Pharmacological inhibition of acid sensing ion channel 1a protects the heart from ischemia-reperfusion injury

Meredith A. Redd[1,9,12], Sarah E. Scheuer[2,3,4,12], Natalie J. Saez[1], Ling Gao[2], Mark Hicks[2,4,5], Jeanette E. Villanueva[2,4], Melissa E. Reichelt[6], Jason N. Peart[7], Louise E. See Hoe[8,9], Han S. Chiu[1], Xiaoli Chen[1], Yuliangzi Sun[1], Jacky Y. Suen[8,9], Snezana Maljevic[10], Walter G. Thomas[6], Gabriel Cuellar-Partida[11], John F. Fraser[8,9], Steven Petrou[10], Glenn F. King[1,13], Peter S. Macdonald[2,3,4,13] & Nathan J. Palpant[1,13]

[1]Institute for Molecular Bioscience, The University of Queensland, St. Lucia, QLD, Australia
[2]Victor Chang Cardiac Research Institute, Sydney, Australia
[3]Cardiopulmonary Transplant Unit, St. Vincent's Hospital, Sydney, Australia
[4]Faculty of Medicine, University of New South Wales, Sydney, Australia
[5]Department of Pharmacology, St. Vincent's Hospital, Sydney, Australia
[6]School of Biomedical Sciences, The University of Queensland, St. Lucia, Australia
[7]School of Medical Science, Griffith University, Southport, Australia
[8]Faculty of Medicine, The University of Queensland, Brisbane, QLD, Australia
[9]Critical Care Research Group, The Prince Charles Hospital and The University of Queensland, Brisbane, QLD, Australia
[10]Florey Institute of Neuroscience and Mental Health, University of Melbourne, Melbourne, Australia
[11]The University of Queensland Diamantina Institute, Faculty of Medicine and Translational Research Institute, Woolloongabba, QLD, Australia
[12]These authors contributed equally
[13]Corresponding authors

Addresses for correspondence:

Dr. Nathan J. Palpant
Institute for Molecular Bioscience
306 Carmody Road, Building 80 (Level 4 North)
St. Lucia, Queensland, Australia, 4072
T: +61 7 3346 2054
E: n.palpant@imb.uq.edu

Professor Glenn F. King
Institute for Molecular Bioscience
306 Carmody Road, Building 80 (Level 2 North)
St. Lucia, Queensland, Australia, 4072
T: +61 7 3346 2025
E: glenn.king@imb.uq.edu.au

Professor Peter Macdonald
Cardiopulmonary Transplant Unit, St. Vincent's Hospital
Darlinghurst, New South Wales, Australia
T: +61 2 8382 2641
E: peter.macdonald@svha.org.au

Abstract

Acid-sensing ion channel 1a (ASIC1a) is a proton-gated channel that is activated by tissue acidosis. We utilized genetic loss of function to demonstrate that ASIC1a mediates the injury response to myocardial ischemia-reperfusion injury (IRI). We show that pharmacological inhibition of ASIC1a affords cardioprotection across a range of rodent and human models of cardiac ischemia, resulting in improved cardiac viability and functional recovery post-IRI. Consistent with this data, we show that polymorphisms in the ASIC1 gene locus are associated with myocardial infarction and small-vessel stroke in humans. This study provides compelling evidence that ASIC1a is a novel target for cardioprotective drugs that could reduce the burden of disease associated with myocardial ischemia.

Introduction

Conditions caused by obstruction of blood flow to the heart, such as myocardial infarction (MI), are the most common emergency manifestation of cardiovascular disease[1]. Although acute reperfusion therapies have improved patient outcomes, mortality remains high[2] and MI is one of the largest attributable risks for heart failure (HF)[3]. Globally, 1 in 5 people develop HF, with an annual estimated $108B in healthcare costs[4,5]. Heart transplantation remains the most effective treatment option for HF[6,7], but 75% of potential donor hearts are discarded, many due to sensitivity of the donor heart to ischemic injury[8]. Myocardial sensitivity to ischemia-reperfusion injury (IRI) therefore remains a primary point of vulnerability underlying cardiovascular disease, which is the leading cause of morbidity and mortality worldwide[9]. Despite decades of preclinical therapeutic development, there are currently no drugs that block the acute injury response to cardiac ischemia[10].

Myocardial IRI is a complex pathophysiological process that underlies cardiac injury sustained during cardiac surgery, heart transplant, MI, and cardiac arrest. During myocardial ischemia, reduced oxygen availability causes a shift from fatty acid metabolism to anaerobic glycolysis[11,12]. The resulting lactic acidosis causes the extracellular pH to fall to as low as 6.0 – 6.5[13,14]. In patients suffering from acute MI, the severity of acidosis strongly correlates with patient mortality, with pH < 7.35 associated with > 60% mortality[15]. Ongoing acidosis results in the activation of several transmembrane ion channels and receptors, including the sodium-hydrogen exchanger (NHE), glutamate-gated NMDA receptors, and TRP family of ion channels, each of which are thought to contribute to calcium overload, metabolic dysfunction, and eventually cell death[16-18]. In line with this, the Expedition Trial showed significant protection against peri-operative MI with the NHE inhibitor, cariporide, in patients undergoing high-risk coronary artery bypass surgery[19]. While this trial failed due to cerebrovascular complications, it demonstrated that pharmacological conditioning has the capacity to reduce the injury response to myocardial IRI.

Acid sensing ion channels (AISCs) are voltage-independent proton-gated cation channels of the degenerin/epithelial sodium channel superfamily. There are six ASIC isoforms derived from four genes (*ACCN1–4*), which assemble into homotrimeric or heterotrimeric channels. The pH sensitivity and kinetics of ASICs are determined by their subunit composition[20,21]. ASICs (similar to TRP channels and GPCRs) are involved in extracellular acidification-induced calcium overload in neurons and cardiomyocytes[22-24]. ASIC1a, a splice variant of the *ACCN1* gene, is the most pH-sensitive ASIC channel. Activation begins at pH ≤ 7, with half-maximal activation at pH 6.6[25,26], and ASIC1a currents are potentiated by numerous associated metabolic events that occur during ischemia including membrane stretch and increased levels of extracellular lactate, pyruvate, and arachidonic acid[27,28]. ASIC1a plays a critical role in mediating neuronal death after cerebral

ischemia, and inhibition of ASIC1a through genetic ablation or pharmacological blockade is neuroprotective in rodent models of ischemic stroke[29-31].

In this study, we used genetic approaches to demonstrate, for the first time, that ASIC1a is a critical mediator of the injury response to myocardial IRI. We show that venom-derived peptide inhibitors of ASIC1a provide robust cardioprotection across multiple species and model systems from the cellular to whole organ level, including models of IRI and donor heart preservation. Our data reveal that ASIC1a is a novel target for cardioprotective drugs, with potential applications for the treatment of MI, cardiac arrest, and the preservation of donor hearts.

Results

Genetic ablation of ASIC1a improves functional recovery following cardiac IRI. We assessed ASIC isoform expression in the adult mouse heart from transcriptomic data of sorted cardiac cell populations after sham and myocardial infarction (MI)[32]. In sham hearts, cardiomyocytes and endothelial cells expressed high levels of *ASIC1*, while endothelial cells, fibroblasts, and leukocytes express both *ASIC1* and *ASIC3*. In all four cell types, *ASIC2* and *ASIC4* had low to undetectable expression (**Fig. 1a**). The expression of *ASIC1* was not differentially expressed between sham and post-MI conditions in cardiomyocytes or endothelial cells (**Supplementary Fig. 1a**).

To determine whether ASIC1a plays a functional role during cardiac ischemia, we assessed IRI tolerance of Langendorff-perfused isolated hearts from ASIC1a-specific knockout (KO) mice. Baseline function and heart rate in ASIC1a KO (ASIC1a$^{-/-}$) isolated hearts were comparable to those measured in wildtype (WT) controls (ASIC1a$^{+/+}$) (**Table 1**). To assess tolerance to IRI, hearts were subjected to 25 minutes of global normothermic zero-flow ischemia, followed by 45 minutes of aerobic reperfusion. WT and ASIC1a KO hearts showed similar initial responses to ischemia with comparable levels of ventricular contracture (**Supplementary Fig. 1b**). During reperfusion, both groups demonstrated gradual recovery of contractile function over time, with markedly improved functional recovery in ASIC1a KO hearts (**Fig. 1b**). By the end of the reperfusion period, ASIC1a KO hearts had significantly improved systolic and diastolic function with higher left ventricular developed pressure (LVDP) (44 ± 5% of baseline) and lower end diastolic pressure (EDP) (38 ± 3 mmHg) compared to WT hearts (LVDP: 23 ± 7% of baseline; EDP: 50 ± 3 mmHg); **Fig. 1c,d**). ASIC1a KO hearts also had improved recovery of coronary flow (CF) rates by the end of reperfusion (KO: 97 ± 6%; WT: 78 ± 3%, percent baseline) (**Fig. 1e** and **Supplementary Fig. 1c,d**). To assess cell death, lactate dehydrogenase (LDH) efflux was measured in perfusate samples collected during reperfusion. LDH efflux from ASIC1a KO hearts was significantly lower compared to WT hearts after 2 min of reperfusion (**Fig. 1f**), with a similar trend at the end of reperfusion (**Supplementary Fig.**

1d). Our data provide strong evidence that ASIC1a has no evident role in maintaining functional homeostasis, but it is a critical mediator of the organ response to myocardial IRI.

ASIC1a inhibitors protect mouse hearts against IRI. We next sought to determine whether pharmacologic inhibition of ASIC1a provides cardioprotection during an acute cardiac ischemic insult. For this purpose, we took advantage of two venom-derived inhibitors of ASIC1a. Hi1a, a 76-residue double-knot peptide isolated from the venom of an Australian funnel-web spider (**Fig. 2a**), is the most potent and selective inhibitor of ASIC1a identified to date, with an IC_{50} of 500 pM[31]. PcTx1, a single-knot peptide from the venom of a tarantula, is also a potent and selective inhibitor of ASIC1a (IC_{50} ~1 nM)[29]. Although the two peptides are closely related (Hi1a is comprised of two PcTx1-like domains joined by a short linker; **Fig. 2b,c**), they have distinct inhibitory modes of action. Hi1a inhibits ASIC1a activation whereas PcTx1 promotes and stabilizes a desensitized state of the channel[31]. We also utilized an analogue of PcTx1 that contains mutations of two pharmacophore residues (R27A/V32A) which dramatically reduces its inhibitory effect on ASIC1a[29].

To examine if these ASIC1a inhibitors are cardioprotective, we assessed tolerance to IRI in Langendorff-perfused isolated mouse hearts with and without peptide treatment. Consistent with genetic ablation of ASIC1a, Hi1a, PcTx1, and PcTx1-R27A/V32A had no effect on baseline contractile function during the first 10 min of peptide infusion prior to ischemia (**Fig. 2d**). Functional tolerance to IRI was greater in hearts exposed to Hi1a or PcTx1 (10 nM) compared to control hearts (**Fig. 2e**). At the end of reperfusion, Hi1a- and PcTx1-treated hearts, but not hearts treated with the PcTx1-R27A/V32A analogue, had markedly improved recovery of LVDP (Hi1a: 60 ± 4%, PcTx1: 49 ± 12%, PcTx1-R27A/V32A: 36 ± 6%) compared to vehicle controls (20 ± 4%) (**Fig. 2f**). Similarly, treatment with Hi1a and PcTx1, but not PcTx1-R27A/V32A, led to reduced EDP after 45 min reperfusion (Hi1a: 35 ± 4 mmHg; PcTx1: 33 ± 8 mmHg; PcTx1-R27A/V32A: 48 ± 3 mmHg) compared to vehicle controls (57 ± 2 mmHg) (**Fig. 2g**). No differences in final CF were observed between groups (**Fig 2h**), although hearts treated with PcTx1, but not Hi1a, displayed significant reactive hyperaemia during early reperfusion, as evidenced by increased CF during the first 5 min of reperfusion (**Supplementary Fig. 2a,b**). Taken together, our data show that ASIC1a inhibitors protect the heart from myocardial IRI and recapitulate the functional benefits of genetic ablation of the channel.

