Independent evolution of lateral inhibition mechanisms in different lineages of land plants: MpFEW RHIZOIDS1 miRNA-mediated lateral inhibition controls rhizoid cell patterning in *Marchantia polymorpha*

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ABSTRACT

Lateral inhibition patterns differentiated cells during development in bacteria, metazoans and land plants. Tip-growing rhizoid cells develop among flat epidermal cells in the epidermis of the early diverging land plant *Marchantia polymorpha*. We show that the majority of rhizoid cells develop individually but some develop in linear, one-dimensional clusters of between two and seven rhizoid cells in wild type plants. The distribution of rhizoid cells can be accounted for within a simple model of lateral inhibition. The model also predicted that, in the absence of lateral inhibition, rhizoid cell clusters would be two-dimensional with larger clusters than those formed with lateral inhibition. Rhizoid differentiation in *Marchantia polymorpha* is positively regulated by the ROOT HAIR DEFECTIVE SIX-LIKE1 (MpRSL1) basic Helix Loop Helix (bHLH) transcription factor which is directly repressed by the FEW RHIZOIDS1 (MpFRH1) miRNA. To test if MpFRH1 miRNA acts during lateral inhibition we generated loss-of-function mutants that did not produce the MpFRH1 miRNA. Two-dimensional clusters of rhizoids develop in Mp*frh1*$^{loss\text{-}of\text{-}function\ (lof)}$ mutants as predicted by the model for plants that lack lateral inhibition. Furthermore, clusters of up to nine rhizoid cells developed in the Mp*frh1*^{lof} mutants compared to a maximum number of seven observed in wild type. The higher steady state levels of MpRSL1 mRNA in Mp*frh1*^{lof} mutants indicate that MpFRH1-mediated lateral inhibition involves the repression of Mp*RSL1* activity. Together the modelling and genetic data indicate that the pattern of cell differentiation in the *M. polymorpha* epidermis is consistent with a lateral inhibition process in which MpFRH1 miRNA represses MpRSL1. This discovery suggests that novel mechanisms of lateral inhibition may operate in different lineages of land plants, unlike metazoans where the conserved Delta-Notch signaling system controls lateral inhibition in diverse metazoan lineages.

35 **INTRODUCTION**

36

37 Spatial arrays of differentiated cells form during the development of multicellular
38 organisms. The patterning of different cell types during animal development involves lateral
39 inhibition, a process in which an individual cell instructs adjacent cells to acquire an identity
40 that is different from the fate of the instructing cell. In metazoans, lateral inhibition involves
41 cell-to-cell signalling carried out by the Delta-Notch ligand-receptor pair (reviewed in
42 Artavanis-Tsakonas et al., 1999; Bray, 2006; Perrimon et al., 2012). Neither Delta nor Notch
43 proteins are present in plants and other mechanisms of lateral inhibition operate in the
44 angiosperm *Arabidopsis thaliana* (The Arabidopsis Genome Initiative, 2000; Wigge and
45 Weigel, 2001; Bowman et al., 2017). Computational modelling suggests that the production
46 of EPIDERMAL PATTERNING FACTOR2 (EPF2) peptide by meristemoids inhibits meristemoid
47 development in adjacent cells during guard cell development in leaves (Horst et al., 2015;
48 Lee et al., 2015; Simmons and Bergmann, 2016). The cell-to-cell movement of Myb
49 transcriptional repressor proteins pattern trichomes (leaf hair cells) among pavement
50 epidermal cells in the *A. thaliana* shoot epidermis (Schellmann et al., 2002). A similar
51 mechanism is required for the maintenance of patterning in the root epidermis (Lee and
52 Schiefelbein, 2002; Savage et al., 2008). The demonstration that there are at least two
53 mechanisms of lateral inhibition in *A. thaliana* alone suggests that, unlike metazoans,
54 diverse mechanisms of lateral inhibition likely function among land plants.

55

56 Rhizoid cells develop from fields of equivalent cells in the epidermis of the liverwort
57 *Marchantia polymorpha,* an early diverging land plant. Flat epidermal cells occupy the
58 space between rhizoid cells in wild type plants. The RSL1 basic-Helix-Loop-Helix
59 transcription factor is necessary and sufficient for rhizoid cell development (Menand et al.,
60 2007; Catarino et al., 2016; Proust et al., 2016). Rhizoid cells do not develop in plants
61 harbouring loss-of-function Mp*rsl1* mutations and flat epidermal cells develop in their place
62 (Proust et al., 2016). Conversely plants harbouring Mp*RSL1* gain-of-function mutations
63 develop supernumerary ectopic rhizoids in place of flat epidermal cells (Proust et al., 2016).
64 MpRSL1 promotes the expression of Mp*FRH1* which encodes a 21-nucleotide microRNA
65 (Honkanen et al., 2018). Evidence that MpFRH1 represses rhizoid cell development by
66 negatively regulating MpRSL1 activity include (i) FRH1 miRNA directly targets RSL1 mRNA for

67 DICERLIKE1-mediated cleavage and (ii) the rhizoidless phenotype of Mp*FRH1* overexpression
68 is suppressed by co-expressing an MpFRH1 miRNA-resistant version of MpRSL1 (Honkanen
69 et al., 2018). Therefore, MpRSL1 and MpFRH1 act together in a network with feedback
70 where the transcription factor is an activator of rhizoid cell development and the miRNA is a
71 repressor of rhizoid cell development.
72
73 Since MpRSL1 and MpFRH1 constitute an activator-repressor network with negative
74 feedback, we hypothesized that the MpFRH1 miRNA could be involved in lateral inhibition
75 during the patterning of rhizoid cells in the epidermis *M. polymorpha* (Honkanen et al.,
76 2018). To test this hypothesis, we first modelled the effect of lateral inhibition on the
77 pattern of rhizoid cell spacing in wild type conditions. This revealed that the spatial
78 distribution of rhizoid cells in wild type was consistent with lateral inhibition being involved
79 in patterning. We then ran the model in the absence of lateral inhibition to predict the
80 rhizoid cell distribution that would be observed in a plant in which lateral inhibition did not
81 operate. To test if MpFRH1 miRNA was involved in lateral inhibition we characterised
82 rhizoid cell distribution in Mp*frh1*$^{loss\text{-}of\text{-}function\ (lof)}$ mutants. The distribution of rhizoid cells in
83 Mp*frh1* mutants was consistent with the predicted distribution from the model without
84 lateral inhibition. These data indicate that MpFRH1 miRNA acts in the lateral inhibition of
85 rhizoid cell development during epidermal development in *M. polymorpha.*

RESULTS

Linear clusters of rhizoid cells develop in wild type *Marchantia polymorpha* gemma epidermis

The *M. polymorpha* gemma epidermis comprises flat epidermal cells that surround rhizoid cells that produce a tip growing projection that is involved in rooting function. To determine if lateral inhibition is involved in the patterning of rhizoid cells, we characterised their spatial arrangement on the dorsal surface of wild type gemmae (Tak-1 and Tak-2). Rhizoid cells develop individually and in clusters in wild type gemmae (Figure 1; Table 1). The majority (~70%) of rhizoid cells develop individually, surrounded by flat epidermal cells in wild type. In the Tak-2 background, approximately 30% of rhizoids develop in clusters of between 2 and 6 rhizoid cells (Table 1); 18.8% 7.3%, 2.6%, 1.0%and 1.0% of rhizoids formed in clusters of 2, 3, 4, 5 and 6 rhizoid cells, respectively. Most rhizoid cells in clusters are arranged in a one-dimensional, linear array, like beads on a string. Rhizoid cells at either end contact one neighbouring rhizoid cell and internal rhizoid cells always contact two rhizoid cells. Consequently, in a linear cluster of three rhizoid cells, there are two rhizoid cells on either end of the cluster with a single rhiziod cell between each of these end initials. Therefore, such a cluster of three develops an average of 1.33 rhizoid cell neighbours ((1 + 2 + 1)/3 = 1.33). The rhizoid cells in 24 of the 25 3-rhizoid cell clusters were one-dimensional. The rhizoid cells in a single 3-rhizoid cell cluster contacted each other and had an average of 2 rhizoid cell neighbours (two-dimensional). The average number of rhizoid cell neighbours in 3-rhizoid cell clusters was therefore 1.36, (SD = 0.13; n= 25). The mean number of rhizoid cell neighbours increases as cluster size increases. For example in a one-dimensional cluster of 4-rhizoid cells there is an average of 1.5 rhizoid cell neighbours ((1+2+2+1)/4 = 1.5). Rhizoid cells in 12 of 13 four-cell cluster were arranged in one-dimensional clusters while rhizoid cells were arranged in a two-dimensional cluster in a single case. Therefore the mean rhizoid cell neighbour number in four-cell cluster is 1.57 (SD = 0.27; n= 13). We observed four 5-rhizoid cell clusters and three 6-cell cluster, and rhizoid cells were arranged in a one-dimension in all seven. Therefore the average rhizoid cell neighbour number was = 1.6 in 5-cell cluster (SD = 0; n= 4 clusters) and 1.66 in 6-cell cluster (SD = 0; n=3) (Figure 1). There was a single, two-dimensional, 7-cell cluster in the Tak-1 background (Figure S1). Hence, for such one-dimensional rhizoid arrays with *n* cells, the average rhizoid cell neighbour number = 2-2/n. The SD is close to zero indicating that this pattern is consistent

