

1 **Research Article**

2

3 **Full Title:**

4 Laboratory testing of very low-copper-treated water to prolong pupation and
5 emerging time of mosquito larvae: an alternative method to delay mosquito
6 breeding capability

7

8 **Short Title:**

9 Effects of exposure to very low concentrations of copper on mosquito larvae

10

11 **Authors:**

12 Mohamad Reza^{a*}, Cimi Ilmiawati^b

13 *^a Department of Biology, Faculty of Medicine, Andalas University, Padang, West
14 Sumatra, Indonesia*

15 *^b Department of Pharmacology, Faculty of Medicine, Andalas University, Padang,
16 West Sumatra, Indonesia*

17

18 ***Corresponding Author:**

19 Mohamad Reza, MD, PhD

20 Department of Biology

21 Faculty of Medicine, Andalas University

22 Main Campus at Limau Manis, Gedung C Lantai 2

23 Pauh, Padang, 25166

24 West Sumatra, Indonesia

25 e-mail: reza@med.unand.ac.id

26

27 **ABSTRACT**

28

29 **Introduction:** Larvicide application in ovitrap is one of the currently available
30 methods used in mosquito eradication campaign because they eliminate the larval
31 stage. We previously reported that copper in liquid form is a promising candidate
32 due to its potent larvicide properties in a laboratory setting and even in the field. In
33 the field study, several larvae survived in the ovitrap due to the dilution of copper
34 concentration by the rain. The surviving larvae were smaller and less motile. This
35 led our interest to study the effect of a sub-lethal dose of copper in ovitrap on
36 larvae development, pupation time and lifespan in the adult stage.

37 **Methods:** First instar larvae of three species of mosquito (*Aedes albopictus*, *Culex*
38 *pipiens* and *Anopheles stephensi*) were bred in copper-treated water at a
39 concentration of 0.60 ppm, 0.30 ppm, and 0.15 ppm and compared with the
40 control group. The surviving larvae were recorded every day in terms of pupa
41 emerging time and adult emerging time. The number of adult mortality was
42 recorded and compared with the control.

43 **Results:** Copper showed potent larvicide effect in the 0.60 ppm concentration and
44 prolonged pupation time and caused a significantly lower number of emerging
45 mosquitoes down to the lowest concentration of 0.15 ppm. The adult lifespan was
46 not different compared to the control.

47 **Conclusion:** This study demonstrates the capability of copper below 1 ppm to
48 prolong the pupation time and the emerging time of mosquito larvae. Our findings
49 open the possibility of copper application to cut mosquito breeding capacity that
50 eventually will reduce disease transmission.

51 **Keywords:** *Aedes albopictus*, *Anopheles stephensi*, *Culex pipiens*, copper, larval
52 control, ovitrap, pupation time

53

54 INTRODUCTION

55 Mosquitoes are causing millions of deaths every year worldwide. More
56 than half of the world's populations live in areas where mosquitoes are present as
57 vectors and transmit malaria, dengue, Zika, Chikungunya, yellow fever and other
58 diseases [1]. Mosquito-borne diseases are causing an enormous burden on the
59 public health system in many countries with limited financial and human
60 resources. Despite being considered as the most conceivable method to eradicate
61 mosquito-borne diseases [1], global funding for malaria vector control is far below
62 the budget endowed for exploring cures and vaccines for these diseases (US\$ 56
63 vs 408 million in 2018) [2]. Lack of funding is considered as one of the challenges
64 in achieving and sustaining malaria control [3].

65 Core methods for vector control, i.e. insecticide-treated nets (ITNs) and
66 indoor residual spraying (IRS), rely on insecticides. Unfortunately, there is an
67 increase in mosquito resistance to insecticides [4]. Therefore, there is a need to
68 develop novel methods for vector control, particularly practical and economical
69 ones, to be combined with the available methods in integrated vector management.
70 Larval source management (LSM) is part of integrated vector control using
71 strategies to modify or manipulate water bodies as the potential larval habitats of
72 mosquitoes to prevent maturation of mosquito developmental stages. LSM
73 strategies also include the introduction of larvicide (chemical and biological
74 agents) and larvivorous fish (natural predator) into larval habitats [5-7]. Previous
75 studies have proposed the utility of using metallic [8, 9] and liquid [5, 10, 11]

76 copper as a potential and affordable larvicide [7]. Copper is effective as a
77 mosquito larvicide at a concentration below 2 ppm [9], the threshold value deemed
78 safe for human drinking water [12], thus making copper a potential candidate for
79 use in the public health setting.

