

End-to-end multitask learning, from protein language to protein features without alignments

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Abstract

Correctly predicting features of protein structure and function from amino acid sequence alone remains a supreme challenge for computational biology. For almost three decades, state-of-the-art approaches combined machine learning and evolutionary information from multiple sequence alignments. Exponentially growing sequence databases make it infeasible to gather evolutionary information for entire microbiomes or meta-proteomics. On top, for many important proteins (e.g. *dark proteome* and *intrinsically disordered proteins*) evolutionary information remains limited. Here, we introduced a novel approach combining recent advances of Language Models (LMs) with multi-task learning to successfully predict aspects of protein structure (secondary structure) and function (cellular component or subcellular localization) without using any evolutionary information from alignments. Our approach fused self-supervised pre-training LMs on an unlabeled big dataset (UniRef50, corresponding to 9.6 billion words) with supervised training on labelled high-quality data in one single end-to-end network. We provided a proof-of-principle for the novel concept through the semi-successful per-residue prediction of protein secondary structure and through per-protein predictions of localization (Q10=69%) and the distinction between integral membrane and water-soluble proteins (Q2=89%). Although these results did not reach the levels obtained by the best available methods using evolutionary information from alignments, these less accurate multi-task predictions have the advantage of speed: they are 300-3000 times faster (where HHblits needs 30-300 seconds on average, our method needed 0.045 seconds). These new results push the boundaries of predictability towards grayer and darker areas of the protein space, allowing to make reliable predictions for proteins which were not accessible by previous methods. On top, our method remains scalable as it removes the necessity to search sequence databases for evolutionary related proteins.

Key words: Machine Learning; Language Modelling; Semi-Supervised Learning; Multi-Task Learning; Protein Secondary Structure Prediction; Protein subcellular-localization Prediction.

Abbreviations used: **1D**, one-dimensional – information representable in a string such as secondary structure or solvent accessibility; **3D**, three-dimensional; **3D structure**, three-dimensional coordinates of protein structure; **DBMTL**, Deep Biology Multi-Task Learning; **NLP**, Natural Language Processing; **PIDE**, percentage of pairwise identical residues;

Introduction

Successful combination of evolutionary information and artificial intelligence. Predicting structural and functional aspects of proteins based on their amino acid sequence has been one of the most challenging problems for computational biology. Researchers have been working on this problem for nearly five decades in order to bridge the sequence-structure/function gap, i.e. the gap between 180M proteins of known sequence (UniProt (Consortium, 2018)) and about 560K proteins with experimental annotations about function (SwissProt (Boutet, et al., 2016)) or 150K with high-resolution experimental structures (PDB (Berman, et al., 2000)). The biggest single improvement in prediction performance was achieved over two decades ago through the combination of machine learning (ML) and evolutionary information, i.e. the profiles extracted from Multiple Sequence Alignments (MSA) of related proteins as input feature for protein secondary structure prediction (Rost and Sander, 1993; Rost and Sander, 1993; Rost and Sander, 1994). Evolutionary information was quickly adopted by the field (Frishman and Argos, 1995; Jones, 1999; Mehta, et al., 1995) and has become the *de facto* standard for encoding protein sequences for most machine learning applications, including the prediction of transmembrane helices (Rost, et al., 1995), solvent accessibility (Pollastri, et al., 2002; Rost and Sander, 1994), protein flexibility (Capriotti, et al., 2005; Radivojac, et al., 2004), inter-residue contacts (Hayat, et al., 2015), protein-protein interactions (Hamp and Rost, 2015; Zhang, et al., 2012), and subcellular localization (Casadio, et al., 2008; Goldberg, et al., 2014; Nair and Rost, 2003).

With protein sequence data exploding exponentially, evolutionary information becomes even more valuable (Jones, 1999; Rost, 2001). However, even today's fastest solutions, HHBlits3 (Steinegger, et al., 2019) and MMSeqs2 (Steinegger and Söding, 2017), can hardly cope with all 2.5B sequences in BFD (Steinegger, et al., 2019). On top, evolutionary information is more tricky to obtain for proteins that are difficult to align such as intrinsically disordered proteins (IDPs), or proteins from the Dark Proteome (Perdigao, et al., 2015).

Mining the wealth of unlabeled bio-data through transfer learning. What if we could replace the search for evolutionary related proteins by semi-supervised multi-task (MT) learning? Such an approach would build upon the recent success of language models (LMs) (Devlin, et al., 2018; Howard and Ruder, 2018; Peters, et al., 2018) applied to protein sequences (Heinzinger, et al., 2019; Rives, et al., 2019). Language models are trained on large unlabeled text-corpora to predict the most probable next word in a sentence, given all previous words in this sentence (auto-regression). As only a sequence of words (sentence) is needed to train, such approaches are referred to as *self-supervision*, i.e. the learning of syntax and semantics through data without requiring expert knowledge. The same idea can be adopted to computational biology by considering single amino acids as words and their sequences as sentences. This simple transfer unleashes the power of big data sets easily outgrowing text-corpora used in NLP such as Wikipedia by orders of magnitude (Heinzinger, et al., 2019; Howard and Ruder, 2018; Rives, et al., 2019).

So far applications of LMs to protein sequences have split the training into two phases (Alley, et al., 2019; Heinzinger, et al., 2019; Howard and Ruder, 2018): (1) self-supervised pre-training on unlabeled data learns vector representations (embeddings), which is followed by (2) supervised training of a second network on labeled data using the embeddings as input. Although transfer learning succeeds to an amazing extent to extract some principles relevant for the understanding the language of life, databases of protein sequences do not capture any explicit information about the molecular constraints shaping proteins in evolution. Not surprisingly, the simple two-step solution self-supervised transfer-learning followed by supervised deep-learning fails to beat the best methods using evolutionary information. For NLP, multi-task (MT) learning merges the two steps in an end-to-end trainable machine, e.g. through transformer models currently reaching the state-of-the-art in NLP (Dai, et al., 2019; Devlin, et al., 2018; Radford, et al., 2019).

