Inferring parameters of the distribution of fitness effects of new mutations when beneficial mutations are strongly advantageous and rare

Tom R. Booker[1,2]

1. Department of Forest and Conservation Sciences, University of British Columbia, Vancouver, Canada.

2. Biodiversity Research Centre, University of British Columbia, Vancouver, Canada

Correspondence: booker@zoology.ubc.ca

Abstract

Characterising the distribution of fitness effects (DFE) for new mutations is central in evolutionary genetics. Analysis of molecular data under the McDonald-Kreitman test has suggested that adaptive substitutions make a substantial contribution to between-species divergence. Methods have been proposed to estimate the parameters of the distribution of fitness effects for positively selected mutations from the unfolded site frequency spectrum (uSFS). However, when beneficial mutations are strongly selected and rare, they may make little contribution to standing variation and will thus be difficult to detect from the uSFS. In this study, I analyse uSFS data from simulated populations subject to advantageous mutations with effects on fitness ranging from mildly to strongly beneficial. When advantageous mutations are strongly selected and rare, there are very few segregating in populations at any one time. Fitting the uSFS in such cases leads to erroneous parameter estimates which may lead researchers to false conclusions regarding the relative contribution adaptive mutations make to molecular evolution. Fortunately, the parameters for the distribution of fitness effects for harmful mutations are typically estimated with high accuracy and precision. The results from this study suggest that the parameters of positively selected mutations obtained by analysis of the uSFS should be treated with caution and suggest that variability at linked sites should be used in conjunction with standing variability to estimate parameters of the distribution of fitness effects in the future.

Introduction

Characterising the distribution of fitness effects for beneficial mutations is central in evolutionary biology. The rate and fitness effects of advantageous mutations may determine important evolutionary processes such as how variation in quantitative traits is maintained (Hill, 2010), the evolution of sex and recombination (Otto, 2009) and the dynamics of evolutionary rescue in changing environments (Orr & Unckless, 2014). However, despite its central role in evolution, relatively little is known about the distribution of fitness effects (DFE) for advantageous mutations in natural populations. The DFE for advantageous mutations can be estimated from data obtained via targeted mutation or from mutation accumulation experiments (e.g. Bank et al., 2014; Böndel et al., 2019), but such efforts may be limited to laboratory systems. Alternatively, estimates of the DFE can be obtained for natural systems using population genetic methods.

When natural selection is effective, beneficial alleles are promoted to eventual fixation while deleterious variants are maintained at low frequencies. Under recurrent mutation, selection and genetic drift interact to shape the distribution of allele frequencies in a population (Wright, 1937). Parameters of the DFE for both advantageous and deleterious mutations can be estimated by modelling population genomic data, specifically the site frequency spectrum (SFS). The SFS is the distribution of allele frequencies present in a sample of individuals drawn from a population. By contrasting the SFS for a class of sites expected to be subject to selection with that of a linked neutral comparator, one can estimate the parameters of the DFE if selected mutations are segregating in the population of interest (reviewed in Eyre-Walker & Keightley, 2007). Typically, the DFE for nonsynonymous sites in protein coding genes is estimated using synonymous sites as the neutral comparator. Several methods have been proposed that estimate the DFE for deleterious mutations from the SFS under the assumption that beneficial mutations contribute

little to standing genetic variation (e.g. Barton & Zeng, 2018; Boyko et al., 2008; Keightley & Eyre-

Walker, 2007; Tataru et al.,, 2017).

The DFE for deleterious mutations can be used when estimating α, the proportion of between-

species divergence attributable to adaptive evolution (Eyre-Walker & Keightley, 2009). α can be

estimated by rearranging the terms of the McDonald-Kreitman test (MK-test), which assesses the

extent of positive selection. Under strong purifying selection, the ratio of divergence at

nonsynonymous sites (d_N) to that of synonymous sites (d_S) should be exactly equal to the ratio of

nucleotide diversity at nonsynonymous (π_N) and synonymous sites (π_S)(McDonald & Kreitman,

1991). Adaptive evolution of protein sequences may contribute to d_N such that $d_N/d_S > \pi_N/\pi_S$.

Smith & Eyre-Walker (2002) suggested that the excess d_N due to positive selection (α) can be

estimated as

79                                  $\alpha = 1 - d_S\pi_N/d_N\pi_S.$

Slightly deleterious alleles may contribute to both standing genetic variation and between-species

divergence, estimates of α may therefore be refined by subtracting the contribution that

deleterious alleles make to both polymorphism and divergence and this can be calculated using

the DFE for harmful mutations (Eyre-Walker & Keightley, 2009). Application of such methods to

natural populations suggest that α is of the order of 0.5 in a large variety of animal taxa (Galtier,

2016). However, if adaptive evolution is as frequent as MK-test analyses suggest, the assumption

that advantageous alleles contribute little to standing variation may be violated and ignoring them

could lead to biased estimates of the DFE (Tataru et al., 2017).

