1 Topology-driven analysis of protein-protein interaction

2 networks detects functional genetic modules regulating

3 reproductive capacity

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10 Abstract

11 Understanding the genetic regulation of organ structure is a fundamental problem in

12 developmental biology. Here, we use egg-producing structures of insect ovaries, called

13 ovarioles, to deduce systems-level gene regulatory relationships from quantitative functional

14 genetic analysis. We previously showed that Hippo signalling, a conserved regulator of animal

15 organ size, regulates ovariole number in Drosophila melanogaster. To comprehensively

16 determine how Hippo signalling interacts with other pathways in this regulation, we screened all

17 known signalling pathway genes, and identified Hpo-dependent and Hpo-independent signalling

18 requirements. Network analysis of known protein-protein interactions among screen results

19 identified independent gene regulatory modules regulating one or both of ovariole number and

20 egg laying. These modules predict involvement of previously uncharacterised genes with higher

21 accuracy than the original candidate screen. This shows that network analysis combining

functional genetic and large-scale interaction data can predict function of novel genes regulating

- 23 development.
- 24

Keywords : *Drosophila melanogaster*, Reproduction, Ovariole, Ovary, Egg laying, Topology,
 Network analysis, Interactome, Hippo signalling.

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Introduction 29

30 The final shape and size of an organ is critical to organismal function and viability. Defects in 31 human organ morphology cause a multitude of pathologies, including cancers, organ 32 hypertrophies and atrophies (e.g. Yang and Xu, 2011). It is thus critical to understand the 33 regulatory mechanisms underlying the stereotypic shape and size of organs. To this end, 34 assessing the genetic regulation of size is significantly facilitated by using quantifiable changes in organ size and shape. 35

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37 The Drosophila melanogaster female reproductive system is a useful paradigm to study 38 quantitative anatomical traits. In these organs, the effects of multiple genes and the environment 39 combine to produce a quantitative phenotype: a species-specific average number of egg-40 producing ovarian tubes called ovarioles. Fruit fly ovaries can contain as few as one and as 41 many as 50 ovarioles per ovary, depending on the species (Kambysellis and Heed, 1971; King, 42 1970; Markow et al., 2009; Sarikaya et al., 2019), with each ovariole capable of producing eggs. 43 Ovariole number, therefore, may affect the reproductive fitness of *Drosophila* species by 44 determining the potential of an adult female to produce eggs (Klepsatel et al., 2013b; R'kha et 45 al., 1997). While ovariole number within a species can vary across temperatures (Azevedo et 46 al., 1996), altitudinal and latitudinal clines (Capy et al., 1994; David and Bocquet, 1975), under 47 constant environmental conditions ovariole number is highly stereotypic (Capy et al., 1993; 48 Klepsatel et al., 2013a; R'Kha et al., 1991; R'kha et al., 1997). The reproducibility of ovariole 49 number thus indicates a strong genetic component (Sarikaya et al., 2019). Genome wide 50 association studies and quantitative trait locus mapping have demonstrated that the ovariole 51 number is a highly polygenic trait (Bergland et al., 2008; Lobell et al., 2017; Orgogozo et al., 52 2006; Wayne et al., 2001; Wayne et al., 1997; Wayne and McIntyre, 2002). In contrast, 53 functional genetic studies have identified only a small number of genes whose activity regulates 54 ovariole number (discussed below). Thus, the complexity of the genetic regulation of this 55 important trait remains largely unknown. 56

57 The determination of ovariole number in *D. melanogaster* occurs during late larval and pupal 58 development (King et al., 1968). Each ovariole in the adult fly arises from a single primordial 59 structure called a terminal filament (TF), which forms in the late third instar larval ovary (Godt 60 and Laski, 1995) by convergent extension (Keller, 2006) of the terminal filament cells (TFCs) 61 (Godt and Laski, 1995; Sahut-Barnola et al., 1996). TFCs are first specified from an anterior

62 population of somatic cells in the larval ovary by the expression of transcription factors including 63 Bric-à-brac 1/2 (bric-à-brac 1/2; bab1/2) and Engrailed (engrailed; en) (Godt and Laski, 1995) 64 (Sahut-Barnola et al., 1995). Initially a loosely arranged group in the anterior of the larval ovary, 65 TFCs undergo morphogenetic movements to give rise to the ordered columns of cells that are 66 TFs. Cell intercalation during convergent extension is dependent on the actin regulators Cofilin 67 (twinstar) and the large Maf factor Traffic Jam (traffic jam; tj), and on E-cadherin dependent 68 adhesion (Chen et al., 2001; Godt and Laski, 1995). Regulation of ovariole number is thus 69 largely dependent on the specification of the TFCs and their rearrangement into TFs (Sarikaya 70 and Extavour, 2015).

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72 We previously showed that the regulation of both TFC and TF number is dependent on the 73 Hippo signalling pathway (Sarikaya and Extavour, 2015), a pan-metazoan regulator of organ and tissue size (Hilman and Gat, 2011; Sebe-Pedros et al., 2012). At the core of the Hippo 74 75 kinase cascade are two protein kinases, Hippo (hippo; hpo) and Warts (warts), which prevent 76 the nuclear localisation of the transcriptional co-activator Yorkie (yorkie; yki). Yki and the 77 transcription factor Scalloped (scalloped) together initiate the transcription of multiple gene 78 targets, including those that promote cell proliferation and survival. In the D. melanogaster larval 79 ovary, loss of Hpo in the somatic cells causes an increase in nuclear Yki, leading to an increase 80 in TFCs, TFs, ovariole number and egg laying in adults (Sarikaya and Extavour, 2015).

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82 Production of fertile eggs from a stereotypic number of ovarioles requires a spatially and 83 temporally coordinated interplay of signalling between the somatic and germ line cells of the 84 ovary. Thus, signalling amongst somatic and germ line cells in the larval ovary is crucial to all 85 stages of ovarian development (Ables and Drummond-Barbosa, 2017; Gilboa, 2015; Green II et 86 al., 2011; LaFever and Drummond-Barbosa, 2005; LaFever et al., 2010; Sarikaya and Extavour, 87 2015). For instance, disruptions in insulin or Tor signalling affect both somatic and germ line cell 88 proliferation (Gancz and Gilboa, 2013; Green II and Extavour, 2012; Hsu and Drummond-89 Barbosa. 2009: LaFever and Drummond-Barbosa. 2005: LaFever et al., 2010: Sarikava et al., 90 2012). Similarly, ecdysone pulses from the prothoracic gland regulate the timely differentiation 91 of the primordial germ cells (PGCs) and the somatic TFCs (Gancz et al., 2011; Hodin and 92 Riddiford, 1998, 2000b). Both Hpo and ecdysone signalling also control the proportion of germ 93 line to somatic cells by differentially regulating proliferation of both cell types (Gancz et al., 94 2011; Sarikaya and Extavour, 2015).

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96 Although it is clear that genes function together in regulatory networks (Gonzalez and Kann, 97 2012), determining how the few genes functionally verified as required for ovariole development 98 and function, work together to coordinate ovariole number and ovarian function more generally, 99 is a challenge because most genes or pathways have been considered individually. An 100 alternative approach that is less often applied to animal developmental genetics, is a systems 101 biology representation of complex biological systems as networks (Barabási and Albert, 1999; 102 Watts and Strogatz, 1998). Protein-protein interaction (PPI) networks are such an example 103 (Albert and Barabási, 2002). The availability of high throughput molecular biology datasets from, 104 for example, yeast two-hybrid, protein CHiP and microarrays has allowed for the emergence of 105 large scale interaction networks representing both functional and physical molecular interactions 106 (Barabási and Oltvai, 2004; Berger et al., 2007; Giot et al., 2003; Gonzalez and Kann, 2012). 107 108 With ample evidence that signalling in the ovary can affect ovarian development, but few genes 109 functionally verified to date, we aimed to identify novel regulators of ovariole development by 110 functionally testing all known members of all characterized *D. melanogaster* signalling 111 pathways. We used tissue-specific RNAi to systematically knock down 463 genes in the larval 112 ovary, and looked for modifiers of the hpo loss of function egg laying and ovariole number 113 phenotypes. To analyse the results of this phenotypic analysis, we used topology-driven 114 network analysis to identify genetic modules regulating these phenotypes, thus generating

hypotheses about the relationships between these modules. With this systems biology approach, we identify not only signalling pathway genes, but also previously untested genes that affect these reproductive traits. Functional testing showed that these novel genes affect ovariole number and/or egg laying, providing us with a novel *in silico* method to identify target genes that affect ovarian development and function. We use these findings to propose putative developmental regulatory modules underlying one or both of ovariole formation and egg laying.

121

122 **Results**

123 An RNAi modifier screen for signalling pathway involvement in ovariole

124 number

125 To systematically ascertain the function of signalling pathway genes and their interactions with

126 Hippo signalling in the development of the *D. melanogaster* ovary, we first curated a list of all

127 known and predicted signalling genes (Gramates et al., 2016; Kanehisa et al., 2010; Mbodj et

al., 2013). We identified 475 genes belonging to the 14 developmental signalling pathways

129 characterised in *D. melanogaster* (Table 1; Table S1), and obtained UAS:RNAi lines for 463 of

130 these genes from the Vienna *Drosophila* RNAi centre (VDRC) or the TRiP collections at the

- 131 Bloomington Drosophila Stock centre (BDSC) (all D. melanogaster genetic lines used are listed
- 132 in Methods).
- 133

134 We previously showed that reducing the levels of *hpo* in the somatic cells of the larval ovary 135 using traffic jam Gal4 (tj:Gal4) driving hpo[RNAi] increased both ovariole number and egg laying 136 of adult female flies (Sarikaya and Extavour, 2015). To identify genes that modify these 137 phenotypes, we used ti: Gal4 to drive simultaneous hpo[RNAi] and RNAi against a signalling 138 candidate gene, and quantified the phenotypic change (Figure 1a-d). We observed that on 139 driving two copies of *hpo*[*RNAi*] using *ti*:*Gal4*, we obtained a further increase in both egg laying 140 and ovariole number (Figure 1e). This indicates that ovaries have further potential to increase 141 ovariole number and egg laying beyond the increase induced by tj:Gal4 driving one copy of 142 hpo[RNAi], and that ti:Gal4 can drive the expression of two RNAi constructs, indicating that our 143 screen could identify both enhancers and suppressors of the *ti:Gal4>hpo[RNAi]* phenotype. 144

145 We proceeded to identify modifiers of the tj:Gal4>hpo[RNAi] phenotype by crossing males of 146 each of the 463 candidate genes RNAis individually with tj:Gal4>hpo[RNAi] females, and 147 performing three phenotypic screens on the offspring. In the first screen (Figure 1a), we 148 measured egg laying of three F1 female offspring (tj:Gal4>hpo[RNAi], signalling 149 candidate[RNAi]) over 5 days. To address batch variation (Figure S1), we standardized egg 150 laying measurements by calculating the Z scores (Z_{aene} = number of standard deviations from 151 the mean) for each candidate line relative to its batch controls. 190 genes had an egg laying 152 $|Z_{gene}|$ below 1. Previous studies have shown that the egg laying of newly eclosed adult mated 153 females correlates with ovariole number during the first five days (Klepsatel et al., 2013b). We 154 therefore eliminated these 190 genes from subsequent screening, because the change in egg 155 laying was so modest that we considered these candidates were unlikely to show changes in 156 ovariole number when compared to controls.

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- 158 In the second screen (Figure 1b), we measured egg laying in a wild-type background
- 159 (tj>signalling candidate[RNAi]) for the 273 remaining candidate genes. For the third screen
- 160 (Figure 1c), we quantified the ovariole number of *tj:Gal4>hpo[RNAi], signalling candidate[RNAi]*

161 F1 adult females for the same 273 candidate genes. To choose candidates from the second and 162 third screens for further study, we wished to account for the fact that the two screens had 163 different effective numbers of data points. This was because eqg laying data were obtained from 164 individual vials of three females over five days, while ovariole numbers were obtained from 20 165 ovaries from ten females (see methods). We therefore selected the 67 genes with a $|Z_{qene}|$ above 166 two for ovariole number (Figure 1c, 1d; Table 2), and the 49 genes with a more conservative 167 |Z_{gene}| above five for egg laying (Figure 1a, 1b, 1d; Table 2), for a total of 116 positive candidates 168 for subsequent analyses.

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170 Ovariole number is weakly correlated with egg laying

171 Standardization of the results from the three screens using Z scores allowed us to compare the 172 effects of individual genes on one or both of egg laying and ovariole number. We performed a 173 pairwise comparison of the Z_{gene} values for all combinations of screens, and considered genes 174 with $|Z_{qene}|$ values that were above the thresholds set for the phenotype in each screen (above 175 two for ovariole number, above five for egg laying; green dots in Figure 2a-c. Across all three 176 screens, loss of function of our positive candidates yielded reductions in ovariole number and 177 egg laying more commonly than increases (Figure 2a-c). Comparing the $|Z_{qene}|$ values of egg laving and ovariole number of ti:Gal4>hpo[RNAi], signalling candidate[RNAi] adult females 178 179 revealed that genes that caused a change in egg laying did not always similarly affect ovariole 180 number, and vice versa (Figure 2a). We therefore hypothesise that egg laying and ovariole 181 number may be regulated by genetically separable mechanisms. This hypothesis 182 notwithstanding, we observed a weak but statistically significant correlation between egg laying and ovariole number (p=1e10⁻⁵; Figure 2d), and this correlation was most significant in adult 183 184 females that had a drastic reduction in both phenotypes (Figure 2a). 185

186 No single signaling pathway dominates regulation of ovariole number or

187 egg laying

- 188 We found that at least some genes from all tested signalling pathways could affect both egg
- 189 laying and ovariole number (Figure 3). To determine if some pathway(s) appeared to play a
- 190 more important role than others in these processes, we asked whether any of our screens were

191 enriched for genes from a specific signalling pathway. To measure enrichment, we compared 192 the distribution of individual pathway genes among the positive candidates in each screen to a 193 bootstrapped null distribution of pathway genes among a group of the same number of genes 194 randomly selected from our curated list of 463 signalling genes (Figure 3a). Involvement of a 195 pathway in the regulation of a phenotype would be reflected in a difference between the 196 representation of pathway genes in an experimentally derived list and a randomly selected 197 group of signalling genes. We found that rather than only one or a few pathways showing 198 functional evidence of regulating ovariole number or egg laying, nearly all pathways affected 199 both phenotypes (Figure 3a). We further tested this result by calculating the hypergeometric p-200 value for the enrichment of each signaling pathway, in each of the three groups of genes. 201 Consistent with the results of the bootstrapping approach (Figure 3a), we found that most 202 pathway members were not significantly enriched for egg laving or ovariole number phenotypes 203 (Figure 3b). The absence of significant enrichment of any specific pathway is not simply 204 attributable to the pool of genes that were screened, because our experimental manipulations of 205 ovariole number and egg laying did cause a change in the distribution of signalling pathway 206 members (Figure S2a). Instead, both phenotypes appeared to be regulated by members of 207 most or all signalling pathways (Figure 3; Figure S2). The only two exceptions to this trend were 208 a greater than twofold enrichment of (1) genes from the Notch signalling pathway in the 209 regulation of ovariole number (p-value < 0.05, pink bar in Figure 3a, b), and (2) members of the 210 Hedgehog (Hh) signaling pathway in the regulation of Hippo-dependent egg laying (p-value < 211 0.05, brown bar in Figure 3a, 3b; Figure S3). In summation, our analyses of the enrichment of 212 signalling pathways within the different screens indicated that both ovariole number and egg 213 laying are regulated by genes from nearly all described animal signalling pathways (Figure 3a), 214 rather than being dominated by any single pathway.

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216 Comparing the results of the Egg Laying screens performed in a wild type background (Figure 217 1b) or in a *hpo[RNAi]* background (Figure 1a), revealed that most of the genes that met a 218 threshold of $|Z_{aene}| > 5$ in one screen, did not meet that threshold in the other screen (Figure 2c). 219 This result suggests the existence of both Hippo-dependent and Hippo-independent 220 mechanisms of regulation of egg laying. The interpretations of separable Hippo-dependent and -221 independent regulation of egg laying, and of the separable regulation of ovariole number and 222 egg laying, was supported by the results of the network analysis described in the following 223 section.

Centrality of genes in the ovarian protein-protein interaction networks canpredict the likelihood of loss of function phenotypic effects

226 The finding that these reproductive traits were regulated by the genes of all signalling pathways 227 led us to consider the broader topology of putative gene regulatory networks in the analysis of 228 our data. Previously characterized genes in the ovary are often pleiotropic and can regulate 229 both ovariole number and egg laying (Gilboa, 2015; Sarikaya and Extavour, 2015). As with 230 proteins in a linear pathway, proteins in a protein-protein interaction (PPI) network are more 231 likely to function in conjunction with genes that are connected to them within the network (e.g. 232 Ideker and Sharan, 2008; Jeong et al., 2001). Centrality is one measure of the connectedness 233 of a gene in the PPI and can be used to identify the most important functional centres within a 234 protein network (Hahn and Kern, 2005; Ma'ayan, 2011). Most centrality measures use path 235 length, which is a measure of the number of other proteins required to link any two proteins in 236 the network. Here we used four commonly used metrics to quantify gene centrality, each 237 measuring slightly different properties (Jalili et al., 2016; Koschutzki and Schreiber, 2008). (1) 238 Degree centrality is proportional to the number of proteins that a given protein directly interacts 239 with. (2) Betweenness centrality measures the number of shortest paths amongst all the 240 shortest paths between all pairs of proteins that require passing through a particular protein. (3) 241 *Closeness centrality* measures the average shortest path that connects a given protein to all 242 other proteins in the network. (4) Eigenvector centrality is a measure of the closeness of a given 243 protein to other highly connected proteins within the network.

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245 We hypothesised that if the candidate genes we identified in our screen as playing roles in 246 ovarian function worked together as a PPI network, then the degree of centrality of a gene might 247 be an indicator of function. To test this hypothesis, we calculated the four centrality measures, 248 described above, for all genes within the *D. melanogaster* PPI (Figure S4). We then rank 249 ordered only the genes tested in each screen by their score for each centrality measure, and 250 asked whether their rank order correlated with the results of the screen, plotting these results as 251 a receiver operating characteristic (ROC) curve. Positive correlations between centrality (a 252 continuous variable) and phenotype (a binary variable: above or below the $|Z_{aene}|$ threshold) are 253 reflected in an area under the curve (AUC) of more than 0.5. We found that the higher the 254 centrality score, the greater the likelihood that a gene had $|Z_{aene}|$ values above our threshold for 255 effects on ovariole number and egg laying (Figure 4a; Table S3). This supports the premise that 256 the positive candidates identified in our screen function together as a network in the regulation

of either ovariole number or egg laying. Interestingly, while the centrality of genes did predict
whether a gene would affect our phenotypes of interest, it could only weakly predict the strength
of that effect (Figure S5).

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261 Genes regulating egg laying and ovariole number regulation form non-262 random interaction modules

263 The centrality analyses above suggested that the genes implicated in ovariole number and egg-264 laying displayed characteristics of a functional network. PPI networks can often be further sorted 265 into a collection of sub-networks. A sub-network is a smaller selection of proteins from the PPI. 266 Examples of such sub-networks could be proteins within the same subcellular organelle (Foster 267 et al., 2006) or genes that are expressed at the same time (Spellman et al., 1998), thus making 268 them likely to function together (Srinivasan et al., 2007). A module is a sub-network that can 269 perform regulatory functions independent of other sub-networks, and has key measurable 270 features (Barabási and Oltvai, 2004; Hartwell et al., 1999; Ravasz et al., 2002; Yook et al., 271 2004). We therefore asked if our sub-networks consisting of genes that showed similar mutant 272 phenotypes might constitute such functional modules. To determine whether genes that were 273 implicated in regulation of ovariole number and egg laying interacted with each other in specific 274 groups more than would be expected by chance, we created four lists of genes, called "seed" 275 lists, based on their individual phenotypic effects based on our screen results: (1) the core seed 276 list, including genes positive in all three screens (Figure 4b); (2) the egg laying seed list, 277 including genes positive in the wild type background egg-laving screen (Figure 1b: Figure 4c): 278 (3) the hpo[RNAi] egg laying seed list, including genes positive in the hpo[RNAi] background 279 egg laying screen (Figure 1a; Figure 4c); and (4) the *hpo[RNAi]* ovariole seed list, including 280 genes positive in the *hpo[RNAi]* background ovariole number screen (Figure 1c; Figure 4c). 281 Interestingly, the core seed list, comprising genes that affected all three measured phenotypes, 282 only consisted of genes that caused a reduction in both ovariole number and egg laying (Figure 283 4b).

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285 Based on published molecular interactions, putative functional modules of genes can be

predicted by algorithms that use either the shortest path method (Bromberg et al., 2008) or the

287 Steiner Tree approach (Huang and Fraenkel, 2009). Such methods identify and predict

288 functional connections between the seed proteins, as well as additional nodes (proteins or 289 genes) that have not been experimentally tested within the given parameters, but are known to 290 interact with the seed genes in the PPI (Albert and Albert, 2004)((Yu et al., 2006). This process 291 results in a predicted module, and subsequent functional genetic testing of this module can 292 confirm its functionality. Given its recent success in predicting gene modules, we used the 293 previously published Seed Connector Algorithm (SCA), a member of the Steiner Tree algorithm 294 family (Wang et al., 2017; Wang and Loscalzo, 2018), to identify putative functional modules 295 formed by genes that had similar phenotypic effects in our screens (Figure 4b, 4c). The SCA 296 connects seed genes and previously untested novel genes (connectors) to each other using a 297 known PPI network, producing the largest possible connected putative module given the data. 298 We compiled a PPI network consisting of all described interactions between D. melanogaster 299 proteins, from the combination of publicly available PPI studies in the DroID database (see 300 Methods). Using this PPI network and the aforementioned four seed lists, we applied a custom 301 python implementation of the SCA (Methods: 04 Seed-Connector.jpynb) to build and extract the 302 largest possible (given our PPI) connected putative modules that regulate egg laying and 303 ovariole number.

