On deleterious mutations in perennials: inbreeding depression, mutation load and life-history evolution

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Abstract. In Angiosperms, perennials typically present much higher levels of inbreeding depression than annuals. The mechanisms leading to this pattern are poorly understood. In fact, despite the potential significance of this pattern for important evolutionary questions, only two hypotheses have been proposed to explain it. Based on the fact that mutations occurring in somatic tissues may be passed onto the offspring in plants, because they do not have a segregated germline, the first hypothesis states that more long-lived species may accumulate more somatic mutations as they grow, thereby generating higher inbreeding depression. The second hypothesis, which is not in contradiction with the first, stems from the observation that inbreeding depression is typically expressed across multiple life stages in Angiosperms. It posits that increased inbreeding depression in more long-lived species could also be explained by the fact that mutations, regardless of whether they are produced during mitosis or meiosis, may differ in the way they affect fitness in annual and perennial populations, through the life stages at which they are expressed. In this study, we aim to investigate the second hypothesis, setting aside somatic mutations accumulation. We combine a physiological growth model and multilocus population genetics approaches in order to describe a full genotype-to-phenotype-to-fitness map, where the phenotype relates to fitness through biological assumptions, so that the fitness landscape emerges from biological assumptions instead of being assumed a priori. We study the behaviour of different types of mutations affecting growth or survival, and explore their consequences in terms of inbreeding depression and mutation load. Then, we discuss the role deleterious mutations maintained at mutation-selection balance may play in the coevolution between growth and survival strategies.

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1 Introduction

Perennials, which make up the majority of Angiosperms ($\sim 70\%$, Munoz et al., 2016), typically present much higher levels of inbreeding depression than annuals. Indeed, metaanalyses found inbreeding depression to span from $\delta \approx 0.2$ on average in short-lived herbaceous species to $\delta \approx 0.5$ in long-lived herbaceous species and shrubs, and $\delta \approx 0.6$ in woody species (Duminil et al., 2009; Angeloni et al., 2011). Inbreeding depression, defined as the reduction in fitness of inbred relative to outbred individuals, is thought to be mainly due to the increased homozygosity of inbred individuals for recessive deleterious mutations segregating at low frequencies in populations (Charlesworth and Charlesworth, 1987; Charlesworth and Willis, 2009). Yet, why inbreeding depression is higher in more long-lived species is poorly understood. In fact, only two hypotheses have been proposed 10 to explain this pattern, despite the potential significance of this pattern for important evolutionary questions, such as mating systems or dispersal rates evolution (Barrett and 12 Harder, 1996; Roze and Rousset, 2005; Epinat and Lenormand, 2009; Duputié and Massol, 13 2013), and for more applied issues, as many cultivated species are perennial (e.g. fruit trees in general) and efforts are being made to develop perennial grain crops (DeHaan and 15 Van Tassel, 2014). 16 The first hypothesis was formally put forward by Scofield and Schultz (2006). In plants, mutations occurring in somatic tissues may be passed onto the offspring, because they do 18 not have a segregated germline. Thus, Scofield and Schultz (2006) proposed that more 19 long-lived species may accumulate more somatic mutations as they grow and transmit them to their offspring, thereby generating the increase in inbreeding depression observed in such species. Phenotypic data in a long-lived clonal shrub (Vaccinium angustifolium,

Bobiwash et al., 2013), and genomic results in Quercus robur (Plomion et al., 2018) demonstrated that some somatic mutations can indeed be passed onto the offspring. However, the number of detected heritable somatic mutations is low (Schmid-Siegert et al., 2017; 25 Plomion et al., 2018), and recent studies have concluded that the number of cell divisions from embryonic cells to gametes production may be much lower than previously thought 27 (Burian et al., 2016; Watson et al., 2016; Burian et al., 2016; Schmid-Siegert et al., 2017; 28 Lanfear, 2018), due for instance to early specification and quiescence mechanisms of axillary meristems cells, resulting in little opportunity for heritable mutations to accumulate 30 during plant growth. Furthermore, intraorganismal selection is expected to efficiently 31 purge deleterious somatic mutations, resulting in little to no somatically generated mutation load at the population level (Otto and Orive, 1995). Hence, although somatic mutations can be inherited and contribute to the mutation load in plants, their relative significance compared with meiotic mutations remains unclear (Schoen and Schultz, 2019). The second hypothesis stems from the observation that inbreeding depression is typi-36 cally expressed across multiple life stages in Angiosperms (Husband and Schemske, 1996; 37 Winn et al., 2011; Angeloni et al., 2011). It posits that increased inbreeding depression in 38 more long-lived species could be explained by the fact that mutations, regardless of whether they are produced during mitosis or meiosis, may differ in the way they affect fitness in annual and perennial populations, through the life stages at which they are expressed. Most theoretical studies of the mutation load focused on the case of mutations affecting 42 fitness on a strictly linear fitness landscape (that is, fitness is the trait, e.g. Charlesworth et al., 1990; Roze, 2015), or through an abstract trait (or set of traits) associated with a gaussian fitness landscape (e.g. Roze and Blanckaert, 2014; Abu Awad and Roze, 2018).

In these cases, inbreeding depression in annual and perennial populations is not expected to differ (Charlesworth, 1980). On the other hand, the dynamics of mutations affecting other aspects of individuals' life cycle, such as survival or growth, were seldom investi-48 gated. Morgan (2001) investigated the dynamics of mutations affecting survival between mating events in a perennial populations. They concluded that inbreeding depression should sharply decrease as life expectancy increases, and even become negative for long-51 lived species (outbreeding depression). However, Morgan (2001) studied mutations with a strong effect on fitness, and assumed no age-structure, that is, individuals did not differ in 53 fecundity or survival probability with age. Strong variations in both survival and fecundity with respect to age are yet observed in perennials. Indeed, while juveniles typically suffer from very high mortality rates, established individuals tend to experience rather low mortalities with particularly slow senescence (Petit and Hampe, 2006). Furthermore, 57 fecundity usually increases dramatically with age in perennials (Franco and Silvertown, 1996), due to the positive scaling of reproductive output with size in plants (Klinkhamer 59 et al., 1985; Weiner et al., 2009). Mutations slowing their bearer's growth could there-60 fore play a role in generating higher inbreeding depression in more long-lived species, as growth delays may impact individuals' fecundities differently depending on their age or size. This latter aspect of age-structuration in perennials was, to our knowledge, never tackled theoretically. The present study aims to study the second hypothesis, that is, we investigate the 65 behaviour of mutations affecting fitness differently with respect to life-history, putting 66 aside somatic mutations accumulation. Namely, we study meiotic mutations affecting growth or survival in a partially selfing population, in which individuals grow as they

age and fecundity is proportional to size, but survival between mating events is assumed to not depend on age. We combine a physiological growth model (West et al., 2001) and multilocus population genetics approaches (Barton and Turelli, 1991; Kirkpatrick et al., 2002) in order to describe a full genotype-to-phenotype-to-fitness map, where the fitness landscape emerges from biological assumptions instead of being assumed a priori. We study the behaviour of different types of mutations affecting growth or survival, and explore their consequences in terms of inbreeding depression and mutation load (Crow, 1958). Then, we discuss the role deleterious mutations maintained at mutation-selection balance may play in the coevolution between growth and survival strategies.

2 Model outline and methods

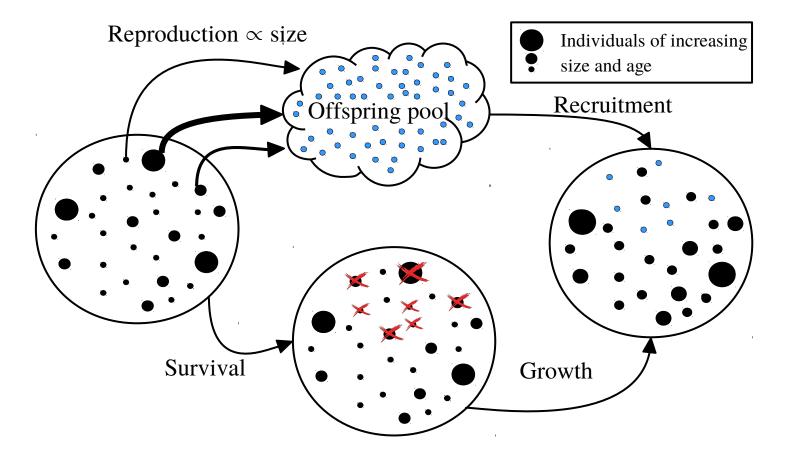


FIGURE 1: Life cycle and demography. Deceased individuals are marked by a red cross, juveniles are depicted in light blue. Larger dots depict larger and older individuals.

Life cycle. We consider a large population of diploid hermaphrodites, which may survive from one mating season to another with a probability S, assumed to be constant with respect to age. If they survive, individuals grow between mating events, following a physiological growth model described briefly in the next paragraph. If they die, juveniles, which are assumed to be produced in large excess compared with the resources available for establishment, are recruited to replace the dead, so that population size is kept constant (Fig. 1). Each juvenile has a probability J of being recruited. During reproduction,

- individuals are assumed to contribute to the gamete pool in proportion to their size (the larger an individual, the larger its contribution to the gamete pool), and to self-fertilise at a fixed rate α .
- Growth model. We consider the growth model developed by West et al. (2001) (see details in Appendix I). The energy available for growth and maintenance at age t, B_t , is assumed to scale as a 3/4-power law of body size as a result of allometry (Peters, 1983), so that

$$B_t = B_0 \ G_t^{3/4},\tag{1}$$

where B_0 is the basal metabolic rate and G_t is body size at age t. This energy can be subdivided into the energy required to maintain the existing body, controlled by a maintenance cost c, and the energy available to produce new body parts, controlled by a production cost ε , so that growth is fully described by the following differential equation

$$\frac{dG}{dt} = \frac{B_0}{\varepsilon} G_t^{3/4} - \frac{c}{\varepsilon} G_t. \tag{2}$$

Under this model, individual size naturally saturates when the energy required to maintain
the existing body equals the available energy (Fig. 2a-2b).

Genetic assumptions. Mutations are assumed to occur at rate U (per haploid genome) at a large number of loci, which recombine at rate $0 \le r \le \frac{1}{2}$. In three separate models, we consider mutations affecting three different traits. Mutations may affect growth by increasing either their bearer's maintenance cost (c) or production cost (ε) , or they may affect its survival. When mutations affect survival, they are assumed to decrease both their

bearer's probability of being recruited as a juvenile (J) and its adult survival probability (S). The effect of mutations is denoted s, with a dominance coefficient $h \leq \frac{1}{2}$. Loci affect traits multiplicatively, so that for any trait z ($z \in \{c, \varepsilon, S\}$), we have

$$z = z_0 (1 \pm s)^{H_o} (1 \pm sh)^{H_e}$$

where H_o (resp. H_e) is the number of homozygous (resp. heterozygous) mutations born by the considered individual.