When advantageous alleles contribute to standing variation, parameters of the DFE for both

deleterious and beneficial mutations can be estimated from the SFS (Schneider et al., 2011; Tataru

et al., 2017). When data from an outgroup species are available, variable sites within a focal

species can be polarised as either ancestral or derived and the *unfolded* SFS (uSFS) can be

obtained. The uSFS is a vector of length $2n$, where n is the number of haploid genome copies

sampled. The ith entry of the uSFS is the count of derived alleles observed at a frequency i in the

sample. Note that when outgroup data are not available, alleles cannot be polarised and the

distribution of minor allele frequencies (known as the *folded* SFS) is analysed. There is limited

power to detect positive selection from the uSFS, so the DFE for beneficial mutations is often

modelled as a discrete class of mutational effects, with one parameter specifying the fitness

effects of beneficial mutations, $\gamma_a = 2N_e s_a$ where N_e is the effective population size and s_a is the

positive selection coefficient in homozygotes, and another specifying the proportion of new

mutations that are advantageous, p_a. Estimates of γ_a and p_a for nonsynonymous sites have only

been obtained a handful of species, and these are summarised in Table 1. The positive selection

parameter estimates that have been obtained for mice and *Drosophila* are fairly similar (Table 1).

Note that the estimates for humans obtained by Castellano et al, (2019) did not provide a

significantly greater fit to the observed data than did a model with no positive selection.

Furthermore, Castellano et al, (2019) estimated the parameters for numerous great ape species,

the parameters shown for humans are representative of the estimates for all taxa they analysed.

Table 1 Estimates of the parameters of positive selection obtained from the uSFS for
nonsynonymous sites.

Common name	Scientific name	γ_a	p_a	Reference	Method used[¶]
House mouse	*Mus musculus castaneus*	14.5	0.0030	Booker & Keightley, (2018)	*DFE-alpha*
Fruit fly	*Drosophila melanogaster*	23.0	0.0045	Keightley et al, (2016)	*DFE-alpha*
Humans	*Homo sapiens*	0.0064[†]	0.000025	Castellano et al, (2019)	*polyDFE*

[¶] - DFE-alpha implements the analysis methods described by Schneider et al., (2011), *polyDFE* implements the
methods described by Tataru et al., (2017)
[†] - Castellano et al., (2019) estimated the mean fitness effect for an exponential distribution of advantageous
mutational effects.

Depending on the rate and fitness effects of beneficial mutations, different aspects of population genomic data may be more or less informative for estimating the parameters of positive selection. As beneficial mutations spread through populations, they may carry linked neutral variants to high frequency, causing selective sweeps (Barton, 2000). On the other hand, if advantageous mutations have mild fitness effects, they may take a long time to reach fixation and make a substantial contribution to standing genetic variation. Campos, et al., (2017) used a model of selective sweeps to analyse the negative correlation observed between d_N and π_S in *Drosophila melanogaster* and estimated γ_a = 250 and p_a = 2.2 x 10^{-4}. An analysis of the uSFS from the same dataset yielded estimates of γ_a = 23 and p_a = 0.0045 for nonsynonymous sites (Keightley et al., 2016). The sharp contrast between the two studies' estimates of the positive selection parameters may be explained if the DFE for advantageous mutations in *D. melanogaster* is bimodal. If this were so, the different methods (i.e. sweep models versus uSFS analysis) may be capturing distinct aspects of the DFE for advantageous mutations. The handful of studies which have attempted to estimate γ_a and p_a from the uSFS have yielded similar estimates of positive selection (Table 1), which may indicate commonalities in the DFE for beneficial mutations across taxa. On the other hand, it could be that uSFS analyses have only found evidence for mildly beneficial mutations because the approach is only powered to detect weakly beneficial mutations. Indeed, it has been argued that rare strongly selected advantageous mutations, which may contribute little to standing variation, will be undetectable by analysis of the uSFS (Booker & Keightley, 2018; Campos et al., 2017).

The studies describing the two most recently proposed methods for estimating the DFE for beneficial mutations from the uSFS (Schneider et al., 2011; Tataru et al., 2017) performed extensive simulations, but did not test the case of rare advantageous mutations (p_a < 0.01) with strong effects on fitness (γ_a > 24). Testing this case is important, as studies have indicated that the DFE for advantageous mutations contains strongly beneficial mutations in a variety of taxa (Booker

141 & Keightley, 2018; Campos et al., 2017; Elyashiv et al., 2016; Nam et al., 2017). Note that Tataru et

142 al., (2017) did simulate a population subject to frequent strongly beneficial mutations (γ_a = 800

143 and p_a = 0.02), but the parameter combination they tested may not be biologically relevant as the

144 proportion of adaptive substitutions it yielded was far higher than is typically estimated from real

145 data (α = 0.99). The limited parameter ranges tested in the simulations performed by Schneider et

146 al., (2011) and Tataru et al., (2017) leave a critical gap in our knowledge as to how uSFS based

147 methods perform when advantageous mutations are strongly selected and infrequent.

149 In this study, I use simulated datasets to fill this gap and examine how uSFS-based analyses

150 perform when beneficial mutations are strongly selected and rare. I simulate populations subject

151 to a range of positive selection parameters, including cases similar to those modelled by Tataru et

152 al., (2017) and cases where beneficial mutations are strongly selected but infrequent. It has been

153 pointed out that estimating selection parameters by modelling within species polymorphism along

154 with between-species divergence makes the assumption that the DFE has remained invariant since

155 the ingroup and outgroup began to diverge (Tataru et al., 2017). By analysing only the

156 polymorphism data, one can potentially avoid that potentially problematic assumption. Using the

157 state-of-the-art package *polyDFE* v2.0 (Tataru & Bataillon, 2019), I analyse the uSFS data and

158 estimate selection parameters for all simulated datasets with or without divergence. The results

159 from this study suggest that, when beneficial mutations are strongly selected and rare, analysis of

160 the uSFS results in spurious parameter estimates and the proportion of adaptive substitutions may

161 be poorly estimated.