This SCA method yielded four putative modules, one for each seed list, which we refer to as the core module (Figure 5b), *hpo[RNAi]* Egg Laying module (Figure S6a), Egg Laying module (Figure S6b), and *hpo[RNAi]* Ovariole Number Module (Figure S6c) respectively. Each of the modules contained seed genes, which had been functionally evaluated in our screens (green and red circles in Figure 5), as well as connector genes, which were genes newly predicted as regulators of these phenotypes (green and red triangles in Figure 5).

310 We then asked whether these four putative modules were more connected than we would 311 expect by chance; in other words, we formally tested them for modularity. Meeting our criteria 312 for modularity would suggest that the genes in these modules operated together as functional 313 sub-networks within the Drosophila PPI. We defined our modularity test using four commonly 314 measured network metrics: (1) Largest Connected Component (LCC) (the number of proteins 315 connected together by at least one interaction), (2) network density (the relative number of 316 edges as compared to the theoretical maximum), (3) total number of edges, and (4) average 317 shortest path (average of the minimum distances connecting any two proteins). We considered 318 a group of genes to form a module if they showed higher LCC, higher network density, more 319 edges, and shorter average shortest path length than a random, bootstrapped selection of the

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322	To determine whether these criteria would correctly identify genes of the same signaling
323	pathway, which are known to function together, as a module, we measured these four
324	parameters in the original set of genes used in this study (Table 1). We found that all the genes
325	of a given signalling pathway were identified as a module based on these parameters (Figure
326	S7a). We then used this approach to test the modularity of the four phenotypic sub-networks, as
327	compared to 1000 bootstrapped "control sub-networks" consisting of a group of the same
328	number of genes as contained in the sub-network, but chosen randomly from among the
329	candidate genes from our initial screen list (Table 1). We found that the four predicted
330	phenotypic modules showed a significantly increased Largest Connected Component (LCC)
331	value, network density, number of edges and decreased average shortest path (Figure S7b),
332	compared to our "control module". This result indicates that these sub-networks are likely to
333	function as modules within the PPI, to regulate one or both of ovariole number or egg laying.

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Low edge densities between modules indicates genetically separable mechanisms of ovariole number and egg laying

337 Our network analysis identified four highly connected networks of genes that regulate two 338 distinct developmental processes, together with or independently of Hippo signalling activity: 339 ovariole number determination, which occurs primarily during larval development, and egg 340 laying, which takes place in adult life (Figure 5). We wished to assess the degree to which there 341 were any shared genetic components between the four modules. To understand potential 342 interactions between the modules in the regulation of both ovariole number and egg laying, we 343 constructed a composite network of all genes in each of the four modules (Figure 5b; Figure 344 S6), which we refer to as the "meta network" (Figure 7a). We then grouped the genes based on 345 their phenotypic effects as measured in the three screens, resulting in seven sub-networks (I-VII 346 in Figure 7a). We then asked if these sub-networks were as connected to each other, as were 347 the genes within each of the sub-networks. To do this, we used an edge density map, which 348 reflects the number of interactions between the genes within a sub-network and between each 349 of the sub-networks (Figure 7b).

350 This analysis yielded three principal findings. First, edge densities between the three sub-351 networks corresponding to the three scored phenotypes (I, II and III in Figure 7a) were very low 352 (Figure 7b). This indicates that genes in each of these sub-networks function as largely 353 independent networks, rather than interacting substantially with any genes in the other non-core 354 sub-networks. Second, the core sub-network (IV in Figure 7a) displayed a higher edge density 355 with the other three sub-networks (I, II and III in Figure 7a) than any of them did with each other 356 (Figure 7b). Consistent with the definition of core module genes as regulating all three scored 357 reproductive phenotypes, this result suggests that the core module genes, in contrast to those 358 from the other three sub-networks, may share substantial functional interactions with genes of 359 the other sub-networks. Finally, three small additional sub-networks emerged from this analysis 360 (V, VI and VII in Figure 7a), suggesting small functional networks of genes that work together to 361 regulate two of the three scored phenotypes. In sum, this meta network analysis supports the 362 hypothesis of three largely independent genetic networks that regulate Hippo-dependent 363 ovariole number, Hippo-dependent egg laying, and Hippo-independent egg laying. Moreover, 364 each of these genetically separable networks included genes in multiple signalling pathways 365 (Figure 7c).

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Network analysis predicts novel genes involved in egg laying and ovariolenumber

369 The four predicted phenotypic modules produced by the SCA approach included connector 370 genes that were not included in our original screen, and thus had not been tested for possible 371 effects on our phenotypes of interest (triangles in Figure 5b; Figure S6). Given that prior work in 372 human disease models showed that predicted disease modules can correctly predict gene 373 involvement in the relevant diseases (Chen et al., 2006; Gonzalez et al., 2007; Wang et al., 374 2017; Wang and Loscalzo, 2018), we asked whether our deployment of the SCA had likewise 375 successfully predicted novel, functionally important genes in each module. To do this, we 376 measured the effects of knocking down each the connector genes (triangles in Figure 5b and 377 Figure S6) on ovariole number and egg laying, using UAS:RNAi lines for each connector, driven 378 by tj:Gal4.

Of the ten predicted novel connectors in the core module, loss of function of several of thesehad significant effects on at least one of the three scored phenotypes. Five affected ovariole

number two affected Hpo-dependent egg laying, and one affected Hpo-independent egg laying.
However, only one of them significantly altered all three scored phenotypes (Figure 6a; Table
S4).

384 The predicted connector genes from two of the other three phenotypic modules showed high 385 positive prediction rates. RNAi against seven out of 18 of the *hpo[RNAi]* Egg Laving module 386 connectors, three out of 11 of the hpo[RNAi] Ovariole Number module connectors, and none of 387 the 11 Egg Laving module connectors, significantly affected the module phenotype (Table S4). 388 Thus, although the Egg Laying module connectors failed to impact this phenotype in our assay, 389 41.2% and 27.3% of the connectors from the other two modules were correctly predicted 390 (Figure 6b; Table S4). These positive hit rates exceed those obtained in our initial candidate 391 screens, where 59/463 (12.7%) and 67/273 (24.5%) tested genes affected hpo-dependent egg 392 laying and *hpo*-dependent ovariole number respectively (Figure 6d; Table 2). In sum, taken 393 across all modules (Figure 6c; Table S4, Table S5), this network analysis correctly identified 394 genes regulating all scored reproductive phenotypes, at rates higher than those obtained in the 395 original screen of 463 members of all known signalling pathways. By this measure, testing 396 network-predicted regulatory modules derived from experimentally obtained data was even 397 more successful than testing signalling pathways as a means of identifying novel genes that 398 regulate ovariole number and egg laying.

399 Discussion

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401 Identification of regulatory modules for ovariole development and egg

402 laying

The *D. melanogaster* ovary is a commonly studied model for organogenesis (Chen et al., 2001;
Godt and Laski, 1995; Lobell et al., 2017; Sarikaya and Extavour, 2015), stem cell maintenance
(Gilboa, 2015) and interactions of development and ecology (Cohet and David, 1978; Hodin and
Riddiford, 2000a; Klepsatel et al., 2013a; Sarikaya et al., 2019). Nevertheless, our
understanding of the genetic mechanisms that regulate these processes remains fragmentary.
In this paper, we have identified four distinct protein interaction modules that regulate ovariole
number and egg laying in the *D. melanogaster* ovary. These modules consist of both novel and

410 previously characterized genes that regulate either ovariole number or egg laying or both, thus

- 411 enhancing our understanding of the genetic underpinnings of this reproductive system.
- 412

413 Of the four modules, the core module affects both ovariole number and egg laying. The core 414 module contains numerous housekeeping genes, including regulators of transcription, 415 translation and cell division such as polo (Llamazares et al., 1991), cyclin K (Edwards et al., 416 1998), nucleosome assembly protein 1 (Ito et al., 1996) and eukaryotic translation release factor 417 1 (Chao et al., 2003). While polo and eukaryotic translation release factor 1 are members of 418 signalling pathways, cyclin K and nucleosome assembly protein 1 are genes predicted by the 419 SCA. Given that the core module largely consists of genes whose loss of function decreases 420 both of these parameters, we hypothesise that these are essential genes for the basic structure 421 and function of the ovaries. Essential genes are more interconnected in a PPI with higher 422 centrality measures (Jeong et al., 2001) and interestingly, we find that the genes in the core 423 module also have higher connectivity than those in the other three modules (Figure S4). 424 425 In addition to genes that regulate basic cellular processes, the core module is enriched for the

426 core components of the Hh signalling cascade, namely patched (ptc), smoothened (smo) and 427 costa (cos) (Lee et al., 2016). However, we find that the loss of Hh ligand, which is expressed in 428 the TF cells in the developing larval ovary (Lai et al., 2017), does not significantly affect either 429 ovariole number or egg laying. Though surprising, ligand-independent activation of Hedgehog 430 signalling has been observed before. For example, in the Drosophila eye, loss of either ptc or 431 cos in clones leads to non-cell autonomous proliferation in wild type cells, as well as growth 432 disadvantages in the mutant tissue (Christiansen et al., 2012). In another example, sufficient 433 intracellular *smo* levels can also activate downstream transcription of Hh pathway targets. 434 showing that Hh itself is not always required to activate the cascade (Jiang et al., 2018).

435 Development of the larval ovary

The *hpo[RNAi]* Ovariole Number module is composed of genes that affect the Hippo signalling activity-dependent determination of ovariole number during development. Establishment of ovariole number occurs largely during the third instar stage of larval development in *D. melanogaster* (Godt and Laski, 1995; Hodin and Riddiford, 1998; King et al., 1968; Sahut-Barnola et al., 1996). During this period, the TFCs are specified in the anterior of the ovary and undergo rearrangement into stacks of cells called TFs, each of which gives rise to an ovariole (Godt and Laski, 1995; Sahut-Barnola et al., 1995). TF specification requires the expression of 443 engrailed (En) (Bolívar et al., 2006) and the transcription factors Bab1 and Bab2, encoded by 444 the bric-à-brac locus (Couderc et al., 2002; Godt et al., 1993). A third transcription factor, 445 Lmx1a, was recently found to be necessary for the specification of the TFCs (Allbee et al., 446 2018). Our hpo[RNAi] Ovariole Number module identifies numerous additional novel 447 transcription factors including bunched (bun) and retinoblastoma-family protein (rbf), which we 448 hypothesize could also be involved in the specification of ovariole number. bun and rbf have 449 been implicated in the migration (Dobens et al., 2005) and endoreplication (Cayirlioglu et al., 450 2003) of the follicle cells during oogenesis, but have not, to our knowledge, been previously 451 identified as playing a role in the context of larval ovary development.

452

453 The TFCs specified in the larval ovary undergo a process of convergent extension to form TFs. 454 This process of convergent extension requires cell intercalation, and the actin depolymerizing 455 factor Cofilin, encoded by the gene *twinstar*, is essential to this process (Chen et al., 2001). 456 During intercalation, the cells also dynamically modify their actin cytoskeleton and their 457 expression of E-cadherin (Godt and Laski, 1995). Our hpo[RNAi] Ovariole Number module 458 further identifies Rho1 (Barrett et al., 1997) and Rho kinase (Rok) (Mizuno et al., 1999) as 459 necessary for correct ovariole number. During the extension of the *D. melanogaster* embryonic 460 germ band, a commonly studied model of convergent extension, the localised activation of the 461 actin-myosin network facilitated by Rho1 and Rok is necessary for cell intercalation (Kasza et 462 al., 2014). Given the known roles of *Rho1* and *Rok* as regulators of the actin cytoskeleton 463 (Ridley, 2006), we propose that TF assembly in the ovary requires both these proteins for 464 correct cell intercalation. A third actin cytoskeleton regulator, *misshapen (msn)*, was also 465 identified by our *hpo*[RNAi] Ovariole Number module. *msn* encodes a MAP kinase previously 466 shown to regulate the polarisation of the actin cytoskeleton during oogenesis (Lewellyn et al., 467 2013), but has not, to our knowledge, been studied to date in the context of larval ovarian 468 development.

469

We propose that the polarity of the somatic cells in the ovary is also necessary for correct larval
ovary development, given the presence of the lateral membrane proteins *discs large 1* (*dlg1*)
and *prickle* (*pk*) in the ovariole module. During the maturation of the TFs during larval

- 473 development, the TFCs undergo significant cell shape changes, coincident with localised
- 474 expression of beta-Catenin and actin to the lateral edges of the TFCs (Godt and Laski, 1995).
- 475 Restriction of the E-cadherin domain in epithelia requires establishment of the basolateral

domain (Harris and Peifer, 2004) and we propose that testing a similar requirement for *dlg1* and *pk* in the larval ovary would be a fruitful avenue for future studies.

478 Network analysis as a tool in developmental biology

479 Using a systems biology approach to analyse RNAi screening data has proven fruitful, providing 480 us with new insights into the development and function of the *D. melanogaster* ovary by 481 identifying novel and previously understudied genes that regulate this process. Systematic 482 analysis of the function of single genes in development has been a historical convention and 483 has provided valuable and precise genetic interaction information (Jansen et al., 2002; von 484 Mering et al., 2002). With the advent of genome-wide analysis, however, we can use data from 485 a larger number of genes to predict the identity of additional functionally significant genes with 486 relative ease (Yu et al., 2006). We note that the novel gene prediction rate ranged from as high 487 as 41.2% from the *hpo[RNAi]* Ovariole Number module to as low as 0% from the Egg Laying 488 module (Figure 6b; Tables S4, S5). We suggest that this may be due to multiple factors. Firstly, 489 the possible incompleteness of the PPI is expected to lead to some areas of the network being 490 sparse or non-existent (von Mering et al., 2002). If the module of interest happens to fall in such 491 regions of the network, prediction algorithms will fail. Secondly, the initial restriction of tested 492 genes to signaling pathway members might have provided a seed list too sparse to usefully 493 predict functional connectors. Finally, it could be the case that "Egg Laving" is such a complex 494 phenotype that its gene regulation cannot be adequately captured within a highly connected 495 network of the type suited for identification by the analyses we have used here. Ovariole 496 number in *D. melanogaster* is the outcome of a discrete developmental process with a clear 497 beginning and end, comprising a specific series of cellular behaviors that take place in the 498 confines of one organ (Godt and Laski, 1995; Hodin and Riddiford, 2000a; Sahut-Barnola et al., 499 1996). Once established during larval life, ovariole number in Drosophila remains unaltered 500 through to and during adulthood, even if oogenesis within those ovarioles suffers congenital or 501 age-related defects (King, 1970). Because previous work suggested that ovariole number in 502 Drosophila could have at least some predictive relation to egg laving (Cohet and David, 1978; 503 Klepsatel et al., 2013b; Sarikaya and Extavour, 2015), we reasoned that scoring the latter 504 phenotype in a primary screen (Figure 1a) could be an effective way to uncover ovariole number 505 regulators (Figure 1c). While our results showed that this was true in many cases, it was also 506 clear that these two traits can vary independently (Figure 2), highlighting the fact that ovariole 507 number is not the only determinant of egg laying. Egg-laying dynamics, even during the limited

508 five day assay used in our study, are likely influenced not just by a single anatomical parameter

- such as ovariole number, but rather by many biological, biomechanical, hormonal and
- 510 behavioural processes. Consequently, the functional module we were able to extract from the
- results of this screen (Figure 1b) might be too coarse to extract novel genes that participate in
- 512 potentially complex gene interactions regulating egg laying.
- 513 The predictive rates of the approach we have used here, although encouraging, are likely
- 514 limited by the degree of noise in the high throughput data used to generate the PPI (Li et al.,
- 515 2010), the sparseness of the PPI, and the degree of misidentification of protein interactions
- 516 (Zhang et al., 2017). Addressing one or more of these parameters could improve the outcomes
- 517 of future network predictions from developmental genetics data. For example, the problem of
- 518 sparseness, which is a paucity of high confidence detectable interactions relative to all
- 519 biologically relevant interactions, has been addressed in other studies by using an "Interolog
- 520 PPI" (Matthews et al., 2001) in place of an organism-specific PPI. The Interolog PPI combines
- 521 known interactions from multiple organisms, and has been used successfully to identify, for
- 522 example, gene modules relevant in squamous carcinoma, based on a starting dataset of
- 523 microarray data on differentially expressed genes between cancer cells and the surrounding
- tissue (Wachi et al., 2005). Future studies applying the Interolog PPI to the outcomes of genetic
- 525 screens for developmental processes of interest could potentially overcome the problem of
- 526 sparseness, as well as the biases towards proteins that are more heavily studied and thus
- 527 better represented in organism-specific PPIs.

528 Tables

529 Table 1

Signalling pathway	Number of genes in screen
EGF	45
FGF	25
FOXO	67
Нірро	60
JAK/STAT	31
JNK	28
МАРК	29
Notch	48
SHH	54
TGF B	52
Toll	36
VEGF	17
Wnt	125
mTOR	36

530

531 **Table 1: Number of candidate genes tested in each signalling pathway.** Candidate genes

are grouped by their reported roles in one or more signalling pathways based on published

533 literature. Genes in this list are not necessarily unique to a single pathway, but may function in

more than one signalling pathway. The list of specific genes per pathway that were included in

the screen for functional analysis (Figure 1) is found in Table S1.

536

537 Table 2

538

Egg Laying Screens	<i>hpo[RNAi]</i> Egg Laying (Figure 1a)	Egg Laying (Figure 1b)	Ovariole Number Screen	<i>hpo[RNAi]</i> Ovariole Number (Figure 1c)
RNAi stocks unavailable	12	0	RNAi stocks unavailable	0
Primary filter ($ Z_{gene} < 1$)	190	N/A	Primary filter ($ Z_{gene} < 1$)	N/A
No effect (-5 < $ Z_{gene} $ < 5)	214	224	No effect (-2< $ Z_{gene} $ < 2)	206
Negative effect ($Z_{gene} < -5$)	48	44	Negative effect ($Z_{gene} < -2$)	54
Positive effect ($Z_{gene} > 5$)	11	5	Positive effect ($Z_{gene} > 2$)	13
Total	475	273	Total	273

539

540 **Table 2: Results of the three functional genetic screens.** Number of genes tested in each screen and cumulative results.

541 "Negative effect" corresponds to a reduction in eggs laid or number of ovarioles below the Z score (*Z*_{gene}) threshold for each

542 phenotype. "Positive effect" indicates an increase above the set Z_{gene} thresholds. Z_{gene} thresholds for each category in each screen

543 are indicated in brackets. The primary filter of $|Z_{gene}| < 1$ was applied only to the *hpo*[*RNAi*] Egg Laying screen shown in Figure 1a.

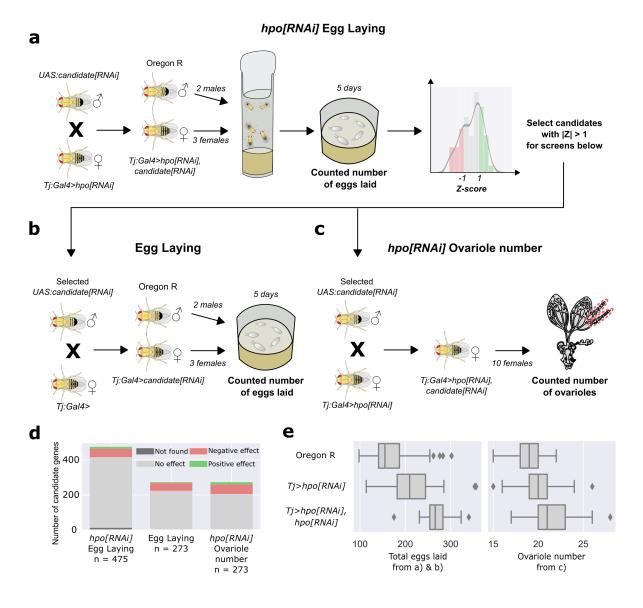
544 The list of specific genes that exceeded our chosen Z_{gene} thresholds for each scored phenotype (Figure 1), and were therefore

545 considered to have a positive or negative effect on the phenotype, is found in Table S1. The 12 genes for which RNAis stocks were

546 unavailable at the time of testing are listed in Table S2.

547 Figures

548 Figure 1

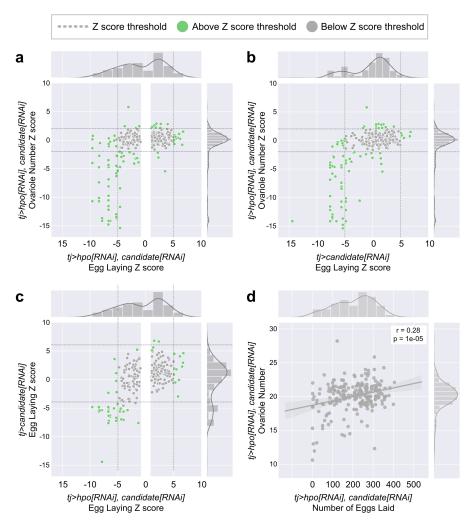


549

550

Figure 1. Screen methodology. a,b,c) Diagrammatic representation of screen workflow. d)
Distributions of results of genes in the three screens. n = number of genes tested in each
screen (see also Table 2). e) Total eggs laid by three female flies over five days (left panel) and
ovariole number (right panel) of Oregon R (top row), *tj:Gal4* driving one copy of UAS:hpo[RNAi]
(middle row), and *tj:Gal4* driving two copies of UAS:hpo[RNAi] (bottom row), showing that, the
previously reported *tj:Gal4>hpo[RNAi]* ovariole number and egg laying phenotypes (Sarikaya
and Extavour, 2015) can be modified by further UAS:RNAi-mediated gene knockdown.