118 in each cluster class. In total, 93.5 % of rhizoid clusters in wiltype are arranged in a one-
119 dimension and only 6.5 % are arranged in a two dimensions (n= 46, Figure 4c). Based on
120 those findings, the maximum number of rhizoid neighbour cells is generally two and clusters
121 comprise one-dimensional linear arrays of rhizoid cells. These data show that wild type
122 rhizoid cells form either individually or in linear clusters where each rhizoid cell has either
123 one or two neighbours and not more than two. This is consistent with the hypothesis that
124 lateral inhibition operates in the patterning of rhizoid cells in the epidermis.
125
126 **Modelling predicts that lateral inhibition in involved in the patterning of rhizoid cells in**
127 **wild type gemmae**
128 A model for the development of rhizoid cells in the gemma epidermis was developed. We
129 initiated a lattice of cells with seeds for rhizoid cells at similar density to measured
130 experimentally. The model had two rules. Rule (1): For each rhizoid, at each simulation step
131 a probability P determined whether the cluster stops expanding (and stays fixed at that cell
132 number for remainder of simulation) or (1-P) that a new neighbouring cell becomes a
133 rhizoid cell. P was estimated from the data from wild type clusters. To incorporate new
134 rhizoid cells in wild type conditions we applied the Rule (2): the new rhizoid cell had to be a
135 neighbour to precisely one existing rhizoid cell (no fewer and no more) (Figure 2a). No
136 other rules or parameters were incorporated into the model. We analysed the number and
137 distribution of clusters produced by the model. With this simple rule, the model replicated
138 the observed distributions of rhizoid cell cluster sizes and the rhizoid cell neighbour
139 frequency as observed experimentally (Figure 2a, b). All rhizoid cell clusters that developed
140 were linear – each rhizoid cell in the cluster developed either one rhizoid cell neighbour (if
141 located at the end of a cluster) or two rhizoid cell neighbours (if located away from cluster
142 ends) and the SD for mean rhizoid cell neighbour number was 0 in each size class. Thus,
143 simple lateral inhibition is consistent with our observed patterns of rhizoid cell development
144 and also with the invariant neighbour number.
145
146 We next considered the scenario whereby rule 2 was relaxed so that a new cell can
147 develop next to one *or more* existing rhizoid cells; *i.e.* we remove the lateral inhibition
148 requirement. The resulting pattern did not resemble the pattern of rhizoid cells in wild type
149 plants (Figure 2c); clusters were two-dimensional instead of one-dimensional. In the

150 absence of lateral inhibition, larger rhizoid cell clusters developed than in wild type due to
151 merging of neighbouring clusters. Furthermore, the variability in the number of rhizoid cell
152 neighbours, measured by SD, for rhizoid cell cluster larger than 3 was always greater than 0,
153 the value predicted for the simulation with lateral inhibition. Other more complex models
154 can be considered. However, our simple Boolean-like lateral inhibition model is consistent
155 with the hypothesis that lateral inhibition functions during the patterning of rhizoids; a
156 rhizoid cell can develop next to flat epidermal cells or a single rhizoid but never more than
157 one rhizoid cell.
158
159 With our given *P*, clusters of 6 or more cells are not seen in the wild type simulations
160 where lateral inhibition is active, which is consistent with what is observed in wild type
161 plants. However, in our simulations with the relaxed Rule 2, we noticed that occasionally
162 clusters can merge; in the absence of lateral inhibition, two rhizoid cell clusters are not
163 inhibited from adding a joining cell. In this case, larger clusters form, comprising up to 12
164 cells in the model, but these are rare (observed once in over 1000 simulations). The model
165 predicts both an increase in the rhizoid cell neighbour number and an increase in the
166 standard deviation in cluster neighbours. Furthermore, these clusters have rounder shapes
167 than the elongated, linear wild type structures (Figure 2e). These arrangements are only
168 rarely observed in the models in which lateral inhibition operates in wild type plants. These
169 modelling results indicate that if lateral inhibition controls the spacing of rhizoid cells in *M.*
170 *polymorpha*, loss of this mechanism will cause development of two-dimensional rhizoid cell
171 clusters and rare large clusters. We emphasise that these quantitative model predictions
172 require no extra parameters; they are simply dependent on the presence/absence of lateral
173 inhibition.
174
175 **Two dimensional clusters developed in Mp*frh1* mutants**
176 If MpFRH1 miRNA mediates lateral inhibition during rhizoid cell development, the model
177 predicted that two-dimensional rhizoid clusters would develop in Mp*frh1*^{lof} mutants while
178 one-dimensional rhizoid clusters would develop in wild type. To determine if MpFRH1
179 miRNA is involved in lateral inhibition we generated mutants in which the entire Mp*FRH1*
180 miRNA coding sequence was deleted. We used a pair of single guide RNAs (sgRNAs) 569 bp
181 apart to delete the MpFRH1 gene using CRISPR/Cas9 technology (Sugano et al., 2014).

182 sgRNA1 was 20 bp long and was complementary to a sequence between 404bp – 384 bp 5'
183 of the Mp*FRH1* miRNA sequence (sgRNA1) and sgRNA2 was 20 bp long and complementary
184 to a sequence between 164 bp – 184 bp 3' of the Mp*FRH1* miRNA sequence (Figure 3a). Five
185 independent mutant lines were identified in which the entire Mp*FRH1* miRNA sequence was
186 deleted. There were deletions of 267 bp, 587 bp, 589 bp and 573 bp in the Mp*FRH1* gene of
187 Mp*frh1*-7, Mp*frh1*-27, Mp*frh1*-34 and Mp*frh1*-36 mutants respectively. In Mp*frh1*-11, a 780
188 bp deletion removed the Mp*FRH1* sequence and a 202 bp sequence inserted 5' of the
189 Mp*FRH1* sequence. Mp*FRH1* miRNA was not detected (Figure 3b) in any of the five deletion
190 mutants consistent with the hypothesis that each harboured complete loss-of-function
191 Mp*frh1* mutations. Consequently, the mutants were designated Mp*frh1*$^{lof-7,}$ Mp*frh1*$^{lof-11}$,
192 Mp*frh1*$^{lof-27}$, Mp*frh1*$^{lof-34}$ and Mp*frh1*$^{lof-36}$ (where lof is loss-of-function and distinguishes
193 them from Mp*FRH1*^{GOF} lines which harbour gain-of-function mutations).
194
195 To determine if the pattern of rhizoids in Mp*frh1*^{lof} mutants was as predicted by the
196 model in the absence of lateral inhibition, the number of rhizoid cell neighbours was
197 measured in rhizoid clusters in each of the mutants. The model predicted that the number
198 of rhiziod cell neighbours in clusters would be greater than in wild type. As predicted by the
199 model, the mean rhizoid cell neighbour number is greater in the mutants than in wild type
200 rhizoid cell clusters for each cluster size class (Figure 4b, Figure 5). In all five Mp*frh1*^{lof}
201 mutants, the number of neighbours in clusters of three rhizoid cells was greater than a 1.36
202 (the average number of neighbours in a cluster of three rhizoid cells in wild type).
203 Furthemore while the SD was close to 0 in wild type indicating that there was little variation
204 in the mean number of neighbour cells per cluster, the SD was always greater than 0 in the
205 mutants. For example, there were on average 1.4 (SD = 0.21) rhizoid cell neighbours in
206 clusters of three rhizoid cells in Mp*frh1*$^{lof-7}$, 1.45 (SD = 0.26) rhizoid cell neighbours in
207 Mp*frh1*$^{lof-27}$, 1.48 (SD = 0.28) rhizoid cell neighbours in Mp*frh1*$^{lof-11}$, 1.56 (SD=0.34) in
208 Mp*frh1*$^{lof-34}$ and 1.6 (SD=0.37) in Mp*frh1*$^{lof-36}$ (Figure 5a, e). The same trend held for clusters
209 of four rhizoid cells where the number of rhizoid cell neighbours was 1.57 (SD = 0.27) in wild
210 type (1.63 for Tak-1 (SD = 0.35), 1.5 for Tak-2 (SD = 0)). Clusters containing four rhizoid cells
211 developed an average 1.73 (SD = 0.37) rhizoid cell neighbours in Mp*frh1*$^{lof-7}$, 1.61 (SD
212 =0.33) in Mp*frh1*$^{lof-11}$, 1.67 (SD = 0.29) in Mp*frh1*$^{lof-34}$ mutants (Figure 5b, e,). Clusters of five
213 rhizoid cells in Mp*frh1*$^{lof-27}$ developed 2.2 (SD = 0.28) rhizoid cell neighbours compared to

214 1.6 (SD = 0) in wild type (Figure 5c, e). Consequently rhizoid cell clusters are two
215 dimensionsal in Mp*frh1*^{lof} mutants rather than one-dimensional as in in wild type.
216 Remarkably, our simple model predicted similar shifts in both the mean and standard
217 deviation under loss of the purported lateral inhibtion (Figure 2).
218
219 To confirm that the increased number of rhizoid cell neigbours resulted in an
220 increased frequency of two-dimensional clusters, as predicted by the model, we measured
221 the frequencecy of one-dimensional and two-dimensional rhizoid clusters in Mp*frh1*^{lof}
222 mutants. 92-96% of rhizoid cluster are one-dimensional and only 4-8% of rhizoid cluster are
223 two-dimensional in wild type. By contrast, 32% of rhizoid cluster are two-dimensional in
224 Mp*frh1*^{lof} mutants. This includes 30% of rhizoid clusters in Mp*frh1*$^{lof-}$ 7 (14 of 46), 25% in
225 Mp*frh1*$^{lof-11}$ (7 of 21), 37% in Mp*frh1*$^{lof-27}$ (9 of 24), 33% in Mp*frh1*$^{lof-34}$ (3 of 9) and 50% in
226 Mp*frh1*$^{lof-36}$ (4 of 8 cluster)(Figure 4c). The higher frequency of two-dimensional clusters in
227 Mp*frh1*^{lof} than in wildtype is consistent with the hypothesis that Mp*FRH1* activity is required
228 for lateral inhibition during rhizoid development. The phenotypes of larger rhizoid cell
229 neighbour number and higher frequency of two-dimensional rhizoid cell clusters in
230 Mp*frh1*^{lof} mutants than in wild type are consistent with the hypothesis that Mp*FRH1* is
231 active in lateral inhibition during rhizoid cell development.
232
233 If MpFRH1 miRNA acts as an inhibitor during lateral inhibition, the model predicted
234 that rhizoid clusters with up to 9 rhizoid cells should develop infrequently in Mp*frh1*^{lof}
235 mutants. Such clusters form from the merger of adjacent clusters; a process which is
236 inhibited in wild type conditions. The maximum number of rhizoid cells in clusters was 7 in
237 wild type (Table 1, Figure 2c). Rhizoid cell clusters with more than 7 cells were found in
238 Mp*frh1*^{lof} mutants (Table 1, Figure 4b). A single 8-rhizoid cell cluster and three 9-rhizoid cell
239 clusters were observed in Mp*frh1*$^{lof-7}$ and a single 8-rhiziod cell cluster was found in
240 Mp*frh1*$^{lof-27.}$ The observation that rare clusters containing up to 9 rhizoid cells developed in
241 Mp*frh1*^{lof} mutants is consistent with the hypothesis that MpFRH1 acts as an inhibitor in
242 lateral inhibition during epidermal develoment.
243

