086371	CG9829	*poly*	*poly*
FBgn0267350	CG10260	*Phosphatidylinositol 4-kinase III alpha*	*PI4KIIIalpha*
FBgn0267698	CG10295	*p21-activated kinase*	*Pak*
FBgn0283462	CG18279	*Immune induced molecule prepropeptide*	*IMPPP*
FBgn0267339	CG33338	*p38c MAP kinase*	*p38c*
FBgn0085506	CG40635	-	*CG40635*

751 **Table S2. 12 signalling candidate genes with no available RNAi lines at either BDSC or**
752 **VDRC at the time of this study.**

754 Table S3

Centrality metric	*hpo[RNAi]* Ovariole Number	*hpo[RNAi]* Egg Laying	Egg Laying
Betweenness	0.603	0.57	0.586
EigenVector	0.632	0.573	0.586
Closeness	0.612	0.551	0.588
Degrees	0.615	0.592	0.599

Table S3: Area under the curve (AUC) of ROC curves. AUC values for the ROC curves for
each centrality measure for the three screens (Figure 4a). AUC values range from 0 to 1. A
score above 0.5 indicates a positive correlation between the continuous variable (centrality) and
the binary variable (above or below the Z score threshold). A score of 0.5 or less indicates no
correlation between the variables.

Table S4

Module	Number of Seeds	Number of Connectors	Number of connector genes above $|Z_{gene}|$ threshold within module phenotype
hpo[RNAi] Egg Laying	58	18	7 (41.2%)
Core	27	10	1 (10.0%)
Egg Laying	49	11	0 (0.0%)
hpo[RNAi] Ovariole Number	66	11	3 (27.3%)

Table S4. Distribution of seed genes and connectors in each module. Two genes that were
above $|Z_{gene}|$ threshold (Table 2) in the *hpo[RNAi]* Egg Laying (CG12147) and *hpo[RNAi]*
Ovariole Number seed list (CG6104) were not found in the PPI, and therefore not included in
the network analysis or in this table (see methods for details). The removal of these two genes
accounts for the difference between the number of positive candidates in Table 2 and the
number of seed genes in these two modules (Table S1 and S4). The proportion of connectors
whose loss of function produced a significant phenotype ($|Z_{gene}|$ above threshold) is in
parentheses and plotted in Figure 6a, 6b). All connectors except *eukaryotic translation initiation*
factor 3 subunit j (*eIF3J*) in the *hpo[RNAi]* Egg Laying Module, for which no RNAi line was
available at the time of testing, were tested. Therefore, the percentages of connectors above the
threshold for the *hpo[RNAi]* Egg Laying Module were calculated out of 17 connectors.

Table S5

Total number of unique connectors in all four modules	Number of connector genes above $|Z_{gene}|$ threshold		
	For Egg Laying Phenotype	**For *hpo[RNAi]* Egg Laying Phenotype**	**For *hpo[RNAi]* Ovariole Number Phenotype**
43	10 (23.8%)	13 (31.0%)	14 (33.3%)

Table S5. Number of unique connector genes above $|Z_{gene}|$ threshold for the three

phenotypic measurements. Percentage of the number of connectors above threshold for each

phenotype from the total number of connectors is in parentheses and plotted in Figure 6c. All

connectors except *eukaryotic translation initiation factor 3 subunit j* (*eIF3J*) in the *hpo[RNAi]* Egg

Laying Module, for which no RNAi line was available at the time of testing, were tested.

Therefore, the percentages of connectors above the threshold were calculated out of 42 unique

connectors.

Methods

LEAD CONTACT AND MATERIALS AVAILABILITY

This study did not generate new unique reagents. This study generated new python3 code available on GitHub: https://github.com/extavourlab/hpo_ova_eggL_screen.

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Cassandra G. Extavour (extavour@oeb.harvard.edu).

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Wild type and mutant lines of *Drosophila melanogaster* were obtained from publicly accessible stock centers and maintained as described in "Fly Stocks" below. Genotypes and provenance are provided in the Key Resource Table. Candidate genes were randomly assigned to batches for screening (see Table S1 for which genes were in each batch). F1 animals from the same cross were randomly assigned to experimental groups for phenotyping in all screens.

METHOD DETAILS

Fly stocks

Flies were reared at 25°C at 60% humidity with standard *Drosophila* food (Sarikaya et al., 2012) containing yeast and in uncrowded conditions as previously defined (Sarikaya and Extavour, 2015). RNAi lines were obtained from the TRiP RNAi collection at the Bloomington *Drosophila* Stock Centre (BDSC) and from the Vienna *Drosophila* Resource Centre (VDRC). See Key Resources Table for complete list of stocks used in this study. Oregon R was used as a wild type strain. The genotype of the *traffic jam:Gal4* line used in the screen was *y w; P{w[+mW.hs] = GawB}NP1624* (Kyoto Stock Center, K104–055; abbreviated hereafter as *tj:Gal4*). The *hippo* RNAi line used in the screen was *y[1] v[1]; P{y[+t7.7]v[+t1.8]=TRiP. HMS00006}attP2* (BDSC:33614; abbreviated hereafter as *hpo[RNAi]*).

Egg and ovariole number counts

Adult egg laying was quantified by crossing three virgin females of the desired genotype (see "*Screen design*" below) with two males in a vial containing standard food and yeast granules (day one) and then transferring them into a fresh food vial without yeast granules for a 24 hour period. Eggs from vials were then counted by visual inspection of the surface of the food in the vial. Males and females were transferred to fresh food vials without yeast granules, every day thereafter until day six. All egg laying measurements reported and analysed in the paper are the sum of the eggs laid by three adult female flies over the five days of this assay (days two

819 through six without yeast granules). Data from any vial in which either a female or male died,
820 during the course of the experiment, were not included in the analysis.
821
Ovariole number was quantified by mating ten virgin adult females with five virgin adult Oregon
R males for three days post eclosion in vials with yeast at 25°C and 60% humidity. After this
three-day mating period, all 20 adult ovaries from the mated females were dissected in 1X PBS
with 0.1% Triton-X-100 and stained with 1ug/ml Hoechst 33321 (1:10,000 of a 10mg/ml stock
solution). Ovarioles were separated from each other with No. 5 forceps (Fine Science Tools)
and counted by counting the number of germaria under a ZEISS Stemi 305 compact stereo
microscope with a NIGHTSEA stereo microscope UV Fluorescence adaptor.

Screen design

In the primary screen (Figure 1a: *hpo[RNAi]* Egg Laying), 463 candidate genes (Table S1) were
screened for the effect of an RNAi-induced loss of gene function in a *hpo[RNAi]* background on
the number of eggs laid in the first five days of mating (see "*Egg and ovariole number counts*"
above) by adult females. These females were the F1 offspring of UAS:*candidate gene* RNAi
males crossed to *P{w[+mW.hs] = GawB}NP1624; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2*
(*tj:Gal4; UAS:hpo[RNAi]*) virgin adult females (Figure 1a: *hpo[RNAi]* Egg Laying). All genes that
yielded an egg laying count with a $|Z_{gene}| > 1$ (see "*Gene selection based on Z score and batch*
standardization" below) were selected to undergo two secondary screenings (n=273, Table 2,
Figure 1d). First, these genes were screened for effects on the egg laying of mated adult female
offspring from a cross of *UAS:candidate gene[RNAi]* males and *tj:Gal4* virgin adult females
(Figure 1b: Egg Laying). Secondly, these genes were screened for effects on ovariole number in
a *hpo[RNAi]* background. All 20 ovaries from ten adult female F1 offspring of a cross between
UAS:candidate gene[RNAi] males to *P{w[+mW.hs] = GawB}NP1624; P{y[+t7.7]*
v[+t1.8]=TRiP.HMS00006}attP2 (*tj:Gal4; UAS:hpo[RNAi]*) virgin adult females were scored for
ovariole number (see "*Egg and ovariole number counts*" above). (Figure 1c: *hpo[RNAi]* Ovariole
Number).

Gene selection based on Z score and batch standardization

Candidate genes were screened in batches with an average size of 50 genes. For each batch,
control flies were the female F1 offspring of Oregon R males crossed to *P{w[+mW.hs] =*
GawB}NP1624; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2 (*tj:Gal4; UAS:hpo[RNAi]*) virgin adult
females. Because the control group in each batch had slightly different distributions of egg
laying and ovariole number values (Figure S1), it was inappropriate to compare absolute mean
values between genes that were scored in different batches. Instead, comparisons of the Z
score of each candidate (Z_{gene}) to its batch control group was used as a discriminant. This
approach standardizes for batch effects and allows the comparison of all genotypes within and
across the primary and secondary screens with a single metric (Z_{gene}).
856
Firstly, the mean and standard deviation of the eggs laid by the control genotype for a batch
were calculated as μ_b and σ_b respectively. Then, using the number of eggs laid by adult females

of a candidate gene RNAi (x_{gene}) of the same batch, the Z score for the egg laying count of that

gene (Z_{gene}) was calculated as $Z_{gene} = \frac{x_{geme} - \mu_b}{\sigma_b}$. The same standardization protocol was

applied to both egg laying and ovariole number counts of every gene and its corresponding

batch control.

Ovariole numbers were derived from counts of the number of ovarioles per ovary for 20 ovaries

per candidate gene, and a threshold of $|Z_{gene}| > 2$ (corresponding to a false positive probability

less than 0.045) was applied for ovariole number phenotype. Egg laying counts were derived

from measurements of three females in a single vial per gene. We therefore chose to be more

conservative in our Z score comparisons for the egg laying phenotype, than for ovariole number

phenotype, and applied a stringent threshold of $|Z_{gene}| > 5$ (corresponding to a false positive

probability less than 0.00006) to select genes of interest. All genes with $|Z_{gene}|$ values above

these thresholds are referred to throughout the study as "positive candidates". (See Ipython

notebooks 02_Z_score_calculation.ipynb and 02.2_Z_score_calculation_prediction.ipynb for

code implementation and calculation of Z scores, and 06_Screen Analysis.ipynb for batch

effects.)

875 Signaling pathway enrichment analysis

To study the enrichment of a particular signaling pathway in a group of candidate genes that

had similar phenotypic effects revealed by the screen, custom scripts (see

07_Signaling_pathway_analysis.ipynb for code implementation) were generated to implement

two different methods (Figure 3a, 3b; Figure S52a-c).

The first method is a numerical method that uses bootstrapping to calculate the null distribution

of the number of members (M) of a signaling pathway (S) that would be expected at random in a

set of genes of size (N). The script randomly sampled N genes from among the 463 tested *D.*

melanogaster signaling genes 10,000 times, and counted the number of genes (M) that were

members of the signaling pathway S. Positive candidates in each of the three screens were

sorted by their presence in signalling pathways and counted. The Z score was then calculated

by comparing the experimentally observed number of positive candidates in each signaling

pathway against the bootstrapped null distribution.

The second method used the hypergeometric p-value to calculate the probability of M members

of a signaling pathway being in a group of N genes, given a starting population of 463 tested *D.*

melanogaster signaling genes, and the known attribution to a pathway S of each gene.

893 Protein-Protein Interaction Network (PPI) building

There is no standard complete Protein-Protein Interaction (PPI) network available for *Drosophila*

melanogaster. However, there exist many smaller networks from different screens, as well as

literature extractions. We therefore combined data from these sources and then created a PPI

for use in the present study, as follows:

Step 1: Several screens assessing protein-protein interactions have been centralized in a
database called DroID: http://www.droidb.org. The version DroID_v2018_08 was used. All
available datasets were first downloaded from that database using this link:
http://www.droidb.org/Downloads.jsp. The description of all of these datasets can be found here:
http://www.droidb.org/DBdescription.jsp

Step 2: We used the datasets from all screens that assessed direct protein-protein interactions
and did not use the interolog database (predicted protein interaction based on mouse human
and yeast PPI). These direct assessment screens were seven in total, as follows:

○ Finley Yeast Two-Hybrid Data (size 2.0 MB)
○ Curagen Yeast Two-Hybrid Data (size 4.6 MB)
○ Hybrigenics Yeast Two-Hybrid Data (size 381 KB)
○ Perrimon co-AP complex (size 108 KB)
○ DPiM co-AP complex (size 6.3 MB)
○ PPI from other databases (size 16.2 MB)
○ PPI curated by FlyBase (size 7.4 MB)

An important element to note is that the PPI curated by FlyBase is a literature-based PPI.
FlyBase protein-protein interactions are experimentally derived physical interactions curated
from the literature by FlyBase, and does not include FlyBase-curated genetic interactions.

Step 3: We concatenated the seven datasets listed above into a single unique database. A
custom python script was created that downloads and reads each of the above seven unique
PPI tables, and generates a single PPI network. From this concatenation, a single edge
undirected network was created and saved. This network is hereafter referred to as **the PPI**
(see 01_PPI_builder.ipynb).

Network metric computations

The centrality of a node is often used as a measure of a node's importance in a network. Within
a PPI, the centrality of a gene reflects the number of interactions in which the gene directly or
indirectly participates. Four different centrality metrics were computed for all genes in the PPI
using the networkx python library:

(1) **Betweenness** reflects the number of shortest paths passing through a gene.
(2) **Eigenvector** is a measure of the influence of a gene in the network.
(3) **Closeness** measures the sum of shortest distance of a gene to all the other genes.
(4) **Degree centrality** corresponds to the normalized number of edges of a gene in the network.

While there exist more centrality measures, these four are commonly used to assess biological
networks. These computed centrality parameters of the genes measured in the screen were

computed with 03_ROC_curve_analysis_of_network_metrics.ipynb, and are reported in the
Table S1 (see 09_Making_the_database_table.ipynb).

Receiver Operating Characteristic (ROC) curves

To check whether the centrality of a gene in the network could predict the phenotypic effect
produced by RNAi against that gene, ROC curves were plotted for the four aforementioned
centrality measures of each gene in each screen. A ROC analysis is used to measure the
correlation between a continuous variable (centrality) and a binary outcome (above or below Z
score threshold). Therefore, for each screen, measured genes were rank-ordered from high
centrality to low centrality, and plotted against the binary outcome of $|Z_{gene}|$ being above or
below the appropriate |Z score| threshold (>5 for egg laying and >2 for ovariole number). The
Area Under the Curve (AUC) measures the extent of correlation between centrality and effect of
a gene on measured phenotype. AUC above or below 0.5 indicates a positive or negative
correlation respectively, while an AUC of 0.5 indicates no correlation of the parameters. The
scikit learn python package was used to calculate the AUC of each ROC curve plotted (see
03_ROC_curve_analysis_of_network_metrics.ipynb).

Building the network modules

Network modules were built using the previously published Seed-Connector algorithm (SCA)
(Wang et al., 2017; Wang and Loscalzo, 2018), implemented here in python (see 04_Seed-
Connector.ipynb) and illustrated in Figure 5a. Creating a module using the SCA requires a list of
seed genes and a PPI. From each of the three screens, we selected the genes whose $|Z_{gene}|$
value was above the threshold and created three seed lists respectively (Figure 4c: Egg laying,
hpo[RNAi] egg laying and *hpo[RNAi]* ovariole 'seed' list). A fourth list consisting of the
intersection of the aforementioned seed lists was also collated and called the core 'seed' list
(Figure 4b). Genes were assigned in the core list if they passed the Z threshold in all 3 screens.
The Seed-Connector algorithm was then executed on each of these seed lists using the PPI.
Not all genes in the four seed lists were found in the PPI network (specifically, CG12147 in the
hpo[RNAi] Egg Laying seed list and CG6104 in the *hpo[RNAi]* Ovariole number seed list were
absent from the PPI) and were therefore eliminated from further network analysis. The removal
of these two genes accounts for the variation in the number of positive candidates in Table 2
and the number of seed genes in the module. Modules were obtained for each seed list (Figure
5b; Figure S6) consisting of the seed genes (circles in Figure 5b and Figure S6) and previously
untested genes added by the SCA (squares in Figure 5b and Figure S6) to increase the LCC
size that we refer to as connector genes (see 04_Seed-Connector.ipynb). The results of the
algorithm are summarized in Table S1.
973
The modularity of the subnetworks was then assessed using four network metrics namely
Largest Connected Component (LCC), number of edges, network density and average shortest
path in the LCC. Each metric for each module was assessed using distance of the network
metric to a null distribution. Initially, the null distribution was calculated by taking 1000 samples
of 463 genes randomly selected from the PPI and calculating the above metrics. We found that

the 463 genes selected in the signalling screen were already more connected than the null
distribution of sets of 463 genes randomly selected from the PPI (Figure S7a). Therefore, to
avoid a false positive detection of modularity, the four experimentally obtained subnetworks
were compared to null distributions obtained by randomly sampling an equal number of genes
from the 463 signalling candidate genes selected for our screen. For each of the four modules,
comparison of the metrics was performed on the seed lists and the sub-network after the SCA.
Most metrics were enriched in the seed group when compared to the null distribution with the
exception of the Average shortest path (Figure S7b, light red line). The sub-networks obtained
from the SCA further increased all four metrics suggesting the modularity of the four sub-
networks (Figure S7b, dark red line; see 05_Network_Module_testing.ipynb for code
implementation).