Methods

Population genomic simulations

I tested the hypothesis that the parameters of infrequent, strongly beneficial mutations are

difficult to estimate by analysis of the uSFS using simulated datasets. Wright-Fisher populations of

N_e = 2,500 diploid individuals were simulated using the forward-in-time package *SLiM* (v3.2; Haller

& Messer, 2019). Simulated chromosomes consisted of seven gene models, each separated by

8,100bp of neutrally evolving sequence. The gene models consisted of five 300bp exons separated

by 100bp neutrally evolving introns. The gene models were based on those used by Campos &

Charlesworth, (2019), but unlike that study, I did not model the untranslated regions of the genes.

Nonsynonymous sites were modelled by drawing the fitness effects for 2/3rds of mutations in

exons from a distribution of fitness effects (DFE), while the remaining 1/3 were strictly neutral and

used to model synonymous sites. The fitness effects of nonsynonymous mutations were beneficial

with probability p_a or deleterious with probability $1 - p_a$. Beneficial mutations had a fixed selection

coefficient of $\gamma_a = 2N_e s_a$. The fitness effects of deleterious mutations were drawn from a gamma

distribution with a mean of $\gamma_d = 2N_e s_d$ = -2,000 and a shape parameter of β = 0.3 (s_d being the

negative selection coefficient in homozygotes). The gamma distribution of deleterious mutational

effects was used for all simulated datasets and was based on results for nonsynonymous sites in

Drosophila melanogaster (Loewe & Charlesworth, 2006). Uniform rates of mutation (μ) and

recombination (r) were set to 1 x 10^{-6} (giving $4N_e r = 4N_e \mu$ = 0.01). Note that μ and r are far higher

than is biologically realistic for most eukaryotes, I scaled up these rates to model a population with

a large N_e using simulations of 2,500 individuals. Across simulations I varied the γ_a and p_a

parameters and performed 2,000 replicates for each combination of parameters. Thus, I simulated

a dataset of 21Mbp of coding sequence for each combination of γ_a and p_a tested.

To model the accumulation of nucleotide substitutions after the split of a focal population with an

outgroup, I recorded all substitutions that occurred in the simulations. Campos & Charlesworth,

(2019) analysed simulations very similar to those that I performed in this study and showed that

populations subject to beneficial mutations with γ_a = 250 and p_a = 0.0002 took $14N_e$ generations

to reach mutation-selection-drift equilibrium. In this study I modelled a range of positive selection

parameters, so to ensure that my simulations reached equilibrium I performed 85,000 ($34N_e$)

generations of burn-in before substitutions were scored. The expected number of neutral

nucleotide substitutions that accumulate per site in T generations is $d_{Neutral} = T\mu$. The point

mutation rate in my simulations was set to $\mu = 1 \times 10^{-6}$ per site per generation, so I ran the

simulations for 50,000 generations beyond the end of the burn-in phase to model a neutral

divergence of $d_{Neutral}$ = 0.05. All variants present in the population sampled at a frequency of 1.0

were also scored as substitutions.

Using the simulated datasets, I constructed 100 bootstraps by sampling the set of 2,000 replicates

with replacement. From each bootstrap sample, I collated variants and constructed the uSFS for

synonymous and nonsynonymous sites for 20 diploid individuals.

Analysis of simulation data

I calculated several summary statistics from the simulated datasets. Firstly, I calculated pairwise

nucleotide diversity at synonymous sites (π_s) and expressed it relative to the neutral expectation

of $\pi_0 = 4N_e\mu$ = 0.01. Secondly, divergence at nonsynonymous sites for both advantageous (dN_a)

and deleterious mutations (dN_d) was used to calculate the observed proportion of adaptive

substitutions, $\alpha_{Obs} = dN_a/(dN_a + dN_d)$. Finally, I recorded the total number of beneficial mutations

segregating in simulated populations, S_{Adv}.

I estimated DFEs from simulated data by analysis of the uSFS using *polyDFE* (v2.0; Tataru &

Bataillon, 2019). *polyDFE* fits an expression for the uSFS expected under a full DFE to data from

putatively neutral and selected classes of sites and estimates parameters by maximum likelihood.

For each set of positive selection parameters, simulated uSFS data were analysed under "Model B"

in *polyDFE* (a gamma distribution of deleterious mutational effects plus a discrete class of

advantageous mutations). Initial parameters for the maximisation were calculated from the data

using the '-e' option and the uSFS was analysed either with or without divergence using the "-w"

option in *polyDFE*. Analysing the uSFS without divergence causes the selection parameters to be

inferred from polymorphism data alone. For each replicate, I tested whether the inclusion of

beneficial mutations in the DFE improved model fit using likelihood ratio tests between the best-

fitting model and a model with p_a set to 0.0. Setting p_a = 0.0 means that positive selection does

not influence the likelihood, so two fewer parameters are being estimated. Twice the difference in

log-likelihood between the full DFE model and the model with p_a = 0.0 was tested against a χ^2

distribution with 2 degrees of freedom. Likelihood surfaces were estimated by running *polyDFE*

using fixed values for DFE parameters.

Data Availability

228   All code and *SLiM* configuration files needed to reproduce the results shown in this study are

229   available at https://github.com/TBooker/PositiveSelection_uSFS.

