Large scale genome-wide association study reveals that drought induced lodging in grain sorghum is associated with plant height and traits linked to carbon remobilisation

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Abstract

Sorghum is generally grown in water limited conditions and often lodges under post-anthesis drought, which reduces yield and quality. Due to its complexity, our understanding on the genetic control of lodging is very limited. We dissected its genetic architecture in grain sorghum through genome-wide association study (GWAS) on 2308 unique hybrids grown in 17 Australian sorghum trials over 3 years. The GWAS detected 213 QTL, the majority of which showed a significant association with leaf senescence and plant height (72% and 71% respectively). Only 16 lodging QTL were associated with neither leaf senescence nor plant height. The high incidence of multi-trait association for the lodging QTL indicates that lodging in grain sorghum is mainly associated with plant height and traits linked to carbohydrate remobilisation. This result supported the selection for stay-green (delayed leaf senescence) to reduce lodging susceptibility, rather than selection for short stature and lodging resistance *per se*, which likely reduces yield. Additionally, our data suggested a protective effect of stay-green on weakening the association between lodging susceptibility and plant height. Our study also showed that lodging resistance might be improved by selection for stem composition but was unlikely to be improved by selection for classical resistance to stalk rots.

Keywords

GWAS, leaf senescence, lignin biosynthesis, lodging, plant height, sorghum, stay-green

Introduction

Water availability is a primary constraint to crop production worldwide . Among many cereal crops, sorghum is renowned for its tolerance to drought and heat (Hadebe *et al.*, 2017). Even though, sorghum crop is often exposed to various types of water stress with drought during grain filling being common (Chapman *et al.*, 2000; Kholová *et al.*, 2013). One of the impacts of water stress during grain filling in sorghum is lodging. Lodging is the permanent displacement of crop stems from their vertical position. It can be caused either by buckling at a basal internode (stem lodging) or by root rotation in the soil (root lodging) (Pinthus, 1974). Stem lodging is the major type of lodging induced by drought. When water stress occurs during grain filling, sorghum plants senesce and subsequently lodge due to stem collapse or breakage at the basal internodes (Henzell *et al.*, 1984). This form of lodging is often observed in grain sorghum hybrids grown in Australia (Henzell *et al.*, 1984), the USA (Rosenow, 1977), and Argentina (Frezzi and Teyssandier, 1980).

The occurrence and severity of lodging depends on the growth environment and the growth stage of the crop. Yield losses due to lodging in grain sorghum in production conditions are not well documented; however, it is believed that lodging occurring following water stress during the grain filling stage causes the most grain loss world-wide, especially in regions where grains are harvested mechanically (Johnson *et al.*, 1997).

The causes of lodging are not well understood. However, many factors may be associated to lodging, including stay-green, plant height, root and stalk rots, and stem characteristics.

Stay-green is an important trait providing tolerance to drought in many cereal crops including sorghum, wheat, maize, and rice (Borrell, Mullet, *et al.*, 2014; Borrell, van Oosterom, *et al.*, 2014; Cha *et al.*, 2002; Christopher *et al.*, 2008; J. Zhang *et al.*, 2019). Stay-green phenotype is expressed as the delayed leaf senescence during grain filling. It is affected by many factors that influence the source-sink relationships within the plant (Borrell *et al.*, 2001; Borrell and Hammer, 2000), such as restricted tiller number and reduced upper leaf size resulting in reduced canopy size and pre-anthesis water demand (Borrell, van Oosterom, *et al.*, 2014), reduced grain number (Borrell and Hammer, 2000), and possibly increased water uptake through narrow

nodal root angle (Mace *et al.*, 2012). Stay-green can increase water availability

during post-anthesis water stress (Borrell, Mullet, *et al.*, 2014), which reduces the

level of carbohydrate remobilisation by ensuring that photosynthesis continues, and

consequently reduces the intensity of carbon remobilisation from stem and leaves

(Borrell and Douglas, 1996)..

Unlike stay-green, plant height not only affects carbohydrate supply/demand but also

can have direct physical effects on lodging. Tall sorghum plants produce a higher

stem biomass per grain than short crops (George-Jaeggli *et al.*, 2011). This indicates

that tall sorghum plants have more stem reserves available for translocation to the

grains during post-anthesis water stress than short sorghum plants, which could

potentially lead to less lodging in tall plants than in short plants. On the other hand,

tall sorghum plants generally have higher yield than short sorghum plants (Jordan *et*

al., 2003), and higher yielding sorghums are more susceptible to terminal drought

stress than are lower yielding sorghums (Rosenow *et al.*, 1983). Additionally the

higher centre of gravity and greater leverage force in tall sorghums means that tall,

high yielding genotypes are more prone to lodging under drought conditions after

flowering.

This already complex interaction of factors is further complicated by the presence of

soil born root and stalk rotting pathogens, such as *Macrophomina phaseolina* and

Fusarium spp., which may accelerate lodging. These fungi are necrotrophic in nature;

while they may invade healthy plants, they are typically unable to colonise healthy

tissue (Dodd, 1980). However, when the stem begins to senesce under water stress

during grain filling, they attack the dying roots and stalks (Srinivasa Reddy *et al.*,

2008). The fungi secrete cell-wall-degrading enzymes (CWDE) to upregulate CWDE-

encoding genes, degrading the integrity of the pith and rind in the basal internodes,

which weakens the stalk and further exacerbates lodging (Bandara *et al.*, 2018).

Lodging is also affected by stem mechanical properties. Associations between stem

lodging and stem strength, rigidity, and chemical composition including lignin density

have been observed (Bashford *et al.*, 1976; Esechie *et al.*, 1977; Gomez *et al.*, 2018,

2017; Murray *et al.*, 2008). Lignin is one of the key biomolecules in vascular plants

and its functions include structural support and water transport (Boerjan *et al.*, 2003;

Higuchi, 2006). Reduced lignin content has been shown to lead to increased lodging

susceptibility in maize (Sun *et al.*, 2018).

Because of its complex nature, limited progress has been made in understanding the

genetic architecture of lodging in sorghum. To date, three studies (Kebede *et al.*,

2001; Murray *et al.*, 2008; Srinivasa Reddy *et al.*, 2008) detected seven QTL

associated with lodging in sorghum with no common QTL being identified across

these studies. Given the complex set of traits known to affect lodging, the lack of

QTL overlap is unsurprising and it is likely that the number of QTL affecting this trait

is large.

The current study aims to dissect the genetic architecture of lodging and the

pleiotropic effect of lodging QTL on stay-green and plant height in large-scale multi-

environment field trials in Australia's northern grain belt. We further investigate the

co-localisation of lodging QTL with other traits including QTL for stalk rot disease

resistance and genes in the lignin biosynthesis pathway.

Results

Phenotypic evaluation

Mean lodging in the 17 trials varied from 2.5% to 37.3%, with heritability varying from 33.3% to 89.4% with a mean of 66.0% (Table 3). Genetic correlations of lodging between trials varied from -0.24 to 0.79, with a mean of 0.29 for AYTM and 0.39 for AYTF trials (Supplemental Table S1).

Visual ratings of leaf senescence were recorded in 9 of the 17 trials and varied in both mean and severity of leaf senescence. Across the nine trials, the range of the leaf senescence ratings varied from 1 to 9, with mean ratings varying from 4.2 to 7.8; difference between the ratings of the most and least senescent genotypes within individual trials varied from 4 to 7 (Table 3). Heritability of the trait was generally high, varying from 45.3% to 75.4% with a mean of 65.5%. Genetic correlations of leaf senescence between trials ranged from 0.30 to 0.92, with a mean of 0.53 for AYTM and 0.39 for AYTF trials (Supplemental Table S1).

Plant height was measured in 11 of the 17 trials, with the range varying from 76 to 210 cm, and with mean plant height varying from 109 to 132.4 cm. Differences between the tallest and shortest plants within individual trials varied from 56 to 112 cm (Table 3). Heritability of plant height was very high, varying from 84.9% to 94.2%, with a mean of 88.7%. Genetic correlations of plant height between trials were also very high, varying from 0.76 to 0.97, with a mean of 0.88 for AYTM and 0.86 for AYTF trials (Supplemental Table S1).