Preconditioning with Hi1a improves functional recovery of isolated rodent hearts after prolonged hypothermic ischemia. In order to evaluate the cardioprotective effect of ASIC1a inhibition in a model of donor heart preservation, we supplemented standard clinical heart preservation solution (Celsior) with Hi1a during 8 h cold storage of isolated rat hearts. The hearts were stabilised for 10 min by retrograde (Langendorff) perfusion and then switched to working-mode in order to assess cardiac function in the presence of physiological afterload. Following assessment

of baseline haemodynamic parameters, the hearts were arrested by infusion of cold (2 – 3°C) Celsior solution with and without 10 nM Hi1a into the coronary circulation. After cardioplegic arrest, hearts were stored under hypothermic conditions for 8 h, followed by reperfusion and functional analysis (**Fig. 3a**). Hearts preserved with Hi1a supplementation had significantly improved recovery of aortic flow (AF) (50 ± 11%) and cardiac output (CO) (57 ± 8%) compared to Celsior only controls (AF: 8 ± 5%; CO: 24 ± 5%) (**Fig. 3b,c**). These results demonstrate that Hi1a is highly cardioprotective against IRI in a model of donor heart preservation with prolonged cold storage.

Hi1a supplementation improves recovery in a postconditioning model of donation after circulatory death. To determine whether pharmacological inhibition of ASIC1a could recover hearts when delivered after a profound ischemic injury, we assessed the therapeutic effect of Hi1a in a rodent model of donation after circulatory death (DCD). Historically, DCD hearts were considered unusable for transplantation due to the severe ischemic injury sustained during organ procurement[33], but recent investigations and clinical case studies have shown that successful transplantation of DCD hearts is possible with effective post-conditioning strategies, namely normothermic *ex vivo* perfusion and pharmacological post-conditioning[34-36]. To mimic a clinical DCD withdrawal in rodents, asphyxiation was induced via tracheal ligation, and the hearts were retrieved after a 10-min warm ischaemic time (WIT) followed by 2-min stand-off. The hearts were then flushed with Celsior with or without supplementation, followed by *ex vivo* reperfusion in Langendorff mode for 30 min and then working mode for an additional 30 min (**Fig. 4a**).

The effect of Hi1a (10 nM) was tested as a single supplement and in combination with two clinically used supplements, glyceryl trinitrate (GTN) and erythropoietin (EPO)[36]. As a positive control, we also evaluated zoniporide (Zon) combined with GTN and EPO, which we have previously shown activates ischemic postconditioning pathways[35]. To assess the viability of hearts post DCD withdrawal, functional parameters and cell death (LDH release) were measured throughout working-mode reperfusion. In Celsior alone controls, LDH levels measured in the coronary effluent increased during the 30 min assessment period. Allografts supplemented with Hi1a, Hi1a + GTN/EPO, or GTN/EPO/Zon, however, had significantly reduced levels of LDH after 30 min, with LDH release in both triple supplemented groups comparable to a heart at baseline, prior to ischemic insult (**Fig. 4b**). Functional parameters of recovery including AF, CO, and pulse pressure (PP) were all significantly improved in supplemented groups compared to Celsior alone (**Fig. 4c-e**). AF, in particular, was significantly increased in hearts supplemented with Hi1a combined with GTN and EPO (32 ± 2 mL/min) compared to Hi1a as a single supplement (18 ± 3 mL/min), suggesting an additive effect of Hi1a with current clinically used cardioprotective agents (**Fig. 4c**).

To investigate the therapeutic benefit of Hi1a in the context of preventing cell death in DCD hearts, we performed western blots to examine the levels of phosphorylated RIP3 (pRIP3) in tissue lysates

collected at the end of working-mode reperfusion. RIP3 is one of the key regulators of the necroptosis pathway that has been implicated in the response to IRI[37,38], and specifically to ASIC1a-mediated cell death during cerebral ischemia[39]. Treatment with Hi1a as a single or triple supplement led to reduced levels of pRIP3 following DCD withdrawal and reperfusion (**Fig. 4f**), suggesting Hi1a blockade of ASIC1a might inhibit necroptosis. Together with our finding that Hi1a improved donor organ viability after prolonged cold storage, the therapeutic efficacy of Hi1a in the DCD model demonstrates significant translational potential for ASIC1a-targeted therapies during clinical donor heart preservation.

ASICs are expressed in the human heart and polymorphisms in *ACCN1* are associated with ischemic disease. Analysis of ASIC expression patterns from mRNA-seq and ribo-seq data collected from human left ventricular cardiac tissue[40] revealed the highest expression of *ASIC1* and *ASIC3* in the human left ventricle and similar levels of *ASIC1* expression in samples from HF and non-HF patients (**Supplementary Fig. 4a,b**).

We next assessed whether natural variation in the genetic locus encoding ASIC1a (*ACCN1*) is associated with ischemic diseases using human population statistical genetics. While fewer than 10% of new molecular entities pass through clinical trials and are approved for therapeutic use, evidence of human genetic association between the gene target and traits sufficiently similar to the indication being treated increase the likelihood of approval to as high as 50%[41]. We utilized summary data from genome-wide association studies (GWAS)[42] and calculated the statistical significance (using fastBAT[43]) of genetic variants encompassing the *ACCN1* locus with human cardiac and cerebral ischemic phenotypes. These data revealed a significant association between genetic variation in *ACCN1* and major coronary heart disease and MI ($P < 0.05$) (**Table 2**). We also found that genetic variation in *ACCN1* associates with small-vessel ischemic stroke ($p = 3.94 \times 10^{-3}$), but not other stroke subtypes (**Table 2**). Taken together, these data provide evidence that *ACCN1* polymorphisms within the human population are associated with ischemic diseases of the heart and brain.

ASIC1a inhibition prevents cell death in *in vitro* human models of IRI. To examine whether pharmacological blockade of ASIC1a might provide therapeutic benefit in the context of human tissue, we used human induced pluripotent stem cell derived cardiomyocytes (hiPSC-CMs). Contractile cardiomyocytes were generated from stem cells using a standard monolayer-based differentiation protocol involving bi-phasic Wnt regulation (**Fig. 5a** and **Supplementary Fig. 3a**). Analysis of mRNA transcripts from single-cell RNA sequencing data over a time course of cardiac differentiation from pluripotency[44] revealed that *ASIC1*, *ASIC3*, and *ASIC4* expression increased starting in day-5 cardiac progenitor populations and peaked in day-15 definitive cardiomyocytes.

ASIC1 was the most highly expressed ASIC subtype, with expression levels comparable to other cardiac ion channels such as $Na_V1.5$ and $Ca_V1.2$ (**Fig. 5b,c**).

Since ASIC1a mediates influx of calcium in addition to sodium[45], we performed calcium imaging after acute addition of the ASIC1a inhibitors Hi1a and PcTx1 to assess whether ASIC1a inhibition alters physiological electro-mechanical coupling in cardiomyocytes. Replated hiPSC-CMs were loaded with a fluorescent calcium dye and calcium transients were recorded before and after peptide addition. Neither Hi1a nor PcTx1 altered calcium amplitudes or spontaneous beating rate at any evaluated concentration (**Fig. 5d**).

To assess whether ASIC1a inhibition is cardioprotective in hiPSC-CMs, we evaluated cell death in response to *in vitro* acidosis, induced by culturing hiPSC-CMs in Hank's buffered sodium salt (HBSS) with pH adjusted to pH 7.4, 6.0, or 5.0. ASIC1 mRNA expression was not significantly altered by low pH treatment, but significant cell death (> 40%), as assessed by LDH secretion, was observed in populations treated overnight at pH 5.0, with minimal cell death occuring at pH 7.4 or pH 6.0 (**Supplementary Fig. 3b,c**). Treatment with either Hi1a or PcTx1 resulted in nearly complete cardioprotection, even at concentrations as low as 1 nM (**Fig. 5e** and **Supplementary Fig. 3d,e**). To further confirm that ASIC1a plays a direct role in mediating cell death, we treated hiPSC-CMs overnight with 20 nM MitTx, a potent agonist of ASIC1a isolated from snake venom[46]. Consistent with ASIC1a mediating the injury response to cardiac ischemia, treatment of hiPSC-CMs with MitTx resulted in increased cell death at both pH 7.4 and pH 6.0 (**Fig. 5f).** We next evaluated the cardioprotective effect of Hi1a and PcTx1 in an *in vitro* model of ischemia/acidosis with reperfusion. To mimic ischemia/acidosis *in vitro*, hiPSC-CMs were incubated overnight in combined hypoxic (0.5% O_2) and acidic (pH 5.0) conditions with or without peptide. After 18 h of incubation, the low-pH medium was replaced with medium at physiological pH 7.4 with or without peptide, and the cells were incubated for an additional hour under normoxic conditions. Significant cell death was observed in control hiPSC-CMs, but this was blocked by either 10 nM Hi1a or PcTx1 (**Fig. 5g,h**). These data provide evidence that ASIC1a mediates cell death responses in human cardiomyocytes and that pharmacological inhibition of ASIC1a confers significant protection against ischemia-induced cell stress.

Discussion

Despite decades of research and promising preclinical data, no pharmacological strategies have led to robust improvement in clinical outcomes for ischemic injuries of the heart[10]. In this study, we showed for the first time that ASIC1a is a key mediator of the cardiomyocyte injury response to ischemia. Our findings are consistent with prior research demonstrating the importance of ASIC1a

during ischemic stroke[29-31], and they extend the potential therapeutic applications of ASIC1a-targeted therapies to myocardial IRI.

Although ASICs, and in particular ASIC1a, are generally associated with the central and peripheral nervous systems[47], our expression analyses revealed that ASIC1 is present in both rodent and human cardiomyocytes, and is not differentially regulated at the transcriptional level as a result of cellular ischemia *in vitro* or *in vivo*. Our evidence of ASIC1 expression in the heart is supported by recent studies that identified ASIC-like currents in hiPSC-CMs[48] and rat cardiomyocytes[22]. In neurons, there are several possible mechanisms by which ASIC1a activation leads to cell death including calcium overload[49], regulation of mitochondrial permeability transition (MPT) pores[50], and the initiation of necroptosis through direct interaction with, and activation of, RIPK1[39]. ASIC1a-induced cell death has not previously been described in cardiomyocytes, but there is evidence that ASIC1a, along with TRPV1, mediates calcium influx in response to extracellular acidosis[22]. Importantly, given the potential for ASIC1a to transport calcium, we showed that genetic ablation or pharmacological inhibition of ASIC1a does not alter normal electro-mechanical coupling in the heart *in vitro* or *in vivo*.

Given that drug targets with genetic evidence of disease association are twice as likely to succeed in clinical trials and lead to approved drug candidates[41], our GWAS analysis of the association between genetic variation in the *ACCN1* locus and ischemic disease points towards the translational potential of ASIC1a-targeted therapies. Indeed, ASIC1a-targeted therapies have the potential to impact a broad scope of clinical applications. Myocardial IRI is evident in many cardiovascular complications such as acute MI, out-of-hospital cardiac arrest (OOHCA), cardiopulmonary bypass, and heart preservation for transplantation[7]. Despite intensive research into therapies aimed at mitigating myocardial IRI in the context of acute MI, none have translated into clinical practice. Drugs that prevent calcium overload by targeting NHE showed translational promise in preclinical studies[51], but they ultimately failed in Phase III clinical trials (The EXPEDITION Trial) due to cerebrovascular complications[19]. Nonetheless, this trial provided strong clinical evidence that myocardial IRI is a threat during coronary artery bypass graft surgery and that NHE inhibitors can reduce post-operative MI[19].

In general, there is a strong association and co-morbidity between stroke and cardiac arrest, with 88% of patients suffering some form of cardiovascular complication within 4 weeks of an ischemic stroke[52]. Likewise, cardiac interventions such as transcatheter aortic valve implantation (TAVI) can cause cerebrovascular complications[53]. We previously used a rat model of ischemic stroke to show that a single dose of Hi1a (2 ng/kg i.c.v) massively reduced infarct size and improved neurological and motor performance even when administered up to 8 h after stroke onset[31]. No other drugs have been shown to provide significant protection beyond 4 h after stroke onset[54]. Thus, ASIC1a inhibitors

might provide a potential major advantage for clinical use due to their ability to protect both the heart and the brain from IRI, which is particularly relevant for OOHCA, for which there is both cardiac and cerebral manifestations.

Our findings suggest that ASIC1a-targeted therapies have promising potential in heart transplantation applications. The predominate source of donor hearts is from brain dead (BD) donors, although recent advances in clinical practice have allowed for the transplantation of DCD hearts which has increased the donor pool[36]. The number of usable BD and DCD hearts, however, continue to be adversely affected by unavoidable injury to the heart that occurs during organ procurement and storage, as well as reperfusion injury in the recipient. Indeed, the relative risk of one-year morbidity is closely correlated with organ ischemic time[7,55]. In general, primary graft dysfunction is a major risk for patients in the first 30 days after transplantation and accounts for 66% of patient deaths, of which 48% are due to IRI[56]. Thus, there is an unmet need for effective, early interventions that reduce injury to the graft and reduce the risk of primary graft dysfunction in the recipient. Our direct comparison between Hi1a and supplements in current clinical use provide strong evidence for the clinical utility of Hi1a for increasing the pool of viable donor hearts for transplantation.