80 Our previous findings on the properties of copper to kill mosquito larvae
81 [10] and laboratory testing on the effect of liquid copper at a concentration of 10
82 ppm on three species of mosquito larvae [11] prompted us to perform a field test
83 in a malaria and dengue-endemic area of West Sumatra, Indonesia. Despite
84 confirming the larvicide effect of copper [7], we observed that dilution by
85 rainwater should be considered as confounding in the field since we noted that
86 some larvae survived in outdoor ovitraps. However, the survived larvae appeared
87 to be less motile and were smaller in size compared to unexposed larvae.
88 Consequently, further study is required to determine whether the survived larvae
89 capable to continue their life cycle until adulthood. Therefore, we performed a
90 laboratory test of sub-lethal concentrations of copper exposure to mosquito larvae
91 to verify its effect on mosquito development and breeding capacity.

92

93 **MATERIALS AND METHODS**

94 **Ethical Approval**

95 This research was approved by the ethics committee of the Faculty of
96 Medicine, Universitas Andalas (Approval No.036/KEP/FK/2019).

97

98 **Mosquito Eggs and Mice**

99 *Anopheles stephensi* (strain SDA 500), were reared in our laboratory under
100 room temperature (26°C), 50-70% relative humidity, and a 13:11 h light: dark

101 cycle. From the day of emergence, mosquitoes were provided with a 5% fructose
102 solution soaked in a filter paper. Females five days after emergence were allowed
103 to feed on anesthetized mice. Three days later, an oviposition dish was placed in a
104 cage containing gravid females. The eggs were laid on a filter paper soaking in
105 the dish. The filter paper with eggs was placed on a 12 x 20 cm hatching tray
106 containing 500 ml of water. After hatching, 3-5 mg of carp food per tray was
107 sprinkled on the surface of the water twice daily. Twelve days later, pupae were
108 collected daily and transferred to cages for adult emergence.

109 Eggs of *Aedes albopictus* and *Culex pipiens* were purchased and the first
110 instar newly hatched larvae were utilized in the experiment. Female BALB/c mice
111 were purchased from SLC (Shizuoka, Japan). The mice were fed *ad libitum* and
112 exposed to a 13:11 h of light: dark cycle. This mosquito-mouse cycle was used to
113 maintain mosquitoes in the laboratory.

114

115 **Preparation of Copper Sulfate Solutions**

116 Four concentrations of CuSO₄ solution were used for the experiment. The
117 copper concentrations were 0.60, 0.30, 0.15, and 0 ppm. Each solution was
118 allocated to a pre-washed square plastic plate and covered with a transparent
119 plastic plate. First, we prepared a standard solution of 100 mM of CuSO₄ by
120 mixing copper sulfate powder with filtered water (Fine Ceramic Filter NGK
121 insulators, Nagoya, Japan). Then, we prepared the four concentrations for the
122 experiment by diluting them from the standard solution and confirming the copper
123 level using a copper measuring device (Hanna Instruments, Tokyo, Japan) and Z-
124 5010 Polarized Zeeman flame atomic absorption spectrophotometer (Hitachi Ltd,
125 Tokyo, Japan).

126

127 **Preparation of Mosquito Larvae**

128 First instar newly hatched larvae were used in this experiment. Fifty larvae
129 were separated and put inside each container. Three containers for each
130 concentration of copper were prepared. Larvae were allowed to hatch and were
131 observed every 24-hours under the microscope for mortality. A fine ceramic filter
132 (NGK Insulators, Nagoya, Japan) was used to remove chlorine ions from tap water
133 for the experiment.