Meta network

To build the meta network, the genes from all four modules were concatenated into one
network. The network was then visually sorted in an approach akin to projecting the network on
a Venn Diagram. The meta network was sorted by which of the three screens the gene was
positive in. The intersections were genes whose $|Z_{gene}|$ value was above the threshold in more
than one and possibly all three of the screening paradigms. For example, if a gene was found in
the *hpo[RNAi]* Ovariole Number and Egg Laying module it is then assigned to the dual positive
group *hpo[RNAi]* Ovariole Number / Egg Laying (Figure 7a, module VI). After applying this
grouping strategy, the connectivity across the groups was studied by calculating the edge
density between all groups ($density = \frac{Edges_{1;2}}{Nodes_1 * Nodes_2}$). Finally, the proportion of each signaling
candidate in each of those groups was calculated by taking the number of members of a
signaling pathway divided by the total members of a group (see Ipython notebook
08_MetaModule_Analysis.ipynb).

QUANTIFICATION AND STATISTICAL ANALYSIS

Number of samples

1005 The number of samples across the different screens were as follows:

hpo[RNAi] Egg Laying and Egg Laying screens

1007 - Controls: five vials of three females and two males
1008 - Sample: one vial of three females and two males

hpo[RNAi] Ovariole number screen

1010 - Controls: 20 flies, two ovaries per fly considered as independent measurements
1011 - Sample: 10 flies, two ovaries per fly considered as independent measurements

Correction of batch effect

Despite best efforts to maintain the exact same condition between each experiment, some variation was measured between the batches. Control flies showed variations in both measured phenotypes, ovariole number and egg laying (Figure S1). In order to compare the values measured across different batches, each sample was standardized by calculating its Z score (Z_{gene}) to the control distribution. For each batch, the measurements for controls were pooled into a distribution, and the mean and standard deviation was computed. Then each sample was compared to its respective batch and its Z score computed (see "*Gene selection based on Z score and batch standardization*" for formula).

Statistical analysis

All statistical analyses were performed using the scipy stats module (https://www.scipy.org/) and scikit learn (https://scikit-learn.org/). Significance thresholds for p-values were set at 0.05. Statistical tests and p-values are reported in the figure legends. All statistical tests can be found in the Ipython notebooks mentioned below.

DATA AND CODE AVAILABILITY

This study generated a series of python3 Ipython notebook files that perform the entire analysis presented in this study. All the results presented in this paper, including the figures with the exception of the network visualizations, which were created using Cytoscape3 (https://cytoscape.org/) can be reproduced by running the aforementioned python3 code. The raw data, calculations made with these data, and code used for calculations and analyses (Ipython notebooks) are available as supplementary information. For ease of access, legibility and reproducibility, the code and datasets have been deposited in a GitHub repository available at https://github.com/extavourlab/hpo_ova_eggL_screen.

1036 KEY RESOURCES TABLES

1037 Software and libraries

1038 All software and libraries used in this study are published under open source licenses and are
1039 therefore publicly available.
1040

Type	Name	Version	Source
Library	matplotlib	3.0.0	https://matplotlib.org/
Library	networkx	2.3	http://networkx.github.io/
Library	numpy	1.11.3	https://www.numpy.org/
Library	pandas	0.20.3	https://pandas.pydata.org /
Library	progressbar	3.38.0	https://github.com/niltonvolpato/python-progressbar
Library	scipy	1.1.0	https://www.scipy.org/
Library	seaborn	0.9.0	https://seaborn.pydata.org/
Software	Cytoscape	3.4.0	https://cytoscape.org/
Software	Inkscape	0.92.3	https://inkscape.org/
Software	Python3	3.7	https://www.python.org/

1041

1042 *Drosophila melanogaster* genetic lines

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Experimental Models: Organisms/Strains		
Description	Stock Center	IDs
D. melanogaster. Expresses dsRNA for RNAi of CanA1 (FBgn0010015) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01871}attP2	Bloomington Drosophila Stock Center	BDSC:25850; FlyBase:FBst0025850;
D. melanogaster. Expresses dsRNA for RNAi of fng (FBgn0011591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01967}attP2	Bloomington Drosophila Stock Center	BDSC:25947; FlyBase:FBst0025947;
D. melanogaster. Expresses dsRNA for RNAi of Aplip1 (FBgn0040281) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02049}attP2	Bloomington Drosophila Stock Center	BDSC:26024; FlyBase:FBst0026024;
D. melanogaster. Expresses dsRNA for RNAi of E(spl)mdelta-HLH (FBgn0002734) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02101}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:26203; FlyBase:FBst0026203;
D. melanogaster. Expresses dsRNA for RNAi of sima (FBgn0266411) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02105}attP2	Bloomington Drosophila Stock Center	BDSC:26207; FlyBase:FBst0026207;
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m8-HLH (FBgn0000591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02096}attP2	Bloomington Drosophila Stock Center	BDSC:26322; FlyBase:FBst0026322;
D. melanogaster. Expresses dsRNA for RNAi of pan (FBgn0085432) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02306}attP2	Bloomington Drosophila Stock Center	BDSC:26743; FlyBase:FBst0026743;
D. melanogaster. Expresses dsRNA for RNAi of CanB (FBgn0010014) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02616}attP2	Bloomington Drosophila Stock Center	BDSC:27307; FlyBase:FBst0027307;
D. melanogaster. Expresses dsRNA for RNAi of mib1 (FBgn0263601) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02629}attP2	Bloomington Drosophila Stock Center	BDSC:27320; FlyBase:FBst0027320;
D. melanogaster. Expresses dsRNA for RNAi of Nct (FBgn0039234) under UAS control in the VALIUM10	Bloomington Drosophila Stock Center	BDSC:27498; FlyBase:FBst0027498;

vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02648}attP2		
D. melanogaster. Expresses dsRNA for RNAi of Cbl (FBgn0020224) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02650}attP2	Bloomington Drosophila Stock Center	BDSC:27500; FlyBase:FBst0027500;
D. melanogaster. Expresses dsRNA for RNAi of Atg12 (FBgn0036255) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02704}attP2	Bloomington Drosophila Stock Center	BDSC:27552; FlyBase:FBst0027552;
D. melanogaster. Expresses dsRNA for RNAi of fz2 (FBgn0016797) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02722}attP2	Bloomington Drosophila Stock Center	BDSC:27568; FlyBase:FBst0027568;
D. melanogaster. Expresses dsRNA for RNAi of Psn (FBgn0284421) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02760}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27681; FlyBase:FBst0027681;
D. melanogaster. Expresses dsRNA for RNAi of Pi3K92E (FBgn0015279) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02770}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27690; FlyBase:FBst0027690;
D. melanogaster. Expresses dsRNA for RNAi of Cdk4 (FBgn0016131) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02795}attP2	Bloomington Drosophila Stock Center	BDSC:27714; FlyBase:FBst0027714;
D. melanogaster. Expresses dsRNA for RNAi of mts (FBgn0004177) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02805}attP2	Bloomington Drosophila Stock Center	BDSC:27723; FlyBase:FBst0027723;
D. melanogaster. Expresses dsRNA for RNAi of Pdk1 (FBgn0020386) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02807}attP2	Bloomington Drosophila Stock Center	BDSC:27725; FlyBase:FBst0027725;
D. melanogaster. Expresses dsRNA for RNAi of ds (FBgn0284247) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02842}attP2	Bloomington Drosophila Stock Center	BDSC:28008; FlyBase:FBst0028008;
D. melanogaster. Expresses dsRNA for RNAi of Hrs (FBgn0031450) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02860}attP2	Bloomington Drosophila Stock Center	BDSC:28026; FlyBase:FBst0028026;

D. melanogaster. Expresses dsRNA for RNAi of mam (FBgn0002643) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02881}attP2	Bloomington Drosophila Stock Center	BDSC:28046; FlyBase:FBst0028046;
D. melanogaster. Expresses dsRNA for RNAi of bun (FBgn0259176) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02954}attP2	Bloomington Drosophila Stock Center	BDSC:28322; FlyBase:FBst0028322;
D. melanogaster. Expresses dsRNA for RNAi of aos (FBgn0004569) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03020}attP2	Bloomington Drosophila Stock Center	BDSC:28383; FlyBase:FBst0028383;
D. melanogaster. Expresses dsRNA for RNAi of pcx (FBgn0003048) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05038}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28552; FlyBase:FBst0028552;
D. melanogaster. Expresses dsRNA for RNAi of Su(fu) (FBgn0005355) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05045}attP2	Bloomington Drosophila Stock Center	BDSC:28559; FlyBase:FBst0028559;
D. melanogaster. Expresses dsRNA for RNAi of Apc2 (FBgn0026598) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05073}attP2	Bloomington Drosophila Stock Center	BDSC:28585; FlyBase:FBst0028585;
D. melanogaster. Expresses dsRNA for RNAi of l(2)tid (FBgn0002174) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05082}attP2	Bloomington Drosophila Stock Center	BDSC:28594; FlyBase:FBst0028594;
D. melanogaster. Expresses dsRNA for RNAi of Su(H) (FBgn0004837) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05110}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28900; FlyBase:FBst0028900;
D. melanogaster. Expresses dsRNA for RNAi of PDZ-GEF (FBgn0265778) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05139}attP2	Bloomington Drosophila Stock Center	BDSC:28928; FlyBase:FBst0028928;
D. melanogaster. Expresses dsRNA for RNAi of wntD (FBgn0038134) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05158}attP2	Bloomington Drosophila Stock Center	BDSC:28947; FlyBase:FBst0028947;
D. melanogaster. Expresses dsRNA for RNAi of Cdk2 (FBgn0004107) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05163}attP2	Bloomington Drosophila Stock Center	BDSC:28952; FlyBase:FBst0028952;

D. melanogaster. Expresses dsRNA for RNAi of ci (FBgn0004859) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01715}attP2	Bloomington Drosophila Stock Center	BDSC:28984; FlyBase:FBst0028984;
D. melanogaster. Expresses dsRNA for RNAi of mgl (FBgn0261260) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02485}attP2	Bloomington Drosophila Stock Center	BDSC:29324; FlyBase:FBst0029324;
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m4-BFM (FBgn0002629) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03310}attP2	Bloomington Drosophila Stock Center	BDSC:29378; FlyBase:FBst0029378;
D. melanogaster. Expresses dsRNA for RNAi of Wnt4 (FBgn0010453) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03378}attP2	Bloomington Drosophila Stock Center	BDSC:29442; FlyBase:FBst0029442;
D. melanogaster. Expresses dsRNA for RNAi of Dif (FBgn0011274) under UAS control in the VALIUM10 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HM05257}attP2	Bloomington Drosophila Stock Center	BDSC:30513; FlyBase:FBst0030513;
D. melanogaster. Expresses dsRNA for RNAi of InR (FBgn0283499) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01482}attP2	Bloomington Drosophila Stock Center	BDSC:31037; FlyBase:FBst0031037;
D. melanogaster. Expresses dsRNA for RNAi of Cdc5 (FBgn0265574) and Roc1b (FBgn0040291) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01517}attP2	Bloomington Drosophila Stock Center	BDSC:31067; FlyBase:FBst0031067;
D. melanogaster. Expresses dsRNA for RNAi of Egfr (FBgn0003731) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01696}attP2	Bloomington Drosophila Stock Center	BDSC:31183; FlyBase:FBst0031183;
D. melanogaster. Expresses dsRNA for RNAi of norpA (FBgn0262738) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01713}attP2	Bloomington Drosophila Stock Center	BDSC:31197; FlyBase:FBst0031197;
D. melanogaster. Expresses dsRNA for RNAi of CenG1A (FBgn0028509) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01807}attP2	Bloomington Drosophila Stock Center	BDSC:31228; FlyBase:FBst0031228;
D. melanogaster. Expresses dsRNA for RNAi of dsh (FBgn0000499) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01254}attP2	Bloomington Drosophila Stock Center	BDSC:31307; FlyBase:FBst0031307;

D. melanogaster. Expresses dsRNA for RNAi of Tl (FBgn0262473) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01276}attP2	Bloomington Drosophila Stock Center	BDSC:31477; FlyBase:FBst0031477;
D. melanogaster. Expresses dsRNA for RNAi of l(2)gl (FBgn0002121) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01073}attP2	Bloomington Drosophila Stock Center	BDSC:31517; FlyBase:FBst0031517;
D. melanogaster. Expresses dsRNA for RNAi of Jra (FBgn0001291) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01184}attP2	Bloomington Drosophila Stock Center	BDSC:31595; FlyBase:FBst0031595;
D. melanogaster. Expresses dsRNA for RNAi of Axn (FBgn0026597) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04012}attP2	Bloomington Drosophila Stock Center	BDSC:31705; FlyBase:FBst0031705;
D. melanogaster. Expresses dsRNA for RNAi of gig (FBgn0005198) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04083}attP2	Bloomington Drosophila Stock Center	BDSC:31770; FlyBase:FBst0031770;
D. melanogaster. Expresses dsRNA for RNAi of Med (FBgn0011655) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02218}attP2	Bloomington Drosophila Stock Center	BDSC:31928; FlyBase:FBst0031928;
D. melanogaster. Expresses dsRNA for RNAi of RagC-D (FBgn0033272) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00333}attP2	Bloomington Drosophila Stock Center	BDSC:32342; FlyBase:FBst0032342;
D. melanogaster. Expresses dsRNA for RNAi of AMPKalpha (FBgn0023169) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00362}attP2	Bloomington Drosophila Stock Center	BDSC:32371; FlyBase:FBst0032371;
D. melanogaster. Expresses dsRNA for RNAi of Rho1 (FBgn0014020) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00375}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32383; FlyBase:FBst0032383;
D. melanogaster. Expresses dsRNA for RNAi of pk (FBgn0003090) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00408}attP2	Bloomington Drosophila Stock Center	BDSC:32413; FlyBase:FBst0032413;
D. melanogaster. Expresses dsRNA for RNAi of RpS6 (FBgn0261592) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00413}attP2	Bloomington Drosophila Stock Center	BDSC:32418; FlyBase:FBst0032418;