This study demonstrates that ASIC1a is a critical mediator of cell death during cardiac ischemia. We show that ASIC1a inhibition with Hi1a affords potent cardioprotection across rodent and human models, and results in improved cardiac viability and functional recovery post-IRI. These findings support the utility of ASIC1a-targeted approaches for the treatment of myocardial IRI with a specific demonstration of efficacy in improving the viability of donor organs for heart transplantation. Development of ASIC1a inhibitors for therapeutic applications has the potential to significantly impact the public health burden of cardiovascular disease, and result in improved quality of life and outcomes for patients with cardiac ischemic injuries.

Acknowledgements. We gratefully acknowledge funding support from The University of Queensland (UQ) Strategic Funding Research Initiative (Grant D17-72571 to NJP, GFK, and JFF), the Australian National Heart Foundation (Grant 101889 to NJP), the Whitaker International Program (Scholar Grant to MAR), the American Australian Association (Grant 441 to MAR), the St Vincent's Clinic Foundation (to PSM), the Australian National Health & Medical Research Council (Program Grant 1074386 to PSM), the Australian Research Council (Discovery Early Career Researcher Award DE180100976 to GCP), and the Australian National Health & Medical Research Council (Principal Research Fellowship APP1136889 and Development Grant APP1158521 to GFK). Thanks to the Integrated Physiology Facility, Melanie Flint, and Mubarak Alzubaidi at UQ for technical support and assistance. Microscopy was performed at the Australian Cancer Research

Foundation (ACRF)/Institute for Molecular Bioscience Cancer Biology Imaging Facility, which was established with the support from ACRF.

Author Contributions

M.A.R. designed, performed, and analysed the experiments presented in **Figs 1**, **2**, and **5**, **Table 1**, and **Supplementary Figs 1 – 3**, and wrote the manuscript. **S.E.S.** designed, performed, and analysed the experiments described in **Figs 3** and **4** and wrote the manuscript. **N.J.S.** produced, purified, and performed QC analyses for peptides used in **Figs 2 – 5 and Supplementary Figs 2** and **3**. **L.G. and J.E.V.** helped design and perform the experiments described in **Figs 3** and **4**. **M.K.** helped design and helped with interpretation of the experiments described in **Figs 3** and **4**. **M.E.R., J.N.P., and L.E.S.H.** conceived ideas and provided valuable insights towards interpretation of results shown in **Figs 1** and **2**. **H.S.C. and X.C.** helped perform the experiments described in **Fig. 5**. **V.S**. performed the computational analysis presented in **Supplementary Fig. 4**. **J.Y.S**. provided valuable intellectual support. **G.C.P** performed GWAS analysis and interpretation presented in **Table 2**. **J.F.F** helped conceive the heart transplant studies and provided intellectual and financial support. **S.M. and S.P** generated the ASIC1a knockout mouse strain used for studies presented in **Fig. 1**. **G.F.K.** conceived of experimental objectives, provided intellectual and financial support, and directly supervised work related to peptide production utilized across all aspects of the study. **P.S.M** conceived of experimental objectives, provided intellectual and financial support, and directly supervised work related to heart transplant modelling. **N.J.P.** conceived of experimental objectives, provided intellectual and financial support, directly supervised work related to mouse IRI, GWAS analysis, human pluripotent stem cell modelling, and oversaw programmatic decisions regarding study design and completion in close collaboration with P.S.M and G.F.K. All authors contributed to revisions and final preparations of the manuscript.

Table 1. Baseline functional parameters for Langendorff-perfused hearts from ASIC1a$^{+/+}$ and ASIC1a$^{-/-}$ mice

Genotype	Heart Rate, bpm	LVDP, mmHg	EDP, mmHg	+dP/dt (mmHg/s)	-dP/dt (mmHg/s)	CF (mL/min/g)
Wildtype (ASIC1a$^{+/+}$)	353 ± 23	105 ± 4	6.8 ± 0.7	3777 ± 139	-2592 ± 144	35 ± 5
KO (ASIC1a$^{-/-}$)	339 ± 17	101 ± 4	5.1 ± 0.7	3647 ± 236	-2702 ± 148	26 ± 3
p-value	*0.67*	*0.54*	*0.13*	*0.66*	*0.64*	*0.25*

All pre-ischemia functional parameters were measured after more than 30 min of stabilization, except for heart rate, which was measured after 15 min of stabilization, prior to ventricular pacing. All values are expressed as mean ± SEM.

Table 2. Association of polymorphisms in *ACCN1* with ischemic conditions. *ACCN1* gene position: chromosome 12, start: 50451419, end: 50477405)

Genotype	No. of SNPs	SNP start	SNP end	χ^2 statistic	Mean p-value
Acute myocardial infarction [1]	75	rs11377593	rs2242507	152.445	1.36E-02
Major coronary heart disease excluding revascularizations [1]	75	rs11377593	rs2242507	199.114	1.79E-03
Myocardial infarction [1]	75	rs11377593	rs2242507	186.588	3.07E-03
Any stroke [2]	61	rs78972052	rs2242507	81.8082	1.45E-01
Any ischemic stroke [2]	62	rs78972052	rs2242507	77.2837	1.98E-01
Large artery stroke [2]	63	rs78972052	rs2242507	67.2951	3.48E-01
Cardioembolic stroke [2]	61	rs78972052	rs2242507	71.6309	2.52E-01
Small vessel stroke [2]	64	rs78972052	rs2242507	147.484	3.94E-03

The number of SNPs inside the *ACCN1* locus was used to calculate the χ^2 statistic and p-value. Source data: GWAS results from [1] UK Biobank from Neale Lab[42] or [2] Malik et al.[57].

Figures

Figure 1. **Genetic knockout of ASIC1a protects mouse hearts from *ex vivo* IRI. (a)** mRNA expression (counts per million, CPM) analysis of sorted cardiac cell populations from P56 adult mouse hearts (data extracted from Ref.[32]). **(b-f)** Langendorff-perfused hearts from ASIC1a KO (ASIC1a$^{-/-}$, n = 10) and WT (ASIC1a$^{+/+}$, n = 5) mice were subjected to 25 min of global ischemia followed by 45 min of reperfusion. **(b)** LVDP, expressed as % of pre-ischemia baseline, over time. **(c-e)** Functional parameters measured or calculated after 45 min of reperfusion. **(c)** LVDP, percent baseline (p = 0.025). **(d)** EDP, p = 0.013. **(e)** CF, percent baseline (p = 0.049). **(f)** Cell death after 2 min of reperfusion (units/mL of LDH normalized to reperfusion flow rate and heart weight, U/min/g, p = 0.006). For LVDP and CF, baseline values were obtained immediately prior to ischemia. All data are expressed as mean ± SEM. Statistical significance was determined using two-tailed unpaired student's *t*-test (*p < 0.05; **p < 0.01).

Figure 2. **ASIC1a inhibitors protect isolated mouse hearts from IRI**. **(a)** Australian funnel-web spider, *Hadronyche infensa*, from which the ASIC1a inhibitor Hi1a was isolated. **(b)** Schematic of the 3D structure of Hi1a (PDB 2N8F[31]) highlighting the two knottin domains. The 3D structure of PcTx1 (green; PDB 2KNI[58]) is overlaid on the N-terminal knottin domain of Hi1a (blue). The disulfide bonds in each structure are shown as maroon tubes. **(c)** Sequence alignment of Hi1a, PcTx1, and the PcTx1-R27A/V32A analogue. Conserved residues are shown in green, except cysteine residues which are highlighted in maroon. Black circles indicate pharmacophore residues of PcTx1. **(d-h)** Langendorff-perfused hearts from adult (12–14 weeks old) male C57BL/6 mice were subjected to 25 min of global ischemia followed by 45 min of reperfusion. Control hearts (no treatment, n = 21) were compared to hearts treated with 10 nM Hi1a (n = 7), 10 nM PcTx1 (n = 5), 10 nM PcTx1-R27A/V32A, n = 11), or 0.1% BSA in water (vehicle control, n = 13). For treated hearts, vehicle or peptide solution was infused for 10 min prior to ischemia and during the first 15 min of reperfusion. **(d)** Pre-ischemia LVDP, expressed as % baseline, after exposure to vehicle or peptide for 10 min. **(e)** LVDP over time. **(f-h)** Functional parameters measured or calculated after 45 min of reperfusion including **(f)** LVDP **(g)** EDP and **(h)** CF. For LVDP and CF, baseline values were obtained prior to peptide/vehicle infusion, or 10 min prior to onset of ischemia (no-infusion controls). All data are expressed as mean ± SEM. Statistical significance was determined using one-way ANOVA (*p < 0.05; **p < 0.01, ***p < 0.001).

Figure 3. **Hi1a protects isolated rat hearts during long-term hypothermic storage**. **(a)** Experimental schematic of 8 h cold storage protocol. **(b-c)** Functional parameters assessed throughout 30 min working mode (post-storage reperfusion) and normalized to pre-storage function. **(b)** Aortic flow (% baseline) over time (left) and after 30 min (right). **(c)** Cardiac output (% baseline) over time (left) and after 30 min (right). All data are expressed as mean ± SEM. Statistical significance was determined using two-tailed unpaired student's t-test (*p < 0.05; **p < 0.01).

Figure 4. **Hi1a protects rat hearts in a preclinical model of DCD**. **(a)** Experimental design to mimic clinical DCD withdrawal. **(b)** Arbitrary LDH units in the coronary effluent throughout 30 min working mode (normalized to coronary flow rate, LDH a.u./mL/min). **(c-e)** Assessment of heart function throughout working mode with **(c)** AF (mL/min), **(d)** CO (mL/min), and **(e)** PP (mmHg) plotted over time (left) and after 30 min in working mode (right). **(f)** Representative pRIP3 western blot on pRIP3-IP lysates collected at the end of reperfusion and quantification of relative signal intensity of upper band (~46 kDa). Individual lanes represent different hearts (n = 3 hearts/group/membrane). All data are expressed as mean ± SEM. For all groups, n = 6 hearts. Statistical significance was determined using one-way ANOVA (*p < 0.05; **p < 0.01, ***p < 0.001).

Figure 5. **Hi1a protects hiPSC-CMs from ischemic injury**. **(a)** Schematic depicting directed differentiation of hiPSC-CMs (adapted from Ref.[44]). **(b-c)** Analysis of published transcriptomic data (scRNAseq) of cardiac differentiation[44]. **(b)** Expression of ASIC1, ASIC2, ASIC3, and ASIC4 at day 0, 2, 5, 15, and 30 of differentiation. **(c)** Gene expression in day-15 hiPSC-CMs. **(d)** Fluorescent imaging of calcium transients (normalized arbitrary fluorescent units (F/F_0)) before and after Hi1a or PcTx1 addition using a FLIPR Tetra system. Representative traces (black dotted line indicates time of peptide addition) and quantification of spontaneous beat rate and calcium amplitude are shown. Amplitude and beat rate are both expressed as a response over baseline (post-addition measurement normalized to pre-addition measurement). **(e)** Cell death (LDH secretion) analysis after overnight treatment in HBSS pH 5.0 with or without Hi1a (top) or PcTx1 (bottom). **(f)** Cell death (LDH) after overnight treatment with 20 nM MitTx in HBSS pH 7.4 or HBSS pH 6.0. **(g-h)** Cell death (LDH) after *in vitro* IRI with overnight hypoxic (0.5% O_2) incubation in HBSS pH 5.0 followed by 1 h reperfusion with HBSS pH 7.4 in normoxic conditions. For panels e – h, data are expressed as percent cell death calculated from LDH levels in low (RPMI + B27) and high (1% Triton X-100 in RPMI + B27) controls. Statistical significance was determined using one-way ANOVA (*p < 0.05; **p < 0.01, ***p < 0.001).

Online Methods

Animals. All animals used in this study received humane care in compliance with National Health and Medical Research Council (Australia) guidelines and the *Guide for the Care and Use of Laboratory Animals* (National Institute of Health, Bethesda, MD). All experiments were performed in accordance with protocols approved by the Animal Ethics Committee of The University of Queensland (IMB/171/18) and Garvan Institute of Medical Research (16/38). To test the therapeutic efficacy of pharmacological inhibitors of ASIC1a, young male C57BL/6 mice (12 – 14 weeks) or male Wistar rats (325 – 395 g) were purchased from the Animal Resource Centre (Canning Vale, Western Australia). For genetic ablation studies, ASIC1a-/- (KO) and ASIC1a+/+ (WT) mice (12 – 14 weeks) were used (see below for generation and validation). Methods for ASIC1a KO mouse will be provided in a subsequent updated version.