Twelve of the 17 trials were harvested and they varied in both the range and mean of grain yield. Across the 12 trials, the range of grain yield varied from 0 to 11.33 t ha^{-1}, with mean yield varying from 2.59 to 7.25 t ha^{-1} (Table 3). Heritability of grain yield varied from 34.9% to 77.1%, with a mean of 56.5%. Genetic correlations of yield between trials varied from -0.07 to 0.71, with a mean of 0.20 for AYTM and 0.11 for AYTF trials (Supplemental Table S1).

Weak to moderate correlations between overall BLUPs for lodging and leaf senescence, and between overall BLUPs for lodging and plant height, were observed, ranging from 0.22 to 0.58, ranged from 0.25 to 0.42 (Figure 1). In individual trials, correlations between lodging and leaf senescence, between lodging

and plant height were also weakly to moderately positive for all five tester
populations (Supplemental Figure S2).

Between-trait correlations of hybrids varied between the testers (Table 2; Figure 1;
Supplemental Figure S2). Comparisons could only be made within female or male
testers but could not be made between male and female testers because even
though they were grown at the same locations, the male and female tester hybrids
were grown in different trials and there were differences in the genetic variance of
the traits in the different parental pools. However, the same general trends were
observed. Testers with low breeding values for leaf senescence in both female and
male trials showed low correlations between leaf senescence and lodging (Table 2;
Figure 1; Supplemental Figure S2). The female tester with the lowest breeding value
for height showed the lowest correlation between hybrid height and lodging. In
contrast, within the male testers the line with the higher breeding value for height
(R986087-2-4-1) displayed a lower association between height and lodging.

QTL identification

Across the 39 tester/trial combinations, a total of 213 unique QTL for lodging were
detected. These QTL were distributed across all 10 chromosomes. Each individual
QTL was detected in multiple tester/trial combinations, with 75% of the QTL
identified in at least five tester/trial combinations (Supplemental Table S2).

The number of QTL identified per tester population differed largely, with the least
number of QTL (i.e. 59 and 64 respectively) detected for the R986087-2-4-1 and
R993396 hybrids of the AYTF trials and the most number of QTL identified for the
B963676 hybrids in the AYTM trials (176 QTL). The majority of the lodging QTL were
in common across tester populations, with 61% (i.e. 129) displaying significant
effects in at least two tester populations (Supplemental Table S2). Of the lodging
QTL which were identified in only one population, the majority (i.e. 60/84) were
identified in the B963676 hybrid trials.

Association of leaf senescence and plant height with lodging QTL

The QTL that affected lodging showed consistent directions of effect for both plant
height and leaf senescence. In both the male and female populations, the QTL
haplotypes with high lodging tended to have high leaf senescence. Similarly, the

QTL haplotypes with high lodging tended to be taller. The 213 QTL by 5 testers

produced 1065 instances where the difference between the extreme lodging

haplotypes could be compared for the other two traits. For leaf senescence in 808 of

1065 instances, the high lodging haplotype corresponded with the high senescence

haplotype. For plant height in 799 of 1065 instances, the high lodging haplotype

corresponded with the tall haplotype. The direction and significance of these

haplotype differences are shown in Supplemental Table S2.

Only 16 lodging QTL were not significantly associated with either plant height or leaf

senescence in at least one tester. In addition, 6 lodging QTL were associated with

leaf senescence or plant height but showed the opposite haplotype effects (i.e. high

lodging haplotypes had lower leaf senescence or were shorter). These 22 QTL were

defined as non-pleiotropic effect QTL (npQTL) (Figure 2; Supplemental Table S2).

Co-localisation with previously identified QTL for lodging and stalk rots

Six of the seven previously identified QTL contributing to lodging resistance (Kebede

et al., 2001; Murray *et al.*, 2008; Srinivasa Reddy *et al.*, 2008) were able to be

projected onto the sorghum genetic linkage consensus map and all 6 overlapped

with lodging QTL identified in this study (Figure 2). These six QTL were identified in

mapping populations of between 93 and 176 individuals and hence had large

confidence intervals.

Two previous studies detected 34 unique QTL for resistance to charcoal rot and/or

Fusarium stalk rot (Adeyanju *et al.*, 2015; Srinivasa Reddy *et al.*, 2008), of which 12

overlapped with lodging QTL detected in this study (Fig 2). Eleven of these 12

overlapping lodging QTL were associated with leaf senescence and/or plant height,

with only one of the charcoal rot QTL co-locating with the npQTL for lodging.

Co-localisation with genes for stem composition

Very few genes for stem composition have been identified and confirmed in sorghum.

Based on phylogenetic relationships with functionally confirmed genes in sorghum

and other species, predicted translational protein functions, enzymatic activity assay,

and gene expression patterns, 21 candidate genes in the lignin biosynthesis

pathways have been identified in sorghum (Bout and Vermerris, 2003; Jun *et al.*,

2017, 2018, Saballos *et al.*, 2012, 2009, Sattler *et al.*, 2009, 2017, Walker *et al.*,

2016, 2013; Wang *et al.*, 2017). Additionally, the major morphological *D* gene, which

changes the structure of the sorghum pith, was recently cloned (Zhang *et al.*, 2018).

Each of these genes was compared to the location of the QTL for lodging identified

in this study.

Twelve of the 21 genes in the lignin biosynthesis pathway in sorghum were located

within the lodging QTL identified in this study, representing a significant enrichment

(χ^2 test, p value = 0.008) (Figure 2). To confirm this enrichment, comparison was

made to multiple sets of 21 randomly selected sorghum house-keeping genes

identified from *Arabidopsis thaliana* (Scheideler *et al.*, 2002) and these were found

not to be significantly enriched within the lodging QTL identified in this study (χ^2 test,

p value = 0.07, average of 20 independent random samplings).

The recently cloned *Dry* gene controls the stem juice content and the composition

and structure of the stem pith, and influences the water content of the stem (Zhang

et al., 2018). For this reason, the gene is likely to affect stem strength. It was found

to be co-located with one of the QTL associated with leaf senescence, QLDGX6.9.

Discussion

Sorghum crops experiencing drought stress during grain filling often lodge, resulting in substantial yield losses. Lodging can be affected by many factors such as plant stature, stay-green and stalk rots. Plant height may further influence lodging by changing the mechanical stress experienced by the stem. If stalk rotting pathogens are present and environmental circumstances are favourable, lodging is further exacerbated by the degradation of the stem. Direct selection for resistance to stalk rots has not proven to be effective in reducing lodging susceptibility. To date, selection for lodging resistance has been based on selection against lodging susceptibility and to some extent selection for stay-green (Jordan *et al.*, 2012). A major challenge to this approach is that genotypes that are more susceptible to lodging also tend to have higher yield potential and hence are more likely to suffer carbohydrate shortages when carbon supply is restricted by drought. As a result, selection for lodging resistance constrains genetic gain for grain yield and vice versa. Little is known about the genetic architecture of the trait due to its complexity. Here, with a view to improving the efficiency of selection for lodging resistance, we present the largest investigation of the genetic architecture of lodging in sorghum to date. The results of this study have revealed the complexity of the genetic architecture of the trait with over 200 QTL identified. Of these QTL, over 90% were also associated with differences in plant height and leaf senescence.

Lodging is under complex genetic control

In this study, we conducted GWAS across 39 separate tester/trial combinations involving 2308 unique elite F_1 hybrids in 17 pre-breeding trials across 9 locations in 3 growing seasons. The populations explored variation in both the male and female parental pools. We identified 213 QTL for lodging, which was equivalent to an average of 5.6 QTL per tester/trial combination. Three quarters of the QTL were detected in at least five and 91 QTL in more than 10 tester/trial combinations (Supplemental Table S2). The large number of QTL identified in this study demonstrates that lodging is a highly quantitative trait, and is one of the most complex traits reported to date.