D. melanogaster. Expresses dsRNA for RNAi of foxo (FBgn0038197) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00422}attP2	Bloomington Drosophila Stock Center	BDSC:32427; FlyBase:FBst0032427;
D. melanogaster. Expresses dsRNA for RNAi of Plc21C (FBgn0004611) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00436}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32438; FlyBase:FBst0032438;
D. melanogaster. Expresses dsRNA for RNAi of Ilp2 (FBgn0036046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00476}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32475; FlyBase:FBst0032475;
D. melanogaster. Expresses dsRNA for RNAi of hh (FBgn0004644) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00492}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32489; FlyBase:FBst0032489;
D. melanogaster. Expresses dsRNA for RNAi of Spred (FBgn0020767) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00637}attP2	Bloomington Drosophila Stock Center	BDSC:32852; FlyBase:FBst0032852;
D. melanogaster. Expresses dsRNA for RNAi of upd3 (FBgn0053542) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00646}attP2	Bloomington Drosophila Stock Center	BDSC:32859; FlyBase:FBst0032859;
D. melanogaster. Expresses dsRNA for RNAi of Ilp1 (FBgn0044051) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00648}attP2	Bloomington Drosophila Stock Center	BDSC:32861; FlyBase:FBst0032861;
D. melanogaster. Expresses dsRNA for RNAi of Ilp7 (FBgn0044046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00649}attP2	Bloomington Drosophila Stock Center	BDSC:32862; FlyBase:FBst0032862;
D. melanogaster. Expresses dsRNA for RNAi of SkpA (FBgn0025637) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00657}attP2	Bloomington Drosophila Stock Center	BDSC:32870; FlyBase:FBst0032870;
D. melanogaster. Expresses dsRNA for RNAi of CG3226 (FBgn0029882) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00662}attP2	Bloomington Drosophila Stock Center	BDSC:32875; FlyBase:FBst0032875;
D. melanogaster. Expresses dsRNA for RNAi of CtBP (FBgn0020496) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00677}attP2	Bloomington Drosophila Stock Center	BDSC:32889; FlyBase:FBst0032889;

D. melanogaster. Expresses dsRNA for RNAi of Hel89B (FBgn0022787) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00684}attP2	Bloomington Drosophila Stock Center	BDSC:32895; FlyBase:FBst0032895 ;
D. melanogaster. Expresses dsRNA for RNAi of Tctp (FBgn0037874) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00701}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32911; FlyBase:FBst0032911 ;
D. melanogaster. Expresses dsRNA for RNAi of jub (FBgn0030530) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00714}attP2	Bloomington Drosophila Stock Center	BDSC:32923; FlyBase:FBst0032923 ;
D. melanogaster. Expresses dsRNA for RNAi of Gug (FBgn0010825) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00756}attP2	Bloomington Drosophila Stock Center	BDSC:32961; FlyBase:FBst0032961 ;
D. melanogaster. Expresses dsRNA for RNAi of sav (FBgn0053193) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00760}attP2	Bloomington Drosophila Stock Center	BDSC:32965; FlyBase:FBst0032965 ;
D. melanogaster. Expresses dsRNA for RNAi of bsk (FBgn0000229) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00777}attP2	Bloomington Drosophila Stock Center	BDSC:32977; FlyBase:FBst0032977 ;
D. melanogaster. Expresses dsRNA for RNAi of Pgcl (FBgn0011822) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00792}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32992; FlyBase:FBst0032992 ;
D. melanogaster. Expresses dsRNA for RNAi of polo (FBgn0003124) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00530}attP2	Bloomington Drosophila Stock Center	BDSC:33042; FlyBase:FBst0033042 ;
D. melanogaster. Expresses dsRNA for RNAi of cnk (FBgn0286070) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00238}attP2	Bloomington Drosophila Stock Center	BDSC:33366; FlyBase:FBst0033366 ;
D. melanogaster. Expresses dsRNA for RNAi of kay (FBgn0001297) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00254}attP2	Bloomington Drosophila Stock Center	BDSC:33379; FlyBase:FBst0033379 ;
D. melanogaster. Expresses dsRNA for RNAi of apolpp (FBgn0087002) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00265}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:33388; FlyBase:FBst0033388 ;

D. melanogaster. Expresses dsRNA for RNAi of Su(var)205 (FBgn0003607) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00278}attP2	Bloomington Drosophila Stock Center	BDSC:33400; FlyBase:FBst0033400;
D. melanogaster. Expresses dsRNA for RNAi of Tak1 (FBgn0026323) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00282}attP2	Bloomington Drosophila Stock Center	BDSC:33404; FlyBase:FBst0033404;
D. melanogaster. Expresses dsRNA for RNAi of hpo (FBgn0261456) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2	Bloomington Drosophila Stock Center	BDSC:33614; FlyBase:FBst0033614;
D. melanogaster. Expresses dsRNA for RNAi of Akt1 (FBgn0010379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00007}attP2	Bloomington Drosophila Stock Center	BDSC:33615; FlyBase:FBst0033615;
D. melanogaster. Expresses dsRNA for RNAi of N (FBgn0004647) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00009}attP2	Bloomington Drosophila Stock Center	BDSC:33616; FlyBase:FBst0033616;
D. melanogaster. Expresses dsRNA for RNAi of csw (FBgn0000382) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00012}attP2	Bloomington Drosophila Stock Center	BDSC:33619; FlyBase:FBst0033619;
D. melanogaster. Expresses dsRNA for RNAi of tor (FBgn0003733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00021}attP2	Bloomington Drosophila Stock Center	BDSC:33627; FlyBase:FBst0033627;
D. melanogaster. Expresses dsRNA for RNAi of Stat92E (FBgn0016917) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00035}attP2	Bloomington Drosophila Stock Center	BDSC:33637; FlyBase:FBst0033637;
D. melanogaster. Expresses dsRNA for RNAi of Dsor1 (FBgn0010269) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00037}attP2	Bloomington Drosophila Stock Center	BDSC:33639; FlyBase:FBst0033639;
D. melanogaster. Expresses dsRNA for RNAi of Pten (FBgn0026379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00044}attP2	Bloomington Drosophila Stock Center	BDSC:33643; FlyBase:FBst0033643;
D. melanogaster. Expresses dsRNA for RNAi of CycD (FBgn0010315) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00059}attP2	Bloomington Drosophila Stock Center	BDSC:33653; FlyBase:FBst0033653;

D. melanogaster. Expresses dsRNA for RNAi of Rel (FBgn0014018) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00070}attP2	Bloomington Drosophila Stock Center	BDSC:33661; FlyBase:FBst0033661;
D. melanogaster. Expresses dsRNA for RNAi of upd1 (FBgn0004956) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00545}attP2	Bloomington Drosophila Stock Center	BDSC:33680; FlyBase:FBst0033680;
D. melanogaster. Expresses dsRNA for RNAi of Ilp3 (FBgn0044050) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00546}attP2	Bloomington Drosophila Stock Center	BDSC:33681; FlyBase:FBst0033681;
D. melanogaster. Expresses dsRNA for RNAi of Ilp4 (FBgn0044049) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00547}attP2	Bloomington Drosophila Stock Center	BDSC:33682; FlyBase:FBst0033682;
D. melanogaster. Expresses dsRNA for RNAi of Ilp5 (FBgn0044048) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00548}attP2	Bloomington Drosophila Stock Center	BDSC:33683; FlyBase:FBst0033683;
D. melanogaster. Expresses dsRNA for RNAi of Ilp6 (FBgn0044047) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00549}attP2	Bloomington Drosophila Stock Center	BDSC:33684; FlyBase:FBst0033684;
D. melanogaster. Expresses dsRNA for RNAi of d (FBgn0262029) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01096}attP2	Bloomington Drosophila Stock Center	BDSC:33754; FlyBase:FBst0033754;
D. melanogaster. Expresses dsRNA for RNAi of Dad (FBgn0020493) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01102}attP2	Bloomington Drosophila Stock Center	BDSC:33759; FlyBase:FBst0033759;
D. melanogaster. Expresses dsRNA for RNAi of inaC (FBgn0004784) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02958}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:33768; FlyBase:FBst0033768;
D. melanogaster. Expresses dsRNA for RNAi of wg (FBgn0284084) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00844}attP2	Bloomington Drosophila Stock Center	BDSC:33902; FlyBase:FBst0033902;
D. melanogaster. Expresses dsRNA for RNAi of srl (FBgn0037248) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00858}attP2	Bloomington Drosophila Stock Center	BDSC:33915; FlyBase:FBst0033915;

D. melanogaster. Expresses dsRNA for RNAi of SkpC (FBgn0026175) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00871}attP2	Bloomington Drosophila Stock Center	BDSC:33925; FlyBase:FBst0033925;
D. melanogaster. Expresses dsRNA for RNAi of SPE (FBgn0039102) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00873}attP2	Bloomington Drosophila Stock Center	BDSC:33926; FlyBase:FBst0033926;
D. melanogaster. Expresses dsRNA for RNAi of Traf6 (FBgn0265464) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00880}attP2	Bloomington Drosophila Stock Center	BDSC:33931; FlyBase:FBst0033931;
D. melanogaster. Expresses dsRNA for RNAi of upd2 (FBgn0030904) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00901}attP2	Bloomington Drosophila Stock Center	BDSC:33949; FlyBase:FBst0033949;
D. melanogaster. Expresses dsRNA for RNAi of Tor (FBgn0021796) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00904}attP2	Bloomington Drosophila Stock Center	BDSC:33951; FlyBase:FBst0033951;
D. melanogaster. Expresses dsRNA for RNAi of dally (FBgn0263930) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00905}attP2	Bloomington Drosophila Stock Center	BDSC:33952; FlyBase:FBst0033952;
D. melanogaster. Expresses dsRNA for RNAi of Rheb (FBgn0041191) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00923}attP2	Bloomington Drosophila Stock Center	BDSC:33966; FlyBase:FBst0033966;
D. melanogaster. Expresses dsRNA for RNAi of Cat (FBgn0000261) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00990}attP2	Bloomington Drosophila Stock Center	BDSC:34020; FlyBase:FBst0034020;
D. melanogaster. Expresses dsRNA for RNAi of CycK (FBgn0025674) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01003}attP2	Bloomington Drosophila Stock Center	BDSC:34032; FlyBase:FBst0034032;
D. melanogaster. Expresses dsRNA for RNAi of wts (FBgn0011739) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00026}attP2	Bloomington Drosophila Stock Center	BDSC:34064; FlyBase:FBst0034064;
D. melanogaster. Expresses dsRNA for RNAi of yki (FBgn0034970) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00041}attP2	Bloomington Drosophila Stock Center	BDSC:34067; FlyBase:FBst0034067;

D. melanogaster. Expresses dsRNA for RNAi of srp (FBgn0003507) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01083}attP2	Bloomington Drosophila Stock Center	BDSC:34080; FlyBase:FBst0034080 ;
D. melanogaster. Expresses dsRNA for RNAi of Fas2 (FBgn0000635) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01098}attP2	Bloomington Drosophila Stock Center	BDSC:34084; FlyBase:FBst0034084 ;
D. melanogaster. Expresses dsRNA for RNAi of eIF4E1 (FBgn0015218) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00969}attP2	Bloomington Drosophila Stock Center	BDSC:34096; FlyBase:FBst0034096 ;
D. melanogaster. Expresses dsRNA for RNAi of Sema1a (FBgn0011259) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01307}attP2	Bloomington Drosophila Stock Center	BDSC:34320; FlyBase:FBst0034320 ;
D. melanogaster. Expresses dsRNA for RNAi of fz (FBgn0001085) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01308}attP2	Bloomington Drosophila Stock Center	BDSC:34321; FlyBase:FBst0034321 ;
D. melanogaster. Expresses dsRNA for RNAi of fj (FBgn0000658) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01310}attP2	Bloomington Drosophila Stock Center	BDSC:34323; FlyBase:FBst0034323 ;
D. melanogaster. Expresses dsRNA for RNAi of eve (FBgn0000606) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01312}attP2	Bloomington Drosophila Stock Center	BDSC:34325; FlyBase:FBst0034325 ;
D. melanogaster. Expresses dsRNA for RNAi of tll (FBgn0003720) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01316}attP2	Bloomington Drosophila Stock Center	BDSC:34329; FlyBase:FBst0034329 ;
D. melanogaster. Expresses dsRNA for RNAi of aPKC (FBgn0261854) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01320}attP2	Bloomington Drosophila Stock Center	BDSC:34332; FlyBase:FBst0034332 ;
D. melanogaster. Expresses dsRNA for RNAi of Vang (FBgn0015838) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01343}attP2	Bloomington Drosophila Stock Center	BDSC:34354; FlyBase:FBst0034354 ;
D. melanogaster. Expresses dsRNA for RNAi of Lst8 (FBgn0264691) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01350}attP2	Bloomington Drosophila Stock Center	BDSC:34361; FlyBase:FBst0034361 ;

D. melanogaster. Expresses dsRNA for RNAi of Lkb1 (FBgn0038167) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01351}attP2	Bloomington Drosophila Stock Center	BDSC:34362; FlyBase:FBst0034362;
D. melanogaster. Expresses dsRNA for RNAi of puc (FBGn0243512) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01386}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34392; FlyBase:FBst0034392;
D. melanogaster. Expresses dsRNA for RNAi of Sxl (FBgn0264270) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00609}attP2	Bloomington Drosophila Stock Center	BDSC:34393; FlyBase:FBst0034393;
D. melanogaster. Expresses dsRNA for RNAi of Cka (FBgn0044323) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00081}attP2	Bloomington Drosophila Stock Center	BDSC:34522; FlyBase:FBst0034522;
D. melanogaster. Expresses dsRNA for RNAi of bnl (FBgn0014135) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01046}attP2	Bloomington Drosophila Stock Center	BDSC:34572; FlyBase:FBst0034572;
D. melanogaster. Expresses dsRNA for RNAi of RagA-B (FBgn0037647) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01064}attP2	Bloomington Drosophila Stock Center	BDSC:34590; FlyBase:FBst0034590;
D. melanogaster. Expresses dsRNA for RNAi of Wbp2 (FBgn0036318) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00563}attP2	Bloomington Drosophila Stock Center	BDSC:34603; FlyBase:FBst0034603;
D. melanogaster. Expresses dsRNA for RNAi of pip (FBgn0003089) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01288}attP2	Bloomington Drosophila Stock Center	BDSC:34613; FlyBase:FBst0034613;
D. melanogaster. Expresses dsRNA for RNAi of dome (FBgn0043903) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01293}attP2	Bloomington Drosophila Stock Center	BDSC:34618; FlyBase:FBst0034618;
D. melanogaster. Expresses dsRNA for RNAi of Stlk (FBgn0046692) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01295}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34620; FlyBase:FBst0034620;
D. melanogaster. Expresses dsRNA for RNAi of slp1 (FBgn0003430) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01107}attP2	Bloomington Drosophila Stock Center	BDSC:34633; FlyBase:FBst0034633;

D. melanogaster. Expresses dsRNA for RNAi of slp2 (FBgn0004567) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01108}attP2	Bloomington Drosophila Stock Center	BDSC:34634; FlyBase:FBst0034634;
D. melanogaster. Expresses dsRNA for RNAi of hth (FBgn0001235) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01112}attP2	Bloomington Drosophila Stock Center	BDSC:34637; FlyBase:FBst0034637;
D. melanogaster. Expresses dsRNA for RNAi of RhoGEF2 (FBgn0023172) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01118}attP2	Bloomington Drosophila Stock Center	BDSC:34643; FlyBase:FBst0034643;
D. melanogaster. Expresses dsRNA for RNAi of Wnt5 (FBgn0010194) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01119}attP2	Bloomington Drosophila Stock Center	BDSC:34644; FlyBase:FBst0034644;
D. melanogaster. Expresses dsRNA for RNAi of spi (FBgn0005672) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01120}attP2	Bloomington Drosophila Stock Center	BDSC:34645; FlyBase:FBst0034645;
D. melanogaster. Expresses dsRNA for RNAi of Galphao (FBgn0001122) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01129}attP2	Bloomington Drosophila Stock Center	BDSC:34653; FlyBase:FBst0034653;
D. melanogaster. Expresses dsRNA for RNAi of shn (FBgn0003396) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01167}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34689; FlyBase:FBst0034689;
D. melanogaster. Expresses dsRNA for RNAi of Ser (FBgn0004197) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01179}attP2	Bloomington Drosophila Stock Center	BDSC:34700; FlyBase:FBst0034700;
D. melanogaster. Expresses dsRNA for RNAi of H (FBgn0001169) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01182}attP2	Bloomington Drosophila Stock Center	BDSC:34703; FlyBase:FBst0034703;
D. melanogaster. Expresses dsRNA for RNAi of opa (FBgn0003002) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01185}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34706; FlyBase:FBst0034706;
D. melanogaster. Expresses dsRNA for RNAi of Pkc53E (FBgn0003091) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01195}attP2	Bloomington Drosophila Stock Center	BDSC:34716; FlyBase:FBst0034716;