244 **MpFRH1-mediated lateral inhibition acts by repressing Mp*RSL1* activity**

245 Since MpFRH1 miRNA targets the MpRSL1 mRNA, we hypothesized that the loss of MpFRH1
246 function would result in an increase in the steady state levels of Mp*RSL1* mRNA. mRNA was
247 isolated from thallus that developed from whole gemmae grown for 7 days. Mp*RSL1* mRNA
248 abundance was normalised to the Mp*APT1* housekeeping gene (Saint-Marcoux et al., 2015).
249 MpRSL1 mRNA is more abundant in Mp*frh1*^{lof} mutants than in wild type. Steady state levels
250 of MpRSL1 mRNA were 2-2.5 times more abundant in Mp*frh1*$^{lof-36}$ and Mp*frh1*$^{lof-34}$ than in
251 wild type (Figure 3c). Steady state levels of MpRSL1 mRNA were approximately 1.5 times
252 higher in Mp*frh1*$^{lof-7}$, Mp*frh1*$^{lof-11}$ and Mp*frh1*$^{lof-27}$ mutants than in wild type. In an
253 independent experiment with fewer replicates, the steady state levels of MpRSL1 mRNA
254 were 3.7, 2.7, 4.23-, 3.77-, and 1.6- fold-higher in Mp*frh1*$^{lof-7}$, Mp*frh1*$^{lof-11}$, Mp*frh1*$^{lof-27}$
255 Mp*frh1*$^{lof-34}$ and Mp*frh1*$^{lof-36}$, respectively, than in wild type (see Supplementary Figure2).
256 These higher steady state levels of MpRSL1 mRNA in Mp*frh1*^{LOF} mutants than wild type is
257 consistent with the hypothesis that MpFRH1 targets MpRSL1 mRNA during lateral inhibition.
258

259 If MpFRH1-mediated lateral inhibition operates by repressing MpRSL1,
260 overexpression of MpRSL1 should overcome the inhibitory effect of MpFRH1. If true, ectopic
261 overexpression of MpRSL1 would increase the mean number of rhizod cell neighbours in
262 clusters and that larger clusters of rhizoid cells would develop than in wild type. That is, the
263 rhizoid cluster phenotypes of the Mp*RSL1*^{GOF} and Mp*frh1*^{lof} mutants would be similar. 3-
264 rhizoid cell clusters in Mp*RSL1*^{GOF} mutants developed an average of 1.59 (SD = 0.34, n = 13)
265 neighbouring rhizoid cells, compared to 1.36 neighbours in wild type (SD = 0.13) (Figure 5a).
266 4-rhizoid cell clusters in Mp*RSL1*^{GOF} mutants developed 1.7 (SD = 0.45, n = 5) rhizoid cell
267 neighbours compared to 1.58 in wild type (SD =0.28) (Figure 5b). 5-rhizoid cell clusters in
268 Mp*RSL1*^{GOF} mutants developed 1.73 (SD = 0.23) neighbours, compared to 1.6 in wild type
269 (SD = 0) (Figure 4c). The larger number and variability of rhizoid cell neighbours in
270 Mp*RSL1*^{GOF} mutants than in wild type indicate that ectopic overexpression of Mp*RSL1*
271 causes similar phenotypic defecst in rhizoid clusters as in Mp*frh1*^{lof} mutants. These data are
272 consistent with the hypothesis that MpFRH1 miRNA represses Mp*RSL1* activity during lateral
273 inhibition as rhizoids develop.
274

275 The model indicates that the maximum cluster size would be greater than 6 in the
276 absence of lateral inhibition. If lateral inhibition were inactive in Mp*RSL1*^{GOF} mutants, the
277 model predicts that there would be more than 6 rhizoid cells in some clusters that develop
278 in plants that ectopically overexress Mp*RSL1*. We counted the number of rhizoid cells in
279 clusters in Mp*RSL1*^{GOF} mutants. While the maximum number of rhizoid cells in wild type
280 clusters was 7, there were up to 21 rhizoid cells in a single cluster in Mp*RSL1*^{GOF} plants
281 (Figure 1b, Supplement Figure 3). We observed three clusters greater than ten cells,
282 included a single 10-cell cluster, a 12-cell cluster and a 21-cell cluster (summarzised to >10
283 cell cluster in Table1). This confirmes that repression of Mp*RSL1* occurs during lateral
284 inhibition. Taken together, the data reported here demonstrates that MpFRH1 is a
285 repressor that acts during lateral inhibition and MpFRH1 miRNA-mediated lateral inhibition
286 acts by repressing Mp*RSL1* during the development of rhizoid cells in the *M. polymorpha*
287 epidermis.

288 **DISCUSSION**

289

290 We report that MpFRH1 miRNA represses rhizoid cell development through lateral
291 inhibition in the *M. polymorpha* epidermis. Rhizoid cells develop individually or as one-
292 dimensional clusters of between 2 and 7 rhizoid cells in the wild type epidermis. There is a
293 high frequency of single rhizoid cells and progressively fewer clusters of 2, 3, 4, 5, 6 and-7
294 rhizoid cells. The spatial arrangement and distribution of rhizoid cells can be accounted for
295 by a simple model that includes lateral inhibition. This model allowed us to predict the
296 phenotype of rhizoid cell patterns that would develop in a mutant in which lateral inhibition
297 was absent. It predicted that rhizoid cells would develop in two-dimensional clusters with
298 larger clusters possible compared to wild type where clusters would be one-dimensional
299 with a maximum of six cells in plants that lacked lateral inhibition. We showed that the
300 rhizoid cell phenotypes that develop in plants that lack the MpFRH1 miRNA are those
301 predicted by the model for an epidermis developing without lateral inhibition; clusters were
302 two-dimensional and there were nine cells in the largest rhizoid cell cluster. These
303 combined modelling and genetic data indicate that lateral inhibition mediated by the
304 MpFRH1 miRNA is required for the patterning of rhizoid cells in *M. polymorpha*.

305

306 We propose that the development of rhizoid cells involves two distinct phases. The
307 first phase involves the specification of individual rhizoid cells from a field of equivalent
308 cells. The probability of a rhizoid cell being specified could be determined by stochastic
309 variation in the expression levels of MpRSL1; higher levels would favour rhizoid cell
310 development and relatively lower levels would favour flat epidermal cell development.
311 MpRSL1 then induces expression of the MpFRH1 miRNA in the developing rhizoid cell.
312 MpFRH1 activity represses rhizoid cell development in adjacent cells through lateral
313 inhibition and these cells develop as flat epidermal cells. Repeated rounds of rhizoid
314 initiated during a competence period results in the development of rare adjacent rhizoid
315 cells which form linear clusters, and the shape of these clusters is determined by lateral
316 inhibition.

317

318 Our modelling and experimental data suggest that being adjacent to two rhizoid cells
319 represses rhizoid cell development. By contrast, being adjacent to a single rhizoid cell is not

320 sufficient to repress rhizoid development. This suggests that there is a threshold of
321 repressor activity. Being adjacent to one rhizoid cell exposes an adjacent cell to sub-
322 threshold levels of repressor while being adjacent to two rhizoid cells exposes a cell to
323 repressor levels above this threshold and represses differentiation. A repressive signal from
324 a rhizoid cell to adjacent cells represses rhizoid cell development if it reaches a threshold
325 level in the receiving cell. MpFRH1 activity is required for this repressive signal. There are at
326 least three potential modes of signalling. First, the repressive signal could be MpFRH1
327 miRNA itself. miRNAs can be mobile and it has been suggested that movement of miRNAs
328 can account for the establishment of tissue boundaries in organs (Carlsbecker et al., 2010).
329 Second, the repressive signal could be produced as a result of MpFRH1 miRNA activity.
330 Since MpRSL1 mRNA is the only confirmed target of MpFRH1, the production of this
331 hypothetical signal would likely also be MpRSL1-dependent (Honkanen et al., 2018). Third,
332 the repressive signal could be transmitted by cell contact through an unknown mechanism.
333 The amount of contact could be proportional to total shared cell surface area shared
334 between neighbouring cells. Accordingly, sharing one cell face would expose the cells to
335 sub-threshold levels of repressor while sharing a cell face with two rhizoid cells would
336 expose the cell to repressor levels above this threshold. Since MpRSL1 mRNA is the only
337 confirmed target of MpFRH1, the production of this hypothetical cell-contact signal would
338 also be MpRSL1-dependent.
339
340 While the molecular mechanisms of lateral inhibition are different between plants
341 (Myb repressor-, EDP2- and MpFRH1-dependent) and metazoans (Delta-Notch) there are
342 similarities in the underlying logic of the process. For example, the Delta ligand represses
343 neuroblast development during Notch-Delta signalling in Drosophila (reviewed in Bray,
344 2006). However, Delta is expressed on future neuroblasts and represses neuroblast
345 development in adjacent cells by non-cell autonomous signalling mediated by the Notch
346 receptor located on the neighbouring cell. That is, the repressor acts at a distance from its
347 site of synthesis to repress the rhizoid cell identity in adjacent cells. This logic operates in
348 Myb repressor- and EDP2-mediated lateral inhibition in *A. thaliana*. Similar logic operates in
349 MpFRH1-mediated lateral inhibition in *M. polymorpha*. MpFRH1 is a repressor of rhizoid cell
350 development that it is expressed in the developing rhizoid cells. Furthermore, our data
351 indicate that MpFRH1 miRNA represses rhizoid cell identity in adjacent cells that would

352 otherwise differentiate as flat epidermal cells in wild type. This leads to the hypothesis that
353 MpFRH1 expression in developing rhizoid cells acts non-cell autonomously to repress rhizoid
354 cell development in epidermal cells next to rhizoid cells. While the molecular mechanism of
355 lateral inhibition is different between plants and animals, the regulatory similarities suggest
356 that there may be underlying similarities in control logic.
357
358 The MpRSL1-MpFRH1 mechanism of lateral inhibition that controls the patterning of
359 rhizoid cells is liverwort-specific. MpFRH1 is a liverwort-specific miRNA and the miRNA
360 target sequence has been identified in the RSL genes of many liverwort taxa but has not
361 been found in RSL genes from any other lineage of land plants (Honkanen et al., 2018). This
362 indicates that the mechanism of lateral inhibition in the liverwort lineage is entirely
363 different from the two mechanisms that has been described among the angiosperms (Myb
364 repressor and EPF2). Since the MpFRH1-lateral inhibition mechanism is restricted to the
365 liverworts it suggests that different lateral inhibition mechanisms control the patterning of
366 tip growing rooting cells, rhizoids and root hairs in liverworts and angiosperms respectively.
367 It also demonstrates that entirely different mechanisms can operate to control lateral
368 inhibition in early diverging groups of land plants (hornworts and mosses) than in
369 angiosperms. This contrasts with animals in which the Notch-Delta mechanism of lateral
370 inhibition is conserved among most metazoan lineages. We propose that different
371 mechanisms of lateral inhibition evolved many times during the course of land plant
372 evolution while a single mechanism has been conserved among metazoans.