Results

Population genomic simulations

I performed simulations that modelled genes subject to mutation-selection-drift balance with

fitness effects drawn from a distribution that incorporated both deleterious and advantageous

mutations. The DFE for harmful mutations was constant, but I varied the fraction (p_a) and fitness

effects (γ_a) of beneficial mutations across simulated datasets (Table 2). For each set of

advantageous mutation parameters, 21Mbp of coding sequences was simulated, of which 14Mbp

were nonsynonymous and 7Mbp were synonymous sites. Variants present in the simulated

populations were used to construct the uSFS for a sample of 20 diploid individuals, a sample size

which is fairly typical of current population genomic datasets (e.g. Castellano et al., 2019; Laenen

et al., 2018; Williamson et al., 2014).

Table 2 Parameters of positive selection assumed in simulations and the proportion of *polyDFE*
runs for which modelling positive selection gave a significantly better fit to the data.

| γ_a | p_a | $\gamma_a\,p_a$ | *Proportion Significant* | |
			With Divergence	*Without Divergence*
10		0.001	0.66	0.09
50		0.005	1	0.13
100	0.0001	0.01	1	0.83
500		0.05	1	0.58
1,000		0.1	1	0.97
10		0.01	1	0.27
50		0.05	1	1
100	0.001	0.1	1	1
500		0.5	1	1
1,000		1	1	1
10		0.1	1	1
50		0.5	1	1
100	0.01	1	1	1
500		5	1	1
1,000		10	1	1

Across simulations, the strength of selection acting on advantageous mutations ranged from γ_a =

10 to γ_a = 1,000. For a given p_a parameter, increasing the strength of selection increased the

observed proportion of adaptive substitutions, α_{Obs} (Figure 1A). This is expected and is due to the

monotonic increasing relationship between fixation probability and the strength of positive

selection first described by Haldane (1927). Additionally, parameter combinations for which $\gamma_a p_a$

were equal had similar proportions of adaptive substitutions, for example compare γ_a = 10 and p_a

= 0.01 to γ_a = 1,000 and p_a = 0.0001 (Figure 1A). This was also expected because the rate of

adaptive substitutions is proportional to $\gamma_a p_a$. In some datasets, particularly when p_a = 0.01 and

advantageous mutations were very strongly selected (i.e. $\gamma_a \geq$ 500), α_{Obs} exceeded 0.75, which is

higher than is typically estimated from empirical data (Galtier, 2016), so these parameter

combinations may not be biologically relevant.

The effects of selection at linked sites varied across simulated datasets. The DFE for deleterious

mutations was kept constant across simulations, so the extent of background selection should be

fairly similar across all parameter sets and thus variation in π_S/π_0 reflects the effects of selective

sweeps. I found that selection at linked sites reduced synonymous site diversity below the

expected value of 0.01 in all simulations (Figure 1B). Increasing the fitness effects or frequency of

advantageous mutations had a strong effect on genetic diversity at synonymous sites, as shown by

π_S/π_0 in Figure 1B. Unlike α_{Obs}, however, π_S/π_0 responded differently to parameter combinations

that had the same $\gamma_a p_a$ (Figure 1). Finally, differences in p_a explained most of the variation in the

number of segregating advantageous mutations ($S_{Adv.}$) across simulated datasets, but $S_{Adv.}$ also

increased with the strength of positive selection (Figure 1C). On the basis of these results, it is

clear that there will be lower power to estimate positive selection on the basis of standing

variation when advantageous mutations are rare (i.e. p_a = 0.0001) than when they are

comparatively frequent (i.e. p_a = 0.01).

273
274
275 **Figure 1** Population genetic summary statistics collated across all simulated genes. α_{Obs} is the
276 observed proportion of substitutions fixed by positive selection. π_S/π_0 is genetic diversity relative
277 to neutral expectation, in the absence of selection, this takes an expected value of 1.0. $S_{Adv.}$ is the
278 number of segregating nonsynonymous sites in the simulated dataset.
279

Analysis of the unfolded site frequency spectrum

Figure 2 shows the observed (bars) and expected (lines) distribution of derived allele frequencies

for beneficial mutations segregating in simulated populations. The three panels of Figure 2

correspond to three parameter combinations (γ_a = 1,000 and p_a = 0.0001, γ_a = 100 and p_a = 0.001

and γ_a = 10 and p_a = 0.01). The three parameter combinations all resulted in similar α_{Obs}, but

different π_S/π_0 and $S_{Adv.}$ (Figure 1). The lines in each of the panels of Figure 2 show the analytical

expectation for the uSFS of advantageous mutations calculated using Equation 2 from Tataru et

al., (2017). The analytical expectation closely matches the observed data for all three parameter

combinations (Figure 2). However, for a given value of p_a, the analytical expectation for models

with increasing fitness effects were very similar, which likely makes it difficult to distinguish

advantageous mutation parameters on the basis of polymorphism alone (Figure 2). For the three

parameter sets shown in Figure 2, the overall contribution that advantageous alleles make to the

uSFS for nonsynonymous sites is small relative to deleterious ones (Figure S1). Accurate estimation

of positive selection parameters from the uSFS requires that the distribution of advantageous

alleles can be distinguished from deleterious variants, so when p_a is small it seems likely that uSFS

analyses will be unable to easily distinguish competing models.

Figure 2 The uSFS for advantageous mutations under different combinations of positive selection
parameters. The three bar charts show the observed uSFS under positive selection parameters
that yield similar α. The lines in each panel show the expected frequency spectra for different
strengths of beneficial mutations and were obtained using Equation 2 from Tataru et al., (2017).
301

When analysing a particular uSFS dataset in *polyDFE*, I either modelled the full DFE (i.e. a gamma

distribution of deleterious mutations and a discrete class of advantageous mutational effects), or

just a gamma DFE for harmful mutations (dDFE). I compared the fit of the two models using

likelihood ratio tests, which tested the null hypothesis that the fit of a full DFE model is similar to

that of a model containing only deleterious mutations. For each of the combinations of positive

selection parameters shown in Table 2, I ran *polyDFE* on uSFS data from 100 bootstrap replicates.