1
2 Here we hypothesized that multi-task (MT) learning into one end-to-end network might enable
3 to fine-tune NLP models to particular supervised problems in computational biology. More
4 specifically, we trained an MT transformer model on five different tasks: (1) The language
5 modeling task was trained on 35M sequences from UniRef50 (Suzek, et al., 2015); (2) The
6 per-residue (word-level) task was the prediction of secondary structure in three (and eight)
7 states derived from DSSP (Kabsch and Sander, 1983); (3) The per-protein (sentence-level)
8 tasks included the predictions of protein subcellular localization in ten classes and a binary
9 classification into membrane-bound and soluble proteins. In order to simplify the comparability
10 of results between different approaches, we used two datasets from recent state-of-the-art
11 publications (Almagro Armenteros, et al., 2017; Klausen, et al., 2019).
12
13

14 **Materials & Methods**

15 **Data**

16 **Language modeling.** The self-supervised, language modelling task was trained on UniRef50
17 a subset of UniProt (Consortium, 2018) clustered at 50% pairwise sequence identity (PIDE).
18 The database contained 33M protein sequences with 9,577,889,953 residues and a
19 vocabulary of 25 words: the standard 20 and two rare amino acids (U and O) along with three
20 letters for ambiguous (B, Z) or unknown amino acids (X). Each protein was treated as a
21 sentence and each amino acid was interpreted as a single word. The model was trained using
22 99.9% of the data, while randomly keeping 0.01% (~33k) of the proteins for validation. In this
23 case, since the goal is self-supervision, homology doesn't play a major role and thus
24 train/validation splits could be drawn at random. The maximal model length was set to 1024;
25 for proteins longer than 1024 residues only the first 1024 residues were used.
26

27 **Per-residue: secondary structure.** Using a data set of protein secondary structure
28 previously published (Klausen, et al., 2019) simplified comparisons of this task to state-of-the-
29 art prediction methods, including: NetSurfP2.0 (Klausen, et al., 2019), Spider3 (Heffernan, et
30 al., 2017), RaptorX (Wang, et al., 2016) and JPred4 (Drozdetskiy, et al., 2015). For training,
31 10,837 sequence-unique (at 25% PIDE) proteins with high-quality 3D structures (<2.5Å or
32 0.25nm) were collected by the PISCES server (Wang and Dunbrack Jr, 2003) from the Protein
33 Data Bank – PDB (Berman, et al., 2000). DSSP (Kabsch and Sander, 1983) assigned
34 secondary structure; residues without atomic coordinates (*REMARK-465*) were removed. The
35 seven DSSP states (+ one for unknown) were mapped to three states as in previous works
36 (Rost and Sander, 1993): [G,H,I] → H: helix, [B,E] → E: strand, all others to O: other). Since
37 NetSurfP-2.0 only published pre-computed profiles, protein sequences were extracted through
38 SIFTS (Velankar, et al., 2012). During this mapping step, 56 proteins were removed from the
39 training and three from the test set due to differences in lengths between SIFTS and NetSurfP-
40 2.0 (two from CB513 (Cuff and Barton, 1999); one from CASP12 (Abriata, et al., 2018); none
41 from TS115 (Yang, et al., 2016)). Proteins longer than 512 residues were also removed due
42 to constraints of our transformer models. Three test sets (also referred to as validation sets)
43 were distinguished: TS115: 115 proteins from high-quality structures (<3Å) released after 2015
44 (and at most 30% PIDE to any protein of known structure in the PDB at the time); CB513: 513
45 non-redundant sequences compiled 20 years ago (511 after SIFTS mapping); CASP12: 21
46 proteins taken from the CASP12 free-modelling targets (20 after SIFTS mapping; all 21 fulfilled
47 a stricter criterion toward non-redundancy than the two other sets; non-redundant with respect
48 to all 3D structures known until May 2018 and all their relatives). The NetSurfP-2.0 authors
49 removed all proteins from training with >25% PIDE to any protein in the test sets.
50