373 **Methods**

374

375 **Plant material and growth conditions**

376 M. polymorpha accessions Tagaragaike-1 (Tak-1, male) and Tagaragaike-2 (Tak-2,
377 female)(Ishizaki et al., 2008) were used as wild type and grown as reported in (Honkanen et
378 al., 2016). Mp*RSL1*^{GOF} were generated by Proust et al., 2016. For RNA extraction plants were
379 grown on medium covered with cellulosic cellophane membrane (AA Packaging Limited,
380 Preston, UK) before harvesting to avoid agar contamination.

381

382 **CRISPR/Cas9 knock out**

383 CRISPR/Cas9 mutations were generated following the protocol described in (Sugano et al.,
384 2014). A modified version of pMpGE_En03 harbouring a second sgRNA scaffolds with a
385 preceding BsmBI restriction site under the pU6 promoter was kindly provided by Holger
386 Breuninger, named pHB453. Primer oAT110 and oAT111 were used to introduce sgRNA1
387 into the BbsI site, primer oAT112 and oAT113 were used to introduce sgRNA2 into BsmBI
388 site. MpFRH1 allele was amplified using primer oAT135 and pAT136 and subsequentially
389 sequenced by Source BioScience.

390

391 **qPCR of MpRSL1 and MpAPT**

392 Total RNA was extracted from 7-day old gemmae using Direct-Zol RNA MINIprep kit (Zymo
393 Research) following manufactures instructions. For each line three biological replicates were
394 extracted unless stated otherwise. To remove DNA, 3ug total RNA were treated with TURBO
395 DNA-free kit following manufacturers instruction. cDNA synthesis was performed according
396 to the First Strand cDNA Synthesis protocol from NEB (#M0368) using ProtoSript II Reverse
397 Transcriptase and Murine RNase Inhibitor. MpRSL1 and MpAPT cDNA was amplified using
398 the primer oAT173/oAT174 and oAT175/176, respectively (as previously reported in
399 Honkanen et al, 2018). qRT-PCT was performed in the Applied Biosystems 7300 Real-Time
400 PCR System (Life Technologies) with SensiMix SYBR Hi-ROX Kit (Bioline) following
401 manufactures instructions. Three technical replicates per biological replicate were
402 performed. qPCR data was first analysed using LinRegPCR v2012.0(Ruijter et al., 2009). The
403 average N_0 value was calculated. Relative RSL1 mRNA abundance was calculated by
404 normalising the N_0 of each replicate against the N_0 value of the reference gene, MpAPT1.

405 **Stem loop PCR for MpFRH1 and MpmiR166**

406 To amplify MpFRH1 and MpmiR166, RNA was extracted from 19 day old gemmae using
407 miRVana miRNA Isolation Kit following manufacturer's instructions. 1ug RNA was treated
408 with DNase according to TURBO DNA-free kit following manufacturers instruction. Stem-
409 loop PCR was carried out as described in (Varkonyi-Gasic et al., 2007) using a MpFRH1
410 specific primer (oAT218) or a MpmiR166 specific primer (oAT219). The reverse transcribed
411 and extended miRNAs were amplified using PCRBIO Ultra Polymerase with a miRNA specific
412 forward primer (oAT221 for MpFRH1, oAT222 for MpmiR166) and a universal reverse
413 primer (oAT220). Amplicons were visualised on a 3% agarose gel containing SYBR Safe.
414

415 **Imaging**

416 Cell walls were stained using propidium iodide (PI). Gemmae were incubated in 5ug/ml PI
417 solution for (5-)10 minutes. PI solution was removed and gemmae washed twice with MilliQ
418 water. Gemmae were mounted on a slide with heated 0.2% agar solution. Once Agar
419 solidified gemmae were imaged immediately. Images were acquired with a Leica SP5
420 confocal laser microscope and the Leica Application Suite (LAS) software using either a Leica
421 HCX PL Fluotar 10x/0.30 or HC PL APO 20x/0.75 IMM CORR CS2 lense. PI and chlorophyll
422 were excited at 543 nm and emission was collected between 561 and 640 nm for PI and
423 between 680-700nm for chlorophyll.
424 For Scanning Electron Micropscopy (SEM), gemmae were fixed in 100% dry methanol for 30
425 minutes and then washed twiced with 100% dry ethanol. Gemmae were stored in 100% dry
426 ethanol at 4 °C upon processing. The samples were dried at the critical point using a
427 Tousimis Autosamdri-815, then mounted on aluminium stubs (Agar Scientific) using double-
428 sided carbon adhesive discs (Agar Scientific), and coated with a mixture of gold and
429 palladium using an SC7640 sputter coater (Quorum Technologies). The samples were
430 imaged immediately with a JSM-5510 scanning electron microscope (JEOL) at an operating
431 voltage of 15kV.

432 **Modelling**

433 The model was implemented as follows. (1) A grid of ~400 hexagonal cells was defined. A
434 predefined density of cells were selected as rhizoid precursors. (2) A probability P defined
435 the probability of a rhizoid cluster to include an additional neighbouring cell. Each rhizoid
436 cluster was tested. If a cluster did not add a new cell (probability 1-P), then that cluster was
437 considered to have finished expansion. (3) The simulation performed step (2) iteratively
438 until all clusters finished expansion. In this simple scenario, the probability of a cluster
439 having *n* cells = $(1-P)^{n-1}$.

440

441 When a rhizoid cluster is selected for expansion, two possible rules were considered. (1) No
442 lateral inhibition; in this case, any neighbouring cell to the cluster can become integrated
443 within the cluster. (2) With lateral inhibition; only cells neighbouring exactly one member of
444 the rhizoid cluster can become integrated within the cluster. We also considered the
445 restriction that the new rhizoid cell has to be a neighbour of an end cell of the cluster. This
446 corresponds to four possible configurations ((i) lateral inhibition with only end joining, (ii)
447 lateral inhibition with new cells allowed so long as only one neighbour in the rhizoid cluster,
448 (iii) no lateral inhibition with only end joining, (iv) no lateral inhibition with new cells
449 allowed so long as only one neighbour in the rhizoid cluster). The results presented in the
450 paper for wild type and mutants correspond to cases (i) and (iv) respectively. However,
451 using rules (ii) and (iii) do not alter the general results, though clusters become more
452 irregular as branching can occur.

453

454 When lateral inhibition was present, two nearby rhizoid clusters cannot merge; any cell
455 between two clusters would have a minimum of two rhizoid neighbours. In the absence of
456 lateral inhibition, such a cell can take on the rhizoid cell fate. Hence, two clusters can merge
457 to form a larger cluster. In this way, although P is unchanged in all simulation, larger clusters
458 are possible in the absence of lateral inhibition.

459

460 The model was encoded in Matlab. All data is produced from averaging the average cluster
461 distributions for 1000 simulations. At each decision step, a uniform random number
462 between 0 and 1 was drawn using the function *rand* and tested against the value *P.*

463

464 **Primer list**
465

oAT110_MpFRH1_sgRNA1_for	CTCGCTAGCAGTTGCTGGAGTTCG
oAT111_MpFRH1_sgRNA1_rev	AAACCGAACTCCAGCAACTGCTAG
oAT112_MpFRH1_sgRNA2_for	CTCGGATCCTCACCTTCCTCACAG
oAT113_MpFRH1_sgRNA2_rev	AAACCTGTGAGGAAGGTGAGGATC
oAT135_FRH1_CRISPRgenotyping_for1	GTACTTCCATCAGCAGACGACATCA
oAT136_FRH1_CRIPSRgenotyping_rev1	ATCTGCATCCTTCAGTGTCCCCT
oAT173_qPCR_RSL1_f	AGATGAGTCTGGGGCAACC
oAT174_qPCR_RSL1_r	GGATGAGCGCTTTAGAGTGG
oAT175_qPCR_APT_f	CGAAAGCCCAAGAAGCTACC
oAT176_qPCR_APT_r	GTACCCCCGGTTGCAATAAG
oAT218_MpFRH1 specific RT primer	GTCGTATCCAGTGCAGGGTCCGAGG TATTCGCACTGGATACGACACATTG
oAT219_MpmiR166 specific RT primer	GTCGTATCCAGTGCAGGGTCCGAGG TATTCGCACTGGATACGACGGGAAT
oAT220_stemloop_universal reverse	CCAGTGCAGGGTCCGAGGTA
oAT221_SLPCR_MpFRH1	CGGCGTGTGTGAGAAGAGGC
oAT222_SLPCR_MpmiR166a	CGCTCTTCGGACCAGGCTTC