When modelling the full uSFS (i.e. with divergence), *polyDFE* identified models containing positive

selection consistently for all but one (p_a = 0.0001 and γ_a = 10) of the parameter combinations

tested (Table 2). When the DFE was inferred from polymorphism data alone (i.e. without

divergence), models containing positive selection were identified less often, particularly when

beneficial mutations were rare (p_a = 0.0001; Table 2).

314
315 **Figure 3** Estimates of the parameters of advantageous mutations and the proportion of adaptive
316 substitutions they imply from simulated datasets. A) γ_a is the inferred selective effect of a new
317 advantageous mutation; B) p_a is the proportion of new mutations that are beneficial; C) α_{DFE} is the
318 proportion of adaptive substitutions expected under the inferred DFE, the dashed lines indicate
319 $\alpha_{Obs,}$ the proportion of adaptive substitutions observed in the simulated datasets.
320

Figures 3A and 3B show the parameters of positive selection estimated by analysis of uSFS from

simulated datasets. I found that when simulated beneficial mutations were mildly advantageous

(γ_a = 10) but relatively frequent (p_a = 0.01), both γ_a and p_a were estimated accurately regardless of

whether divergence was modelled or not (Figures 3A-B). This finding is consistent with both

Schneider et al., (2011) and Tataru et al., (2017). When p_a = 0.01 and γ_a > 10, the analysis of the

uSFS with or without divergence yielded very similar parameter estimates, but in both cases, the

strength of positive selection seemed to be positively correlated with the estimated p_a (Figure 3).

In all cases, when beneficial mutations had $\gamma_a \geq 50$, neither γ_a nor p_a were accurately estimated

(Figure 3).

Tataru et al., (2017) pointed out that, if one had an estimate of the full DFE (i.e. with divergence),

the proportion of adaptive substitutions could be obtained by taking the ratio of the fixation

probability for a new beneficial mutation over the fixation probability for a random mutation

integrating over the full DFE (Equation 10; Tataru et al., 2017). The proportion of adaptive

substitutions obtained in this way is denoted α_{DFE}. When modelling the full uSFS, α_{DFE} was

estimated with high accuracy, but with a slight upward bias (Figure 3C). When the DFE was

inferred without divergence α_{DFE} was underestimated when beneficial mutations were strongly

selected and rare (Figure 3).

In the presence of infrequent, strongly beneficial mutations the parameters of the DFE for

deleterious mutations estimated by *polyDFE* were very accurate (Figure S1). Estimates of the DFE

for harmful mutations were less accurate when beneficial mutations occurred with $p_a \geq 0.001$ and

$\gamma_a \geq 100$. This is presumably because in such cases recurrent selective sweeps eliminate a large

amount of neutral diversity and distort the distribution of standing genetic variation at

nonsynonymous sites. However, as stated above, the parameter range where the DFE for harmful

mutations was poorly estimated in this study may not be biologically relevant.

351   Model Identifiability

353
354   **Figure 4** The likelihood surface for the γ_a and p_a parameters for three simulated datasets. Hue
355   indicates differences in log likelihood between a particular parameter combination and the best-
356   fitting model. Best fitting models are indicated by red points and the true parameters are given
357   above the plots and indicated by the white plus signs on the likelihood surface. The relation $\gamma_a p_a$ =
358   0.1 is shown as a turquoise line and is constant across the three datasets shown.
359

It is very difficult to tease apart the parameters of positive selection from the uSFS by maximum

likelihood. Figure 4 shows the likelihood surface for three sets of positive selection parameters.

The three parameter sets all satisfy the condition that $\gamma_a p_a$ = 0.1. The proportion of adaptive

substitutions is largely determined by the product $\gamma_a p_a$ (Kimura & Ohta, 1971) and, as expected,

the three parameter combinations shown in Figure 4 all exhibit a similar α_{Obs} (Figure 1A). However,

the extent by which neutral genetic diversity is reduced and the number of segregating

advantageous mutations differ substantially across the three parameter combinations (Figure 1).

The top row of panels in Figure 4 shows that when modelling the full uSFS, the likelihood surface

closely tracks the relation $\gamma_a p_a$ = 0.1. Focussing on the top panel in Figure 4A, the maximum

likelihood estimates (MLEs) of the positive selection parameters (the red dot) are far from the true

parameter values (indicated by the plus sign), but the MLEs obtained satisfy $\gamma_a p_a$ = 0.1. The ridge

in the likelihood surface was described in both Schneider et al., (2011) and Tataru et al., (2017). It

comes about because between-species divergence carries information about α, and α is

proportional to $\gamma_a p_a$.

Inferring the parameters of the DFE from polymorphism alone avoids the assumption of an

invariant DFE, but when doing so it may be difficult to distinguish competing models. Indeed,

across the three parameter combinations shown, values close to the truth were only obtained

from simulated data when γ_a = 10 and p_a = 0.01 (bottom panel Figure 4C). In the case of γ_a = 1000

and p_a = 0.0001, the likelihood surface about the true parameters was very flat (Figure 4A).