558 Figure 2

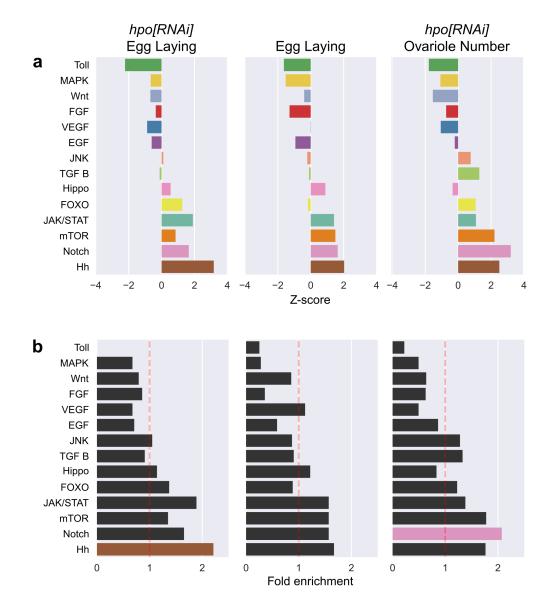


559 560

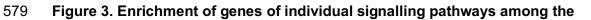
561 Figure 2. Relationship between Egg Laying and Ovariole Number phenotypes generated in the screens. a) Scatter plots of the Z score for each gene (Z_{gene}) of egg laying versus the 562 563 ovariole number of adult tj>hpo[RNAi], candidate[RNAi] females. b) Scatter plots of the Z score 564 for each gene (Z_{gene}) of egg laying of adult tj>candidate[RNAi] females versus the ovariole 565 number of adult tj>hpo[RNAi], candidate[RNAi] females. c) Scatter plots of the Z score for each 566 gene (Z_{eene}) of egg laying of adult ti>candidate[RNAi] females versus egg laying of adult 567 *tj>hpo[RNAi], candidate[RNAi]* females. In **a**, **b** and **c**, bar graphs on the top and right sides of 568 each panel show the distribution of genes in each axis of the adjacent scatter plots. Green dots 569 = genes that meet the Z_{gene} threshold for the indicated phenotype. Grey dots = genes that do not 570 meet the Z_{gene} threshold for the indicated phenotype. Dark grey dotted lines = thresholds for 571 each phenotype: $|Z_{gene}| > 5$ for Egg Laying and $|Z_{gene}| > 2$ for Ovariole Number. In **a** and **c**, the 572 white vertical bar removes all genes in the $t_j > hpo[RNAi]$, candidate[RNAi] with a $|Z_{gene}| < 1$ for 573 egg laying. These genes were not measured in the other two conditions and are therefore not 574 represented in the scatter plots. d) Correlation between non-zero Ovariole Number and Egg 575 Laying values.

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576 Figure 3



577 578



580 **experimentally obtained positive candidates of each screen. a)** Depletion/enrichment

analysis to identify over- or under- represented members of individual signalling pathways

among positive candidates of each screen. Positive Z scores represent an enrichment, and
 negative Z scores represent depletion, of genes of a pathway among those genes that

experimentally affected the phenotype Enrichment and depletion are defined relative to a null

585 distribution of the expected number of members of a signalling pathway among a group

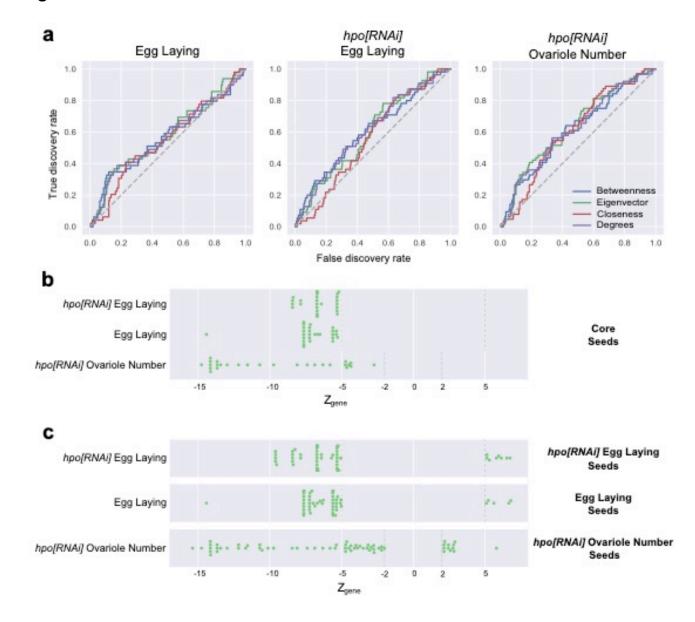
586 containing the same number of randomly selected signalling genes. **b)** Fold enrichment and

587 hypergeometric p-value calculation to identify over- or under-representation of the genes of a

588 pathway in each screen. Significantly enriched pathways (colored bars: brown = Hedgehog;

589 pink = Notch) are defined by having a hypergeometric p-value less than 0.05.

590 Figure 4



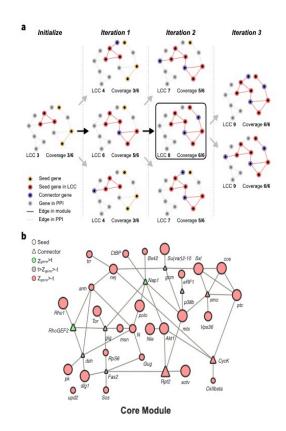
591

592

593 Figure 4. Screened genes function as a network. a) Receiver operating characteristic (ROC) 594 curves of genes ordered by rank for each of four network centrality metrics (Betweenness 595 centrality, Eigenvector centrality, Closeness centrality and Degree centrality) versus a binary 596 outcome (above or below Z score threshold) for each of the three screens. For each screen and 597 metric, the Area Under the Curve (AUC) is > 0.5 (Table S3). **b**) Genes whose $|Z_{gene}|$ value was 598 above the threshold (green dots; Table 2) in all three screens were assigned to the Core seed 599 list. c) Genes whose $|Z_{gene}|$ value was above the threshold (green dots; Table 2) in each screen 600 were assigned to the corresponding seed list.

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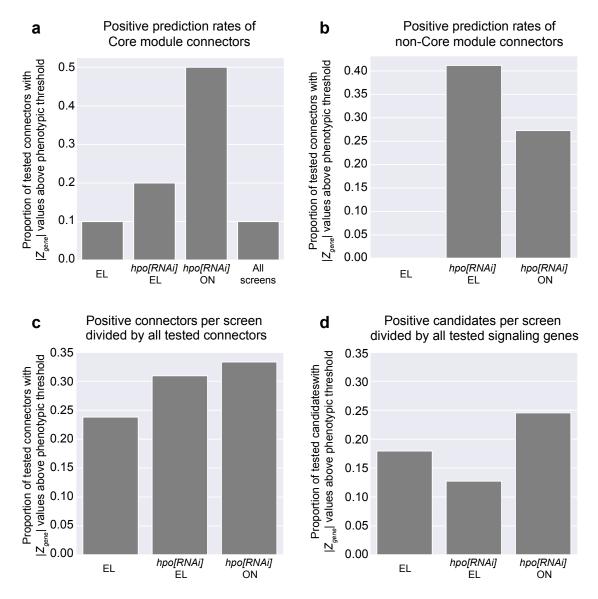
601 Figure 5



602

603 Figure 5. Representation of Seed Connector Algorithm and output. a) Schematic 604 representation of the seed connector algorithm. The algorithm initializes by creating a 605 subnetwork of seed genes from the PPI, and computes the Largest Connected Component 606 (LCC) and coverage (number of genes from the seed set in the LCC). At each iteration, genes 607 in the direct neighborhood of the LCC (distance = 1) are added one at a time to the seed set, 608 and the coverage and LCC are recomputed. This process is repeated for each gene in the direct 609 neighborhood, each time restarting from the seed set of the preceding iteration. If any gene 610 outside the seed set but in the direct neighbourhood is found to maximize coverage while 611 minimizing the LCC, it is added to the seed set as a connector gene. Black arrows indicate the 612 path taken by the algorithm for which the criteria of maximal coverage and minimal LCC are 613 met; such a path would be used to proceed to the subsequent iteration. Grey arrows indicate 614 paths that fail to meet these criteria; such paths would be disregarded. The iteration repeats 615 until the coverage cannot be increased; in this schematic example, this state is achieved in 616 iteration 3. b) The Core Module generated by the Seed Connector Algorithm (SCA) based on 617 the results of the genetic screens (Figure 1a-c). The size of the shapes indicate the relative Z_{gene} 618 score of the gene. Circles indicate seed genes (functionally tested in the screen; Table 2; Table 619 S1) while triangles are connector genes (novel predicted genes; Tables S1, S4, S5). Green = 620 genes with a positive Z_{gene} score above the threshold; red = genes with a negative Z_{gene} score 621 above the threshold; grey = genes with Z_{gene} values below the threshold.

622 Figure 6



623

624

Figure 6. Positive prediction rates of the connector genes in each of the four modules. a)

Proportion of core module connector genes with $|Z_{gene}|$ above the threshold in each of the three screens. The "All phenotypes" category includes the genes with $|Z_{gene}|$ above the threshold in all

three screens. **b)** Proportion of tested connector genes in each of the three modules with $|Z_{gene}|$

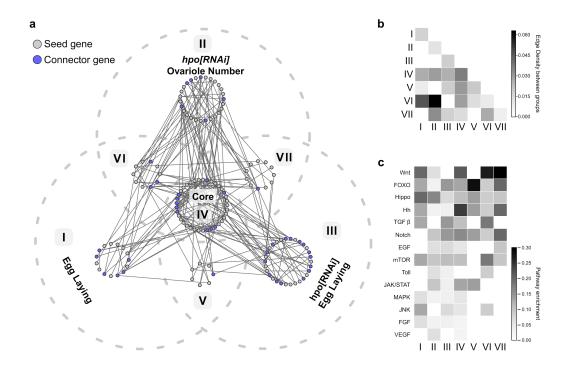
above the threshold within their corresponding screen. **c)** Proportion of all unique connector genes predicted by all four modules with $|Z_{gene}|$ above the respective threshold in any of the

631 three screens. **d**) Proportion of positive candidate genes emerging from the three original

signalling candidate screens with $|Z_{gene}|$ above the threshold relative to the total number of

633 genes tested in each screen.

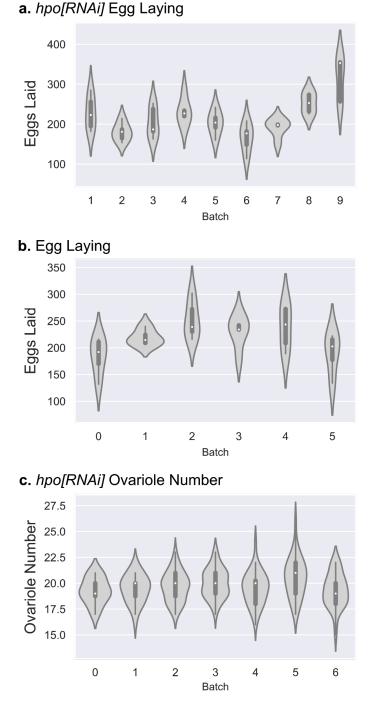




635 636

Figure 7. Phenotypically separable sub-networks formed by analysis of the combined genes from all modules. The meta 637 638 network is generated by the union of the genes in the four phenotypic modules: *hpo[RNAi]* Egg Laving (Figure S6a), Egg Laving 639 (Figure S6b), hpo[RNAi] Ovariole Number (Figure S6c) and Core (Figure 5b). a) The meta network is represented as a Venn 640 diagram, in which each grey dotted outline represents the screen in which a given gene was identified as affecting the scored 641 phenotype. Within each sub-network, grey circles indicate seed genes, and blue circles indicate connector genes. A single gene, sloppy paired 1, was a seed in the Egg Laying module and also a connector in the hpo[RNAi] Egg Laying module; it fell within 642 643 sub-network VII in the meta network, and is marked as a seed (grey) in this figure. Solid grey lines indicate interactions between 644 genes in the meta network from the PPI. b) Edge densities between the seven sub-networks of the meta-network. c) Relative 645 enrichment of screened members of the 14 tested developmental signalling pathways within the seven sub-networks of the meta-646 network.

647 Figure S1



- 648
- 649

650 Figure S1. Violin plots of egg laying and ovariole number of controls in each screen

651 **batch. a)** Distribution of number of eggs laid by five replicates of three *tj:Gal4>hpo[RNAi]*

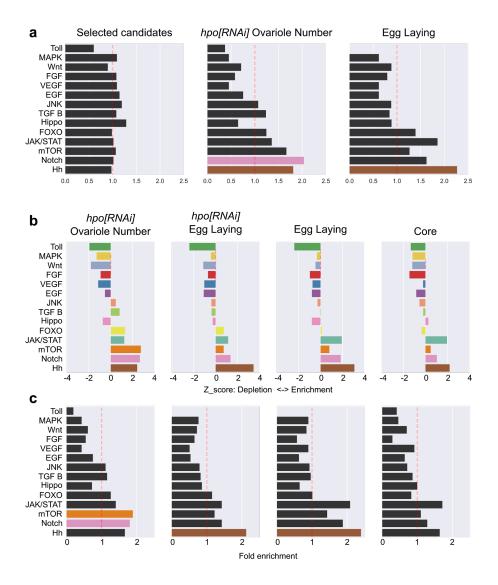
652 females over five days for each batch. **b)** Distribution of number of eggs laid by five replicates of

three *tj:Gal4* females over five days for each batch. **c)** Distribution of number of ovarioles per

654 ovary in 20 ovaries from ten *tj:Gal4>hpo[RNAi]* females in each batch.

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655 Figure S2

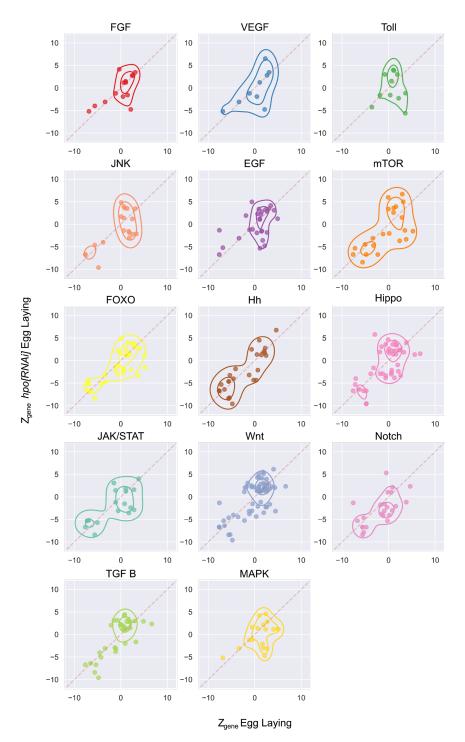


656 657

658 Figure S2. Biological aspects of the network modules. a) Enrichment/depletion analysis of 659 the 273 signalling pathway genes above the threshold $|Z_{qene}| > 1$ (Figure 1a) against all 660 signalling candidates. We also measured the enrichment/depletion of positive signalling 661 candidate genes in the hpo[RNAi] Ovariole (Figure 1c) and Egg Laying (Figure 1b) screens from 662 the 273 genes tested in those screens. b) Signalling pathway depletion enrichment analysis. For 663 each module, a null distribution of the expected number of members of a signalling pathway 664 from a group of the same number of randomly selected signalling pathway genes was 665 calculated. The Z score from the expected distribution was then calculated. Negative Z scores 666 represent a depletion, while positive Z scores represent an enrichment. No single pathway is 667 enriched in any of those modules. c) Fold enrichment and hypergeometric p-value calculation 668 for each pathway in the four modules. Pathway members in color (orange = mTor; brown = 669 Hedgehog; pink = Notch) have a p-value < 0.05.

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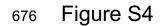
670 Figure S3

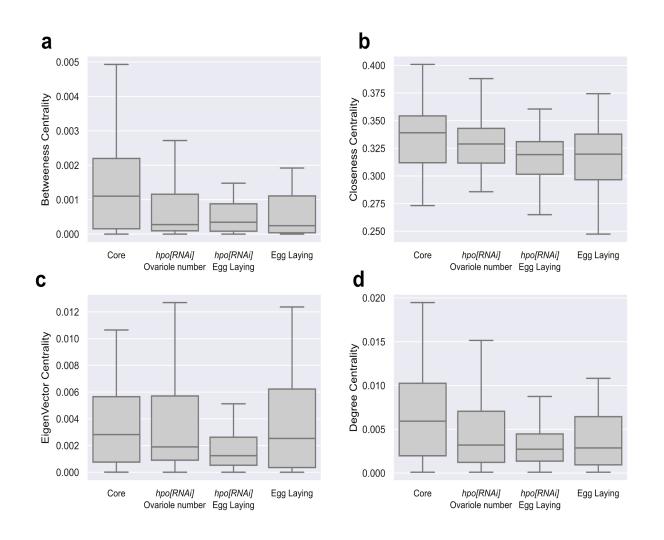


671

Figure S3. Comparison of egg laying candidate genes by pathway. *Z*_{gene} of egg laying of

- 673 adult females of *tj>hpo[RNAi],candidate[RNAi]* plotted against Z_{gene} of egg laying of
- 674 *tj>candidate[RNAi]* adult females displayed by pathway. Contour plots indicate a 2D gaussian
- 675 kernel density estimation.





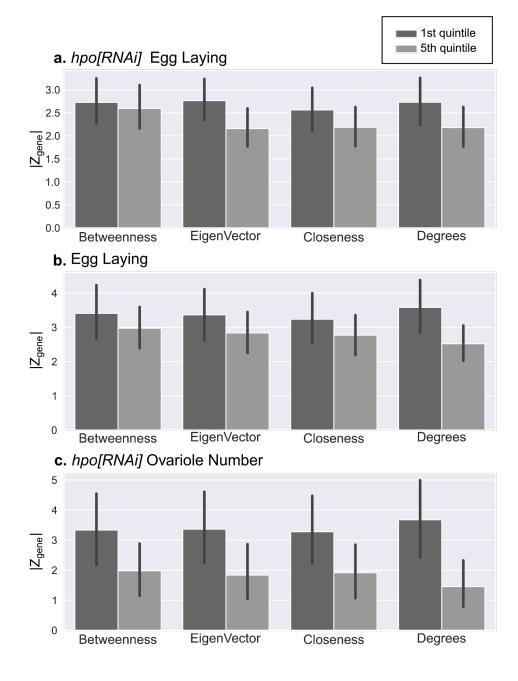
677

Figure S4. Box plots of the four centrality measures calculated for the genes in each of the four phenotypic modules. See
 modules in Figures 5 and S6.

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680

681 Figure S5

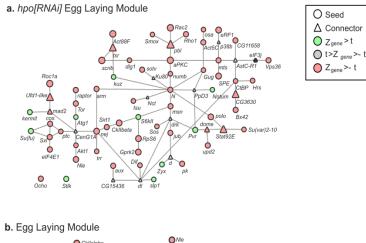


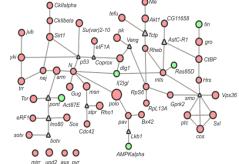
682 683

Figure S5. Comparisons of the Z_{gene} scores of the positive candidate genes sorted by

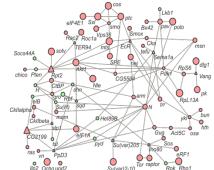
685 **centrality metrics.** In each screen (a, b, c), the $|Z_{gene}|$ values of the first (dark grey) and fifth 686 (light grey) quintiles of positive candidate genes ordered by rank for each of the four chosen 687 centrality metrics, are plotted as a bar plot. Bars indicate standard error.

688 Figure S6









689

690 Figure S6. Three modules generated by the Seed Connector Algorithm (SCA). a)

hpo[RNAi] Egg Laying Module **b)** Egg Laying Module **c)** *hpo[RNAi]* Ovariole Number Module.

692 The size of the shapes indicate the Z_{gene} score of the gene. Circles = seed genes; triangles =

693 connector genes. Green = genes with a positive Z_{gene} above the threshold. Red = genes with a

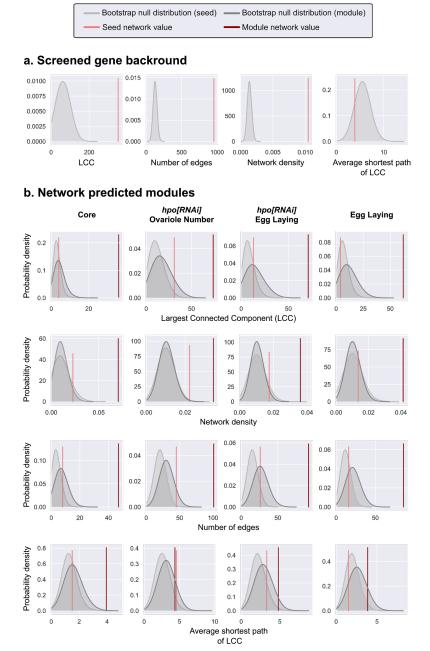
694 negative Z_{gene} above threshold. Grey = genes with Z_{gene} values below the threshold. All

695 connectors were phenotypically tested (Table S1) except *eukaryotic translation initiation factor* 3

696 *subunit j* (*eIF3J*), in the *hpo[RNAi]* Egg Laying Module (black triangle), for which no RNAi stock 697 as available at the time of testing.

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698 Figure S7



699



701 **Network) application of the seed connector algorithm. a)** Comparison of network metrics of

all screened genes (red line) to a null distribution of network metrics derived by bootstrapping an

equal number of randomly selected genes in the PPI (grey curve). **b)** Comparisons of the

Largest Connected Component (LCC), network density, number of edges and average shortest

path between the seed network (light red line) and the module network (dark red line). The

bootstrapped null distribution (1000 bootstraps) of both the seed network (light grey curve) and

the module network (dark grey curve) are indicated..

708 Supplementary tables

709 Table S1

- 710 **Table S1: Tabulation of raw data and analysis for every gene in the screen.**
- 711 <u>https://github.com/extavourlab/hpo_ova_eggL_screen/blob/master/Results/MasterTable.c</u>
 712 sv.