466

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477 **References**

478 Artavanis-Tsakonas S. 1999. Notch Signaling: Cell Fate Control and Signal Integration in
479 Development. *Science (80-)* **284**:770–776. doi:10.1126/science.284.5415.770
480 Bowman JL, Kohchi T, Yamato KT, Jenkins J, Shu S, Ishizaki K, Yamaoka S, Nishihama R,
481 Nakamura Y, Berger F, Adam C, Aki SS, Althoff F, Araki T, Arteaga-Vazquez MA,
482 Balasubrmanian S, Barry K, Bauer D, Boehm CR, Briginshaw L, Caballero-Perez J,
483 Catarino B, Chen F, Chiyoda S, Chovatia M, Davies KM, Delmans M, Demura T,
484 Dierschke T, Dolan L, Dorantes-Acosta AE, Eklund DM, Florent SN, Flores-Sandoval E,
485 Fujiyama A, Fukuzawa H, Galik B, Grimanelli D, Grimwood J, Grossniklaus U, Hamada T,
486 Haseloff J, Hetherington AJ, Higo A, Hirakawa Y, Hundley HN, Ikeda Y, Inoue K, Inoue S
487 ichiro, Ishida S, Jia Q, Kakita M, Kanazawa T, Kawai Y, Kawashima T, Kennedy M, Kinose
488 K, Kinoshita T, Kohara Y, Koide E, Komatsu K, Kopischke S, Kubo M, Kyozuka J,
489 Lagercrantz U, Lin SS, Lindquist E, Lipzen AM, Lu CW, De Luna E, Martienssen RA,
490 Minamino N, Mizutani Masaharu, Mizutani Miya, Mochizuki N, Monte I, Mosher R,
491 Nagasaki H, Nakagami H, Naramoto S, Nishitani K, Ohtani M, Okamoto T, Okumura M,
492 Phillips J, Pollak B, Reinders A, Rövekamp M, Sano R, Sawa S, Schmid MW, Shirakawa
493 M, Solano R, Spunde A, Suetsugu N, Sugano S, Sugiyama A, Sun R, Suzuki Y, Takenaka
494 M, Takezawa D, Tomogane H, Tsuzuki M, Ueda T, Umeda M, Ward JM, Watanabe Y,
495 Yazaki K, Yokoyama R, Yoshitake Y, Yotsui I, Zachgo S, Schmutz J. 2017. Insights into
496 Land Plant Evolution Garnered from the Marchantia polymorpha Genome. *Cell*
497 **171**:287-304.e15. doi:10.1016/j.cell.2017.09.030
498 Bray SJ. 2006. Notch signalling: A simple pathway becomes complex. *Nat Rev Mol Cell Biol*
499 **7**:678–689. doi:10.1038/nrm2009
500 Carlsbecker A, Lee J-Y, Roberts CJ, Dettmer J, Lehesranta S, Zhou J, Lindgren O, Moreno-
501 Risueno M a, Vatén A, Thitamadee S, Campilho A, Sebastian J, Bowman JL, Helariutta Y,
502 Benfey PN. 2010. Cell signalling by microRNA165/6 directs gene dose-dependent root
503 cell fate. *Nature* **465**:316–321. doi:10.1038/nature08977
504 Catarino B, Hetherington AJ, Emms DM, Kelly S, Dolan L. 2016. The Stepwise Increase in the
505 Number of Transcription Factor Families in the Precambrian Predated the
506 Diversification of Plants on Land. *Mol Biol Evol* **33**:2815–2819.
507 doi:10.1093/molbev/msw155
508 Honkanen S, Jones VAS, Morieri G, Champion C, Hetherington AJ, Kelly S, Proust H, Saint-
509 Marcoux D, Prescott H, Dolan L. 2016. The Mechanism Forming the Cell Surface of Tip-
510 Growing Rooting Cells Is Conserved among Land Plants. *Curr Biol* **26**:3238–3244.
511 doi:10.1016/j.cub.2016.09.062
512 Honkanen S, Thamm A, Arteaga-Vazquez MA, Dolan L. 2018. Negative regulation of
513 conserved RSL class I bHLH transcription factors evolved independently among land
514 plants. *Elife* **7**. doi:10.7554/eLife.38529
515 Horst RJ, Fujita H, Lee JS, Rychel AL, Garrick JM, Kawaguchi M, Peterson KM, Torii KU. 2015.
516 Molecular Framework of a Regulatory Circuit Initiating Two-Dimensional Spatial
517 Patterning of Stomatal Lineage. *PLoS Genet* **11**:1–23.
518 doi:10.1371/journal.pgen.1005374
519 Ishizaki K, Chiyoda S, Yamato KT, Kohchi T. 2008. Agrobacterium-mediated transformation
520 of the haploid liverwort Marchantia polymorpha L., an emerging model for plant
521 biology. *Plant Cell Physiol* **49**:1084–1091. doi:10.1093/pcp/pcn085
522 Lee JS, Hnilova M, Maes M, Lin YCL, Putarjunan A, Han SK, Avila J, Torii KU. 2015.
523 Competitive binding of antagonistic peptides fine-tunes stomatal patterning. *Nature*

524 **522**:439–443. doi:10.1038/nature14561
525 Lee MM, Schiefelbein J. 2002. Cell pattern in the Arabidopsis root epidermis determined by
526 lateral inhibition with feedback. *Plant Cell* **14**:611–618. doi:10.1105/tpc.010434
527 Menand B, Yi K, Jouannic S, Hoffmann L, Ryan E, Linstead P, Schaefer DG, Dolan L. 2007. An
528 Ancient Mechanism Controls the Development of Cells with a Rooting Function in Land
529 Plants. *Science (80-)* **316**:1477–1480. doi:10.1126/science.1142618
530 Perrimon N, Pitsouli C, Shilo BZ. 2012. Signaling mechanisms controlling cell fate and
531 embryonic patterning. *Cold Spring Harb Perspect Biol* **4**:1–18.
532 doi:10.1101/cshperspect.a005975
533 Proust H, Honkanen S, Jones VAS, Morieri G, Prescott H, Kelly S, Ishizaki K, Kohchi T, Dolan L.
534 2016. RSL Class I Genes Controlled the Development of Epidermal Structures in the
535 Common Ancestor of Land Plants. *Curr Biol* **26**:93–99. doi:10.1016/j.cub.2015.11.042
536 Ruijter JM, Ramakers C, Hoogaars WMH, Karlen Y, Bakker O, van den hoff MJB, Moorman
537 AFM. 2009. Amplification efficiency: Linking baseline and bias in the analysis of
538 quantitative PCR data. *Nucleic Acids Res* **37**. doi:10.1093/nar/gkp045
539 Saint-Marcoux D, Proust H, Dolan L, Langdale JA. 2015. Identification of reference genes for
540 real-time quantitative PCR experiments in the liverwort marchantia polymorpha. *PLoS*
541 *One* **10**:1–14. doi:10.1371/journal.pone.0118678
542 Savage N Saint, Walker T, Wieckowski Y, Schiefelbein J, Dolan L, Monk NAM. 2008. A Mutual
543 Support Mechanism through Intercellular Movement of CAPRICE and GLABRA3 Can
544 Pattern the Arabidopsis Root Epidermis. *PLoS Biol* **6**:e235.
545 doi:10.1371/journal.pbio.0060235
546 Schellmann S, Schnittger A, Kirik V, Wada T, Okada K, Beermann A, Thumfahrt J, Jürgens G,
547 Hülskamp M. 2002. TRIPTYCHON and CAPRICE mediate lateral inhibition during
548 trichome and root hair patterning in Arabidopsis. *EMBO J* **21**:5036–5046.
549 doi:10.1093/emboj/cdf524
550 Simmons AR, Bergmann DC. 2016. Transcriptional control of cell fate in the stomatal
551 lineage. *Curr Opin Plant Biol* **29**:1–8. doi:10.1016/j.pbi.2015.09.008
552 Sugano SS, Shirakawa M, Takagi J, Matsuda Y, Shimada T, Hara-Nishimura I, Kohchi T. 2014.
553 CRISPR/Cas9-Mediated Targeted Mutagenesis in the Liverwort Marchantia polymorpha
554 L. *Plant Cell Physiol* **55**:475–481. doi:10.1093/pcp/pcu014
555 The Arabidopsis Genome Initiative. 2000. Analysis of the genome sequence of Arabidopsis
556 thaliana. *Nature* **408**:796–815.
557 Varkonyi-Gasic E, Wu R, Wood M, Walton EF, Hellens RP. 2007. Protocol: a highly sensitive
558 RT-PCR method for detection and quantification of microRNAs. *Plant Methods* **3**:12.
559 doi:10.1186/1746-4811-3-12
560 Wigge PA, Weigel D. 2001. Arabidopsis genome: Life without notch. *Curr Biol* **11**:112–114.
561 doi:10.1016/S0960-9822(01)00043-4
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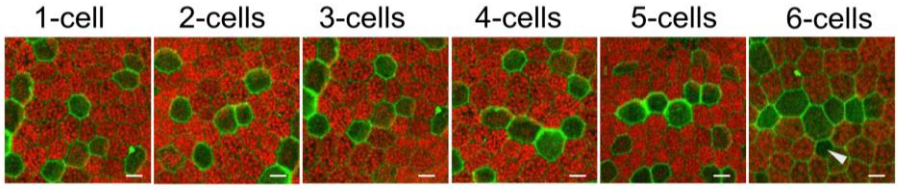


Tak-1	72.8 %	18.5 %	4.3 %	3.1 %	0.8 %	0.4 %
Tak-2	69.1 %	18.8 %	7.3 %	2.6 %	1 %	1 %
Mean RCN	0 ± 0	1 ± 0	1.36 ± 0.13	1.57 ± 0.27	1.6 ± 0	1.67 ± 0

Figure 1 Organisation of rhizoid clusters in the *M. polymorpha* gemma epidermis

Representative examples of individual rhizoid cells and clusters with between 2 and 6 rhizoid cells on mature gemmae of Tak-1 and Tak-2 wild type accessions. Chlorophyll autofluorescence is shown in red, cell walls of rhizoid cells are stained with propidium iodide (green). Tabular presentation of the frequencies of each size class of rhizoid cells cluster. The values represent the frequency of rhizoid cluster developing of each class as a percentage of the total number of rhizoid cell cluster. The mean rhizoid cell neighbour number (RCN) for each size class is represented with the standard deviation. (n [Tak-1] = 22 gemmae, n [Tak-2] = 23). Scale bar 20 µm, arrowhead indicates the position of an oil body cell which is smaller than a rhizoid cell.