134

135 **RESULTS**

136 To determine the effect of very low concentrations of copper on mosquito
137 larval mortality, larvae of *A. albopictus*, *A. stephensi* and *C. pipiens* were exposed
138 to 0.60, 0.30 and 0.15 ppm of copper solutions (CuSO₄) and were observed daily.
139 Larvae of *A. stephensi* and *C. pipiens* exposed to 0.60 ppm of copper showed
140 100% mortality within a week, while a small number of *A. albopictus* larvae
141 survived. Larvae from three species roughly showed 50% mortality when exposed
142 to 0.30 ppm of copper for seven days. CuSO₄ at 0.15 ppm only had a statistically
143 significant effect on *C. pipiens* larval mortality compared to control, started on the
144 third day of exposure (**Figure 1**).

145

146 **Figure 1. The observed mortality of *A. albopictus*, *A. stephensi*, and *C. pipiens***
147 **larvae on exposure to 0.15, 0.30, and 0.60 ppm of CuSO₄. *significantly**
148 **different from control, p<0.05 (t-test or Mann-Whitney U test)**

149

150 To figure out the effect of very low concentrations of copper exposure to
151 larval pupation time, the number of emerging pupae from the three species were
152 counted. Pupae started to emerge at day eight to 10, depending on the mosquito
153 species. Surviving larvae exposed to 0.15 and 0.30 ppm of copper showed
154 prolonged pupation time in all species observed compared to the control group.
155 We also observed that the surviving larvae of *Ae. albopictus* on 0.60 ppm were not
156 turned into pupae (**Figure 2**).

157

158 **Figure 2. The number of emerging pupae of *A. albopictus*, *A. stephensi*, and *C.***
159 ***pipiens* on exposure to 0.15, 0.30, and 0.60 ppm of CuSO₄. *significantly**
160 **different from control, p<0.05 (Mann-Whitney U test)**

161

162 To determine the effect of very low concentrations of copper exposure on
163 mosquito ability to reach adulthood, we observed and counted the number of
164 emerging adults in three exposed species. Surviving pupae of all observed species
165 exposed to 0.15 ppm of CuSO₄ become adult mosquitoes, albeit at a statistically
166 significantly lower number compared to the control group. Adult mosquito of all
167 observed species emerged at later days compared to control and survived up to 22
168 days of observation. Almost none of the pupae of all observed species exposed to
169 0.30 ppm of CuSO₄ emerged as an adult mosquito (**Figure 3**).

170

171 **Figure 3. The number of emerging *A. albopictus*, *A. stephensi*, and *C. pipiens***
172 **mosquito on exposure to 0.15, 0.30, and 0.60 ppm of CuSO₄. *significantly**
173 **different from control, p<0.05 (Mann-Whitney U test)**

174

175 It is noteworthy that all adults emerged in copper-treated groups and
176 control appeared to be viable and showed no difference in term of their lifespan. It
177 seemed that the copper effect on the survived larvae was limited to prolonging the
178 emergence of pupae and was not carried on to the adult stage.

179

180 **DISCUSSION**

181 The results of the present study demonstrate that copper adequately lead to
182 a high rate of larval mortality in at a concentration below 1 ppm (0.60 ppm). At a
183 further lower concentration (0.30 ppm), copper still kill half of the larvae, and
184 together with the lowest concentration of 0.15 ppm, prolongs pupation time and
185 the emerging time of adult mosquitoes compared to the control group. The number
186 of mosquitoes that emerged afterward is also significantly lower than those in the
187 control group.

188 However, the copper effect is limited in the larvae and pupae stage. Copper
189 shows no effect on the adult stage. The effect on the larvae probably caused by the
190 copper effect to commensal bacteria in the larval midgut, which disrupts their
191 intestinal function [13]. This prevents them to eat properly and to collect enough
192 energy for pupation and emerge as an adult mosquito. Once the larvae become
193 adult, this copper effect no longer plays a role in their life cycle.

194 This is an interesting finding, since at a very low concentration copper
195 prolongs breeding time of mosquito. This might lead to the possibility to utilize
196 copper in a tap water to jeopardize mosquitoes breeding capability. The
197 mosquitoes will need a longer time to breed, which then reduce their capability to
198 transmit mosquito-borne diseases such as dengue hemorrhagic fever.