Increasing the p_a parameter increased likelihood for all strengths of selection, so that the MLEs

shown in Figure 4A are simply the values with the highest p_a in the range tested (the vertical red

line in Figure 4A). When γ_a = 100 and p_a = 0.001, the likelihood surface about the estimates was

steep, but the selection parameters identified by maximum likelihood were incorrect (Figure 4B).

Discussion

In this study, I analysed simulated datasets modelling a range of positive selection parameter

combinations. I found that estimates of positive selection parameters obtained by analysis of the

uSFS were only accurate when beneficial mutations had $\gamma_a \leq$ 50, under stronger selection the

individual parameters of positive selection were not accurately estimated (Figure 3). This is not

particularly surprising and is consistent with verbal arguments made in published studies (Booker

& Keightley, 2018; Campos et al., 2017). However, it is troubling that when beneficial mutations

are strongly selected and rare, the uSFS often contained significant signal of positive selection but

erroneous parameter estimates were obtained. If one were to analyse an empirical dataset and

estimate parameters of positive selection of the order $\gamma_a \sim 10$ and $p_a \sim 0.01$, it would be difficult to

know whether those were reflective of the true underlying parameters or an artefact of strong

selection. Furthermore, α is accurately estimated when modelling the full uSFS with divergence,

but doing so assumes that the DFE has remained invariant over time and that assumption may be

problematic. Estimates of α obtained by analysing polymorphism data without divergence avoid

this assumption, but may underestimate the true values if beneficial mutations are strongly

selected and rare.

On the basis of this study, it seems that researchers should treat parameters of positive selection

obtained by analysis of the uSFS with caution. The expected uSFS for advantageous mutations is

very similar when DFE models share the same p_a parameter, and in such cases differing models

can only be distinguished by the density of high frequency derived variants (Figure 2). Polarization

error when estimating the uSFS can generate an excess in the number of high frequency variants

(Keightley & Jackson, 2018), so may generate a spurious signal of strong positive selection.

Analysis methods have been proposed which attempt to estimate the rate of polarisation error

when modelling the uSFS (Barton & Zeng, 2018; Tataru et al., 2017), but further study is required

to determine whether such methods reduce the signal of positive selection in uSFS-based

analyses. Parameters of the DFE for harmful mutations were robustly estimated across the

simulated datasets.

Estimates of α based on analysis of the uSFS may be biased when beneficial mutations are strongly

selected and infrequent. Calculating α using the rearranged MK-test makes the problematic

assumption that the DFE has remained invariant in the time since the focal species began to

diverge from the outgroup (Tataru et al., 2017). However, Tataru et al., (2017) pointed out that

one can avoid that assumption if α_{DFE} is calculated from a DFE estimated without divergence data.

In this study, estimates of α_{DFE} obtained when the full uSFS was analysed were very precise, but

with a slight upward bias (Figure 3). When simulated beneficial mutations were strongly selected

and rare, the parameters inferred using polymorphism data alone (i.e. without divergence) yielded

spurious estimates of α_{DFE} (Figure 3). When analysing datasets from real populations, α_{DFE} may not

capture the contribution that strongly beneficial mutations make to molecular evolution. This may

make it difficult to contrast α_{DFE} between species with large differences in population size, because

the number of segregating advantageous mutations and thus ability to accurately estimate

selection parameters will depend on N_e.

The simulations I performed in this study generated the ideal dataset for estimating parameters of

selection from the uSFS. I simulated datasets containing 21Mbp of sites in which genotypes and

whether sites were selected or not was unambiguously known. When analysing real data this is

not the case and researchers often have to filter a large proportion of sites out of their analyses or

choose to analyse a subset of genes that have orthology with outgroups or biological properties of

interest. In addition, the populations I simulated were randomly mating and had constant sizes

over time. The results I present in this study suggest that even with perfect knowledge of a

population that adheres to the assumptions of a Wright-Fisher model, it is inherently difficult to

infer the parameters of strongly beneficial mutations from the uSFS, particularly so when

beneficial mutation occur infrequently.

Estimating parameters of positive selection from the uSFS versus

estimates from patterns of diversity

As discussed above, studies based on analysis of the uSFS and those based on selective sweep

models have yielded vastly different estimates of the parameters of positive selection. Patterns of

neutral genetic diversity in both humans and wild mice cannot be explained by the effects of

background selection alone, and in both species it has been suggested that strongly beneficial

mutations are required to explain the observed patterns (Booker & Keightley, 2018; Nam et al.,

2017). In the case of wild house mice, positive selection parameters obtained by analysis of the

uSFS do not explain dips in nucleotide diversity around functional elements (Booker & Keightley,

2018). Recently, Castellano et al. (2019) analysed the uSFS for nonsynonymous sites in great ape

species but did not find significant evidence for positive selection. In their dataset, Castellano et al.

(2019) had 8 haploid genome sequences for each of great ape species they analysed, and they

argued that they were underpowered to detect positive selection on the basis of the uSFS.

However, the results given in this study suggest that even with larger samples (recall that I

analysed 40 haploid genomes from my simulated populations), parameters of strongly beneficial

mutations are very difficult to resolve by analysis of the uSFS.

In this study, I modelled beneficial mutations using a discrete class of selection coefficients when,

in reality, there is likely a continuous distribution of fitness effects. Indeed, studies in both humans

and *D. melanogaster* have found evidence for a bimodal distribution containing both strongly and

weakly beneficial mutations contributing to adaptive evolution using methods which incorporate

linkage information but do not explicitly estimate selection parameters (Elyashiv et al., 2016;

Uricchio, Petrov, & Enard, 2019). There are currently no methods that estimate the DFE using an

analytical expression for the uSFS expected under the combined effects of BGS and sweeps.