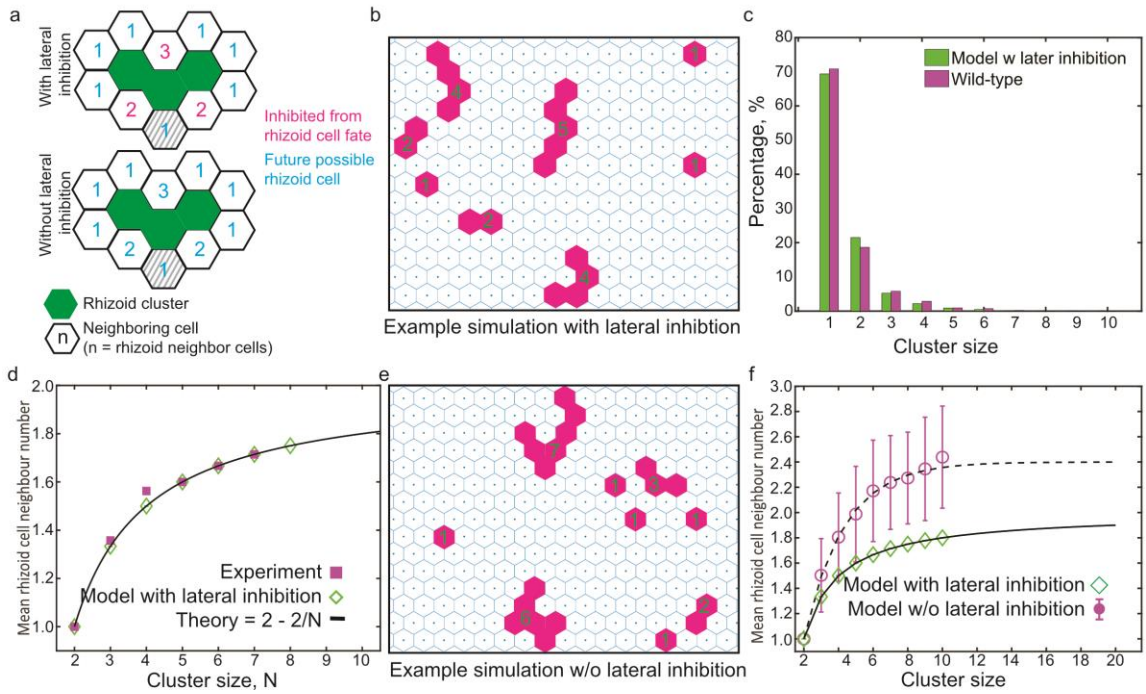


Figure 2 Modeling indicates that lateral inhibition can account for the patterning of rhizoid cells in the *M. polymorpha* gemma

a. Schematic of lateral inhibition model of rhizoid development. (Top) Lateral inhibition restricts the addition of a new cell to a rhizoid cluster to those only neighbouring a single rhizoid cell. (Bottom) In the absence of lateral inhibition, any cell can differentiate as a rhizoid cell. Grey cell represents a new possible rhizoid cell that is not adjacent to the end of the rhizoid cluster. We exclude such a possibility in the simulations, but including this does not alter the statistical results (Supplementary Information).
b. Output of the lateral inhibition model showing rhizoid cell distribution (magenta) and flat epidermal cells (white) in the gemma epidermis. In the presence of lateral inhibition, each cluster forms a linear array with cells arranged like beads on a string.
c. Validation of lateral inhibition model by comparing the frequency distribution of rhizoid cell clusters produced by the model (green, n = 500 simulations) and those observed in wild type gemma epidermis (magenta, “experiment”, n = 45 gemmae).
d. Validation of lateral inhibition model by comparing the relationship between mean rhizoid cell neighbour number and rhizoid cluster size as predicted by the model (green) with data from wild type gemmae (magenta).
e. Output of the model without lateral inhibition, showing the distribution of rhizoid cells (magenta) and flat epidermal cells (white) in the gemma epidermis. In the absence of lateral inhibition, clusters can develop in two-dimensional arrays.
f. Predicted relationship between mean rhizoid cell neighbour number and rhizoid cluster size in the presence and absence of lateral inhibition. The lower curve, with green points, indicates the output from the model that includes lateral inhibition. The upper curve with magenta points, indicates the output of the model without lateral inhibition. The main differences are than the mean rhizoid cell number is greater in the absence of lateral inhibition than with lateral inhibition for e each rhizoid cell cluster size and while these values are invariant in the presence of lateral inhibition they are variable in the absence of lateral inhibition.

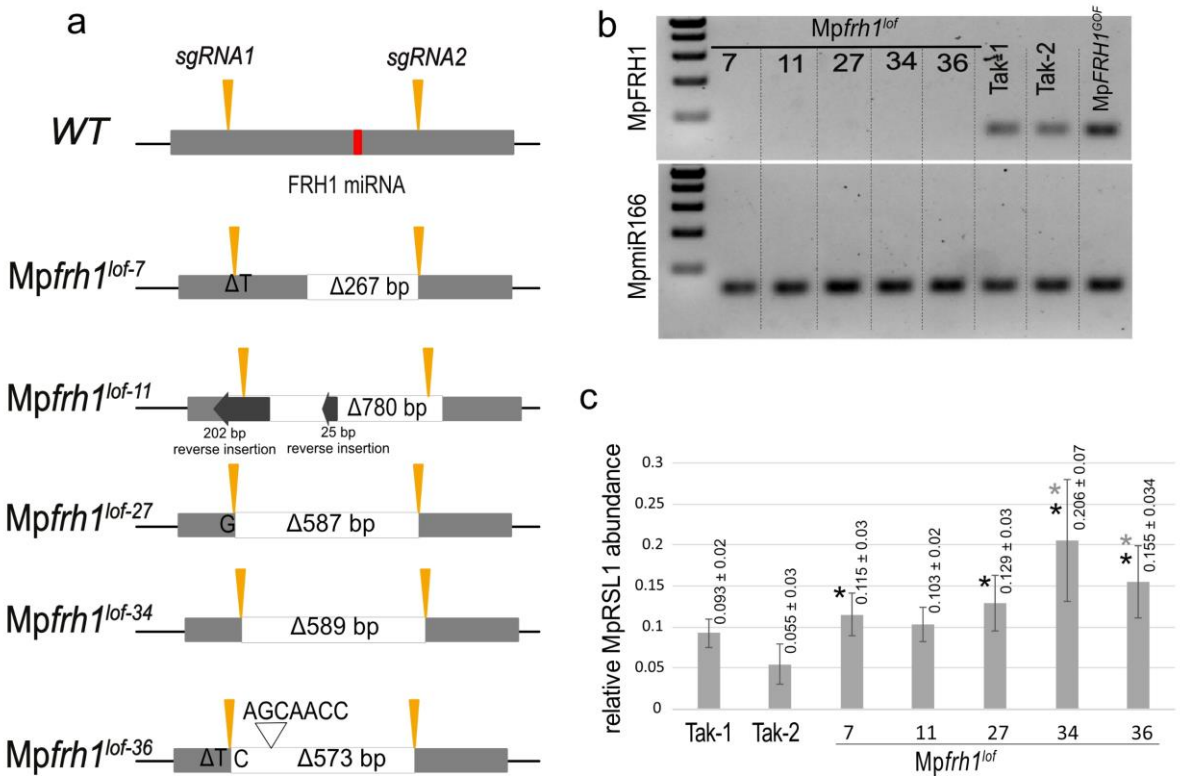


Figure 3 Genomic organisation of Mp*frh1* mutants and expression of Mp*RSL1* and Mp*FRH1* in Mp*frh1* loss of function mutants

a. Genomic organisation of the Mp*FRH1* gene in wild type (WT) and Mp*frh1*^{lof} mutants. The sequence for the 1.2kb MpFRH1 transcript is represented with a grey box. The 21 nucleotides corresponding to the MpFRH1 miRNA are indicated in red. The orange arrow heads indicate the position of the two single guide RNAs (sgRNAs) used to generate deletion mutants. sgRNA1 is complementary to a sequence that is between 384 and 404 bp 5' of the MpFRH1 sequence. SgRNA2 is complementary to a sequence that is 164-184 3' of the MpFRH1 sequence. The entire Mp*FRH1* miRNA sequence was deleted in each of the mutants. In Mp*frh1*$^{lof-7}$ there was a 267bp deletion between sgRNA1 and sgRNA2 and T deletion. In Mp*frh1*$^{lof-11}$ there was a deletion of 780bp and reverse insertions of 202bp and 26bp. In Mp*frh1* $^{lof-27}$ there was a 587bp deletion and a G insertion. In Mp*frh1*$^{lof-34}$ there was a 589bp deletion. In Mp*frh1*$^{lof-36}$ there was a 573bp deletion and substitution of T into C at the sgRNA1 target site and a 7bp insertion.

b. Amplification of MpFRH1 miRNA (upper row) and MpmiR166 control (lower row) using stem loop PCR on RNA isolated from 7-day old thalli grown from gemmae in Mp*frh1*^{lof} mutants, Tak-1, Tak-2, and Mp*FRH1*^{GOF} mutant.

c. Amplification of MpRSL1 mRNA from RNA isolated from in 7-day old thalli grown from gemmae in wild type (Tak-1, Tak-2) plants and Mp*frh1*^{lof} mutants; mRNA levels were normalized to the housekeeping gene Mp*APT1*. Three biological replicates per genotype, except for Mp*frh1*$^{lof-7}$(n=2). There were at least three technical replicates per biological replicate. Grey asterisk: significant difference to Tak-1, black asterisk: significant difference to Tak-2. $P<0.05$