All previous QTL mapping studies for lodging resistance in sorghum were conducted in small bi-parental populations (93-176 individuals) in a limited number of

environments (1-3). These studies have identified a total of 7 QTL, on average 2.3

QTL/study, none of which overlapped between studies (Kebede *et al.*, 2001; Murray

et al., 2008; Srinivasa Reddy *et al.*, 2008). However, 6 of the 7 QTL projected onto

the sorghum consensus map (Mace *et al.*, 2009) overlapped with QTL identified in

the current study (Figure 2). It was very likely that larger population sizes, larger

genetic diversity of breeding lines, and bigger number of growth environment all

contributed to the identification of more QTL for lodging in this study.

Carbohydrate remobilisation and plant height are the major drivers of lodging

Leaf senescence rated at maturity in sorghum subjected to water stress during grain

filling provides a measure of the degree of remobilisation required to fill the

developing grain (Borrell and Hammer, 2000). The degree of senescence is related

to the imbalance between the demand and supply of carbohydrate within the plant.

Remobilisation of carbohydrate reserves from the stem and leaves ultimately results

in tissue death and predisposes the plant to lodging. The degree of lodging can be

further modified by constitutive factors of the stem and by pathogen invasion

(Henzell *et al.*, 1984). In this study, we found that the lodging score was positively

correlated (p<0.001) with leaf senescence, across environments and genetic

background of testers (Table 2; Figure 1; Supplemental Figure S2A). Sixty-nine

percent of the 213 lodging QTL identified showed a significant positive association

between lodging score and leaf senescence (Supplemental Table S2). This clearly

suggests an important role of carbohydrate remobilisation in lodging. This is in line

with the earlier studies (Henzell *et al.*, 1984; Rosenow, 1980; Rosenow and Clark,

1995) that showed increased lodging was associated with increased leaf

senescence.

This and other studies led to the adoption of the stay-green trait (delayed leaf

senescence) as an additional protective trait in Australia and the USA, where it has

led to significant increases in grain yield as well as reduced lodging (Henzell *et al.*,

1992; Henzell and Hare, 1996; Jordan *et al.*, 2012; Rosenow *et al.*, 1997). The

results from both our study and previous studies indicate that the stay-green trait, a

delayed leaf senescence, is a very useful trait in reducing stalk lodging.

This study also detected a significant positive association between lodging score and

plant height regardless of the environments and genetic background of testers

(Figure 1; Supplemental Figure S2B). A positive association between lodging and

plant height has also been reported by earlier studies (Esechie *et al.*, 1977; Gomez

et al., 2017, 2018; Henzell *et al.*, 1984; Srinivasa Reddy *et al.*, 2008). In agreement

with the phenotypic association between plant height and lodging score, the

haplotype ANOVA results revealed that 66% of the 213 lodging QTL identified in this

study significantly affected lodging susceptibility with taller plants being more

susceptible to lodging (Supplemental Table S2). However, similar to stay-green,

differences in the association between plant height and lodging were observed

amongst the testers (Table 2; Figure 1). Testers with more stay-green were less

susceptible to negative effects of increased plant height on lodging. In fact, the male

tester with a lower leaf senescence value (R986087-2-4-1) produced hybrids with

higher average height but showed a lower association between height and lodging.

Similar protective effects of stay-green were observed when the female testers were

compared. Compared to B010054, the less leaf senescent female tester B963676

that also produced taller hybrids on average showed a lower association between

height and lodging.

We also investigated the impact of yield potential on lodging because high yielding

genotypes tend to remobilise more stem reserves for grain filling, which potentially

causes more lodging. Our data suggest that yield potential is an important factor in

lodging. Overall BLUPs for lodging were predicted to get a measure of lodging

propensity for each hybrid and single trial BLUPs for grain yield were predicted for

each hybrid in individual trials. We calculated the correlation between overall BLUPs

for lodging and single trial BLUPs for yield for trials where lodging did not occur and

observed significant, positive correlations in 24 of the 45 tester/trial combinations

and significant, negative correlations in only 4 combinations (Supplemental Table

S3).

The impact of stalk rotting pathogens

Stalk rots can also accelerate lodging of infected plants. Very few studies have

identified QTL for resistance to stalk rot in sorghum to date. Twelve of the 34 unique

stalk rot resistance QTL previously detected (Adeyanju *et al.*, 2015; Srinivasa Reddy

et al., 2008) overlapped with lodging QTL in this study, 11 of which were associated

with leaf senescence and/or plant height (Figure 2). Given that *Fusarium* spp. and

Macrophomina phaseolina are necrotrophs (Dodd, 1980) and the observation of the

substantial association with remobilisation and/or plant height, it seems likely that

classical resistance genes are of limited importance in the selection for enhanced

lodging resistance.

The impact of stem composition

Twelve of the 21 genes involved in lignin biosynthesis (Bout and Vermerris, 2003;

Jun *et al.*, 2017, 2018, Saballos *et al.*, 2012, 2009, Sattler *et al.*, 2009, 2017, Walker

et al., 2013, 2016; Wang *et al.*, 2017) were located within genomic regions

associated with lodging in this study (Figure 2), representing a significant enrichment

(χ^2 test, p value = 0.008). Lodging resistant sorghum generally have more

lignification, both in the number of cells with lignified walls and in the degree of

lignification in the epidermis, sub-epidermis, and vascular bundles (Schertz and

Rosenow, 1977). Similar results have been previously observed in maize, with

reduced lignin content in the stem resulting in increased stalk lodging (Sun *et al.*,

2018). More recently, Z. Zhang et al. (2019) reported that downregulation of a maize

gene, *stiff1*, resulted in thicker stalk cell walls with increased cellulose and lignin,

which improved stalk strength and reduced lodging.

Previous studies have reported that sorghum lines with both increased (e.g.

SbMyb60 overexpression plants) and reduced lignin content (e.g. *bmr* lines) can

provide the same level of or higher resistance to *Fusarium* spp. and *Macrophomina*

phaseolina than sorghum lines with normal lignin content (Funnell-harris *et al.*, 2010,

2014, 2017, 2018, 2019). This might be due to the inhibition the infection of stalk

rotting pathogens by cell-wall-bound phenolic compounds from monolignol

biosynthesis that can induce defensible pathways (Funnell-harris *et al.*, 2017).

Therefore, the role of lignin in lodging resistance may act in two ways, either by

providing mechanical support to the stem (Bashford *et al.*, 1976) or by restricting the

infection of stalk rotting pathogens.

The recently cloned *Dry* gene controls the stem juice content and the composition

and structure of the stem pith, and influences the water content of the stem (Zhang

et al., 2018). We found that the *Dry* gene co-located with one of the QTL associated

with leaf senescence (QLDGX6.9). Using markers within the *Dry* gene, four

haplotypes were identified and they differed in lodging severity (data not presented).

For the above reasons, we believe that the *Dry* gene is likely to affect stem strength

and thus lodging, which may or may not be associated with stalk rots as reported by

Funnell-Harris et al. (2016) that both susceptible and resistant sweet sorghum lines

to *F.* spp. and *M. phaseolina* had been identified (Funnell-harris *et al.*, 2016).

These two co-localisations with lodging QTL suggest stem composition plays an

important role in lodging, and could be a useful target for selection.

360 Implications for breeding and future research

The identification of 213 QTL in this study demonstrates that lodging is a highly

quantitative trait with multiple QTL each of small effect.

Lodging is affected by many traits with confounding effects. However, one primary

cause of lodging is the imbalance between the supply and demand of carbohydrate

during grain filling. A breeding program selecting for lodging resistance *per se* is

likely to be of limited effectiveness because of the trade-off with yield. Our study

showed that there is a strong association between grain yield potential and lodging

susceptibility (Supplemental Table S3). This suggests that sorghum breeders must

be cautious when selecting directly for lodging resistance as it is likely to significantly

constrain genetic gain for yield. The results support the current practice of direct

selection for increased yield and indirect selection for lodging resistance by selecting

for delayed leaf senescence (stay-green). In addition to its role in reducing lodging,

the integrating stay-green phenotype has been shown to be associated with

increased yield in most Australian sorghum environments (Borrell *et al.*, 2000; Borrell,

van Oosterom, *et al.*, 2014; Henzell *et al.*, 1992; Henzell and Hare, 1996; Jordan *et*

al., 2012) and the USA (Rosenow *et al.*, 1997).

