Title: Demographic history and genomics of local adaptation in blue tit populations

Running title: Population genomics in blue tit

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Abstract

Understanding the genomic processes underlying local adaptation is a central aim of

modern evolutionary biology. This task requires identifying footprints of local selection

but also estimating spatio-temporal variation in population demography and variation in

recombination rate and diversity along the genome. Here, we investigated these

parameters in blue tit populations inhabiting deciduous *versus* evergreen forests and

insular *versus* mainland areas, in the context of a previously described strong

phenotypic differentiation. Neighboring population pairs of deciduous and evergreen

habitats were weakly genetically differentiated (F_{ST} = 0.004 on average), nevertheless

with a statistically significant effect of habitat type on the overall genetic structure. This

low differentiation was consistent with the strong and long-lasting gene flow between

populations, inferred by demographic modeling. In turn, insular and mainland

populations were moderately differentiated (F_{ST} = 0.08 on average), in line with the

inference of moderate ancestral migrations, followed by isolation since the end of the

last glaciation. Effective population sizes were overall large, yet smaller on the island

than on the mainland. Weak and non-parallel footprints of divergent selection between

deciduous and evergreen populations were consistent with their high connectivity and

the probable polygenic nature of local adaptation in these habitats. In turn, stronger

footprints of divergent selection were identified between long isolated insular *versus*

mainland birds, and were more often found in regions of low recombination as expected

from theory. Lastly, we identified a genomic inversion on the mainland, spanning

2.8Mb. These results provide insights into the demographic history and genetic

architecture of local adaptation in blue tit populations at multiple geographic scales.

Introduction

Local adaptation in heterogeneous environments imposing divergent selection on

counterparts of a same species has fascinated scientists for decades (Kawecki & Ebert

2004; Blanquart *et al.* 2013). However, it is only with the recent advance in sequencing

technologies that numerous recent empirical studies started discovering the genomic

features and processes underlying local adaptation in heterogeneous environments

(Savolainen *et al.* 2013; Tiffin & Ross-Ibarra 2014; Manel *et al.* 2016; Hoban *et al.* 2016).

Especially, many studies used so-called F_{ST} genome scans between populations (Excoffier

et al. 2009; Narum & Hess 2011; de Villemereuil *et al.* 2014) to detect genes under

selection, and potentially implicated in local adaptations (Hohenlohe et al. 2010;

Lamichhaney et al. 2016; Reid et al. 2016; Perrier et al. 2017). Nonetheless, genome

scans need improvements for example for studying polygenic variation (Rockman 2012;

Wellenreuther & Hansson 2016). Genome scans also need to be interpreted in the light

of recombination variation along the genome (Stapley et al. 2017, Booker et al. 2020)

since it shapes the potential extent of diversity and of differentiation in the different

regions of the genome (Cutter & Payseur 2013; Tine et al. 2014; Burri et al. 2015;

Rougemont et al. 2019). Genome scans results need to be interpreted in the light of the

demographic history of populations since i) spatio-temporal variation in gene flow and

of effective population size affect the possibility of local adaptations (Lenormand 2002)

and the level of false positive due to genetic drift, and ii) regions implicated in

reproductive isolation accumulated during allopatry showing increased differentiation

upon secondary contacts can be misinterpreted as genuine footprints of recent

divergent selection (Bierne et al. 2011; 2013). Lastly, structural variants such as

inversions also require special attention, since they protect adaptive gene sets from

recombination (Kirkpatrick 2006; 2010; Stapley *et al.* 2017; Wellenreuther *et al.* 2019)

and hence enable their persistence and rapid redeployment in heterogeneous

environments despite high gene flow (Lowry & Willis 2010; Sinclair-Waters et al. 2017;

Todesco et al. 2019).

Here, we aimed at investigating various demographic and genomic aspects of the

adaptive divergence of a well-studied passerine bird, the blue tit (*Cyanistes caeruleus*).

Populations of small insectivorous passerines have long been used to study local

adaptation, in both quantitative genetics and population genetics frameworks

(Carbonell et al. 2003; Broggi et al. 2005; Laaksonen et al. 2015). In particular, several

blue tit populations breeding in a heterogeneous habitat in Southern France (Figure 1A)

offer an ideal context to study local adaptation. Four of them (two deciduous and two

evergreen, circled in black in Figure 1A) have been subject to a long term project

spanning more than 40 years (Blondel et al. 2006; Charmantier et al. 2016). These

populations show marked quantitative phenotypic differences (Figure 1B&C), notably in

morphological (e.g. tarsus length and body mass), life-history (lay date and clutch size)

and behavioural traits (e.g. song characteristics and handling aggression) (Charmantier

et al 2016). These phenotypic differences were found at two spatial scales. First, birds

breeding in deciduous forest habitats are taller, more aggressive, and lay larger and

earlier broods than birds in evergreen forests (see table 1 in Charmantier et al 2016).

Strikingly, neighbouring populations in deciduous and evergreen habitats are weakly

genetically differentiated (Porlier et al. 2012b; Szulkin et al. 2016; Dubuc-Messier et al.

2018) despite the short spatial scale, which questioned the mechanisms of persistence

of the observed phenotypic differentiation against presumably large gene flow. Second,

insular blue tits from Corsica, that might have been isolated since the sea level rise after

the last glacial maximum (the sea level raised of 120 m from 17,000 to 5,000 years ago

(Lambeck & Bard 2000; Jouet et al. 2006)), are smaller and more colourful than their

mainland counterparts (again, see table 1 in Charmantier et al 2016) and are listed as

two subspecies (*Cyanistes caeruleus caeruleus* on mainland Europe and *Cyanistes*

caeruleus ogliastrae mainly in Corsica and Sardinia). Overall, traits displaying these

strong phenotypic differences had heritability ranging from 0.20-0.43 (eg for lay date,

(Caro et al. 2009)) to 0.42-0.60 (eg for tarsus length, (Teplitsky et al. 2014; Delahaie et

al. 2017; Perrier et al. 2018)), are classically related with fitness, and hence could be

involved in a local adaptation process. In particular, the heritability of lay date and the

breeding time gap between populations could result in reproductive isolation by

breeding time, limiting gene flow and favouring local adaptation. The studied traits were

typically quantitative (Charmantier et al. 2016), hence probably controlled by a

polygenic architecture (Perrier et al. 2018) composed of many loci with small individual

effects, as found in similar traits for other passerine birds (Santure et al. 2013; Bosse et

al. 2017; Hansson et al. 2018; Lundregan et al. 2018). Overall, given their phenotypic,

demographic and genetic characteristics, these blue tit populations provide an ideal

framework to study the genomic architecture of polygenic adaptation in heterogeneous

environments.

We investigated genome wide patterns of genetic diversity and differentiation

and the demographic history between several populations of blue tit from Southern

France, in heterogeneous forest habitats (deciduous versus evergreen) and in insular

(Corsica island) and mainland areas (Mainland France) in order to better understand the

determinants of their local adaptation. The analysis was based on Resequencing of birds

sampled in four sites studied in the context of a long-term project (Blondel et al. 2006;

Charmantier et al. 2016) together with three additional pairs of deciduous and

evergreen forests in order to test for parallel evolution (see Figure 1A). First, we

investigated variation in genetic diversity and differentiation in order to verify that

habitat type and geographic distance explained a significant proportion of the genetic

structure between populations (Szulkin et al. 2016). Second, we investigated the

historical and contemporary demography of each deciduous and evergreen population

pair in order to better understand the origin of their differentiation, and the

demography between the populations on mainland France and on the Corsica Island in

order to estimate their split time and their subsequent connectivity and effective

population size. Third, we screened the genome for F_{ST} outlier loci and inversions

potentially implicated in adaptation in the two habitat types and between Corsica and

Mainland France. Rather than identifying the genes under selection, we wanted to test

whether the adaptation had a polygenic or oligogenic architecture and whether outlier

loci would be more frequent in regions with low recombination rate. We discuss our

results in the light of the recent literature in population genomics, to decipher the role

of genomic and demographic variations on the genetic and phenotypic divergence

between blue tits in deciduous *versus* evergreen habitats and in mainland *versus* insular

contexts.