IRI in Langendorff-perfused mouse hearts. Isolated hearts were assessed for tolerance to IRI as previously described[59,60]. Mice were anesthetized via an intraperitoneal injection of 10 mg/mL ketamine and 1.6 mg/mL xylazil. The heart was excised via thoracotomy and the aorta cannulated. The hearts were retrograde perfused under constant hydrostatic pressure (80 mmHg) with oxygenated (95% O_2; 5% CO_2) modified Krebs-Henseleit buffer (composition in mM: 119 NaCl, 22 $NaHCO_3$, 4.7 KCL, 1.2 $MgCl_2$, 1.2 KH_2PO_4, 0.5 EDTA, 1.85 $CaCl_2$, 11 D-(+)-glucose, 2 Na^+ pyruvate, all from Sigma-Aldrich). Temperature was continuously measured with thermocouple needle microprobes (SDR Scientific) and maintained at 37°C with circulating water baths. Contractile function was measured via a fluid-filled balloon inserted in the left ventricle and connected to a pressure transducer (ADInstruments Pty Ltd.). Coronary flow was measured with an in-line Doppler flow probe positioned above the aortic cannulae (Transonic Systems Inc.). All functional data were recorded on a four-channel MacLab system (ADInstruments Pty Ltd.). Following 15 – 30 min of equilibration, hearts were switched to ventricular pacing at 420 beats/min (BPM) using a SD9 stimulator (Grass Instruments, Inc.). Baseline measurements were made for 10 – 20 min followed by 25 min of global normothermic ischemia and 45 min of reperfusion. Pacing was stopped during ischemia and resumed after 15 min of reperfusion.

For peptide-treated experimental groups, 1 μM peptide solution was infused with a syringe pump (World Precision Instruments, LLC.) into the buffer, directly upstream of the cannula, at a rate of 1% of CF, for a final working concentration of 10 nM. Peptide was infused for 10 min prior to the onset of ischemia and during the first 15 min of reperfusion. To assess cell death, effluent was collected at 2 and 45 min after the start of reperfusion. Effluent levels of LDH were measured using a cytotoxicity detection kit (Roche). Normalized absorbance values (absorbance at 492 nm minus absorbance at 690 nm) were measured using a PHERAStar FS microplate reader (BMG Labtech). Standard curves were generated with bovine LDH (Sigma-Aldrich) and used to convert sample

absorbance values to units of LDH. Data were normalized to CF and heart weight and then expressed as U/min/g. For all analyses, hearts were excluded if they met the following criteria: abnormally high coronary flow (> 8 mL/min), delayed onset of ischemic contracture (time to onset of ischemic contracture (TOIC) > 20 min), poor contractile function after equilibration (significant arrhythmias and/or left ventricular systolic pressure < 80 mmHg), or technical issues with the perfusion rig.

Isolated working model of donor heart preservation after prolonged cold storage. We used an isolated working heart model of donor heart preservation that we previously described [61-63]. Rodents were anesthetized with an intraperitoneal injection of ketamine (80 mg/kg; Cenvet, Australia) and xylazine (10 mg/kg; Provet, Australia), with supplemental doses given as necessary until adequate anaesthesia was achieved. An incision was then made in the abdomen of the animal and a bolus injection of 300 IU of heparin administered into the renal vein. A sternotomy was subsequently performed, then the heart was rapidly excised and arrested by immersion in ice-cold Krebs-Henseleit buffer (composition in mM: 118 NaCl, 4.7 KCl, 1.2 $MgSO_4$, 1.2 KH_2PO_4, 25 $NaHCO_3$, 1.4 $CaCl_2$, 11 glucose, all from Sigma-Aldrich). Following aortic cannulation, hearts were stabilised for 10 min by retrograde (Langendorff) perfusion with Krebs-Henseleit buffer (37°C), at a hydrostatic pressure of 100 cm H_2O. During this period, the left atrial appendage was cannulated. This nonworking Langendorff system was then converted to a working preparation by switching the supply of perfusate from the aorta to the left atrial cannula at a hydrostatic pressure of 15 cm H_2O (preload). The working heart ejected perfusate into the aortic cannula against the maintained fixed hydrostatic pressure of 100 cm H_2O (afterload). Aortic pressure was monitored via a side arm of the aortic cannula with a pressure transducer (Ohmeda, Pty Ltd., Singapore). Aortic flow (AF) was measured by an in-line flowmeter (Transonics Instruments Inc., Ithaca, NY). Aortic pressure and flow were recorded using PowerLab/4e (ADInstruments Pty Ltd, Sydney, Australia) with heart rate (HR) calculated from the flow trace. Coronary flow (CF) was measured by timed collection of effluent draining from the heart via a small incision in the pulmonary artery (PA). Cardiac output (CO) was calculated from the sum of AF and CF.

Figure 3a illustrates the experimental timeline. The heart was maintained in working mode for 15 min, at which time functional parameters (AF, CF, CO and HR) were recorded and defined as the baseline value. Any hearts having a baseline AF < 35 mL/min, HR < 200 bpm, or CF < 10 mL/min were excluded at this stage. The heart was then arrested by infusion of a preservation flush containing Celsior solution (2 – 3°C) ± 10 nM Hi1a via the aortic cannula (60 cm H_2O infusion pressure over 3 min), immersed in 100 mL Celsior ± 10 nM Hi1a, and kept on ice for 8 h. The heart was then remounted on the perfusion apparatus and perfused with fresh Krebs-Henseleit buffer for 15 min in Langendorff mode, followed by 30 min in working mode. Celsior solution was obtained

from the Institut Georges Lopez (Lissieu, France). Cardiac functional indices after 30 min of working-mode reperfusion were expressed as percent recovery of baseline values.

Donor heart preservation following circulatory death in rodents. The experimental model of donation after circulatory death (DCD) in rodents (**Fig. 4a**) was developed by our laboratory to closely mimic the events during withdrawal of life support (WLS) in the clinical scenario. Rodents were anesthetized with an intraperitoneal injection of ketamine (80 mg/kg; Cenvet, Australia) and xylazine (10 mg/kg; Provet, Australia), with supplemental doses given as necessary until adequate anaesthesia was achieved. Non-invasive monitoring was then established, consisting of measurement of pulse oximetry (Rad-5v, Masimo Inc., Irvine, CA) and continuous electrocardiogram (P55 AC pre-amplifier, Grass Instrument Co., RI). A midline incision was then made in the neck of the animal to expose the right carotid artery, which was cannulated for invasive monitoring of arterial pressure via a pressure transducer (Ohmeda, Pty Ltd., Singapore). Once baseline monitoring parameters were recorded, a dose of 250 IU for antemortem heparin was delivered via a side port in the arterial pressure monitoring line. The animal was then subjected to WLS via asphyxiation. A fixed warm ischemic time of 10 min followed by a 2-min stand-off period (to mimic the regulated standoff period in a clinical setting) was chosen. Upon completion of the stand-off period, the heart and lungs were swiftly excised *en bloc* and immersed in ice-cold Krebs-Henseleit buffer. The aorta and left atrium were then fashioned for instrumentation on an *ex-vivo* Langendorff perfusion apparatus. Once the aorta was cannulated onto the *ex-vivo* circuit, the pulmonary veins were rapidly ligated, and the preservation flush (containing Celsior solution (2 – 3°C) ± supplements) was commenced via the aortic cannula at a flow rate of 15 – 20 mL/min up to a total volume of 100 mL. The following concentrations of supplements were used: Hi1a: 10 nM; GTN: 100 mg/L; EPO: 5U/mL; zoniporide: 1 μM. Left atriotomy was performed to prevent left ventricle distension and to facilitate cannulation on the *ex-vivo* apparatus for functional assessment during the working phase of the experiment.

Following the preservation flush, Langendorff reperfusion was commenced for a period of 30 min, using oxygenated Krebs-Henseleit buffer (37°C) to allow stabilization and recovery of the heart. As previously described, the apparatus was then converted to a working mode via switching the supply of perfusate from the aortic to the left atrial cannula at a hydrostatic pressure of 15 cm H_2O (preload). Functional parameters (AF, CO, PP and HR) were measured continuously, and CF measured at 5-min intervals, during working mode. Coronary effluent and left ventricular free-wall samples were collected as per the protocol for the isolated working heart model.

To assess cell death throughout working mode, effluent LDH levels were measured as an index of structural cardiomyocyte injury using the TOX 7 assay kit (Sigma-Aldrich). Duplicate 75 μl aliquots

were assayed according to the manufacturer's instructions. The resulting absorbance of the tetrazolium product was measured at 492 nm using a 96-well plate reader and expressed as arbitrary units, normalised to coronary flow per minute.

Peptide production. Recombinant Hi1a was produced by expression as a His_6-maltose-binding protein (MBP) fusion protein in the periplasm of *Escherichia coli* as previously described[31], but with optimisation of the expression and purification conditions to improve the yield of Hi1a. Briefly, *E. coli* BL21(DE3) cells were grown at 30°C in Terrific Broth until the optical density at 600 nm reached 2.0 − 2.5, at which point the temperature was reduced to 17°C and after a 15 min delay expression of the Hi1a fusion protein was induced with 1 mM IPTG. Cells were harvested after a further 21 h growth. PcTx1 and the PcTx1-R27A/V32A analogue were produced in the same manner with minor modifications to the expression protocol. *E. coli* were grown at 37°C for the entire expression period and harvested approximately 4 − 5 h after induction.

Cell pellets were resuspended in 50 mM Tris, 300 mM NaCl, 5% glycerol, 15 mM imidazole at pH 8.3 for Hi1a or pH 8.0 for PcTx1, then the cells were lysed by high-pressure cell disruption (Constant Systems Limited, United Kingdom). The His_6-MBP-peptide fusion proteins were purified from the clarified soluble lysate over a nickel affinity resin, then the resin was washed with the same buffer to elute weakly bound proteins before eluting the Hi1a fusion protein with the same buffer containing 300 mM imidazole. Fusion proteins were then exchanged into low (< 30 mM) imidazole buffer using an Amicon Ultra-15 centrifugal concentrators (Merck Millipore, Germany) with a 30 kDa molecular weight cut-off in preparation for liberation of Hi1a from the fusion tag using tobacco etch virus (TEV) protease. The fusion proteins were cleaved in redox buffer containing 3 mM reduced glutathione and 0.3 mM oxidised glutathione at pH 8.3 for Hi1a or pH 8.0 for PcTx1, using ~1 mg of TEV protease per 50 mg of fusion protein. For Hi1a the cleavage reaction was allowed to proceed at 4°C over 3 − 6 days. For the PcTx1 peptides, cleavage was performed at room temperature for a minimum of 16 h. The recombinant peptides each contain a non-native N-terminal serine residue, which is a vestige of the TEV protease recognition site. The released peptides were purified from the cleavage reaction solutions using reverse phase high-performance liquid chromatography.

Masses of the purified peptides were confirmed by electrospray-ionisation mass spectrometry then the pure peptides were lyophilised prior to confirmation of ASIC1a inhibitory activity using two electrode voltage clamp electrophysiology, as described previously for both Hi1a[31] and PcTx1[29,58,64]. Unless otherwise noted, lyophilized stocks of peptide were reconstituted in sterile deionised water prior to use.

Western blot. Left ventricular tissue samples (~60 mg) from hearts (N = 6/group) collected at the end of the DCD protocol were homogenized in ice-cold lysis buffer (150 mM NaCl, 50 mM Tris-HCl,

1% Triton X-100, 1 mM sodium orthovanadate, 1 mM glycerophosphate, 5 mM dithiothreitol, 15% Roche cocktail protease inhibitors; pH 7.4). Samples were then centrifuged at 10,000 rpm (9,400 *g*) for 5 min at 4°C. The protein concentration of each lysate was measured using a Bradford assay kit (Pierce Biotechnology, Rockford, IL), with bovine serum albumin used as standard. If required, immunoprecipitation (IP) was performed (see below). Protein samples (40 μg) were heated at 95°C in Laemmli sample loading buffer for 10 min and electrophoretically separated on 4 – 20% graduated precast gels (Bio-Rad) and transferred to polyvinylidene difluoride membranes. After blocking with 5% non-fat dry milk in Tris-buffered saline (TBS) containing 0.1% Tween-20, membranes were incubated overnight at 4°C with primary antibody (pRIP3 Ser227, Cell Signalling Technology, USA). Membranes were incubated with secondary anti-rabbit (1:2000 in 5% non-fat dry milk + TBS-T) or anti-mouse (1:5000 in 1% BSA + TBS-T) IgG conjugated to horseradish peroxidase (GE healthcare, Ryadalmere, Australia) for 1 h at 25°C. Protein bands were visualized using SuperSignal West Pico Chemiluminescent Substrate (Life Technologies, Scoresby, VIC, Australia), digitized, then quantified using Image J software (Version 1.52a, National Institute of Health, USA). Within each blot, the signal intensity from each lane was normalized to the Celsior only group average from the same membrane. Blots were repeated to confirm results. All antibodies, unless otherwise stated, were sourced from Cell Signalling Technology (Beverly, MA).