Our study confirmed the undocumented observation by Australian sorghum industry

personnel that increased plant height is associated with increased propensity to

lodging (Table 2; Figure 1; Supplemental Figure S2B). However, as with yield,

sorghum breeders need to be cautious when selecting for lodging resistance through

the selection for short plants because taller plants tend to have higher potential grain

yield (Jordan *et al.*, 2003). Our research indicates that the traits associated with the

integrating stay-green phenotype provides protective effects, weakening the

association between plant height and lodging (Table 2; Figure 1). Therefore, there

would appear to be opportunities to increase genetic yield gain by increasing height

while reducing lodging susceptibility through selection for traits associated with non-

senescence.

Although stalk rots can promote the development and the severity of lodging,

previous studies suggest stalk rot susceptibility is affected by source-sink

relationships and considered to be a problem under post-anthesis stresses (Dodd,

1980; Henzell *et al.*, 1984; Rosenow, 1984; Rosenow and Clark, 1995). Consistent

with these early studies, we found that 12 of the 34 QTL for stalk rot resistance co-

located with lodging QTL in this study, 11 of which were related to leaf senescence

and/or plant height (Figure 2). Direct selection for resistance to stalk rots has not

been proven to be effective in reducing lodging susceptibility. Our data indicates that

classical resistance genes to stalk rots are likely to be of limited importance in

breeding for increased lodging resistance.

Co-localisations of the genes for stem composition with lodging QTL in the study

highlight the importance of stem strength and composition in lodging resistance.

More research is required to investigate the relationship between morphological,

mechanical, and composite traits of the stem and lodging resistance. The

identification of traits related to physical stem strength that can improve resistance to

lodging may prove to be a more effective and efficient way by reducing the

dependency of screening lodging in drought conditions during grain filling.

Materials and Methods

Breeding Trials

This study made use of data from 17 trials of the Australia sorghum pre-breeding program run by the University of Queensland, the Department of Agriculture and Fisheries in Queensland, and the Grains Research and Development Corporation (UQ/DAF/GRDC). The trials were grown in a three year period from 2015-2017 and sampled the major sorghum growing region of Australia from northern New South Wales to central Queensland (from 22.295°S to 31.268°S; from 147.517°E to 152.105°E; Table 1). The 17 trials were selected from a larger set of 38 trials grown by the UQ/DAF/GRDC pre-breeding program during this period on the basis that lodging was recorded in the trial. Lodging is a critical trait for sorghum in Australia and usually scored whenever it occurs.

The trials formed part of the advanced yield testing (AYT) component of the pre-breeding program and contained hybrids that had already been subjected to some degree of selection for performance in preliminary trials. The 17 trials comprised 11 different sowing dates extending from 23 September to 18 February. Typical practice within the breeding program is for two trials to be grown at each site; one AYTM trial consisted of F_1 hybrids derived from crosses between a large number of male parent lines and three female testers (i.e. B010054, B963676, and B986604), while the other AYTF trial consisted of F_1 hybrids derived from crosses between a large number of female parent lines and two male testers (i.e. R986087-2-4-1 and R993396). The five testers varied in levels of stay-green (measured by delayed leaf senescence), plant height, and lodging (Table 2). Although the aim within each trial was to have the same number of hybrids between test lines and the respective testers, the numbers varied because of seed availability. Similarly, it was intended that each AYTM or AYTF trial series contained the same hybrids at each site in a particular year but this was also not achieved due to seed constraints. Since the trials were part of an active pre-breeding program, hybrids were added and dropped from the trial series between seasons. As a result, only a proportion of hybrids were in common between each year within the greatest coincidence being 195 in common between consecutive years.

The total number of hybrids grown per trial varied from 408 to 925 depending on

season and location (Table 1), with entries of the trials including both the test hybrids

detailed above and a range of commercial hybrids. All trials were arranged in a

partially replicated row column trial design; on average between 20-30% of hybrids

were replicated at least twice per trial, while the remaining hybrids were only grown

once. All plots consisted of two rows of 5-metre length. The trials used a solid row

configuration. Row spacing was 0.76 or 1 metre depending on trial. Trials were

managed according to local management practices. The trial identifiers used were a

combination of trial type ("AYTM" or "AYTF"), year (such as "15" for 2015), and

location (e.g. "CAP" for "Capella").

Phenotypes

Lodging was visually scored as the percentage of plants lodged in a plot. Leaf

senescence, as a measure of stay-green, was visually rated from 1 to 9, with 1

indicating less than 10% senescent leaves and 9 indicating over 90% senescent

leaves in a plot. As stay-green (i.e. delayed leaf senescence) was only expressed in

trials suffering from water stress after flowering (Jordan *et al.*, 2012), leaf

senescence ratings were recorded in only 9 of the 17 trials. Plant height was

measured in centimetres from the base of the plant to the tip of the panicle after

maturity and was measured in 11 of the 17 trials. Only 12 of the 17 trials were

harvested; the other 5 trials could not be harvested due to severe lodging just before

harvest. Grains were harvested for individual plots with a small plot harvester with

integrated load cells; grain yield of each plot was later converted to tonnes per

hectare.

Statistical analysis

Phenotypes were analysed using linear mixed models implemented in the asreml-R

package (Butler *et al.*, 2009) using the R software version 3.3.2 (R Core Team,

2016). Genotype was included as a random effect and Best Linear Unbiased

Predictors (BLUPs) for lodging, leaf senescence and plant height were predicted for

all test hybrids in individual trials after accounting for spatial variation and

randomisation processes used in the design. The concurrence of hybrids between

locations and seasons allowed the trials to be included in multi-environment trials

(MET) analysis. As there were no genotypes in common between AYTM and AYTF,

MET analyses were conducted separately for the two types of trials. Factor analytic

(FA) models were implemented in the MET analyses. The FA models accounted for

spatial variation in individual trials and used between-trial correlations as

environmental loadings through a variance matrix across trials (Smith *et al.*, 2001).

An overall BLUP for each genotype across trials was subsequently predicted.

Broad-sense heritability for each trait in individual trials was estimated according to

the formula developed by Cullis et al. (2006) to account for spatial variability.

Genotyping

DNA from mixed leaf blade tissue of two-week-old seedlings of each line was

extracted using procedures described by Diversity Arrays Technology

(http://www.diversityarrays.com). Samples were genotyped with the DArTseq^{TM}

platform, which represents a combination of DArT complexity reduction methods

based on methyl filtration and next-generation sequencing platforms (Kilian *et al.*,

2012). A total of 24,135 SNPs were identified for all female and male lines across

three years, of which 11,435 SNPs were polymorphic. Missing data were imputed

with Beagle version 4.1 (Browning and Browning, 2016), with an accuracy of 94%,

determined through multiple rounds of randomly masking genotypes, imputing

genotype calls and correlating with genotype calls before masking. Following

imputation and removal of SNPs with a minor allele count in less than 5 homozygous

sample, the final marker data sets consisted of between 3,661 and 7,577 SNPs for

the individual AYTM trials and between 3,290 and 3,622 SNPs for the individual

ATYF trails.

Genome-wide association study (GWAS)

GWAS for lodging was conducted separately on parent lines within each tester/trial

combination, in order to avoid the confounding effect of the differences in the genetic

background of the testers and the differences in environments, using FarmCPU (Liu

et al., 2016). As no population structure was observed for hybrids in combination

with each tester, population structure was not included as a covariate in GWAS.