Approximation of the expected number of mutations, inbreeding depression and mutation load. We use two approaches to study our model. In the first approach, 109 we make the assumption that selective pressures acting on mutations at various life-stages 110 can be summarized into a single lifetime fitness expression, so that the population can 111 be studied as an adequately rescaled annual population (the Lifetime Fitness approach, 112 LF). For each type of mutation, this approach allows us to gain insights into the way 113 selection acts on mutations, by summarising it into a single lifetime selection coefficient. This coefficient is denoted \bar{s}_z (where z = c, ε or S depending on the considered trait). 115 Besides, reasoning in terms of lifetime fitness is paramount to compute key quantities 116 such as inbreeding depression or the mutation load. However, the LF approach fails to account for genetic associations correctly. Thus, in order to obtain approximations of the 118 expected number of mutations per haploid genome accounting for genetic associations, we 119 also study each step of the life cycle successively (the Life Cycle approach, LC). We do so under the assumption that the phenotypic effect of mutations is weak and that the number 121 of segregating mutations is large, following the work of Roze (2015) that we adapted to the case of many mutations affecting a trait rather than fitness directly, in an age- and size-structured population.

Individual-centered simulations were run assuming individuals depicted 125 by two linear chromosomes of length λ (in cM), along which mutations occur stochasti-126 cally (Roze and Michod, 2010). The number of mutations occurring is sampled from a Poisson distribution with mean U, and their position on chromosomes are sampled from 128 a uniform distribution. Recombination is modeled by exchanging segments between chro-129 mosomes. Similar to mutations, the number of crossing-overs is sampled from a Poisson 130 distribution with mean λ , while their position along chromosomes are randomly drawn 131 in a uniform distribution. The population has a constant size N. At each timestep, an 132 individual survives with probability S, which can depend on its genotype in the case of mutations affecting survival. If the individual does survive, it grows deterministically 134 depending on its age and individual physiological growth costs (Fig. 2, Equation A4). 135 If it does not, it is replaced by an offspring generated from the population, in which 136 parents are chosen with a probability proportional to their size. The offspring is pro-137 duced by self-fertilisation with probability α , and by random mating otherwise. We mea-138 sure the average of the trait affected by mutations, the average number of mutations 139 per haploid genome in the population, and inbreeding depression. Inbreeding depres-140 sion is measured as the relative difference in lifetime reproductive success of selfed and 141 outcrossed individuals. Individuals lifetime reproductive success is obtained by counting 142 the number of times they are chosen as parents before they die. The mutation load in 143 the sense of Crow (1958), that is the decrease in mean fitness of the population com-144 pared with a population with no mutations, is measured using the average of the trait

affected by mutations. This average is used to compute the expected mean lifetime fitness
in the population (Equation (A16)). Then, this quantity is compared to the expected
mean lifetime fitness when mutations are absent. All programs are available from GitHub
(https://github.com/Thomas-Lesaffre/On deleterious mutations in perennials).

3 Results

3.1 Mutation-selection equilibrium

The intensity of selection acting on mutations affecting a trait depends on how the change 150 they cause in the trait modifies the fitness of their bearer, and how the fitness of their bearer then compares with the fitnesses of all the individuals they are competing with, 152 that is the distribution of fitness in the population. When mutations segregate at many 153 loci, individuals almost always carry mutations at several loci. Hence, the intensity of 154 selection acting on mutations at a given locus may depend on the genetic composition 155 of the population at other loci, because other loci may affect both the distribution of 156 phenotypes in the population (and therefore of fitness), and how the phenotypic effect of 157 a mutation at the considered locus changes their bearer's fitness. 158 In most population genetics models interested in mutation load dynamics, mutations 159 directly affecting fitness are considered, that is, fitness is the trait. In other words, the 160 phenotype-to-fitness relationship (the fitness landscape) is strictly linear. In that specific 161

directly affecting fitness are considered, that is, fitness is the trait. In other words, the
phenotype-to-fitness relationship (the fitness landscape) is strictly linear. In that specific
case, the genetic composition of the population at other loci does not affect the intensity
of selection at a given locus in the way described above, because mutations always reduce
fitness by exactly the same proportion, irrespective of their bearer's position on the fitness
landscape (Roze, 2015). This is no longer the case as soon as the fitness landscape is

not assumed to be strictly linear. An example of such non-linear fitness landscape is

Fisher's Geometric Model (Fisher, 1930), which depicts at trait (or set of traits) under 167 stabilizing selection on a gaussian fitness landscape centered on the optimal trait value. 168 This model has been considered in mutation load dynamics studies (e.g. Abu Awad and 169 Roze, 2018). However, these studies assumed that mutations have weak additive effects 170 and occur symetrically in both directions (i.e. increasing or decreasing the trait), so that 171 the mean fitness is little affected by mutations, and the intensity of selection is not affected by the genetic composition of the population in this case either. 173 In our model, we do not impose any constraint on the shape of the fitness landscape. 174 Instead, we let the genotype-to-phenotype-to-fitness map arise from biological assump-175 tions. The resulting fitness landscape is non-linear, convex, and has a singularity at the 176 origin (Equation A16 in Appendix III.1, Fig. S1). It is therefore required that we ac-177 count for the change in the distribution of the trait caused by mutations at other loci when 178 quantifying selection at a given locus. When the population is large enough, this can be 179 done by following the change in the trait average as mutations segregate in the population 180

$$\begin{cases}
\Delta p_i(\bar{z}) = 0 \\
\bar{z}(p_i) = z_0 \times e^{\pm s \sum_i \overline{Z}_i} \left(1 + s^2 \sum_{j \neq i} \frac{\overline{\Delta Z_i \times \Delta Z_j}}{2} \right),
\end{cases} \tag{3}$$

In the first line of Equation (3), Δp_i is the change in the frequency of the mutant allele at the i^{th} locus, which depends on the trait average \bar{z} . On the second line, \bar{z} is the average of the trait in the population, which depends on the number of mutation per haploid genome $\sum_i p_i$. On this line, the term $\sum_i \overline{Z}_i$ quantifies the effect on the trait of

(Equation 3, Appendix II).

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the $\sum_i p_i$ mutations per haploid genome born on average by individuals, neglecting the effects of genetic associations between loci, while the term $\overline{\Delta Z_i \times \Delta Z_j}$ quantifies the effect of pairwise associations between loci on the trait average. Associations of higher order are neglected. Solving of Equation (3) for p_i and \bar{z} allows us to obtain predictions for the average of the trait and for the average number of mutations per haploid genome maintained at mutation-selection equilibrium, which we then use to compute the inbreeding depression and mutation load at expected equilibrium.

193 3.2 Fitness effect of mutations neglecting genetic associations

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To leading order, genetic associations can be neglected so that Equation (3) simplifies into

where $F = \frac{\alpha}{2-\alpha}$ is the inbreeding coefficient. The first line of Equation (4) shows

$$\begin{cases}
U - \bar{s}_z(\bar{z}) \left[h \sum_i p_i + (1 - h)F \sum_i p_i \right] = 0 \\
\bar{z}(p_i) = z_0 \times \exp\left[\pm s \left(2h \sum_i p_i + (1 - 2h)F \sum_i p_i \right) \right],
\end{cases} \tag{4}$$

that when we neglect genetic associations between loci, the selective pressures acting on 196 mutations are encapsulated in a single lifetime selection coefficient \bar{s}_z (Appendix III.1.2). 197 These coefficients depend on the population average of the trait, owing to the mechanisms 198 described in the former section. 199 The phenotypic effect of each type of mutation (Fig. 2, top row), and the resulting 200 lifetime selection coefficients are presented in Fig. 2 (bottom row). For clarity, the plot-201 ted \bar{s}_z coefficients were obtained assuming mutations are absent (i.e. $\bar{z}=z_0$), because 202 although they vary quantitatively when mutations segregate in the population, their qual-203 itative behaviour with respect to life expectancy remains unchanged. In other words, the

coefficients plotted in Fig. 2 (bottom row) represent the intensity of selection a mutation

206 would face if a single locus was modeled.

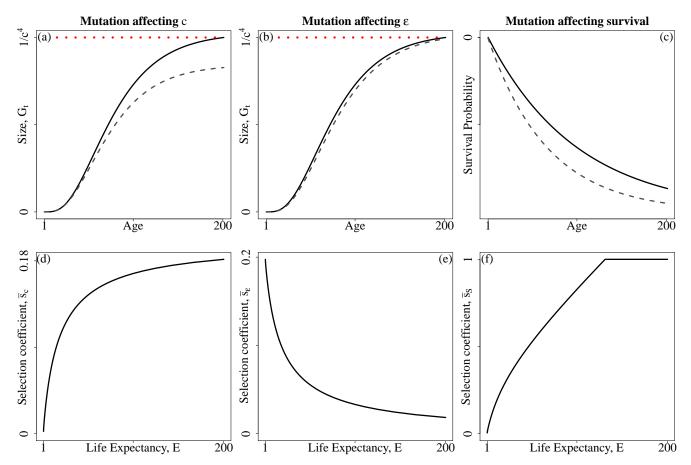


FIGURE 2: Phenotypic and leading order fitness effect of a mutation. Top row: phenotypic effects on growth or survival are presented as a function of age, the dashed line depicts the mutated phenotype while the solid line depicts the unmutated (c=0.001, $\varepsilon=0.01$ and s=0.05 for mutations affecting growth, and S=0.99, s=0.005 for mutations affecting survival). Bottom row: Resulting effects of mutations on lifetime fitness as a function of life expectancy. The dotted red lines depict maximal size. Note that the phenotypic effect of mutations (s) differs between mutations affecting growth and survival.

Mutations affecting the maintenance cost c cause growth delays to increase as individuals age (Fig. 2a). Hence, their fitness effect increases with life expectancy (Fig. 2d). On the contrary, mutations affecting the production cost ε do not affect individuals' maximal size, as they asymptotically tend to the same size, but rather the speed at which they reach it (Fig. 2b). Therefore, the growth delay mutated individuals accumulate in early

years fades away in older individuals. This causes selection against mutations affecting ε to decrease with respect to life expectancy, because they become gradually neutral in 213 older age-classes. Mutations affecting survival cause individuals to perform less mating 214 events in a lifetime. Furthermore, they cause mutated individuals to perform less well during mating events, because they tend to be younger than unmutated individuals (Fig. 216 2c). Thus, age-structure increases selection against mutations affecting survival. Further-217 more, selection against mutations affecting survival strongly increases as life expectancy increases (Fig. 2f), contrary to mutations affecting growth costs, whose fitness effects 219 remain moderate and of the same order of magnitude (Fig. 2d-e). Overall, the results 220 described in Fig. 2 show that mutations affecting different traits on the same genotype-221 to-phenotype-to-fitness map face very different selective pressures, both in magnitude and 222 in the way they vary with life expectancy.