713

- This table contains a summary representation of the data generated by the three screens as
- 715 well as results from the analysis. Each line corresponds to an independent measurement of a
- particular RNAi line. Some genes which did not pass the first filter of $|Z_{gene}| > 1$ in the *hpo*[RNAi]
- Fight Fight
- have been independently measured again. The Z scores have been rounded up to 4 significant
- 719 digits in this table and the Centrality metrics rounded up to 10 significant digits due to their low
- values, but the full values for both are available in the raw data files provided in the
- supplementary files in Data/Screens for the Z scores and Results for the centrality values.
- 722 Moreover this is a summary table and does not contain values for controls as well as batch
- numbers, all are available in the supplementary files in Data/Screens.
- 724
- 725 FbID: FlyBase ID of the tested gene.
- 726 **CG number**: CG Number of the tested gene.
- **NAME**: Common name (as per FlyBase nomenclature) of the gene if existing, else it is a -.
- **SYMBOL**: Symbol (as per FlyBase nomenclature) of the gene if existing, else CG number
- 729 [ScreenName]_[Variable]_(Metric)_Count: Within the screen [ScreenName], the count of the
- 730 measured variable [*Variable*]. Optional: (*metric*) will indicate if a particular operation was done
- 731 over the data, such as sum, mean or standard deviation.
- e.g. [HippoRNAi_EggL]_[Day_4_Egg]_Count is the count of eggs, on day 4, of the *hpo[RNAi]*Egg Laying screen.
- [ScreenName]_[Variable]_(Metric)_Zscore: Within the screen [ScreenName], the Z score of
 the measured variable [Variable] as calculated to batch control. Optional: (metric) will indicate if
- a particular operation was done over the data, such as sum, mean or standard deviation.
- e.g. [EggL]_[All_Days_Egg]_(Sum)_Zscore is the Z score of the sum of eggs count, of the EggLaying screen.
- PPI_[*Metric*]_centrality: Within the PPI used in this paper, the calculated centrality value for
 the metric [*Metric*].
- 741 [*ModuleName*]_Network: Presence of absence of a gene in the module [*ModuleName*]. If the
- gene is in the module, this value is True, if it is absent it is False. (An exception is made for the
- 743 Meta Network displayed in Figure 7 where instead of True/False, the group assignment I-VII is744 written)
- 745 [ModuleName]_Connector: Status of a gene in the module [ModuleName] as a connector. If
- True, the gene is a connector, else if False, the gene is not a connector.
- 747 [PathwayName]_Pathway: Participation of a gene to the signalling pathway [PathwayName].
- 748 If the gene participates in the pathway the value is 1, else it is 0.

749 Table S2

FbID	CG Number	Name	Symbol
FBgn0283468	CG3412	supernumerary limbs	slmb
FBgn0267821	CG5102	daughterless	da
FBgn0266724	CG5161	TRAPP subunit 20	Trs20
FBgn0267378	CG7085	sauron	sau
FBgn0267487	CG9181	Protein tyrosine phosphatase 61F	Ptp61F
FBgn0267912	CG9819	Calcineurin A at 14F	CanA-14F
FBgn0086371	CG9829	poly	poly
FBgn0267350	CG10260	Phosphatidylinositol 4- kinase III alpha	PI4KIIIalpha
FBgn0267698	CG10295	p21-activated kinase	Pak
FBgn0283462	CG18279	Immune induced molecule prepropeptide	IMPPP
FBgn0267339	CG33338	p38c MAP kinase	p38c
FBgn0085506	CG40635	-	CG40635

750

- 751 Table S2. 12 signalling candidate genes with no available RNAi lines at either BDSC or
- 752 **VDRC at the time of this study.**

753

754 Table S3

755

Centrality metric	<i>hpo[RNAi]</i> Ovariole Number	<i>hpo[RNAi]</i> Egg Laying	Egg Laying
Betweenness	0.603	0.57	0.586
EigenVector	0.632	0.573	0.586
Closeness	0.612	0.551	0.588
Degrees	0.615	0.592	0.599

756

757 **Table S3: Area under the curve (AUC) of ROC curves.** AUC values for the ROC curves for

each centrality measure for the three screens (Figure 4a). AUC values range from 0 to 1. A

score above 0.5 indicates a positive correlation between the continuous variable (centrality) and

the binary variable (above or below the Z score threshold). A score of 0.5 or less indicates no

761 correlation between the variables.

762 Table S4

763

Module	Number of Seeds	Number of Connectors	Number of connector genes above Z _{gene} threshold within module phenotype
<i>hpo[RNAi]</i> Egg Laying	58	18	7 (41.2%)
Core	27	10	1 (10.0%)
Egg Laying	49	11	0 (0.0%)
<i>hpo[RNAi]</i> Ovariole Number	66	11	3 (27.3%)

764

765	Table S4. Distribution of seed genes and connectors in each module. Two genes that were
766	above Z _{gene} threshold (Table 2) in the <i>hpo[RNAi]</i> Egg Laying (CG12147) and <i>hpo[RNAi]</i>
767	Ovariole Number seed list (CG6104) were not found in the PPI, and therefore not included in
768	the network analysis or in this table (see methods for details). The removal of these two genes
769	accounts for the difference between the number of positive candidates in Table 2 and the
770	number of seed genes in these two modules (Table S1 and S4). The proportion of connectors
771	whose loss of function produced a significant phenotype ($ Z_{gene} $ above threshold) is in
772	parentheses and plotted in Figure 6a, 6b). All connectors except eukaryotic translation initiation
773	factor 3 subunit j (eIF3J) in the hpo[RNAi] Egg Laying Module, for which no RNAi line was
774	available at the time of testing, were tested. Therefore, the percentages of connectors above the

threshold for the *hpo[RNAi]* Egg Laying Module were calculated out of 17 connectors.

776 Table S5

777

Total number of unique connectors	Number of connector genes above Z _{gene} threshold		
in all four modules	For Egg Laying Phenotype	For <i>hpo[RNAi]</i> Egg Laying Phenotype	For <i>hpo[RNAi]</i> Ovariole Number Phenotype
43	10 (23.8%)	13 (31.0%)	14 (33.3%)

778

Table S5. Number of unique connector genes above $|Z_{gene}|$ threshold for the three

780 **phenotypic measurements.** Percentage of the number of connectors above threshold for each

781 phenotype from the total number of connectors is in parentheses and plotted in Figure 6c. All

782 connectors except eukaryotic translation initiation factor 3 subunit j (eIF3J) in the hpo[RNAi] Egg

783 Laying Module, for which no RNAi line was available at the time of testing, were tested.

Therefore, the percentages of connectors above the threshold were calculated out of 42 unique

785 connectors.

787 Methods

788 LEAD CONTACT AND MATERIALS AVAILABILITY

This study did not generate new unique reagents. This study generated new python3 code
available on GitHub: <u>https://github.com/extavourlab/hpo_ova_eggL_screen.</u>

791

Further information and requests for resources and reagents should be directed to and will be

fulfilled by the Lead Contact, Cassandra G. Extavour (<u>extavour@oeb.harvard.edu</u>).

794 EXPERIMENTAL MODEL AND SUBJECT DETAILS

Wild type and mutant lines of *Drosophila melanogaster* were obtained from publicly accessible
stock centers and maintained as described in "Fly Stocks" below. Genotypes and provenance
are provided in the Key Resource Table. Candidate genes were randomly assigned to batches
for screening (see Table S1 for which genes were in each batch). F1 animals from the same
cross were randomly assigned to experimental groups for phenotyping in all screens.

800 METHOD DETAILS

801 Fly stocks

802 Flies were reared at 25°C at 60% humidity with standard *Drosophila* food (Sarikaya et al., 2012)

803 containing yeast and in uncrowded conditions as previously defined (Sarikaya and Extavour,

804 2015). RNAi lines were obtained from the TRiP RNAi collection at the Bloomington Drosophila

805 Stock Centre (BDSC) and from the Vienna *Drosophila* Resource Centre (VDRC). See Key

806 Resources Table for complete list of stocks used in this study. Oregon R was used as a wild

807 type strain. The genotype of the *traffic jam:Gal4* line used in the screen was *y w; P{w[+mW.hs]*

- 808 = *GawB*}*NP1624* (Kyoto Stock Center, K104–055; abbreviated hereafter as *tj:Gal4*). The *hippo*
- 809 RNAi line used in the screen was y[1] v[1]; $P\{y[+t7.7]v[+t1.8]=TRiP. HMS00006\}attP2$
- 810 (BDSC:33614; abbreviated hereafter as *hpo[RNAi]*).

811 Egg and ovariole number counts

812 Adult egg laying was quantified by crossing three virgin females of the desired genotype (see

813 *"Screen design"* below) with two males in a vial containing standard food and yeast granules

814 (day one) and then transferring them into a fresh food vial without yeast granules for a 24 hour

period. Eggs from vials were then counted by visual inspection of the surface of the food in the

vial. Males and females were transferred to fresh food vials without yeast granules, every day

817 thereafter until day six. All egg laying measurements reported and analysed in the paper are the

sum of the eggs laid by three adult female flies over the five days of this assay (days two

through six without yeast granules). Data from any vial in which either a female or male died, 819

820 during the course of the experiment, were not included in the analysis.

821

822 Ovariole number was quantified by mating ten virgin adult females with five virgin adult Oregon 823 R males for three days post eclosion in vials with yeast at 25°C and 60% humidity. After this 824 three-day mating period, all 20 adult ovaries from the mated females were dissected in 1X PBS 825 with 0.1% Triton-X-100 and stained with 1ug/ml Hoechst 33321 (1:10,000 of a 10mg/ml stock

- 826 solution). Ovarioles were separated from each other with No. 5 forceps (Fine Science Tools)
- 827 and counted by counting the number of germaria under a ZEISS Stemi 305 compact stereo
- 828 microscope with a NIGHTSEA stereo microscope UV Fluorescence adaptor.

Screen design 829

830 In the primary screen (Figure 1a: hpo[RNAi] Egg Laying), 463 candidate genes (Table S1) were

831 screened for the effect of an RNAi-induced loss of gene function in a hpo[RNAi] background on 832 the number of eggs laid in the first five days of mating (see "Egg and ovariole number counts"

833

above) by adult females. These females were the F1 offspring of UAS: candidate gene RNAi 834 males crossed to P{w[+mW.hs] = GawB}NP1624; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2

835 (tj:Gal4; UAS:hpo[RNAi]) virgin adult females (Figure 1a: hpo[RNAi] Egg Laying). All genes that

836 yielded an egg laying count with a $|Z_{qene}| > 1$ (see "Gene selection based on Z score and batch

837 standardization" below) were selected to undergo two secondary screenings (n=273, Table 2,

838 Figure 1d). First, these genes were screened for effects on the egg laying of mated adult female

839 offspring from a cross of UAS:candidate gene[RNAi] males and tj:Gal4 virgin adult females

840 (Figure 1b: Egg Laying). Secondly, these genes were screened for effects on ovariole number in

841 a hpo[RNAi] background. All 20 ovaries from ten adult female F1 offspring of a cross between

842 UAS:candidate gene[RNAi] males to P{w[+mW.hs] = GawB}NP1624; P{y[+t7.7]

843 v[+t1.8]=TRiP.HMS00006}attP2 (tj:Gal4; UAS:hpo[RNAi]) virgin adult females were scored for

844 ovariole number (see "Egg and ovariole number counts" above). (Figure 1c: hpo[RNAi] Ovariole 845 Number).

Gene selection based on Z score and batch standardization 846

847 Candidate genes were screened in batches with an average size of 50 genes. For each batch,

848 control flies were the female F1 offspring of Oregon R males crossed to P{w[+mW.hs] =

849 GawB}NP1624; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2 (tj:Gal4; UAS:hpo[RNAi]) virgin adult

850 females. Because the control group in each batch had slightly different distributions of egg

851 laying and ovariole number values (Figure S1), it was inappropriate to compare absolute mean

852 values between genes that were scored in different batches. Instead, comparisons of the Z

853 score of each candidate (Z_{gene}) to its batch control group was used as a discriminant. This

854 approach standardizes for batch effects and allows the comparison of all genotypes within and 855 across the primary and secondary screens with a single metric (Z_{ene}).

856

857 Firstly, the mean and standard deviation of the eggs laid by the control genotype for a batch 858 were calculated as μ_b and σ_b respectively. Then, using the number of eggs laid by adult females

- of a candidate gene RNAi (x_{gene}) of the same batch, the Z score for the egg laying count of that gene (Z_{gene}) was calculated as $Z_{gene} = \frac{x_{geme} - \mu_b}{\sigma_b}$. The same standardization protocol was
- applied to both egg laying and ovariole number counts of every gene and its corresponding batch control.
- 863

864 Ovariole numbers were derived from counts of the number of ovarioles per ovary for 20 ovaries 865 per candidate gene, and a threshold of $|Z_{gene}| > 2$ (corresponding to a false positive probability 866 less than 0.045) was applied for ovariole number phenotype. Egg laying counts were derived 867 from measurements of three females in a single vial per gene. We therefore chose to be more 868 conservative in our Z score comparisons for the egg laying phenotype, than for ovariole number 869 phenotype, and applied a stringent threshold of $|Z_{gene}| > 5$ (corresponding to a false positive 870 probability less than 0.00006) to select genes of interest. All genes with $|Z_{gene}|$ values above 871 these thresholds are referred to throughout the study as "positive candidates". (See lpython 872 notebooks 02 Z score calculation.jpynb and 02.2 Z score calculation prediction.jpynb for 873 code implementation and calculation of Z scores, and 06 Screen Analysis.jpynb for batch

874 effects.)

875 Signaling pathway enrichment analysis

To study the enrichment of a particular signaling pathway in a group of candidate genes that

877 had similar phenotypic effects revealed by the screen, custom scripts (see

878 07_Signaling_pathway_analysis.ipynb for code implementation) were generated to implement

two different methods (Figure 3a, 3b; Figure S52a-c).

880

881 The first method is a numerical method that uses bootstrapping to calculate the null distribution 882 of the number of members (M) of a signaling pathway (S) that would be expected at random in a 883 set of genes of size (N). The script randomly sampled N genes from among the 463 tested D. 884 melanogaster signaling genes 10.000 times, and counted the number of genes (M) that were 885 members of the signaling pathway S. Positive candidates in each of the three screens were 886 sorted by their presence in signalling pathways and counted. The Z score was then calculated 887 by comparing the experimentally observed number of positive candidates in each signaling 888 pathway against the bootstrapped null distribution.

889

The second method used the hypergeometric p-value to calculate the probability of M members of a signaling pathway being in a group of N genes, given a starting population of 463 tested *D*.

892 *melanogaster* signaling genes, and the known attribution to a pathway S of each gene.

893 Protein-Protein Interaction Network (PPI) building

894 There is no standard complete Protein-Protein Interaction (PPI) network available for *Drosophila* 895 *melanogaster*. However, there exist many smaller networks from different screens, as well as

literature extractions. We therefore combined data from these sources and then created a PPI

897 for use in the present study, as follows:

898 899 Step 1: Several screens assessing protein-protein interactions have been centralized in a 900 database called DroID: http://www.droidb.org. The version DroID v2018 08 was used. All 901 available datasets were first downloaded from that database using this link: 902 http://www.droidb.org/Downloads.jsp. The description of all of these datasets can be found here: 903 http://www.droidb.org/DBdescription.jsp 904 905 Step 2: We used the datasets from all screens that assessed direct protein-protein interactions 906 and did not use the interolog database (predicted protein interaction based on mouse human 907 and yeast PPI). These direct assessment screens were seven in total, as follows: 908 909 • Finley Yeast Two-Hybrid Data (size 2.0 MB) 910 • Curagen Yeast Two-Hybrid Data (size 4.6 MB) 911 • Hybrigenics Yeast Two-Hybrid Data (size 381 KB) 912 • Perrimon co-AP complex (size 108 KB) 913 • DPiM co-AP complex (size 6.3 MB) 914 • PPI from other databases (size 16.2 MB) 915 • PPI curated by FlyBase (size 7.4 MB) 916 917 An important element to note is that the PPI curated by FlyBase is a literature-based PPI. 918 FlyBase protein-protein interactions are experimentally derived physical interactions curated 919 from the literature by FlyBase, and does not include FlyBase-curated genetic interactions. 920 921 Step 3: We concatenated the seven datasets listed above into a single unique database. A

922 custom python script was created that downloads and reads each of the above seven unique 923 PPI tables, and generates a single PPI network. From this concatenation, a single edge 924 undirected network was created and saved. This network is hereafter referred to as the PPI

925 (see 01 PPI builder.ipynb).

Network metric computations 926

927 The centrality of a node is often used as a measure of a node's importance in a network. Within 928 a PPI, the centrality of a gene reflects the number of interactions in which the gene directly or 929 indirectly participates. Four different centrality metrics were computed for all genes in the PPI 930 using the networkx python library:

- 931
- 932 (1) **Betweenness** reflects the number of shortest paths passing through a gene.
- 933 (2) **Eigenvector** is a measure of the influence of a gene in the network.
- 934 (3) **Closeness** measures the sum of shortest distance of a gene to all the other genes.
- 935 (4) **Degree centrality** corresponds to the normalized number of edges of a gene in the network.
- 936
- 937 While there exist more centrality measures, these four are commonly used to assess biological
- 938 networks. These computed centrality parameters of the genes measured in the screen were

939 computed with 03_ROC_curve_analysis_of_network_metrics.ipynb, and are reported in the

940 Table S1 (see 09_Making_the_database_table.ipynb).

941 Receiver Operating Characteristic (ROC) curves

942 To check whether the centrality of a gene in the network could predict the phenotypic effect 943 produced by RNAi against that gene, ROC curves were plotted for the four aforementioned 944 centrality measures of each gene in each screen. A ROC analysis is used to measure the 945 correlation between a continuous variable (centrality) and a binary outcome (above or below Z 946 score threshold). Therefore, for each screen, measured genes were rank-ordered from high 947 centrality to low centrality, and plotted against the binary outcome of $|Z_{gene}|$ being above or 948 below the appropriate |Z score| threshold (>5 for egg laying and >2 for ovariole number). The 949 Area Under the Curve (AUC) measures the extent of correlation between centrality and effect of 950 a gene on measured phenotype. AUC above or below 0.5 indicates a positive or negative 951 correlation respectively, while an AUC of 0.5 indicates no correlation of the parameters. The 952 scikit learn python package was used to calculate the AUC of each ROC curve plotted (see 953 03 ROC curve analysis of network metrics.ipynb).

954 Building the network modules

955 Network modules were built using the previously published Seed-Connector algorithm (SCA) 956 (Wang et al., 2017; Wang and Loscalzo, 2018), implemented here in python (see 04 Seed-957 Connector.ipynb) and illustrated in Figure 5a. Creating a module using the SCA requires a list of 958 seed genes and a PPI. From each of the three screens, we selected the genes whose $|Z_{eene}|$ 959 value was above the threshold and created three seed lists respectively (Figure 4c: Egg laying, 960 hpo[RNAi] egg laying and hpo[RNAi] ovariole 'seed' list). A fourth list consisting of the 961 intersection of the aforementioned seed lists was also collated and called the core 'seed' list 962 (Figure 4b). Genes were assigned in the core list if they passed the Z threshold in all 3 screens. 963 The Seed-Connector algorithm was then executed on each of these seed lists using the PPI. 964 Not all genes in the four seed lists were found in the PPI network (specifically, CG12147 in the 965 hpo[RNAi] Egg Laving seed list and CG6104 in the hpo[RNAi] Ovariole number seed list were 966 absent from the PPI) and were therefore eliminated from further network analysis. The removal 967 of these two genes accounts for the variation in the number of positive candidates in Table 2 968 and the number of seed genes in the module. Modules were obtained for each seed list (Figure 969 5b; Figure S6) consisting of the seed genes (circles in Figure 5b and Figure S6) and previously 970 untested genes added by the SCA (squares in Figure 5b and Figure S6) to increase the LCC 971 size that we refer to as connector genes (see 04 Seed-Connector.ipynb). The results of the 972 algorithm are summarized in Table S1.

973

The modularity of the subnetworks was then assessed using four network metrics namely

975 Largest Connected Component (LCC), number of edges, network density and average shortest

976 path in the LCC. Each metric for each module was assessed using distance of the network

- 977 metric to a null distribution. Initially, the null distribution was calculated by taking 1000 samples
- 978 of 463 genes randomly selected from the PPI and calculating the above metrics. We found that

the 463 genes selected in the signalling screen were already more connected than the null 979 980 distribution of sets of 463 genes randomly selected from the PPI (Figure S7a). Therefore, to 981 avoid a false positive detection of modularity, the four experimentally obtained subnetworks 982 were compared to null distributions obtained by randomly sampling an equal number of genes 983 from the 463 signalling candidate genes selected for our screen. For each of the four modules, 984 comparison of the metrics was performed on the seed lists and the sub-network after the SCA. 985 Most metrics were enriched in the seed group when compared to the null distribution with the 986 exception of the Average shortest path (Figure S7b, light red line). The sub-networks obtained 987 from the SCA further increased all four metrics suggesting the modularity of the four sub-988 networks (Figure S7b, dark red line; see 05 Network Module testing.ipynb for code 989 implementation).

990 Meta network

991 To build the meta network, the genes from all four modules were concatenated into one 992 network. The network was then visually sorted in an approach akin to projecting the network on 993 a Venn Diagram. The meta network was sorted by which of the three screens the gene was 994 positive in. The intersections were genes whose $|Z_{gene}|$ value was above the threshold in more 995 than one and possibly all three of the screening paradigms. For example, if a gene was found in 996 the hpo[RNAi] Ovariole Number and Egg Laying module it is then assigned to the dual positive 997 group hpo[RNAi] Ovariole Number / Egg Laying (Figure 7a, module VI). After applying this 998 grouping strategy, the connectivity across the groups was studied by calculating the edge density between all groups (density $= \frac{Edges_{1;2}}{Nodes_1*Nodes_2}$). Finally, the proportion of each signaling 999 1000 candidate in each of those groups was calculated by taking the number of members of a 1001 signaling pathway divided by the total members of a group (see lpython notebook 1002 08 MetaModule Analysis.ipynb).

1003 QUANTIFICATION AND STATISTICAL ANALYSIS

- 1004 Number of samples
- 1005 The number of samples across the different screens were as follows:

1006 *hpo[RNAi]* Egg Laying and Egg Laying screens

- 1007 Controls: five vials of three females and two males
- 1008 Sample: one vial of three females and two males
- 1009 *hpo[RNAi]* Ovariole number screen
- 1010 Controls: 20 flies, two ovaries per fly considered as independent measurements
- 1011 Sample: 10 flies, two ovaries per fly considered as independent measurements

1012 Correction of batch effect

1013 Despite best efforts to maintain the exact same condition between each experiment, some

- 1014 variation was measured between the batches. Control flies showed variations in both measured
- 1015 phenotypes, ovariole number and egg laying (Figure S1). In order to compare the values
- 1016 measured across different batches, each sample was standardized by calculating its Z score
- 1017 (Z_{gene}) to the control distribution. For each batch, the measurements for controls were pooled into
- 1018 a distribution, and the mean and standard deviation was computed. Then each sample was
- 1019 compared to its respective batch and its Z score computed (see "Gene selection based on Z
- 1020 score and batch standardization" for formula).