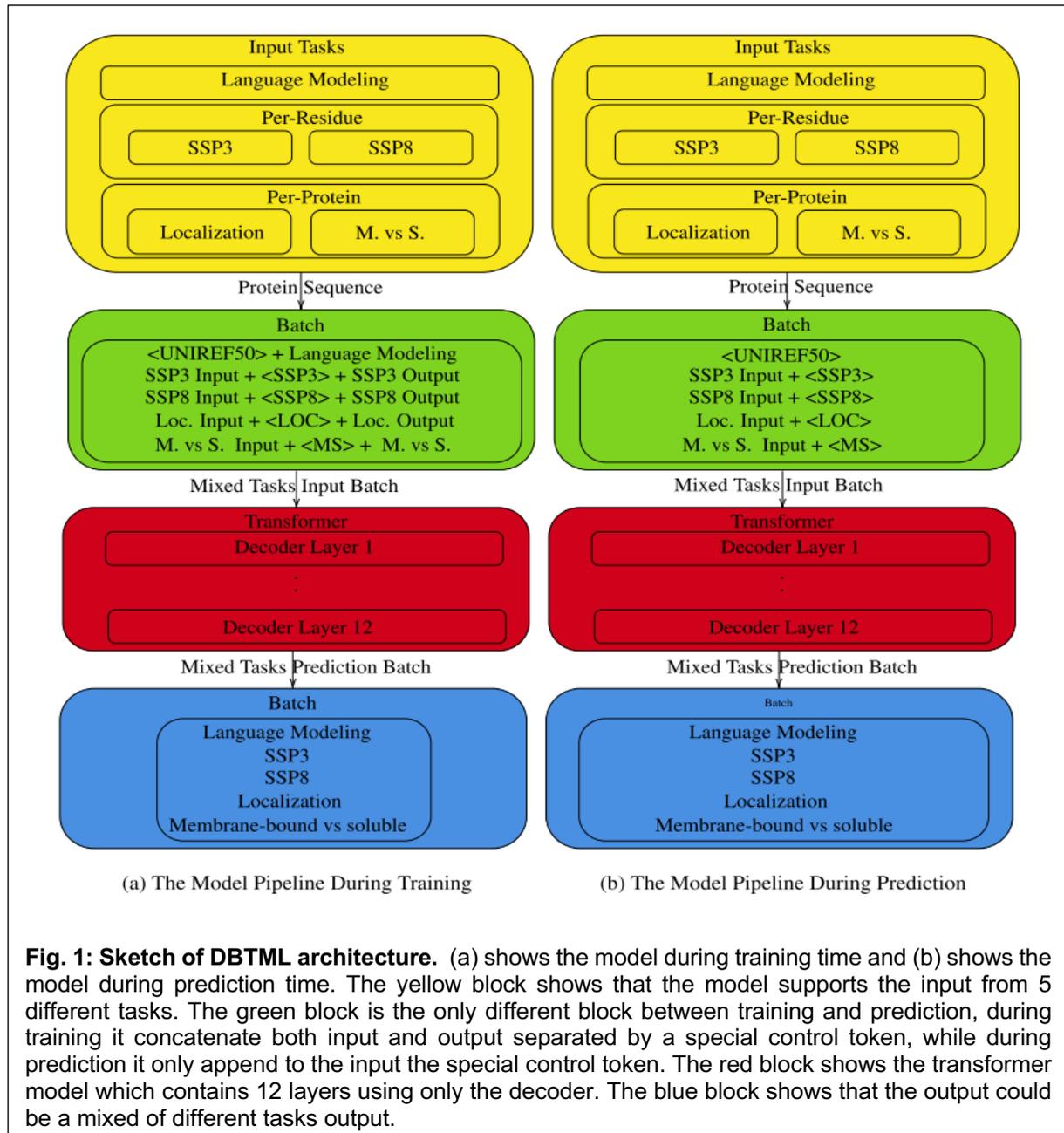
1 **Per-protein data: subcellular localization and membrane/soluble.** The prediction of
2 subcellular localization (also referred to as *location* or *cellular component*) was trained and
3 evaluated through data from the authors of DeepLoc (Almagro Armenteros, et al., 2017), one
4 of the state-of-the-art prediction methods for this task. The performance of several other
5 methods was also evaluated on this set, namely: LocTree2 (Goldberg, et al., 2012), MultiLoc2
6 (Blum, et al., 2009), SherLoc2 (Briesemeister, et al., 2009), CELLO (Yu, et al., 2006), iLoc-
7 Euk (Chou, et al., 2011), WoLF PSORT (Horton, et al., 2007) and YLoc (Briesemeister, et al.,
8 2010). This data set pools proteins with experimental annotation (code: ECO:0000269) from
9 UniProtKB/Swiss-Prot (release: 2016_04, (Consortium, 2018)). To facilitate training and
10 evaluation, the authors of DeepLoc mapped the plain localization annotations to ten classes,
11 removing all proteins with multiple annotations. Additionally, all proteins were labelled as
12 *water-soluble* or *membrane-bound* (some with ambiguous annotations as *unknown*). The
13 resulting 13,858 proteins clustered into 8,464 representatives with PSI-CD-HIT (Fu, et al.,
14 2012; Li and Godzik, 2006) (v4.0; at 30% PIDE or E-val<10⁻⁶; for alignments covering>80%
15 of the shorter protein). The remaining proteins were split into training and testing by using the
16 same proteins for testing as DeepLoc. Similar to DeepLoc, proteins longer than the maximum
17 processed by the method (1000 for DeepLoc, 1024 for the method presented here) remained
18 in the data set and were simply cut (keeping the first and the last 512 residues).

19
20 **Evaluation measures.** For simplicity of comparison, the evaluation measures followed
21 previous publications (Almagro Armenteros, et al., 2017; Klausen, et al., 2019), namely three-
22 state per-residue accuracy (Q3 (Rost and Sander, 1993)) as the percentage of residues
23 correctly predicted in three secondary structure states (helix, strand, other), the corresponding
24 eight-state per-residue accuracy (Q8), the two-state per-protein accuracy (Q2) to describe the
25 percentage of proteins correctly predicted as membrane-bound/water-soluble, and the ten-
26 state per-protein accuracy (Q10) for the ten classes of localization. All numbers reported
27 constituted averages over all proteins in the final test sets.

29 Deep biology multi-task learning (DBMTL)

30 **Concept.** Thus far, successful applications of NLP techniques in computational biology (Alley,
31 et al., 2019; Heinzinger, et al., 2019; Howard and Ruder, 2018; Rives, et al., 2019) used a two-
32 step approach: Step 1: train a self-supervised model on protein sequences without annotations
33 to learn vector representations (embeddings). Step 2: input embeddings to train supervised
34 models on annotated data (e.g. secondary structure or localization). DBMTL merged the two
35 steps into one single end-to-end model. This was achieved by applying a joint loss consisting
36 of the four supervised tasks ((I) 3- and (II) 8-state secondary structure prediction, (III) 10-state
37 per-protein localization and (IV) 2-state per-protein membrane/globular) and (V) the semi-
38 supervised language modeling task. Thereby, the model could learn biochemical and
39 biophysical properties of amino acids together (I, II & V) with the global properties of the
40 macromolecules they constitute (III & IV). The idea was borrowed from the NLP transformer
41 model (Vaswani, et al., 2017). These models consist of two building blocks, namely an encoder
42 and a decoder. Here, we tested only the first approach (decoder only (Radford, et al., 2019)).