Immunoprecipitation. Lysate samples from individual hearts prepared as detailed above were centrifuged for 10 min at 10,000 *g* (4°C). Lysates were then diluted with lysis buffer without dithiothreitol to make a final volume of 0.5 mL containing 1 mg of total proteins (1 μg/μl). Pre-clearing of the samples were then performed by adding 60 μl of 50% Protein A agarose beads to 1 mL of diluted lysate from above. Samples were then tumbled for 30 min at 4°C in a tube rotator, prior to being centrifuged for 5 min at 1,000 *g* (4°C). Supernatants were then collected for immunoprecipitation. The cleared lysates were then divided into two 400 μl aliquots and 1:100 specific primary antibody added to form the Ag/Ab complex (pRIP3 Ser227, Cell Signalling Technology, USA). 50 μl of 50% Protein A beads were then added followed by gentle rocking for 2 h at 4°C. Following centrifugation for 30 s at 10,000 *g*, 4°C, the supernatant (containing unbound proteins) was aspirated, and either discarded or snap frozen at –80°C for sequential immunoprecipitation of other antigens. The remaining pellet was then washed five times with 500 μl of ice-cold lysis buffer. For each wash, the beads were re-suspended by inversion and then centrifuged for 30 s at 1000 *g* at 4°C, prior to the supernatant being aspirated and discarded. The final pellet was then resuspended with 50 μl of 2 x SDS sample buffer without β-mercaptoethanol. The samples were mixed vigorously and then heat denatured for 5 min at 95°C. Following a 30 s centrifugation at 10,000 *g*, the supernatants were transferred to new vials and β-mercaptoethanol

was added. The prepared samples were then loaded on 4 – 20% graduated precast gels (Bio-Rad) and analysed by Western blotting as detailed above.

Generation of cardiomyocytes from human induced pluripotent stem cells. All hiPSC studies were performed with consent from the University of Queensland's Institutional Human Research Ethics approval (HREC#: 2015001434). Cardiomyocytes generated in this study were derived from the WTC-11 hiPSC line (Gladstone Institute of Cardiovascular Disease, UCSF)[65,66]. Undifferentiated hiPSCs were maintained on Vitronectin XF (5 µg/mL, Stem Cell Technologies) coated tissue culture dishes as per manufacturer recommendation with either mTeSR medium with supplementation or mTeSR PLUS medium with supplementation (Stem Cell Technologies). Contractile cardiomyocytes were differentiated using a high-density monoloyer format as previously described[44]. On day -1 of differentiation, hiPSCs were dissociated with 0.5 mM EDTA solution supplemented with 1.1 mM D-glucose. Single-cell suspensions were plated at a density of 1.2 x 10^5 cells/cm^2 and cultured overnight in mTeSR medium supplemented with 10 µM Y-27632 dihydrochloride (Stem Cell Technologies). Once the monolayer reached approximately 80% confluence (usually the following day), differentiation was induced (day 0). The cells were quickly washed with phosphate-buffered saline (PBS) followed by a change in medium to RPMI (ThermoFisher) containing 3 µM CHIR99021 (Stem Cell Technologies), 500 µg/mL BSA (Sigma Aldrich), and 213 µg/mL ascorbic acid (Sigma Aldrich). After 3 days of culture, the medium was exchanged to RPMI containing 500 µg/mL BSA, 213 µg/mL ascorbic acid, and 5 µM Xav-939 (Stem Cell Technologies). On day 5, the medium was replaced with RPMI containing BSA and ascorbic acid as on day 3. Starting on day 7, the cells were fed every other day with RPMI containing 1x B27 supplement with insulin (Life Technologies). Spontaneous beating was typically observed between days 9 and 11 of differentiation.

In vitro *ischemia-acidosis injury model with hiPSC-CMs*. Differentiated cardiomyocytes were replated on either day 15 or day 17 of differentiation for *in vitro* ischemia/acidosis assays. At the time of replating, a subset of cells (~500,000) was set aside for flow cytometry analysis of cardiomyocyte purity (see below). For all experiments, only cell preparations with greater than 80% sarcomeric α-actinin-positive cardiomyocytes were used. After re-plating, the cells were maintained for an additional 7 days in RPMI + B27. To prepare media for ischemia/acidosis injury, 10x HBSS without sodium bicarbonate (Sigma) was diluted to 1x concentration in sterile tissue culture-grade water. Solutions were buffered with either 12 mM HEPES (for pH 7.4 media, Sigma Aldrich) or 12 mM MES (for pH < 6.5, Sigma Aldrich) and the pH adjusted accordingly with 1 M NaOH. The medium was sterile filtered with 0.22 µm syringe filters (Millipore). Unless otherwise noted, the re-plated cells were treated overnight (18 h) in HBSS with or without peptide under either normoxic (~18.5% O_2; 5% CO_2) or hypoxic (0.5% O_2; 5% CO_2) culture conditions. For reperfusion experiments, the medium

was replaced with HBSS pH 7.4 (with or without peptide) after overnight incubation and cultured for 1 h under in normoxic conditions.

To assess cell death, supernatant was collected and LDH levels were measured using a cytotoxicity detection kit (Roche). For all cell culture experiments, percent cell death was calculated using low and high controls. For low control (LC), cardiomyocytes were cultured overnight in standard culture media (RPMI + B27). For high control (HC), cells were cultured in RPMI + B27 containing 1% Triton X-100 (Sigma-Aldrich).

Flow cytometry. To assess the cardiomyocyte purity of differentiated cell populations, cells were fixed with 4% paraformaldehyde (Sigma Aldrich), permeabilized in 0.75% saponin (Sigma Aldrich), and labelled with Phycoerythrin (PE)-conjugated sarcomeric α-actinin (SA) antibody (Miltenyi Biotec Australia Pty) or PE-conjugated mouse isotype (IgG) control (Miltenyi Biotec Australia Pty). Stained samples were analysed on a FACS CANTO II (Becton Dickinson) machine with FACSDiva software (BD Biosciences). Data were analysed was using FlowJo software and cardiac populations were determined with population gating from isotype controls.

Immunohistochemisty/TUNEL stain. After overnight treatment at low pH, replated hiPSC-CMs were fixed in 4% paraformaldehyde for 10 min, washed in PBS, and incubated for 1 h at room temperature in blocking solution (PBS containing 2% heat-inactivated sheep serum (HISS) and 0.05% Triton X-100). Cells were then incubated overnight at 4°C with primary antibody followed by incubation with secondary antibody for 1 h at room temperature. Nuclei were counterstained with 1 µg/mL DAPI. The samples were washed with PBS and TUNEL stain (Abcam) was performed as per manufacturer protocol. Stained samples were imaged within one week. All antibody dilutions were prepared in blocking solution. The following antibodies were used: mouse anti-α-actinin (Clone EA-53, Sigma Aldrich, 1:100) and donkey anti-mouse AlexaFluor 647 (ThermoFisher, 1:100). High-resolution images were obtained using a Zeiss LSM710 AiryScan confocal microscope with 20x 0.8 NA or 40x 1.3 NA Plan Apochromat objectives running Zeiss Zen Black. All image processing and quantitation was performed in FIJI [67].

Quantitative real-time PCR. To assess mRNA transcript levels in hiPSC-CM populations, total RNA was extracted using a RNeasy Mini Kit (QIAGEN). Superscript III First Strand Synthesis (ThermoFisher) was used to generate cDNA and quantitative real-time (qRT) PCR was performed on ViiA 7 Real-Time PCR Machine (Applied Biosystems) with SYBR Green PCR Master Mix (ThermoFisher). Transcript copy numbers were calculated using the double delta ct method relative to the housekeeping gene *HPRT1*. Primers used for qRT-PCR analysis were as follows:

Gene name	Forward Primer (5’ → 3’)	Reverse Primer (5’ → 3’)

HPRT1	TGACACTGGCAAAACAATGCA	GGTCCTTTTCACCAGCAAGCT
TNNI1	CCCAGCTCCACGAGGACTGAACA	TTTGCGGGAGGCAGTGATCTTGG
TNNI3	GGAACCTCGCCCTGCACCAG	GCGCGGTAGTTGGAGGAGCG
MYH7	GGGCAACAGGAAAGTTGGC	ACGGTGGTCTCTCCTTGGG
MYH6	CAAGTTGGAAGACGAGTGCT	ATGGGCCTCTTGTAGAGCTT
ASIC1	GGATGGAGGTCTACCCTGGA	GACCTCAGCTTCTGCCTGTCA
ASIC2	GCCCCTTTTTGTACCCCTGA	TGTAGCGGGTTAGGTTGCAG
ASIC3	CCTCTGTGAGGTGTTCCGAG	CAGGCTGAGGTGGGGAAC
ASIC4	ACGCTGTTGACATGTCCTGAA	CCTCGATCGTCTCCTTGTGG

hiPSC-CM calcium analysis*.* Cardiomyocytes were replated (2 x 10^4 per well) in a 384 well plate (CellBIND black with clear bottom, Corning) and cultured for 7 days in RPMI + B27. On the day of the experiment, the cells were loaded for 1.5 h at 37°C with FLIPR calcium 4 dye (Molecular Devices) diluted in HBSS pH 7.4. After loading, the plate was transferred to a FLIPR Tetra fluorescent plate reader (Molecular Devices). Calcium transients were measured with excitation wavelengths at 470 – 495 nm and emission at 515 – 575 nm. For each plate, the camera gain and intensity were adjusted to optimize signal intensity. Data for each well was expressed as normalized arbitrary fluorescence units. All data were acquired at 0.5 s per read, with baseline measurements for 45 s followed by at least 100 s of data collection after each peptide addition. To assess peptide effects on calcium handling in normal physiological conditions, concentrated peptide dilutions (in HBSS pH 7.4) were added to each well to give a final concentration of 1 nM, 10 nM, 100 nM, or 1 µM. Calcium amplitude, maximum calcium, minimum calcium, and spontaneous beating rate were analysed using ScreenWorks software (Molecular Devices) and normalized to baseline measurements.

Analysis of ASIC1 from genome wide associated studies of relevant conditions*.* To assess whether genetic variation of *ASIC1* associates with cardiovascular disease and stroke, we performed a gene-based level test on GWAS summary data using fastBAT[43] implemented in the Complex-Traits Genetics Virtual Lab (CTG-VL)[68]. GWAS summary data contains the statistical information of the association of all the genetic variants included in a GWAS against a particular trait. fastBat uses these data to estimate the association of a set of genetic variants within 50 kb from a gene (*ACCN1* in this case) while accounting for the correlation between genetic variants (i.e. linkage disequilibrium). Specifically, we performed analyses using GWAS summary data for acute

MI (N_{cases} =5,948, $N_{controls}$ = 354,176), major coronary heart disease (N_{cases} = 10,157, $N_{controls}$ = 351,037) and MI (N_{cases} = 7,018, $N_{controls}$ = 354,176) from Neale's UK Biobank GWAS database[42] and stroke (including any type of stroke, ischemic stroke, large artery stroke, cardioembolic stroke and small vessel stroke[57] (N_{cases} =40,585, $N_{controls}$ = 406,111).

Computational analysis of human translatome data. Transcriptional (mRNA-seq) and translational (Ribo-seq) raw data of human heart were obtained from Hubner Lab website (http://shiny.mdc-berlin.de/cardiac-translatome/)[40]. Quality control checks for both RNA-seq and Ribo-seq data sets were performed to ensure reasonable quality for downstream analysis. Genes that were expressed in > 99% of samples were retained. After quality control, 47,094 and 43,183 genes were retained in the RNA-seq and Ribo-seq datasets, respectively. DESeq2 normalization was applied to normalize sequencing depth and RNA composition. The detailed normalization workflow performed by DESeq2[69] involved four steps. First, calculated row-wise geometric mean (pseudo-reference sample) by square root of the product of all expression values across samples for each gene. Second, calculated ratio of each sample to the row-wise geometric mean, which was dividing expression value of every gene by their row-wise geometric mean within a sample. Third, the normalization factor was calculated for each sample, which is the median value of all ratios for a sample. Lastly, the normalized count was calculated using the normalization factor, which divided each raw count in a sample by the normalization factor of that sample, and this was performed for every gene in every sample. The normalized counts were log-transformed to eliminate the effect of extreme values.