D. melanogaster. Expresses dsRNA for RNAi of hkb (FBgn0261434) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01216}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34736; FlyBase:FBst0034736;
D. melanogaster. Expresses dsRNA for RNAi of gig (FBgn0005198) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01217}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34737; FlyBase:FBst0034737;
D. melanogaster. Expresses dsRNA for RNAi of ken (FBgn0011236) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01219}attP2	Bloomington Drosophila Stock Center	BDSC:34739; FlyBase:FBst0034739;
D. melanogaster. Expresses dsRNA for RNAi of cact (FBgn0000250) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00084}attP2	Bloomington Drosophila Stock Center	BDSC:34775; FlyBase:FBst0034775;
D. melanogaster. Expresses dsRNA for RNAi of Bx42 (FBgn0004856) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00086}attP2	Bloomington Drosophila Stock Center	BDSC:34777; FlyBase:FBst0034777;
D. melanogaster. Expresses dsRNA for RNAi of Rpt2 (FBgn0015282) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00104}attP2	Bloomington Drosophila Stock Center	BDSC:34795; FlyBase:FBst0034795;
D. melanogaster. Expresses dsRNA for RNAi of Skp2 (FBgn0037236) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00116}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34807; FlyBase:FBst0034807;
D. melanogaster. Expresses dsRNA for RNAi of raptor (FBgn0029840) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00124}attP2	Bloomington Drosophila Stock Center	BDSC:34814; FlyBase:FBst0034814;
D. melanogaster. Expresses dsRNA for RNAi of Sos (FBgn0001965) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00149}attP2	Bloomington Drosophila Stock Center	BDSC:34833; FlyBase:FBst0034833;
D. melanogaster. Expresses dsRNA for RNAi of Gprk2 (FBgn0261988) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00161}attP2	Bloomington Drosophila Stock Center	BDSC:34843; FlyBase:FBst0034843;
D. melanogaster. Expresses dsRNA for RNAi of rl (FBgn0003256) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00173}attP2	Bloomington Drosophila Stock Center	BDSC:34855; FlyBase:FBst0034855;

D. melanogaster. Expresses dsRNA for RNAi of mwh (FBgn0264272) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00180}attP2	Bloomington Drosophila Stock Center	BDSC:34862; FlyBase:FBst0034862;
D. melanogaster. Expresses dsRNA for RNAi of Apc (FBGn0015589) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00188}attP2	Bloomington Drosophila Stock Center	BDSC:34869; FlyBase:FBst0034869;
D. melanogaster. Expresses dsRNA for RNAi of Tao (FBgn0031030) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01226}attP2	Bloomington Drosophila Stock Center	BDSC:34881; FlyBase:FBst0034881;
D. melanogaster. Expresses dsRNA for RNAi of 14-3-3epsilon (FBgn0020238) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01229}attP2	Bloomington Drosophila Stock Center	BDSC:34884; FlyBase:FBst0034884;
D. melanogaster. Expresses dsRNA for RNAi of Sara (FBgn0026369) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01239}attP2	Bloomington Drosophila Stock Center	BDSC:34894; FlyBase:FBst0034894;
D. melanogaster. Expresses dsRNA for RNAi of gbb (FBgn0024234) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01243}attP2	Bloomington Drosophila Stock Center	BDSC:34898; FlyBase:FBst0034898;
D. melanogaster. Expresses dsRNA for RNAi of Atg8b (FBgn0038539) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01245}attP2	Bloomington Drosophila Stock Center	BDSC:34900; FlyBase:FBst0034900;
D. melanogaster. Expresses dsRNA for RNAi of Rac1 (FBgn0010333) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01258}attP2	Bloomington Drosophila Stock Center	BDSC:34910; FlyBase:FBst0034910;
D. melanogaster. Expresses dsRNA for RNAi of ato (FBgn0010433) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01278}attP2	Bloomington Drosophila Stock Center	BDSC:34929; FlyBase:FBst0034929;
D. melanogaster. Expresses dsRNA for RNAi of dl (FBgn0260632) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00028}attP2	Bloomington Drosophila Stock Center	BDSC:34938; FlyBase:FBst0034938;
D. melanogaster. Expresses dsRNA for RNAi of Mer (FBgn0086384) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00459}attP2	Bloomington Drosophila Stock Center	BDSC:34958; FlyBase:FBst0034958;

D. melanogaster. Expresses dsRNA for RNAi of mats (FBgn0038965) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00475}attP2	Bloomington Drosophila Stock Center	BDSC:34959; FlyBase:FBst0034959;
D. melanogaster. Expresses dsRNA for RNAi of ex (FBgn0004583) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00874}attP2	Bloomington Drosophila Stock Center	BDSC:34968; FlyBase:FBst0034968;
D. melanogaster. Expresses dsRNA for RNAi of ft (FBgn0001075) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00932}attP2	Bloomington Drosophila Stock Center	BDSC:34970; FlyBase:FBst0034970;
D. melanogaster. Expresses dsRNA for RNAi of msk (FBgn0026252) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01408}attP2	Bloomington Drosophila Stock Center	BDSC:34998; FlyBase:FBst0034998;
D. melanogaster. Expresses dsRNA for RNAi of baz (FBgn0000163) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01412}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35002; FlyBase:FBst0035002;
D. melanogaster. Expresses dsRNA for RNAi of arm (FBgn0000117) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01414}attP2	Bloomington Drosophila Stock Center	BDSC:35004; FlyBase:FBst0035004;
D. melanogaster. Expresses dsRNA for RNAi of Stam (FBgn0027363) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01429}attP2	Bloomington Drosophila Stock Center	BDSC:35016; FlyBase:FBst0035016;
D. melanogaster. Expresses dsRNA for RNAi of Gadd45 (FBgn0033153) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01436}attP2	Bloomington Drosophila Stock Center	BDSC:35023; FlyBase:FBst0035023;
D. melanogaster. Expresses dsRNA for RNAi of htl (FBgn0010389) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01437}attP2	Bloomington Drosophila Stock Center	BDSC:35024; FlyBase:FBst0035024;
D. melanogaster. Expresses dsRNA for RNAi of tsh (FBgn0003866) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01443}attP2	Bloomington Drosophila Stock Center	BDSC:35030; FlyBase:FBst0035030;
D. melanogaster. Expresses dsRNA for RNAi of Socs36E (FBgn0041184) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01450}attP2	Bloomington Drosophila Stock Center	BDSC:35036; FlyBase:FBst0035036;

D. melanogaster. Expresses dsRNA for RNAi of Cad99C (FBgn0039709) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01451}attP2	Bloomington Drosophila Stock Center	BDSC:35037; FlyBase:FBst0035037;
D. melanogaster. Expresses dsRNA for RNAi of pnt (FBgn0003118) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01452}attP2	Bloomington Drosophila Stock Center	BDSC:35038; FlyBase:FBst0035038;
D. melanogaster. Expresses dsRNA for RNAi of numb (FBgn0002973) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01459}attP2	Bloomington Drosophila Stock Center	BDSC:35045; FlyBase:FBst0035045;
D. melanogaster. Expresses dsRNA for RNAi of stan (FBgn0024836) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01464}attP2	Bloomington Drosophila Stock Center	BDSC:35050; FlyBase:FBst0035050;
D. melanogaster. Expresses dsRNA for RNAi of dco (FBgn0002413) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00001}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35134; FlyBase:FBst0035134;
D. melanogaster. Expresses dsRNA for RNAi of CkIIalpha (FBgn0264492) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00003}attP2	Bloomington Drosophila Stock Center	BDSC:35136; FlyBase:FBst0035136;
D. melanogaster. Expresses dsRNA for RNAi of gish (FBgn0250823) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00005}attP2	Bloomington Drosophila Stock Center	BDSC:35138; FlyBase:FBst0035138;
D. melanogaster. Expresses dsRNA for RNAi of Mkk4 (FBgn0024326) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00010}attP2	Bloomington Drosophila Stock Center	BDSC:35143; FlyBase:FBst0035143;
D. melanogaster. Expresses dsRNA for RNAi of CkIalpha (FBgn0015024) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00021}attP2	Bloomington Drosophila Stock Center	BDSC:35153; FlyBase:FBst0035153;
D. melanogaster. Expresses dsRNA for RNAi of lic (FBgn0261524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00022}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35154; FlyBase:FBst0035154;
D. melanogaster. Expresses dsRNA for RNAi of Btk29A (FBgn0003502) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00027}attP2	Bloomington Drosophila Stock Center	BDSC:35159; FlyBase:FBst0035159;

D. melanogaster. Expresses dsRNA for RNAi of alc (FBgn0260972) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00029}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35161; FlyBase:FBst0035161;
D. melanogaster. Expresses dsRNA for RNAi of Pka-C1 (FBgn0000273) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00038}attP2	Bloomington Drosophila Stock Center	BDSC:35169; FlyBase:FBst0035169;
D. melanogaster. Expresses dsRNA for RNAi of hpo (FBgn0261456) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00046}attP2	Bloomington Drosophila Stock Center	BDSC:35176; FlyBase:FBst0035176;
D. melanogaster. Expresses dsRNA for RNAi of IKKbeta (FBgn0024222) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00058}attP2	Bloomington Drosophila Stock Center	BDSC:35186; FlyBase:FBst0035186;
D. melanogaster. Expresses dsRNA for RNAi of Aduk (FBgn0037679) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00059}attP2	Bloomington Drosophila Stock Center	BDSC:35187; FlyBase:FBst0035187;
D. melanogaster. Expresses dsRNA for RNAi of put (FBgn0003169) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00069}attP2	Bloomington Drosophila Stock Center	BDSC:35195; FlyBase:FBst0035195;
D. melanogaster. Expresses dsRNA for RNAi of hep (FBgn0010303) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00089}attP2	Bloomington Drosophila Stock Center	BDSC:35210; FlyBase:FBst0035210;
D. melanogaster. Expresses dsRNA for RNAi of brm (FBgn0000212) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00090}attP2	Bloomington Drosophila Stock Center	BDSC:35211; FlyBase:FBst0035211;
D. melanogaster. Expresses dsRNA for RNAi of pyd (FBgn0262614) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00109}attP2	Bloomington Drosophila Stock Center	BDSC:35225; FlyBase:FBst0035225;
D. melanogaster. Expresses dsRNA for RNAi of hyx (FBgn0037657) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00123}attP2	Bloomington Drosophila Stock Center	BDSC:35238; FlyBase:FBst0035238;
D. melanogaster. Expresses dsRNA for RNAi of p38a (FBgn0015765) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00131}attP2	Bloomington Drosophila Stock Center	BDSC:35244; FlyBase:FBst0035244;

D. melanogaster. Expresses dsRNA for RNAi of p38b (FBgn0024846) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00140}attP2	Bloomington Drosophila Stock Center	BDSC:35252; FlyBase:FBst0035252;
D. melanogaster. Expresses dsRNA for RNAi of app (FBgn0260941) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00181}attP2	Bloomington Drosophila Stock Center	BDSC:35280; FlyBase:FBst0035280;
D. melanogaster. Expresses dsRNA for RNAi of sdt (FBgn0261873) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00193}attP2	Bloomington Drosophila Stock Center	BDSC:35291; FlyBase:FBst0035291;
D. melanogaster. Expresses dsRNA for RNAi of aux (FBgn0037218) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00213}attP2	Bloomington Drosophila Stock Center	BDSC:35310; FlyBase:FBst0035310;
D. melanogaster. Expresses dsRNA for RNAi of CaMKII (FBgn0264607) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00237}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35330; FlyBase:FBst0035330;
D. melanogaster. Expresses dsRNA for RNAi of Ask1 (FBgn0014006) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00238}attP2	Bloomington Drosophila Stock Center	BDSC:35331; FlyBase:FBst0035331;
D. melanogaster. Expresses dsRNA for RNAi of SAK (FBgn0026371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00244}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35335; FlyBase:FBst0035335;
D. melanogaster. Expresses dsRNA for RNAi of sgg (FBgn0003371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00277}attP2	Bloomington Drosophila Stock Center	BDSC:35364; FlyBase:FBst0035364;
D. melanogaster. Expresses dsRNA for RNAi of hop (FBgn0004864) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00305}attP2	Bloomington Drosophila Stock Center	BDSC:35386; FlyBase:FBst0035386;
D. melanogaster. Expresses dsRNA for RNAi of Mekk1 (FBgn0024329) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00322}attP2	Bloomington Drosophila Stock Center	BDSC:35402; FlyBase:FBst0035402;
D. melanogaster. Expresses dsRNA for RNAi of aop (FBgn0000097) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00324}attP2	Bloomington Drosophila Stock Center	BDSC:35404; FlyBase:FBst0035404;

D. melanogaster. Expresses dsRNA for RNAi of Ras85D (FBgn0003205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00336}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35414; FlyBase:FBst0035414;
D. melanogaster. Expresses dsRNA for RNAi of Nap1 (FBgn0015268) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00370}attP2	Bloomington Drosophila Stock Center	BDSC:35445; FlyBase:FBst0035445;
D. melanogaster. Expresses dsRNA for RNAi of osa (FBgn0261885) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00372}attP2	Bloomington Drosophila Stock Center	BDSC:35447; FlyBase:FBst0035447;
D. melanogaster. Expresses dsRNA for RNAi of key (FBgn0041205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00088}attP2	Bloomington Drosophila Stock Center	BDSC:35572; FlyBase:FBst0035572;
D. melanogaster. Expresses dsRNA for RNAi of Stat92E (FBgn0016917) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00437}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:35600; FlyBase:FBst0035600;
D. melanogaster. Expresses dsRNA for RNAi of Gcn5 (FBgn0020388) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00439}attP40	Bloomington Drosophila Stock Center	BDSC:35601; FlyBase:FBst0035601;
D. melanogaster. Expresses dsRNA for RNAi of TER94 (FBgn0261014) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00448}attP2	Bloomington Drosophila Stock Center	BDSC:35608; FlyBase:FBst0035608;
D. melanogaster. Expresses dsRNA for RNAi of wek (FBgn0001990) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21045}attP2	Bloomington Drosophila Stock Center	BDSC:35680; FlyBase:FBst0035680;
D. melanogaster. Expresses dsRNA for RNAi of bab2 (FBgn0025525) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21085}attP2	Bloomington Drosophila Stock Center	BDSC:35720; FlyBase:FBst0035720;
D. melanogaster. Expresses dsRNA for RNAi of Patj (FBgn0067864) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01489}attP2	Bloomington Drosophila Stock Center	BDSC:35747; FlyBase:FBst0035747;
D. melanogaster. Expresses dsRNA for RNAi of Mtl (FBgn0039532) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01500}attP2	Bloomington Drosophila Stock Center	BDSC:35754; FlyBase:FBst0035754;

D. melanogaster. Expresses dsRNA for RNAi of gro (FBgn0001139) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01506}attP2	Bloomington Drosophila Stock Center	BDSC:35759; FlyBase:FBst0035759 ;
D. melanogaster. Expresses dsRNA for RNAi of Wnt6 (FBgn0031902) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00457}attP2	Bloomington Drosophila Stock Center	BDSC:35808; FlyBase:FBst0035808 ;
D. melanogaster. Expresses dsRNA for RNAi of CycE (FBgn0010382) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00511}attP40	Bloomington Drosophila Stock Center	BDSC:36092; FlyBase:FBst0036092 ;
D. melanogaster. Expresses dsRNA for RNAi of Myd88 (FBgn0033402) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00183}attP2	Bloomington Drosophila Stock Center	BDSC:36107; FlyBase:FBst0036107 ;
D. melanogaster. Expresses dsRNA for RNAi of Cul1 (FBgn0015509) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00561}attP2	Bloomington Drosophila Stock Center	BDSC:36601; FlyBase:FBst0036601 ;
D. melanogaster. Expresses dsRNA for RNAi of chico (FBgn0024248) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01553}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36665; FlyBase:FBst0036665 ;
D. melanogaster. Expresses dsRNA for RNAi of Rbf2 (FBgn0038390) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01586}attP2	Bloomington Drosophila Stock Center	BDSC:36697; FlyBase:FBst0036697 ;
D. melanogaster. Expresses dsRNA for RNAi of rictor (FBgn0031006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01588}attP2	Bloomington Drosophila Stock Center	BDSC:36699; FlyBase:FBst0036699 ;
D. melanogaster. Expresses dsRNA for RNAi of sty (FBgn0014388) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01599}attP2	Bloomington Drosophila Stock Center	BDSC:36709; FlyBase:FBst0036709 ;
D. melanogaster. Expresses dsRNA for RNAi of Zyx (FBgn0011642) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01606}attP40	Bloomington Drosophila Stock Center	BDSC:36716; FlyBase:FBst0036716 ;
D. melanogaster. Expresses dsRNA for RNAi of Wnt2 (FBgn0004360) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01613}attP2	Bloomington Drosophila Stock Center	BDSC:36722; FlyBase:FBst0036722 ;