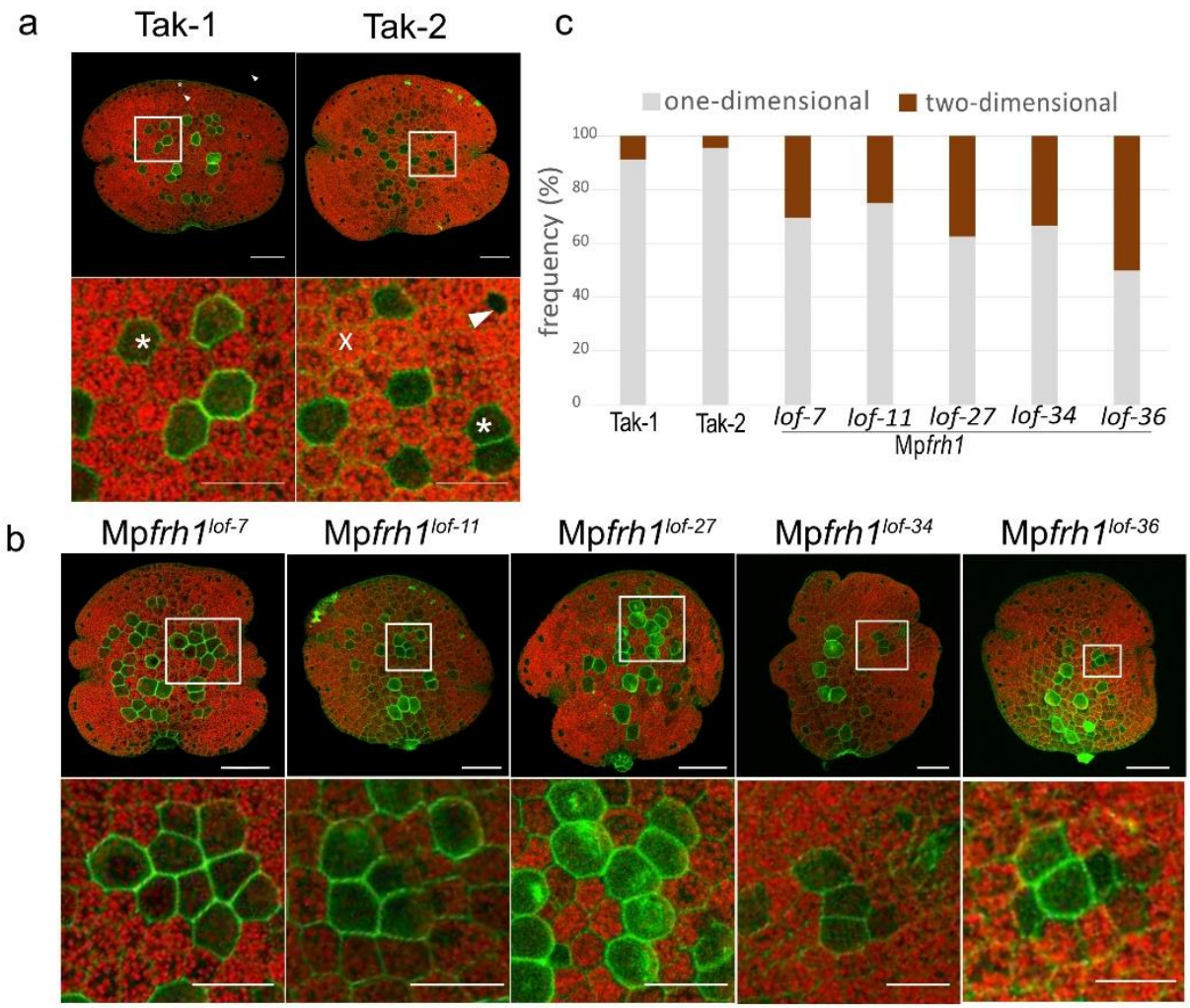
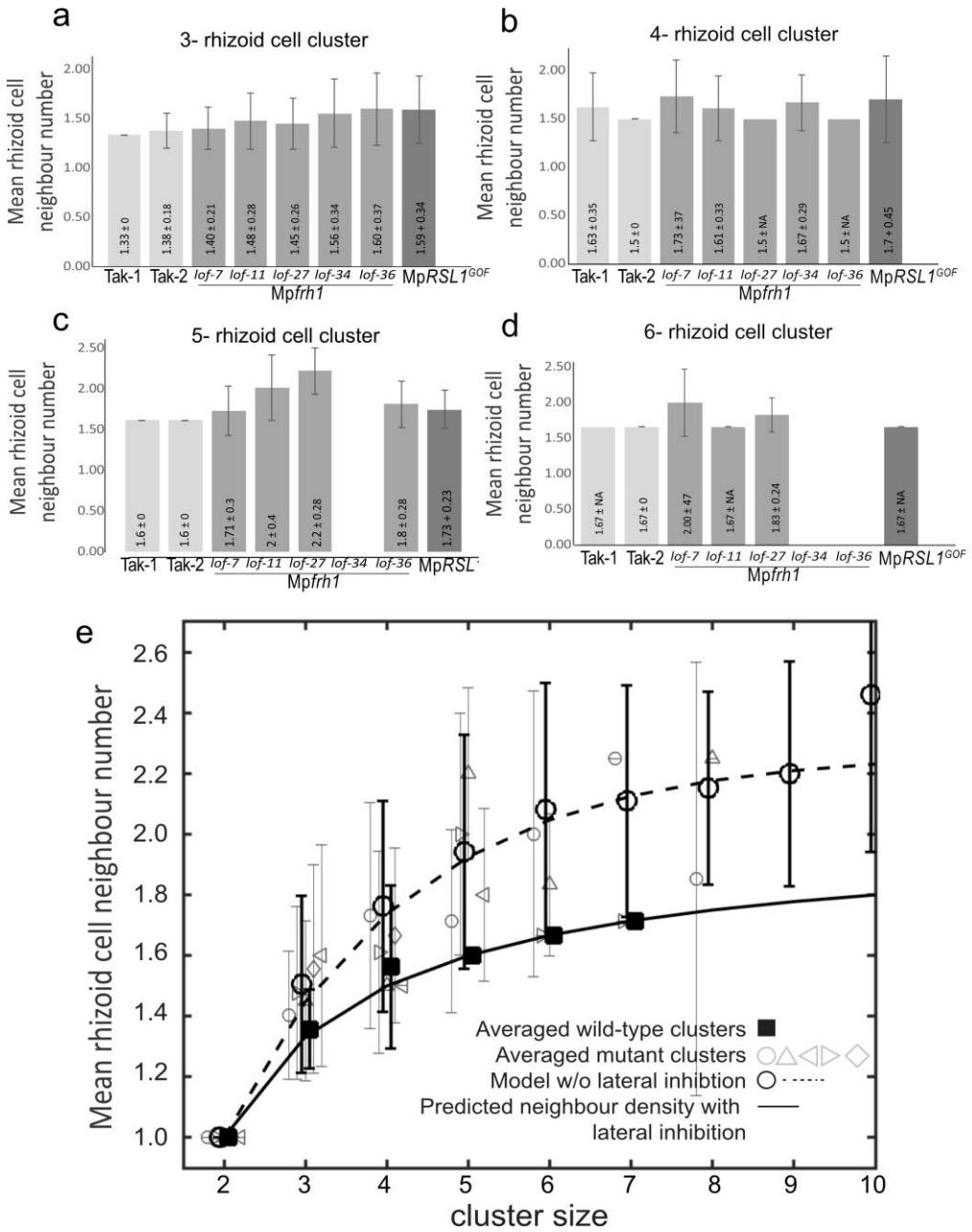


Figure 4 Two-dimensional rhizoid cell clusters develop in Mp*frh1*^{lof} and Mp*RSL1*^{GOF} mutants

a. Rhizoid patterning in gemmae of wild type (Tak-1, Tak-2). Upper row, overview of representative gemma (scalebar = 100 µm). Lower row shows the higher magnification of inset highlighted above, showing representative rhizoid clusters (Scalebar = 50µm).

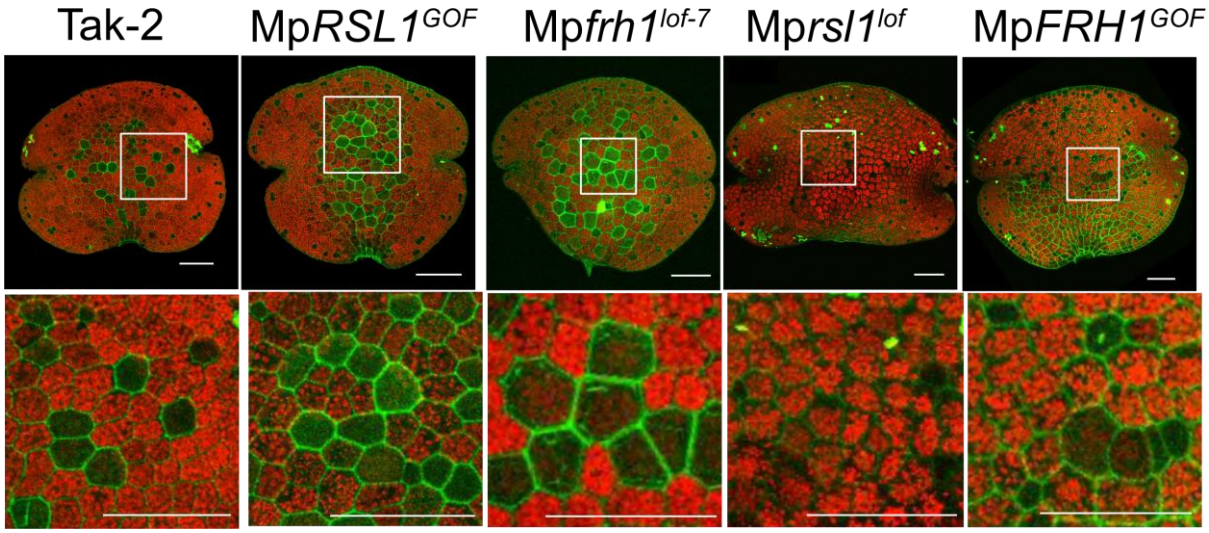
b. Rhizoid patterning in gemmae of Mp*frh1*$^{lof-7}$, Mp*frh1*$^{lof-11}$, Mp*frh1*$^{lof-27}$, Mp*frh1*$^{lof-34}$, Mp*frh1*$^{lof-36}$ mutants. Upper row, overview of representative gemma (scalebar = 100 µm). Lower row, magnification of highlighted square showing representative rhizoid cluster (scalebar = 50µm).

c. Frequency (in %) of one-dimensional and two-dimensional rhizoid cluster in wildtype (Tak-1, Tak-2) and Mp*frh1*^{lof} mutants. All rhizoid cluster larger than three cells were combined (n[Tak-1] = 23, n[Tak-2=23], n[Mp*frh1*$^{lof-7}$] = 46, n[Mp*frh1*$^{lof-11}$] = 28, n[Mp*frh1*$^{lof-27}$] = 24, n[Mp*frh1*$^{lof-34}$] = 9, n[Mp*frh1*$^{lof-36}$]=8).



643

644 **Figure 5 More rhizoid cell neighbours develop in Mp*frh1*^{lof} and Mp*RSL1*^{GOF} mutants**
645 a-d. Mean rhizoid cell neighbour number in 3-rhizoid cell clusters (a), 4-rhizoid cell clusters
646 (b), 5-rhizoid cell clusters (c), and 6-rhizoid cell clusters (d) in wild type (Tak-1, Tak-2),
647 Mp*frh1*$^{lof-7}$, Mp*frh1*$^{lof-11}$, Mp*frh1*$^{lof-27}$, Mp*frh1*$^{lof-34}$, Mp*frh1*$^{lof-36}$ and Mp*RSL1*^{GOF}.
648 e. The relationship between mean rhizoid cell neighbour numbers and cluster size predicted
649 by the model with lateral inhibition (solid line). The relationship predicted by the model
650 without lateral inhibition (open circles, n= 500 simulations; dashed line represents fit to
651 $2.4 - 1.4e^{-\frac{(N-2)}{2.3}}$ where *N* is cluster size). The experimentally observed relationship between
652 mean rhizoid cell neighbour numbers and cluster size in wild type (filled black square) and
653 Mp*frh1*^{lof} mutants (Mp*frh1*$^{lof-7}$ = grey circle, Mp*frh1*$^{lof-11}$ = grey triangle pointing to right,
654 Mp*frh1*$^{lof-27}$ = grey triangle pointing up, Mp*frh1*$^{lof-34}$ = grey diamond, Mp*frh1*$^{lof-36}$ = grey
655 triangle pointing to left). Errorbars are SD.