Methods

Sites and sampling

947 blue tits (*Cyanistes caeruleus*) were captured in ten locations (Figure 1A, Table 1). Three locations are located in the South of mainland France (codes staring with "A": A1D, A2D, A2E) and seven in the Corsica Island (staring with "B": B3E, B4D, B4E, B5D, B5E, B6D, B6E). The numbers from 1 to 6 indicated the sampling area and hence the population pairs. Half of these sites were dominated by the deciduous downy oak *Quercus pubescens* (indicated by "D" in the code) and the other half are dominated by the evergreen holm oak *Q. ilex* (indicated by "E"). Four of these locations (A1D, B3E, B4D, B4E) are monitored as part of a long-term research program started in 1976 and have been described in previous studies (Blondel et al. 2006; Charmantier et al. 2016). The 6 other sites (A2D, A2E, B5D, B5E, B6D & B6E) were sampled in order to obtain further replicates of neighboring deciduous and evergreen populations. Capture and handling of the birds was conducted under permits provided by the *Centre de Recherches sur la Biologie des Populations d'Oiseaux* (CRBPO) and by the *Direction Départementale des Services Vétérinaires* (DDSV). Birds were captured during the reproductive period, from early April to late June, on their nesting territory. Birds were captured either in nest boxes during the feeding of their young (in the four sites studied on the long term), or using nets in the vicinity of their nest (in the other sites). Birds were banded with a unique metal ring provided by the CRBPO. Five to 20µl of blood were sampled from a neck or a wing vein from breeding blue tits. Blood was stored at 4°C in Queen's buffer (Seutin et al. 1991).

Molecular biology and sequencing

DNA extractions were achieved using Qiagen DNeasy Blood & Tissue kits and were

randomized across sites. DNA was quantified using first a NanoDrop ND8000

spectrophotometer and then a Qubit 2.0 fluorometer with the DNA HS assay kit (Life

Technologies). DNA quality was examined on agarose gels. Library preparation using

RAD-seq (restriction-site-associated DNA sequencing; (Baird et al. 2008)) with the

enzyme *SbfI* was done by Montpellier GenomiX (MGX) platform (CNRS, Montpellier).

Each individual was identified using a unique 6 nucleotides tag, individuals were

multiplexed in equimolar proportions by libraries of 36 individuals, and each library was

sequenced on a lane of an Illumina HiSeq 2000. Single-end sequencing was used to

produce 100bp reads. This design was used to obtain an average read depth of 50x. The

DNA of three individuals was extracted twice and processed in different libraries to test

for reliability of the genotyping process.

Bioinformatics and data filtering

Raw sequences were inspected with *FastQC* (Andrews 2010) for quality controls.

Potential fragments of Illumina adapters were trimmed with *Cutadapt* (Martin 2011),

allowing for a 10% mismatch in the adapter sequence. Reads were filtered for overall

quality, demultiplexed and trimmed to 85bp using *process_radtags,* from the *Stacks*

software pipeline 1.39 (Catchen et al. 2013), allowing for one mismatch in the barcode

sequence. Sequencing RAD-tags resulted in a median value of 5,449,564 reads per

individual. *BWA-MEM* 0.7.13 (Li & Durbin 2009) was used to map individual sequences

against the reference genome of the great tit *Parus major* (Laine et al. 2016) and to

produce *sam* files using default options. Indeed, although great tit and blue tit diverged

about 7 to 14My ago (Päckert et al 2007), the use of a reference genome over a de-novo

approach is very often highly recommended (See e.g. Rochette et Catchen 2017). It is

also interesting to note that synteny is typically highly conserved in birds (Backström et

al 2008), suggesting that aligning reads on a divergent species would still allow analyses

based on SNP position along the genome. On average, 93% of the raw reads were

mapped against the genome (Supplementary table 1). *Samtools* 0.1.19 (Li et al. 2009)

was used to build and sort *bam* files. We used *pstacks* to treat *bam* files, align the reads

as assembled loci and call SNPs in each locus. We used a minimum depth of coverage

(m) of 5, the SNP model, and alpha = 0.05 (chi square significance level required to call a

heterozygote or homozygote). *cstacks* was used to build the catalogue of loci, allowing

3 mismatches between sample loci when building the catalog. *sstacks* was used to

match loci against the catalog. Lastly, *populations* program in *Stacks* was used to

genotype individuals. In this program, relatively permissive filters were applied, to retain

SNP genotyped in at least 50% of individuals (all individuals from all sites grouped) and

with a minimum read depth of 4. 350,941 SNP and 947 individuals were obtained in this

"GeneralDataset".

We hence applied additional filters to this GeneralDataset using the programs

VCFtools (Danecek et al. 2011) and *Plink* (Purcell et al. 2007). We filtered for a minimum

average read depth of 10 and a maximum average read depth of 100 (corresponding

approximately to the 5%-95% distribution of read depth). SNP genotyped in less than

80% of the individuals were removed. We removed SNPs with observed heterozygosity

≥ 0.65 among individuals on Corsica or on mainland to reduce the potential occurrence

of stacked paralogues. Individuals below 85% genotyping rate were removed. Identity-

By-State (IBS) was estimated for three replicated individuals (from the DNA extraction to

SNP-calling) in order to investigate reliability of the entire genotyping process. The IBS

measured between replicates were high, ranging from 0.9989 to 0.9999, indicating very

low genotyping error rate. These replicates were then removed from the dataset. Using

the R packages *gdsfmt* and *SNPRelate* (Zheng et al. 2012), we measured the realized

genomic relatedness (GRM) between individuals in order to remove highly related

individuals (*i.e.* full-sibs and parent-offspring). For each pair of individuals with

relatedness ≥ 0.35, we removed one individual. This procedure was applied in order to

limit biases due to highly related individuals. Indeed, since we sampled breeding birds at

their nests for the four sites studied on the long-term and that they tend to disperse

relatively close to their nests, we expected a higher percentage of related individuals

than there actually is in the population. We then limited the number of individuals to

60, chosen at random, in populations (A1D, B3E, B4D & B4E) in which a large number of

individuals were genotyped. This limitation was achieved in order to limit differences in

analyses precision between populations due to unequal sample sizes (all individuals will

be used in another ongoing study of the genomic architecture of quantitative adaptive

traits). We hence removed potential monomorphic SNPs and created the dataset

"FilteredDataset". 454 individuals (Table 1) and 144,773 SNPs were kept in this final

dataset used for all the genomic analyses excepted analyses of demographic history

using ABC. The median genotyping rate across all SNPs for these individuals was 0.981.

The median read depth across genotypes (SNPs x Individual) was 49.2x. The number of

SNPs per chromosome ranged from 143 (LGE22) to 15,738 (chromosome 2). Further

filtering (e.g. MAF) was often operated, depending on the analyses, and thereafter

mentioned if it was the case. To perform analyses of demographic history using ABC

analyses, we produced a haplotype VCF file for these 454 aforementioned individuals

with the populations module of Stacks and we filtered it as explained in the

supplementary note 1.

Analysis of genetic structure and effects of environmental variables

We used PCAs with the function *snpgdsPCA* from *snprelate* to depict genetic structure

between individuals. PCAs were run for the entire dataset, for mainland and Corsican

populations separately and for each pairs of deciduous and evergreen populations. We

inferred admixture proportions for each individual using *Admixture* 1.23 (Alexander et

al. 2009) with K-values ranging from 1 to 12 and 1,000 bootstraps. The different

clustering solutions were inferred by plotting cross-validations errors and by plotting

individual admixture proportion. The effect of environmental variables, forest

phenology (E versus D), geographic distance (latitude & longitude) and altitude, on

genomic differentiation was measured using a redundancy analysis (RDA) (Legendre &

Fortin 2010; Forester et al. 2015) as implemented in the R package *Vegan* (Oksanen et

al. 2007). We investigated the proportion of genetic variability explained by a

constraining covariance matrix consisting of phenology, latitude, longitude, and altitude

for each individual. We tested the global significance of the model using 1,000

permutations. We ran marginal effects permutations to address the significance of each

variable. Then we focused on the effect of phenology alone, using partial RDA to take

into account the effect of latitude, longitude and altitude. Significance was tested

running 1,000 permutations. For the PCA, the admixture analysis and the RDA, we

selected from the FilteredDataset the SNPs with more than 95% genotyping rate, MAF >

0.05, retaining one SNP per locus and we remove SNP in linkage disequilibrium using the

Plink command "indep 50 5 2". Genome wide differentiation between each sampling

location was measured with Weir and Cockerham's F_{ST} estimator (Weir & Cockerham

1984) implemented in *StAMPP* (Pembleton et al. 2013). F_{ST} was estimated for all the

SNPs, the ones on the autosomes and the ones on the sex chromosome Z separately.

Significance was assessed using 1,000 bootstraps replicates.