199 It is also known that *Aedes sp.* are resistant to insecticide in indoor or
200 outdoor spraying. A study conducted in Sri Lanka showed *Aedes aegypti* and
201 *Aedes albopictus* were highly resistant to DDT and they can oviposit indoors and
202 outdoors [14]. Another study in Bangladesh reported abundant potential larval
203 habitats for *Aedes sp.* in containers or jars spread around the city [15]. These
204 situations need another approach of vector control, such as laying ovitraps or
205 another effort to cut or delay mosquito breeding capability, or even provide
206 insecticide in the tap water if possible.

207 The US EPA is suggesting 1.3 ppm as a safe limit of copper concentration
208 in the drinking water [16], thus making the application of very low 0.15 ppm of
209 copper might be logically acceptable in the future.

210 We believed that, together with the application of copper in the ovitraps,
211 the concentration of 0.15 ppm of copper in the tap water in houses and large scale
212 water reservoir might be a potential strategy in cutting mosquito breeding
213 capability and their ability to transmit mosquito-borne diseases, especially dengue
214 hemorrhagic fever, which mainly breed in the city water reservoir and container.
215 This result might be a chance to re-evaluate the allowance of copper in drinking
216 water as one of the possible strategies in eradicating mosquito-borne diseases in
217 the future.

218

219 **Authors` contributions:** MR conceived and designed the study protocol. MR and
220 CI formulated the study proposal and obtained funding. MR conducted research in
221 the laboratory. MR carried out data analysis and interpretation and wrote the
222 manuscript, while CI conducted the statistical analysis. All authors read and
223 approved the final manuscript.

224

225 **Acknowledgment:** The authors thank Prof. Hirotomo Kato, Ph.D for his kind
226 permit to use his laboratory in Japan for this research.

227

228 **Conflict of interest:** The authors report no conflicts of interest.

229

230 **References**

231

232 1. WHO. Mosquito-borne diseases. 2019. Available from:
233 [https://www.who.int/neglected_diseases/vector_ecology/mosquito-](https://www.who.int/neglected_diseases/vector_ecology/mosquito-borne-diseases/en/)
234 [borne-diseases/en/](https://www.who.int/neglected_diseases/vector_ecology/mosquito-borne-diseases/en/)

235

236 2. WHO. Global malaria report 2019. Geneva: World Health Organization;
237 2019.

238

239 3. Bastiaens GJH, Bousema T, Leslie T. Scale-up of malaria rapid
240 diagnostic tests and artemisinin-based combination therapy: challenges
241 and perspectives in Sub-Saharan Africa. PLoS Med. 2014;11(1):
242 e1001590. doi:10.1371/journal.pmed.1001590.

243

244 4. Liu N. Insecticide resistance in mosquitoes: impact, mechanisms, and
245 research directions. Annu Rev Entomol. 2015; 60:537-559.

246

247 5. Reza M, Yamamoto DS, Matsuoka H. Laboratory testing of larvivoracious
248 fish Japanese medaka (*Oryzias latipes*) predation ability to copper-

- 249 treated *Anopheles stephensi* larvae: an alternative method for vector
250 control under low concentration of copper. Med Entomol Zool.
251 2013;64(2):67-71.
- 252
- 253 6. WHO. Larval source management: a supplementary measure for malaria
254 vector control: an operational manual. Geneva: World Health
255 Organization; 2013.
- 256
- 257 7. Reza M, Ilmiawati C, Matsuoka H. Application of copper-based ovitraps
258 in local houses in West Sumatera, Indonesia: a field test of a simple and
259 affordable larvicide for mosquito control. Trop Med Health. 2016;44:11.
260 doi:10.1186/s13071-016-0007-8.
- 261
- 262 8. Romi R, Di Luca M, Raineri W, Pesce M, Rey A, Giovannangeli S,
263 Zanasi F, Bella A. Laboratory and field evaluation of metallic copper on
264 *Aedes albopictus* (Diptera: Culicidae) larval development. J Med
265 Entomol. 2000; 37(2):281-285.
- 266
- 267 9. Becker N, Oo TT, Schork N. Metallic copper spray – a new control
268 technique to combat invasive container-inhabiting mosquitoes. Parasites
269 Vectors. 2015;8:575. doi:10.1186/s13071-015-1180-z.
- 270
- 271 10. Reza M, Yamamoto DS, Matsuoka H. Low concentration copper
272 solution jeopardizes larval movement and ability to survive predation:

273 new insight into malaria eradication via vector control. Med Entomol
274 Zool. 2012;63:217-222.