3.3 Average number of mutations, inbreeding depression and mutation load

The intensity of selection acting on mutations does not depend on the values of c and ε , but on the ratio $\frac{c}{\varepsilon}$ (Appendix III.1.2). Thus, in what follows, results are described using this ratio. FIGURE 3 presents results for the average number of mutations per haploid genome maintained at equilibrium, and the resulting inbreeding depression and mutation load for h = 0.25 and $\frac{c}{\varepsilon} = 1$ (other parameter sets are shown in appendix as results are qualitatively similar, FIG. S3 to S10). Analytical predictions, which are depicted by solid lines, are obtained by solving Equation (3) numerically using the LC approach and accounting for pairwise genetic associations, and dots depict simulations results. For mutations affecting

the maintenance cost and survival, the average number of mutations per haploid genome 232 maintained at equilibrium $(n = \sum_i p_i)$ decreases as life expectancy increases (Fig. 3a-c), 233 because selection against mutations increases. This effect is more marked for mutations 234 affecting survival because selection is stronger in this case (Fig. 2d-f). Conversely, nincreases as life expectancy increases for mutations affecting the production cost, because 236 selection against these mutations weakens as life expectancy increases (Fig. 2e). In every 237 case, n decreases as the selfing rate increases due to the purging effect of self-fertilisation. Large differences in n do not translate into strong variations in inbreeding depression 239 with respect to life expectancy (Fig. 3, middle row). However, these variations differ be-240 tween mutation types. Indeed, inbreeding depression always decreases with life expectancy 241 when mutations affect survival, irrespective of the selfing rate (Fig. 3f), contrary to mu-242 tations affecting growth. Mutations affecting c generate higher inbreeding depression in 243 more short-lived species at low selfing rate, but this pattern is reversed at higher selfing 244 rates (Fig. 3d). Conversely, mutations affecting ε generate higher inbreeding depression 245 in more long-lived species for low α and this pattern is reversed for high α (Fig. 3e). This is the result of the interaction between life expectancy, the magnitude of selection and the 247 selfing rate. 248 The mutation load is lower when mutations are more numerous for all three muta-249 tions types (Fig. 3, bottom row). This result is not a mere reflection of differences in 250 the absolute strength of selection acting on mutations in populations with different life 251 expectancies. Otherwise, changing the phenotypic effect of mutations in a given model 252 and for a given parameters set would also change the mean phenotypic deviation from the 253 optimum, and therefore the mutation load. This is not what we observe. Indeed, Figure

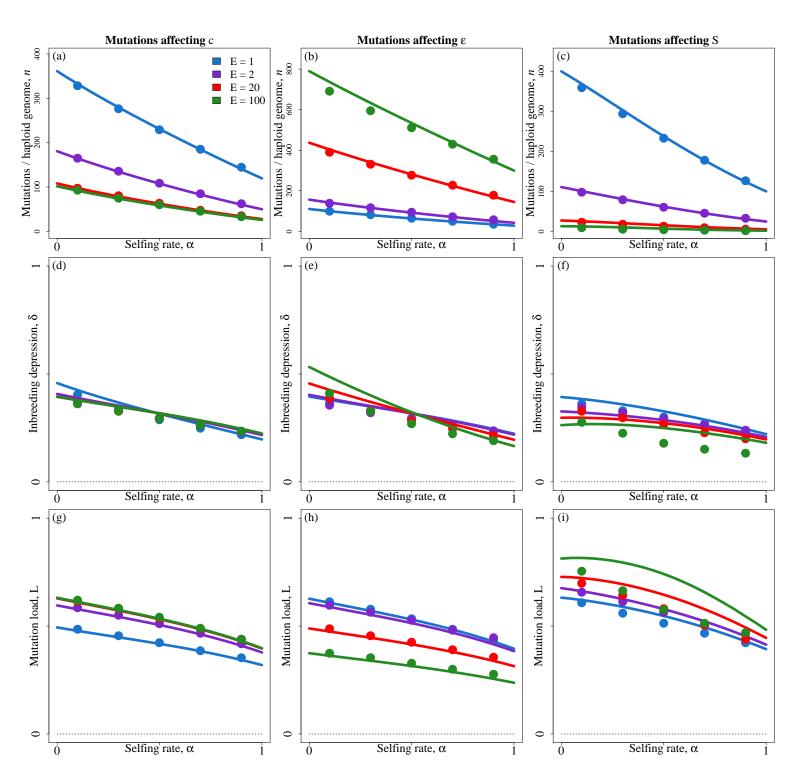


FIGURE 3: Average number of mutations per haploid genome (n, top row), inbreeding depression $(\delta, \text{ middle row})$, and mutation load (L, bottom row) as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lines: analytical predictions accounting for genetic associations between loci. Parameters shown here are $\frac{c}{\varepsilon} = 1, U = 0.5, s = 0.005, h = 0.25$.

S2 shows that when the effect of mutations is made ten times larger, this does not cause differences in the equilibrium phenotypic deviation, although the number of segregating mutations at equilibrium was considerably lower (because selection was stronger). This observation is consistent with results obtained by previous authors (Bataillon and Kirkpatrick, 2000), who showed that when populations exceeds a particular size, the mutation load becomes independent from the strength of selection acting on mutations. We argue that the differences we observe in terms of mutation load are imputable to differences in the shape of the fitness landscapes (Fig. S1), which are not fully captured by differences in the efficacy of selection.

3.4 Consequences for life-history evolution

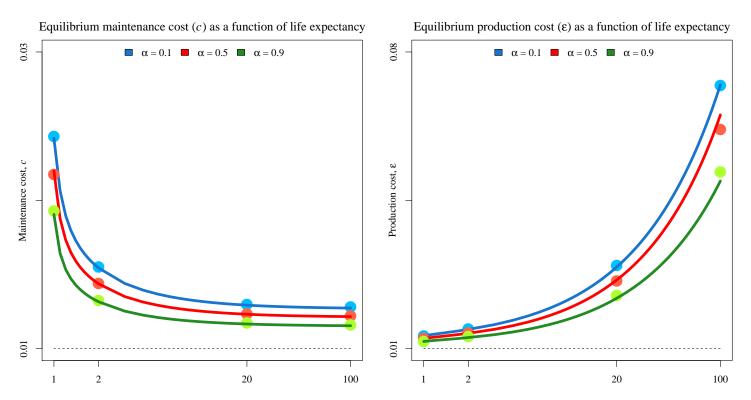


FIGURE 4: Equilibrium maintenance (left) and production (right) costs as function of life expectancy (log-scaled), for various selfing rates. Dots depict simulation results while lines depict analytical predictions. Parameters shown here are $c = \varepsilon = 0.01$, h = 0.25, s = 0.005.

The mutations segregating in the population cause the mean phenotype to deviate from its initial value. As the intensity of selection acting on mutations depends on this mean 265 phenotype, this generates a coevolutionary process, which leads to a joint equilibrium for 266 both the mean phenotype and the number of mutations segregating in the population (Appendix II, III). This equilibrium varies with respect to life expectancy and mating 268 system. Increasing the selfing rate slightly decreases the deviation of the mean phenotype 269 from its initial value, because selfing induces a better purging of mutations. More importantly, the equilibrium phenotypes vary significantly between populations with different 271 life expectancies. Indeed, as species become more long-lived, the maintenance cost de-272 creases, so that maximal size increases, and the production cost increases, so that growth 273 is slowed down. This means that if, for any reason, life expectancy changes in a species, 274 the growth strategy should also be changed as a consequence of the selective pressures 275 acting on deleterious mutations maintained at mutation-selection balance being altered.

4 Discussion

The phenotypic dimension of the genotype-to-phenotype-to-fitness map is usually overlooked in mutation load dynamics studies, as mutations directly affecting fitness are considered most of the time (e.g. Kondrashov, 1985; Lande and Schemske, 1985; Charlesworth
et al., 1990; Lande et al., 1994; Roze, 2015). Besides, studies which do integrate a phenotypic dimension do so by making use of Fisher's Geometric Model (Fisher, 1930), thereby
assuming a Gaussian (or quasi-Gaussian) distribution of fitness effect of mutations (e.g.
Wright, 1950; Bulmer, 1971; Roze and Blanckaert, 2014; Abu Awad and Roze, 2018).
While this assumption is both classical and valid for quantitative traits under stabilizing

selection underlay by a large amount of loci, not all traits fit this description, and this
constraint imposed on the shape of the fitness landscape remains somewhat arbitrary. In
the present paper, we incorporated phenotype in a different way. Indeed, we based our
model on physiological assumptions, letting the fitness effect of mutations arise from them,
without imposing any constraint on the shape of the fitness landscape.

Inbreeding depression. Our initial aim was to investigate the proposition that higher 290 levels of inbreeding depression in more long-lived species may be explained by mutations 291 affecting fitness differently in such species. To do so, we studied mutations affecting growth 292 and survival. Prior to the present study, the dynamics of mutations affecting survival had 293 been studied by Morgan (2001), in a perennial but not age-structured population. They 294 concluded that inbreeding depression should quickly decrease as life expectancy increases, and that significant outbreeding depression should be observed in long-lived species. They 296 argued this result could be attributed to the greater variance in fitness observed among the 297 offspring produced by self-fertilisation, which led to a higher mean lifetime fitness among them. Although they did not state it explicitly, this result stems from the fact that a 290 very low amount of mutations were maintained at mutation-selection balance in long-lived 300 species under the parameter sets they investigated. Indeed, Morgan (2001) considered very 301 large phenotypic effects of mutations (s = 0.1 and s = 1.0), which resulted in tremendous 302 fitness effects and therefore in the maintenance of almost no mutation at equilibrium. This 303 led variance in fitness to dominate over other aspects. Here, we investigated mutations 304 with much lower phenotypic effects (s = 0.005), and did not observe the same patterns. 305 In fact, we show that although inbreeding depression still decreases with life expectancy, 306 differences are considerably reduced, and outbreeding depression is no longer observed. Thus, if inbreeding depression is observed on traits related to adult survival in long-lived species, we conclude that this should be caused by mutations with very weak phenotypic effects.