Analysis of demographic history

Alternative models of divergence history including the effects of selection at linked sites

affecting *Ne* and of differential introgression (*m*) were compared using an ABC

framework modified from Roux et al. (2016). Linked selection either under the form of

hitchhiking of neutral loci linked to a selective sweep (Maynard Haigh & Smith, 1974) or

under the form of background selection (Charlesworth et al. 1993), have strong effects

in regions of low recombination and have been shown to influence model choice and

parameter estimates (Ewing & Jensen, 2015; Schrider et al. 2016). The same is true

when populations accumulate reproductive incompatibilities during the divergence

process: the resulting barrier to gene flow reduces the effective migration rate along the

genome (Barton & Bengtsson, 1986) and not accounting for it can affect demographic

model choice and parameter estimates (Roux et al. 2014; Souza et al. 2013). Moreover,

including selected loci in demographic inferences can reveal the deeper origins of

population divergence (Bierne et al. 2013). Six scenarios were compared for the 4 pairs

of deciduous and evergreen populations in order to test whether the divergence

between different habitats was not due to a divergence in different historical refugia

but to a contemporary ecological divergence. We included a model of panmixia (PAN), a

model of equilibrium corresponding to the island model with two populations (EQ), a

model of isolation with migration (IM), a model of secondary contact (SC), a model of

divergence with migration during the first generations, *i.e.* ancestral migration (AM),

and a model of strict isolation (SI). The prior and details of the simulation pipeline are

fully described in the supplementary note 1. The PAN model assumes that the two focal

populations descent from a single panmictic population characterized by its effective

size (Ne). The EQ model (equivalent to the island model) assumes that the population is

subdivided into two discrete populations of sizes N_{pop1} and N_{pop2} that are connected by

continuous gene flow at a constant rate each generation. In this model, the divergence

time is not a parameter. The IM, SI, SC, and AM models all assume that an ancestral

population of size N_{ANC} splits at T_{split} into two daughter populations of sizes N_{pop1} and

N_{pop2}. Under SI no subsequent gene flow occurs. Under AM model, gene flow occurs

from T_{split} to T_{am} and is followed by a period without gene flow. Under IM, gene flow is

continuous after T_{split}. Under, the SC model, T_{split} is followed by a period of strict

isolation, after which a secondary contact starts T_{sc} generations. The EQ, IM, AM and SC

models included migration as $M = 4 N_0.m,$ with $M_{1 \leftarrow 2}$ being the number of migrants from

population 2 to population 1 and $M_{2 \leftarrow 1}$ being the reverse. The effect of linked selection

and barriers to gene flow were accounted for, by modeling heterogeneous population

size (Ne) and heterogeneous migration (m) respectively. Such heterogeneity was

modeled using beta distributions as hyper-prior on each of these two parameters. These

resulted in four alternative versions for models with gene flow EQ, AM, IM, SC

(NhomoMhomo, NhomoMhetero, NheteroMhomo, NheteroMhetero) and two versions

for PAN and SI (Nhomo and Nhetero). We used a modified ABC (Csilléry et al. 2010)

pipeline from (Rougemont & Bernatchez 2018) to perform model selection, compute

robustness and to estimate posterior probabilities of parameters.

We also investigated the historical demography of the populations from Corsica

as compared to the ones from the mainland. Gene flow has probably been impossible

since the last de-glaciation and sea-level rise. Therefore, we compared models of

ancient migration (AM) and Strict Isolation (SI). The pipeline described in the above

section, integrating linked selection and barriers to gene flow, was run between the A1D

samples (chosen on the mainland because it had the largest sample size) and B4D

(chosen in Corsica because it had both a large sample size and was from the same

habitat as A1D). We used the same ABC pipeline for the model selection procedure and

parameter estimation as described in the above section. Finally, we attempted to

convert demographic parameter into biological units assuming a mutation rate of 1e-8

mutations/bp/generations.

Genomic diversity

Genome wide genetic diversity was inferred for each sampling location in the dataset by

measuring observed heterozygosity (Ho), proportion of polymorphic loci and MAF

spectrums. For each population and chromosome, and subsequently on average for the

entire genome, Linkage Disequilibrium (LD) decay was measured with *Plink* and

smoothed in R. To contrast with long term Ne estimates from coalescent simulation in

our ABC modeling (see "Analysis of demographic history"), we also inferred recent Ne

for each population using *SNeP* V1.1 (Barbato et al. 2015), which uses LD data, with a

MAF ≥ 0.05 filter per population. We investigated the nature of SNP variation, *i.e.*

synonymous or non-synonymous, blasting all rad sequences on the reference genome

and the transcriptome of the great tit (ftp://ftp.ncbi.nih.gov/genomes/Parus_major/;

Santure et al 2011) using *blastx* (McGinnis & Madden 2004). We kept hits with at least

90% similarity and a minimum amino acid sequence length alignment of 25. We kept

only SNPs for which both the alternative and reference allele yields the same score.

Finally, we tested for differences in the distribution of run of homozygosity (ROH)

between the mainland and Corsica that may have resulted from smaller Ne and larger

inbreeding in Corsica versus the mainland. We used plink 1.9 to estimate the length and

number of ROH. We required a window of 500 kb to be homozyguous in order to be

considered as a ROH, and with a maximum of 100 SNP using the following parameters: --

homozyg-density   50,--homozyg-gap   100,--homozyg-kb   500,--homozyg-snp   100,--

homozyg-window-het 1,--homozyg-window-snp 100,--homozyg-window-threshold 0.05,

and--homozyg-window-missing 20.

Identification of genomic footprints of selection

We used three methods to search for outlier SNPs potentially under divergent selection

between blue tit populations. First, we used *Bayescan* V2.1 (Foll & Gaggiotti 2008) to

search for SNPs potentially under divergent selection at two geographic levels: i)

between the mainland and Corsican populations and ii) between each local pairs of

evergreen (E) and deciduous (D) populations, *i.e.* A2D vs A2E, B4D vs B4E, B5D vs B5E,

B6D vs B6E. We filtered each of the five datasets for a minimum MAF of 0.05. We used

default parameters except for prior odds that were set at 10,000 in order to limit false

positives. We investigated the parallelism across pairs of D-E environments. Second, we

estimated F_{ST} along the genome using either a 200kb sliding average with *VCFtools*, or

the function "snpgdsSlidingWindow" from the package SNPrelate to estimate F_{ST} in

blocks of 50 SNPs moving by 5 SNPs. The second window strategy was used in order to

compensate for lower SNP density in regions of low recombination that tend to

exaggerate the contribution of individual SNPs in these regions and to dilute the

individual SNP contribution in regions of high recombination (see Perrier & Charmantier

2019 for a broader comment on this). Third, we used an RDA as an alternative method

to search for SNPs putatively implicated in multilocus adaptation i) between populations

in deciduous and evergreen habitats, and ii) between populations in Corsica and on the

mainland. As suggested by Forester et al (2018), such a multivariate method may be

more suitable than univariate ones to detect weaker footprints of adaptation that are

expected in polygenic adaptations in response to complex environmental heterogeneity.

Using a similar procedure as described earlier in the methods, we used two RDAs

constrained to investigate the effect of phenology (i) or of the geography (ii). We then

used a 3 standard deviation cutoff as suggested by Forester et al (2018) to list loci with

outlier loading scores on the first RDA axes. We compared the loci found using these

different methods. We reported in which genes these outliers were found (the list of

genes can be found together with the genome published by Laine et al 2016 on NCBI).

Variation of genomic differentiation with recombination rate

We investigated variation of F_{ST} with local recombination rate and whether SNP outliers

were more often found in regions of low recombination than elsewhere in the genome.

We estimated local recombination rates using a coalescent method implemented in

Ldhat (McVean 2004) using linkage disequilibrium signal. Following these authors'

recommendation, the dataset was split in blocks of 2,000 SNPs with 500 overlapping

SNPs. A MAF of 0.05 and a maximum of 5% of missing data were allowed. The local

recombination rate, $\rho = 4Ne\ r$, was then estimated in each block independently with the

Bayesian reversible jump MCMC scheme implemented in interval. We used a block

penalty of 5, with 30 millions MCMC iterations sampled every 5,000 iterations. The first

250,000 iterations were discarded. To speed up computations we used the pre-

computed two locus likelihood table for n = 190 and assuming theta = 0.001. We

estimated ρ in a composite dataset with individuals from every population, habitats and

sex. We tested for correlations between SNP F_{ST} and recombination rate using linear

models, for Corsica-mainland and for deciduous-evergreen differentiation,

independently, and we represented the correlation using a LOESS fit. We tested

whether outliers found using Bayescan and the RDA method for both Corsica-mainland

and deciduous-evergreen differentiation were more often found in regions of low

recombination than elsewhere in the genome using chi-square ($\chi 2$) tests, and we

represented the pattern using histograms.