A total of 80 samples were analysed, including a dilated cardiomyopathy group (65 samples) and control group (15 samples). We utilized R package ggplot2 (v3.2.1)[70] with box and violin plot to generate the two conditions in one graph for the genes encoding ASIC1, ASIC2, ASIC3, and ASIC4, in order to visualize the distribution of gene expression within one group and compare expression differences between groups.

References

1 Australian Institute of Health and Welfare. National Hospital Mortality Database. (2018).

2 Cung, T. T. *et al.* Cyclosporine before PCI in patients with acute myocardial infarction. *N Engl J Med* **373**, 1021–1031, doi:10.1056/NEJMoa1505489 (2015).

3 Roger, V. L. Epidemiology of heart failure. *Circ Res* **113**, 646-659, doi:10.1161/CIRCRESAHA.113.300268 (2013).

4 Cook, C., Cole, G., Asaria, P., Jabbour, R. & Francis, D. P. The annual global economic burden of heart failure. *Int J Cardiol* **171**, 368-376, doi:10.1016/j.ijcard.2013.12.028 (2014).

5 Hellermann, J. P. *et al.* Heart failure after myocardial infarction: a review. *The American Journal of Medicine* **113**, 324-330, doi:10.1016/s0002-9343(02)01185-3 (2002).

6 Savarese, G. & Lund, L. H. Global Public Health Burden of Heart Failure. *Cardiac Failure Review* **03**, doi:10.15420/cfr.2016:25:2 (2017).

7 See Hoe, L. E., Bartnikowski, N., Wells, M. A., Suen, J. Y. & Fraser, J. F. Hurdles to Cardioprotection in the Critically Ill. *Int J Mol Sci* **20**, doi:10.3390/ijms20153823 (2019).

8 ANZOD Registry, 2017 Annual Report, Section 12: Organ Waiting List. (Adelaide, Australia, 2017).

9 Benjamin, E. J. *et al.* Heart Disease and Stroke Statistics-2019 Update: A Report From the American Heart Association. *Circulation* **139**, e56-e528, doi:10.1161/CIR.0000000000000659 (2019).

10 Hausenloy, D. J. & Yellon, D. M. Ischaemic conditioning and reperfusion injury. *Nat Rev Cardiol* **13**, 193-209, doi:10.1038/nrcardio.2016.5 (2016).

11 Sansbury, B. E. *et al.* Metabolomic analysis of pressure-overloaded and infarcted mouse hearts. *Circ Heart Fail* **7**, 634-642, doi:10.1161/CIRCHEARTFAILURE.114.001151 (2014).

12 Stanley, W. C., Recchia, F. A. & Lopaschuk, G. D. Myocardial substrate metabolism in the normal and failing heart. *Physiol Rev* **85**, 1093-1129, doi:10.1152/physrev.00006.2004 (2005).

13 Cobbe, S. The time of onset and severity of acidosis in myocardial ischaemia. *Journal of Molecular and Cellular Cardiology* **12**, 745-760, doi:10.1016/0022-2828(80)90077-2 (1980).

14 Gabel, S. A., Cross, H. R., London, R. E., Steenbergen, C. & Murphy, E. Decreased intracellular pH is not due to increased H+ extrusion in preconditioned rat hearts. *Am J Physiol* **273**, H2257-2262 (1997).

15 Gandhi, A. A. & Akholkar, P. J. Metabolic acidosis in acute myocardial infarction. *International Journal of Advances in Medicine* **2**, 260-263, doi:10.18203/2349-3933.ijam20150556 (2015).

16 Bhowmick, S. *et al.* Acidotoxicity via ASIC1a Mediates Cell Death during Oxygen Glucose Deprivation and Abolishes Excitotoxicity. *ACS Chemical Neuroscience* **8**, 1204-1212, doi:10.1021/acschemneuro.6b00355 (2017).

17 Conklin, D. J. *et al.* TRPA1 channel contributes to myocardial ischemia-reperfusion injury. *American Journal of Physiology-Heart and Circulatory Physiology* **316**, H889-H899, doi:10.1152/ajpheart.00106.2018 (2019).

18 El Banani, H. *et al.* Changes in intracellular sodium and pH during ischaemia-reperfusion are attenuated by trimetazidine: Comparison between low- and zero-flow ischaemia. *Cardiovascular Research* **47**, 688-696, doi:10.1016/S0008-6363(00)00136-X (2000).

19 Mentzer, R. M., Jr. *et al.* Sodium-hydrogen exchange inhibition by cariporide to reduce the risk of ischemic cardiac events in patients undergoing coronary artery bypass grafting: results of the EXPEDITION study. *Ann Thorac Surg* **85**, 1261-1270, doi:10.1016/j.athoracsur.2007.10.054 (2008).

20 Gründer, S. & Chen, X. Structure, function, and pharmacology of acid-sensing ion channels (ASICs): focus on ASIC1a. *Int J Physiol Pathophysiol Pharmacol* **2**, 73-94 (2010).

21 Wemmie, J. A., Taugher, R. J. & Kreple, C. J. Acid-sensing ion channels in pain and disease. *Nat Rev Neurosci* **14**, 461-471, doi:10.1038/nrn3529 (2013).

22 Hu, Y.-L. *et al.* Multiple H+ sensors mediate the extracellular acidification-induced [Ca2+]i elevation in cultured rat ventricular cardiomyocytes. *Scientific Reports* **7**, 44951-44951, doi:10.1038/srep44951 (2017).

23 Li, Z. *et al.* TRPV6 protects ER stress-induced apoptosis via ATF6α-TRPV6-JNK pathway in human embryonic stem cell-derived cardiomyocytes. *Journal of Molecular and Cellular Cardiology* **120**, 1-11, doi:10.1016/j.yjmcc.2018.05.008 (2018).

24 Lipski, J. *et al.* Involvement of TRP-like channels in the acute ischemic response of hippocampal CA1 neurons in brain slices. *Brain Res* **1077**, 187-199, doi:10.1016/j.brainres.2006.01.016 (2006).

25 Babini, E., Paukert, M., Geisler, H. S. & Grunder, S. Alternative splicing and interaction with di- and polyvalent cations control the dynamic range of acid-sensing ion channel 1 (ASIC1). *J Biol Chem* **277**, 41597-41603, doi:10.1074/jbc.M205877200 (2002).

26 Li, M. *et al.* Acid-sensing ion channels in acidosis-induced injury of human brain neurons. *J Cereb Blood Flow Metab* **30**, 1247-1260, doi:10.1038/jcbfm.2010.30 (2010).
27 Immke, D. C. & McCleskey, E. W. Lactate enhances the acid-sensing Na+ channel on ischemia-sensing neurons. *Nature neuroscience* **4**, 869-870, doi:10.1038/nn0901-869 (2001).
28 Allen, N. J. & Attwell, D. Modulation of ASIC channels in rat cerebellar Purkinje neurons by ischaemia-related signals. *J Physiol* **543**, 521-529, doi:10.1113/jphysiol.2002.020297 (2002).
29 McCarthy, C. A., Rash, L. D., Chassagnon, I. n. R., King, G. F. & Widdop, R. E. PcTx1 affords neuroprotection in a conscious model of stroke in hypertensive rats via selective inhibition of ASIC1a. *Neuropharmacology* **99**, 650-657, doi:10.1016/j.neuropharm.2015.08.040 (2015).
30 Xiong, Z.-G. *et al.* Neuroprotection in Ischemia: Blocking Calcium-Permeable Acid-Sensing Ion Channels. *Cell* **118**, 687-698, doi:10.1016/j.cell.2004.08.026 (2004).
31 Chassagnon, I. R. *et al.* Potent neuroprotection after stroke afforded by a double-knot spider-venom peptide that inhibits acid-sensing ion channel 1a. *Proceedings of the National Academy of Sciences* **114**, 201614728-201614728, doi:10.1073/pnas.1614728114 (2017).
32 Quaife-Ryan, G. A. *et al.* Multicellular transcriptional analysis of mammalian heart regeneration. *Circulation* **136**, 1123-1139, doi:10.1161/CIRCULATIONAHA.117.028252 (2017).
33 Macdonald, P. S. Heart Transplantation From DCD donors: From the Bedside to the Bench. *Transplantation* **101**, 1753-1754, doi:10.1097/TP.0000000000001829 (2017).
34 Iyer, A. *et al.* Normothermic ex vivo perfusion provides superior organ preservation and enables viability assessment of hearts from DCD donors. *Am J Transplant* **15**, 371-380, doi:10.1111/ajt.12994 (2015).
35 Iyer, A. *et al.* Increasing the tolerance of DCD hearts to warm ischemia by pharmacological postconditioning. *Am J Transplant* **14**, 1744-1752, doi:10.1111/ajt.12782 (2014).
36 Dhital, K. K. *et al.* Adult heart transplantation with distant procurement and ex-vivo preservation of donor hearts after circulatory death: a case series. *The Lancet* **385**, 2585-2591, doi:10.1016/s0140-6736(15)60038-1 (2015).
37 Linkermann, A. *et al.* Two independent pathways of regulated necrosis mediate ischemia-reperfusion injury. *Proc Natl Acad Sci U S A* **110**, 12024-12029, doi:10.1073/pnas.1305538110 (2013).
38 Zhu, H. & Sun, A. Programmed necrosis in heart disease: Molecular mechanisms and clinical implications. *J Mol Cell Cardiol* **116**, 125-134, doi:10.1016/j.yjmcc.2018.01.018 (2018).
39 Wang, Y. Z. *et al.* Tissue acidosis induces neuronal necroptosis via ASIC1a channel independent of its ionic conduction. *eLife* **4**, 1-21, doi:10.7554/eLife.05682 (2015).
40 van Heesch, S. *et al.* The Translational Landscape of the Human Heart. *Cell* **178**, 242-260 e229, doi:10.1016/j.cell.2019.05.010 (2019).
41 King, E. A., Davis, J. W. & Degner, J. F. Are drug targets with genetic support twice as likely to be approved? Revised estimates of the impact of genetic support for drug mechanisms on the probability of drug approval. *BioRxiv*, doi:10.1101/513945 (2019).
42 *The UK Biobank*, <http://www.nealelab.is/uk-biobank> (2018).
43 Bakshi, A. *et al.* Fast set-based association analysis using summary data from GWAS identifies novel gene loci for human complex traits. *Sci Rep* **6**, 32894, doi:10.1038/srep32894 (2016).
44 Friedman, C. E. *et al.* Single-Cell Transcriptomic Analysis of Cardiac Differentiation from Human PSCs Reveals HOPX-Dependent Cardiomyocyte Maturation. *Cell Stem Cell* **23**, 586-598.e588, doi:10.1016/j.stem.2018.09.009 (2018).
45 Bässler, E. L., Ngo-Anh, T. J., Geisler, H. S., Ruppersberg, J. P. & Gründer, S. Molecular and Functional Characterization of Acid-sensing Ion Channel (ASIC) 1b. *Journal of Biological Chemistry* **276**, 33782-33787, doi:10.1074/jbc.M104030200 (2001).
46 Bohlen, C. J. *et al.* A heteromeric Texas coral snake toxin targets acid-sensing ion channels to produce pain. *Nature* **479**, 410-414, doi:10.1038/nature10607 (2011).
47 Besson, T., Lingueglia, E. & Salinas, M. Pharmacological modulation of Acid-Sensing Ion Channels 1a and 3 by amiloride and 2-guanidine-4-methylquinazoline (GMQ). *Neuropharmacology* **125**, 429-440, doi:10.1016/j.neuropharm.2017.08.004 (2017).
48 Zhang, X.-h., Saric, T., Mehrjardi, N.-Z., Hamad, S. & Morad, M. Acid Sensitive Ion channels are expressed in human induced pluripotent stem cell-derived cardiomyocytes. *Stem Cells and Development*, scd.2018.0234-scd.2018.0234, doi:10.1089/scd.2018.0234 (2019).
49 Herrera, Y. *et al.* sigma-1 receptor modulation of acid-sensing ion channel a (ASIC1a) and ASIC1a-induced Ca2+ influx in rat cortical neurons. *J Pharmacol Exp Ther* **327**, 491-502, doi:10.1124/jpet.108.143974 (2008).