Parameter method.bin was set to "optimum" with the default settings of bin.size and

bin.selection to optimise the possible QTN (quantitative trait nucleotide) window size

and the number of possible QTNs selected into the FarmCPU model. Thirty-nine

separate GWAS were conducted for the thirty-four tester/trial combinations and

hybrids in combination with each of the five testers across trials (five tester

populations). To integrate the GWAS output across the 39 separate analyses, a

comparable method to the mpQTL approach described previously (Mace *et al.*, 2013)

was employed to calculate an integrated probability statistic, to identify regions of the

genome that have a significant contribution to the variation of lodging in the

tester/trial combinations studied. Specifically, the most significant marker for each

tester/trial combination was selected from a sliding window of length 2 cM around

each marker, based on the genetic distances calculated using the sorghum genetic

linkage consensus map (Mace *et al.*, 2009). This process was performed on each

tester/trial combination separately to generate a series of probability values centred

on each marker along each chromosome. The sum of these probability values

across all trial combinations for each tester was then calculated, not including values

where p>0.05 and plotted across the genome for each tester (Supplemental Figure

S1), with a threshold of sum(-log10(p)) >= 5.2 used to identify significance. This

threshold is equivalent to a minimum of 4 trials with significance for lodging at p <

0.05 for each tester, and is more stringent than the criteria of using the Bonferroni

corrected p value (0.05/Number of SNPs) for each single tester/trial combination.

The peak position above the defined significance threshold for each genomic region

across each tester was identified and the results combined across the five testers.

Peaks within 2 cM of each other were considered as the same QTL across testers,

with the peak position considered as the position with the highest sum(-log10(p))

within a 2 cM window. The confidence interval (CI) of a QTL was set to its peak

position ± 1 cM. All QTL identified across the five tester populations were projected

onto the sorghum consensus map (Mace *et al.*, 2009) and visualised with MapChart

v2.32 (Voorrips, 2002).

QTL for lodging identified in the current study were identified as "QLDGX", followed

by chromosome number, a full-stop, and a number suffix indicating the instance of

QTL occurrence on each chromosome, i.e. "QLDGX1.3" refers to the third QTL for

lodging detected on chromosome 1.

QTL haplotype identification

SNPs within the QTL confidence interval of each QTL identified were used for

haplotype identification within all male lines crossed to all three female testers and all

female lines crossed to the two male testers separately. There was only one or no SNP within the 2 cM confidence interval of 5 QTL for female lines and of 2 QTL for male lines; hence, these regions were extended to 4 cM to include a minimum of two SNPs. The final number of SNPs within individual QTL ranged from 2 to 93 depending on the QTL and tester population.

Given the co-ancestry of the breeding populations, the haplotypes represent regions that are identical-by-descent. Haplotypes were identified for both male and female lines using SNPs within a QTL using k-means clustering, with the number of clusters estimated by optimum average Silhouette width implemented in the "pamk" function of the "fpc" R package (Hennig, 2018). Haplotype information based on all male or female lines were then extracted for each tester population.

Mean values of overall BLUPs for lodging score, leaf senescence, and plant height of each haplotype of each QTL × tester population were subsequently calculated. Analysis of variance (ANOVA) of the difference among the means of each trait for the haplotypes within each QTL/tester population was performed to test the effects of lodging QTL on leaf senescence and plant height. Haplotypes whose sample sizes were less than the number of haplotypes plus one were removed to reduce false positives. In order to determine the directions of the effects of lodging QTL on leaf senescence and plant height, mean values of leaf senescence and plant height of the two extreme haplotypes, which had the maximum and minimum average lodging scores, were calculated for each QTL/tester population. Differences in the mean leaf senescence and plant height of the two extreme haplotypes were compared to the corresponding difference in the mean lodging score for each QTL/tester population.

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Author contributions

X.W., D.J., and G.H. conceived the project. X.W., D.J., E.M, Y. T., and A.C. contributed to the development of the analytical framework and the synthesis of the results. X.W. and C.H. conducted the statistical analyses. X.W. conducted GWAS analysis. X.W. wrote the manuscript. All authors read and revised the manuscript.

Conflict of Interest Statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be constructed as a potential conflict of interest.

References

Adeyanju A, Little C, Yu J, Tesso T (2015) Genome-Wide Association Study on
Resistance to Stalk Rot Diseases in Grain Sorghum. *G3 Genes| Genomes| Genetics*
5: 1165–1175.

Bandara YMAY, Weerasooriya DK, Liu S, Little CR (2018) The Necrotrophic Fungus
Macrophomina phaseolina Promotes Charcoal Rot Susceptibility in Grain Sorghum
Through Induced Host Cell-Wall-Degrading Enzymes. *Phytopathology* **108**: 948–956.

Bashford LL, Maranville JW, Weeks SA, Campbell R (1976) Mechanical Properties
Affecting Lodging of Sorghum. *Transactions of the ASAE* **19**: 962–966.

Boerjan W, Ralph J, Baucher M (2003) LIGNIN BIOSYNTHESIS. *Annual Review of*
Plant Biology **54**: 519–546.

Borrell AK, Douglas ACL (1996) Maintaining green leaf area in grain sorghum
increases yield in a water-limited environment. In: Proceedings of the third Australian
sorghum conference. Tamworth, NSW, 315–322.

Borrell AK, Hammer GL (2000) Nitrogen dynamics and the physiological basis of
stay-green in Sorghum. *Crop Science* **40**: 1295–1307.

Borrell AK, Hammer GL, Douglas ACL (2000) Does maintaining green leaf area in
sorghum improve yield under drought? II. Dry Matter Production and Yield. *Crop*
Science **40**: 1026–1037.

Borrell A, Hammer G, Van Oosterom E (2001) Stay-green: A consequence of the
balance between supply and demand for nitrogen during grain filling? *Annals of*
Applied Biology **138**: 91–95.

Borrell AK, Mullet JE, George-Jaeggli B, Van Oosterom EJ, Hammer GL, Klein PE,
Jordan DR (2014a) Drought adaptation of stay-green sorghum is associated with
canopy development, leaf anatomy, root growth, and water uptake. *Journal of*
Experimental Botany **65**: 6251–6263.

Borrell AK, van Oosterom EJ, Mullet JE, George-Jaeggli B, Jordan DR, Klein PE,
Hammer GL (2014b) Stay-green alleles individually enhance grain yield in sorghum
under drought by modifying canopy development and water uptake patterns. *New*
Phytologist **203**: 817–830.

Bout S, Vermerris W (2003) A candidate-gene approach to clone the sorghum *Brown*
midrib gene encoding caffeic acid O-methyltransferase. *Molecular Genetics and*
Genomics **269**: 205–214.

Browning BL, Browning SR (2016) Genotype imputation with millions of reference
samples. *The American Journal of Human Genetics* **98**: 116–126.

Butler DG, Cullis BR, Gilmour AR, Gogel BJ (2009) *ASReml-R reference manual*.

Cha KW, Lee YJ, Koh HJ, Lee BM, Nam YW, Paek NC (2002) Isolation,
characterization, and mapping of the stay green mutant in rice. *Theoretical and*
Applied Genetics **104**: 526–532.

Chapman SC, Cooper M, Hammer GL, Butler DG (2000) Genotype by environment
interactions affecting grain sorghum. II. Frequencies of different seasonal patterns of
drought stress are related to location effects on hybrid yields. *Australian Journal of*
Agricultural Research **51**: 209–221.

Christopher JT, Manschadi AM, Hammer GL, Borrell AK (2008) Developmental and
physiological traits associated with high yield and stay-green phenotype in wheat.
Australian Journal of Agricultural Research **59**: 354–364.

Cullis BR, Smith AB, Coombes NE (2006) On the Design of Early Generation Variety.
Journal of Agricultural, Biological, and Environmental Statistic **11**: 381–393.

Dodd JL (1980) The photosynthetic stress-translocation balance concept of sorghum
stalk rots. In: In Sorghum diseases, a world review: Proceedings of the international
workshop on sorghum diseases, sponsored jointly by Texas A&M University, USA,
and ICRISAT, Patancheru, A.P. 502324, India. pp 300–305.

Esechie HA, Maranville JW, Ross WM (1977) Relationship of Stalk Morphology and
Chemical Composition to Lodging Resistance in Sorghum. *Crop Science* **17**: 609–
612.

Frezzi M, Teyssandier EE (1980) Summay and historical review of sorghum in
Argentina. In: Sorghum Diseases - A World Review. Proceedings of The
International Workshop on Sorghum Diseases. pp 11–14.