Rather, nuisance parameters or demographic models are used to correct for the contribution that

selection at linked sites may make to the shape of the SFS (Eyre-Walker, Woolfit, & Phelps, 2006;

Galtier, 2016; Tataru et al., 2017). However, as this study shows, the parameters of positive

selection may not be reliably detected when analysing the uSFS alone. A way forward may be in

using approximate Bayesian computation (ABC) or machine-learning approaches to make use of all

of the available data, while not necessitating an expression for the uSFS expected under the

combined effects of BGS, sweeps, population size change and direct selection. An advance in this

direction has recently been made by Uricchio et al., (2019) who developed an ABC method for

estimating α which makes use of the distortions to the uSFS generated by BGS and sweeps. By

applying their method to data from humans, Uricchio et al., (2019) found that α = 0.13 for

nonsynonymous sites, 72% of which was generated by mildly beneficial mutations and 28% by

strongly beneficial mutations. Unfortunately, their method does not provide exact estimates for

positive selection parameters, but indicates that by using computational approaches, progress

could be made in our estimation and understanding of the DFE for both advantageous and

deleterious mutations.

Acknowledgements

479 I wish to extend gratitude to Peter Keightley, Michael Whitlock and Sam Yeaman for valuable

480 advice and mentorship. Thanks to Sam Yeaman and Peter Keightley for comments on the

481 manuscript. Thanks to Paula Tataru and Thomas Bataillon for help with *polyDFE*.

References

Bank, C., Hietpas, R. T., Wong, A., Bolon, D. N., & Jensen, J. D. (2014). A bayesian MCMC approach

to assess the complete distribution of fitness effects of new mutations: uncovering the

potential for adaptive walks in challenging environments. *Genetics*, *196*(3), 841–852.

https://doi.org/10.1534/genetics.113.156190

Barton, H. J., & Zeng, K. (2018). New methods for inferring the distribution of fitness effects for

INDELs and SNPs. *Molecular Biology and Evolution*, *35*(6), 1536–1546.

https://doi.org/10.1093/molbev/msy054

Barton, N. H. (2000). Genetic hitchhiking. *Philos Trans R Soc Lond B Biol Sci*, *355*(1403), 1553–1562.

https://doi.org/10.1098/rstb.2000.0716

Böndel, K. B., Kraemer, S. A., Samuels, T., McClean, D., Lachapelle, J., Ness, R. W., … Keightley, P. D.

(2019). Inferring the distribution of fitness effects of spontaneous mutations in

Chlamydomonas reinhardtii. *PLOS Biology*, *17*(6), e3000192.

https://doi.org/10.1371/journal.pbio.3000192

Booker, T. R., & Keightley, P. D. (2018). Understanding the Factors That Shape Patterns of

Nucleotide Diversity in the House Mouse Genome. *Mol. Biol. Evol.*, *35*(12), 2971–2988.

Boyko, A. R., Williamson, S. H., Indap, A. R., Degenhardt, J. D., Hernandez, R. D., Lohmueller, K. E.,

… Bustamante, C. D. (2008). Assessing the evolutionary impact of amino acid mutations in the

human genome. *PLoS Genet*, *4*(5), e1000083. https://doi.org/10.1371/journal.pgen.1000083

Campos, J L, Zhao, L., & Charlesworth, B. (2017). Estimating the parameters of background

selection and selective sweeps in Drosophila in the presence of gene conversion. *Proc Natl*

Acad Sci, *114*(24), E4762–E4771. https://doi.org/10.1073/pnas.1619434114

10.5061/dryad.vs264)

Campos, José Luis, & Charlesworth, B. (2019). The effects on neutral variability of recurrent

selective sweeps and background selection. *Genetics*, *212*(1), 287–303.

https://doi.org/10.1534/genetics.119.301951

Castellano, D., Macià, M. C., Tataru, P., Bataillon, T., & Munch, K. (2019). Comparison of the Full

Distribution of Fitness Effects of New Amino Acid Mutations Across Great Apes. *Genetics*,

genetics.302494.2019. https://doi.org/10.1534/genetics.119.302494

Elyashiv, E., Sattath, S., Hu, T. T., Strutsovsky, A., McVicker, G., Andolfatto, P., … Sella, G. (2016). A

genomic map of the effects of linked selection in Drosophila. *PLoS Genet*, *12*(8), e1006130.

https://doi.org/10.1371/journal.pgen.1006130

Eyre-Walker, A., & Keightley, P. D. (2007). The distribution of fitness effects of new mutations. *Nat*

Rev Genet, *8*(8), 610–618. https://doi.org/10.1038/nrg2146

Eyre-Walker, A., & Keightley, P. D. (2009). Estimating the rate of adaptive molecular evolution in

the presence of slightly deleterious mutations and population size change. *Mol Biol Evol*,

26(9), 2097–2108. https://doi.org/10.1093/molbev/msp119

Eyre-Walker, A., Woolfit, M., & Phelps, T. (2006). The distribution of fitness effects of new

deleterious amino acid mutations in humans. *Genetics*, *173*(2), 891–900.

https://doi.org/10.1534/genetics.106.057570

Galtier, N. (2016). Adaptive Protein Evolution in Animals and the Effective Population Size

Hypothesis. *PLoS Genet.*, *12*(1), e1005774.