43
44 **Architecture and training.** The model used for DBMTL consists only of the decoder part of
45 the transformer model (Fig. 1a: training, Fig. 1b: prediction/testing). We used 12 attention
46 layers with maximum length of 1024 for inputs and outputs. The number of hidden layers was
47 set to 1024, while the number of heads was set to 16. A dropout rate of 0.2 reduced the risk of
48 over-fitting. For this model, the input differed between training and prediction (Fig. 1). During
49 training, both the input and the output of each task were concatenated inserting a control token
50 between them. During prediction, the model saw only the input followed by the control token.
51 Each task had its own control token allowing the model to learn what to predict next:



1 <UNIREF50> (Language Modelling), <SSP3> and <SSP8> (secondary structure prediction in
2 3 and 8 states), <LOC> (localization), <MS> (membrane-bound vs. water-soluble proteins).

3 The model was trained on 6 Titan GPUs (local batch size: 512; global batch: 3072)
4 using the Adam optimizer with 0.0002 learning rate, warm-up rate of 16k steps, and learning
5 rate decay for 250k steps. The model was trained for a total number of 500K steps.
6

7 Results

8 **Per-residue: secondary structure prediction competitive.** DBMTL did not reach to the
9 level of performance obtained by the top performers using evolutionary information from
10 multiple sequence alignments (MSAs), such as NetSurfP-2.0 (Klausen, et al., 2019) or Spider3
11 (Heffernan, et al., 2017) and RaptorX (Wang, et al., 2016). However, it appeared to reach close

1 and appeared to outperform other methods not using evolutionary information. In particular
 2 DeepSeqVec (Heinzinger, et al., 2019) and *ProtVec* (Asgari and Mofrad, 2015), the latter being
 3 one of the first approaches to successfully adopt NLP solutions (namely Google's Word2vec
 4 (Mikolov, et al., 2013)) to problems in computational biology by extracting rules from languages
 5 that are context-independent. These results were not shown because we discovered a major
 6 problem with how Tensor2Tensor (T2T) (Vaswani, et al., 2018) utilizes the concept of teacher
 7 forcing for per-residue predictions: essentially, what works for the field of NLP or the per-
 8 protein prediction is not applicable. Instead, T2T automatically uses some of the observation
 9 for prediction (Discussion for detail). This problem did not affect the per-protein predictions.

10
 11 **Per-protein performance high but not top.** For both per-protein prediction tasks explored
 12 (localization and membrane-bound/water-soluble globular) DBMTL **did not** reach the top
 13 performance level (Fig. 2ab). For the prediction of localization, the best method DeepLoc
 14 reached Q10=78%, while the approach introduced here (DBMTL) remained nine percentage
 15 points below Q10=69% (Fig. 2a, Table SOM_3). DBMTL was numerically higher than other
 16 popular prediction methods using evolutionary information, namely (sorted by date): WoLF
 17 PSORT (Horton, et al., 2007), CELLO (Yu, et al., 2006), MultiLoc2 (Blum, et al., 2009),
 18 SherLoc2 (Briesemeister, et al., 2009), YLoc (Briesemeister, et al., 2010), iLoc-Euk (Chou, et
 19 al., 2011) and LocTree2 (Goldberg, et al., 2012). However, for localization prediction, the end-
 20 to-end solution DBMTL hardly outperformed the two-step approach realized earlier through
 21 SeqVec (Heinzinger, et al., 2019) embeddings (DeepSeqVec Fig. 2a). In contrast, the
 22 Word2vec-like approach ProtVec (Asgari and Mofrad, 2015) realized in DeepProtVec again
 23 performed much worse (over 20 percentage points: Fig. 2a).

24 The situation was similar for the other per-protein task, namely the binary classification
 25 into membrane-bound and water-soluble, globular proteins. Although for this task, DBMTL
 26 missed the top level only by a smaller margin of three percentage points: Q2(DeepLoc)=92%
 27 vs. Q2(DBMTL)=89% (Fig. 2b, Table SOM_4). Once again, the novel end-to-end solution
 28 realized by DBMTL outperformed other solutions not using evolutionary information, e.g. the
 29 two-stage approach DeepSeqVec and the non-contextualized transfer-learning approach
 30 DeepProtVec by (Fig. 2b).