Funnell-Harris DL, Gfbru U, States U (2014) Response of *Fusarium thapsinum* to
Sorghum *brown midrib* Lines and to Phenolic Metabolites. *Plant Disease* **98**: 1300–

1308.

Funnell-Harris DL, Neill PMO, Gfbru U, States U (2016) Response of Sweet

Sorghum Lines to Stalk Pathogens *Fusarium thapsinum* and *Macrophomina*

phaseolina. *Plant Disease* **100**: 896–903.

Funnell-Harris DL, Neill PMO, Sattler SE (2018) Field response of near-isogenic

brown midrib sorghum lines to fusarium stalk rot, and response of wildtype lines to

controlled water deficit. *Plant Pathology* **67**: 1474–1482.

Funnell-Harris DL, Neill PMO, Sattler SE, Gries T, Berhow MA, Pedersen JF (2017)

Response of sorghum stalk pathogens to brown midrib plants and soluble phenolic

extracts from near isogenic lines. *European Journal of Plant Pathology* **148**: 941–953.

Funnell-Harris DL, Pedersen JF, Sattler SE (2010) Alteration in Lignin Biosynthesis

Restricts Growth of *Fusarium* spp. in Brown Midrib Sorghum. *Phytopathology* **100**:

671–681.

Funnell-Harris DL, Sattler SE, Neill PMO, Gries T, Tetreault HM, Clemente TE (2019)

Response of Sorghum Enhanced in Monolignol Biosynthesis to Stalk Rot Pathogens.

Plant Disease **103**: 2277–2287.

George-Jaeggli B, Jordan DR, van Oosterom EJ, Hammer GL (2011) Decrease in

sorghum grain yield due to the *dw3* dwarfing gene is caused by reduction in shoot

biomass. *Field Crops Research* **124**: 231–239.

Gomez FE, Muliana AH, Niklas KJ, Rooney WL (2017) Identifying Morphological and

Mechanical Traits Associated with Stem Lodging in Bioenergy Sorghum (*Sorghum*

bicolor). *Bioenergy Research* **10**: 635–647.

Gomez FE, Muliana AH, Rooney WL (2018) Predicting stem strength in diverse

bioenergy sorghum genotypes. *Crop Science* **58**: 739–751.

Hadebe ST, Modi AT, Mabhaudhi T (2017) Drought Tolerance and Water Use of

Cereal Crops: A Focus on Sorghum as a Food Security Crop in Sub-Saharan Africa.

Journal of Agronomy and Crop Science **203**: 177–191.

Hennig C (2018) fpc: Flexible Procedures for Clustering. R package version 2.1-11.1.

Henzell R, Brengman RL, Fletcher DS, McCosker AN (1992) Relationship between

yield and staygreen in some grain sorghum hybrids grown under terminal drought

stress. In: Proceedings of the Second Australian Sorghum Conference. 355–359.

Henzell RG, Dodman RL, Done AA, Brengman RL, Mayers PE (1984) Lodging, Stalk

Rot, and Root Rot in Sorghum in Australia. In: Sorghum Root and Stalk Rots, A

Critical Reivew: Proceedings of the Consultative Group Discussion on Research

Needs and Strategies for Control of Sorghum Root and Stalk Rot Diseases

(Mughogho L, Rosenberg G, eds) pp 225–236.

Henzell RG, Hare BW (1996) Sorghum Breeding in Austrlaia - Public and Private

Endeavours. In: Proceedings of the Third Australian Conference. pp 159–171.

Higuchi T (2006) Look back over the studies of lignin biochemistry. *Journal of Wood*

Science **52**: 2–8.

Johnson JW, Stegmeier WD, Andrews DJ, Rosenow DT, Henzell RG, Monk RL

(1997) Genetic Resistance to Lodging. In: Proceedings of The International

Conference on Genetic Improvement of Sorghum and Pearl Millet. Lubbock, Texas,

pp 481–489.

Jordan DR, Hunt CH, Cruickshank AW, Borrell AK, Henzell RG (2012) The

Relationship Between the Stay-Green Trait and Grain Yield in Elite Sorghum Hybrids

Grown in a Range of Environments. *Crop Science* **52**: 1153–1161.

Jordan DR, Tao Y, Godwin ID, Henzell RG, Cooper M, McIntyre CL (2003)

Prediction of hybrid performance in grain sorghum using RFLP markers. *Theoretical*

and Applied Genetics **106**: 559–567.

Jun S-Y, Sattler SA, Cortez GS, Vermerris W, Sattler SE (2018) Biochemical and

Structural Analysis of Substrate Specificity of a Phenylalanine Ammonia-Lyase. *Plant*

Physiology **176**: 1452–1468.

Jun S-Y, Walker AM, Kim H, Ralph J, Vermerris W, Sattler SE (2017) The Enzyme

Activity and Substrate Specificity of Two Major Cinnamyl Alcohol Dehydrogenases in

Sorghum (*Sorghum bicolor*), SbCAD2 and SbCAD4. *Plant Physiology* **174**: 2128–

2145.

Kebede H, Subudhi PK, Rosenow DT, Nguyen HT (2001) Quantitative trait loci

influencing drought tolerance in grain sorghum (*Sorghum bicolor* L. Moench).

Theoretical and Applied Genetics **103**: 266–276.

Kholová J, Mclean G, Vadez V, Craufurd P, Hammer GL (2013) Drought stress characterization of post-rainy season (rabi) sorghum in India. *Field Crops Research* **141**: 38–46.

Kilian A, Wenzl P, Huttner E, Carling J, Xia L, Blois H, Caig V, Heller-Uszynska K, Jaccoud D, Hopper C, *et al* (2012) Diversity Arrays Technology: A Generic Genome Profiling Technology on Open Platforms. In: Data Production and Analysis in Population Genomics. Methods in Molecular Biology (Methods and Protocols) (Pompanon F, Bonin A, eds), pp 67–89. Totowa: Humana Press.

Liu X, Huang M, Fan B, Buckler ES, Zhang Z (2016) Iterative Usage of Fixed and Random Effect Models for Powerful and Efficient Genome-Wide Association Studies. *PLoS Genetics*: 1–24.

Mace ES, Hunt CH, Jordan DR (2013) Supermodels: sorghum and maize provide mutual insight into the genetics of flowering time. *Theoretical and Applied Genetics* **126**: 1377–1395.

Mace ES, Rami J-FF, Bouchet S, Klein PE, Klein RR, Kilian A, Wenzl P, Xia L, Halloran K, Jordan DR (2009) A consensus genetic map of sorghum that integrates multiple component maps and high-throughput Diversity Array Technology (DArT) markers. *BMC plant biology* **9**: 13.

Mace ES, Singh V, Van Oosterom EJ, Hammer GL, Hunt CH, Jordan DR (2012) QTL for nodal root angle in sorghum (*Sorghum bicolor* L. Moench) co-locate with QTL for traits associated with drought adaptation. *Theoretical and Applied Genetics* **124**: 97–109.

Murray SC, Rooney WL, Mitchell SE, Sharma A, Klein PE, Mullet JE, Kresovich S (2008) Genetic improvement of sorghum as a biofuel feedstock: II. QTL for stem and leaf structural carbohydrates. *Crop Science* **48**: 2180–2193.

Pinthus MJ (1974) Lodging in wheat, barley, and oats: The phenomenom, its causes, and preventive measures. *Advances in Agronomy* **25**: 209–263.

R Core Team (2016) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-

project.org/.

Rosenow DT (1977) Breeding for lodging resistance in sorghum. In: Proceedings of
the 32nd Annual Corn and Sorghum Research Conference. Dec. 6-8, Chicago, IL,
171–185.