Haldane, J. B. S. (1927). A Mathematical Theory of Natural and Artificial Selection, Part V: Selection

and Mutation. *Mathematical Proceedings of the Cambridge Philosophical Society*, *23*(7), 838–

844. https://doi.org/10.1017/S0305004100015644

Haller, B. C., & Messer, P. W. (2019). {SLiM} 3: Forward Genetic Simulations Beyond the {Wright--

Fisher} Model. *Mol. Biol. Evol.*, *36*(3), 632–637.

Hill, W. G. (2010, January 12). Understanding and using quantitative genetic variation.

Philosophical Transactions of the Royal Society B: Biological Sciences, Vol. 365, pp. 73–85.

https://doi.org/10.1098/rstb.2009.0203

Keightley, P D, Campos, J. L., Booker, T. R., & Charlesworth, B. (2016). Inferring the frequency

spectrum of derived variants to quantify adaptive molecular evolution in protein-coding

genes of Drosophila melanogaster. *Genetics*, *203*(2), 975–984.

https://doi.org/10.1534/genetics.116.188102

Keightley, P D, & Eyre-Walker, A. (2007). Joint inference of the distribution of fitness effects of

deleterious mutations and population demography based on nucleotide polymorphism

frequencies. *Genetics*, *177*(4), 2251–2261. https://doi.org/10.1534/genetics.107.080663

Keightley, Peter D, & Jackson, B. C. (2018). Inferring the Probability of the Derived vs. the Ancestral

Allelic State at a Polymorphic Site. *Genetics*, *209*(3), 897–906.

Kimura, M., & Ohta, T. (1971). *Theoretical aspects of population genetics*. Princeton Univ. Press.

Laenen, B., Tedder, A., Nowak, M. D., Toräng, P., Wunder, J., Wötzel, S., … Slotte, T. (2018).

Demography and mating system shape the genome-wide impact of purifying selection in

Arabis alpina. *Proceedings of the National Academy of Sciences of the United States of*

America, *115*(4), 816–821. https://doi.org/10.1073/pnas.1707492115

Loewe, L., & Charlesworth, B. (2006). Inferring the distribution of mutational effects on fitness in

Drosophila. *Biol Lett*, *2*(3), 426–430. https://doi.org/10.1098/rsbl.2006.0481

McDonald, J. M., & Kreitman, M. (1991). Adaptive protein evolution at the Adh locus in

Drosophila. *Nature*, *351*.

Nam, K., Munch, K., Mailund, T., Nater, A., Greminger, M. P., Krützen, M., … Schierup, M. H.

(2017). Evidence that the rate of strong selective sweeps increases with population size in the

great apes. *Proceedings of the National Academy of Sciences of the United States of America*,

114(7), 1613–1618. https://doi.org/10.1073/pnas.1605660114

Orr, H. A., & Unckless, R. L. (2014). The Population Genetics of Evolutionary Rescue. *PLoS Genetics*,

10(8). https://doi.org/10.1371/journal.pgen.1004551

Otto, S. P. (2009). The evolutionary enigma of sex. *American Naturalist*, *174*(SUPPL. 1).

https://doi.org/10.1086/599084

Schneider, A., Charlesworth, B., Eyre-Walker, A., & Keightley, P. D. (2011). A method for inferring

the rate of occurrence and fitness effects of advantageous mutations. *Genetics*, *189*(4), 1427–

1437. https://doi.org/10.1534/genetics.111.131730

Smith, N. G. C., & Eyre-Walker, A. (2002). Adaptive protein evolution in Drosophila. *Nature*,

415(6875), 1022–1024. https://doi.org/10.1038/4151022a

Tataru, P., & Bataillon, T. (2019). polyDFEv2.0: testing for invariance of the distribution of fitness

effects within and across species. *Bioinformatics*, *35*(16), 2868–2869.

https://doi.org/10.1093/bioinformatics/bty1060

Tataru, P., Mollion, M., Glemin, S., & Bataillon, T. (2017). Inference of distribution of fitness effects

and proportion of adaptive substitutions from polymorphism data. *Genetics*, *207*(3), 1103–

1119. https://doi.org/10.1534/genetics.117.300323

Uricchio, L. H., Petrov, D. A., & Enard, D. (2019). Exploiting selection at linked sites to infer the rate

and strength of adaptation. *Nat Ecol Evol*, *3*(6), 977–984.

Williamson, R. J., Josephs, E. B., Platts, A. E., Hazzouri, K. M., Haudry, A., Blanchette, M., & Wright,

S. I. (2014). Evidence for widespread positive and negative selection in coding and conserved

noncoding regions of Capsella grandiflora. *PLoS Genetics*, *10*(9), e1004622.

https://doi.org/10.1371/journal.pgen.1004622

Wright, S. (1937). The Distribution of Gene Frequencies in Populations. *Proceedings of the*

National Academy of Sciences, *23*(6), 307–320. https://doi.org/10.1073/pnas.23.6.307

Supplementary Material

Figure S1 The observed uSFS for nonsynonymous sites for three sets of positive selection
parameters. The distribution of deleterious mutations is shown in orange and the distribution of
advantageous mutations is shown in blue. For the purposes of visualising the data on a log scale,
the number of segregating sites is shown +1.

597 **Figure S2** Parameter estimates for the DFE for deleterious mutations obtained from simulated
598 datasets. A) the mean effect of a deleterious mutation and b) the shape parameter of the gamma
599 distribution. Error bars indicate the 95% range of 100 bootstrap replicates.