1021 Statistical analysis

- 1022 All statistical analyses were performed using the scipy stats module (<u>https://www.scipy.org/</u>) and
- scikit learn (<u>https://scikit-learn.org/</u>). Significance thresholds for p-values were set at 0.05.
- 1024 Statistical tests and p-values are reported in the figure legends. All statistical tests can be found
- 1025 in the lpython notebooks mentioned below.

1026 DATA AND CODE AVAILABILITY

1027 This study generated a series of python3 lpython notebook files that perform the entire analysis

- 1028 presented in this study. All the results presented in this paper, including the figures with the
- 1029 exception of the network visualizations, which were created using Cytoscape3
- 1030 (https://cytoscape.org/) can be reproduced by running the aforementioned python3 code. The
- 1031 raw data, calculations made with these data, and code used for calculations and analyses
- 1032 (Ipython notebooks) are available as supplementary information. For ease of access, legibility
- 1033 and reproducibility, the code and datasets have been deposited in a GitHub repository available
- 1034 at <u>https://github.com/extavourlab/hpo_ova_eggL_screen.</u>

1036 KEY RESOURCES TABLES

1037 Software and libraries

1038 All software and libraries used in this study are published under open source licenses and are

1039 therefore publicly available.

1040

Туре	Name	Version	Source
Library	matplotlib	3.0.0	https://matplotlib.org/
Library	networkx	2.3	http://networkx.github.io/
Library	numpy	1.11.3	https://www.numpy.org/
Library	pandas	0.20.3	https://pandas.pydata.org /
Library	progressbar	3.38.0	https://github.com/niltonvolpato/python-progressbar
Library	scipy	1.1.0	https://www.scipy.org/
Library	seaborn	0.9.0	https://seaborn.pydata.org/
Software	Cytoscape	3.4.0	https://cytoscape.org/
Software	Inkscape	0.92.3	https://inkscape.org/
Software	Python3	3.7	https://www.python.org/

1042 Drosophila melanogaster genetic lines

Description	Stock Center	IDs
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CanA1 (FBgn0010015) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01871}attP2	Bloomington Drosophila Stock Center	BDSC:25850; FlyBase:FBst0025850 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fng (FBgn0011591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01967}attP2	Bloomington Drosophila Stock Center	BDSC:25947; FlyBase:FBst0025947 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Aplip1 (FBgn0040281) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02049}attP2	Bloomington Drosophila Stock Center	BDSC:26024; FlyBase:FBst0026024 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of E(spl)mdelta-HLH (FBgn0002734) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02101}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:26203; FlyBase:FBst0026203 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sima (FBgn0266411) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02105}attP2	Bloomington Drosophila Stock Center	BDSC:26207; FlyBase:FBst0026207 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of E(spl)m8-HLH (FBgn0000591) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02096}attP2	Bloomington Drosophila Stock Center	BDSC:26322; FlyBase:FBst0026322 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pan (FBgn0085432) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02306}attP2	Bloomington Drosophila Stock Center	BDSC:26743; FlyBase:FBst0026743 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CanB (FBgn0010014) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02616}attP2	Bloomington Drosophila Stock Center	BDSC:27307; FlyBase:FBst0027307 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of mib1 (FBgn0263601) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02629}attP2	Bloomington Drosophila Stock Center	BDSC:27320; FlyBase:FBst0027320 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Nct (FBgn0039234) under UAS control in the VALIUM10	Bloomington Drosophila Stock Center	BDSC:27498; FlyBase:FBst0027498 ;

	-	
vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02648}attP2		
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cbl (FBgn0020224) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02650}attP2	Bloomington Drosophila Stock Center	BDSC:27500; FlyBase:FBst0027500 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Atg12 (FBgn0036255) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02704}attP2	Bloomington Drosophila Stock Center	BDSC:27552; FlyBase:FBst0027552 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fz2 (FBgn0016797) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02722}attP2	Bloomington Drosophila Stock Center	BDSC:27568; FlyBase:FBst0027568 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Psn (FBgn0284421) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02760}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27681; FlyBase:FBst0027681 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pi3K92E (FBgn0015279) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02770}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:27690; FlyBase:FBst0027690 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cdk4 (FBgn0016131) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02795}attP2	Bloomington Drosophila Stock Center	BDSC:27714; FlyBase:FBst0027714 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of mts (FBgn0004177) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02805}attP2	Bloomington Drosophila Stock Center	BDSC:27723; FlyBase:FBst0027723 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pdk1 (FBgn0020386) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02807}attP2	Bloomington Drosophila Stock Center	BDSC:27725; FlyBase:FBst0027725 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ds (FBgn0284247) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02842}attP2	Bloomington Drosophila Stock Center	BDSC:28008; FlyBase:FBst0028008 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Hrs (FBgn0031450) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02860}attP2	Bloomington Drosophila Stock Center	BDSC:28026; FlyBase:FBst0028026 ;

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<i>D. melanogaster</i> . Expresses dsRNA for RNAi of mam (FBgn0002643) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02881}attP2	Bloomington Drosophila Stock Center	BDSC:28046; FlyBase:FBst0028046 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of bun (FBgn0259176) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02954}attP2	Bloomington Drosophila Stock Center	BDSC:28322; FlyBase:FBst0028322 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of aos (FBgn0004569) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03020}attP2	Bloomington Drosophila Stock Center	BDSC:28383; FlyBase:FBst0028383 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pcx (FBgn0003048) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05038}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28552; FlyBase:FBst0028552 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Su(fu) (FBgn0005355) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05045}attP2	Bloomington Drosophila Stock Center	BDSC:28559; FlyBase:FBst0028559 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Apc2 (FBgn0026598) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05073}attP2	Bloomington Drosophila Stock Center	BDSC:28585; FlyBase:FBst0028585 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of I(2)tid (FBgn0002174) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05082}attP2	Bloomington Drosophila Stock Center	BDSC:28594; FlyBase:FBst0028594 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Su(H) (FBgn0004837) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05110}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:28900; FlyBase:FBst0028900 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of PDZ- GEF (FBgn0265778) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05139}attP2	Bloomington Drosophila Stock Center	BDSC:28928; FlyBase:FBst0028928 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wntD (FBgn0038134) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05158}attP2	Bloomington Drosophila Stock Center	BDSC:28947; FlyBase:FBst0028947 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cdk2 (FBgn0004107) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM05163}attP2	Bloomington Drosophila Stock Center	BDSC:28952; FlyBase:FBst0028952 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ci (FBgn0004859) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01715}attP2	Bloomington Drosophila Stock Center	BDSC:28984; FlyBase:FBst0028984 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of mgl (FBgn0261260) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02485}attP2	Bloomington Drosophila Stock Center	BDSC:29324; FlyBase:FBst0029324 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of E(spl)m4-BFM (FBgn0002629) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03310}attP2	Bloomington Drosophila Stock Center	BDSC:29378; FlyBase:FBst0029378 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Wnt4 (FBgn0010453) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF03378}attP2	Bloomington Drosophila Stock Center	BDSC:29442; FlyBase:FBst0029442 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Dif (FBgn0011274) under UAS control in the VALIUM10 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HM05257}attP2	Bloomington Drosophila Stock Center	BDSC:30513; FlyBase:FBst0030513 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of InR (FBgn0283499) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01482}attP2	Bloomington Drosophila Stock Center	BDSC:31037; FlyBase:FBst0031037 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cdc5 (FBgn0265574) and Roc1b (FBgn0040291) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01517}attP2	Bloomington Drosophila Stock Center	BDSC:31067; FlyBase:FBst0031067 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Egfr (FBgn0003731) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01696}attP2	Bloomington Drosophila Stock Center	BDSC:31183; FlyBase:FBst0031183 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of norpA (FBgn0262738) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01713}attP2	Bloomington Drosophila Stock Center	BDSC:31197; FlyBase:FBst0031197 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CenG1A (FBgn0028509) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01807}attP2	Bloomington Drosophila Stock Center	BDSC:31228; FlyBase:FBst0031228 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dsh (FBgn0000499) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01254}attP2	Bloomington Drosophila Stock Center	BDSC:31307; FlyBase:FBst0031307 ;

<i>D. melanogaster.</i> Expresses dsRNA for RNAi of TI (FBgn0262473) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01276}attP2	Bloomington Drosophila Stock Center	BDSC:31477; FlyBase:FBst0031477 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of I(2)gl (FBgn0002121) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01073}attP2	Bloomington Drosophila Stock Center	BDSC:31517; FlyBase:FBst0031517 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Jra (FBgn0001291) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF01184}attP2	Bloomington Drosophila Stock Center	BDSC:31595; FlyBase:FBst0031595 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Axn (FBgn0026597) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04012}attP2	Bloomington Drosophila Stock Center	BDSC:31705; FlyBase:FBst0031705 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gig (FBgn0005198) under UAS control in the VALIUM1 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HM04083}attP2	Bloomington Drosophila Stock Center	BDSC:31770; FlyBase:FBst0031770 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Med (FBgn0011655) under UAS control in the VALIUM10 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02218}attP2	Bloomington Drosophila Stock Center	BDSC:31928; FlyBase:FBst0031928 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of RagC- D (FBgn0033272) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00333}attP2	Bloomington Drosophila Stock Center	BDSC:32342; FlyBase:FBst0032342 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of AMPKalpha (FBgn0023169) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00362}attP2	Bloomington Drosophila Stock Center	BDSC:32371; FlyBase:FBst0032371 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Rho1 (FBgn0014020) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00375}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32383; FlyBase:FBst0032383 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pk (FBgn0003090) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00408}attP2	Bloomington Drosophila Stock Center	BDSC:32413; FlyBase:FBst0032413 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of RpS6 (FBgn0261592) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00413}attP2	Bloomington Drosophila Stock Center	BDSC:32418; FlyBase:FBst0032418 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of foxo (FBgn0038197) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00422}attP2	Bloomington Drosophila Stock Center	BDSC:32427; FlyBase:FBst0032427 ;
D. melanogaster. Expresses dsRNA for RNAi of Plc21C (FBgn0004611) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00436}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32438; FlyBase:FBst0032438 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp2 (FBgn0036046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00476}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32475; FlyBase:FBst0032475 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hh (FBgn0004644) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00492}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32489; FlyBase:FBst0032489 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Spred (FBgn0020767) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00637}attP2	Bloomington Drosophila Stock Center	BDSC:32852; FlyBase:FBst0032852 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of upd3 (FBgn0053542) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00646}attP2	Bloomington Drosophila Stock Center	BDSC:32859; FlyBase:FBst0032859 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of llp1 (FBgn0044051) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00648}attP2	Bloomington Drosophila Stock Center	BDSC:32861; FlyBase:FBst0032861 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp7 (FBgn0044046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00649}attP2	Bloomington Drosophila Stock Center	BDSC:32862; FlyBase:FBst0032862 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of SkpA (FBgn0025637) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00657}attP2	Bloomington Drosophila Stock Center	BDSC:32870; FlyBase:FBst0032870 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG3226 (FBgn0029882) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00662}attP2	Bloomington Drosophila Stock Center	BDSC:32875; FlyBase:FBst0032875 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CtBP (FBgn0020496) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00677}attP2	Bloomington Drosophila Stock Center	BDSC:32889; FlyBase:FBst0032889 ;

D. melanogaster. Expresses dsRNA for RNAi of Hel89B		
(FBgn0022787) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00684}attP2	Bloomington Drosophila Stock Center	BDSC:32895; FlyBase:FBst0032895 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Tctp (FBgn0037874) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00701}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32911; FlyBase:FBst0032911 ;
D. melanogaster. Expresses dsRNA for RNAi of jub (FBgn0030530) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00714}attP2	Bloomington Drosophila Stock Center	BDSC:32923; FlyBase:FBst0032923 ;
D. melanogaster. Expresses dsRNA for RNAi of Gug (FBgn0010825) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00756}attP2	Bloomington Drosophila Stock Center	BDSC:32961; FlyBase:FBst0032961 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sav (FBgn0053193) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00760}attP2	Bloomington Drosophila Stock Center	BDSC:32965; FlyBase:FBst0032965 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of bsk (FBgn0000229) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00777}attP2	Bloomington Drosophila Stock Center	BDSC:32977; FlyBase:FBst0032977 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pgcl (FBgn0011822) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00792}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:32992; FlyBase:FBst0032992 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of polo (FBgn0003124) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00530}attP2	Bloomington Drosophila Stock Center	BDSC:33042; FlyBase:FBst0033042 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of cnk (FBgn0286070) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00238}attP2	Bloomington Drosophila Stock Center	BDSC:33366; FlyBase:FBst0033366 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of kay (FBgn0001297) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00254}attP2	Bloomington Drosophila Stock Center	BDSC:33379; FlyBase:FBst0033379 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of apolpp (FBgn0087002) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00265}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:33388; FlyBase:FBst0033388 ;

D. melanogaster. Expresses dsRNA for RNAi of		
Su(var)205 (FBgn0003607) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00278}attP2	Bloomington Drosophila Stock Center	BDSC:33400; FlyBase:FBst0033400 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Tak1 (FBgn0026323) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00282}attP2	Bloomington Drosophila Stock Center	BDSC:33404; FlyBase:FBst0033404 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of hpo (FBgn0261456) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00006}attP2	Bloomington Drosophila Stock Center	BDSC:33614; FlyBase:FBst0033614 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Akt1 (FBgn0010379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00007}attP2	Bloomington Drosophila Stock Center	BDSC:33615; FlyBase:FBst0033615 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of N (FBgn0004647) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00009}attP2	Bloomington Drosophila Stock Center	BDSC:33616; FlyBase:FBst0033616 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of csw (FBgn0000382) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00012}attP2	Bloomington Drosophila Stock Center	BDSC:33619; FlyBase:FBst0033619 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tor (FBgn0003733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00021}attP2	Bloomington Drosophila Stock Center	BDSC:33627; FlyBase:FBst0033627 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Stat92E (FBgn0016917) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00035}attP2	Bloomington Drosophila Stock Center	BDSC:33637; FlyBase:FBst0033637 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Dsor1 (FBgn0010269) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00037}attP2	Bloomington Drosophila Stock Center	BDSC:33639; FlyBase:FBst0033639 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pten (FBgn0026379) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00044}attP2	Bloomington Drosophila Stock Center	BDSC:33643; FlyBase:FBst0033643 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CycD (FBgn0010315) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00059}attP2	Bloomington Drosophila Stock Center	BDSC:33653; FlyBase:FBst0033653 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rel (FBgn0014018) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00070}attP2	Bloomington Drosophila Stock Center	BDSC:33661; FlyBase:FBst0033661 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of upd1 (FBgn0004956) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00545}attP2	Bloomington Drosophila Stock Center	BDSC:33680; FlyBase:FBst0033680 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp3 (FBgn0044050) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00546}attP2	Bloomington Drosophila Stock Center	BDSC:33681; FlyBase:FBst0033681 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp4 (FBgn0044049) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00547}attP2	Bloomington Drosophila Stock Center	BDSC:33682; FlyBase:FBst0033682 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp5 (FBgn0044048) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00548}attP2	Bloomington Drosophila Stock Center	BDSC:33683; FlyBase:FBst0033683 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp6 (FBgn0044047) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00549}attP2	Bloomington Drosophila Stock Center	BDSC:33684; FlyBase:FBst0033684 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of d (FBgn0262029) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01096}attP2	Bloomington Drosophila Stock Center	BDSC:33754; FlyBase:FBst0033754 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Dad (FBgn0020493) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01102}attP2	Bloomington Drosophila Stock Center	BDSC:33759; FlyBase:FBst0033759 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of inaC (FBgn0004784) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02958}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:33768; FlyBase:FBst0033768 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wg (FBgn0284084) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00844}attP2	Bloomington Drosophila Stock Center	BDSC:33902; FlyBase:FBst0033902 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of srl (FBgn0037248) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00858}attP2	Bloomington Drosophila Stock Center	BDSC:33915; FlyBase:FBst0033915 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of SkpC		
(FBgn0026175) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00871}attP2	Bloomington Drosophila Stock Center	BDSC:33925; FlyBase:FBst0033925 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of SPE (FBgn0039102) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00873}attP2	Bloomington Drosophila Stock Center	BDSC:33926; FlyBase:FBst0033926 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Traf6 (FBgn0265464) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00880}attP2	Bloomington Drosophila Stock Center	BDSC:33931; FlyBase:FBst0033931 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of upd2 (FBgn0030904) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00901}attP2	Bloomington Drosophila Stock Center	BDSC:33949; FlyBase:FBst0033949 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Tor (FBgn0021796) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00904}attP2	Bloomington Drosophila Stock Center	BDSC:33951; FlyBase:FBst0033951 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dally (FBgn0263930) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00905}attP2	Bloomington Drosophila Stock Center	BDSC:33952; FlyBase:FBst0033952 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rheb (FBgn0041191) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00923}attP2	Bloomington Drosophila Stock Center	BDSC:33966; FlyBase:FBst0033966 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cat (FBgn0000261) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00990}attP2	Bloomington Drosophila Stock Center	BDSC:34020; FlyBase:FBst0034020 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CycK (FBgn0025674) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01003}attP2	Bloomington Drosophila Stock Center	BDSC:34032; FlyBase:FBst0034032 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wts (FBgn0011739) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00026}attP2	Bloomington Drosophila Stock Center	BDSC:34064; FlyBase:FBst0034064 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of yki (FBgn0034970) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00041}attP2	Bloomington Drosophila Stock Center	BDSC:34067; FlyBase:FBst0034067 ;

<i>D. melanogaster.</i> Expresses dsRNA for RNAi of srp (FBgn0003507) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01083}attP2	Bloomington Drosophila Stock Center	BDSC:34080; FlyBase:FBst0034080 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Fas2 (FBgn0000635) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01098}attP2	Bloomington Drosophila Stock Center	BDSC:34084; FlyBase:FBst0034084 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eIF4E1 (FBgn0015218) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00969}attP2	Bloomington Drosophila Stock Center	BDSC:34096; FlyBase:FBst0034096 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Sema1a (FBgn0011259) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01307}attP2	Bloomington Drosophila Stock Center	BDSC:34320; FlyBase:FBst0034320 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fz (FBgn0001085) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01308}attP2	Bloomington Drosophila Stock Center	BDSC:34321; FlyBase:FBst0034321 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fj (FBgn0000658) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01310}attP2	Bloomington Drosophila Stock Center	BDSC:34323; FlyBase:FBst0034323 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eve (FBgn0000606) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01312}attP2	Bloomington Drosophila Stock Center	BDSC:34325; FlyBase:FBst0034325 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tll (FBgn0003720) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01316}attP2	Bloomington Drosophila Stock Center	BDSC:34329; FlyBase:FBst0034329 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of aPKC (FBgn0261854) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01320}attP2	Bloomington Drosophila Stock Center	BDSC:34332; FlyBase:FBst0034332 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Vang (FBgn0015838) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01343}attP2	Bloomington Drosophila Stock Center	BDSC:34354; FlyBase:FBst0034354 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Lst8 (FBgn0264691) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01350}attP2	Bloomington Drosophila Stock Center	BDSC:34361; FlyBase:FBst0034361 ;

<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Lkb1 (FBgn0038167) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01351}attP2	Bloomington Drosophila Stock Center	BDSC:34362; FlyBase:FBst0034362 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of puc (FBgn0243512) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01386}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34392; FlyBase:FBst0034392 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Sxl (FBgn0264270) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00609}attP2	Bloomington Drosophila Stock Center	BDSC:34393; FlyBase:FBst0034393 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Cka (FBgn0044323) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00081}attP2	Bloomington Drosophila Stock Center	BDSC:34522; FlyBase:FBst0034522 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of bnl (FBgn0014135) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01046}attP2	Bloomington Drosophila Stock Center	BDSC:34572; FlyBase:FBst0034572 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of RagA- B (FBgn0037647) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01064}attP2	Bloomington Drosophila Stock Center	BDSC:34590; FlyBase:FBst0034590 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Wbp2 (FBgn0036318) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00563}attP2	Bloomington Drosophila Stock Center	BDSC:34603; FlyBase:FBst0034603 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pip (FBgn0003089) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01288}attP2	Bloomington Drosophila Stock Center	BDSC:34613; FlyBase:FBst0034613 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of dome (FBgn0043903) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01293}attP2	Bloomington Drosophila Stock Center	BDSC:34618; FlyBase:FBst0034618 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Stlk (FBgn0046692) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01295}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34620; FlyBase:FBst0034620 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of slp1 (FBgn0003430) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01107}attP2	Bloomington Drosophila Stock Center	BDSC:34633; FlyBase:FBst0034633 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of slp2 (FBgn0004567) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01108}attP2	Bloomington Drosophila Stock Center	BDSC:34634; FlyBase:FBst0034634 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of hth (FBgn0001235) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01112}attP2	Bloomington Drosophila Stock Center	BDSC:34637; FlyBase:FBst0034637 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of RhoGEF2 (FBgn0023172) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01118}attP2	Bloomington Drosophila Stock Center	BDSC:34643; FlyBase:FBst0034643 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Wnt5 (FBgn0010194) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01119}attP2	Bloomington Drosophila Stock Center	BDSC:34644; FlyBase:FBst0034644 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of spi (FBgn0005672) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01120}attP2	Bloomington Drosophila Stock Center	BDSC:34645; FlyBase:FBst0034645 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Galphao (FBgn0001122) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01129}attP2	Bloomington Drosophila Stock Center	BDSC:34653; FlyBase:FBst0034653 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of shn (FBgn0003396) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01167}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34689; FlyBase:FBst0034689 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ser (FBgn0004197) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01179}attP2	Bloomington Drosophila Stock Center	BDSC:34700; FlyBase:FBst0034700 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of H (FBgn0001169) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01182}attP2	Bloomington Drosophila Stock Center	BDSC:34703; FlyBase:FBst0034703 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of opa (FBgn0003002) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01185}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34706; FlyBase:FBst0034706 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Pkc53E (FBgn0003091) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01195}attP2	Bloomington Drosophila Stock Center	BDSC:34716; FlyBase:FBst0034716 ;