Rosenow DT (1980) Stalk rot resistance breeding in Texas. In: Sorghum Diseases, A
Wrold Review: Proceedings of the International Workshop on Sorghum Diseases.
(ICRISAT, ed), pp 306–314. Hyderabad, India

Rosenow DT (1984) Breeding for resistance to root and stalk rots in Texas. In:
Interanational Crops Research Institute for the Semi-Arid Tropics, ed. Sorghum Root
and Stalk Rots, A Critical Reivew: Proceedings of the Consultative Group Discussion
on Research Needs and Strategies for Control of Sorghum Root and Stalk Rot
Diseases, 27 Nov - 2 Dec 1983, Bellagio, Italy. Patancheru, A.P. 502324, India: ICR.
209–218.

Rosenow DT, Clark LE (1995) Drought and lodging resistance for a quality sorghum
crop. In: Fiftieth annual corn and sorghum industry research conference, December
6-7, 1995. Chicago, IL: American Seed Trade Association, 82–97.

Rosenow DT, Ejeta G, Clark LE, Gilbert ML, Henzell RG, Borrell AK, Muchow RC
(1997) Breeding for Pre- and Post-Flowering Drought Stress Resistance in Sorghum.
In: Proceedings of The International Conference on Genetic Improvement of
Sorghum and Pearl Millet. 400–411.

Rosenow DT, Quisenberry JE, Wendt CW, Clark LE (1983) Drought tolerant
sorghum and cotton germplasm. *Agricultural Water Management* **7**: 207–222.

Saballos A, Ejeta G, Sanchez E, Kang C, Vermerris W (2009) A Genomewide
Analysis of the Cinnamyl Alcohol Dehydrogenase Family in Sorghum [*Sorghum
bicolor* (L.) Moench] Identifies *SbCAD2* as the *Brown midrib6* Gene. *Genetics* **181**:
783–795.

Saballos A, Sattler SE, Sanchez E, Foster TP, Xin Z, Kang C, Pedersen JF,
Vermerris W (2012) *Brown midrib2* (*Bmr2*) encodes the major 4-coumarate□:
coenzyme A ligase involved in lignin biosynthesis in sorghum (*Sorghum bicolor* (L.)
Moench). *The Plant Journal* **70**: 818–830.

Sattler SE, Saathoff AJ, Haas EJ, Palmer NA, Funnell-Harris DL, Sarath G,

Pedersen JF (2009) A Nonsense Mutation in a Cinnamyl Alcohol Dehydrogenase

Gene Is Responsible for the Sorghum *brown midrib6* Phenotype. *Plant Physiology*

150: 584–595.

Sattler SA, Walker AM, Vermerris W, Sattler SE, Kang C (2017) Structural and

Biochemical Characterization of Cinnamoyl-CoA Reductases. *Plant Physiology* **173**:

1031–1044.

Scheideler M, Schlaich NL, Fellenberg K, Beissbarth T, Hauser NC, Vingron M,

Slusarenko AJ, Hoheisel JD (2002) Monitoring the switch from housekeeping to

pathogen defense metabolism in Arabidopsis thaliana using cDNA arrays. *Journal of*

Biological Chemistry **277**: 10555–10561.

Schertz KF, Rosenow DT (1977) Anatomical Variation in Stalk Internodes of

Sorghum. *Crop Science* **17**: 628–631.

Smith A, Cullis B, Thompson R (2001) Analyzing Variety by Environment Data Using

Multiplicative Mixed Models and Adjustments. *Biometrics* **57**: 1138–1147.

Srinivasa Reddy P, Fakrudin B, Rajkumar, Punnuri SM, Arun SS, Kuruvinashetti MS,

Das IK, Seetharama N (2008) Molecular mapping of genomic regions harboring

QTLs for stalk rot resistance in sorghum. *Euphytica* **159**: 191–198.

Sun Q, Liu X, Yang J, Liu W, Du Q, Wang H, Fu C, Li W (2018) microRNA528

Affects Lodging Resistance of Maize by Regulating Lignin Biosynthesis under

Nitrogen-Luxury Conditions. *Molecular Plant* **11**: 806–814.

Voorrips RE (2002) MapChart: Software for the Graphical Presentation of Linkage

Maps and QTLs. *Journal of Heredity* **93**: 77–78.

Walker AM, Hayes RP, Youn B, Vermerris W, Sattler SE, Kang C (2013) Elucidation

of the Structure and Reaction Mechanism of Sorghum Hydroxycinnamoyltransferase

and Its Structural Relationship to Other Coenzyme A-Dependent Transferases and

Synthases. *Plant Physiology* **162**: 640–651.

Walker AM, Sattler SA, Regner M, Jones JP, Ralph J, Vermerris W, Sattler SE, Kang

C (2016) The Structure and Catalytic Mechanism of *Sorghum bicolor* Caffeoyl-CoA

O-Methyltransferase. *Plant Physiology* **172**: 78–92.

Wang J, Feng J, Jia W, Fan P, Bao H, Li S, Li Y (2017) Genome-Wide Identification
of *Sorghum bicolor* Laccases Reveals Potential Targets for Lignin Modification.
Frontiers in Plant Science **8**.

Zhang J, Fengler KA, Van Hemert JL, Gupta R, Mongar N, Sun J, Allen WB, Wang Y,
Weers B, Mo H, Lafitte R, Hou Z, Bryant A, Ibraheem F, Arp J, Swaminathan K,
Moose SP, Li B and Shen B (2019) Identification and characterization of a novel
stay-green QTL that increases yield in maize. *Plant Biotechnology Journal* **17**: 2272–
2285.

Zhang L-M, Leng C-Y, Luo H, Wu X-Y, Liu Z-Q, Zhang Y-M, Zhang H, Xia Y, Shang
L, Liu C-M, Hao D-Y, Zhou Y, Chu C, Cai H-W, Jing H-C (2018) Sweet Sorghum
Originated through Selection of *Dry* , a Plant-Specific NAC Transcription Factor
Gene. *The Plant Cell* **30**: 2286–2307.

Zhang Z, Zhang X, Lin Z, Wang J, Liu H, Zhou L (2019) A Large Transposon
Insertion in the *stiff1* Promoter Increases Stalk Strength in Maize. *The Plant Cell*.
DOI: 10.1105/tpc.19.00486

Figure Legends

Figure 1 Correlations between overall BLUPs for leaf senescence, plant height, and lodging by tester.

B010054, B963676, B986604, R986087-2-4-1, and R993396 are the five tester parents of the hybrids grown in the 17 yield testing trials.

Figure 2 Projection of the 213 lodging QTL identified across testers onto the sorghum consensus genetic map and their co-localisation with previously identified QTL for lodging and stalk rots and genes in lignin biosynthesis pathway.

The 213 lodging QTL identified as horizontal segments on each chromosome and colour-coded as follows: lodging QTL that significantly affect leaf senescence (green), lodging QTL that significantly affect plant height (pink), lodging QTL that significantly affect both leaf senescence and plant height (black), QTL did not affect either leaf senescence or plant height (red). Brown vertical bars to the right of each chromosome marked as "LDG" represent the locations of QTL previously detected for lodging (Kebede *et al.*, 2001; Murray *et al.*, 2008; Srinivasa Reddy *et al.*, 2008); the black bars indicate the location of a QTL previously reported for charcoal rot and/or Fusarium stalk rot (Adeyanju *et al.*, 2015; Srinivasa Reddy *et al.*, 2008). The location of genes in the lignin biosynthesis pathways that co-located with lodging QTL detected in this study are highlighted in green to the right of each chromosome, and those that did not co-locate with lodging QTL are highlighted in blue (Bout and Vermerris, 2003; Jun *et al.*, 2017, 2018, Saballos *et al.*, 2012, 2009, Sattler *et al.*, 2009, 2017, Walker *et al.*, 2013, 2016; Wang *et al.*, 2017). The location of the recently cloned *Dry* gene is highlighted in red (Zhang *et al.*, 2018).

Supplementary Information

Figure S1 Diagram of the method to integrate the GWAS output across the 39

separate analyses, a comparable method to the mpQTL approach describe by Mace

et al. (2013).