Detection of genomic inversions

We searched for potential genomic inversions using a variety of descriptive statistics. First, we searched for genomic regions having a particularly low recombination rate and large long-distance linkage disequilibrium, nevertheless associated to a high density of SNPs and therefore unlikely to correspond to pericentromeric regions but rather to local suppression of recombination that may be due to inversions. Second, we implemented a PCA sliding window analysis in order to identify portions of the genome with individuals carrying an inversion at the homozygous or heterozygous state or individuals exempt from the inversion (Ma & Amos 2012). We first used 10Mb windows sliding by 1Mb and then 1Mb windows sliding by 100kb, in order to use enough SNPs to perform PCAs. We then used *Lostruct* (Li & Ralph 2019), with k=2, sliding by 100 SNPs, to identify particular blocks of linked SNPs explaining an abnormally high proportion of variance between 2 groups of individuals (e.g. inverted and non-inverted).

We detected one putative inversion. We verified, using admixture with K = 2 for analyzing SNPs from the inversion, that putative heterozygous individuals had an admixture ratio close to 1:1 of both putative inverted and non-inverted homozygous clusters. We then inspected variations of F_{ST} (per SNP) and π (per 10kb window, using *vcftools*) along the genome between individuals that were inverted homozygous, non-inverted homozygous, and heterozygous for the inversion, looking for potentially reduced diversity and increased differentiation at the inversion. We looked for potential salient variations in read depth in and around the putative inversion. We also tested whether the detected inversion was at Hardy-Weinberg equilibrium (using a common

χ2-test) and whether the frequency of the inversion varied geographically and between

evergreen and deciduous habitats.

To study the history of the putative inversion identified, we aimed at measuring

intra- and inter-specific genetic distance and absolute divergence, at the inversion and

for the entire genome, for several blue tits and great tits. To do that, we first generated

a new SNP dataset by running stacks with the same pipeline and parameters as

explained earlier, with 2 mainland great tits, 2 insular great tits, 4 mainland blue tits

homozygous for the inversion, 4 mainland blue tits not carrying the inversion, and 4

Corsican blue tits not carrying the inversion (the inversion was not found in Corsica).

Second, we selected the SNPs from the region of the inversion. Third, Dxy was measured

between the 5 aforementioned groups of individuals using *PopGenome* (Pfeifer et al

2014). Using this measure of Dxy, we estimated approximately the inversion apparition

time using $T = Dxy / 2\mu$, assuming a standard mutation rate of 1e-8 and with the

simplifying assumption of no gene flow and no introgression. Lastly, we represented the

divergence between these individuals using an UPGMA tree of bitwise distance using

the R package *poppr* 1.1.1 (Kamvar et al 2014).

Gene ontology

We used the R package topGO (Alexa & Rahnenführer, 2009) to investigate the potential

gene ontologies (GO) that were statistically enriched for the sets of genes identified

among outliers and the inversion, compared to the entire list of genes in which all the

SNPs from the entire dataset were found. For the outlier tests, we used GO analyses for

each gene list obtained using the different outlier identification methods but also for

aggregated lists of all of the gene identified for deciduous-evergreen tests or for Corsica-

Mainland tests. We used the GO referenced for the zebra finch, T. Guttata

(tguttata_gene_ensembl). We report a Fisher enrichment test p-value and a p-value

after applying a Benjamini and Hochberg (BH) correction for multiple testing to control

for false discovery rate (FDR).

Results

Genetic structure and effects of environmental variables

The admixture analysis suggested the existence of 2 main distinct genetic groups

corresponding to mainland and island populations (Figure 2A). Increasing K value

contributed to delineate the populations, showing the existence of a weak structure

between the 6 main areas. However, the coefficient of variation increased with K. There

was very little evidence for genetic structure within each of the four pairs of D- and E-

populations. PCAs revealed a clear structure between individuals from the mainland and

the island but little structure within each of these 2 groups (Supplementary Figure 1).

The full RDA model was significant (p < 0.001, Supplementary table 2, Figure 2B), as well

as the effects of each variable tested in the model: phenology (*i.e.* E versus D, p = 0.006),

latitude (p < 0.001), longitude (p < 0.001) and altitude (p < 0.001). The first axis of the

global RDA explained 4.54% of the variance, and was correlated mainly with latitude (-

0.987) and longitude (+0.999). The fourth axis of this global RDA displayed the strongest

correlation with phenology (-0.856), and explained 0.27% of the variance. The partial

RDA model conditioning for the effects of latitude, longitude and altitude, was globally

significant (p < 0.001), as well as the effect of phenology (p <0.001). The axis of the

partial RDA explained 0.29% of the variance and was correlated with phenology (0.922).

All pairwise F_{ST} comparisons were significantly different from zero (Table 2).

Average F_{ST} calculated between island and mainland populations was 0.081. It was on

average 0.003 between close populations within the mainland and 0.005 within the

island. F_{ST} was low, on average 0.003, for each of the four pairs of D- and E- populations,

ranging from 0.0006 between B6D and B6E to 0.0079 between B4D and B4E. The F_{ST}

estimated on the Z chromosome were on average 1.52 times higher than on the

autosomes; 1.62 times higher when comparing populations from the mainland versus

the Corsica Island and 1.44 times higher when comparing neighboring populations in the

island or in the mainland.

Demographic history

When deciphering the historical demography of deciduous and evergreen population

pairs, the hierarchical model choice procedure strongly rejected models of strict

isolation (SI), ancient migration and of panmixia (PAN), which were associated with a

posterior probability of 0 (Supplementary table 3). Instead, models with gene flow were

highly supported. In three of the four pairs, the equilibrium model (EQ) received the

highest posterior probability with P(EQ) = 0.99 and 0.95, 0,94 in the B6D vs B6E, B5D vs

B5E and B4D vs B4E comparisons, respectively. In the A2D vs A2E comparisons, the best

supported model was the isolation with migration model, P(IM) = 0,74 with the second

best model being SC, with P(SC) = 0.23. Comparing the two models against each other

while excluding all remaining models yield unambiguous support for IM, with P(IM)

=0.99 (Supplementary Table S3). Across all models, comparisons with heterogeneous

gene flow and heterogeneous effective population size were not supported, indicating

that, if genetic barriers or linked selection were at play they could not be detected.

Demographic parameters were estimated for each pair of populations under the best

model (Supplementary table 4). Posterior distributions were well differentiated from

their prior indicating that estimated parameters were confidently estimated

(Supplementary Figure 2). Effective population size (N_e) where slightly higher in

deciduous than in evergreen habitats (Ne1 in Table 2, Figure 3A, Supplementary table 4)

and tended to be higher on the mainland than on the island. In a number of

comparisons, our migration estimates reached the prior upper bounds. Therefore, we

ran a new set of simulations with wider priors (Supplementary Note 1). Migration rates

were not different between deciduous and evergreen habitats: in two instances, the

rate of migration was higher from the deciduous to the evergreen and in two other

instances, the reverse was true (Supplementary Figure 3).

Deciphering the historical demography between Corsica and the mainland, the

ancient migration models (AM) strongly outperformed the strict isolation models (SI),

with p(AM) = 0.99 (Supplementary table 5). For the AM model, the simplest model

without linked selection or barriers to gene flow was the best supported, with p = 0.76.

Parameters estimates revealed posterior generally well differentiated from the prior

providing increased confidence (Supplementary Figure 4). Effective population size of

the mainland was 397,685 [CI = 244,375 -- 600,105], around 7.5 times greater than the

one of the island, 52,990 [CI = 33,065 -- 78,475] (Figure 3B, Ne1 in Table 2,

Supplementary table 6). Our analysis also indicated a strong population size change

during the process of population isolation, since the ancestral Ne was estimated at only

240 [CI = 65 -- 685]. Split time was estimated at 1,417,480 [CI = 740,000 -- 2,394,000]

generations ago, hence around 3.2M years ago (assuming a 2.3 years generation time

(Charmantier et al. 2004)) and gene flow subsequently stopped 4540 [CI = 2500 -- 6360]

generations ago, hence around 10,000 years ago, corresponding closely with the end of

the last glacial maximum. The ancestral migration rate between Corsica and the

mainland, from the split in two populations to the end of the latest gene flow between

these populations, was inferred asymmetric, with a 5 times larger migration rate from

the island to the mainland ($m \sim 1.9e\text{-}4$) than the other way around ($m \sim 3.8e\text{-}5$).

However, this asymmetry did not generate a strong difference in gene flow between the

two groups, given the lower effective population size on the island compared to the

mainland, (i.e. respective number of migrants of 10 and 15 from Corsica to the mainland

and in the reverse direction, with overlapping credible intervals, see details in

Supplementary table 6).