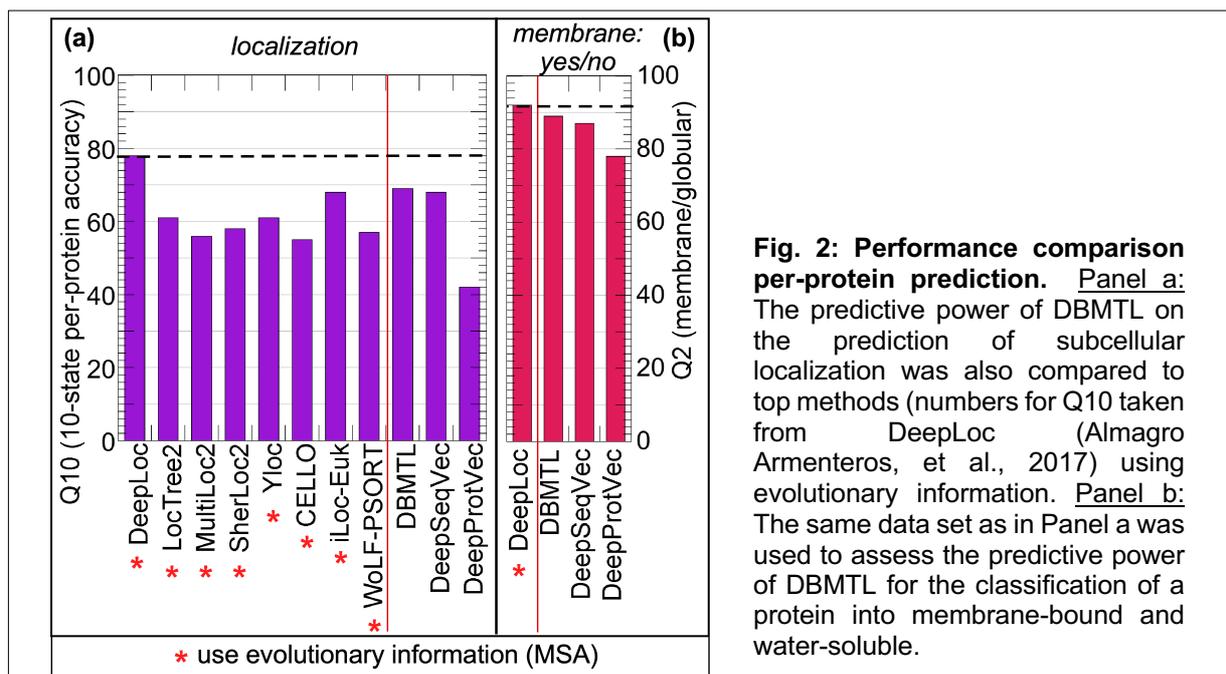


Fig. 2: Performance comparison per-protein prediction. Panel a: The predictive power of DBMTL on the prediction of subcellular localization was also compared to top methods (numbers for Q10 taken from DeepLoc (Almagro Armenteros, et al., 2017) using evolutionary information. Panel b: The same data set as in Panel a was used to assess the predictive power of DBMTL for the classification of a protein into membrane-bound and water-soluble.

1 **Blazingly fast predictions.** One of the main advantages of the transformer model over
2 LSTM-based models such as ELMo (Peters, et al., 2018) is speed reached by processing input
3 tokens in parallel through self-attention mechanisms (Bahdanau, et al., 2014). Using a single
4 1080Ti GPU, the model took ~35 seconds to load, and with a batch size of 32, the prediction
5 for a 1024-residue protein took, on average, 0.033 seconds. This was similar to the LSTM-
6 based transfer-learning model SeqVec (Heinzinger, et al., 2019). In comparison to methods
7 that use evolutionary information and have to first build the MSA, DBMTL reached a speed-up
8 of approximately 110-fold compared to NetSurfP-2.0 using MMSeqs2 for creating alignments.
9
10

11 Discussion

12 **Reporting negative results.** Since the submission of the first version of this work, the authors
13 spent all their resources on making the model openly available for the community to use. After
14 trying to do so using machine learning toolkits (T2T (Vaswani, et al., 2018)), and failing to
15 obtain speedy fixes by the community, the authors decided to re-engineer the underlying deep
16 learning model. During the process of re-engineering the model, the authors discovered a
17 fundamental problem with how the model calculates loss on secondary structure predictions,
18 which undermines the authors' confidence in the results initially reported. Since re-engineering
19 the model with an open system and reproducing all experiments and results is time demanding
20 and in-progress, the authors considered it important to update the initial version of the
21 manuscript by removing results that are no longer sustained, until these can safely be verified
22 or nullified. Additionally, the authors considered it important to update the preprint describing
23 the shortfalls that emerged during re-engineering, so as to support fellow researchers not to
24 commit the same mistakes.
25

26 **Flawed per-residue predictions.** The core of DBMTL consists of the decoder part of a
27 transformer model (Vaswani, et al., 2018; Vaswani, et al., 2017). Essentially, this model applies
28 a stack of self-attention layers (Bahdanau, et al., 2014) followed by a linear transformation to
29 predict the next token in a sequence, given all previous tokens in this sequence (self-
30 supervision). These tokens do not necessarily need to come from the same language as for
31 example in machine translation (Bahdanau, et al., 2014). Feeding a sentence concatenated
32 with its translation to a self-supervised model like DBMTL or SeqVec (Heinzinger, et al., 2019)
33 allows the translation to be conditioned upon the source language and in addition also upon
34 those parts of the sentence that had already been translated. This produces more coherent
35 translations and resembles the idea of teacher forcing (Williams and Zipser, 1989). Teacher
36 forcing is used for problems which require sequential output generation like question
37 answering or machine translation. During training, teacher forcing randomly uses predictions
38 of previous states or ground truth labels which improves generalization. During inference,
39 however, it relies solely on predictions as no translation exists, yet. The problem of the latter
40 approach is efficiency: it requires LSTM-like sequential processing of a sentence where one
41 token is predicted at a time in order to be able to give predictions of previous time steps to the
42 model. With transformer models and their parallel processing capabilities being on the rise in
43 the field of NLP, this time-consuming step was removed from the framework used here by
44 using only ground truth labels for teacher forcing during training and evaluation/testing. Using
45 this approach during evaluation approximates the actual performance during inference.
46 However, this approximation might lead to overestimation because during inference, the model
47 will have to rely on its own predictions instead of the high-quality ground truth data.
48

49 Applying the basic concept of translation and teacher forcing to protein sequences and
50 their 'translation' to secondary structure, highlighted a pitfall of this tradeoff between reliability
51 and computational overhead: the inherently well-structured nature of protein secondary
structure allowed the model to reach state-of-the-art performance by always only replicating

1 the secondary structure element of the previous residue. By feeding only ground truth labels,
2 the model will just learn to replicate this annotation, since most secondary structure elements
3 naturally occur in patches. Put simply, this is equal to shifting all ground truth labels in the
4 secondary structure annotation by one, which in fact results in 85% 3-state secondary structure
5 prediction performance. As a result, during inference the model will simply replicate the
6 secondary structure prediction for the first residue because the model never learnt to switch
7 from one secondary structure state to the other but just to replicate the teacher forcing signal.
8 Obviously, the results are over-estimated performance values for all methods using the type
9 of teacher forcing described here, namely all our implementations of per-residue predictions.