D. melanogaster. Expresses dsRNA for RNAi of Rbf (FBgn0015799) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS03004}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36744; FlyBase:FBst0036744;
D. melanogaster. Expresses dsRNA for RNAi of dlg1 (FBgn0001624) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02287}attP2	Bloomington Drosophila Stock Center	BDSC:36771; FlyBase:FBst0036771;
D. melanogaster. Expresses dsRNA for RNAi of dpp (FBgn0000490) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02455}attP2	Bloomington Drosophila Stock Center	BDSC:36779; FlyBase:FBst0036779;
D. melanogaster. Expresses dsRNA for RNAi of Dl (FBgn0000463) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00520}attP40	Bloomington Drosophila Stock Center	BDSC:36784; FlyBase:FBst0036784;
D. melanogaster. Expresses dsRNA for RNAi of HDAC1 (FBgn0015805) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01005}attP40	Bloomington Drosophila Stock Center	BDSC:36800; FlyBase:FBst0036800;
D. melanogaster. Expresses dsRNA for RNAi of Su(dx) (FBgn0003557) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01077}attP2	Bloomington Drosophila Stock Center	BDSC:36836; FlyBase:FBst0036836;
D. melanogaster. Expresses dsRNA for RNAi of pbl (FBgn0003041) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01092}attP2	Bloomington Drosophila Stock Center	BDSC:36841; FlyBase:FBst0036841;
D. melanogaster. Expresses dsRNA for RNAi of Sod2 (FBgn0010213) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01015}attP40	Bloomington Drosophila Stock Center	BDSC:36871; FlyBase:FBst0036871;
D. melanogaster. Expresses dsRNA for RNAi of Art1 (FBgn0037834) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01072}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36891; FlyBase:FBst0036891;
D. melanogaster. Expresses dsRNA for RNAi of CG10924 (FBgn0034356) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00200}attP2	Bloomington Drosophila Stock Center	BDSC:36915; FlyBase:FBst0036915;
D. melanogaster. Expresses dsRNA for RNAi of trr (FBgn0023518) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01019}attP2	Bloomington Drosophila Stock Center	BDSC:36916; FlyBase:FBst0036916;

D. melanogaster. Expresses dsRNA for RNAi of Cdc42 (FBgn0010341) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00620}attP40	Bloomington Drosophila Stock Center	BDSC:37477; FlyBase:FBst0037477 ;
D. melanogaster. Expresses dsRNA for RNAi of nej (FBGn0261617) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01507}attP2	Bloomington Drosophila Stock Center	BDSC:37489; FlyBase:FBst0037489 ;
D. melanogaster. Expresses dsRNA for RNAi of Pvr (FBgn0032006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01662}attP40	Bloomington Drosophila Stock Center	BDSC:37520; FlyBase:FBst0037520 ;
D. melanogaster. Expresses dsRNA for RNAi of cno (FBgn0259212) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00633}attP40	Bloomington Drosophila Stock Center	BDSC:38194; FlyBase:FBst0038194 ;
D. melanogaster. Expresses dsRNA for RNAi of pygo (FBgn0043900) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00647}attP40	Bloomington Drosophila Stock Center	BDSC:38208; FlyBase:FBst0038208 ;
D. melanogaster. Expresses dsRNA for RNAi of ed (FBgn0000547) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00648}attP40	Bloomington Drosophila Stock Center	BDSC:38209; FlyBase:FBst0038209 ;
D. melanogaster. Expresses dsRNA for RNAi of Nrg (FBgn0264975) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00656}attP40	Bloomington Drosophila Stock Center	BDSC:38215; FlyBase:FBst0038215 ;
D. melanogaster. Expresses dsRNA for RNAi of aph-1 (FBgn0031458) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01693}attP40	Bloomington Drosophila Stock Center	BDSC:38249; FlyBase:FBst0038249 ;
D. melanogaster. Expresses dsRNA for RNAi of cbt (FBgn0043364) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01726}attP40	Bloomington Drosophila Stock Center	BDSC:38276; FlyBase:FBst0038276 ;
D. melanogaster. Expresses dsRNA for RNAi of Vps36 (FBgn0086785) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01739}attP40	Bloomington Drosophila Stock Center	BDSC:38286; FlyBase:FBst0038286 ;
D. melanogaster. Expresses dsRNA for RNAi of crb (FBgn0259685) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01842}attP40	Bloomington Drosophila Stock Center	BDSC:38373; FlyBase:FBst0038373 ;

D. melanogaster. Expresses dsRNA for RNAi of tws (FBgn0004889) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00670}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38899; FlyBase:FBst0038899;
D. melanogaster. Expresses dsRNA for RNAi of S (FBgn0003310) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00686}attP2	Bloomington Drosophila Stock Center	BDSC:38914; FlyBase:FBst0038914;
D. melanogaster. Expresses dsRNA for RNAi of Pvf3 (FBgn0085407) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01876}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38962; FlyBase:FBst0038962;
D. melanogaster. Expresses dsRNA for RNAi of CanB2 (FBgn0015614) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01886}attP2	Bloomington Drosophila Stock Center	BDSC:38971; FlyBase:FBst0038971;
D. melanogaster. Expresses dsRNA for RNAi of Pi3K21B (FBgn0020622) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01907}attP40	Bloomington Drosophila Stock Center	BDSC:38991; FlyBase:FBst0038991;
D. melanogaster. Expresses dsRNA for RNAi of par-6 (FBgn0026192) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01928}attP40	Bloomington Drosophila Stock Center	BDSC:39010; FlyBase:FBst0039010;
D. melanogaster. Expresses dsRNA for RNAi of Pvf1 (FBgn0030964) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01958}attP40	Bloomington Drosophila Stock Center	BDSC:39038; FlyBase:FBst0039038;
D. melanogaster. Expresses dsRNA for RNAi of Pka-C3 (FBgn0000489) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01970}attP2	Bloomington Drosophila Stock Center	BDSC:39050; FlyBase:FBst0039050;
D. melanogaster. Expresses dsRNA for RNAi of DAAM (FBgn0025641) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01978}attP2	Bloomington Drosophila Stock Center	BDSC:39058; FlyBase:FBst0039058;
D. melanogaster. Expresses dsRNA for RNAi of scrib (FBgn0263289) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01993}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:39073; FlyBase:FBst0039073;
D. melanogaster. Expresses dsRNA for RNAi of sina (FBgn0003410) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02008}attP40	Bloomington Drosophila Stock Center	BDSC:40842; FlyBase:FBst0040842;

D. melanogaster. Expresses dsRNA for RNAi of btl (FBgn0285896) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02038}attP2	Bloomington Drosophila Stock Center	BDSC:40871; FlyBase:FBst0040871 ;
D. melanogaster. Expresses dsRNA for RNAi of Pp2B-14D (FBgn0011826) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02039}attP2	Bloomington Drosophila Stock Center	BDSC:40872; FlyBase:FBst0040872 ;
D. melanogaster. Expresses dsRNA for RNAi of Smurf (FBgn0029006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02153}attP40	Bloomington Drosophila Stock Center	BDSC:40905; FlyBase:FBst0040905 ;
D. melanogaster. Expresses dsRNA for RNAi of CycB (FBgn0000405) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02163}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:40915; FlyBase:FBst0040915 ;
D. melanogaster. Expresses dsRNA for RNAi of fu (FBgn0001079) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00705}attP40	Bloomington Drosophila Stock Center	BDSC:41588; FlyBase:FBst0041588 ;
D. melanogaster. Expresses dsRNA for RNAi of ru (FBgn0003295) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01129}attP2	Bloomington Drosophila Stock Center	BDSC:41593; FlyBase:FBst0041593 ;
D. melanogaster. Expresses dsRNA for RNAi of ksr (FBgn0015402) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01134}attP2	Bloomington Drosophila Stock Center	BDSC:41598; FlyBase:FBst0041598 ;
D. melanogaster. Expresses dsRNA for RNAi of slpr (FBgn0030018) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01187}attP2	Bloomington Drosophila Stock Center	BDSC:41605; FlyBase:FBst0041605 ;
D. melanogaster. Expresses dsRNA for RNAi of p53 (FBgn0039044) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01220}attP40	Bloomington Drosophila Stock Center	BDSC:41638; FlyBase:FBst0041638 ;
D. melanogaster. Expresses dsRNA for RNAi of Smox (FBgn0025800) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02203}attP40	Bloomington Drosophila Stock Center	BDSC:41670; FlyBase:FBst0041670 ;
D. melanogaster. Expresses dsRNA for RNAi of rho (FBgn0004635) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02264}attP40	Bloomington Drosophila Stock Center	BDSC:41699; FlyBase:FBst0041699 ;

D. melanogaster. Expresses dsRNA for RNAi of S6k (FBgn0283472) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02267}attP2	Bloomington Drosophila Stock Center	BDSC:41702; FlyBase:FBst0041702;
D. melanogaster. Expresses dsRNA for RNAi of snk (FBgn0003450) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02289}attP2	Bloomington Drosophila Stock Center	BDSC:41723; FlyBase:FBst0041723;
D. melanogaster. Expresses dsRNA for RNAi of Ufd1-like (FBgn0036136) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01251}attP2	Bloomington Drosophila Stock Center	BDSC:41823; FlyBase:FBst0041823;
D. melanogaster. Expresses dsRNA for RNAi of RasGAP1 (FBgn0004390) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01258}attP2	Bloomington Drosophila Stock Center	BDSC:41830; FlyBase:FBst0041830;
D. melanogaster. Expresses dsRNA for RNAi of 14-3-3zeta (FBgn0004907) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01310}attP40	Bloomington Drosophila Stock Center	BDSC:41878; FlyBase:FBst0041878;
D. melanogaster. Expresses dsRNA for RNAi of tkv (FBgn0003716) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01338}attP2	Bloomington Drosophila Stock Center	BDSC:41904; FlyBase:FBst0041904;
D. melanogaster. Expresses dsRNA for RNAi of homer (FBgn0025777) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02301}attP2	Bloomington Drosophila Stock Center	BDSC:41908; FlyBase:FBst0041908;
D. melanogaster. Expresses dsRNA for RNAi of pll (FBgn0010441) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02332}attP40	Bloomington Drosophila Stock Center	BDSC:41935; FlyBase:FBst0041935;
D. melanogaster. Expresses dsRNA for RNAi of elB (FBgn0004858) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02357}attP2	Bloomington Drosophila Stock Center	BDSC:41960; FlyBase:FBst0041960;
D. melanogaster. Expresses dsRNA for RNAi of ea (FBgn0000533) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02358}attP2	Bloomington Drosophila Stock Center	BDSC:41961; FlyBase:FBst0041961;
D. melanogaster. Expresses dsRNA for RNAi of CycB3 (FBgn0015625) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02377}attP2	Bloomington Drosophila Stock Center	BDSC:41979; FlyBase:FBst0041979;

D. melanogaster. Expresses dsRNA for RNAi of lgs (FBgn0039907) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02381}attP2	Bloomington Drosophila Stock Center	BDSC:41983; FlyBase:FBst0041983;
D. melanogaster. Expresses dsRNA for RNAi of Actbeta (FBgn0024913) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02057}attP40	Bloomington Drosophila Stock Center	BDSC:42493; FlyBase:FBst0042493;
D. melanogaster. Expresses dsRNA for RNAi of CG5059 (FBgn0037007) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02058}attP40	Bloomington Drosophila Stock Center	BDSC:42494; FlyBase:FBst0042494;
D. melanogaster. Expresses dsRNA for RNAi of msn (FBgn0010909) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02084}attP40	Bloomington Drosophila Stock Center	BDSC:42518; FlyBase:FBst0042518;
D. melanogaster. Expresses dsRNA for RNAi of Rassf (FBgn0039055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02102}attP40	Bloomington Drosophila Stock Center	BDSC:42534; FlyBase:FBst0042534;
D. melanogaster. Expresses dsRNA for RNAi of sax (FBgn0003317) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02118}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:42546; FlyBase:FBst0042546;
D. melanogaster. Expresses dsRNA for RNAi of et (FBgn0031055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02213}attP40	Bloomington Drosophila Stock Center	BDSC:42557; FlyBase:FBst0042557;
D. melanogaster. Expresses dsRNA for RNAi of nmo (FBgn0011817) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02229}attP40	Bloomington Drosophila Stock Center	BDSC:42570; FlyBase:FBst0042570;
D. melanogaster. Expresses dsRNA for RNAi of CG12147 (FBgn0037325) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02447}attP2	Bloomington Drosophila Stock Center	BDSC:42612; FlyBase:FBst0042612;
D. melanogaster. Expresses dsRNA for RNAi of Act5C (FBgn0000042) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02487}attP2	Bloomington Drosophila Stock Center	BDSC:42651; FlyBase:FBst0042651;
D. melanogaster. Expresses dsRNA for RNAi of Act87E (FBgn0000046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02488}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:42652; FlyBase:FBst0042652;

D. melanogaster. Expresses dsRNA for RNAi of emc (FBgn0000575) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00724}attP2	Bloomington Drosophila Stock Center	BDSC:42768; FlyBase:FBst0042768;
D. melanogaster. Expresses dsRNA for RNAi of the Stellate gene family (FBgn0003523) plus Ste12DOR and SteXh:CG42398 (FBgn0044817 and FBgn0259817) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01156}attP2	Bloomington Drosophila Stock Center	BDSC:42786; FlyBase:FBst0042786;
D. melanogaster. Expresses dsRNA for RNAi of Socs44A (FBgn0033266) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02515}attP2	Bloomington Drosophila Stock Center	BDSC:42830; FlyBase:FBst0042830;
D. melanogaster. Expresses dsRNA for RNAi of gcm (FBgn0014179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02582}attP40	Bloomington Drosophila Stock Center	BDSC:42889; FlyBase:FBst0042889;
D. melanogaster. Expresses dsRNA for RNAi of CkIIbeta (FBgn0000259) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02636}attP40	Bloomington Drosophila Stock Center	BDSC:42943; FlyBase:FBst0042943;
D. melanogaster. Expresses dsRNA for RNAi of smo (FBgn0003444) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01472}attP2	Bloomington Drosophila Stock Center	BDSC:43134; FlyBase:FBst0043134;
D. melanogaster. Expresses dsRNA for RNAi of dock (FBgn0010583) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01519}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43176; FlyBase:FBst0043176;
D. melanogaster. Expresses dsRNA for RNAi of Mad (FBgn0011648) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01527}attP40	Bloomington Drosophila Stock Center	BDSC:43183; FlyBase:FBst0043183;
D. melanogaster. Expresses dsRNA for RNAi of Pp2A-29B (FBgn0260439) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01921}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43283; FlyBase:FBst0043283;
D. melanogaster. Expresses dsRNA for RNAi of CG11658 (FBgn0036196) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02671}attP40	Bloomington Drosophila Stock Center	BDSC:43298; FlyBase:FBst0043298;