Genetic diversity

The mainland populations displayed significantly higher patterns of observed

heterozygosity (on average 0.169, table 2) than Island populations (on average 0.147),

all t-tests' p-values < 0.001. On the contrary there was no significant difference of

observed heterozygosity at smaller spatial scale among populations within the island

(either same or different tree phenology, E- and D-) or within the mainland (all t-tests' p-

value > 0.05). More SNPs were polymorphic in mainland populations (77,306 to 80,616

SNPs for a MAF > 0.05) than in island populations (62,728 to 65,876 SNPs for a MAF >

0.05). The MAF spectrum showed enrichment of variants with smaller frequencies in

mainland populations versus island populations (Supplementary Figure 5A). LD decayed

rapidly in the first 5kb and was lower in populations from the mainland (especially A1D)

compared to island populations (Supplementary Figure 5B). This pattern of rapid LD

decay was similar between chromosomes (see for example chromosomes 1, 2 and Z,

Supplementary Figure 6). Contemporary N_e inferred from LD varied from 142 (in A2E) to

355 (in A1D) (Ne2 in Table 2, Supplementary Figure 5C). N_e values were rather similar

within pairs of E- and D- populations, although Ne was in average smaller in evergreen

populations (245 in average) than in deciduous ones (261 in average). The four largest

N_e (with an average of 318) were found for the four populations monitored on the long

term in forest in which hundreds of artificial nest boxes have been installed for several

decades (the N_e in the 6 other populations was in average of 210). In the entire dataset,

2599 SNPs were identified as non-synonymous while 6751 SNPs were identified as

synonymous. We observed 1.18 times lower allele frequencies for non-synonymous

variants (average MAF = 0.09) than for synonymous ones (average MAF = 0.11) (t.test p-

value < 0.000001). This overall lower frequency of non-synonymous compared to

synonymous mutations was similar between populations from Corsica (1.19) and the

mainland (1.16), as well as between deciduous (1.18) and evergreen (1.19) habitats (See

supplementary table 7). The Z chromosome harbored a proportion of non-synonymous

mutations 1.83 times higher than the average for the autosomes. Finally, we observed

significantly slightly longer ROH on Corsica compared to mainland (mean $_{island}$ = 44,769

kb *vs* mean $_{mainland}$ = 41,586 kb, Wilcoxon rank sum test W = 32,695, p <2e-16,

Supplementary Figure 7&8). We also observed significant differences in the count of

ROH among populations from the mainland versus those from the island but not among

populations within the mainland or the island (ANOVA p <2e-16; Supplementary Table 8

for Tukey HSD test).

Footprints of selection

Manhattan plots of F_{ST} per SNP revealed no clear evidence for high peaks of F_{ST} for the

five comparisons considered (First panels of Figures 4 & 5). Throughout the five

Bayescan tests (Corsica vs mainland populations and the 4 D-4 population pairs), we

identified 40 SNPs with a log10(BF) > 0 among which 18 SNPs had a log10(BF) > 1

(Supplementary table 9, Supplementary Figure 9). None of these SNPs were detected

twice among the different tests conducted with Bayescan. Among these 40 SNPs, 27

were found for the Corsica vs mainland test, 2 were found for A2D vs A2E, 9 were found

for B4D vs B4E, 0 were found for B5 vs B5E and 2 were found for B6D vs B6E. Among

these 40 SNPs, 19 genes were identified (Supplementary table 9). Sliding windows of F_{ST}

along the genome showed a few outlier windows of modest intensities (last panels of

Figure 4 & 5), with almost no parallelism between the tests (Supplementary Table 10).

Expectedly, outlier windows found for the Corsica *vs* mainland comparison depicted

larger F_{ST} than for deciduous vs evergreen comparisons. The outlier F_{ST} windows falling

in the top 1% of the F_{ST} windows distribution are reported in Supplementary Table 10.

Using the RDA to identify outlier SNPs with extreme loading values to axes, we identified

229 outlier SNPs associated to habitat type (deciduous vs evergreen), and 227 SNPs

associated to geography (longitude & latitude, essentially representing a Corsica-

Mainland comparison) (Supplementary table 11, Supplementary Figure 10). For the

deciduous vs evergreen test, only one of the RDA outliers was also found outlier in

Bayescan tests. For the Corsica-mainland test, 8 of the RDA outliers were also found

outliers in the Bayescan test. Genes in which the outliers were found are reported in the

supplementary table 13.

Variation of genomic differentiation with recombination rate

F_{ST} between mainland and Corsican populations was negatively correlated to

recombination rate (Figure 6A, linear model p < 2e-16; Supplementary Figure 11). In

contrast, F_{ST} between each pair of deciduous and evergreen populations was not

correlated to recombination rate. F_{ST} outlier SNPs between Corsica and the mainland

populations and identified by Bayescan or by the RDA were more often found in regions

of low recombination (χ2 test p-values < 0.01, Figure 6C) than observed for the entire

SNPs (Figure 6B). Average recombination rate was on average twice lower for these F_{ST}

outlier SNPs compared to the rest of the SNPs (t-test p-values < 1e-6). In contrast, F_{ST}

outlier SNPs between deciduous and evergreen populations were not preferentially

found in regions of low recombination (Figure 6D).

Genomic inversions

We detected one putative inversion on chromosome 3, spanning 2.8Mb, from position

11,838,789 to 14,661,550, and containing 390 SNPs. The estimated recombination rate

was on average 9 times lower (t-test: p<2.2e-16) in this region of the chromosome

compared to the rest of the chromosome (Figure 7A). However, the SNP density did not

appear reduced at the location of this putative inversion (Figure 7A), suggesting a recent

drop of recombination (*i.e.* dissimilar to what is expected in a peri-centromeric

location). We did not find evidence for an increase of read depth at specific loci in the

putative inversion or its vicinity in inverted individuals compared to normal ones. We

did however notice the absence (read depth = 0) of 3 loci from the inversion, in the four

inverted individuals. The sliding PCA detected either 2 groups (individuals from Corsica

and from mainland) outside of the putative inversion, or 4 groups at the putative

inversion location, separating individuals from Corsica and 3 groups for mainland

individuals (Figure 7B). We assumed that the inverted segment was the one with the

lower frequency on the mainland and that was absent from Corsica. The Lostruct

analysis confirmed for this same region the presence of four successive blocks of 100

SNPs highly discriminating individuals (Figure 7C). The average F_{ST} between the three

groups of mainland individuals was much higher (t-test: p<2.2e-16, Figure 7D) for SNPs

in the putative inversion (F_{ST} = 0.26) than for SNPs outside of this region (F_{ST} = 5.8e-4). π

was lower (t-test: p=1.9e-5, Figure 7E) for SNPs located in the putative inversion for

inverted homozygous individuals (π = 2.5e-5) compared to non-inverted homozygous

individuals (π = 5.3e-5). Both inverted and non-inverted homozygous individuals had

lower π at this inversion compared to heterozygous individuals at this region (π = 7.6e-

5) and compared to the three types of individuals for the rest of the chromosome 3 (π

ranging from 7.4e-5 to 7.6e-5). The PCA (Figure 7F) and the admixture analysis (Figure

7G) using the SNPs from the inversion clearly delineated inverted, non-inverted and

heterozygous individuals, with heterozygous individuals falling at intermediate

eigenvalue between the two categories of homozygous individuals. LD was higher

(Figure 7H) for the region of the putative inversion (Little LD was found when only

homozygous non-inverted individuals were kept in the analysis). For the region of the

inversion, Dxy was 0.0020 between normal blue tits from the mainland and from Corsica

as well as between great tits from the mainland and from Corsica. Dxy was 0.0044

between inverted and normal blue tits from either the mainland or from Corsica, and

ranged from 0.0149 to 0.0152 for interspecific comparisons. Using the formula T = Dxy /

2μ, the inversion likely appeared approximately 220,000 generations ago (around

506,000 years ago). The tree of genetic distance illustrated the divergence of inverted

blue tits from non-inverted blue tits from both mainland and Corsica (Figure 7i). This

region contains 21 genes listed in supplementary table 12. Out of 113 individuals from

the mainland, we found 4 inverted homozygous, 23 heterozygous, and 86 non-inverted

homozygous, hence we observed no deviation from the Hardy-Weinberg equilibrium ($\chi 2$

test p-value > 0.1). The percentage of each genotype was similar across all mainland

locations and both habitat types (the inverted segment was at 11% in A1D, 16% in A2D,

17% in A2E, and 14% across these 3 locations).