10 The problem might be addressed through at least two different approaches: remove
11 teacher forcing at the cost of less coherent predictions, or use a model based on the encoder
12 side of the transformer, e.g. Bert (Devlin, et al., 2018). The latter would replace the auto-
13 regressive next-token prediction by an autoencoder-like training which tries to reconstruct
14 corrupted or masked input.

15
16 **Limited gain for per-protein prediction tasks.** For both per-protein tasks explored
17 (localization and membrane-bound/not), the best competitors using evolutionary information
18 still performed substantially better than the novel end-to-end solution (DBMTL; Fig. 2ab). For
19 localization prediction, our approach still outperformed many popular solutions that use MSA
20 (Fig. 2a). However, a recent publication might suggest this view to be possibly slightly distorted
21 by the choice of data set and number of states (Savojardo, et al., 2018). Another reason for
22 this result could be attributed to the technical limitation forcing proteins to be chopped to 1024
23 residues (using only the first and last 512 residues for protein longer than 512 amino acids).
24 This clearly affected the per-protein data sets more than the per-residue data sets as the latter
25 were taken from the PDB (Consortium, 2018), which tends to contain domain-like, aka. shorter
26 fragments of long proteins. However, the same pre-processing (cutting long proteins) was
27 performed for the classification into membrane-bound/not, whilst here performance was not as
28 much below the top-performer (DeepLoc). This could be explained by two observations. Firstly,
29 the signal of membrane-bound proteins is among the most dominant signals captured during
30 language modeling (Heinzinger, et al., 2019). Secondly, subcellular localization relies heavily
31 on certain short sequence motifs which might be removed when chopping the sequences
32 (Almagro Armenteros, et al., 2017).

33 Another possible explanation as to why the end-to-end solution DBMTL improved less
34 over the two-step approach for localization and membrane classification w.r.t. secondary
35 structure prediction might be the smaller data sets for the former. All per-protein tasks had
36 many orders of magnitude fewer samples than the per-residue tasks. Possibly, these were too
37 few to tap into the full potential of the end-to-end solutions with many free parameters.

38
39 **How can semi-supervised multi-task learning be so successful?** Two factors may have
40 contributed to the impressive improvement in secondary structure prediction: (1) semi-
41 supervised learning and (2) model type. (1) *Semi-supervised learning*: previously, models
42 tapping into the power of transfer learning capabilities of self-supervised language modeling
43 might have pushed what can be achieved from single sequences to the limit without end-to-
44 end training, i.e. coupling the pre-training phase and the task-specific fine-tuning phase. The
45 DBMTL model tapped into the possibility of leveraging large amounts of unlabeled sequence
46 (big data) while transferring knowledge between annotated proteins and proteins without any
47 labels. This might have allowed the model to transfer the knowledge not only from the language
48 model task to other tasks, but between tasks. (2) *Model type*: In NLP, the transformer model
49 is currently achieving state-of-the-art results in various sequence to sequence problems (Dai,
50 et al., 2019; Devlin, et al., 2018; Yang, et al., 2019). Leveraging this concept for biological
51 sequences helped to push the boundaries of methods which do not rely on evolutionary
52 information. The core module of all transformer models is the attention mechanism. This
53 mechanism allows transformers to learn which regions in the input space (aka. sequence)

1 contribute most to a prediction at a certain position (residue). Simply put, each output of the
2 self-attention module is a weighted sum over all its inputs. One crucial advantage of this
3 approach is that the length of the computational graph computing the dependency between
4 any two residue positions i and j in a protein is independent of their sequential distance $|i-j|$,
5 allowing to better capture long-range dependencies (Dai, et al., 2019).
6

7 **Speed-up.** Most importantly, DBMTL speeds up over other state-of-the-art methods by not
8 having to compile MSAs. How much time is gained through this step depends on how the
9 MSAs are built. Compared to NetSurfP-2.0 we increase the speed $\sim 3000x$ (HHblits3 is used
10 to compute alignments) – $110x$ (MMseqs2 (Steinegger and Söding, 2017) is used to compute
11 the alignments) On top, the advantage of using alignment-free predictors is growing hand-in-
12 hand with the growth of bio-databases. These figures explain why, for many applications, even
13 slightly less accurate, but faster predictors might be preferable to users when the top predictors
14 become inexecutable due to limited computing resources.
15
16

17 Conclusion

18 We presented a new method, DBMTL (Fig. 1), realizing end-to-end multi-task (MT) with the
19 objective to push machine learning to a level at which it can compete with methods using
20 evolutionary information in an alignment-free manner. DBMTL demonstrated how semi-
21 supervised learning can be applied to the language of life, distilled in the form of protein
22 sequences. By fusing self-supervised language modelling and supervised fine-tuning into one
23 model which is trained with a joint loss that characterizes various protein properties, knowledge
24 is transferred between different tasks, allowing the model to build a multi-modal understanding
25 of proteins. In order to share as much knowledge between the tasks as possible, while still
26 enabling the network to learn task-specific features, the MT implementation used in this work
27 provides task-specific input signals. We implemented four supervised tasks: two on the level
28 of residues (word-level), namely secondary structure prediction in 3- and 8-states, and two on
29 the level of proteins (sentence-level), namely subcellular localization prediction and the
30 classification membrane-vs-soluble proteins. For the time being, the performance of the per-
31 residue (word-level) tasks cannot be trusted due to problems arising from adopting T2T
32 (Vaswani, et al., 2018; Vaswani, et al., 2017) to this problem (Discussion). For the per-protein
33 (sentence-level) tasks, DBMTL performed similar to many methods using evolutionary
34 information from MSAs, however, it remained substantially below the level of today's top
35 performers (e.g. DeepLoc (Almagro Armenteros, et al., 2017)). Nevertheless, this is achieved
36 without the costly step of building MSAs, thus our new method speeds up over competitors by
37 about two orders of magnitude. Thus, semi-supervised MT approaches, as the one described
38 in this work, may become a new frontier in computational biology and medicine, in particular
39 when a wealth of unannotated data (sequences) contrasts with very constrained annotations
40 (about protein structure and function).
41
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4 References