D. melanogaster. Expresses dsRNA for RNAi of Myc (FBgn0262656) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01314}attP40	Bloomington Drosophila Stock Center	BDSC:43962; FlyBase:FBst0043962;
D. melanogaster. Expresses dsRNA for RNAi of pav (FBGn0011692) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01316}attP40	Bloomington Drosophila Stock Center	BDSC:43963; FlyBase:FBst0043963;
D. melanogaster. Expresses dsRNA for RNAi of eIF4EHP (FBgn0053100) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02703}attP40	Bloomington Drosophila Stock Center	BDSC:43990; FlyBase:FBst0043990;
D. melanogaster. Expresses dsRNA for RNAi of Atg1 (FBgn0260945) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02750}attP40	Bloomington Drosophila Stock Center	BDSC:44034; FlyBase:FBst0044034;
D. melanogaster. Expresses dsRNA for RNAi of tefu (FBgn0045035) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02790}attP40	Bloomington Drosophila Stock Center	BDSC:44073; FlyBase:FBst0044073;
D. melanogaster. Expresses dsRNA for RNAi of bi (FBgn0000179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02815}attP2	Bloomington Drosophila Stock Center	BDSC:44095; FlyBase:FBst0044095;
D. melanogaster. Expresses dsRNA for RNAi of mad2 (FBgn0035640) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01381}attP2	Bloomington Drosophila Stock Center	BDSC:44430; FlyBase:FBst0044430;
D. melanogaster. Expresses dsRNA for RNAi of ebi (FBgn0263933) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01413}attP40	Bloomington Drosophila Stock Center	BDSC:44443; FlyBase:FBst0044443;
D. melanogaster. Expresses dsRNA for RNAi of dx (FBgn0000524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01607}attP2	Bloomington Drosophila Stock Center	BDSC:44455; FlyBase:FBst0044455;
D. melanogaster. Expresses dsRNA for RNAi of fz3 (FBgn0027343) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01626}attP2	Bloomington Drosophila Stock Center	BDSC:44468; FlyBase:FBst0044468;
D. melanogaster. Expresses dsRNA for RNAi of cos (FBgn0000352) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02347}attP2	Bloomington Drosophila Stock Center	BDSC:44472; FlyBase:FBst0044472;

D. melanogaster. Expresses dsRNA for RNAi of disp (FBgn0029088) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02877}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:44633; FlyBase:FBst0044633;
D. melanogaster. Expresses dsRNA for RNAi of spen (FBGn0016977) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01647}attP40	Bloomington Drosophila Stock Center	BDSC:50529; FlyBase:FBst0050529;
D. melanogaster. Expresses dsRNA for RNAi of dlp (FBgn0041604) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01658}attP40	Bloomington Drosophila Stock Center	BDSC:50540; FlyBase:FBst0050540;
D. melanogaster. Expresses dsRNA for RNAi of wgn (FBgn0030941) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01716}attP2	Bloomington Drosophila Stock Center	BDSC:50594; FlyBase:FBst0050594;
D. melanogaster. Expresses dsRNA for RNAi of RpL8 (FBgn0261602) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02977}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50610; FlyBase:FBst0050610;
D. melanogaster. Expresses dsRNA for RNAi of Act42A (FBgn0000043) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02992}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50625; FlyBase:FBst0050625;
D. melanogaster. Expresses dsRNA for RNAi of tin (FBgn0004110) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03064}attP2	Bloomington Drosophila Stock Center	BDSC:50663; FlyBase:FBst0050663;
D. melanogaster. Expresses dsRNA for RNAi of EcR (FBgn0000546) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03114}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50712; FlyBase:FBst0050712;
D. melanogaster. Expresses dsRNA for RNAi of aru (FBgn0029095) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02966}attP2	Bloomington Drosophila Stock Center	BDSC:50730; FlyBase:FBst0050730;
D. melanogaster. Expresses dsRNA for RNAi of eIF4E4 (FBgn0035709) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21052}attP40	Bloomington Drosophila Stock Center	BDSC:50951; FlyBase:FBst0050951;
D. melanogaster. Expresses dsRNA for RNAi of pont (FBgn0040078) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21078}attP40	Bloomington Drosophila Stock Center	BDSC:50972; FlyBase:FBst0050972;

D. melanogaster. Expresses dsRNA for RNAi of dpn (FBgn0010109) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03154}attP2	Bloomington Drosophila Stock Center	BDSC:51440; FlyBase:FBst0051440;
D. melanogaster. Expresses dsRNA for RNAi of Spn27A (FBgn0028990) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03159}attP2	Bloomington Drosophila Stock Center	BDSC:51445; FlyBase:FBst0051445;
D. melanogaster. Expresses dsRNA for RNAi of ttv (FBgn0265974) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03225}attP40	Bloomington Drosophila Stock Center	BDSC:51480; FlyBase:FBst0051480;
D. melanogaster. Expresses dsRNA for RNAi of Mipp2 (FBgn0026060) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03229}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:51482; FlyBase:FBst0051482;
D. melanogaster. Expresses dsRNA for RNAi of kibra (FBgn0262127) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03256}attP2	Bloomington Drosophila Stock Center	BDSC:51499; FlyBase:FBst0051499;
D. melanogaster. Expresses dsRNA for RNAi of tld (FBgn0003719) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03275}attP2	Bloomington Drosophila Stock Center	BDSC:51507; FlyBase:FBst0051507;
D. melanogaster. Expresses dsRNA for RNAi of S6kII (FBgn0262866) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03140}attP40	Bloomington Drosophila Stock Center	BDSC:51694; FlyBase:FBst0051694;
D. melanogaster. Expresses dsRNA for RNAi of ast (FBgn0015905) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03173}attP2	Bloomington Drosophila Stock Center	BDSC:51700; FlyBase:FBst0051700;
D. melanogaster. Expresses dsRNA for RNAi of ras (FBgn0003204) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03250}attP2	Bloomington Drosophila Stock Center	BDSC:51717; FlyBase:FBst0051717;
D. melanogaster. Expresses dsRNA for RNAi of E(spl)mgamma-HLH (FBgn0002735) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03315}attP2	Bloomington Drosophila Stock Center	BDSC:51762; FlyBase:FBst0051762;
D. melanogaster. Expresses dsRNA for RNAi of Src64B (FBgn0262733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03327}attP40	Bloomington Drosophila Stock Center	BDSC:51772; FlyBase:FBst0051772;

D. melanogaster. Expresses dsRNA for RNAi of brk (FBgn0024250) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03345}attP2	Bloomington Drosophila Stock Center	BDSC:51789; FlyBase:FBst0051789 ;
D. melanogaster. Expresses dsRNA for RNAi of Ext2 (FBGn0029175) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03621}attP40	Bloomington Drosophila Stock Center	BDSC:52883; FlyBase:FBst0052883 ;
D. melanogaster. Expresses dsRNA for RNAi of pen-2 (FBgn0053198) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03648}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:52908; FlyBase:FBst0052908 ;
D. melanogaster. Expresses dsRNA for RNAi of Tsc1 (FBgn0026317) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03672}attP40	Bloomington Drosophila Stock Center	BDSC:52931; FlyBase:FBst0052931 ;
D. melanogaster. Expresses dsRNA for RNAi of NT1 (FBgn0261526) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21720}attP40	Bloomington Drosophila Stock Center	BDSC:53003; FlyBase:FBst0053003 ;
D. melanogaster. Expresses dsRNA for RNAi of arr (FBgn0000119) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03571}attP40	Bloomington Drosophila Stock Center	BDSC:53342; FlyBase:FBst0053342 ;
D. melanogaster. Expresses dsRNA for RNAi of kermit (FBgn0010504) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03578}attP40	Bloomington Drosophila Stock Center	BDSC:53349; FlyBase:FBst0053349 ;
D. melanogaster. Expresses dsRNA for RNAi of Sirt1 (FBgn0024291) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21708}attP40	Bloomington Drosophila Stock Center	BDSC:53697; FlyBase:FBst0053697 ;
D. melanogaster. Expresses dsRNA for RNAi of tow (FBgn0035719) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21747}attP40	Bloomington Drosophila Stock Center	BDSC:53704; FlyBase:FBst0053704 ;
D. melanogaster. Expresses dsRNA for RNAi of eIF4E3 (FBgn0265089) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21195}attP40	Bloomington Drosophila Stock Center	BDSC:53880; FlyBase:FBst0053880 ;
D. melanogaster. Expresses dsRNA for RNAi of hppy (FBgn0263395) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21199}attP40	Bloomington Drosophila Stock Center	BDSC:53884; FlyBase:FBst0053884 ;

D. melanogaster. Expresses dsRNA for RNAi of dome (FBgn0043903) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21208}attP40	Bloomington Drosophila Stock Center	BDSC:53890; FlyBase:FBst0053890;
D. melanogaster. Expresses dsRNA for RNAi of Ocho (FBgn0040296) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21588}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:54851; FlyBase:FBst0054851;
D. melanogaster. Expresses dsRNA for RNAi of sqd (FBgn0263396) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03848}attP40	Bloomington Drosophila Stock Center	BDSC:55169; FlyBase:FBst0055169;
D. melanogaster. Expresses dsRNA for RNAi of mago (FBgn0002736) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03947}attP40	Bloomington Drosophila Stock Center	BDSC:55260; FlyBase:FBst0055260;
D. melanogaster. Expresses dsRNA for RNAi of egr (FBgn0033483) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03963}attP40	Bloomington Drosophila Stock Center	BDSC:55276; FlyBase:FBst0055276;
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m3-HLH (FBgn0002609) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03989}attP2	Bloomington Drosophila Stock Center	BDSC:55302; FlyBase:FBst0055302;
D. melanogaster. Expresses dsRNA for RNAi of Coprox (FBgn0021944) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04005}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:55318; FlyBase:FBst0055318;
D. melanogaster. Expresses dsRNA for RNAi of Rab23 (FBgn0037364) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04039}attP40	Bloomington Drosophila Stock Center	BDSC:55352; FlyBase:FBst0055352;
D. melanogaster. Expresses dsRNA for RNAi of tsu (FBgn0033378) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04055}attP40	Bloomington Drosophila Stock Center	BDSC:55367; FlyBase:FBst0055367;
D. melanogaster. Expresses dsRNA for RNAi of Notum (FBgn0044028) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04067}attP40	Bloomington Drosophila Stock Center	BDSC:55379; FlyBase:FBst0055379;
D. melanogaster. Expresses dsRNA for RNAi of sd (FBgn0003345) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04092}attP40	Bloomington Drosophila Stock Center	BDSC:55404; FlyBase:FBst0055404;

D. melanogaster. Expresses dsRNA for RNAi of Raf (FBgn0003079) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03854}attP2	Bloomington Drosophila Stock Center	BDSC:55679; FlyBase:FBst0055679;
D. melanogaster. Expresses dsRNA for RNAi of Mo25 (FBGn0017572) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03865}attP2	Bloomington Drosophila Stock Center	BDSC:55681; FlyBase:FBst0055681;
D. melanogaster. Expresses dsRNA for RNAi of ptc (FBgn0003892) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03872}attP40	Bloomington Drosophila Stock Center	BDSC:55686; FlyBase:FBst0055686;
D. melanogaster. Expresses dsRNA for RNAi of Pka-C2 (FBgn0000274) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04129}attP2	Bloomington Drosophila Stock Center	BDSC:55859; FlyBase:FBst0055859;
D. melanogaster. Expresses dsRNA for RNAi of sev (FBgn0003366) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04136}attP2	Bloomington Drosophila Stock Center	BDSC:55866; FlyBase:FBst0055866;
D. melanogaster. Expresses dsRNA for RNAi of shf (FBgn0003390) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04137}attP2	Bloomington Drosophila Stock Center	BDSC:55867; FlyBase:FBst0055867;
D. melanogaster. Expresses dsRNA for RNAi of babo (FBgn0011300) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04142}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:55871; FlyBase:FBst0055871;
D. melanogaster. Expresses dsRNA for RNAi of Shark (FBgn0015295) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04146}attP2	Bloomington Drosophila Stock Center	BDSC:55874; FlyBase:FBst0055874;
D. melanogaster. Expresses dsRNA for RNAi of Drl-2 (FBgn0033791) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04172}attP2	Bloomington Drosophila Stock Center	BDSC:55893; FlyBase:FBst0055893;
D. melanogaster. Expresses dsRNA for RNAi of Takl2 (FBgn0039015) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04181}attP2	Bloomington Drosophila Stock Center	BDSC:55899; FlyBase:FBst0055899;
D. melanogaster. Expresses dsRNA for RNAi of Takl1 (FBgn0046689) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04186}attP2	Bloomington Drosophila Stock Center	BDSC:55903; FlyBase:FBst0055903;

D. melanogaster. Expresses dsRNA for RNAi of Doa (FBgn0265998) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04193}attP2	Bloomington Drosophila Stock Center	BDSC:55908; FlyBase:FBst0055908;
D. melanogaster. Expresses dsRNA for RNAi of fus (FBGn0023441) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04208}attP40	Bloomington Drosophila Stock Center	BDSC:55921; FlyBase:FBst0055921;
D. melanogaster. Expresses dsRNA for RNAi of grk (FBgn0001137) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04213}attP40	Bloomington Drosophila Stock Center	BDSC:55926; FlyBase:FBst0055926;
D. melanogaster. Expresses dsRNA for RNAi of PpD3 (FBgn0005777) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS04508}attP40	Bloomington Drosophila Stock Center	BDSC:57307; FlyBase:FBst0057307;
D. melanogaster. Expresses dsRNA for RNAi of Nct (FBgn0039234) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04812}attP40	Bloomington Drosophila Stock Center	BDSC:57497; FlyBase:FBst0057497;
D. melanogaster. Expresses dsRNA for RNAi of CG15436 (FBgn0031610) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04637}attP40	Bloomington Drosophila Stock Center	BDSC:57867; FlyBase:FBst0057867;
D. melanogaster. Expresses dsRNA for RNAi of Su(var)2-10 (FBgn0003612) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21959}attP40	Bloomington Drosophila Stock Center	BDSC:58067; FlyBase:FBst0058067;
D. melanogaster. Expresses dsRNA for RNAi of Atg8a (FBgn0052672) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ22416}attP40	Bloomington Drosophila Stock Center	BDSC:58309; FlyBase:FBst0058309;
D. melanogaster. Expresses dsRNA for RNAi of spz (FBgn0003495) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ22258}attP40	Bloomington Drosophila Stock Center	BDSC:58499; FlyBase:FBst0058499;
D. melanogaster. Expresses dsRNA for RNAi of AstC-R1 (FBgn0036790) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ23767}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:62372; FlyBase:FBst0062372;
D. melanogaster. Expresses dsRNA for RNAi of Ku80 (FBgn0041627) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ24057}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:62513; FlyBase:FBst0062513;

D. melanogaster. Expresses dsRNA for RNAi of CG2199 (FBgn0035213) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ30228}attP40	Bloomington Drosophila Stock Center	BDSC:63661; FlyBase:FBst0063661;
D. melanogaster. Expresses dsRNA for RNAi of tsr (FBgn0011726) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00534}attP2	Bloomington Drosophila Stock Center	BDSC:65055; FlyBase:FBst0065055;
D. melanogaster. Expresses dsRNA for RNAi of CG3630 (FBgn0023540) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC06220}attP2	Bloomington Drosophila Stock Center	BDSC:65945; FlyBase:FBst0065945;
D. melanogaster. Expresses dsRNA for RNAi of tub (FBgn0003882) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS05426}attP40	Bloomington Drosophila Stock Center	BDSC:66960; FlyBase:FBst0066960;
D. melanogaster. Expresses dsRNA for RNAi of eIF4E-6 (FBgn0039622) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17580
D. melanogaster. Expresses dsRNA for RNAi of gd (FBgn0000808) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14892
D. melanogaster. Expresses dsRNA for RNAi of Act88F (FBgn0000047) under UAS control.	Vienna Drosophila Resource Center	VDRC:v9780
D. melanogaster. Expresses dsRNA for RNAi of eRF1 (FBgn0036974) under UAS control.	Vienna Drosophila Resource Center	VDRC:v45027
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m2-BFM (FBgn0002592) under UAS control.	Vienna Drosophila Resource Center	VDRC:v30115
D. melanogaster. Expresses dsRNA for RNAi of tsl (FBgn0003867) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14430
D. melanogaster. Expresses dsRNA for RNAi of wbl (FBgn0004003) under UAS control.	Vienna Drosophila Resource Center	VDRC:v13864
D. melanogaster. Expresses dsRNA for RNAi of boss (FBgn0000206) under UAS control.	Vienna Drosophila Resource Center	VDRC:v4365
D. melanogaster. Expresses dsRNA for RNAi of CG32396 (FBgn0020251) under UAS control.	Vienna Drosophila Resource Center	VDRC:v41896
D. melanogaster. Expresses dsRNA for RNAi of Rac2 (FBgn0014011) under UAS control.	Vienna Drosophila Resource Center	VDRC:v28926
D. melanogaster. Expresses dsRNA for RNAi of ihog (FBgn0031872) under UAS control.	Vienna Drosophila Resource Center	VDRC:v29898
D. melanogaster. Expresses dsRNA for RNAi of sog (FBgn0003463) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37405
D. melanogaster. Expresses dsRNA for RNAi of CG9314 (FBgn0032061) under UAS control.	Vienna Drosophila Resource Center	VDRC:v44647