Gene ontology

None of the gene lists gathered with the different tests (each outlier test among

deciduous and evergreen environment and between Corsica and the mainland and the

inversion test) yielded any significantly enriched GO term after correction multiple

testing (Supplementary table 14). The most promising GO (uncorrected p.value <0.0025)

included: i) for the deciduous-evergreen outlier tests, GO:0060385, axonogenesis

involved in innervation; GO:2001013, epithelial cell proliferation; GO:0007194, negative

regulation of adenylate cyclase; GO:0090647, modulation of age-related behavioral;

GO:1901379, regulation of potassium ion transmembrane transport, ii) for the Corsica-

Mainland outlier tests, GO:0008344, adult locomotory behavior; GO:2001224, positive

regulation of neuron migration, iii) for the inversion, GO:0016446, somatic

hypermutation of immunoglobulin; GO:0045910, negative regulation of DNA

recombination; GO:0006298, mismatch repair; GO:0008340, determination of adult

lifespan.

Discussion

In this study, we investigated demographic history and genome wide patterns of genetic

diversity and differentiation between several populations of blue tit presenting strong

phenotypic differences between deciduous *versus* evergreen forest habitats as well as

insular *versus* mainland contexts. Between populations in deciduous and evergreen

forest habitats, demographic modelling showed large gene flow and large effective

population sizes (Ne), explaining the low genetic differentiation between these

populations. Demographic modeling also revealed that each pair of deciduous and

evergreen populations most likely never diverged and maintained high connectivity

through migration. We identified slight and mostly unrepeated footprints of divergent

selection across these evergreen and deciduous population pairs, which is consistent

with their demography and the likely polygenic nature of several traits implicated in

their local adaptation. In both insular and mainland populations, we found large Ne,

although smaller for insular populations than mainland ones, resulting in different

distributions and lengths of ROH. Gene flow between Corsica and the mainland most

likely stopped at the end of the last glaciation. Both large Ne and gene flow until the end

of the last glaciation hence explained moderate genome-wide differentiation. We

identified several genomic footprints of selection, enriched in regions of low

recombination only in the case of the mainland/island divergence. Finally, we identified

a putative genomic inversion spanning 2.8Mb, polymorphic in mainland populations

only. We discuss these results in the context of the vast scientific knowledge acquired

on these blue tit populations during the last four decades and more generally in the

context of genomics of adaptation with gene flow.

Divergence between populations in deciduous *versus* evergreen habitats

Although we found a significant effect of habitat on genetic structure, the genetic

differentiation between neighbouring deciduous and evergreen populations was low

(F_{ST} ranging from 0.0006 to 0.0079). This result is in line with the primary observations

realized earlier on a smaller set of populations (Porlier et al. 2012b; Szulkin et al. 2016).

Accordingly, we found high gene flow from deciduous to evergreen populations. Yet,

this quantification of high gene flow and low genetic structure contrasted with the

demographic knowledge collected on blue tit. Indeed, demographic studies suggested

restricted dispersal between these populations, with 4 dispersal events observed

between B4D and B4E (5.6 km apart) and none between the B3E and either B4D or B4E

(24.1 km), among a total of 2788 males, 2672 females and 25,158 nestling ringed in the

three main Corsican sites between 1976 and 2018, with a mean recruitment rate of 6%

(Charmantier, com pers). Our interpretation of this contrast between gene flow

estimations gathered from population genomic *versus* recapture data is quadruple. First,

dispersal estimation on the field using capture-mark-recapture is very challenging and

may require more data than currently collected, to detect rare dispersal events, even

though these affect population genetic parameters. Moreover, since natal dispersal in

blue tit classically ranges between 330 m and 4 km (see (Tufto et al. 2005; Ortego et al.

2011)), the long-term monitoring sites in Corsica equipped with nest-boxes (black circled

dots in Figure 1C) are not ideally spaced to identify the origin of immigrants and the

destination of emigrants, and only a small fraction of the landscape favourable to blue

tit breeding is covered. Second, only a few migrants are sufficient to decrease the

genetic distance between populations, measured through the F_{ST} (Marko & Hart 2011;

Cayuela et al. 2018). In that regard, our results may be compatible with the few

dispersal events recorded throughout the years. Third, it is important to note that the

number of migrants estimated using demographic analyses represents an average over

historical time scales on the order of Ne generations and may have varied widely during

contemporary times. Fourth, the large effective population size found both using

coalescence and a LD method might be explained by the existence of large

"metapopulations" connected by high gene flow and such large Ne might largely

contribute to limit the genetic divergence between populations.

Ne was on average slightly larger in deciduous than in evergreen populations.

This could be explained by the higher productivity of deciduous forests compared to

evergreen forests, resulting in larger clutches and more fledglings in deciduous habitats

(Table 1 in Charmantier et al 2016). However, the very high gene flow between

deciduous and evergreen populations and the low genetic differentiation between these

populations limits further interpretations. In addition, populations monitored on the

long term, for which hundreds of artificial nest boxes have been installed, tended to

have larger Ne than non-monitored populations. Breeding density in the nest-box areas

studied (Figure 1A) was around 1 to 1.3 pairs per ha (Blondel et al 2006), which is most

probably 3 to 5 times higher than natural densities for blue tit when these secondary

hole-nesting birds rely on natural cavities only. It is hence possible that the recent

availability of nest boxes locally increased productivity, effective population size and

heterozygosity.

Demographic modeling indicated that secondary gene flow between previously

isolated populations, which could have eroded most genome-wide differentiation

outside regions implicated in local adaptation and/or acting as reproductive barriers in

particular in low recombining areas (Bierne et al. 2013), was very unlikely. Secondary

contacts were strongly rejected in favor of an island model supporting ongoing gene-

flow without divergence (EQ model) in 3 out of 4 population pairs, the last one

supporting an isolation with migration model (IM). As is often the case, our

demographic estimates should be interpreted cautiously, particularly in the absence of a

well-documented mutation rate in blue tit. The relatively narrow credible intervals

around parameter estimates nevertheless provide confidence in their biological

relevance. These results suggest that the focal population pairs occupying

deciduous/evergreen habitats have been continuously connected by relatively high gene

flow. Hence, genetic divergence between populations in each pair did not originate or

build up on pre-existing genetic divergence accumulated during an allopatric phase but

rather is supported as a case of genuine recent ecological divergence (Pinho & Hey

2010; Wang & Bradburd 2014).

We did not find strong footprints of divergent selection between any of the four

pairs of deciduous and evergreen populations (Figure 4). Moreover, the observed

outliers were not found repeatedly across the 4 deciduous-evergreen comparisons but

rather each outlier was found only once, and they were not enriched in regions of low

recombination. First, this result is in line with the high migration rates observed. High

gene flow indeed most likely limits the potential for local adaptation since it limits the

accumulation of allelic differentiation, even with high selection coefficients (Lenormand

2002). Second, this pattern is consistent with the best demographic model being the

equilibrium model and not a secondary contact. Indeed, the later would more often

create repeated and strong outliers located in regions implicated in reproductive

isolation between divergent populations (eg (Rougemont et al. 2017)). Third, this

pattern is congruent with a model of local polygenic adaptation involving a transient

genetic architecture with multiple alleles of small effects underlying (multiple)

quantitative characters (Yeaman & Whitlock 2011; Yeaman 2015) and displaying low F_{ST}

among loci under divergent selection. Fourth, most documented traits that are involved

in the adaptation of blue tit to the deciduous *versus* evergreen habitat types, such as

clutch size or laying date (Lambrechts et al. 1997; Blondel et al. 1998) are quantitative

and most likely rely on a polygenic architecture with alleles of small effects (see e.g.

(Santure et al. 2013)). However, some statistical issues may limit the interpretation of

this result. First, currently available genome-scan methods to detect alleles of small

effect are statistically limited, especially when applied to relatively small datasets

(Rockman 2012; Hoban *et al.* 2016). Second, a potential lack of power in detecting

outliers could arise from the combination of the use of RADsequencing and a rapid LD

decay along the genome, that may result in too low resolution especially in regions with

larger recombination rates (Lowry et al. 2017). Further analyses of the potential outliers

found here, as well as a new analysis with higher marker density and more individuals,

are needed to better document and discuss the potential genes and biological processes

implicated. Particularly, quantitative genomics will be useful to establish links between

the level of divergent selection on these putative targets and their effect size on

phenotypic trait variation (Stinchcombe & Hoekstra 2007; Gagnaire & Gaggiotti 2016).

This would notably reveal whether genes under divergent selection are also responsible

for the observed phenotypic variation, and contribute to assessing of how much of

phenotypic variation is adaptive.