- 5 Abriata, L.A., *et al.* Assessment of hard target modeling
6 in CASP12 reveals an emerging role of alignment
7 based contact prediction methods. *Proteins*
8 *Structure, Function, and Bioinformatics* 2018;86:97
9 112. 67
- 10 Alley, E.C., *et al.* Unified rational protein engineering
11 with sequence-based deep representation learning
12 *Nat Methods* 2019:1-8. 70
- 13 Almagro Armenteros, J.J., *et al.* DeepLoc: prediction of
14 protein subcellular localization using deep learning
15 *Bioinformatics* 2017;33(24):4049. 73
- 16 Asgari, E. and Mofrad, M.R. Continuous Distributed
17 Representation of Biological Sequences for Deep
18 Proteomics and Genomics. *PloS one*
19 2015;10(11):e0141287. 77
- 20 Bahdanau, D., Cho, K. and Bengio, Y. Neural machine
21 translation by jointly learning to align and translate
22 *arXiv preprint arXiv:1409.0473* 2014. 80
- 23 Berman, H.M., *et al.* The protein data bank. *Nucleic*
24 *acids research* 2000;28(1):235-242. 82
- 25 Blum, T., Briesemeister, S. and Kohlbacher, O.
26 MultiLoc2: integrating phylogeny and Gene Ontology
27 terms improves subcellular protein localization
28 prediction. *BMC Bioinformatics* 2009;10:274. 86
- 29 Boutet, E., *et al.* UniProtKB/Swiss-Prot, the Manually
30 Annotated Section of the UniProt KnowledgeBase
31 How to Use the Entry View. *Methods Mol Biol*
32 2016;1374:23-54. 90
- 33 Briesemeister, S., *et al.* SherLoc2: a high-accuracy
34 hybrid method for predicting subcellular localization of
35 proteins. *J Proteome Res* 2009;8(11):5363-5366. 93
- 36 Briesemeister, S., Rahnenfuhrer, J. and Kohlbacher, O.
37 YLoc - an interpretable web server for predicting
38 subcellular localization. *Nucleic Acids Res* 2010;38
39 Suppl:W497-502. 97
- 40 Capriotti, E., Fariselli, P. and Casadio, R. I-Mutant2
41 predicting stability changes upon mutation from the
42 protein sequence or structure. *Nucleic Acids Res*
43 2005;33(Web Server issue):W306-310. 101
- 44 Casadio, R., Martelli, P.L. and Pierleoni, A. The
45 prediction of protein subcellular localization from
46 sequence: a shortcut to functional genome
47 annotation. *Brief Funct Genomic Proteomic*
48 2008;7(1):63-73. 106
- 49 Chou, K.C., Wu, Z.C. and Xiao, X. iLoc-Euk: a multi-
50 label classifier for predicting the subcellular
51 localization of singleplex and multiplex eukaryotic
52 proteins. *PloS one* 2011;6(3):e18258. 110
- 53 Consortium, U. UniProt: the universal protein
54 knowledgebase. *Nucleic acids research*
55 2018;46(5):2699. 113
- 56 Cuff, J.A. and Barton, G.J. Evaluation and improvement
57 of multiple sequence methods for protein secondary
58 structure prediction. *Proteins: Structure, Function*
59 *and Genetics* 1999;34(4):508-519. 117
- 60 Dai, Z., *et al.* Transformer-xl: Attentive language models
61 beyond a fixed-length context. *arXiv preprint*
62 *arXiv:1901.02860* 2019.
- Devlin, J., *et al.* Bert: Pre-training of deep bidirectional
transformers for language understanding. *arXiv*
preprint arXiv:1810.04805 2018.
- Drozdetskiy, A., *et al.* JPred4: a protein secondary
structure prediction server. *Nucleic acids research*
2015;43(W1):W389-W394.
- Frishman, D. and Argos, P. Knowledge-based protein
secondary structure assignment. *Proteins: Structure,*
Function, and Genetics 1995;23:566-579.
- Fu, L., *et al.* CD-HIT: accelerated for clustering the next-
generation sequencing data. *Bioinformatics*
2012;28(23):3150-3152.
- Goldberg, T., Hamp, T. and Rost, B. LocTree2 predicts
localization for all domains of life. *Bioinformatics*
2012;28(18):i458-i465.
- Goldberg, T., *et al.* LocTree3 prediction of localization.
Nucleic Acids Res 2014;42(Web Server issue):W350-
355.
- Hamp, T. and Rost, B. Evolutionary profiles improve
protein-protein interaction prediction from sequence.
Bioinformatics 2015;31(12):1945-1950.
- Hayat, S., *et al.* All-atom 3D structure prediction of
transmembrane β -barrel proteins from sequences.
Proceedings of the National Academy of Sciences
2015;112(17):5413-5418.
- Heffernan, R., *et al.* Capturing non-local interactions by
long short-term memory bidirectional recurrent neural
networks for improving prediction of protein
secondary structure, backbone angles, contact
numbers and solvent accessibility. *Bioinformatics*
2017;33(18):2842-2849.
- Heinzinger, M., *et al.* Modeling aspects of the language
of life through transfer-learning protein sequences.
BMC Bioinformatics 2019;20:723.
- Heinzinger, M., *et al.* Modeling the Language of Life-
Deep Learning Protein Sequences. *bioRxiv*
2019:614313.
- Horton, P., *et al.* WoLF PSORT: protein localization
predictor. *Nucleic Acids Res* 2007;35(Web Server
issue):W585-587.
- Howard, J. and Ruder, S. Universal language model
fine-tuning for text classification. *arXiv preprint*
arXiv:1801.06146 2018.
- Jones, D.T. Protein secondary structure prediction
based on position-specific scoring matrices. *Journal*
of Molecular Biology 1999;292(2):195-202.
- Kabsch, W. and Sander, C. Dictionary of protein
secondary structure: pattern recognition of hydrogen
bonded and geometrical features. *Biopolymers*
1983;22:2577-2637.
- Klausen, M.S., *et al.* NetSurfP-2.0: Improved prediction
of protein structural features by integrated deep
learning. *Proteins* 2019.
- Li, W. and Godzik, A. Cd-hit: a fast program for
clustering and comparing large sets of protein or
nucleotide sequences. *Bioinformatics*
2006;22(13):1658-1659.