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<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hkb (FBgn0261434) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01216}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34736; FlyBase:FBst0034736 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gig (FBgn0005198) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01217}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34737; FlyBase:FBst0034737 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ken (FBgn0011236) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01219}attP2	Bloomington Drosophila Stock Center	BDSC:34739; FlyBase:FBst0034739 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of cact (FBgn0000250) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00084}attP2	Bloomington Drosophila Stock Center	BDSC:34775; FlyBase:FBst0034775 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Bx42 (FBgn0004856) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00086}attP2	Bloomington Drosophila Stock Center	BDSC:34777; FlyBase:FBst0034777 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rpt2 (FBgn0015282) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00104}attP2	Bloomington Drosophila Stock Center	BDSC:34795; FlyBase:FBst0034795 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Skp2 (FBgn0037236) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00116}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:34807; FlyBase:FBst0034807 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of raptor (FBgn0029840) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00124}attP2	Bloomington Drosophila Stock Center	BDSC:34814; FlyBase:FBst0034814 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Sos (FBgn0001965) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00149}attP2	Bloomington Drosophila Stock Center	BDSC:34833; FlyBase:FBst0034833 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Gprk2 (FBgn0261988) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00161}attP2	Bloomington Drosophila Stock Center	BDSC:34843; FlyBase:FBst0034843 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of rl (FBgn0003256) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00173}attP2	Bloomington Drosophila Stock Center	BDSC:34855; FlyBase:FBst0034855 ;

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D. melanogaster. Expresses dsRNA for RNAi of mwh (FBgn0264272) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00180}attP2	Bloomington Drosophila Stock Center	BDSC:34862; FlyBase:FBst0034862 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Apc (FBgn0015589) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00188}attP2	Bloomington Drosophila Stock Center	BDSC:34869; FlyBase:FBst0034869 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Tao (FBgn0031030) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01226}attP2	Bloomington Drosophila Stock Center	BDSC:34881; FlyBase:FBst0034881 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of 14-3- 3epsilon (FBgn0020238) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01229}attP2	Bloomington Drosophila Stock Center	BDSC:34884; FlyBase:FBst0034884 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Sara (FBgn0026369) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01239}attP2	Bloomington Drosophila Stock Center	BDSC:34894; FlyBase:FBst0034894 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gbb (FBgn0024234) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01243}attP2	Bloomington Drosophila Stock Center	BDSC:34898; FlyBase:FBst0034898 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Atg8b (FBgn0038539) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01245}attP2	Bloomington Drosophila Stock Center	BDSC:34900; FlyBase:FBst0034900 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rac1 (FBgn0010333) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01258}attP2	Bloomington Drosophila Stock Center	BDSC:34910; FlyBase:FBst0034910 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ato (FBgn0010433) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01278}attP2	Bloomington Drosophila Stock Center	BDSC:34929; FlyBase:FBst0034929 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dl (FBgn0260632) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00028}attP2	Bloomington Drosophila Stock Center	BDSC:34938; FlyBase:FBst0034938 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Mer (FBgn0086384) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00459}attP2	Bloomington Drosophila Stock Center	BDSC:34958; FlyBase:FBst0034958 ;

D. melanogaster. Expresses dsRNA for RNAi of mats		
(FBgn0038965) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00475}attP2	Bloomington Drosophila Stock Center	BDSC:34959; FlyBase:FBst0034959 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of ex (FBgn0004583) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00874}attP2	Bloomington Drosophila Stock Center	BDSC:34968; FlyBase:FBst0034968 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of ft (FBgn0001075) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00932}attP2	Bloomington Drosophila Stock Center	BDSC:34970; FlyBase:FBst0034970 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of msk (FBgn0026252) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01408}attP2	Bloomington Drosophila Stock Center	BDSC:34998; FlyBase:FBst0034998 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of baz (FBgn0000163) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01412}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35002; FlyBase:FBst0035002 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of arm (FBgn0000117) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01414}attP2	Bloomington Drosophila Stock Center	BDSC:35004; FlyBase:FBst0035004 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Stam (FBgn0027363) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01429}attP2	Bloomington Drosophila Stock Center	BDSC:35016; FlyBase:FBst0035016 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Gadd45 (FBgn0033153) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01436}attP2	Bloomington Drosophila Stock Center	BDSC:35023; FlyBase:FBst0035023 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of htl (FBgn0010389) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01437}attP2	Bloomington Drosophila Stock Center	BDSC:35024; FlyBase:FBst0035024 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tsh (FBgn0003866) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01443}attP2	Bloomington Drosophila Stock Center	BDSC:35030; FlyBase:FBst0035030 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Socs36E (FBgn0041184) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01450}attP2	Bloomington Drosophila Stock Center	BDSC:35036; FlyBase:FBst0035036 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Cad99C (FBgn0039709) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01451}attP2	Bloomington Drosophila Stock Center	BDSC:35037; FlyBase:FBst0035037 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pnt (FBgn0003118) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01452}attP2	Bloomington Drosophila Stock Center	BDSC:35038; FlyBase:FBst0035038 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of numb (FBgn0002973) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01459}attP2	Bloomington Drosophila Stock Center	BDSC:35045; FlyBase:FBst0035045 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of stan (FBgn0024836) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01464}attP2	Bloomington Drosophila Stock Center	BDSC:35050; FlyBase:FBst0035050 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dco (FBgn0002413) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00001}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35134; FlyBase:FBst0035134 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CkIIalpha (FBgn0264492) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00003}attP2	Bloomington Drosophila Stock Center	BDSC:35136; FlyBase:FBst0035136 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gish (FBgn0250823) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00005}attP2	Bloomington Drosophila Stock Center	BDSC:35138; FlyBase:FBst0035138 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mkk4 (FBgn0024326) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00010}attP2	Bloomington Drosophila Stock Center	BDSC:35143; FlyBase:FBst0035143 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CkIalpha (FBgn0015024) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00021}attP2	Bloomington Drosophila Stock Center	BDSC:35153; FlyBase:FBst0035153 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of lic (FBgn0261524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00022}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35154; FlyBase:FBst0035154 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Btk29A (FBgn0003502) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00027}attP2	Bloomington Drosophila Stock Center	BDSC:35159; FlyBase:FBst0035159 ;

D. melanogaster. Expresses dsRNA for RNAi of alc		
<pre>// Interanogaster: Expresses dsRNA for RNA of alc (FBgn0260972) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00029}attP2/TM3, Sb[1]</pre>	Bloomington Drosophila Stock Center	BDSC:35161; FlyBase:FBst0035161 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pka-C1 (FBgn0000273) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00038}attP2	Bloomington Drosophila Stock Center	BDSC:35169; FlyBase:FBst0035169 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hpo (FBgn0261456) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00046}attP2	Bloomington Drosophila Stock Center	BDSC:35176; FlyBase:FBst0035176 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of IKKbeta (FBgn0024222) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00058}attP2	Bloomington Drosophila Stock Center	BDSC:35186; FlyBase:FBst0035186 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Aduk (FBgn0037679) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00059}attP2	Bloomington Drosophila Stock Center	BDSC:35187; FlyBase:FBst0035187 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of put (FBgn0003169) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00069}attP2	Bloomington Drosophila Stock Center	BDSC:35195; FlyBase:FBst0035195 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hep (FBgn0010303) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00089}attP2	Bloomington Drosophila Stock Center	BDSC:35210; FlyBase:FBst0035210 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of brm (FBgn0000212) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00090}attP2	Bloomington Drosophila Stock Center	BDSC:35211; FlyBase:FBst0035211 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pyd (FBgn0262614) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00109}attP2	Bloomington Drosophila Stock Center	BDSC:35225; FlyBase:FBst0035225 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hyx (FBgn0037657) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00123}attP2	Bloomington Drosophila Stock Center	BDSC:35238; FlyBase:FBst0035238 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of p38a (FBgn0015765) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00131}attP2	Bloomington Drosophila Stock Center	BDSC:35244; FlyBase:FBst0035244 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of p38b (FBgn0024846) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00140}attP2	Bloomington Drosophila Stock Center	BDSC:35252; FlyBase:FBst0035252 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of app (FBgn0260941) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00181}attP2	Bloomington Drosophila Stock Center	BDSC:35280; FlyBase:FBst0035280 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sdt (FBgn0261873) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00193}attP2	Bloomington Drosophila Stock Center	BDSC:35291; FlyBase:FBst0035291 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of aux (FBgn0037218) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00213}attP2	Bloomington Drosophila Stock Center	BDSC:35310; FlyBase:FBst0035310 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CaMKII (FBgn0264607) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00237}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35330; FlyBase:FBst0035330 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ask1 (FBgn0014006) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00238}attP2	Bloomington Drosophila Stock Center	BDSC:35331; FlyBase:FBst0035331 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of SAK (FBgn0026371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00244}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35335; FlyBase:FBst0035335 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sgg (FBgn0003371) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00277}attP2	Bloomington Drosophila Stock Center	BDSC:35364; FlyBase:FBst0035364 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hop (FBgn0004864) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00305}attP2	Bloomington Drosophila Stock Center	BDSC:35386; FlyBase:FBst0035386 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mekk1 (FBgn0024329) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00322}attP2	Bloomington Drosophila Stock Center	BDSC:35402; FlyBase:FBst0035402 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of aop (FBgn0000097) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00324}attP2	Bloomington Drosophila Stock Center	BDSC:35404; FlyBase:FBst0035404 ;

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<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ras85D (FBgn0003205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00336}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:35414; FlyBase:FBst0035414 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Nap1 (FBgn0015268) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00370}attP2	Bloomington Drosophila Stock Center	BDSC:35445; FlyBase:FBst0035445 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of osa (FBgn0261885) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00372}attP2	Bloomington Drosophila Stock Center	BDSC:35447; FlyBase:FBst0035447 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of key (FBgn0041205) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00088}attP2	Bloomington Drosophila Stock Center	BDSC:35572; FlyBase:FBst0035572 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Stat92E (FBgn0016917) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00437}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:35600; FlyBase:FBst0035600 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Gcn5 (FBgn0020388) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00439}attP40	Bloomington Drosophila Stock Center	BDSC:35601; FlyBase:FBst0035601 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of TER94 (FBgn0261014) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00448}attP2	Bloomington Drosophila Stock Center	BDSC:35608; FlyBase:FBst0035608 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wek (FBgn0001990) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21045}attP2	Bloomington Drosophila Stock Center	BDSC:35680; FlyBase:FBst0035680 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of bab2 (FBgn0025525) under UAS control in the VALIUM21 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLV21085}attP2	Bloomington Drosophila Stock Center	BDSC:35720; FlyBase:FBst0035720 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Patj (FBgn0067864) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01489}attP2	Bloomington Drosophila Stock Center	BDSC:35747; FlyBase:FBst0035747 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mtl (FBgn0039532) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01500}attP2	Bloomington Drosophila Stock Center	BDSC:35754; FlyBase:FBst0035754 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gro (FBgn0001139) under UAS control in the VALIUM20	Plaamington	PDSC:25750:
vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01506}attP2	Bloomington Drosophila Stock Center	BDSC:35759; FlyBase:FBst0035759 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Wnt6 (FBgn0031902) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00457}attP2	Bloomington Drosophila Stock Center	BDSC:35808; FlyBase:FBst0035808 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CycE (FBgn0010382) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00511}attP40	Bloomington Drosophila Stock Center	BDSC:36092; FlyBase:FBst0036092 ;
D. melanogaster. Expresses dsRNA for RNAi of Myd88 (FBgn0033402) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00183}attP2	Bloomington Drosophila Stock Center	BDSC:36107; FlyBase:FBst0036107 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Cul1 (FBgn0015509) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00561}attP2	Bloomington Drosophila Stock Center	BDSC:36601; FlyBase:FBst0036601 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of chico (FBgn0024248) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01553}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36665; FlyBase:FBst0036665 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rbf2 (FBgn0038390) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01586}attP2	Bloomington Drosophila Stock Center	BDSC:36697; FlyBase:FBst0036697 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of rictor (FBgn0031006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01588}attP2	Bloomington Drosophila Stock Center	BDSC:36699; FlyBase:FBst0036699 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sty (FBgn0014388) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01599}attP2	Bloomington Drosophila Stock Center	BDSC:36709; FlyBase:FBst0036709 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Zyx (FBgn0011642) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01606}attP40	Bloomington Drosophila Stock Center	BDSC:36716; FlyBase:FBst0036716 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Wnt2 (FBgn0004360) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01613}attP2	Bloomington Drosophila Stock Center	BDSC:36722; FlyBase:FBst0036722 ;

D. melanogaster. Expresses dsRNA for RNAi of Rbf		
(FBgn0015799) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS03004}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36744; FlyBase:FBst0036744 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of dlg1 (FBgn0001624) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02287}attP2	Bloomington Drosophila Stock Center	BDSC:36771; FlyBase:FBst0036771 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dpp (FBgn0000490) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.JF02455}attP2	Bloomington Drosophila Stock Center	BDSC:36779; FlyBase:FBst0036779 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of DI (FBgn0000463) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00520}attP40	Bloomington Drosophila Stock Center	BDSC:36784; FlyBase:FBst0036784 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of HDAC1 (FBgn0015805) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01005}attP40	Bloomington Drosophila Stock Center	BDSC:36800; FlyBase:FBst0036800 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Su(dx) (FBgn0003557) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01077}attP2	Bloomington Drosophila Stock Center	BDSC:36836; FlyBase:FBst0036836 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of pbl (FBgn0003041) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01092}attP2	Bloomington Drosophila Stock Center	BDSC:36841; FlyBase:FBst0036841 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Sod2 (FBgn0010213) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01015}attP40	Bloomington Drosophila Stock Center	BDSC:36871; FlyBase:FBst0036871 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Art1 (FBgn0037834) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01072}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:36891; FlyBase:FBst0036891 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG10924 (FBgn0034356) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00200}attP2	Bloomington Drosophila Stock Center	BDSC:36915; FlyBase:FBst0036915 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of trr (FBgn0023518) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01019}attP2	Bloomington Drosophila Stock Center	BDSC:36916; FlyBase:FBst0036916 ;

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D. melanogaster. Expresses dsRNA for RNAi of Cdc42 (FBgn0010341) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00620}attP40	Bloomington Drosophila Stock Center	BDSC:37477; FlyBase:FBst0037477 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of nej (FBgn0261617) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01507}attP2	Bloomington Drosophila Stock Center	BDSC:37489; FlyBase:FBst0037489 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pvr (FBgn0032006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01662}attP40	Bloomington Drosophila Stock Center	BDSC:37520; FlyBase:FBst0037520 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of cno (FBgn0259212) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00633}attP40	Bloomington Drosophila Stock Center	BDSC:38194; FlyBase:FBst0038194 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pygo (FBgn0043900) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00647}attP40	Bloomington Drosophila Stock Center	BDSC:38208; FlyBase:FBst0038208 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ed (FBgn0000547) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00648}attP40	Bloomington Drosophila Stock Center	BDSC:38209; FlyBase:FBst0038209 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Nrg (FBgn0264975) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00656}attP40	Bloomington Drosophila Stock Center	BDSC:38215; FlyBase:FBst0038215 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of aph-1 (FBgn0031458) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01693}attP40	Bloomington Drosophila Stock Center	BDSC:38249; FlyBase:FBst0038249 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of cbt (FBgn0043364) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01726}attP40	Bloomington Drosophila Stock Center	BDSC:38276; FlyBase:FBst0038276 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Vps36 (FBgn0086785) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01739}attP40	Bloomington Drosophila Stock Center	BDSC:38286; FlyBase:FBst0038286 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of crb (FBgn0259685) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01842}attP40	Bloomington Drosophila Stock Center	BDSC:38373; FlyBase:FBst0038373 ;

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D. melanogaster. Expresses dsRNA for RNAi of tws (FBgn0004889) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00670}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38899; FlyBase:FBst0038899 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of S (FBgn0003310) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL00686}attP2	Bloomington Drosophila Stock Center	BDSC:38914; FlyBase:FBst0038914 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pvf3 (FBgn0085407) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01876}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:38962; FlyBase:FBst0038962 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CanB2 (FBgn0015614) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01886}attP2	Bloomington Drosophila Stock Center	BDSC:38971; FlyBase:FBst0038971 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pi3K21B (FBgn0020622) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01907}attP40	Bloomington Drosophila Stock Center	BDSC:38991; FlyBase:FBst0038991 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of par-6 (FBgn0026192) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01928}attP40	Bloomington Drosophila Stock Center	BDSC:39010; FlyBase:FBst0039010 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pvf1 (FBgn0030964) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01958}attP40	Bloomington Drosophila Stock Center	BDSC:39038; FlyBase:FBst0039038 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pka-C3 (FBgn0000489) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01970}attP2	Bloomington Drosophila Stock Center	BDSC:39050; FlyBase:FBst0039050 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of DAAM (FBgn0025641) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01978}attP2	Bloomington Drosophila Stock Center	BDSC:39058; FlyBase:FBst0039058 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of scrib (FBgn0263289) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01993}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:39073; FlyBase:FBst0039073 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sina (FBgn0003410) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02008}attP40	Bloomington Drosophila Stock Center	BDSC:40842; FlyBase:FBst0040842 ;

<i>D. melanogaster.</i> Expresses dsRNA for RNAi of btl (FBgn0285896) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02038}attP2	Bloomington Drosophila Stock Center	BDSC:40871; FlyBase:FBst0040871 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pp2B- 14D (FBgn0011826) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02039}attP2	Bloomington Drosophila Stock Center	BDSC:40872; FlyBase:FBst0040872 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Smurf (FBgn0029006) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02153}attP40	Bloomington Drosophila Stock Center	BDSC:40905; FlyBase:FBst0040905 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CycB (FBgn0000405) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02163}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:40915; FlyBase:FBst0040915 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fu (FBgn0001079) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00705}attP40	Bloomington Drosophila Stock Center	BDSC:41588; FlyBase:FBst0041588 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ru (FBgn0003295) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01129}attP2	Bloomington Drosophila Stock Center	BDSC:41593; FlyBase:FBst0041593 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ksr (FBgn0015402) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01134}attP2	Bloomington Drosophila Stock Center	BDSC:41598; FlyBase:FBst0041598 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of slpr (FBgn0030018) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01187}attP2	Bloomington Drosophila Stock Center	BDSC:41605; FlyBase:FBst0041605 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of p53 (FBgn0039044) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01220}attP40	Bloomington Drosophila Stock Center	BDSC:41638; FlyBase:FBst0041638 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Smox (FBgn0025800) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02203}attP40	Bloomington Drosophila Stock Center	BDSC:41670; FlyBase:FBst0041670 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of rho (FBgn0004635) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02264}attP40	Bloomington Drosophila Stock Center	BDSC:41699; FlyBase:FBst0041699 ;

D. melanogaster. Expresses dsRNA for RNAi of S6k		
(FBgn0283472) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02267}attP2	Bloomington Drosophila Stock Center	BDSC:41702; FlyBase:FBst0041702 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of snk (FBgn0003450) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02289}attP2	Bloomington Drosophila Stock Center	BDSC:41723; FlyBase:FBst0041723 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ufd1- like (FBgn0036136) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01251}attP2	Bloomington Drosophila Stock Center	BDSC:41823; FlyBase:FBst0041823 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of RasGAP1 (FBgn0004390) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01258}attP2	Bloomington Drosophila Stock Center	BDSC:41830; FlyBase:FBst0041830 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of 14-3- 3zeta (FBgn0004907) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01310}attP40	Bloomington Drosophila Stock Center	BDSC:41878; FlyBase:FBst0041878 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tkv (FBgn0003716) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01338}attP2	Bloomington Drosophila Stock Center	BDSC:41904; FlyBase:FBst0041904 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of homer (FBgn0025777) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02301}attP2	Bloomington Drosophila Stock Center	BDSC:41908; FlyBase:FBst0041908 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pll (FBgn0010441) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02332}attP40	Bloomington Drosophila Stock Center	BDSC:41935; FlyBase:FBst0041935 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of elB (FBgn0004858) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02357}attP2	Bloomington Drosophila Stock Center	BDSC:41960; FlyBase:FBst0041960 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ea (FBgn0000533) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02358}attP2	Bloomington Drosophila Stock Center	BDSC:41961; FlyBase:FBst0041961 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CycB3 (FBgn0015625) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02377}attP2	Bloomington Drosophila Stock Center	BDSC:41979; FlyBase:FBst0041979 ;

D. melanogaster. Expresses dsRNA for RNAi of Igs		
(FBgn0039907) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02381}attP2	Bloomington Drosophila Stock Center	BDSC:41983; FlyBase:FBst0041983 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Actbeta (FBgn0024913) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02057}attP40	Bloomington Drosophila Stock Center	BDSC:42493; FlyBase:FBst0042493 ;
D. melanogaster. Expresses dsRNA for RNAi of CG5059 (FBgn0037007) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02058}attP40	Bloomington Drosophila Stock Center	BDSC:42494; FlyBase:FBst0042494 ;
D. melanogaster. Expresses dsRNA for RNAi of msn (FBgn0010909) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02084}attP40	Bloomington Drosophila Stock Center	BDSC:42518; FlyBase:FBst0042518 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rassf (FBgn0039055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02102}attP40	Bloomington Drosophila Stock Center	BDSC:42534; FlyBase:FBst0042534 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of sax (FBgn0003317) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02118}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:42546; FlyBase:FBst0042546 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of et (FBgn0031055) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02213}attP40	Bloomington Drosophila Stock Center	BDSC:42557; FlyBase:FBst0042557 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of nmo (FBgn0011817) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ02229}attP40	Bloomington Drosophila Stock Center	BDSC:42570; FlyBase:FBst0042570 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG12147 (FBgn0037325) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02447}attP2	Bloomington Drosophila Stock Center	BDSC:42612; FlyBase:FBst0042612 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Act5C (FBgn0000042) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02487}attP2	Bloomington Drosophila Stock Center	BDSC:42651; FlyBase:FBst0042651 ;
D. melanogaster. Expresses dsRNA for RNAi of Act87E (FBgn0000046) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02488}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:42652; FlyBase:FBst0042652 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of emc (FBgn0000575) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL00724}attP2	Bloomington Drosophila Stock Center	BDSC:42768; FlyBase:FBst0042768 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of the Stellate gene family (FBgn0003523) plus Ste12DOR and SteXh:CG42398 (FBgn0044817 and FBgn0259817) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01156}attP2	Bloomington Drosophila Stock Center	BDSC:42786; FlyBase:FBst0042786 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Socs44A (FBgn0033266) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02515}attP2	Bloomington Drosophila Stock Center	BDSC:42830; FlyBase:FBst0042830 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gcm (FBgn0014179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02582}attP40	Bloomington Drosophila Stock Center	BDSC:42889; FlyBase:FBst0042889 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CkIlbeta (FBgn0000259) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02636}attP40	Bloomington Drosophila Stock Center	BDSC:42943; FlyBase:FBst0042943 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of smo (FBgn0003444) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01472}attP2	Bloomington Drosophila Stock Center	BDSC:43134; FlyBase:FBst0043134 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dock (FBgn0010583) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GL01519}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43176; FlyBase:FBst0043176 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mad (FBgn0011648) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01527}attP40	Bloomington Drosophila Stock Center	BDSC:43183; FlyBase:FBst0043183 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Pp2A- 29B (FBgn0260439) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS01921}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:43283; FlyBase:FBst0043283 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of CG11658 (FBgn0036196) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02671}attP40	Bloomington Drosophila Stock Center	BDSC:43298; FlyBase:FBst0043298 ;