D. melanogaster. Expresses dsRNA for RNAi of sgl (FBgn0261445) under UAS control.	Vienna Drosophila Resource Center	VDRC:v29434
D. melanogaster. Expresses dsRNA for RNAi of mirr (FBgn0014343) under UAS control.	Vienna Drosophila Resource Center	VDRC:v50134
D. melanogaster. Expresses dsRNA for RNAi of eIF-4B (FBgn0020660) under UAS control.	Vienna Drosophila Resource Center	VDRC:v31364
D. melanogaster. Expresses dsRNA for RNAi of rasp (FBgn0024194) under UAS control.	Vienna Drosophila Resource Center	VDRC:v6459
D. melanogaster. Expresses dsRNA for RNAi of PGRP-SA (FBgn0030310) under UAS control.	Vienna Drosophila Resource Center	VDRC:v5594
D. melanogaster. Expresses dsRNA for RNAi of sinah (FBgn0259794) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17118
D. melanogaster. Expresses dsRNA for RNAi of lft (FBgn0032230) under UAS control.	Vienna Drosophila Resource Center	VDRC:v32146
D. melanogaster. Expresses dsRNA for RNAi of pyr (FBgn0033649) under UAS control.	Vienna Drosophila Resource Center	VDRC:v36524
D. melanogaster. Expresses dsRNA for RNAi of Ssl (FBgn0015300) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17282
D. melanogaster. Expresses dsRNA for RNAi of Pvf2 (FBgn0031888) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7628
D. melanogaster. Expresses dsRNA for RNAi of spz4 (FBgn0032362) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7679
D. melanogaster. Expresses dsRNA for RNAi of IM23 (FBgn0034328) under UAS control.	Vienna Drosophila Resource Center	VDRC:v15384
D. melanogaster. Expresses dsRNA for RNAi of botv (FBgn0027535) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37186
D. melanogaster. Expresses dsRNA for RNAi of G6P (FBgn0031463) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7261
D. melanogaster. Expresses dsRNA for RNAi of stet (FBgn0020248) under UAS control.	Vienna Drosophila Resource Center	VDRC:v7434
D. melanogaster. Expresses dsRNA for RNAi of RanBPM (FBgn0262114) under UAS control.	Vienna Drosophila Resource Center	VDRC:v45981
D. melanogaster. Expresses dsRNA for RNAi of Src42A (FBgn0264959) under UAS control.	Vienna Drosophila Resource Center	VDRC:v26019
D. melanogaster. Expresses dsRNA for RNAi of nkd (FBgn0002945) under UAS control.	Vienna Drosophila Resource Center	VDRC:v3004
D. melanogaster. Expresses dsRNA for RNAi of boi (FBgn0040388) under UAS control.	Vienna Drosophila Resource Center	VDRC:v3060
D. melanogaster. Expresses dsRNA for RNAi of CR45683; Tehao (FBgn0026760) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17903

D. melanogaster. Expresses dsRNA for RNAi of CG45087; Pepck (FBgn0003067) under UAS control.	Vienna Drosophila Resource Center	VDRC:v20529
D. melanogaster. Expresses dsRNA for RNAi of RpL13A (FBgn0037351) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101369
D. melanogaster. Expresses dsRNA for RNAi of Usp7 (FBgn0030366) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110324
D. melanogaster. Expresses dsRNA for RNAi of tsg (FBgn0003865) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108750
D. melanogaster. Expresses dsRNA for RNAi of nec (FBgn0002930) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108366
D. melanogaster. Expresses dsRNA for RNAi of Nle (FBgn0021874) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110728
D. melanogaster. Expresses dsRNA for RNAi of Brd (FBgn0000216) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107929
D. melanogaster. Expresses dsRNA for RNAi of Shc (FBgn0015296) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103906
D. melanogaster. Expresses dsRNA for RNAi of Hs6st (FBgn0038755) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110424
D. melanogaster. Expresses dsRNA for RNAi of fz4 (FBgn0027342) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102339
D. melanogaster. Expresses dsRNA for RNAi of bib (FBgn0000180) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103327
D. melanogaster. Expresses dsRNA for RNAi of Wnt10 (FBgn0031903) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100867
D. melanogaster. Expresses dsRNA for RNAi of Tom (FBgn0026320) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101652
D. melanogaster. Expresses dsRNA for RNAi of Pli (FBgn0025574) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106776
D. melanogaster. Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
D. melanogaster. Expresses dsRNA for RNAi of por (FBgn0004957) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100780
D. melanogaster. Expresses dsRNA for RNAi of wls (FBgn0036141) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103812
D. melanogaster. Expresses dsRNA for RNAi of CG6843 (FBgn0036827) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109411
D. melanogaster. Expresses dsRNA for RNAi of spz3 (FBgn0031959) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102871
D. melanogaster. Expresses dsRNA for RNAi of kuz (FBgn0259984) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107036

D. melanogaster. Expresses dsRNA for RNAi of Hs3st-B (FBgn0031005) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110601
D. melanogaster. Expresses dsRNA for RNAi of Tace (FBgn0039734) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106335
D. melanogaster. Expresses dsRNA for RNAi of eIF-1A (FBgn0026250) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100611
D. melanogaster. Expresses dsRNA for RNAi of Socs16D (FBgn0030869) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100568
D. melanogaster. Expresses dsRNA for RNAi of E(spl)malpha-BFM (FBgn0002732) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109384
D. melanogaster. Expresses dsRNA for RNAi of E(spl)m6-BFM (FBgn0002632) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101965
D. melanogaster. Expresses dsRNA for RNAi of VhaM8.9 (FBgn0037671) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105281
D. melanogaster. Expresses dsRNA for RNAi of melt (FBgn0023001) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105110
D. melanogaster. Expresses dsRNA for RNAi of SkpB (FBgn0026176) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106521
D. melanogaster. Expresses dsRNA for RNAi of CkIIbeta2 (FBgn0026136) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102633
D. melanogaster. Expresses dsRNA for RNAi of spz6 (FBgn0035056) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100897
D. melanogaster. Expresses dsRNA for RNAi of Rok (FBgn0026181) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104675
D. melanogaster. Expresses dsRNA for RNAi of CG9962 (FBgn0031441) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108721
D. melanogaster. Expresses dsRNA for RNAi of spz5 (FBgn0035379) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102389
D. melanogaster. Expresses dsRNA for RNAi of Act57B (FBgn0000044) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102129
D. melanogaster. Expresses dsRNA for RNAi of ndl (FBgn0002926) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102818
D. melanogaster. Expresses dsRNA for RNAi of vn (FBgn0003984) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109437
D. melanogaster. Expresses dsRNA for RNAi of ECSIT (FBgn0028436) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106141
D. melanogaster. Expresses dsRNA for RNAi of SkpE (FBgn0031074) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109539
D. melanogaster. Expresses dsRNA for RNAi of SkpF (FBgn0034863) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106572

D. melanogaster. Expresses dsRNA for RNAi of kek1 (FBgn0015399) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101166
D. melanogaster. Expresses dsRNA for RNAi of ths (FBgn0033652) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102441
D. melanogaster. Expresses dsRNA for RNAi of Ilp8 (FBgn0036690) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102604
D. melanogaster. Expresses dsRNA for RNAi of CG15800 (FBgn0034904) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110049
D. melanogaster. Expresses dsRNA for RNAi of IM3 (FBgn0040736) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104908
D. melanogaster. Expresses dsRNA for RNAi of Roc1a (FBgn0025638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106315
D. melanogaster. Expresses dsRNA for RNAi of dod (FBgn0015379) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110593
D. melanogaster. Expresses dsRNA for RNAi of hipk (FBgn0035142) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108254
D. melanogaster. Expresses dsRNA for RNAi of ave (FBgn0050476) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101471
D. melanogaster. Expresses dsRNA for RNAi of boca (FBgn0004132) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108406
D. melanogaster. Expresses dsRNA for RNAi of gskt (FBgn0046332) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107429
D. melanogaster. Expresses dsRNA for RNAi of stumps (FBgn0020299) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105603
D. melanogaster. Expresses dsRNA for RNAi of CG31431 (FBgn0051431) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104697
D. melanogaster. Expresses dsRNA for RNAi of scw (FBgn0005590) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105303
D. melanogaster. Expresses dsRNA for RNAi of fry (FBgn0016081) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103569
D. melanogaster. Expresses dsRNA for RNAi of Krn (FBgn0052179) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104299
D. melanogaster. Expresses dsRNA for RNAi of pxb (FBgn0053207) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102240
D. melanogaster. Expresses dsRNA for RNAi of cv-c (FBgn0285955) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105435
D. melanogaster. Expresses dsRNA for RNAi of cic (FBgn0262582) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103805
D. melanogaster. Expresses dsRNA for RNAi of dia (FBgn0011202) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103914

D. melanogaster. Expresses dsRNA for RNAi of SkpD;SkpC (FBgn0026174) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109181
D. melanogaster. Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
D. melanogaster. Expresses dsRNA for RNAi of botv (FBgn0027535) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37186

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Reagents

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Name	Catalogue number	Usage
Hoechst 33321	H1399 (ThermoFisher)	1ug/ml final concentration

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References

Ables, E.T., Drummond-Barbosa, D., 2017. Steroid Hormones and the Physiological Regulation of Tissue-Resident Stem Cells: Lessons from the *Drosophila* Ovary. Current Stem Cell Reports 3, 9-18.

Albert, I., Albert, R., 2004. Conserved network motifs allow protein-protein interaction prediction. Bioinformatics 20, 3346-3352.

Albert, R., Barabási, A.-L., 2002. Statistical mechanics of complex networks. Reviews of Modern Physics 74, 47-97.

Allbee, A.W., Rincon-Limas, D.E., Biteau, B., 2018. *Lmx1a* is required for the development of the ovarian stem cell niche in *Drosophila*. Development 145, dev163394.

Azevedo, R.B.R., French, V., Partridge, L., 1996. Thermal evolution of egg size in *Drosophila melanogaster*. Evolution 50, 2338-2345.

Barabási, A.L., Albert, R., 1999. Emergence of scaling in random networks. Science 286, 509-512.

Barabási, A.L., Oltvai, Z.N., 2004. Network biology: understanding the cell's functional organization. Nat. Rev. Genet. 5, 101-113.

Barrett, K., Leptin, M., Settleman, J., 1997. The Rho GTPase and a putative RhoGEF mediate a signaling pathway for the cell shape changes in *Drosophila* gastrulation. Cell 91, 905-915.

Berger, S.I., Posner, J.M., Ma'ayan, A., 2007. Genes2Networks: connecting lists of gene symbols using mammalian protein interactions databases. BMC Bioinformatics 8, 372.

Bergland, A.O., Genissel, A., Nuzhdin, S.V., Tatar, M., 2008. Quantitative trait loci affecting phenotypic plasticity and the allometric relationship of ovariole number and thorax length in *Drosophila melanogaster*. Genetics 180, 567-582.

Bolívar, J., Pearson, J., López-Onieva, L., González-Reyes, A., 2006. Genetic dissection of a stem cell niche: the case of the *Drosophila* ovary. Dev. Dyn. 235, 2969-2979.

Bromberg, K.D., Ma'ayan, A., Neves, S.R., Iyengar, R., 2008. Design logic of a cannabinoid receptor signaling network that triggers neurite outgrowth. Science 320, 903-909.

Capy, P., Pla, E., David, J.R., 1993. Phenotypic and genetic variability of morphometrical traits in natural populations of *Drosophila melanogaster* and *D. simulans.* I. Geographic variations. Genet. Sel. Evol. 25, 517-536.

Capy, P., Pla, E., David, J.R., 1994. Phenotypic and genetic variability of morphometrical traits in natural populations of *Drosophila melanogaster* and *D simulans*. II. Within-population variability. Genetics Selection Evolution 26, 15-28.

Cayirlioglu, P., Ward, W.O., Silver Key, S.C., Duronio, R.J., 2003. Transcriptional repressor functions of *Drosophila* E2F1 and E2F2 cooperate to inhibit genomic DNA synthesis in ovarian follicle cells. Mol. Cell. Biol. 23, 2123-2134.

Chao, A.T., Dierick, H.A., Addy, T.M., Bejsovec, A., 2003. Mutations in eukaryotic release
factors 1 and 3 act as general nonsense suppressors in *Drosophila*. Genetics 165, 601-612.

Chen, J., Godt, D., Gunsalus, K., Kiss, I., Goldberg, M., Laski, F.A., 2001. Cofilin/ADF is
required for cell motility during *Drosophila* ovary development and oogenesis. Nature Cell Biol.
3, 204-209.

Chen, J.Y., Shen, C., Sivachenko, A.Y., 2006. Mining Alzheimer disease relevant proteins from
integrated protein interactome data. Pacific Symposium on Biocomputing 11, 367-378.

Christiansen, A.E., Ding, T., Bergmann, A., 2012. Ligand-independent activation of the
Hedgehog pathway displays non-cell autonomous proliferation during eye development in
Drosophila. Mech. Dev. 129, 98-108.

Cohet, Y., David, J.R., 1978. Control of the Adult reproductive Potential by Preimaginal thermal
Conditions. Oecologia 36, 295-306.

Couderc, J.L., Godt, D., Zollman, S., Chen, J., Li, M., Tiong, S., Cramton, S.E., Sahut-Barnola,
I., Laski, F.A., 2002. The *bric a brac* locus consists of two paralogous genes encoding BTB/POZ
domain proteins and acts as a homeotic and morphogenetic regulator of imaginal development
in *Drosophila*. Development 129, 2419-2433.

David, J.R., Bocquet, C., 1975. Similarities and differences in latitudinal adaptation of two
Drosophila sibling species. Nature 257, 588-590.

Dobens, L., Jaeger, A., Peterson, J.S., Raftery, L.A., 2005. Bunched sets a boundary for Notch
signaling to pattern anterior eggshell structures during *Drosophila* oogenesis. Dev. Biol. 287,
425-437.

Edwards, M.C., Wong, C., Elledge, S.J., 1998. Human cyclin K, a novel RNA polymerase II-
associated cyclin possessing both carboxy-terminal domain kinase and Cdk-activating kinase
activity. Mol. Cell. Biol. 18, 4291-4300.

Foster, L.J., de Hoog, C.L., Zhang, Y., Zhang, Y., Xie, X., Mootha, V.K., Mann, M., 2006. A
mammalian organelle map by protein correlation profiling. Cell 125, 187-199.

Gancz, D., Gilboa, L., 2013. Insulin and Target of rapamycin signaling orchestrate the
development of ovarian niche-stem cell units in *Drosophila*. Development 140, 4145-4154.

Gancz, D., Lengil, T., Gilboa, L., 2011. Coordinated regulation of niche and stem cell precursors
by hormonal signaling. PLoS Biol. 9, e1001202.