- 1 Mehta, P.K., Heringa, J. and Argos, P. A simple and fast
2 approach to prediction of protein secondary structure
3 from multiply aligned sequences with accuracy above
4 70%. *Protein Science* 1995;4:2517-2525. 50
- 5 Mikolov, T., et al. Efficient estimation of word
6 representations in vector space. *ArXiv*
7 2013;arXiv:1301.3781. 53
- 8 Nair, R. and Rost, B. Better prediction of sub-cellular
9 localization by combining evolutionary and structural
10 information. *Proteins: Structure, Function, and*
11 *Bioinformatics* 2003;53(4):917-930. 57
- 12 Perdigao, N., et al. Unexpected features of the dark
13 proteome. *Proceedings of the National Academy of*
14 *Sciences of the United States of America* 2015. 60
- 15 Peters, M.E., et al. Deep contextualized word
16 representations. *arXiv* 2018;arXiv:1802.05365. 62
- 17 Pollastri, G., et al. Prediction of coordination number
18 and relative solvent accessibility in proteins. *Proteins*
19 2002;47(2):142-153. 65
- 20 Radford, A., et al. Language models are unsupervised
21 multitask learners. *OpenAI Blog* 2019;1(8). 67
- 22 Radivojac, P., et al. Protein flexibility and intrinsic
23 disorder. *Protein Science* 2004;13:71-80. 69
- 24 Rives, A., et al. Biological structure and function emerge
25 from scaling unsupervised learning to 250 million
26 protein sequences. *bioRxiv* 2019:622803. 72
- 27 Rost, B. Protein secondary structure prediction
28 continues to rise. *Journal of Structural Biology*
29 2001;134:204-218. 75
- 30 Rost, B., et al. Transmembrane helix prediction at 95%
31 accuracy. *Protein Science* 1995;4:521-533. 77
- 32 Rost, B. and Sander, C. Improved prediction of protein
33 secondary structure by use of sequence profiles and
34 neural networks. *Proceedings of the National*
35 *Academy of Sciences* 1993;90:7558-7562. 81
- 36 Rost, B. and Sander, C. Prediction of protein secondary
37 structure at better than 70% accuracy. *Journal of*
38 *Molecular Biology* 1993;232:584-599. 84
- 39 Rost, B. and Sander, C. Combining evolutionary
40 information and neural networks to predict protein
41 secondary structure. *Proteins: Structure, Function,*
42 *and Genetics* 1994;19:55-72. 88
- 43 Rost, B. and Sander, C. Conservation and prediction of
44 solvent accessibility in protein families. *Proteins*
45 *Structure, Function, and Genetics* 1994;20(3):216-
46 226. 92
93
94
- Savojardo, C., et al. BUSCA: an integrative web server
to predict subcellular localization of proteins. *Nucleic*
Acids Res 2018;46(W1):W459-W466.
- Steinegger, M., et al. HH-suite3 for fast remote
homology detection and deep protein annotation.
BMC Bioinformatics 2019;20(1):473.
- Steinegger, M., Mirdita, M. and Söding, J. Protein-level
assembly increases protein sequence recovery from
metagenomic samples manyfold. *Nat Methods*
2019:1.
- Steinegger, M. and Söding, J. MMseqs2 enables
sensitive protein sequence searching for the analysis
of massive data sets. *Nature biotechnology*
2017;35(11):1026.
- Suzek, B.E., et al. UniRef clusters: a comprehensive
and scalable alternative for improving sequence
similarity searches. *Bioinformatics* 2015;31(6):926-
932.
- Vaswani, A., et al. Tensor2Tensor for neural machine
translation. *arXiv* 2018;1803.07416.
- Vaswani, A., et al. Attention is all you need. In,
Advances in neural information processing systems.
2017. p. 5998-6008.
- Velankar, S., et al. SIFTS: structure integration with
function, taxonomy and sequences resource. *Nucleic*
acids research 2012;41(D1):D483-D489.
- Wang, G. and Dunbrack Jr, R.L. PISCES: a protein
sequence culling server. *Bioinformatics*
2003;19(12):1589-1591.
- Wang, S., et al. RaptorX-Property: a web server for
protein structure property prediction. *Nucleic acids*
research 2016;44(W1):W430-W435.
- Williams, R. and Zipser, D. A learning algorithm for
continually running fully recurrent neural networks.
Neural Computation 1989;1:270-280.
- Yang, Y., et al. Sixty-five years of the long march in
protein secondary structure prediction: the final
stretch? *Briefings in bioinformatics* 2016;19(3):482-
494.
- Yang, Z., et al. XLNet: Generalized Autoregressive
Pretraining for Language Understanding. *arXiv*
preprint arXiv:1906.08237 2019.
- Yu, C.S., et al. Prediction of protein subcellular
localization. *Proteins* 2006;64(3):643-651.
- Zhang, Q.C., et al. Structure-based prediction of
protein-protein interactions on a genome-wide scale.
Nature 2012;490(7421):556-560.