D. melanogaster. Expresses dsRNA for RNAi of Myc		
(FBgn0262656) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01314}attP40	Bloomington Drosophila Stock Center	BDSC:43962; FlyBase:FBst0043962 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pav (FBgn0011692) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GL01316}attP40	Bloomington Drosophila Stock Center	BDSC:43963; FlyBase:FBst0043963 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eIF4EHP (FBgn0053100) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02703}attP40	Bloomington Drosophila Stock Center	BDSC:43990; FlyBase:FBst0043990 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Atg1 (FBgn0260945) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02750}attP40	Bloomington Drosophila Stock Center	BDSC:44034; FlyBase:FBst0044034 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tefu (FBgn0045035) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02790}attP40	Bloomington Drosophila Stock Center	BDSC:44073; FlyBase:FBst0044073 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of bi (FBgn0000179) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02815}attP2	Bloomington Drosophila Stock Center	BDSC:44095; FlyBase:FBst0044095 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of mad2 (FBgn0035640) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01381}attP2	Bloomington Drosophila Stock Center	BDSC:44430; FlyBase:FBst0044430 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ebi (FBgn0263933) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01413}attP40	Bloomington Drosophila Stock Center	BDSC:44443; FlyBase:FBst0044443 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dx (FBgn0000524) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01607}attP2	Bloomington Drosophila Stock Center	BDSC:44455; FlyBase:FBst0044455 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fz3 (FBgn0027343) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01626}attP2	Bloomington Drosophila Stock Center	BDSC:44468; FlyBase:FBst0044468 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of cos (FBgn0000352) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02347}attP2	Bloomington Drosophila Stock Center	BDSC:44472; FlyBase:FBst0044472 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of disp (FBgn0029088) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02877}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:44633; FlyBase:FBst0044633 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of spen (FBgn0016977) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01647}attP40	Bloomington Drosophila Stock Center	BDSC:50529; FlyBase:FBst0050529 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dlp (FBgn0041604) under UAS control in the VALIUM22 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01658}attP40	Bloomington Drosophila Stock Center	BDSC:50540; FlyBase:FBst0050540 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wgn (FBgn0030941) under UAS control in the VALIUM22 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.GLC01716}attP2	Bloomington Drosophila Stock Center	BDSC:50594; FlyBase:FBst0050594 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of RpL8 (FBgn0261602) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02977}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50610; FlyBase:FBst0050610 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Act42A (FBgn0000043) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC02992}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50625; FlyBase:FBst0050625 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tin (FBgn0004110) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03064}attP2	Bloomington Drosophila Stock Center	BDSC:50663; FlyBase:FBst0050663 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of EcR (FBgn0000546) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03114}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:50712; FlyBase:FBst0050712 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of aru (FBgn0029095) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS02966}attP2	Bloomington Drosophila Stock Center	BDSC:50730; FlyBase:FBst0050730 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eIF4E4 (FBgn0035709) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21052}attP40	Bloomington Drosophila Stock Center	BDSC:50951; FlyBase:FBst0050951 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pont (FBgn0040078) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21078}attP40	Bloomington Drosophila Stock Center	BDSC:50972; FlyBase:FBst0050972 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dpn (FBgn0010109) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03154}attP2	Bloomington Drosophila Stock Center	BDSC:51440; FlyBase:FBst0051440 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Spn27A (FBgn0028990) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03159}attP2	Bloomington Drosophila Stock Center	BDSC:51445; FlyBase:FBst0051445 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ttv (FBgn0265974) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03225}attP40	Bloomington Drosophila Stock Center	BDSC:51480; FlyBase:FBst0051480 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mipp2 (FBgn0026060) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03229}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:51482; FlyBase:FBst0051482 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of kibra (FBgn0262127) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03256}attP2	Bloomington Drosophila Stock Center	BDSC:51499; FlyBase:FBst0051499 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tld (FBgn0003719) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03275}attP2	Bloomington Drosophila Stock Center	BDSC:51507; FlyBase:FBst0051507 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of S6kII (FBgn0262866) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03140}attP40	Bloomington Drosophila Stock Center	BDSC:51694; FlyBase:FBst0051694 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ast (FBgn0015905) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03173}attP2	Bloomington Drosophila Stock Center	BDSC:51700; FlyBase:FBst0051700 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ras (FBgn0003204) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03250}attP2	Bloomington Drosophila Stock Center	BDSC:51717; FlyBase:FBst0051717 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of E(spl)mgamma-HLH (FBgn0002735) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03315}attP2	Bloomington Drosophila Stock Center	BDSC:51762; FlyBase:FBst0051762 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Src64B (FBgn0262733) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03327}attP40	Bloomington Drosophila Stock Center	BDSC:51772; FlyBase:FBst0051772 ;

D. melanogaster. Expresses dsRNA for RNAi of brk		
(FBgn0024250) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03345}attP2	Bloomington Drosophila Stock Center	BDSC:51789; FlyBase:FBst0051789 ;
D. melanogaster. Expresses dsRNA for RNAi of Ext2 (FBgn0029175) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03621}attP40	Bloomington Drosophila Stock Center	BDSC:52883; FlyBase:FBst0052883 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pen-2 (FBgn0053198) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03648}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:52908; FlyBase:FBst0052908 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Tsc1 (FBgn0026317) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03672}attP40	Bloomington Drosophila Stock Center	BDSC:52931; FlyBase:FBst0052931 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of NT1 (FBgn0261526) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21720}attP40	Bloomington Drosophila Stock Center	BDSC:53003; FlyBase:FBst0053003 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of arr (FBgn0000119) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03571}attP40	Bloomington Drosophila Stock Center	BDSC:53342; FlyBase:FBst0053342 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of kermit (FBgn0010504) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03578}attP40	Bloomington Drosophila Stock Center	BDSC:53349; FlyBase:FBst0053349 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Sirt1 (FBgn0024291) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21708}attP40	Bloomington Drosophila Stock Center	BDSC:53697; FlyBase:FBst0053697 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tow (FBgn0035719) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21747}attP40	Bloomington Drosophila Stock Center	BDSC:53704; FlyBase:FBst0053704 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eIF4E3 (FBgn0265089) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21195}attP40	Bloomington Drosophila Stock Center	BDSC:53880; FlyBase:FBst0053880 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of hppy (FBgn0263395) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21199}attP40	Bloomington Drosophila Stock Center	BDSC:53884; FlyBase:FBst0053884 ;

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<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dome (FBgn0043903) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21208}attP40	Bloomington Drosophila Stock Center	BDSC:53890; FlyBase:FBst0053890 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Ocho (FBgn0040296) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21588}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:54851; FlyBase:FBst0054851 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of sqd (FBgn0263396) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03848}attP40	Bloomington Drosophila Stock Center	BDSC:55169; FlyBase:FBst0055169 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of mago (FBgn0002736) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03947}attP40	Bloomington Drosophila Stock Center	BDSC:55260; FlyBase:FBst0055260 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of egr (FBgn0033483) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03963}attP40	Bloomington Drosophila Stock Center	BDSC:55276; FlyBase:FBst0055276 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of E(spl)m3-HLH (FBgn0002609) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03989}attP2	Bloomington Drosophila Stock Center	BDSC:55302; FlyBase:FBst0055302 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Coprox (FBgn0021944) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04005}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:55318; FlyBase:FBst0055318 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rab23 (FBgn0037364) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04039}attP40	Bloomington Drosophila Stock Center	BDSC:55352; FlyBase:FBst0055352 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tsu (FBgn0033378) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04055}attP40	Bloomington Drosophila Stock Center	BDSC:55367; FlyBase:FBst0055367 ;
<i>D. melanogaster.</i> Expresses dsRNA for RNAi of Notum (FBgn0044028) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04067}attP40	Bloomington Drosophila Stock Center	BDSC:55379; FlyBase:FBst0055379 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sd (FBgn0003345) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04092}attP40	Bloomington Drosophila Stock Center	BDSC:55404; FlyBase:FBst0055404 ;

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Raf (FBgn0003079) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03854}attP2	Bloomington Drosophila Stock Center	BDSC:55679; FlyBase:FBst0055679 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Mo25 (FBgn0017572) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03865}attP2	Bloomington Drosophila Stock Center	BDSC:55681; FlyBase:FBst0055681 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ptc (FBgn0003892) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC03872}attP40	Bloomington Drosophila Stock Center	BDSC:55686; FlyBase:FBst0055686 ;
D. melanogaster. Expresses dsRNA for RNAi of Pka-C2 (FBgn0000274) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04129}attP2	Bloomington Drosophila Stock Center	BDSC:55859; FlyBase:FBst0055859 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sev (FBgn0003366) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04136}attP2	Bloomington Drosophila Stock Center	BDSC:55866; FlyBase:FBst0055866 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of shf (FBgn0003390) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04137}attP2	Bloomington Drosophila Stock Center	BDSC:55867; FlyBase:FBst0055867 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of babo (FBgn0011300) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04142}attP2/TM3, Sb[1]	Bloomington Drosophila Stock Center	BDSC:55871; FlyBase:FBst0055871 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Shark (FBgn0015295) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04146}attP2	Bloomington Drosophila Stock Center	BDSC:55874; FlyBase:FBst0055874 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Drl-2 (FBgn0033791) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04172}attP2	Bloomington Drosophila Stock Center	BDSC:55893; FlyBase:FBst0055893 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Takl2 (FBgn0039015) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04181}attP2	Bloomington Drosophila Stock Center	BDSC:55899; FlyBase:FBst0055899 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Takl1 (FBgn0046689) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04186}attP2	Bloomington Drosophila Stock Center	BDSC:55903; FlyBase:FBst0055903 ;

D. melanogaster. Expresses dsRNA for RNAi of Doa		
(FBgn0265998) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04193}attP2	Bloomington Drosophila Stock Center	BDSC:55908; FlyBase:FBst0055908 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fus (FBgn0023441) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04208}attP40	Bloomington Drosophila Stock Center	BDSC:55921; FlyBase:FBst0055921 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of grk (FBgn0001137) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04213}attP40	Bloomington Drosophila Stock Center	BDSC:55926; FlyBase:FBst0055926 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of PpD3 (FBgn0005777) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS04508}attP40	Bloomington Drosophila Stock Center	BDSC:57307; FlyBase:FBst0057307 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Nct (FBgn0039234) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04812}attP40	Bloomington Drosophila Stock Center	BDSC:57497; FlyBase:FBst0057497 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG15436 (FBgn0031610) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC04637}attP40	Bloomington Drosophila Stock Center	BDSC:57867; FlyBase:FBst0057867 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Su(var)2-10 (FBgn0003612) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ21959}attP40	Bloomington Drosophila Stock Center	BDSC:58067; FlyBase:FBst0058067 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Atg8a (FBgn0052672) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ22416}attP40	Bloomington Drosophila Stock Center	BDSC:58309; FlyBase:FBst0058309 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of spz (FBgn0003495) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ22258}attP40	Bloomington Drosophila Stock Center	BDSC:58499; FlyBase:FBst0058499 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of AstC- R1 (FBgn0036790) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ23767}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:62372; FlyBase:FBst0062372 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ku80 (FBgn0041627) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ24057}attP40/CyO	Bloomington Drosophila Stock Center	BDSC:62513; FlyBase:FBst0062513 ;

D. malanamatar, European de DNA far DNA; of		
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG2199 (FBgn0035213) under UAS control in the VALIUM20 vector.: y[1] v[1]; P{y[+t7.7] v[+t1.8]=TRiP.HMJ30228}attP40	Bloomington Drosophila Stock Center	BDSC:63661; FlyBase:FBst0063661 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tsr (FBgn0011726) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS00534}attP2	Bloomington Drosophila Stock Center	BDSC:65055; FlyBase:FBst0065055 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG3630 (FBgn0023540) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMC06220}attP2	Bloomington Drosophila Stock Center	BDSC:65945; FlyBase:FBst0065945 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tub (FBgn0003882) under UAS control in the VALIUM20 vector.: y[1] sc[*] v[1] sev[21]; P{y[+t7.7] v[+t1.8]=TRiP.HMS05426}attP40	Bloomington Drosophila Stock Center	BDSC:66960; FlyBase:FBst0066960 ;
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eIF4E-6 (FBgn0039622) under UAS control.	Vienna Drosophila Resource Center	VDRC:v17580
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gd (FBgn0000808) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14892
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Act88F (FBgn0000047) under UAS control.	Vienna Drosophila Resource Center	VDRC:v9780
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of eRF1 (FBgn0036974) under UAS control.	Vienna Drosophila Resource Center	VDRC:v45027
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of E(spl)m2-BFM (FBgn0002592) under UAS control.	Vienna Drosophila Resource Center	VDRC:v30115
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tsl (FBgn0003867) under UAS control.	Vienna Drosophila Resource Center	VDRC:v14430
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wbl (FBgn0004003) under UAS control.	Vienna Drosophila Resource Center	VDRC:v13864
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of boss (FBgn0000206) under UAS control.	Vienna Drosophila Resource Center	VDRC:v4365
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG32396 (FBgn0020251) under UAS control.	Vienna Drosophila Resource Center	VDRC:v41896
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Rac2 (FBgn0014011) under UAS control.	Vienna Drosophila Resource Center	VDRC:v28926
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of ihog (FBgn0031872) under UAS control.	Vienna Drosophila Resource Center	VDRC:v29898
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of sog (FBgn0003463) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37405
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG9314 (FBgn0032061) under UAS control.	Vienna Drosophila Resource Center	VDRC:v44647

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Vienna Drosophila Resource Center	VDRC:v29434
Vienna Drosophila Resource Center	VDRC:v50134
Vienna Drosophila Resource Center	VDRC:v31364
Vienna Drosophila Resource Center	VDRC:v6459
Vienna Drosophila Resource Center	VDRC:v5594
Vienna Drosophila Resource Center	VDRC:v17118
Vienna Drosophila Resource Center	VDRC:v32146
Vienna Drosophila Resource Center	VDRC:v36524
Vienna Drosophila Resource Center	VDRC:v17282
Vienna Drosophila Resource Center	VDRC:v7628
Vienna Drosophila Resource Center	VDRC:v7679
Vienna Drosophila Resource Center	VDRC:v15384
Vienna Drosophila Resource Center	VDRC:v37186
Vienna Drosophila Resource Center	VDRC:v7261
Vienna Drosophila Resource Center	VDRC:v7434
Vienna Drosophila Resource Center	VDRC:v45981
Vienna Drosophila Resource Center	VDRC:v26019
Vienna Drosophila Resource Center	VDRC:v3004
Vienna Drosophila Resource Center	VDRC:v3060
Vienna Drosophila Resource Center	VDRC:v17903
	Vienna Drosophila Resource CenterVienna Drosophila Resource Center

D. melanogaster. Expresses dsRNA for RNAi of	Vienna Drosophila	
CG45087; Pepck (FBgn0003067) under UAS control.	Resource Center	VDRC:v20529
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of RpL13A (FBgn0037351) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101369
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Usp7 (FBgn0030366) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110324
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of tsg (FBgn0003865) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108750
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of nec (FBgn0002930) under UAS control.	Vienna Drosophila Resource Center	VDRC:v108366
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Nle (FBgn0021874) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110728
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Brd (FBgn0000216) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107929
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Shc (FBgn0015296) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103906
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Hs6st (FBgn0038755) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110424
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fz4 (FBgn0027342) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102339
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of bib (FBgn0000180) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103327
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Wnt10 (FBgn0031903) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100867
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Tom (FBgn0026320) under UAS control.	Vienna Drosophila Resource Center	VDRC:v101652
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Pli (FBgn0025574) under UAS control.	Vienna Drosophila Resource Center	VDRC:v106776
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of por (FBgn0004957) under UAS control.	Vienna Drosophila Resource Center	VDRC:v100780
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of wls (FBgn0036141) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103812
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG6843 (FBgn0036827) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109411
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of spz3 (FBgn0031959) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102871
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of kuz (FBgn0259984) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107036

Vienna Drosophila Resource Center	VDRC:v110601
Vienna Drosophila Resource Center	VDRC:v106335
Vienna Drosophila Resource Center	VDRC:v100611
Vienna Drosophila Resource Center	VDRC:v100568
Vienna Drosophila Resource Center	VDRC:v109384
Vienna Drosophila Resource Center	VDRC:v101965
Vienna Drosophila Resource Center	VDRC:v105281
Vienna Drosophila Resource Center	VDRC:v105110
Vienna Drosophila Resource Center	VDRC:v106521
Vienna Drosophila Resource Center	VDRC:v102633
Vienna Drosophila Resource Center	VDRC:v100897
Vienna Drosophila Resource Center	VDRC:v104675
Vienna Drosophila Resource Center	VDRC:v108721
Vienna Drosophila Resource Center	VDRC:v102389
Vienna Drosophila Resource Center	VDRC:v102129
Vienna Drosophila Resource Center	VDRC:v102818
Vienna Drosophila Resource Center	VDRC:v109437
Vienna Drosophila Resource Center	VDRC:v106141
Vienna Drosophila Resource Center	VDRC:v109539
Vienna Drosophila Resource Center	VDRC:v106572
	Vienna Drosophila Resource Center Vienna Drosophila Resource Center

<i>D. melanogaster</i> . Expresses dsRNA for RNAi of kek1	Vienna Drosophila	
(FBgn0015399) under UAS control.	Resource Center	VDRC:v101166
D. melanogaster. Expresses dsRNA for RNAi of ths	Vienna Drosophila	
(FBgn0033652) under UAS control.	Resource Center	VDRC:v102441
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Ilp8 (FBgn0036690) under UAS control.	Vienna Drosophila Resource Center	VDRC:v102604
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of	Vienna Drosophila	
CG15800 (FBgn0034904) under UAS control.	Resource Center	VDRC:v110049
D. melanogaster. Expresses dsRNA for RNAi of IM3	Vienna Drosophila	
(FBgn0040736) under UAS control.	Resource Center	VDRC:v104908
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Roc1a	Vienna Drosophila	
(FBgn0025638) under UAS control.	Resource Center	VDRC:v106315
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dod (FBgn0015379) under UAS control.	Vienna Drosophila Resource Center	VDRC:v110593
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of hipk	Vienna Drosophila	
(FBgn0035142) under UAS control.	Resource Center	VDRC:v108254
D. melanogaster. Expresses dsRNA for RNAi of ave	Vienna Drosophila	
(FBgn0050476) under UAS control.	Resource Center	VDRC:v101471
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of boca	Vienna Drosophila	
(FBgn0004132) under UAS control.	Resource Center	VDRC:v108406
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of gskt (FBgn0046332) under UAS control.	Vienna Drosophila Resource Center	VDRC:v107429
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of stumps	Vienna Drosophila	
(FBgn0020299) under UAS control.	Resource Center	VDRC:v105603
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of CG31431 (FBgn0051431) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104697
D. melanogaster. Expresses dsRNA for RNAi of scw	Vienna Drosophila	
(FBgn0005590) under UAS control.	Resource Center	VDRC:v105303
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of fry (FBgn0016081) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103569
		VDRC.V103509
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of Krn (FBgn0052179) under UAS control.	Vienna Drosophila Resource Center	VDRC:v104299
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of pxb	Vienna Drosophila	
(FBgn0053207) under UAS control.	Resource Center	VDRC:v102240
D. melanogaster. Expresses dsRNA for RNAi of cv-c	Vienna Drosophila	
(FBgn0285955) under UAS control.	Resource Center	VDRC:v105435
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of cic (FBgn0262582) under UAS control.	Vienna Drosophila Resource Center	VDRC:v103805
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of dia	Vienna Drosophila	
(FBgn0011202) under UAS control.	Resource Center	VDRC:v103914

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<i>D. melanogaster</i> . Expresses dsRNA for RNAi of SkpD;SkpC (FBgn0026174) under UAS control.	Vienna Drosophila Resource Center	VDRC:v109181
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of drk (FBgn0004638) under UAS control.	Vienna Drosophila Resource Center	VDRC:v105498
<i>D. melanogaster</i> . Expresses dsRNA for RNAi of botv (FBgn0027535) under UAS control.	Vienna Drosophila Resource Center	VDRC:v37186

1044

1045 Reagents

Name	Catalogue number	Usage
Hoechst 33321	H1399 (ThermoFisher)	1ug/ml final concentration

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1048 **References**

Ables, E.T., Drummond-Barbosa, D., 2017. Steroid Hormones and the Physiological Regulation
of Tissue-Resident Stem Cells: Lessons from the *Drosophila* Ovary. Current Stem Cell Reports
3, 9-18.