To identify genomic regions that have a significant contribution to the variation of

lodging, an integrated probability statistic was calculated by summing the -log10(p)

of the p values (5th to 10th columns) of the most significant SNP for each tester

(B010054 in this example) and trial combination from a sliding window of length 2 cM

around each SNPs (A). The process was performed on each tester/trial combination

separately to generate a series of probability values centred on each SNP along

each chromosome. The sum of these probability values across all trial combinations

for each tester was then calculated, excluding values where p>0.05 and plotted

across the genome for each tester (B). The position of the SNP in the centred of the

2cM sliding window (shaded in grey) is highlighted in orange and the corresponding

sum(-log10(p)) within the window highlighted in green; values highlighted in yellow

indicates the most significant p values for the corresponding tester/trial combination

within the 2cM window, whereas blue highlighted values indicates p values that are <

0.05 but not included in the calculation of sum(-log10(p)).

Figure S2 Correlations between BLUPs for lodging and leaf senescence (A), lodging

and plant height (B) in individual trials by tester.

Table S1 Between-trial genetic correlations of lodging, leaf senescence, plant height,

and grain yield.

Table S2 Details of lodging QTL detected across tester populations.

Table S3 Correlation between overall BLUPs for lodging score and single trial

BLUPs for grain yield in all 38 trials by tester. Mean lodging score, mean leaf

senescence rating, and mean yield of the tester/trial combinations are also

presented. Five trials, AYTM15CUR, AYTM16KIL, AYTM17DAL, AYTF16EME, and

AYTF17DAL, were not harvested due to lodging; therefore, yield BLUPs in these

trials could not be predicted and correlations in these trials could not be calculated

Table 1 Description of the 17 field trials in the 2015-17 growing seasons.

Trial code	Trial	Sowing date	Location	GPS coordinates (Lat; Long)	Altitude (m asl)	No. of unique hybrids[†]	No. of test hybrid derived from				
							B010054	B963676	B986604	R986087-2-4-1	R993396
1	AYTM15CUR	28/10/2014	Curlewis	-31.268 150.222	294	580	222	292	-[‡]	-	-
2	AYTM15WAR	3/12/2014	Warwick	-28.214 152.105	465	582	217	296	-	-	-
3	AYTM15JIM	17/12/2014	Jimbour	-27.029 151.198	347	575	214	296	-	-	-
4	AYTM15CAP	11/01/2015	Capella	-23.222 148.114	235	590	216	303	-	-	-
5	AYTM15EME	12/01/2015	Emerald	-23.529 148.213	161	611	228	309	-	-	-
6	AYTM16CRO	23/09/2015	Croppa Creek	-29.046 150.332	277	710	-	397	293	-	-
7	AYTF16CRO	23/09/2015	Croppa Creek	-29.046 150.332	277	493	-	-	-	253	220
8	AYTM16WAR	21/12/2015	Warwick	-28.211 152.104	459	925	-	457	447	-	-
9	AYTF16WAR	21/12/2015	Warwick	-28.211 152.104	459	532	-	-	-	269	240
10	AYTM16ORI	15/02/2016	Orion	-24.231 148.411	234	706	-	393	289	-	-
11	AYTF16ORI	15/02/2016	Orion	-24.231 148.411	234	467	-	-	-	236	206
12	AYTM16EME	16/02/2016	Emerald	-23.529 148.213	161	835	-	418	393	-	-
13	AYTF16EME	16/02/2016	Emerald	-23.529 148.213	161	478	-	-	-	245	208
14	AYTM16KIL	18/02/2016	Kilcummin	-22.295 147.517	280	527	-	289	214	-	-
15	AYTF16KIL	18/02/2016	Kilcummin	-22.295 147.517	280	408	-	-	-	203	180
16	AYTM17DAL	13/10/2016	Dalby Box	-27.350 151.176	343	543	-	277	240	-	-
17	AYTF17DAL	13/10/2016	Dalby Box	-27.350 151.176	343	881	-	-	-	360	503

[†]No. of unique hybrids included both the test hybrids and a range of commercial hybrids; [‡]"-" indicates hybrids derived from the tester were not grown in the corresponding trial.

Table 2 Overall Best Linear Unbiased Predictors (BLUPs) for lodging score (%), leaf senescence rating (1-9), plant height (cm), and

grain yield (t ha^{-1}), and the between trait correlations of the female and male testers across multiple trials.

Tester	Tester type	Lodging score (%)	Leaf senescence rating	Plant height (cm)	Yield (t ha^{-1})	Correlation coefficient between lodging and leaf senescence rating	Correlation coefficient between lodging and plant height
B010054	Female	9.9	6.4	120.1	5.13	0.57	0.42
B963676	Female	7.7	6.0	121.3	5.56	0.34	0.37
B986604	Female	7.0	6.2	116.2	5.53	0.22	0.32
R986087-2-4-1	Male	10.1	6.4	125.2	5.22	0.57	0.25
R993396	Male	10.8	6.7	122.0	5.07	0.58	0.28

Table 3 Trait characteristics of hybrids grown in the 17 advanced yield testing (AYT) trials for males (AYTM) and females (AYTF) lines.Trials were sorted in ascending order for mean lodging score (%).

Trial code	Trial	Sowing date	Lodging (%)			Leaf senescence			Height (cm)			Yield (t ha^{-1})		
			Range	Mean	H^2 (%)	Range	Mean	H^2 (%)	Range	Mean	H^2 (%)	Range	Mean	H^2 (%)
4	AYTM15CAP	11/01/2015	0 - 75	2.5	84.4	5 - 9	7.4	67.1	84 - 182	121.8	88.7	1.08 - 4.57	3.01	34.9
3	AYTM15JIM	17/12/2014	0 - 80	5.4	89.4	3 - 9	6.3	75.3	-	-	-	1.47 - 7.03	4.33	69.7
11	AYTF16ORI	15/02/2016	2 - 50	9.0	36.5	-	-	-	76 - 146	115.7	84.9	0.84 - 5.80	2.86	36.1
12	AYTM16EME	16/02/2016	0 - 95	10.7	60.1	1 - 8	4.2	52.0	80 - 164	109.3	85.1	0.00 - 8.07	2.63	54.2
8	AYTM16WAR	21/12/2015	10 - 70	12.6	33.3	3 - 9	6.0	74.1	92 - 200	123.5	90.3	2.64 - 11.33	7.25	63.6
10	AYTM16ORI	15/02/2016	2 - 90	14.7	62.5	-	-	-	76 - 160	109.0	90.2	0.07 - 6.39	2.90	56.9
5	AYTM15EME	12/01/2015	0 - 80	15.6	85.4	4 - 9	7.4	72.3	96 - 168	121.0	88.8	1.00 - 6.11	3.13	68.0
15	AYTF16KIL	18/02/2016	0 - 95	17.4	57.0	-	-	-	92 - 148	124.9	91.4	0.00 - 6.22	2.59	61.1
6	AYTM16CRO	23/09/2015	10 - 80	19.0	53.8	-	-	-	-	-	-	3.18 - 7.76	5.78	37.4
2	AYTM15WAR	3/12/2014	10 - 90	23.4	79.3	-	-	-	98 - 210	132.4	86.2	3.05 - 10.22	7.00	77.1
7	AYTF16CRO	23/09/2015	10 - 80	25.3	67.3	-	-	-	-	-	-	1.50 - 7.84	5.40	44.5
9	AYTF16WAR	21/12/2015	10 - 90	25.9	57.9	5 - 9	7.8	75.4	100 - 164	129.7	87.7	3.13 - 10.64	6.66	74.6
16	AYTM17DAL	13/10/2016	10 - 90	28.2	68.4	4 - 9	6.3	65.5	-	-	-	-	-	-
14	AYTM16KIL	18/02/2016	0 - 100	31.3	64.0	-	-	-	88 - 172	117.3	94.2	-	-	-
13	AYTF16EME	16/02/2016	0 - 100	32.2	67.2	2 - 8	6.0	45.3	84 - 144	121.6	87.8	-	-	-
17	AYTF17DAL	13/10/2016	10 - 90	33.1	72.9	3 - 9	6.0	62.5	-	-	-	-	-	-
1	AYTM15CUR	28/10/2014	10 - 90	37.3	82.7	-	-	-	-	-	-	-	-	-