Mutations affecting growth followed qualitatively different patterns, but differences in 311 the magnitude of inbreeding depression with respect to life expectancy were quantitatively 312 small in every investigated case, even for low dominance coefficients and high mutation 313 rates. Thus, we argue that although differences in fitness effects of mutations may contribute to explain the sharp increase in inbreeding depression in more long-lived species 315 observed empirically (Duminil et al., 2009; Angeloni et al., 2011), they are unlikely to con-316 stitute a sufficient explanation. Besides, since long-lived Angiosperms are predominantly 317 outcrossing (Barrett and Harder, 1996; Munoz et al., 2016), of all the mutation types 318 we modeled, only mutations affecting the production cost yielded inbreeding depression 319 patterns consistent with empirical evidence, that is increasing inbreeding depression with 320 life expectancy. This is due to the fact that selection against such mutations weakens 321 as life expectancy increases. In the light of this result, we may restate our initial ques-322 tion. Rather than asking whether mutations affecting fitness differently may contribute 323 to higher inbreeding depression in more long-lived species, we may now ask on what traits 324 is selection expected to be weaker in such species. Whether or not somatic mutations 325 accumulation could explain this empirical pattern is an open question. 326

Mutation load. We showed that more mutations maintained at mutation-selection balance led to a lower mutation load. This result is not a straigthforward consequence of differences in the intensity of selection acting on mutations, in agreement with results obtained by previous authors (Bataillon and Kirkpatrick, 2000). Instead, it is a conse-

quence of differences in the shape of fitness landscapes between species with different life expectancies on the one hand, and of the interaction between the mean phenotype and the 332 intensity of selection on the other hand. This result highlights the fact that incorporating a 333 phenotypic dimension to population genetics studies may lead to counter-intuitive results. In particular, when fitness landscapes are obtained as the result of biological assumptions, 335 and not arbitrarily assumed to be of a particular shape, unusual interactions with vari-336 ous aspects of species life-histories may result in novel predictions. For instance, in the present study, we showed that the mutation load may behave very differently depending 338 on the trait affected by mutations and life expectancy. Therefore, we conclude that com-339 paring mutation loads between species with contrasting life-histories may sometimes be 340 misleading, and would likely require trait-specific approaches. 341

Life-history evolution. Plants vary widely in life expectancy and stature, with life ex-342 pectancies ranging from a few weeks to hundreds, possibly thousands of years, and stature 343 spanning several orders of magnitude across Tracheophytes (Ehrlén and Lehtilä, 2002). These variations are correlated. Indeed, long-lived species tend to grow slower than short-345 lived species (Salguero-Gómez et al., 2016). In life-history traits evolution theory, this 346 type of correlation is usually interpreted in terms of trade-offs, with populations evolving towards the evolutionarily stable allocation of resources between growth, survival and re-348 production, given a number of constraints (Stearns, 1992). In this paper, we have shown 349 that the equilibrium maintenance and production costs differ between life expectancies, 350 causing more long-lived species to grow slower but ultimately larger than more short-lived 351 species, as commonly expected. However, the mechanism underlying this result is com-352 pletely different. Indeed, in our model, life-history traits do not coevolve in response to 353

trade-offs. Instead, the equilibrium growth costs are modified when life expectancy varies

because the selective pressures acting on the many mutations affecting these traits are al-

tered, leading to a more or less efficient purging of said mutations and thereby phenotypic

of differences at mutation-selection equilibrium. Thus, our results suggest that life-history

traits may sometimes coevolve regardless of trade-offs, because a change in a given trait

may alter the efficiency of purging of deleterious mutations affecting other traits.

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References

Abu Awad, D. and Roze, D. (2018). Effects of partial selfing on the equilibrium genetic

variance, mutation load and inbreeding depression under stabilizing selection. Evolution,

72:751-769.

Angeloni, F., Ouborg, N., and Leimu, R. (2011). Meta-analysis on the association of popu-

lation size and life history with inbreeding depression in plants. Biological Conservation,

21

144:35-43.

- Barrett, S. C. and Harder, L. D. (1996). The comparative biology of pollination and mating in flowering plants. *Philosophical Transactions of the Royal Society of London B*, 351:1271–1280.
- Barton, N. and Turelli, M. (1991). Natural and sexual selection on many loci. *Genetics*, 127:229–255.
- Bataillon, T. and Kirkpatrick, M. (2000). Inbreeding depression due to mildly deleterious mutations in finite populations: size does matter. *Genetics Research*, 75:75–81.
- Bobiwash, K., Schultz, S., and Schoen, D. (2013). Somatic deleterious mutation rate in a woody plant: estimation from phenotypic data. *Heredity*, 111:338–344.
- Bulmer, M. (1971). The effect of selection on genetic variability. *The American Naturalist*, 105:201–211.
- Burian, A., Barbier de Reuille, P., and Kuhlemeier, C. (2016). Patterns of stem cell divisions contribute to plant longevity. *Current Biology*, 26:1385–1394.
- Charlesworth, B. (1980). Evolution in age-structured populations. Cambridge Studies in Mathematical Biology.
- Charlesworth, D. and Charlesworth, B. (1987). Inbreeding depression and its evolutionary consequences. *Annual Review of Ecology and Systematics*, 18:237–268.
- Charlesworth, D., Morgan, M., and Charlesworth, B. (1990). Inbreeding depression, genetic load, and the evolution of outcrossing rates in a multilocus system with no linkage.

 Evolution, 44:1469–1489.

- Charlesworth, D. and Willis, J. (2009). The genetics of inbreeding depression. *Nature reviews genetics*, 10:783–796.
- Crow, J. (1958). Some possibilities for measuring selection intensities in man. *Human Biology*, 61:763–775.
- DeHaan, L. and Van Tassel, D. (2014). Useful insights from evolutionary biology for developing perennial grain crops. American Journal of Botany, 101:1801–1819.
- Duminil, J., Hardy, O., and Petit, R. (2009). Plant traits correlated with generation time directly affect inbreeding depression and mating system and indirectly genetic structure. BMC Evolutionary Biology, 9:177.
- Duputié, A. and Massol, F. (2013). An empiricist's guide to theoretical predictions on the evolution of dispersal. *Interface Focus*, 40:20130028.
- Ehrlén, J. and Lehtilä, K. (2002). How perennial are perennial plants? *Oikos*, 98:1070–1072.
- Enquist, B., Brown, J., and West, G. (1998). Allometric scaling of plant energetics and population density. *Nature*, 395:163–165.
- Epinat, G. and Lenormand, T. (2009). The evolution of assortative mating and selfing with in- and outbreeding depression. *Evolution*, 63:2047–2060.
- Fisher, R. (1930). The Genetical Theory of Natural Selection. Oxford University Press.
- Franco, M. and Silvertown, J. (1996). Life-history variation in plants: an exploration of the fast-slow continuum hypothesis. *Philosophical Transactions of the Royal Society of London B*, 351:1341–1348.

- Husband, B. and Schemske, D. W. (1996). Evolution of the magnitude and timing of inbreeding depression in plants. *Evolution*, 50:54–70.
- Kirkpatrick, M., Johnson, T., and Barton, N. (2002). General models of multilocus evolution. *Genetics*, 161:1727–1750.
- Klinkhamer, P., Meelis, E., De Jong, T., and Weiner, J. (1985). On the analysis of size-dependent reproductive output in plants. *Functional Ecology*, 6:308–316.
- Kondrashov, A. (1985). Deleterious mutations as an evolutionary factor. II. facultative apomixis and selfing. *Genetics*, 111:635–653.
- Lande, R. and Schemske, D. (1985). The evolution of self-fertilization and inbreeding depression in plants. I. genetic models. *Evolution*, 39:24–40.
- Lande, R., Schemske, D., and Schultz, S. (1994). High inbreeding depression, selective interference among loci, and the threshold selfing rate for purging recessive lethal mutations. *Evolution*, 48:965–978.
- Lanfear, R. (2018). Do plants have a segregated germline? PloS Biology, 16.
- Morgan, M. (2001). Consequences of life history for inbreeding depression and mating system evolution in plants. *Proceedings of the Royal Society of London B*, 268:1817–1824.
- Munoz, F., Violle, C., and Cheptou, P.-O. (2016). CSR ecological strategies and plant mating systems: outcrossing increases with competitiveness but stress-tolerance is related to mixed mating. *Oikos*, 125:1296–1303.

- Otto, S. and Orive, M. (1995). Evolutionary consequences of mutation and selection within an individual. *Genetics*, 141:1173–1187.
- Peters, R. (1983). The ecological implications of body size. Cambridge studies in ecology.
- Petit, R. and Hampe, A. (2006). Some evolutionary consequences of being a tree. Annual Review of Ecology, Evolution, and Systematics, 37:187–214.
- Plomion, C., Aury, J.-M., Amselem, J., Leroy, T., Murat, F., Duplessis, S., Faye, S., Francillonne, N., Labadie, K., Le Provost, G., Lesur, I., Bartholomé, J., Faivre-Rampant, P., Kohler, A., Leplé, J.-C., Chantret, N., Chen, J., Diévrat, A., Alaeitabar, T., Barbe, V., Belser, C., Bergès, H., Bodénès, C., Bogeat-Triboulot, M.-B., Bouffaud, M.-L., Brachi, B., Chancerel, E., Cohen, D., Couloux, A., Da Silva, C., Dossat, C., Ehrenmann, F. Gaspin, C., Grima-Pettenati, J., Guichoux, E., Hecker, A., Herrmann, S., Hugueney, P., Hummel, I., Klopp, C., Lalanne, C., Lascoux, M., Lasserre, E., Lemainque, A. Desprez-Loustau, M.-L., Luyten, I., Madoui, M.-A., Mangenot, S., Marchal, C., Maumus, F., Mercier, J., Michotey, C., Panaud, O., Picault, N., Rouhier, N., Rué, O., Rustenholz, C., Salin, F., Soler, M., Tarkka, M., Velt, A., Zanne, A., Martin, F., Wincker, P., Quesneville, H., Kremer, A., and Salse, J. (2018). Oak genome reveals facets of long lifespan. Nature Plants, 4:440–452.
- Roze, D. (2015). Effects of interference between selected loci on the mutation load, inbreeding depression, and heterosis. *Genetics*, 201:745–757.
- Roze, D. and Blanckaert, A. (2014). Epistasis, pleiotropy, and the mutation load in sexual and asexual populations. *Evolution*, 86:137–149.

- Roze, D. and Michod, R. (2010). Deleterious mutations and selection for sex in finite, diploid populations. Genetics, 184:1095–1112.
- Roze, D. and Rousset, F. (2005). Inbreeding depression and the evolution of dispersal rates: a multilocus model. *The Amrican Naturalist*, 166:708–721.
- Salguero-Gómez, R., Jones, O. R., Jongejans, E., Blomberg, S. P., Hodgson, D. J., Mbeau-Ache, C., Zuidema, P. A., de Kroon, H., and Buckley, Y. M. (2016). Fast—slow continuum and reproductive strategies structure plant life-history variation worldwide. *Proceedings of the National Academy of Sciences*, 113(1):230–235.
- Schmid-Siegert, E., Sarkar, N., Iseli, C., Calderon, S., Gouhier-Darimont, C., Chrast, J., Cattaneo, P., Schütz, F., Farinelli, L., Pagni, M., Schneider, M., Voumard, J., Jaboyed-off, L., Fankhauser, C., Hardtke, C., Keller, L., Pannell, J., Reymond, A., Robinson-Rechavi, M., Xenarios, I., and Reymond, P. (2017). Low number of fixed somatic mutations in a long-lived oak tree. *Nature Plants*, 12:926–929.
- Schoen, D. and Schultz, S. (2019). Somatic mutation and evolution in plants. *Annual Review of Ecology, Evolution and Systematics*, 50:2.1–2.25.
- Scofield, D. and Schultz, S. (2006). Mitosis, stature and evolution of plant mating systems: low- ϕ and high- ϕ plants. Proceedings of the Royal Society of London B, 273:275–282.
- Stearns, S. (1992). The Evolution of Life Histories. Oxford University Press.
- Watson, J., Platzer, A., Kazda, A., Akimcheva, S., Valuchova, S., Nizhynska, V., Nordborg, M., and Riha, K. (2016). Germline replications and somatic mutation accumula-

tion are independent of vegetative life span in arabidopsis. *Proceedings of the National Academy of Sciences*, 113:12226–12231.