- Albert, I., Albert, R., 2004. Conserved network motifs allow protein-protein interaction prediction.Bioinformatics 20, 3346-3352.
- Albert, R., Barabási, A.-L., 2002. Statistical mechanics of complex networks. Reviews of Modern
 Physics 74, 47-97.
- 1056 Allbee, A.W., Rincon-Limas, D.E., Biteau, B., 2018. *Lmx1a* is required for the development of 1057 the ovarian stem cell niche in *Drosophila*. Development 145, dev163394.
- Azevedo, R.B.R., French, V., Partridge, L., 1996. Thermal evolution of egg size in *Drosophila melanogaster*. Evolution 50, 2338-2345.
- 1060 Barabási, A.L., Albert, R., 1999. Emergence of scaling in random networks. Science 286, 509-1061 512.
- Barabási, A.L., Oltvai, Z.N., 2004. Network biology: understanding the cell's functionalorganization. Nat. Rev. Genet. 5, 101-113.
- Barrett, K., Leptin, M., Settleman, J., 1997. The Rho GTPase and a putative RhoGEF mediate a
 signaling pathway for the cell shape changes in *Drosophila* gastrulation. Cell 91, 905-915.
- 1066 Berger, S.I., Posner, J.M., Ma'ayan, A., 2007. Genes2Networks: connecting lists of gene 1067 symbols using mammalian protein interactions databases. BMC Bioinformatics 8, 372.
- Bergland, A.O., Genissel, A., Nuzhdin, S.V., Tatar, M., 2008. Quantitative trait loci affecting
 phenotypic plasticity and the allometric relationship of ovariole number and thorax length in *Drosophila melanogaster*. Genetics 180, 567-582.
- Bolívar, J., Pearson, J., López-Onieva, L., González-Reyes, A., 2006. Genetic dissection of a
 stem cell niche: the case of the *Drosophila* ovary. Dev. Dyn. 235, 2969-2979.
- Bromberg, K.D., Ma'ayan, A., Neves, S.R., Iyengar, R., 2008. Design logic of a cannabinoid
 receptor signaling network that triggers neurite outgrowth. Science 320, 903-909.
- 1075 Capy, P., Pla, E., David, J.R., 1993. Phenotypic and genetic variability of morphometrical traits
 1076 in natural populations of *Drosophila melanogaster* and *D. simulans*. I. Geographic variations.
 1077 Genet. Sel. Evol. 25, 517-536.
- 1078 Capy, P., Pla, E., David, J.R., 1994. Phenotypic and genetic variability of morphometrical traits
 1079 in natural populations of *Drosophila melanogaster* and *D simulans*. II. Within-population
 1080 variability. Genetics Selection Evolution 26, 15-28.

Cayirlioglu, P., Ward, W.O., Silver Key, S.C., Duronio, R.J., 2003. Transcriptional repressor
functions of *Drosophila* E2F1 and E2F2 cooperate to inhibit genomic DNA synthesis in ovarian
follicle cells. Mol. Cell. Biol. 23, 2123-2134.

- 1084 Chao, A.T., Dierick, H.A., Addy, T.M., Bejsovec, A., 2003. Mutations in eukaryotic release 1085 factors 1 and 3 act as general nonsense suppressors in *Drosophila*. Genetics 165, 601-612.
- 1086 Chen, J., Godt, D., Gunsalus, K., Kiss, I., Goldberg, M., Laski, F.A., 2001. Cofilin/ADF is
 1087 required for cell motility during *Drosophila* ovary development and oogenesis. Nature Cell Biol.
 1088 3, 204-209.
- 1089 Chen, J.Y., Shen, C., Sivachenko, A.Y., 2006. Mining Alzheimer disease relevant proteins from 1090 integrated protein interactome data. Pacific Symposium on Biocomputing 11, 367-378.
- 1091 Christiansen, A.E., Ding, T., Bergmann, A., 2012. Ligand-independent activation of the
 1092 Hedgehog pathway displays non-cell autonomous proliferation during eye development in
 1093 Drosophila. Mech. Dev. 129, 98-108.
- 1094 Cohet, Y., David, J.R., 1978. Control of the Adult reproductive Potential by Preimaginal thermal 1095 Conditions. Oecologia 36, 295-306.
- 1096 Couderc, J.L., Godt, D., Zollman, S., Chen, J., Li, M., Tiong, S., Cramton, S.E., Sahut-Barnola,
- 1097 I., Laski, F.A., 2002. The *bric a brac* locus consists of two paralogous genes encoding BTB/POZ 1098 domain proteins and acts as a homeotic and morphogenetic regulator of imaginal development
- 1099 in *Drosophila*. Development 129, 2419-2433.
- 1100 David, J.R., Bocquet, C., 1975. Similarities and differences in latitudinal adaptation of two 1101 *Drosophila* sibling species. Nature 257, 588-590.
- Dobens, L., Jaeger, A., Peterson, J.S., Raftery, L.A., 2005. Bunched sets a boundary for Notch
 signaling to pattern anterior eggshell structures during *Drosophila* oogenesis. Dev. Biol. 287,
 425-437.
- Edwards, M.C., Wong, C., Elledge, S.J., 1998. Human cyclin K, a novel RNA polymerase IIassociated cyclin possessing both carboxy-terminal domain kinase and Cdk-activating kinase
 activity. Mol. Cell. Biol. 18, 4291-4300.
- Foster, L.J., de Hoog, C.L., Zhang, Y., Zhang, Y., Xie, X., Mootha, V.K., Mann, M., 2006. A
 mammalian organelle map by protein correlation profiling. Cell 125, 187-199.
- 1110 Gancz, D., Gilboa, L., 2013. Insulin and Target of rapamycin signaling orchestrate the 1111 development of ovarian niche-stem cell units in *Drosophila*. Development 140, 4145-4154.
- Gancz, D., Lengil, T., Gilboa, L., 2011. Coordinated regulation of niche and stem cell precursorsby hormonal signaling. PLoS Biol. 9, e1001202.
- Gilboa, L., 2015. Organizing stem cell units in the *Drosophila* ovary. Curr. Op. Genet. Dev. 32C,31-36.
- 1116 Giot, L., Bader, J.S., Brouwer, C., Chaudhuri, A., Kuang, B., Li, Y., Hao, Y.L., Ooi, C.E.,
- 1117 Godwin, B., Vitols, E., Vijayadamodar, G., Pochart, P., Machineni, H., Welsh, M., Kong, Y.,
- 1118 Zerhusen, B., Malcolm, R., Varrone, Z., Collis, A., Minto, M., Burgess, S., McDaniel, L.,
- 1119 Stimpson, E., Spriggs, F., Williams, J., Neurath, K., Ioime, N., Agee, M., Voss, E., Furtak, K.,
- 1120 Renzulli, R., Aanensen, N., Carrolla, S., Bickelhaupt, E., Lazovatsky, Y., DaSilva, A., Zhong, J.,
- 1121 Stanyon, C.A., Finley, R.L., Jr., White, K.P., Braverman, M., Jarvie, T., Gold, S., Leach, M.,

1122 Knight, J., Shimkets, R.A., McKenna, M.P., Chant, J., Rothberg, J.M., 2003. A protein 1123 interaction map of *Drosophila melanogaster*. Science 302, 1727-1736.

Godt, D., Couderc, J.L., Cramton, S.E., Laski, F.A., 1993. Pattern formation in the limbs of *Drosophila: bric à brac* is expressed in both a gradient and a wave-like pattern and is required
for specification and proper segmentation of the tarsus. Development 119, 799-812.

- 1127 Godt, D., Laski, F.A., 1995. Mechanisms of cell rearrangement and cell recruitment in
- 1128 *Drosophila* ovary morphogenesis and the requirement of *bric à brac*. Development 121, 173-1129 187.
- 1130 Gonzalez, G., Uribe, J.C., Tari, L., Brophy, C., Baral, C., 2007. Mining gene-disease
- relationships from biomedical literature: weighting protein-protein interactions and connectivity measures. Pacific Symposium on Biocomputing 12, 28-39.
- 1133 Gonzalez, M.W., Kann, M.G., 2012. Chapter 4: Protein interactions and disease. PLoS Comput 1134 Biol 8, e1002819.
- 1135 Gramates, L.S., Marygold, S.J., Santos, G.D., Urbano, J.M., Antonazzo, G., Matthews, B.B.,
- 1136 Rey, A.J., Tabone, C.J., Crosby, M.A., Emmert, D.B., Falls, K., Goodman, J.L., Hu, Y., Ponting,
- 1137 L., Schroeder, A.J., Strelets, V.B., Thurmond, J., Zhou, P., the FlyBase, C., 2016. FlyBase at 1138 25: looking to the future. Nucleic Acide Res. 45, D663, D671
- 1138 25: looking to the future. Nucleic Acids Res. 45, D663-D671.
- Green II, D.A., Extavour, C.G., 2012. Convergent Evolution of a Reproductive Trait Through
 Distinct Developmental Mechanisms in *Drosophila*. Dev. Biol. 372, 120-130.
- Green II, D.A., Sarikaya, D.P., Extavour, C.G., 2011. Counting in oogenesis. Cell Tissue Res.344, 207-212.
- Hahn, M.W., Kern, A.D., 2005. Comparative genomics of centrality and essentiality in three
 eukaryotic protein-interaction networks. Mol. Biol. Evol. 22, 803-806.
- Harris, T.J., Peifer, M., 2004. Adherens junction-dependent and -independent steps in the establishment of epithelial cell polarity in *Drosophila*. J. Cell Biol. 167, 135-147.
- Hartwell, L.H., Hopfield, J.J., Leibler, S., Murray, A.W., 1999. From molecular to modular cellbiology. Nature 402, C47-C52.
- Hilman, D., Gat, U., 2011. The evolutionary history of YAP and the hippo/YAP pathway. Mol.Biol. Evol. 28, 2403-2417.
- Hodin, J., Riddiford, L.M., 1998. The ecdysone receptor and ultraspiracle regulate the timing
 and progression of ovarian morphogenesis during *Drosophila* metamorphosis. Dev. Genes Evol.
 208, 304-317.
- Hodin, J., Riddiford, L.M., 2000a. Different mechanisms underlie phenotypic plasticity and
 interspecific variation for a reproductive character in Drosophilids (Insecta: Diptera). Evolution 5,
 1638-1653.

- Hodin, J., Riddiford, L.M., 2000b. Parallel alterations in the timing of ovarian ecdysone receptor
- and ultraspiracle expression characterize the independent evolution of larval reproduction in two
 species of gall midges (Diptera: Cecidomyiidae). Dev. Genes Evol. 210, 358-372.
- Hsu, H.-J., Drummond-Barbosa, D., 2009. Insulin levels control female germline stem cell
 maintenance via the niche in *Drosophila*. Proc. Natl. Acad. Sci. USA 106, 1117-1121.
- Huang, S.S., Fraenkel, E., 2009. Integrating proteomic, transcriptional, and interactome data reveals hidden components of signaling and regulatory networks. Science Signaling 2, ra40.
- 1164 Ideker, T., Sharan, R., 2008. Protein networks in disease. Genome Res. 18, 644-652.
- 1165 Ito, T., Bulger, M., Kobayashi, R., Kadonaga, J.T., 1996. *Drosophila* NAP-1 is a core histone 1166 chaperone that functions in ATP-facilitated assembly of regularly spaced nucleosomal arrays.
- 1167 Mol. Cell. Biol. 16, 3112-3124.
- 1168 Jalili, M., Salehzadeh-Yazdi, A., Gupta, S., Wolkenhauer, O., Yaghmaie, M., Resendis-Antonio,
- 1169 O., Alimoghaddam, K., 2016. Evolution of Centrality Measurements for the Detection of 1170 Essential Proteins in Biological Networks, Erentions in Physiology 7, 375
- 1170 Essential Proteins in Biological Networks. Frontiers in Physiology 7, 375.
- Jansen, R., Greenbaum, D., Gerstein, M., 2002. Relating whole-genome expression data withprotein-protein interactions. Genome Res. 12, 37-46.
- 1173 Jeong, H., Mason, S.P., Barabási, A.L., Oltvai, Z.N., 2001. Lethality and centrality in protein 1174 networks. Nature 411, 41-42.
- 1175 Jiang, K., Liu, Y., Zhang, J., Jia, J., 2018. An intracellular activation of Smoothened that is 1176 independent of Hedgehog stimulation in *Drosophila*. J. Cell Sci. 131, jcs211367.
- 1177 Kambysellis, M.P., Heed, W.B., 1971. Studies of Oogenesis in Natural Populations of
- 1178 Drosophilidae. I. Relation of ovarian development and ecological habitats of the Hawaiian 1179 species. Am. Nat. 941, 31-49.
- Kanehisa, M., Goto, S., Furumichi, M., Tanabe, M., Hirakawa, M., 2010. KEGG for
 representation and analysis of molecular networks involving diseases and drugs. Nucleic Acids
 Res. 38, D355-360.
- 1183 Kasza, K.E., Farrell, D.L., Zallen, J.A., 2014. Spatiotemporal control of epithelial remodeling by 1184 regulated myosin phosphorylation. Proc. Natl. Acad. Sci. USA 111, 11732-11737.
- 1185 Keller, R., 2006. Mechanisms of elongation in embryogenesis. Development 133, 2291-2302.
- King, R.C., 1970. Ovarian Development in *Drosophila melanogaster*. Academic Press, NewYork.
- King, R.C., Aggarwal, S.K., Aggarwal, U., 1968. The Development of the Female *Drosophila*Reproductive System. J. Morphol. 124, 143-166.
- 1190 Klepsatel, P., Galikova, M., De Maio, N., Huber, C.D., Schlotterer, C., Flatt, T., 2013a. Variation
- in thermal performance and reaction norms among populations of *Drosophila melanogaster*.Evolution 67, 3573-3587.

- 1193 Klepsatel, P., Galikova, M., De Maio, N., Ricci, S., Schlotterer, C., Flatt, T., 2013b. Reproductive 1194 and post-reproductive life history of wild-caught *Drosophila melanogaster* under laboratory
- 1195 conditions. J. Evol. Biol. 26, 1508-1520.
- Koschutzki, D., Schreiber, F., 2008. Centrality analysis methods for biological networks and
 their application to gene regulatory networks. Gene Regulation and Systems Biology 2, 193201.
- 1199 LaFever, L., Drummond-Barbosa, D., 2005. Direct control of germline stem cell division and cyst 1200 growth by neural insulin in *Drosophila*. Science 309, 1071-1073.
- LaFever, L., Feoktistov, A., Hsu, H.J., Drummond-Barbosa, D., 2010. Specific roles of Target of
 rapamycin in the control of stem cells and their progeny in the *Drosophila* ovary. Development
 137, 2117-2126.
- Lai, C.M., Lin, K.Y., Kao, S.H., Chen, Y.N., Huang, F., Hsu, H.J., 2017. Hedgehog signaling
 establishes precursors for germline stem cell niches by regulating cell adhesion. J. Cell Biol.
 216, 1439-1453.
- 1207 Lee, R.T., Zhao, Z., Ingham, P.W., 2016. Hedgehog signalling. Development 143, 367-372.
- Lewellyn, L., Cetera, M., Horne-Badovinac, S., 2013. Misshapen decreases integrin levels to promote epithelial motility and planar polarity in *Drosophila*. J. Cell Biol. 200, 721-729.
- Li, X., Wu, M., Kwoh, C.K., Ng, S.K., 2010. Computational approaches for detecting protein complexes from protein interaction networks: a survey. BMC Genomics 11 Suppl 1, S3.
- Llamazares, S., Moreira, A., Tavares, A., Girdham, C., Spruce, B.A., Gonzalez, C., Karess,
 R.E., Glover, D.M., Sunkel, C.E., 1991. *polo* encodes a protein kinase homolog required for
 mitosis in *Drosophila*. Genes Dev. 5, 2153-2165.
- 1215 Lobell, A.S., Kaspari, R.R., Serrano Negron, Y.L., Harbison, S.T., 2017. The Genetic
- 1216 Architecture of Ovariole Number in *Drosophila melanogaster*. Genes with Major, Quantitative, 1217 and Pleiotropic Effects. G3 7, 2391-2403.
- 1218 Ma'ayan, A., 2011. Introduction to network analysis in systems biology. Sci Signal 4, tr5.
- 1219 Markow, T.A., Beall, S., Matzkin, L.M., 2009. Egg size, embryonic development time and 1220 ovoviviparity in *Drosophila* species. J. Evol. Biol. 22, 430-434.
- Matthews, L.R., Vaglio, P., Reboul, J., Ge, H., Davis, B.P., Garrels, J., Vincent, S., Vidal, M.,
 2001. Identification of potential interaction networks using sequence-based searches for
 conserved protein-protein interactions or "interologs". Genome Res. 11, 2120-2126.
- 1224 Mbodj, A., Junion, G., Brun, C., Furlong, E.E., Thieffry, D., 2013. Logical modelling of 1225 *Drosophila* signalling pathways. Molecular BioSystems 9, 2248-2258.
- Mizuno, T., Amano, M., Kaibuchi, K., Nishida, Y., 1999. Identification and characterization of *Drosophila* homolog of Rho-kinase. Gene 238, 437-444.

- Orgogozo, V., Broman, K.W., Stern, D.L., 2006. High-resolution quantitative trait locus mapping
 reveals sign epistasis controlling ovariole number between two *Drosophila* species. Genetics
 173, 197-205.
- R'Kha, S., Capy, P., David, J.R., 1991. Host-plant specialization in the *Drosophila melanogaster*species complex: A physiological, behavioral, and genetic analysis. Proc. Natl. Acad. Sci. USA
 88, 1835-1839.
- 1234 R´kha, S., Moreteau, B., Coyne, J.A., David, J.R., 1997. Evolution of a lesser fitness trait: egg 1235 production in the specialist *Drosophila sechellia*. Genetical research 69, 17-23.
- Ravasz, E., Somera, A.L., Mongru, D.A., Oltvai, Z.N., Barabási, A.L., 2002. Hierarchical
 organization of modularity in metabolic networks. Science 297, 1551-1555.
- Ridley, A.J., 2006. Rho GTPases and actin dynamics in membrane protrusions and vesicletrafficking. Trends Cell Biol. 16, 522-529.
- 1240 Sahut-Barnola, I., Dastugue, B., Couderc, J.-L., 1996. Terminal filament cell organization in the
- 1241 larval ovary of *Drosophila melanogaster*: ultrastructural observations and pattern of divisions.
- 1242 Roux's Archives of Developmental Biology 205, 356-363.
- Sahut-Barnola, I., Godt, D., Laski, F.A., Couderc, J.-L., 1995. *Drosophila* Ovary Morphogenesis:
 Analysis of Terminal Filament Formation and Identification of a Gene Required for This Process.
 Dev. Biol. 170, 127-135.
- Sarikaya, D.P., Belay, A.A., Ahuja, A., Green II, D.A., Dorta, A., Extavour, C.G., 2012. The roles
 of cell size and cell number in determining ovariole number in *Drosophila*. Dev. Biol. 363, 279289
- 1249 Sarikaya, D.P., Church, S.H., Lagomarsino, L.P., Magnacca, K.M., Montgomery, S.L., Price,
- 1250 D.P., Kaneshiro, K.Y., Extavour, C.G., 2019. Reproductive capacity evolves in response to 1251 ecology through common developmental mechanisms in Hawai'ian *Drosophila*. Curr. Biol. 29,
- 1252 1877-1884.
- 1253 Sarikaya, D.P., Extavour, C.G., 2015. The Hippo pathway regulates homeostatic growth of stem 1254 cell niche precursors in the *Drosophila* ovary. PLoS Genetics 11, e1004962.
- 1255 Sebe-Pedros, A., Zheng, Y., Ruiz-Trillo, I., Pan, D., 2012. Premetazoan origin of the hippo 1256 signaling pathway. Cell Reports 1, 13-20.
- 1257 Spellman, P.T., Sherlock, G., Zhang, M.Q., Iyer, V.R., Anders, K., Eisen, M.B., Brown, P.O.,
- Botstein, D., Futcher, B., 1998. Comprehensive identification of cell cycle-regulated genes of the yeast *Saccharomyces cerevisiae* by microarray hybridization. Mol. Biol. Cell 9, 3273-3297.
- Srinivasan, B.S., Shah, N.H., Flannick, J.A., Abeliuk, E., Novak, A.F., Batzoglou, S., 2007.
 Current progress in network research: toward reference networks for key model organisms.
- 1262 Brief. Bioinform. 8, 318-332.
- 1263 von Mering, C., Krause, R., Snel, B., Cornell, M., Oliver, S.G., Fields, S., Bork, P., 2002.
- 1264 Comparative assessment of large-scale data sets of protein-protein interactions. Nature 417, 1265 399-403.

- Wachi, S., Yoneda, K., Wu, R., 2005. Interactome-transcriptome analysis reveals the high
 centrality of genes differentially expressed in lung cancer tissues. Bioinformatics 21, 4205-4208.
- Wang, R.S., Hall, K.T., Giulianini, F., Passow, D., Kaptchuk, T.J., Loscalzo, J., 2017. Network
 analysis of the genomic basis of the placebo effect. JCI Insight 2, 93911.
- Wang, R.S., Loscalzo, J., 2018. Network-Based Disease Module Discovery by a Novel Seed
 Connector Algorithm with Pathobiological Implications. J. Mol. Biol. 430, 2939-2950.
- Watts, D.J., Strogatz, S.H., 1998. Collective dynamics of 'small-world' networks. Nature 393,440-442.
- Wayne, M.L., Hackett, J.B., Dilda, C.L., Nuzhdin, S.V., Pasyukova, E.G., Mackay, T.F., 2001.
 Quantitative trait locus mapping of fitness-related traits in *Drosophila melanogaster*. Genetical
- 1276 research 77, 107-116.
- Wayne, M.L., Hackett, J.B., Mackay, T.F.C., 1997. Quantitative Genetics of Ovariole Number in
 Drosophila melanogaster. I. Segregating Variation and Fitness. Evolution 4, 1156-1163.
- Wayne, M.L., McIntyre, L.M., 2002. Combining mapping and arraying: An approach to candidategene identification. Proc. Natl. Acad. Sci. USA 99, 14903-14906.
- Yang, X., Xu, T., 2011. Molecular mechanism of size control in development and humandiseases. Cell Research 21, 715-729.
- 1283 Yook, S.H., Oltvai, Z.N., Barabási, A.L., 2004. Functional and topological characterization of 1284 protein interaction networks. Proteomics 4, 928-942.
- Yu, H., Paccanaro, A., Trifonov, V., Gerstein, M., 2006. Predicting interactions in protein
 networks by completing defective cliques. Bioinformatics 22, 823-829.
- Zhang, Y., Lin, H., Yang, Z., Wang, J., Liu, Y., 2017. An uncertain model-based approach for
 identifying dynamic protein complexes in uncertain protein-protein interaction networks. BMC
 Genomics 18, 743.

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