50 Wang, Y. Z. *et al.* Intracellular ASIC1a regulates mitochondrial permeability transition-dependent neuronal death. *Cell Death and Differentiation* **20**, 1359-1369, doi:10.1038/cdd.2013.90 (2013).
51 Mentzer, R. M., Lasley, R. D., Jessel, A. & Karmazyn, M. Intracellular sodium hydrogen exchange inhibition and clinical myocardial protection. *The Annals of Thoracic Surgery* **75**, S700-S708, doi:10.1016/s0003-4975(02)04700-8 (2003).
52 Ghanekar, S., Corey, S., Lippert, T. & Borlongan, C. V. Pathological links between stroke and cardiac arrest. *Chinese Neurosurgical Journal* **2**, doi:10.1186/s41016-016-0056-1 (2016).
53 Fanning, J. P. *et al.* Neuron-Specific Enolase and Matrix Metalloproteinase 9 Signal Perioperative Silent Brain Infarction During or After Transcatheter Aortic Valve Implantation. *Am J Cardiol* **123**, 434-439, doi:10.1016/j.amjcard.2018.10.022 (2019).
54 Fisher, M. & Saver, J. L. Future directions of acute ischaemic stroke therapy. *The Lancet Neurology* **14**, 758-767, doi:10.1016/s1474-4422(15)00054-x (2015).
55 Stehlik, J. *et al.* The Registry of the International Society for Heart and Lung Transplantation: 29th official adult heart transplant report--2012. *J Heart Lung Transplant* **31**, 1052-1064, doi:10.1016/j.healun.2012.08.002 (2012).
56 Kobashigawa, J. *et al.* Report from a consensus conference on primary graft dysfunction after cardiac transplantation. *J Heart Lung Transplant* **33**, 327-340, doi:10.1016/j.healun.2014.02.027 (2014).
57 Malik, R. *et al.* Multiancestry genome-wide association study of 520,000 subjects identifies 32 loci associated with stroke and stroke subtypes. *Nat Genet* **50**, 524-537, doi:10.1038/s41588-018-0058-3 (2018).
58 Saez, N. J. *et al.* A dynamic pharmacophore drives the interaction between Psalmotoxin-1 and the putative drug target acid-sensing ion channel 1a. *Mol Pharmacol* **80**, 796-808, doi:10.1124/mol.111.072207 (2011).
59 See Hoe, L. E. *et al.* Regulation of the beta-Adrenergic Receptor Signaling Pathway in Sustained Ligand-Activated Preconditioning. *J Pharmacol Exp Ther* **369**, 37-46, doi:10.1124/jpet.118.251660 (2019).
60 Reichelt, M. E., Willems, L., Hack, B. a., Peart, J. N. & Headrick, J. P. Cardiac and coronary function in the Langendorff-perfused mouse heart model. *Experimental physiology* **94**, 54-70, doi:expphysiol.2008.043554 [pii]\n10.1113/expphysiol.2008.043554 (2009).
61 Gao, F., Zhang, J., Wang, F., Xin, X. & Sha, D. Cyclosporin A-related cerebral venous sinus thrombosis: A case report. *Medicine (Baltimore)* **97**, e11642, doi:10.1097/MD.0000000000011642 (2018).
62 Gao, L., Hicks, M. & MacDonald, P. S. Improved preservation of the rat heart with celsior solution supplemented with cariporide plus glyceryl trinitrate. *Am J Transplant* **5**, 1820-1826, doi:10.1111/j.1600-6143.2005.00967.x (2005).
63 Kumarasinghe, G. *et al.* Improved heart function from older donors using pharmacologic conditioning strategies. *J Heart Lung Transplant* **35**, 636-646, doi:10.1016/j.healun.2015.12.020 (2016).
64 Saez, N. J. *et al.* Molecular dynamics and functional studies define a hot spot of crystal contacts essential for PcTx1 inhibition of acid-sensing ion channel 1a. *Br J Pharmacol* **172**, 4985-4995, doi:10.1111/bph.13267 (2015).
65 Miyaoka, Y. *et al.* Isolation of single-base genome-edited human iPS cells without antibiotic selection. *Nat Methods* **11**, 291-293, doi:10.1038/nmeth.2840 (2014).
66 Kreitzer, F. R. *et al.* A robust method to derive functional neural crest cells from human pluripotent stem cells. *Am J Stem Cell* **2**, 119-131 (2013).
67 Schindelin, J. *et al.* Fiji: an open-source platform for biological-image analysis. *Nat Methods* **9**, 676-682, doi:10.1038/nmeth.2019 (2012).
68 Cuéllar-Partida, G. *et al.* Complex-Traits Genetics Virtual Lab: A community-driven web platform for post-GWAS analyses. *BioRxiv*, doi:10.1101/518027 (2019).
69 Love, M. I., Huber, W. & Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* **15**, 550, doi:10.1186/s13059-014-0550-8 (2014).
70 Wickham, H. *ggplot2: Elegant Graphics for Data Analysis*. (Springer-Verlag 2016).

Fig. 1

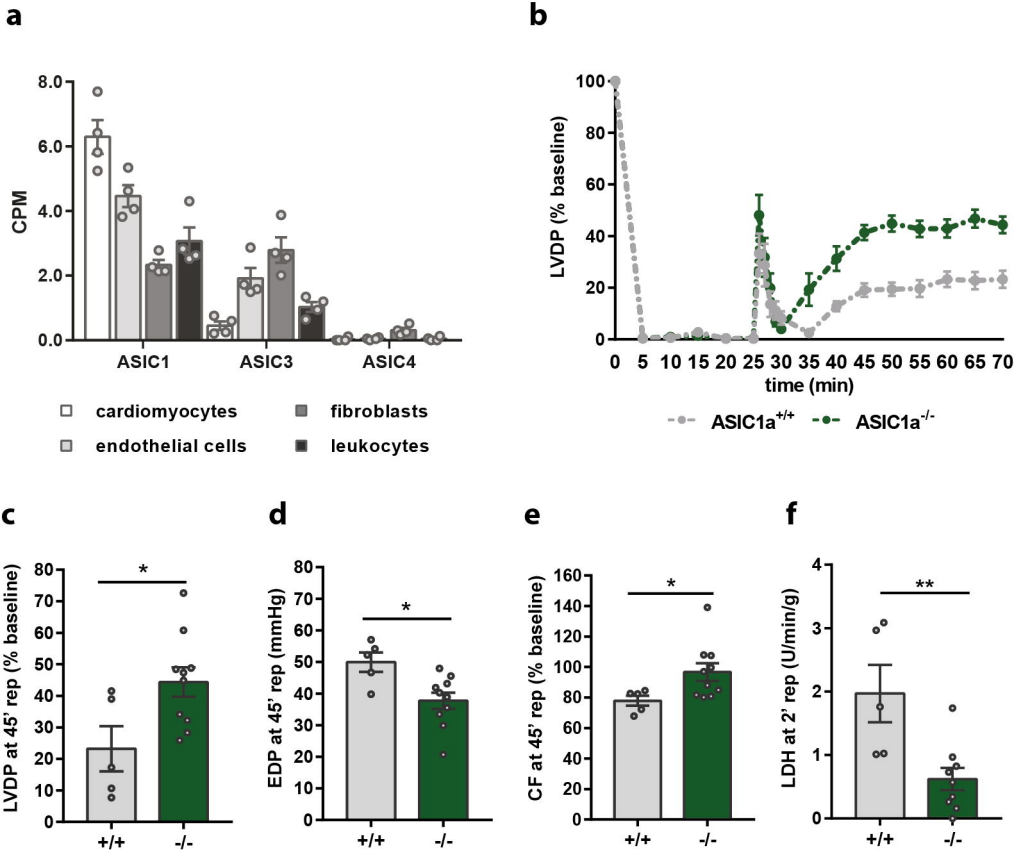

a
CPM
8.0
6.0
4.0
2.0
0.0
ASIC1
ASIC3
ASIC4
cardiomyocytes
fibroblasts
endothelial cells
leukocytes
b
LVDP (% baseline)
100
80
60
40
20
0
0 5 10 15 20 25 30 35 40 45 50 55 60 65 70
time (min)
ASIC1a+/+
ASIC1a-/-
c
LVDP at 45' rep (% baseline)
*
+/+
-/-
d
EDP at 45' rep (mmHg)
*
+/+
-/-
e
CF at 45' rep (% baseline)
*
+/+
-/-
f
LDH at 2' rep (U/min/g)
**
+/+
-/-

Fig. 2

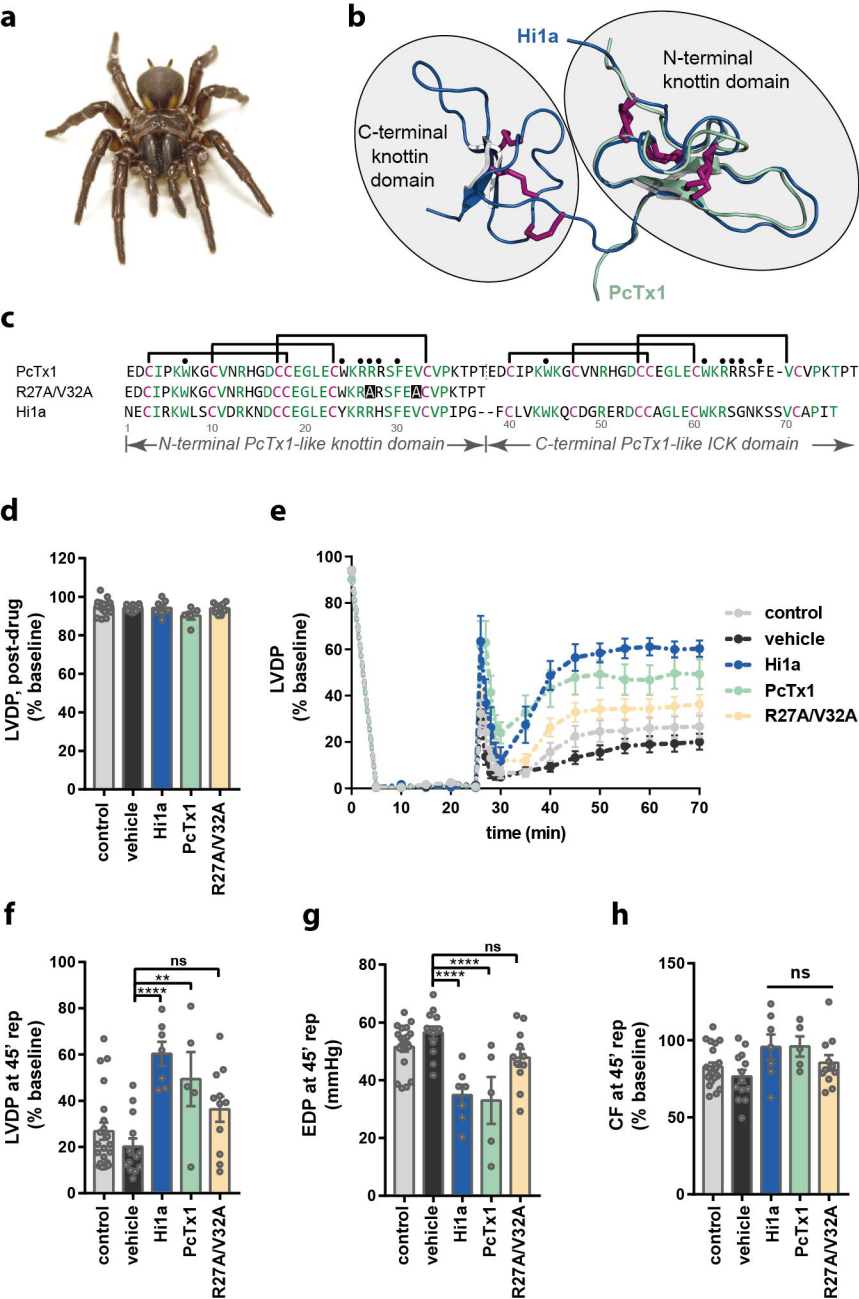
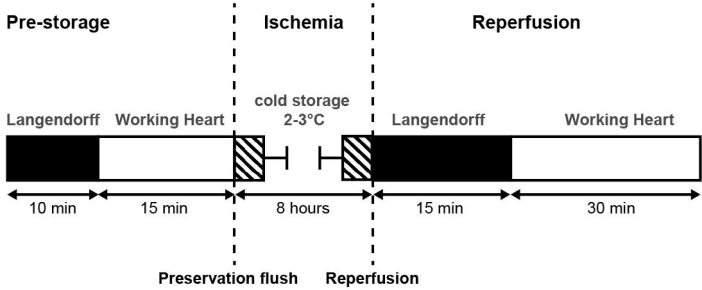
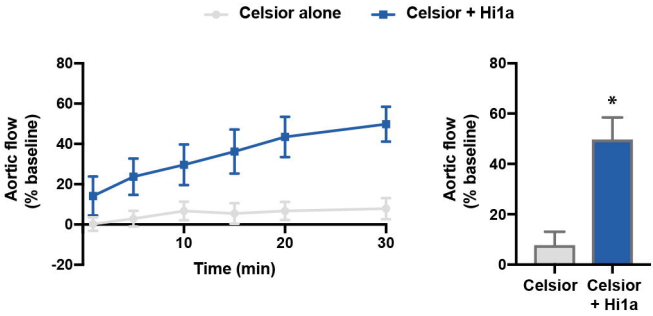

a
b
Hi1a
N-terminal knottin domain
C-terminal knottin domain
PcTx1
c
PcTx1 EDCIPKWKGCVNRHGDCCEGLECWKRRRSFEVCVPKTPTEDCIPKWKGCVNRHGDCCEGLECWKRRRSFE-VCVPKTPT
R27A/V32A EDCIPKWKGCVNRHGDCCEGLECWKRARSFEACVPKTPT
Hi1a NECIRKWLSCVDRKNDCCEGLECYKRRHSFEVCVPIPG--FCLVKWKQCDGRERDCCAGLECWKRSGNKSSVCAPIT
1 10 20 30 40 50 60 70
N-terminal PcTx1-like knottin domain
C-terminal PcTx1-like ICK domain
d
LVDP, post-drug (% baseline)
control
vehicle
Hi1a
PcTx1
R27A/V32A
e
LVDP (% baseline)
time (min)
control
vehicle
Hi1a
PcTx1
R27A/V32A
f
LVDP at 45' rep (% baseline)
ns
**
****
g
EDP at 45' rep (mmHg)
ns
****
****
h
CF at 45' rep (% baseline)
ns