- Weiner, J., Campbell, L., Pino, J., and Echarte, L. (2009). The allometry of reproduction within plant populations. *Journal of Ecology*, 97:1220–1233.
- West, G., Brown, J., and Enquist, B. (1997). A general model for the origin of allometric scaling laws in biology. *Science*, 276:122–126.
- West, G., Brown, J., and Enquist, B. (1999). A general model for the structure, function and allometry of plant vascular systems. *Nature*, 400:664–667.
- West, G., Brown, J., and Enquist, B. (2001). A general model for ontogenetic growth.

 Nature, 413:628–631.
- Winn, A., Elle, E., Kalisz, S., Cheptou, P.-O., Eckert, C., Goodwillie, C., Johnston, M., Moeller, D., Sargent, R., and Vallejo-Marin, M. (2011). Analysis of inbreeding depression in mixed-mating plants provides evidence for selective interference and stable mixed mating. *Evolution*, 65:3339–3359.
- Wright, S. (1950). The genetical structure of populations. Annals of Eugenics.

APPENDIX

I Growth model

At any age t, we assume that an individual is composed of G_t identical units (say, branches or buds), and has a resting metabolic rate B_t . This metabolic rate can be subdivided into energies spent on maintenance and growth as follows (West et al., 2001),

$$B_t = c G_t + \varepsilon \frac{dG_t}{dt},\tag{A1}$$

where c is the energy required to maintain a single unit, and ε is the energy required to create a new unit. Moreover, the resting metabolic rate can be expressed as

$$B_t = B_0 G_t^x, (A2)$$

where B_0 is the species' basal metabolic rate and x is the allometric coefficient (Peters, 1983). In vascular plants and animals, the value $x = \frac{3}{4}$ is generally used in growth models on the basis of both theoretical (e.g. West et al., 1997, 1999), and empirical arguments (e.g. Peters, 1983; Enquist et al., 1998; West et al., 2001), and will thus be used in what follows. Different exponent values would however likely yield qualitatively similar results, provided that they remain positive and smaller than one. Using Equation (A2), Equation (A1) can be rearranged into the following differential equation

$$\frac{dG}{dt} = \frac{1}{\varepsilon} \left(B_0 G_t^{\frac{3}{4}} - cG_t \right). \tag{A3}$$

Solving Equation (A3) for G_t , and setting $B_0 = 1$ for convenience, we obtain

$$G_t = \frac{e^{-\frac{c}{\varepsilon}(t+1)}}{c^4} \left(e^{\frac{c}{4\varepsilon}(t+1)} - 1 \right)^4. \tag{A4}$$

This growth function naturally saturates, when the energy required to maintain existing units becomes too large for new units to be produced. The size at which it saturates is given by

$$\lim_{t \to \infty} G_t = \frac{1}{c^4}.$$

II General recursions for the effects of selection and reproduction under partial selfing

II.1 Selection

In this paper, we use two different approaches to study our model. The first approach makes the assumption that age-structured populations can be viewed as a rescaled annual population (the Lifetime Fitness approach, LF), so that individuals first undergo lifetime selection, then reproduce. The second method proceeds in a more detailed manner, by studying each step of the life cycle successively (the Life Cycle approach, LC). Irrespective of the method, one may derive a general approximation of the relative fitness for an individual as a function of its genotype during any given selection stage. In this section, we derive such approximation assuming fitness is a function of a trait z, using the theoretical framework introduced by Barton and Turelli (1991) and generalized by Kirkpatrick et al. (2002). We then compute general recursions for the effects of selection on allelic frequencies and genetic associations.

II.1.1 Approximating relative fitnesses

Let X_i and X_i be the indicator variables associated with the paternally and maternally inherited alleles at the i^{th} locus. These variables are worth 1 when allele a is present at the position they are associated with, and 0 otherwise, so that for any trait z, assuming mutations affect this trait multiplicatively, an individual's trait value can be written as

$$z = z_0 \times \prod_i \left(1 \pm sh\left(X_i + \overset{*}{X_i}\right) \pm s(1 - 2h)X_i \overset{*}{X_i} \right), \tag{A5}$$

where \pm is replaced with + or - depending on whether mutations are assumed to increase (e.g. physiological growth costs) or decrease (e.g. survival probability) the trait, s is the phenotypic effect of the mutant allele, and h is its dominance coefficient. The log-value of the trait is then

$$\ln z = \ln z_0 + \sum_{i} \ln \left(1 \pm sh \left(X_i + X_i^* \right) \pm s(1 - 2h) X_i X_i^* \right),$$

which, under the assumption that selection is weak, i.e. that s is small, can be approximated by

$$\ln z \approx \ln z_0 \pm sh \sum_i \left(X_i + \overset{*}{X_i} \right) \pm s(1 - 2h) \sum_i X_i \overset{*}{X_i}.$$

Let us now define the centered variables associated with the paternally and maternally inherited alleles at the i^{th} locus, ζ_i and $\overset{*}{\zeta_i}$, as

$$\zeta_i = X_i - \mathbb{E}[X_i] = X_i - p_i \text{ and, } \overset{*}{\zeta_i} = \overset{*}{X_i} - \mathbb{E}[\overset{*}{X_i}] = \overset{*}{X_i} - p_i,$$

where $\mathbb{E}[X_i] = \mathbb{E}[\overset{*}{X_i}] = p_i$ is allele a's frequency at the i^{th} locus. The expectation of products of these centered variables allows one to quantify genetic associations between positions in the genome. For example, the expectation $\mathbb{E}\left[\zeta_i \times \overset{*}{\zeta_i}\right]$ measures to what extent the homozygosity deviates from the panmictic expectation at locus i. To make recursions clearer, let us introduce the condensed notation

$$D_{\mathbb{U},\mathbb{V}} = \mathbb{E}\left[\zeta_{\mathbb{U},\mathbb{V}}\right] = \mathbb{E}\left[\prod_{i\in\mathbb{U}} \left(\zeta_{\mathbb{U}}\right) \times \prod_{i\in\mathbb{V}} \left(\overset{*}{\zeta_{\mathbb{V}}}\right)\right],$$

where \mathbb{U} and \mathbb{V} are sets of positions paternally and maternally inherited, respectively (in the previous example, $\mathbb{U} = \{i\}$ and $\mathbb{V} = \{i\}$), so that the excess in homozygotes at locus i is denoted $D_{i,i}$. Repeated indexes sometimes appear in recursions. They can be dealt with using the relationship

$$D_{\mathbb{U}ii,\mathbb{V}} = (1 - 2p_i)D_{\mathbb{U}i,\mathbb{V}} + p_i q_i D_{\mathbb{U},\mathbb{V}}.$$

Injecting ζ -variables into Equation (II.1.1), it can then be rearranged into

$$\ln z \approx \ln z_0 \pm s \sum_i Z_i$$

with $Z_i = h \sum_i \left(\zeta_i + \overset{*}{\zeta_i} + 2p_i \right) + (1 - 2h) \sum_i \left(\zeta_{i,i} + p_i \left(\zeta_i + \overset{*}{\zeta_i} \right) + p_i^2 \right)$. Hence, the mean log-value is given by

$$\overline{\ln z} = \ln z_0 \pm s \sum_i \overline{Z}_i \text{ with, } \overline{Z}_i = 2h \sum_i p_i + (1 - 2h) \sum_i \left(D_{i,i} + p_i^2 \right),$$

and one may express the trait value z as

$$z = z_0 e^{\pm s \sum_i Z_i} = z_0 e^{\pm s \sum_i (\overline{Z}_i + \Delta Z_i)},$$

with $\Delta Z_i = Z_i - \overline{Z}_i$. Therefore, the trait average, \overline{z} , is simply

$$\bar{z} \approx z_0 \ e^{\pm s \sum_i \overline{Z}_i}.$$
 (A6)

All else being fixed, an individual's fitness during a given selection stage, w(z), is a function

of its trait value z. The mean fitness is a constant, given by $\bar{w} = w(\bar{z})$, and the relative fitness $\frac{w(z)}{\bar{w}}$, to second order in s, is given by

$$\frac{w}{\bar{w}}(z) = 1 + \frac{w'(\bar{z})}{\bar{w}} \sum_{i} \Delta Z_i + \frac{w''(\bar{z})}{2 \bar{w}} \sum_{i} \sum_{j} \Delta Z_i \times \Delta Z_j, \tag{A7}$$

where w' and w'' are the first and second derivatives of w with respect to ΔZ_i . Neglecting terms in i=j, and denoting $\bar{s}_z(\bar{z})=\frac{w'(\bar{z})}{\bar{w}}$ and $\hat{\bar{s}}_z(\bar{z})=\frac{w''(\bar{z})}{2\;\bar{w}}$, this is

$$\frac{w}{\overline{w}}(z) \approx 1 - \bar{s}_z(\bar{z}) \sum_{i} \left(h\left(\zeta_i + \zeta_i^*\right) + (1 - 2h)\left(\zeta_{i,i} - D_{i,i}\right) \right)
+ \hat{\bar{s}}_z(\bar{z}) \left[h^2 \sum_{i < j} (\zeta_i + \zeta_i^*)(\zeta_j + \zeta_j^*) + h(1 - 2h) \sum_{i \neq j} \left(\zeta_i + \zeta_i^*\right)(\zeta_{j,j} - D_{j,j}) \right]
+ (1 - 2h)^2 \sum_{i < j} \left((\zeta_{i,i} - D_{i,i})\left(\zeta_{j,j} - D_{j,j}\right) - (D_{ij,ij} - D_{i,i}D_{j,j}) \right) \right].$$
(A8)

The leading order selection coefficient (\bar{s}) encapsulates the effects of selection acting directly on loci, while the second order selection coefficient (\hat{s}) quantifies the effects of indirect selection between pairs of loci. We neglect higher order selection coefficients.

Allelic frequencies change. Using Equation (A8), the change in allelic frequencies at the i^{th} selected locus owing to selection at a given stage is

$$\Delta^s p_i = \mathbb{E}\left[\frac{w}{\overline{w}} \ \frac{X_i + \overset{*}{X_i}}{2}\right] - p_i,$$

which, to second order in s, yields

$$\Delta^{s} p_{i} = -\bar{s}_{z} \left(h p_{i} + (1 - h) D_{i,i} \right) - \bar{s}_{z} (1 - 2h) \sum_{j \neq i} D_{ij,j}$$

$$+ \hat{\bar{s}}_{z} (1 - h) (1 - 2h) \sum_{j \neq i} \left(D_{ij,ij} - D_{i,i} D_{j,j} \right) + o\left(s^{3}\right),$$
(A9)

Effect of selection on associations between selected loci. We consider the effect of selection on associations between selected loci to leading order in s (first line in Equation A8). Following selection, we have

$$D_{ij,j}^s = \mathbb{E}\left[\frac{w}{\overline{w}} \zeta_{ij,j}\right] - \Delta^s p_i D_{j,j}^s \text{ and, } D_{i,i}^s = \mathbb{E}\left[\frac{w}{\overline{w}} \zeta_{i,i}\right] + (\Delta^s p_i)^2.$$

To leading order, $\Delta^s p_i$ simplifies to

$$\Delta^{s} p_{i} = -\bar{s}_{z} \left(h \ p_{i} - (1 - h) D_{i,i} \right) + o \left(s^{2} \right).$$

Hence, $D_{i,i}^s$ and $D_{ij,j}^s$ are given by

$$D_{i,i}^{s} = D_{i,i} (1 - \bar{s}_z) - \bar{s}_z (1 - 2h) \sum_{j \neq i} (D_{ij,ij} - D_{i,i} D_{j,j}), \qquad (A10)$$

and,

$$D_{ij,j}^{s} = D_{ij,j} - \bar{s}_{z}(1-h) \left(D_{ij,ij} - D_{i,i}D_{j,j} \right). \tag{A11}$$

We neglect the effects of selection on $D_{ij,ij}$.