656

657 **Figure 6 Larger rhizoid cell clusters with altered shape develop in Mp*frh1*^{lof} and Mp*RSL1*^{GOF}**
658 **mutants than in wild type**
659 **Upper row shows distribution of rhizoid cells and rhizoid cell clusters on representative**
660 **mature gemmae of wild type (Tak-2), Mp*RSL1*^{GOF}, Mp*frh1*$^{lof-7}$, Mp*rsl1*^{lof},**
661 **Mp*FRH1*^{GOF}mutants. Scalebar = 100 µm. The lower row shows the higher magnification of**
662 **the inset highlighted on the image in the row above. There are 12 rhizoid cells in the**
663 **Mp*RSL1*^{GOF} rhizoid cell cluster and 8 cells in the Mp*frh1*$^{lof-7}$ rhizoid cell cluster. There are**
664 **no rhizoids in either the Mp*rsl1*^{lof} or Mp*FRH1*^{GOF} mutants. Chlororphyll autofluorescence is**
665 **red, and cell walls of rhizoids are stained with propidium iodide (green). Scalebar = 50 µm.**
666 **White arrowhead indicates the location of oil body cells.**

667 **Table 1 Frequency of individual rhizoid cells and rhizoid cluster in wildtype, Mp*frh1*^{lof} and**
668 **Mp*RSL1*^{GOF} mutants**

669

genotype	1	2	3	4	5	6	7	8	9	≥10	n cluster	n gemmae
Tak-1	72.5%	18.4%	4.3%	3.1%	0.8%	0.4%	0.4%				255	22
Tak-2	69.1%	18.8%	7.3%	2.6%	1.0%	1.0%					191	23
Mp*frh1* $^{lof-7}$	47.1%	23.2%	12.3%	9.0%	4.5%	1.3%		0.6%	1.9%		155	42
Mp*frh1* $^{lof-11}$	61.4%	23.8%	7.4%	4.8%	1.6%	0.5%	0.5%				189	27
Mp*frh1* $^{lof-27}$	63.2%	23.0%	9.8%	0.6%	1.1%	1.1%	0.6%	0.6%			174	22
Mp*frh1* $^{lof-34}$	48.0%	40.0%	8.0%	4.0%							75	9
Mp*frh1* $^{lof-36}$	69.4%	19.4%	6.9%	1.4%	2.8%						72	8
Mp*RSL1* ^{GOF}	58.6%	14.1%	13.1%	5.1%	3.0%	1.0%	2.0%			3.0%	99	10

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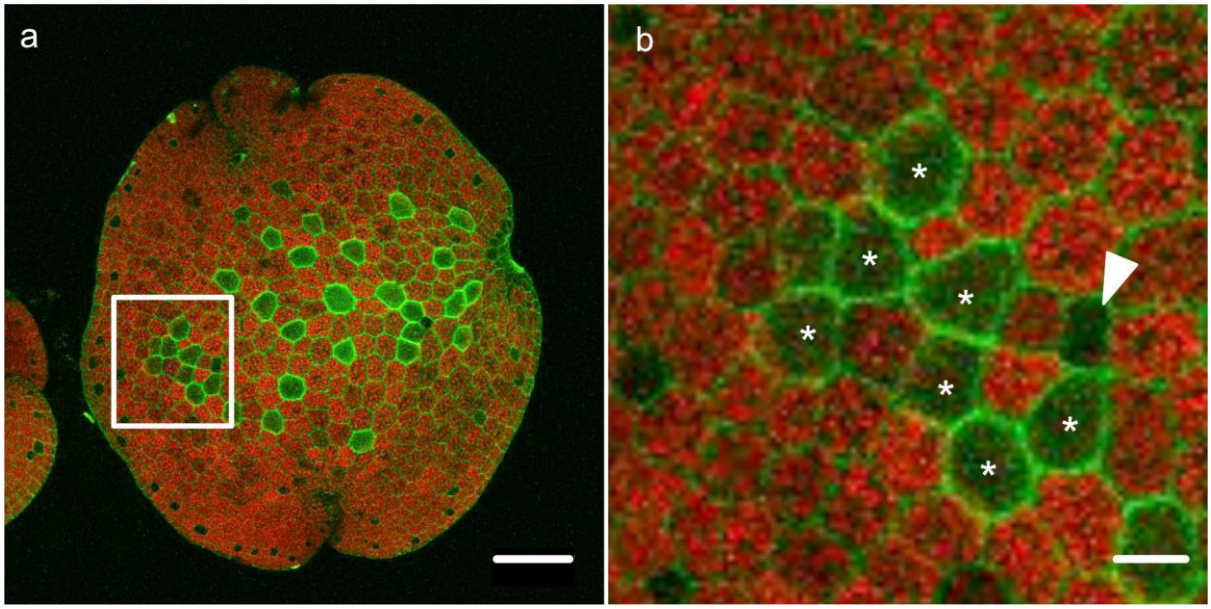
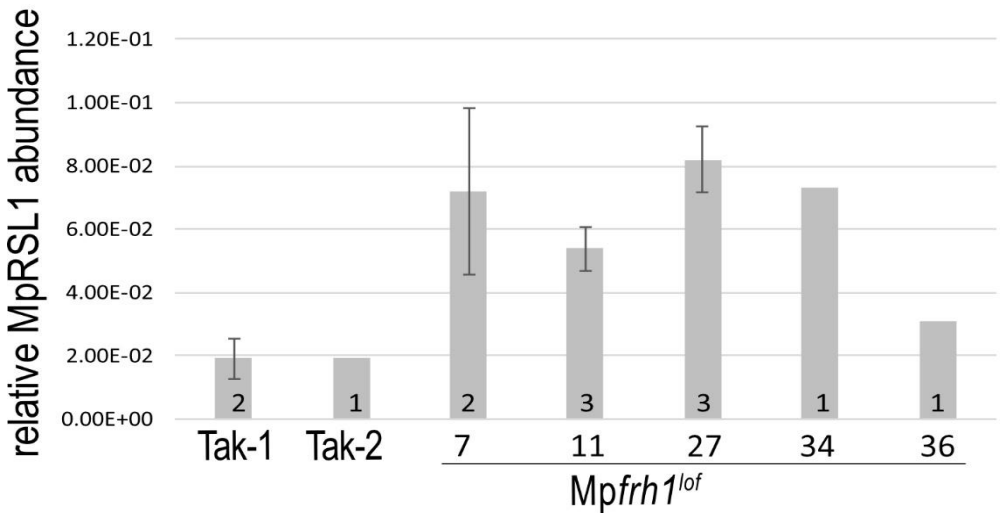


674

675 **Supplementary Figure 1 One 7-cell rhizoid cluster develop on Tak-2 gemma**

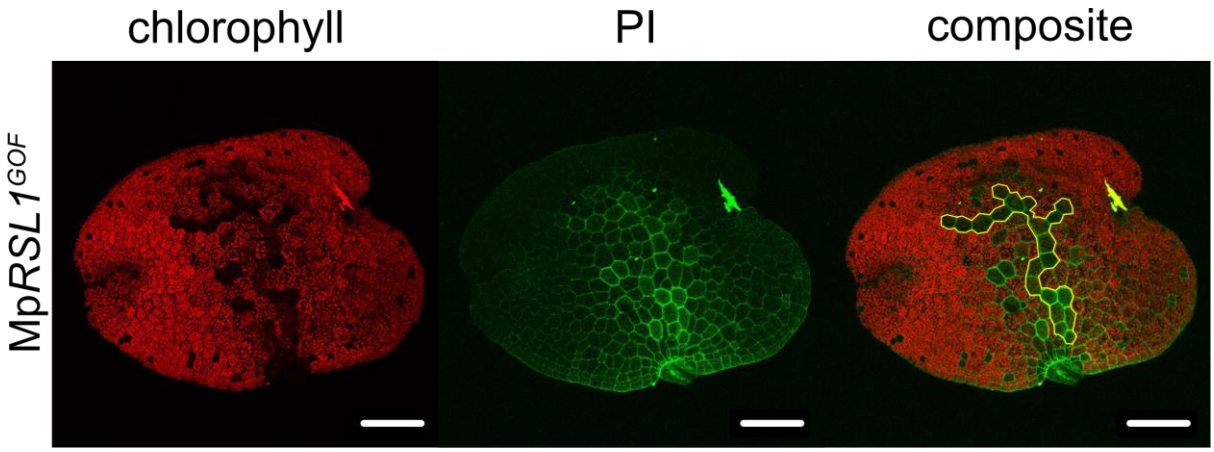
676 a. Rhizoid patterning of a Tak-2 gemma showing individual rhizoid cells, smaller rhizoid
677 cluster and a 7-cell rhizoid cell cluster (chlorophyll in red, PI in green, scalebar = 100 µm).
678 b. Magnification of the 7-cell rhizoid cluster highlighted in a (scalebar = 20 µm).



679

680 **Supplementary Figure 2 expression of MpRSL1 in Mpfrh1^{lof} mutants**

681 Amplification of MpRSL1 mRNA from RNA isolated from in 7-day old thalli grown from
682 gemmae in wild type (Tak-1, Tak-2) plants and Mp*frh1*^{lof} mutants; mRNA levels were
683 normalized to the housekeeping gene Mp*APT1*. n(Tak-1) = 2, n (Tak-2) = 1, n(Mp*frh1*$^{lof-7}$) = 2,
684 n(Mp*frh1*$^{lof-11}$) = 3, n(Mp*frh1*$^{lof-27}$) = 3, n(Mp*frh1*$^{lof-34}$) = 1, n(Mp*frh1*$^{lof-36}$) = 1.



Supplementary Figure 3 21-cell rhizoid cluster develop on Mp*RSL1*^{GOF} gemma

Rhizoid patterning of a Mp*RSL1*^{GOF} gemma showing a 21-cell rhizoid cell cluster highlighted as yellow perimeter (first panel = chlorophyll, second panel = PI, third panel = composite, scalebar = 100 µm).