Last, we searched for inversions potentially associated with phenotypic variation

and/or segregating in both habitats. We report multiple evidences (Figure 6) for a

putative inversion on chromosome 3 in mainland populations. However, the proportion

of inversions in the genomes of deciduous- *versus* evergreen-breeding birds did not

differ, which most probably precludes a putative role of the inversion in local adaptation

in these habitats. Besides, this polymorphism followed the Hardy-Weinberg equilibrium

and we found 4 homozygous inverted individuals, suggesting this inversion does not

involve an accumulation of lethal recessive deleterious alleles (Jay et al. 2019). The

functional consequences of this inversion on variation in life history traits in blue tit call

for further investigations, as achieved in other songbirds (Tuttle et al. 2016; Knief et al.

2017; Kim et al. 2017). In order to genotype this inversion in more birds and to link it to

putative phenotypic variation, we could use a PCR-RFLP approach (da Silva et al. 2019).

Divergence between blue tit populations in mainland *versus* insular contexts

Our demographic modelling approach of Corsican and mainland populations revealed

that a model with ancient migration was the most probable, which is coherent with the

history previously reconstructed for the blue tit complex. Split time of the ancestral

populations in two mainland *versus* island populations connected by gene flow (~3M

years) was relatively coherent with the diversification time found in the literature for

the entire blue tit complex (5M (Illera et al. 2011)). Gene flow between the island and

the mainland likely stopped around 10,000 years, which is compatible with a gene flow

break due to the rise of the flooded stretch between Corsica and the mainland during

ice melting and the sea level rise after the last glaciation, during from 17,000 to 5,000

years ago (Lambeck & Bard 2000; Jouet et al. 2006). Both the signal of population

expansion during the diversification period, and the gene flow five times larger from the

island to the mainland than from the mainland to the island, were coherent with a

suspected recolonisation of the mainland from the islands, as proposed by Illera et al.

(2011) based on analyses of nuclear and mitochondrial DNA sequences. The larger

effective population size on the mainland than on the island may be due to multiple

sources of colonisation on the mainland (Taberlet et al. 1992) and to the much larger

mainland area and hence metapopulation size compared to the island. Expanding this

study with a sampling from the South-East of Europe, one from Sardinia and one from

the Iberian Peninsula, could help determine whether the large size estimated for the

mainland meta-population studied here might be explained by a mixed recolonisation

from these refugium (Kvist et al. 1999; 2004) and whether insular populations

consistently have reduced effective population size. It would also be interesting to

explore a model allowing for multiple cycles of isolations and contacts that could have

resulted from successive glacial cycles (Hewitt 2004).

Given the demographic parameters inferred, it is not surprising that the genetic

differentiation between populations from the mainland and the island was moderate

(F_{ST} = 0.08). However, this differentiation value contrasted with the much larger

divergence at mitochondrial DNA observed for these populations (OST = 0.67 between

A1D and B4D (Kvist et al. 2004)). This discrepancy between whole nuclear genome and

mitochondrial estimates of differentiation could first be explained by the typically four

times smaller Ne in mitochondrial DNA compared to nuclear DNA in diploid organisms,

resulting in larger drift and divergence (Smith & Klicka 2013). Second, accumulation of

cytonuclear incompatibilities could limit its introgression (Burton & Barreto 2012).

Alternatively, gene flow may have been greater for males than females during the

colonisation process from Corsica to the mainland, at the end of the last glacial period.

However, this hypothesis is not supported by the general observation that females

disperse on average further away than males and that rare long distance dispersals are

also achieved by females (Tufto et al. 2005). Inspecting mitochondrial DNA variation for

the birds studied here would be very useful to compare mitochondrial differentiation in

our sample to the previous analysis which also included Corsican birds (Kvist et al. 2004).

In any case, one should bear in mind that our results stem from integrating coalescent

patterns observed across thousands of loci, therefore providing increased resolution to

investigate the determinants of demographic divergence compared to approaches

based on fewer mitochondrial and nuclear loci.

The contemporary demography inferred between the populations from Corsica

and mainland France, especially the absence of gene flow for approximately 4500

generations and the large effective population size, likely provides a suitable context for

the build-up of local adaptation. Moreover, the level of divergence (F_{ST} = 0.08) was

probably low enough to detect divergent outlier loci potentially implicated in local

adaptation. We identified several outlier SNPs and outlier 200 kb windows showing

elevated differentiation, more often in regions of lower recombination than elsewhere

in the genome (contrary to the outliers found for the deciduous vs evergreen

populations). Furthermore, F_{ST} was on average higher in these regions of low

recombination compared to elsewhere in the genome (which was not the case for

deciduous vs evergreen populations). These patterns of increased divergence in regions

with low recombination between divergent populations are commonly observed in

other species (Nachman & Payseur 2011; Cruickshank & Hahn 2014; Gagnaire et al.

2018), including other bird species (Burri et al. 2015; Spurgin et al. 2019). It is well

documented that such increased differentiation in regions with low recombination is

not necessarily due to positive selection, or at least not alone, and that it is largely

influenced by the effect of recombination in interaction with background selection

(Charlesworth et al. 1993; Burri et al. 2015; Perrier & Charmantier 2019). Moreover, a

uniform F_{ST} sliding window, sized in Kb, is expected to dilute signatures of selection in

regions of the genome where LD is fast decaying while it is expected to over-represent

regions where LD decays much more slowly, hence increasing even more the potential

false positives in regions with low recombination (Beissinger *et al.* 2015; Perrier &

Charmantier 2019). Although perfectible, our sliding window approach using a SNP unit

attempted to fix this issue and successfully captured several new outlier regions outside

of deserts of recombination. This second window approach should however be

improved, for example by estimating the local neutral enveloped, which computation

should integrate local variations in LD, recombination, diversity, but also the

demography of populations. As mentioned for the study of adaptive divergence

between deciduous and evergreen populations, complementary analyses integrating

both genome scans and quantitative genomics would improve our comprehension of

the genomic and phenotypic divergence observed between Corsican and mainland blue

tits (Stinchcombe & Hoekstra 2007; Gagnaire & Gaggiotti 2016).

While the putative genomic inversion on chromosome 3 was absent from Corsica

and detected in mainland individuals, its level of divergence from the non-inverted

sequence indicated that it was likely twice older than the beginning of divergence

between blue tit populations from mainland and Corsica. This could first suggest that

this polymorphism emerged in mainland blue tit populations and then did not introgress

the Corsican populations, maybe due to a local disadvantage and/or genetic

incompatibilities or simply due to drift coupled to little gene flow. A second hypothesis

could be that this inversion was present in Corsican populations but had been purged

out due to a local disadvantage. Such a disadvantage can be due to the typical

accumulation of deleterious mutations in such non-recombining inverted sequences

(*e.g.* Jay et al 2019). Lastly, it is possible that this inversion has been acquired in

mainland populations only recently, after the last period of contact between mainland

and Corsican populations, *via* gene flow from a distinct refugium with which

connectivity could have been enhanced after deglaciation, hence after the stop of gene

flow with Corsica. The age of this inversion, its origin, its biological effects and the

potential accumulation of deleterious mutations need to be inferred more thoroughly

via genotyping full inversion sequences from individuals in diverse locations and using

advanced statistical methods (Lohse *et al.* 2015).

Conclusion and perspectives

Our study demonstrated the usefulness of demographic modeling and of the analysis of

the variation of genomic diversity and recombination along the genome to uncover the

genetic determinants of local adaptation in a small passerine with a large distribution,

and occupying different forest habitats. Especially, demographic modeling rejected the

hypothesis of a secondary contact between deciduous and evergreen populations and

favoured a situation with continuous gene flow. These results support the idea that blue

tits have adapted to their habitats despite ongoing gene flow, while contextualising how

large gene flow most probably constrained local adaptation (Lenormand 2002) and

favoured its architecture based on alleles of small effects (Yeaman 2015). The genomic

modelling also refined our knowledge about the divergence between insular and

mainland meta-populations, that have been likely unconnected by gene flow for the last

ten thousand years. We also verified the relationship between local recombination rate

and differentiation, that should probably be integrated in genome scans looking for

footprints of selection (Beissinger *et al.* 2015; Burri *et al.* 2015; Berner & Roesti 2017;

Perrier & Charmantier 2019, Booker et al 2020). Future investigations will require

increased sample sizes and marker density (Lotterhos & Whitlock 2015; Hoban *et al.*

2016) in order to better detect loci with small effects that contribute to the quantitative

phenotypic variation and local adaptation in blue tit. Lastly, the putative inversion found

here would need further analyses since this type of structural variation is often

implicated in phenotypic variation (Kirkpatrick 2006; 2010; Stapley *et al.* 2017;

Wellenreuther *et al.* 2019).