II.2 Reproduction and mutation under partial selfing

As we model infinitely many loci, we may assume that mutations almost never occur two times at the same locus. Since they are not affected by recombination and self-fertilisation, allelic frequencies following reproduction are simply given by

$$p_i^r = p_i + U(1 - p_i) \approx U + p_i, \tag{A12}$$

where p_i here depicts allelic frequencies just before reproduction. On the other hand, while we neglect the effects of mutation on genetic associations, they are affected by recombination and selfing. Following reproduction, denoting $F = \frac{\alpha}{2-\alpha}$, two-way within loci associations are given by

$$D_{i,i}^{r} = \frac{F}{1+F} \left(D_{i,i} + p_{i}q_{i} \right), \tag{A13}$$

As for three- and four-way associations, we have

$$D_{ij,j}^{r} = \frac{F}{1+F} \left(D_{ij,j} + (1-r)D_{ij} + rD_{i,j} \right) \approx \frac{F}{1+F} D_{ij,j}, \tag{A14}$$

and,

$$D_{ij,ij}^{r} = \frac{F}{1+F} \left[(1-R) \left(D_{ij,ij} + p_i q_i p_j q_j \right) + R \left(p_i q_i D_{j,j} + p_j q_j D_{i,i} \right) \right], \text{ with } R = 2r(1-r).$$
(A15)

III Fitnesses and selection coefficients

In Appendix II.1, we derived general recursions for the effects of selection at any given selection stage. We showed that to second order, the intensity of selection can be summarised using selection coefficients, \bar{s}_z and $\hat{\bar{s}}_z$. In this section, our aim is to derive the selection coefficients that arise from our model.

III.1 Lifetime fitness approach

III.1.1 Fitness expression

In the LF approach, we make the assumption that all the selective pressures acting on mutations can be summarised into a single lifetime fitness expression. In this section, we compute the lifetime reproductive output of an individual, given its phenotype. This quantity is used as a measure of lifetime fitness.

The size of an individual at age t (G_t) is the solution of Equation (2). Given the individual's maintenance and production costs, c and ε , it is

$$G_t = \frac{e^{-\frac{c}{\varepsilon}(t+1)}}{c^4} \left(e^{-\frac{c}{4\varepsilon}(t+1)} - 1 \right)^4.$$

Hence, the contribution to reproduction of an individual up to age τ , \bar{G}_{τ} , is given by

$$\bar{G}_{\tau} = \sum_{t=0}^{\tau} G_t.$$

Furthermore, the probability that, given its survival probability S, an individual lives up

to exactly age τ then dies (\bar{S}_{τ}) is

$$\bar{S}_{\tau} = S^{\tau}(1 - S).$$

Hence, the lifetime fitness of an individual (W) is given by

$$W = J \times \sum_{\tau=0}^{\infty} \bar{S}_{\tau} \bar{G}_{\tau} = \frac{J}{c^4} \left(\frac{1}{1-S} - \frac{4}{e^{\frac{c}{4\varepsilon}} - S} + \frac{6}{e^{\frac{c}{2\varepsilon}} - S} - \frac{4}{e^{\frac{3c}{4\varepsilon}} - S} + \frac{1}{e^{\frac{c}{\varepsilon}} - S} \right), \quad (A16)$$

where J is the recruitment probability of the individual as a juvenile.

III.1.2 Leading order lifetime selection coefficients

Here, we write down the leading order lifetime selection coefficients obtained with the method described in Appendix II.1. For any trait z, these coefficients are obtained by applying Equation (A7) to Equation (A16). We show that they depend on the ratio $\frac{c}{\varepsilon}$, rather than c and ε absolute values. In what follows, we set J=1 for convenience.

Mutations altering c. The leading order lifetime selection coefficient for mutations altering c is given by

$$\bar{s}_c = s \frac{4V_c - c \frac{\partial V_c}{\partial c}}{V_c} \quad \text{with,} \quad V_c = \frac{1}{1 - S} - \frac{4}{e^{\frac{\bar{c}}{4\bar{c}}} - S} + \frac{6}{e^{\frac{\bar{c}}{2\bar{c}}} - S} - \frac{4}{e^{\frac{3\bar{c}}{4\bar{c}}} - S} + \frac{1}{e^{\frac{\bar{c}}{\bar{c}}} - S}. \quad (A17)$$

Making the variable change $c = \varepsilon \varphi$ in Equation (A17), so that it now depends of φ instead of c, we obtain that

$$\frac{\partial \bar{s}_c}{\partial \varepsilon} = 0,$$

which demonstrates that \bar{s}_c depends only on the ratio $\varphi = \frac{c}{\varepsilon}$. The same reasoning is followed for all three mutation types and will thus not be repeated.

Mutations altering ε . In this case, the leading order lifetime selection coefficient is

$$\bar{s}_{\varepsilon} = -s \frac{\bar{\varepsilon}}{V_{\varepsilon}} \frac{\partial V_{\varepsilon}}{\partial \varepsilon} \text{ with, } V_{\varepsilon} = \frac{1}{1 - S} - \frac{4}{e^{\frac{c}{4\bar{\varepsilon}}} - S} + \frac{6}{e^{\frac{c}{2\bar{\varepsilon}}} - S} - \frac{4}{e^{\frac{3c}{4\bar{\varepsilon}}} - S} + \frac{1}{e^{\frac{c}{\bar{\varepsilon}}} - S}$$
 (A18)

Mutations altering S. Finally, the leading order lifetime selection coefficient for mutations affecting survival is given by

$$\bar{s}_S = s \frac{V_S + S \frac{\partial V_S}{\partial S}}{V_S} \quad \text{with,} \quad V_S = \frac{1}{1 - \bar{S}} - \frac{4}{e^{\frac{c}{4\bar{\varepsilon}}} - \bar{S}} + \frac{6}{e^{\frac{c}{2\bar{\varepsilon}}} - \bar{S}} - \frac{4}{e^{\frac{3c}{4\bar{\varepsilon}}} - \bar{S}} + \frac{1}{e^{\frac{c}{\bar{\varepsilon}}} - \bar{S}} \quad (A19)$$

III.2 Life cycle approach

In the LC approach, selection at different stages is considered separately, thus we denote the selection coefficients associated with selection phase k, when mutations affect trait z, as \bar{s}_z^k and \hat{s}_z^k . The stages at which selection occurs in our model are reproduction (denoted by k=g), juvenile recruitment (denoted k=j) and adult survival (k=s).

III.2.1 Contribution to reproduction

Selection on contribution to reproduction occurs in all three models (k = g). Size at age $t(G_t)$ is the solution of Equation (2). It is given by

$$G_t = \frac{e^{-\frac{c}{\varepsilon}(t+1)}}{c^4} \left(e^{-\frac{c}{4\varepsilon}(t+1)} - 1 \right)^4.$$

Thus, the average size of an individual given its genotype, which gives its fitness during this selection phase, is

$$w_g = \sum_{t=0}^{\infty} S^t (1 - S) \ G_t = \frac{1 - S}{c^4} \left(\frac{1}{1 - S} - \frac{4}{e^{\frac{c}{4\varepsilon}} - S} + \frac{6}{e^{\frac{c}{2\varepsilon}} - S} - \frac{4}{e^{\frac{3c}{4\varepsilon}} - S} + \frac{1}{e^{\frac{c}{\varepsilon}} - S} \right). \tag{A20}$$

For any trait z, the selection coefficients at this stage, \bar{s}_z^g and \hat{s}_z^g are obtained by applying Equation (A7) to Equation (A20).

III.2.2 Survival and recruitment

Selection on survival occurs in juveniles (k=j) and in adults (k=s). Individual fitnesses during survival selection stages are given by their survival probability. Hence, when mutations affect the maintenance or the production cost (i.e. not survival), we have $\bar{s}_c^j = \bar{s}_c^s = \hat{s}_c^j = \hat{s}_c^s = 0$ and $\bar{s}_\varepsilon^j = \bar{s}_\varepsilon^s = \hat{s}_\varepsilon^j = \hat{s}_\varepsilon^s = 0$. On the other hand, when mutations affect survival given by $\bar{s}_S^j = s$ and $\hat{s}_S^j = s^2$ in juveniles, and $\bar{s}_S^s = s$ and $\hat{s}_S^s = s^2$ in adults.

IV Analytical equilibrium results

At mutation-selection balance, we may obtain leading order approximations for equilibrium genetic associations. These associations differ between our two approaches.

IV.1 QLE associations in the LF approach

Using the LF approach, selection is encapsulated into a leading and second order selection coefficients, \bar{s}_z and $\hat{\bar{s}}_z$. The genetic associations at QLE are computed to leading order. Excess in homozygotes at the i^{th} locus is given by

$$D_{i,i}^* = F \left[1 - \bar{s}_z \left(2F + h(1 - F) \right) \right] p_i, \tag{A21}$$

while three- and four-way associations are given by

$$D_{ij,j}^* = -\bar{s}_z(1-h)\frac{F^2(1-F)(1-R(1-F))}{1+RF}p_ip_j,$$
(A22)

and,

$$D_{ij,ij}^* = \frac{F}{1 + RF} \left[1 - R(1 - 2F) - 2\bar{s}_z \left(RF(1 + F) + [1 + R(1 - 2F)] \left(F + h(1 - F) + \frac{F(1 - R)}{1 + RF} \right) \right) \right] p_i p_j.$$
(A23)

IV.2 QLE associations in the LC approach

In the LC approach, we consider selection occurring at the different stages of the life cycle separately. This allows us to incorporate the effects of selection more precisely. In particular, selection occurring before syngamy (i.e. in adults), impacts genetic associations

differently than following syngamy (in juveniles). The excess in homozygotes at QLE, at the i^{th} locus, is given by

$$D_{i,i}^* = F \left[1 - \bar{s}_z^g \left(F + h(1 - F) \right) \right] p_i, \tag{A24}$$

while other associations are given by

$$D_{ij,j}^* = -(1-h)G\left(\bar{s}_z^s \frac{1+F}{1-\bar{S}} + \bar{s}_z^g F\right) p_i p_j \tag{A25}$$

and,

$$D_{ij,ij}^* = \frac{F}{1 + RF} \left[1 - R(1 - 2F) - 2\bar{s}_z^g \left(h + F(1 - h) \right) \left(1 - R(1 - F) \right) \right] p_i p_j, \quad (A26)$$

where \bar{S} is the average survival probability in the population.