Fig. 3

a


Pre-storage
Ischemia
Reperfusion
cold storage
2-3°C
Langendorff
Working Heart
Langendorff
Working Heart
10 min
15 min
8 hours
15 min
30 min
Preservation flush
Reperfusion

b


Celsior alone
Celsior + Hi1a
Aortic flow
(% baseline)
Time (min)
80
60
40
20
0
-20
10
20
30
*
Celsior
Celsior
+ Hi1a

c

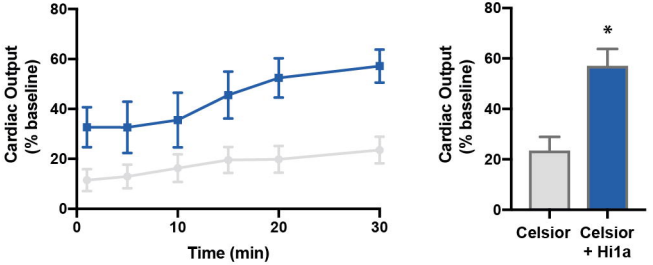

Cardiac Output
(% baseline)
Time (min)
80
60
40
20
0
0
10
20
30
*
Celsior
Celsior
+ Hi1a

Fig. 4

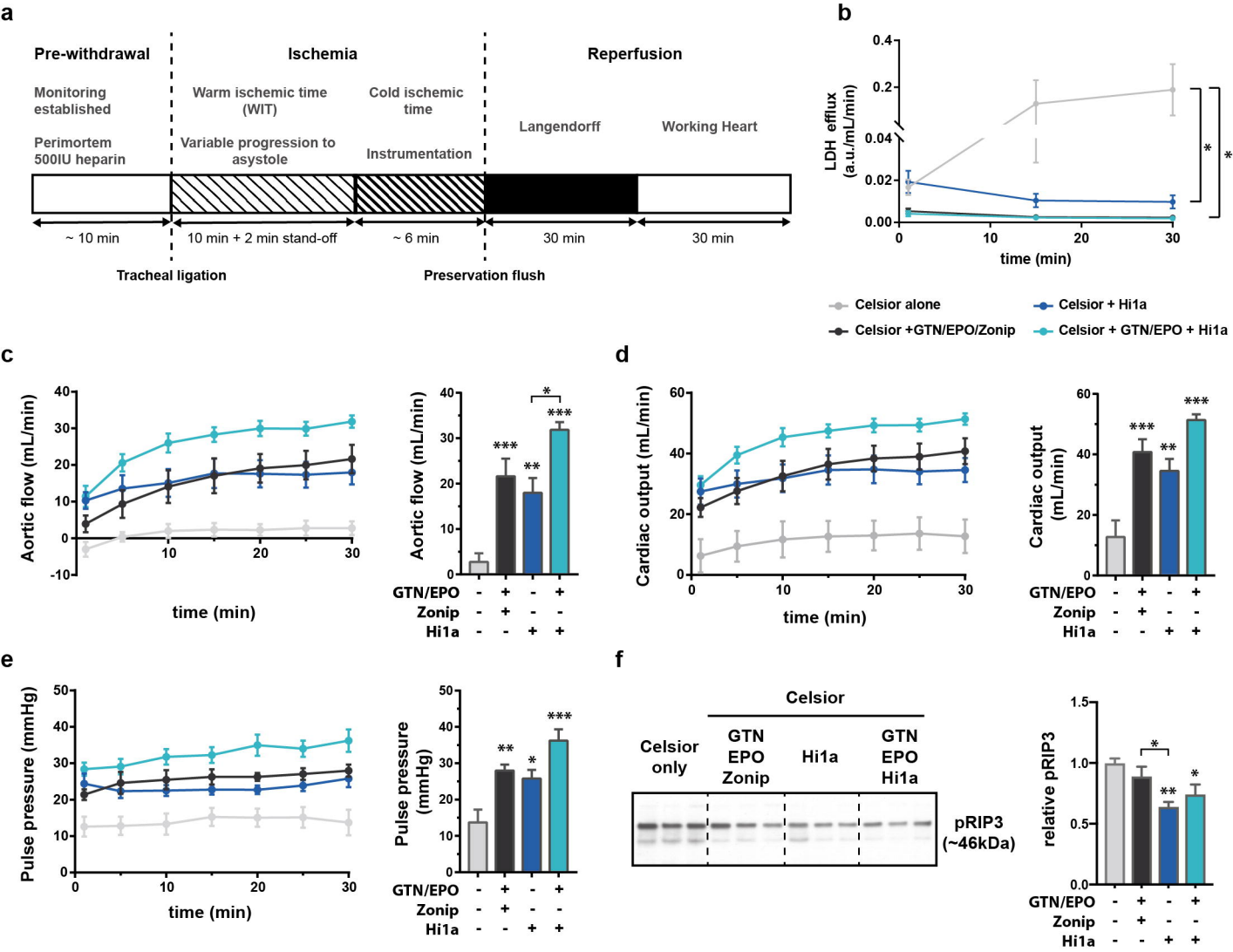
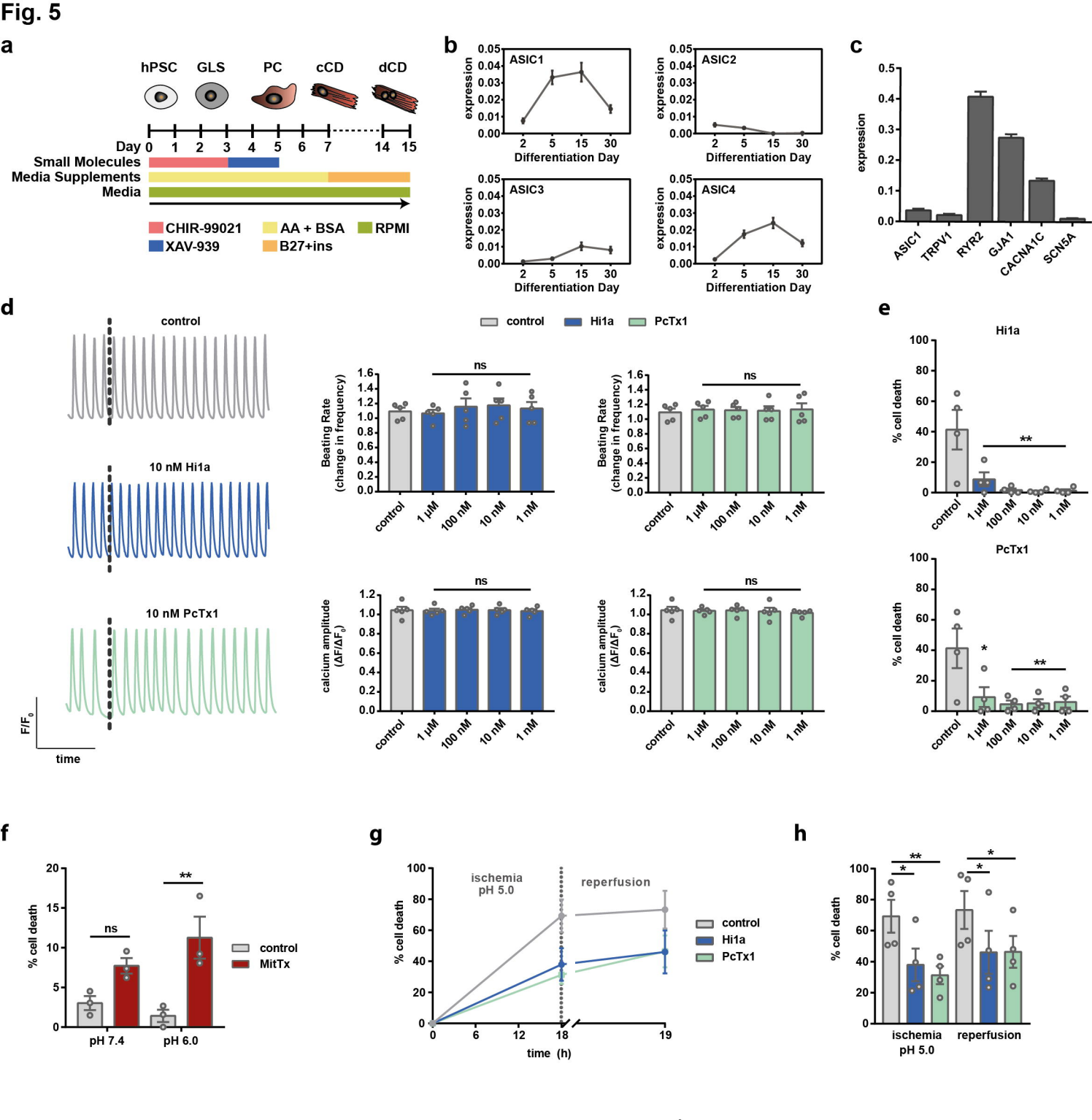

a
Pre-withdrawal
Ischemia
Reperfusion
Monitoring established
Perimortem 500IU heparin
Warm ischemic time (WIT)
Variable progression to asystole
Cold ischemic time
Instrumentation
Langendorff
Working Heart
~ 10 min
10 min + 2 min stand-off
~ 6 min
30 min
30 min
Tracheal ligation
Preservation flush
b
LDH efflux (a.u./mL/min)
time (min)
Celsior alone
Celsior + Hi1a
Celsior +GTN/EPO/Zonip
Celsior + GTN/EPO + Hi1a
c
Aortic flow (mL/min)
time (min)
GTN/EPO
Zonip
Hi1a
d
Cardiac output (mL/min)
time (min)
e
Pulse pressure (mmHg)
time (min)
f
Celsior
Celsior only
GTN EPO Zonip
Hi1a
GTN EPO Hi1a
pRIP3 (~46kDa)
relative pRIP3

Fig. 5


a
hPSC
GLS
PC
cCD
dCD
Day 0 1 2 3 4 5 6 7 14 15
Small Molecules
Media Supplements
Media
CHIR-99021
XAV-939
AA + BSA
B27+ins
RPMI
b
ASIC1
ASIC2
ASIC3
ASIC4
expression
Differentiation Day
c
expression
ASIC1
TRPV1
RYR2
GJA1
CACNA1C
SCN5A
d
control
10 nM Hi1a
10 nM PcTx1
F/F0
time
control
Hi1a
PcTx1
Beating Rate (change in frequency)
calcium amplitude (ΔF/ΔF0)
ns
control
1 μM
100 nM
10 nM
1 nM
e
Hi1a
PcTx1
% cell death
**
*
f
% cell death
ns
**
control
MitTx
pH 7.4
pH 6.0
g
ischemia pH 5.0
reperfusion
% cell death
control
Hi1a
PcTx1
time (h)
h
% cell death
**
*
ischemia pH 5.0
reperfusion