Gilboa, L., 2015. Organizing stem cell units in the *Drosophila* ovary. Curr. Op. Genet. Dev. 32C,
31-36.

Giot, L., Bader, J.S., Brouwer, C., Chaudhuri, A., Kuang, B., Li, Y., Hao, Y.L., Ooi, C.E.,
Godwin, B., Vitols, E., Vijayadamodar, G., Pochart, P., Machineni, H., Welsh, M., Kong, Y.,
Zerhusen, B., Malcolm, R., Varrone, Z., Collis, A., Minto, M., Burgess, S., McDaniel, L.,
Stimpson, E., Spriggs, F., Williams, J., Neurath, K., Ioime, N., Agee, M., Voss, E., Furtak, K.,
Renzulli, R., Aanensen, N., Carrolla, S., Bickelhaupt, E., Lazovatsky, Y., DaSilva, A., Zhong, J.,
Stanyon, C.A., Finley, R.L., Jr., White, K.P., Braverman, M., Jarvie, T., Gold, S., Leach, M.,

Knight, J., Shimkets, R.A., McKenna, M.P., Chant, J., Rothberg, J.M., 2003. A protein interaction map of *Drosophila melanogaster*. Science 302, 1727-1736.

Godt, D., Couderc, J.L., Cramton, S.E., Laski, F.A., 1993. Pattern formation in the limbs of *Drosophila*: *bric à brac* is expressed in both a gradient and a wave-like pattern and is required for specification and proper segmentation of the tarsus. Development 119, 799-812.

Godt, D., Laski, F.A., 1995. Mechanisms of cell rearrangement and cell recruitment in *Drosophila* ovary morphogenesis and the requirement of *bric à brac*. Development 121, 173-187.

Gonzalez, G., Uribe, J.C., Tari, L., Brophy, C., Baral, C., 2007. Mining gene-disease relationships from biomedical literature: weighting protein-protein interactions and connectivity measures. Pacific Symposium on Biocomputing 12, 28-39.

Gonzalez, M.W., Kann, M.G., 2012. Chapter 4: Protein interactions and disease. PLoS Comput Biol 8, e1002819.

Gramates, L.S., Marygold, S.J., Santos, G.D., Urbano, J.M., Antonazzo, G., Matthews, B.B., Rey, A.J., Tabone, C.J., Crosby, M.A., Emmert, D.B., Falls, K., Goodman, J.L., Hu, Y., Ponting, L., Schroeder, A.J., Strelets, V.B., Thurmond, J., Zhou, P., the FlyBase, C., 2016. FlyBase at 25: looking to the future. Nucleic Acids Res. 45, D663-D671.

Green II, D.A., Extavour, C.G., 2012. Convergent Evolution of a Reproductive Trait Through Distinct Developmental Mechanisms in *Drosophila*. Dev. Biol. 372, 120-130.

Green II, D.A., Sarikaya, D.P., Extavour, C.G., 2011. Counting in oogenesis. Cell Tissue Res. 344, 207-212.

Hahn, M.W., Kern, A.D., 2005. Comparative genomics of centrality and essentiality in three eukaryotic protein-interaction networks. Mol. Biol. Evol. 22, 803-806.

Harris, T.J., Peifer, M., 2004. Adherens junction-dependent and -independent steps in the establishment of epithelial cell polarity in *Drosophila*. J. Cell Biol. 167, 135-147.

Hartwell, L.H., Hopfield, J.J., Leibler, S., Murray, A.W., 1999. From molecular to modular cell biology. Nature 402, C47-C52.

Hilman, D., Gat, U., 2011. The evolutionary history of YAP and the hippo/YAP pathway. Mol. Biol. Evol. 28, 2403-2417.

Hodin, J., Riddiford, L.M., 1998. The ecdysone receptor and ultraspiracle regulate the timing and progression of ovarian morphogenesis during *Drosophila* metamorphosis. Dev. Genes Evol. 208, 304-317.

Hodin, J., Riddiford, L.M., 2000a. Different mechanisms underlie phenotypic plasticity and interspecific variation for a reproductive character in Drosophilids (Insecta: Diptera). Evolution 5, 1638-1653.

Hodin, J., Riddiford, L.M., 2000b. Parallel alterations in the timing of ovarian ecdysone receptor
and ultraspiracle expression characterize the independent evolution of larval reproduction in two
species of gall midges (Diptera: Cecidomyiidae). Dev. Genes Evol. 210, 358-372.

Hsu, H.-J., Drummond-Barbosa, D., 2009. Insulin levels control female germline stem cell
maintenance via the niche in *Drosophila*. Proc. Natl. Acad. Sci. USA 106, 1117-1121.

Huang, S.S., Fraenkel, E., 2009. Integrating proteomic, transcriptional, and interactome data
reveals hidden components of signaling and regulatory networks. Science Signaling 2, ra40.

Ideker, T., Sharan, R., 2008. Protein networks in disease. Genome Res. 18, 644-652.

Ito, T., Bulger, M., Kobayashi, R., Kadonaga, J.T., 1996. *Drosophila* NAP-1 is a core histone
chaperone that functions in ATP-facilitated assembly of regularly spaced nucleosomal arrays.
Mol. Cell. Biol. 16, 3112-3124.

Jalili, M., Salehzadeh-Yazdi, A., Gupta, S., Wolkenhauer, O., Yaghmaie, M., Resendis-Antonio,
O., Alimoghaddam, K., 2016. Evolution of Centrality Measurements for the Detection of
Essential Proteins in Biological Networks. Frontiers in Physiology 7, 375.

Jansen, R., Greenbaum, D., Gerstein, M., 2002. Relating whole-genome expression data with
protein-protein interactions. Genome Res. 12, 37-46.

Jeong, H., Mason, S.P., Barabási, A.L., Oltvai, Z.N., 2001. Lethality and centrality in protein
networks. Nature 411, 41-42.

Jiang, K., Liu, Y., Zhang, J., Jia, J., 2018. An intracellular activation of Smoothened that is
independent of Hedgehog stimulation in *Drosophila*. J. Cell Sci. 131, jcs211367.

Kambysellis, M.P., Heed, W.B., 1971. Studies of Oogenesis in Natural Populations of
Drosophilidae. I. Relation of ovarian development and ecological habitats of the Hawaiian
species. Am. Nat. 941, 31-49.

Kanehisa, M., Goto, S., Furumichi, M., Tanabe, M., Hirakawa, M., 2010. KEGG for
representation and analysis of molecular networks involving diseases and drugs. Nucleic Acids
Res. 38, D355-360.

Kasza, K.E., Farrell, D.L., Zallen, J.A., 2014. Spatiotemporal control of epithelial remodeling by
regulated myosin phosphorylation. Proc. Natl. Acad. Sci. USA 111, 11732-11737.

Keller, R., 2006. Mechanisms of elongation in embryogenesis. Development 133, 2291-2302.

King, R.C., 1970. Ovarian Development in *Drosophila melanogaster*. Academic Press, New
York.

King, R.C., Aggarwal, S.K., Aggarwal, U., 1968. The Development of the Female *Drosophila*
Reproductive System. J. Morphol. 124, 143-166.

Klepsatel, P., Galikova, M., De Maio, N., Huber, C.D., Schlotterer, C., Flatt, T., 2013a. Variation
in thermal performance and reaction norms among populations of *Drosophila melanogaster*.
Evolution 67, 3573-3587.

Klepsatel, P., Galikova, M., De Maio, N., Ricci, S., Schlotterer, C., Flatt, T., 2013b. Reproductive
and post-reproductive life history of wild-caught *Drosophila melanogaster* under laboratory
conditions. J. Evol. Biol. 26, 1508-1520.

Koschutzki, D., Schreiber, F., 2008. Centrality analysis methods for biological networks and
their application to gene regulatory networks. Gene Regulation and Systems Biology 2, 193-
201.

LaFever, L., Drummond-Barbosa, D., 2005. Direct control of germline stem cell division and cyst
growth by neural insulin in *Drosophila*. Science 309, 1071-1073.

LaFever, L., Feoktistov, A., Hsu, H.J., Drummond-Barbosa, D., 2010. Specific roles of Target of
rapamycin in the control of stem cells and their progeny in the *Drosophila* ovary. Development
137, 2117-2126.

Lai, C.M., Lin, K.Y., Kao, S.H., Chen, Y.N., Huang, F., Hsu, H.J., 2017. Hedgehog signaling
establishes precursors for germline stem cell niches by regulating cell adhesion. J. Cell Biol.
216, 1439-1453.

Lee, R.T., Zhao, Z., Ingham, P.W., 2016. Hedgehog signalling. Development 143, 367-372.

Lewellyn, L., Cetera, M., Horne-Badovinac, S., 2013. Misshapen decreases integrin levels to
promote epithelial motility and planar polarity in *Drosophila*. J. Cell Biol. 200, 721-729.

Li, X., Wu, M., Kwoh, C.K., Ng, S.K., 2010. Computational approaches for detecting protein
complexes from protein interaction networks: a survey. BMC Genomics 11 Suppl 1, S3.

Llamazares, S., Moreira, A., Tavares, A., Girdham, C., Spruce, B.A., Gonzalez, C., Karess,
R.E., Glover, D.M., Sunkel, C.E., 1991. *polo* encodes a protein kinase homolog required for
mitosis in *Drosophila*. Genes Dev. 5, 2153-2165.

Lobell, A.S., Kaspari, R.R., Serrano Negron, Y.L., Harbison, S.T., 2017. The Genetic
Architecture of Ovariole Number in *Drosophila melanogaster*: Genes with Major, Quantitative,
and Pleiotropic Effects. G3 7, 2391-2403.

Ma'ayan, A., 2011. Introduction to network analysis in systems biology. Sci Signal 4, tr5.

Markow, T.A., Beall, S., Matzkin, L.M., 2009. Egg size, embryonic development time and
ovoviviparity in *Drosophila* species. J. Evol. Biol. 22, 430-434.

Matthews, L.R., Vaglio, P., Reboul, J., Ge, H., Davis, B.P., Garrels, J., Vincent, S., Vidal, M.,
2001. Identification of potential interaction networks using sequence-based searches for
conserved protein-protein interactions or "interologs". Genome Res. 11, 2120-2126.

Mbodj, A., Junion, G., Brun, C., Furlong, E.E., Thieffry, D., 2013. Logical modelling of
Drosophila signalling pathways. Molecular BioSystems 9, 2248-2258.

Mizuno, T., Amano, M., Kaibuchi, K., Nishida, Y., 1999. Identification and characterization of
Drosophila homolog of Rho-kinase. Gene 238, 437-444.

Orgogozo, V., Broman, K.W., Stern, D.L., 2006. High-resolution quantitative trait locus mapping
reveals sign epistasis controlling ovariole number between two *Drosophila* species. Genetics
173, 197-205.

R'Kha, S., Capy, P., David, J.R., 1991. Host-plant specialization in the *Drosophila melanogaster*
species complex: A physiological, behavioral, and genetic analysis. Proc. Natl. Acad. Sci. USA
88, 1835-1839.

R´kha, S., Moreteau, B., Coyne, J.A., David, J.R., 1997. Evolution of a lesser fitness trait: egg
production in the specialist *Drosophila sechellia*. Genetical research 69, 17-23.

Ravasz, E., Somera, A.L., Mongru, D.A., Oltvai, Z.N., Barabási, A.L., 2002. Hierarchical
organization of modularity in metabolic networks. Science 297, 1551-1555.

Ridley, A.J., 2006. Rho GTPases and actin dynamics in membrane protrusions and vesicle
trafficking. Trends Cell Biol. 16, 522-529.

Sahut-Barnola, I., Dastugue, B., Couderc, J.-L., 1996. Terminal filament cell organization in the
larval ovary of *Drosophila melanogaster*: ultrastructural observations and pattern of divisions.
Roux's Archives of Developmental Biology 205, 356-363.

Sahut-Barnola, I., Godt, D., Laski, F.A., Couderc, J.-L., 1995. *Drosophila* Ovary Morphogenesis:
Analysis of Terminal Filament Formation and Identification of a Gene Required for This Process.
Dev. Biol. 170, 127-135.

Sarikaya, D.P., Belay, A.A., Ahuja, A., Green II, D.A., Dorta, A., Extavour, C.G., 2012. The roles
of cell size and cell number in determining ovariole number in *Drosophila*. Dev. Biol. 363, 279-
289

Sarikaya, D.P., Church, S.H., Lagomarsino, L.P., Magnacca, K.M., Montgomery, S.L., Price,
D.P., Kaneshiro, K.Y., Extavour, C.G., 2019. Reproductive capacity evolves in response to
ecology through common developmental mechanisms in Hawai'ian *Drosophila*. Curr. Biol. 29,
1877-1884.

Sarikaya, D.P., Extavour, C.G., 2015. The Hippo pathway regulates homeostatic growth of stem
cell niche precursors in the *Drosophila* ovary. PLoS Genetics 11, e1004962.

Sebe-Pedros, A., Zheng, Y., Ruiz-Trillo, I., Pan, D., 2012. Premetazoan origin of the hippo
signaling pathway. Cell Reports 1, 13-20.

Spellman, P.T., Sherlock, G., Zhang, M.Q., Iyer, V.R., Anders, K., Eisen, M.B., Brown, P.O.,
Botstein, D., Futcher, B., 1998. Comprehensive identification of cell cycle-regulated genes of the
yeast *Saccharomyces cerevisiae* by microarray hybridization. Mol. Biol. Cell 9, 3273-3297.

Srinivasan, B.S., Shah, N.H., Flannick, J.A., Abeliuk, E., Novak, A.F., Batzoglou, S., 2007.
Current progress in network research: toward reference networks for key model organisms.
Brief. Bioinform. 8, 318-332.

von Mering, C., Krause, R., Snel, B., Cornell, M., Oliver, S.G., Fields, S., Bork, P., 2002.
Comparative assessment of large-scale data sets of protein-protein interactions. Nature 417,
399-403.

Wachi, S., Yoneda, K., Wu, R., 2005. Interactome-transcriptome analysis reveals the high
centrality of genes differentially expressed in lung cancer tissues. Bioinformatics 21, 4205-4208.

Wang, R.S., Hall, K.T., Giulianini, F., Passow, D., Kaptchuk, T.J., Loscalzo, J., 2017. Network
analysis of the genomic basis of the placebo effect. JCI Insight 2, 93911.

Wang, R.S., Loscalzo, J., 2018. Network-Based Disease Module Discovery by a Novel Seed
Connector Algorithm with Pathobiological Implications. J. Mol. Biol. 430, 2939-2950.

Watts, D.J., Strogatz, S.H., 1998. Collective dynamics of 'small-world' networks. Nature 393,
440-442.

Wayne, M.L., Hackett, J.B., Dilda, C.L., Nuzhdin, S.V., Pasyukova, E.G., Mackay, T.F., 2001.
Quantitative trait locus mapping of fitness-related traits in *Drosophila melanogaster*. Genetical
research 77, 107-116.

Wayne, M.L., Hackett, J.B., Mackay, T.F.C., 1997. Quantitative Genetics of Ovariole Number in
Drosophila melanogaster. I. Segregating Variation and Fitness. Evolution 4, 1156-1163.

Wayne, M.L., McIntyre, L.M., 2002. Combining mapping and arraying: An approach to candidate
gene identification. Proc. Natl. Acad. Sci. USA 99, 14903-14906.

Yang, X., Xu, T., 2011. Molecular mechanism of size control in development and human
diseases. Cell Research 21, 715-729.

Yook, S.H., Oltvai, Z.N., Barabási, A.L., 2004. Functional and topological characterization of
protein interaction networks. Proteomics 4, 928-942.

Yu, H., Paccanaro, A., Trifonov, V., Gerstein, M., 2006. Predicting interactions in protein
networks by completing defective cliques. Bioinformatics 22, 823-829.

Zhang, Y., Lin, H., Yang, Z., Wang, J., Liu, Y., 2017. An uncertain model-based approach for
identifying dynamic protein complexes in uncertain protein-protein interaction networks. BMC
Genomics 18, 743.

1290