IV.3 Solution at mutation-selection balance

The intensity of selection is dependent on the ratio $\frac{c}{\varepsilon}$ and on the survival probability S. Hence, when mutations are numerous, their effect on the population average of the trait they affect has to be accounted for in order to obtain accurate approximations. To do so, one has to find the joint equilibrium of the number of mutations and the average trait value. This can be done by solving

$$\begin{cases}
\Delta p_i(\bar{z}) = 0 \\
\bar{z} = z_0 \times e^{\pm s \sum_i \overline{Z}_i} \left(1 + s^2 \sum_{j \neq i} \frac{\overline{\Delta Z_i \times \Delta Z_j}}{2} \right),
\end{cases} \tag{A27}$$

for p_i and \bar{z} . However, the solution of this system cannot be written explicitly. It can however be approached numerically, by injecting a leading order approximation for \bar{z} into $\Delta p_i(\bar{z})$, and looking for the smallest value of p_i for which $\Delta p_i(\bar{z})$ becomes negative. Technical details are not given here, but the Mathematica scripts we use to obtain numerical approximations are available from GitHub and commented (https://github.com/Thomas-Lesaffre/On_deleterious_mutations_in_perennials). If the reader has any question regarding these methods, feel free to contact the corresponding author.

In the main text, the analytical approximations obtained using the LF method are not presented. they are presented in a lighter color in all the figures of Appendix VII, which present results for a variety of dominance coefficient and growth costs values. The two methods yield the same results to leading order but differ when accounting for genetic associations when mutations affect survival. In this case, the LF approach is less precise than the LC method. This has to do with the timing of selection. Indeed, in this case, selection occurs both before and after syngamy, that is in adults and juveniles, respectively, or in other words before and after homozygosity is changed by self-fertilisation. Hence, the effects of selection on interactions are not accounted for correctly in the LF approach, and the population cannot simply be assimilated to a rescaled annual population.

V Fitness landscapes

In Figure S1, lifetime fitness is presented as a function of phenotypic deviation ΔP . For each trait z, we have $z = z_0 (1 + \Delta P)$. The resulting fitness landscape for each set of parameters is rescaled by its maximal value, which is always obtained for $\Delta P = 0$.

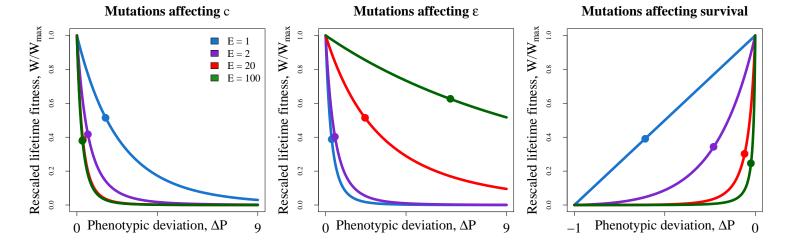


FIGURE S1: Rescaled fitness landscapes for each mutation type, for initial life expectancies $(E=1;\ 2;\ 20;\ 100)$. Lines depict fitness, while dots depict the mutation-selection equilibrium fitness reached for $\varphi=1,\ h=0.25,\ s=0.005,$ and U=0.5.

VI Comparison of mean phenotypic deviation for different magnitude of effect of mutations

Comparison of the phenotypic deviation for different effects of mutations

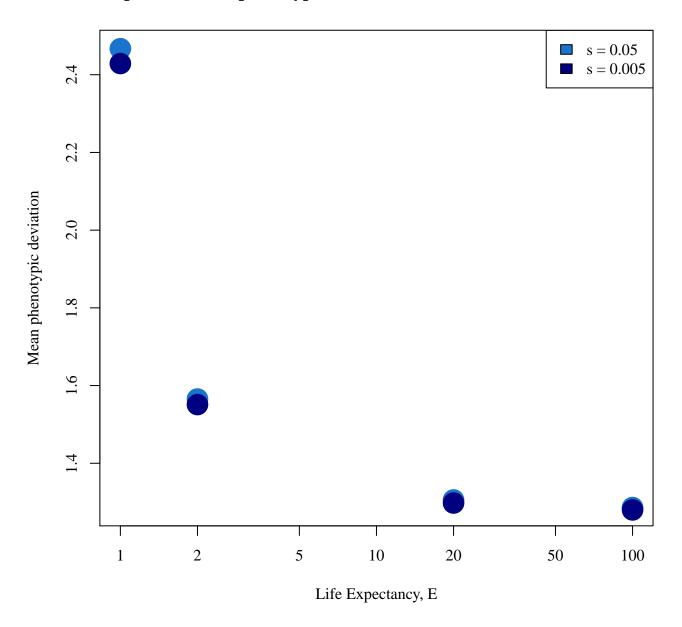


FIGURE S2: Comparison of the mean phenotypic deviation, for mutations affecting the maintenance cost (c), for two different magnitude of effect of mutations (s=0.05 and s=0.005). Parameters used are $\varphi=1,\,h=0.25,\,U=0.5$.

VII Mutations per haploid genome, inbreeding depression and mutation load for other parameter values

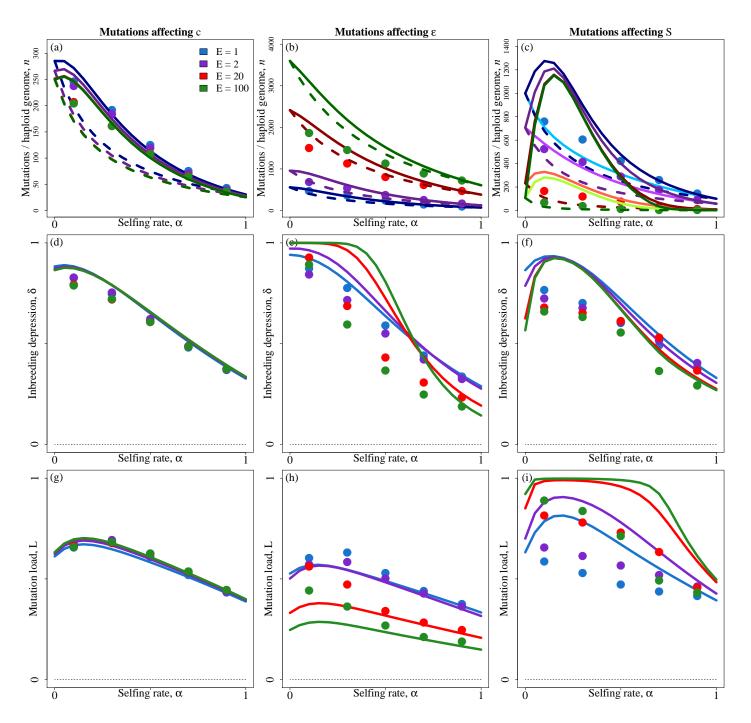


FIGURE S3: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta, bottom row)$ as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 10, U = 0.5, s = 0.005, h = 0.10$.

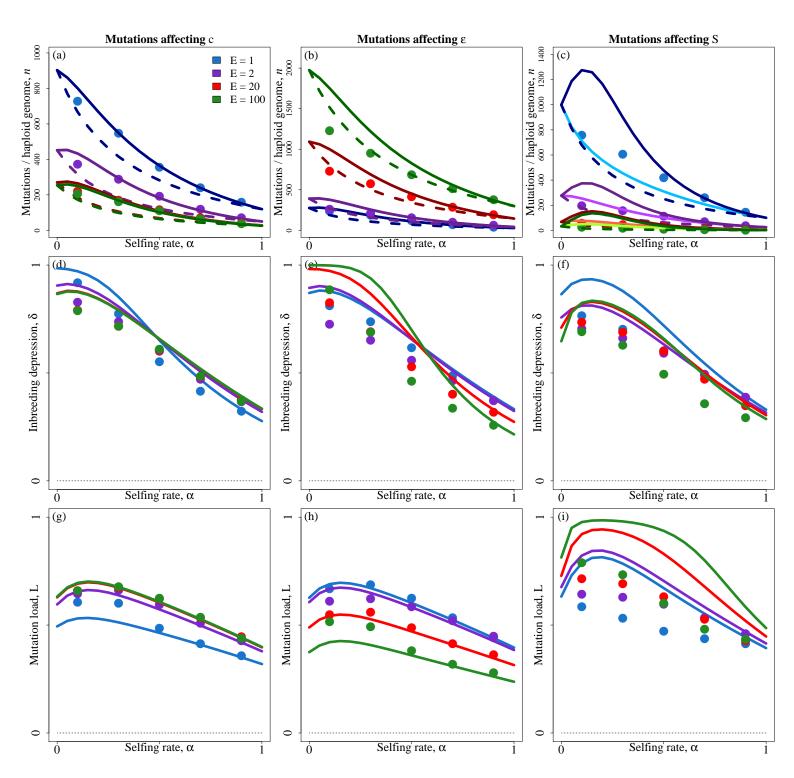


FIGURE S4: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta,$ bottom row) as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha=0.1,\ 0.3,\ 0.5,\ 0.7,\ 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi=1,\ U=0.5,\ s=0.005,\ h=0.10$.

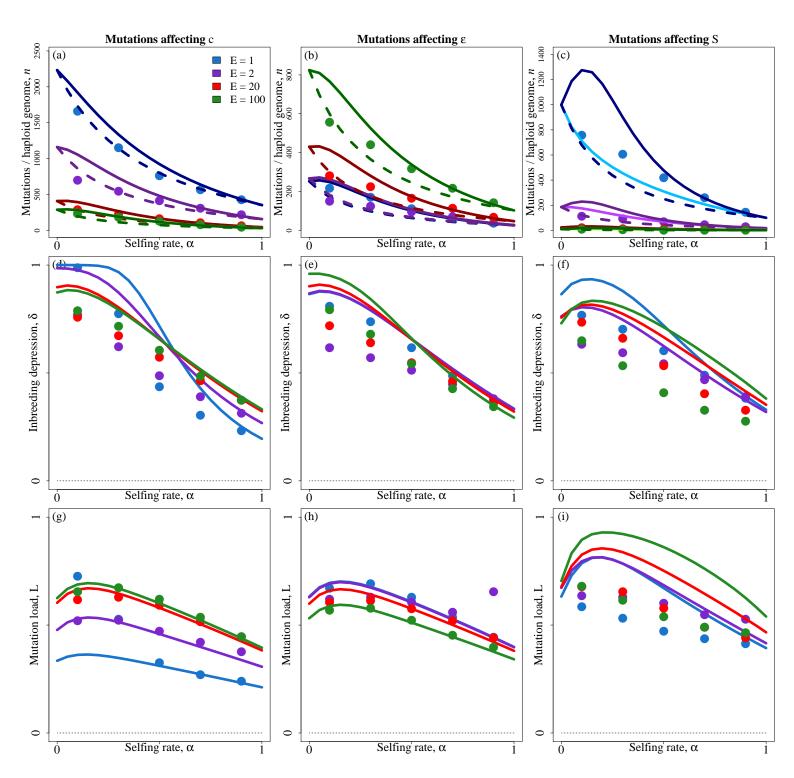


FIGURE S5: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta, bottom row)$ as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 0.1, U = 0.5, s = 0.005, h = 0.10$.