 Tables

 Table 1. Study populations' names, abbreviations, habitat types, locations, approximate forest area (ha), altitude (m), and sample

 sizes.

Name	Abbreviation	Region	Site	Type of oaks	Latitude	Longitude	Forest area	Altitude	N-GeneralDataset	N-FilteredDataset
D-Rouviere	A1D	Mainland	Rouviere	Deciduous	43.664000	3.668000	465	248	203	60
D-Quissac	A2D	Mainland	Quissac	Deciduous	43.891036	3.996931	23	104	35	27
E-Quissac	A2E	Mainland	Quissac	Evergreen	43.926673	3.989759	415	116	31	24
E-Pirio	B3E	Corsica	Pirio	Evergreen	42.376000	8.750000	400	145	180	60
D-Muro	B4D	Corsica	Muro	Deciduous	42.551000	8.923000	188	283	215	60
E-Muro	B4E	Corsica	Muro	Evergreen	42.589000	8.967000	80	99	116	60
D-Zilia	B5D	Corsica	Zilia	Deciduous	42.523685	8.873689	13	102	42	38
E-Zilia	B5E	Corsica	Zilia	Evergreen	42.520633	8.884057	17	267	38	38
D-OlmiCappella	B6D	Corsica	OlmiCappella	Deciduous	42.540078	8.993915	20	1027	44	44
E-OlmiCappella	B6E	Corsica	OlmiCappella	Evergreen	42.529414	9.021652	25	881	43	43
Total/average									947	454

Table 2. Diversity and differentiation across populations. Fixation index between populations (F_{ST}), effective population size (Ne) estimated with ABC (Ne1) using models for Deciduous-Evergreen population pairs or for Corsica-Mainland (A1D-B4D), Ne estimated with a LD method (Ne2), and observed heterozygosity (Ho). Color gradients indicate differentiation intensity (Darker red for higher differentiation and darker blue for lowest differentiation). Underlined values indicate deciduous and evergreen pairs.

| | | | | F_{ST} | | | | | | | Ne1 | | Ne2 | Ho |
	A1D	A2D	A2E	B3E	B4D	B4E	B5D	B5E	B6D	B6E	Deciduous-Evergreen	Corsica-Mainland		
A1D		0.0045	0.0043	0.0812	0.0830	0.0853	0.0796	0.0804	0.0785	0.0784	Na	397,685	355	0.171
A2D	0.0045		0.0029	0.0832	0.0849	0.0872	0.0799	0.0810	0.0793	0.0792	400,005	na	161	0.167
A2E	0.0043	0.0029		0.0828	0.0846	0.0871	0.0796	0.0807	0.0790	0.0787	100,055	na	142	0.168
B3E	0.0812	0.0832	0.0828		0.0058	0.0089	0.0067	0.0071	0.0056	0.0058	Na	Na	334	0.149
B4D	0.0830	0.0849	0.0846	0.0058		0.0079	0.0063	0.0067	0.0061	0.0062	43,070	52,990	310	0.149
B4E	0.0853	0.0872	0.0871	0.0089	0.0079		0.0093	0.0097	0.0083	0.0085	37,100	na	271	0.148
B5D	0.0796	0.0799	0.0796	0.0067	0.0063	0.0093		0.0025	0.0035	0.0032	46,520	na	210	0.145
B5E	0.0804	0.0810	0.0807	0.0071	0.0067	0.0097	0.0025		0.0036	0.0033	42,800	na	216	0.146
B6D	0.0785	0.0793	0.0790	0.0056	0.0061	0.0083	0.0035	0.0035		0.0006	46,885	na	268	0.146
B6E	0.0784	0.0792	0.0787	0.0058	0.0062	0.0085	0.0032	0.0033	0.0006		162,940	na	260	0.146

 Figures captions

 Figure 1. Map of the sampling locations of blue tit populations on the mainland and in

 Corsica (A) and phenotypic differences (B). In red, habitats dominated by deciduous

 oaks and in green, habitats dominated by evergreen oaks. Sites with long-term

 monitoring are circled in black on the map (A1D is D-Rouvière, B3E is D-Pirio, D4D is D-

 Muro and B4E is E-Muro). On figures B & C are presented illustrative phenotypic

 differences (with standard deviations) between the four main populations with long

 term monitoring. Are shown laying date (days), clutch size and male tarsus length (mm)

 (data from Table 1 in Charmantier et al 2016 Evol Appl, see this reference for detailed

 information for these measures and for differences in many other traits).

 Figure 2. Population structure between populations. A) Admixture for K = 2, 6 and 10

 (Vertical lines correspond to individual admixture; colors correspond to distinct genetic

 groups). **B)** RDA showing the influence of latitude, longitude, altitude and phenology on

 genetic structure.

 Figure 3. Demographic parameters A) for each of the four pairs of blue tit populations

 in deciduous and evergreen habitats, estimated using equilibrium models, and **B)** for the

 divergence between populations on the mainland and Corsica, estimated using an

 ancient migration model. In panel A, circle size is proportional to effective population

 size (Ne) and arrow width is proportional to migration rate. In panel B, rectangle width is

 proportional to log10 (effective population size), arrow width is proportional to

migration rate, and split time and time of ancient migration are indicated in number of

generations.

Figure 4. F_{ST} Manhattan plots between the 4 population pairs in deciduous and

evergreen forests. F_{ST} are given either by SNP (four first graphs), by 200kb windows

(graphs 5 & 6) or by 50 SNPs windows (graphs 7 & 8). For graphical simplicity, the 3 pairs

of populations in Corsica were grouped for the sliding window F_{ST} Manhattan plots (BD

vs BE). Dots alternate colors between chromosomes. F_{ST} for unplaced loci are shown at

the end of each of the first four Manhattan plots.

Figure 5. F_{ST} Manhattan plots between Corsica and the mainland. F_{ST} are given either

by SNP, by 200kb windows or by 50 SNPs windows. Dots alternate colors between

chromosomes. F_{ST} for unplaced loci are shown at the end of the first Manhattan plot.

Figure 6. Relationship between recombination rate and divergence. A) Correlation

between local recombination rate and F_{ST} between populations on the mainland and in

Corsica; **B)** SNP frequency per class of recombination rate; **C)** Outlier SNP frequency for

the mainland-Corsica comparison, per class of recombination rate; **D)** Outlier SNP

frequency for the deciduous-evergreen comparisons, per class of recombination rate. In

panels C & D, outliers are given for the Bayescan and the RDA tests.

Figure 7. Detection and characteristics of a putative inversion on chromosome 3. A)

Recombination rate and SNP density along chromosome 3; B) individual values for

eigenvector 1 of a PCA of all the blue tit individuals from Corsica and the mainland; C)

MDS 1 values from Lostruct analysis; D) F_{ST} between inverted, non-inverted and

heterozygous individuals from the mainland; E) Nucleotide diversity for inverted (in red),

non-inverted (in green) and heterozygous individuals (in blue) from the mainland; F)

PCA results for the region of the putative inversion for all the blue tit individuals; G)

Mainland blue tits' admixture proportions for K = 2 for the region of the putative

inversion (individual ordered by decreasing ancestry value for the inverted cluster); H)

Linkage disequilibrium for SNPs found in the putative inversion (plus 1 Mb each side of

the inversion), for mainland blue tit; I) genetic distances at the inversion, between 4

inverted blue tits, 8 non-inverted blue tits, and 4 great tits (sites of capture are indicated

for each individual).

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Authors' contributions:

CP conceived the study, conducted field-work, lab analyses, bioinformatics and

statistical analyses, interpreted the results and wrote the manuscript. QR conducted

bioinformatics and statistical analyses (especially demographic analyses using ABC),

interpreted the results and edited the manuscript. AC obtained the ERC grant, conceived

the study, performed field-work, interpreted the results and edited the manuscript.

Data accessibility:

Individual sequences with metadata are available on NCBI with the SRA accession

PRJNA630135. A filtered VCF file with metadata are available on Dryad with the

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1284

A-Admixture

K=2
CV=0.302

K=6
CV=0.307

K=10
CV=0.314

A1D A2D A2E B3E B4D B4E B5D B5E B6D B6E

B-RDA

Mainland Corsica
A1D   B3E
A2D   B4D
A2E   B4E
          B5D
          B5E
          B6D
          B6E

PHENOLOGY

LATITUDE   LONGITUDE

ALTITUDE

RDA2 / RDA1

ALTITUDE

PHENOLOGY

RDA4 / RDA3