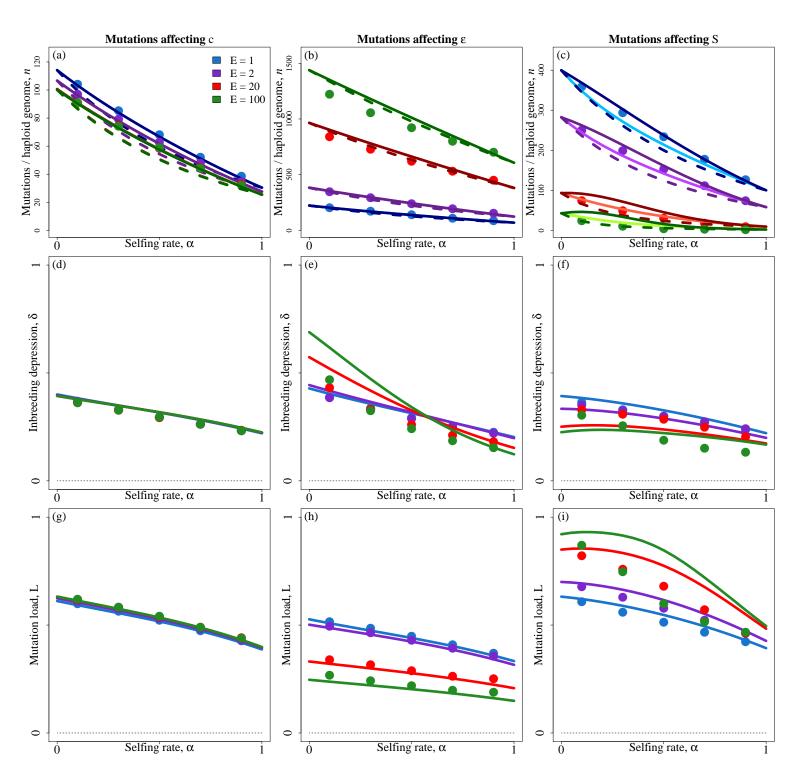


FIGURE S6: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta, bottom row)$ as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 10, U = 0.5, s = 0.005, h = 0.25$.

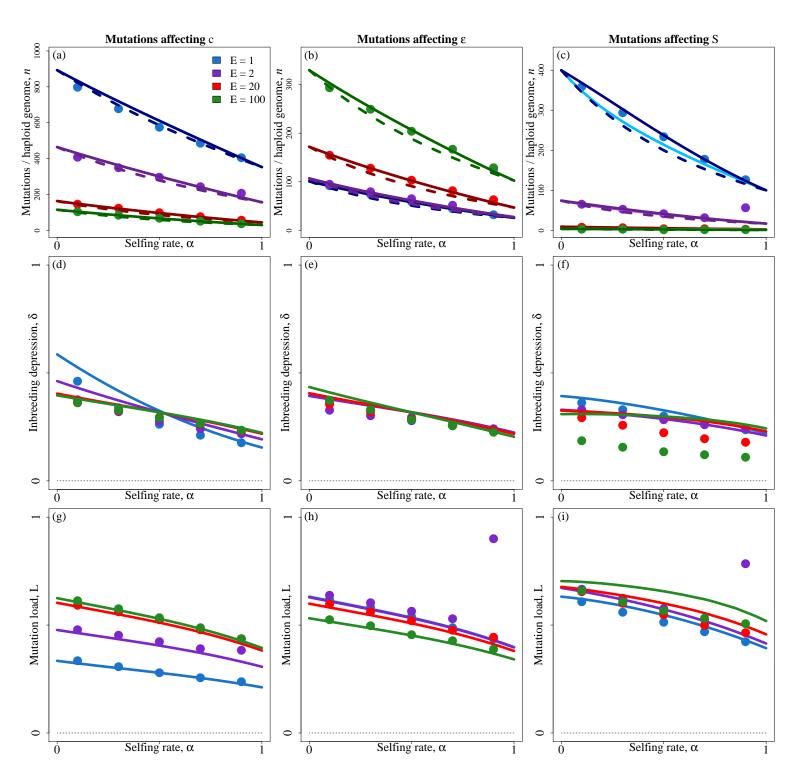


FIGURE S7: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta, bottom row)$ as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 0.1, U = 0.5, s = 0.005, h = 0.250$.

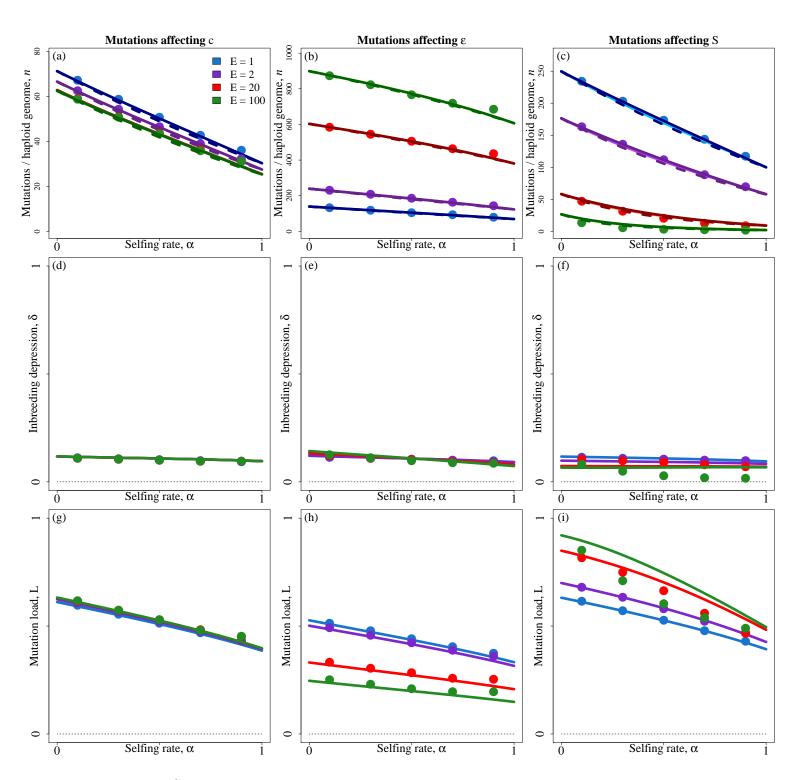


FIGURE S8: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta, bottom row)$ as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 10, U = 0.5, s = 0.005, h = 0.40$.

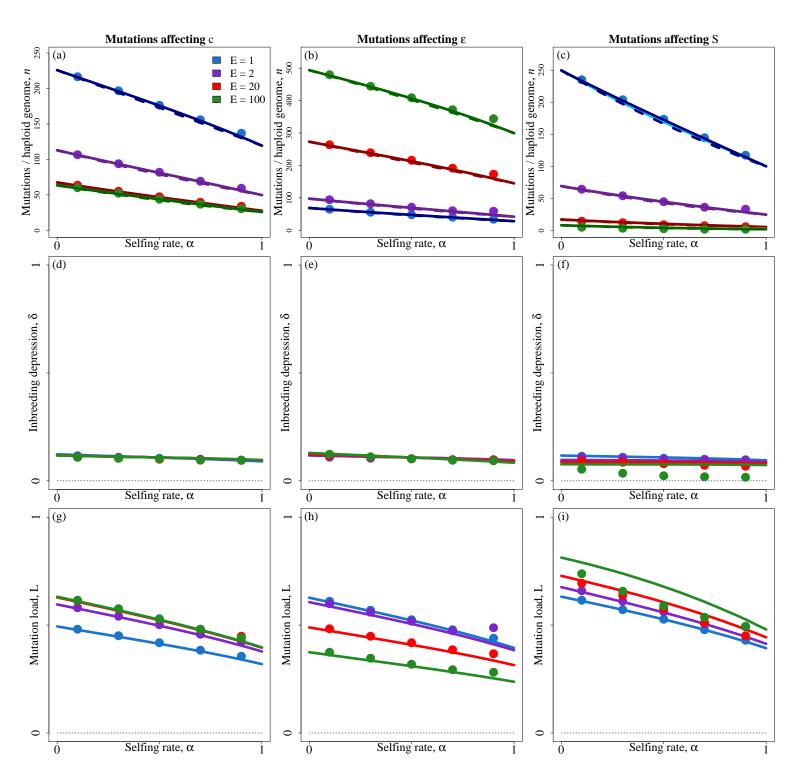


FIGURE S9: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta,$ bottom row) as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha=0.1,\ 0.3,\ 0.5,\ 0.7,\ 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi=1,\ U=0.5,\ s=0.005,\ h=0.40$.

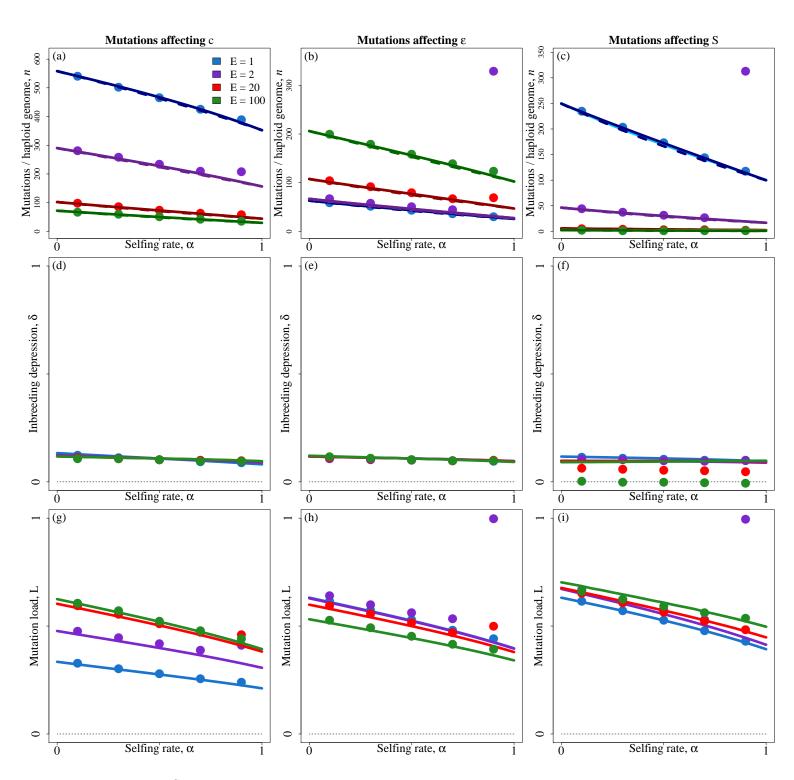


FIGURE S10: Average number of mutations per haploid genome (n, top row) and inbreeding depression $(\delta,$ bottom row) as a function of the selfing rate (α) , for various life expectancies (colors). Each column corresponds to one type of mutation. Dots: simulation results for $\alpha = 0.1, 0.3, 0.5, 0.7, 0.9$. Lighter lines: LF approach predictions. Darker lines: LC approach predictions. Dashed lines: leading order approximations (neglecting interactions). Solid lines: approximations accounting for pairwise interactions between loci. Parameters shown here are $\varphi = 0.1, U = 0.5, s = 0.005, h = 0.40$.