275

276 11. Reza M, Yamamoto DS, Matsuoka H. Larvicidal and ovipositional
277 preference test of copper solution for mosquitos. Med Entomol Zool.
278 2014; 65(3):147-150.

279

280 12. WHO. Trace elements in human nutrition and health. Geneva: World
281 Health Organization; 1996.

282

283 13. Beaty BJ, Mackie RS, Mattingly KS, Carlson JO, Rayms-Keller A. The
284 midgut epithelium of aquatic arthropods: a critical target organ in
285 environmental toxicology. Environ Health Perspect.
286 2002;110(suppl6):911-914.

287

288 14. Dharsini S, Vinobaba M, Jude PJ et al. Prevalence and insecticide
289 susceptibility of dengue vectors in the district of Batticaloa in Eastern Sri
290 Lanka. Trop Med Health. 2011;39:47-52.

291

292 15. Ferdousi F, Yoshimatsu S, Ma E et al. Identification of essential
293 containers for *Aedes* larval breeding to control dengue in Dhaka,
294 Bangladesh. Trop Med Health. 2015;43(4):253-264.

295 16. US EPA. Drinking Water Requirements for States and Public Water
296 Systems: Lead and Copper Rule. Available from:
297 <https://www.epa.gov/dwreginfo/lead-and-copper-rule>

298

299

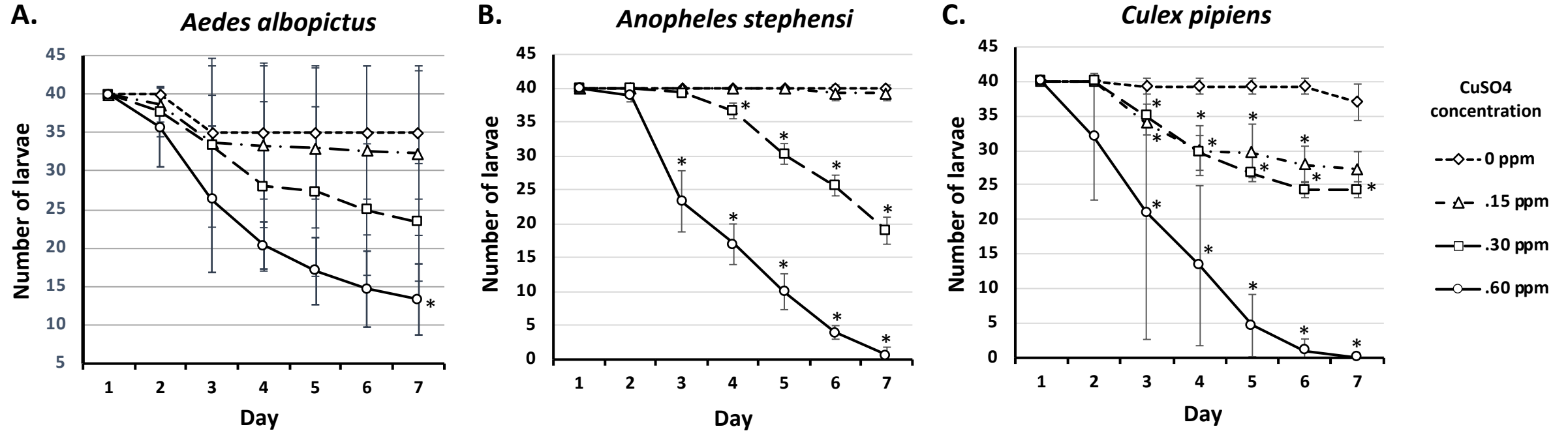


Figure 1. The observed mortality of *A. albopictus*, *A. stephensi*, and *C. pipiens* larvae on exposure to 0.15, 0.30, and 0.60 ppm of CuSO_4 . *significantly different from control, $p < 0.05$ (t-test or Mann-Whitney U test)

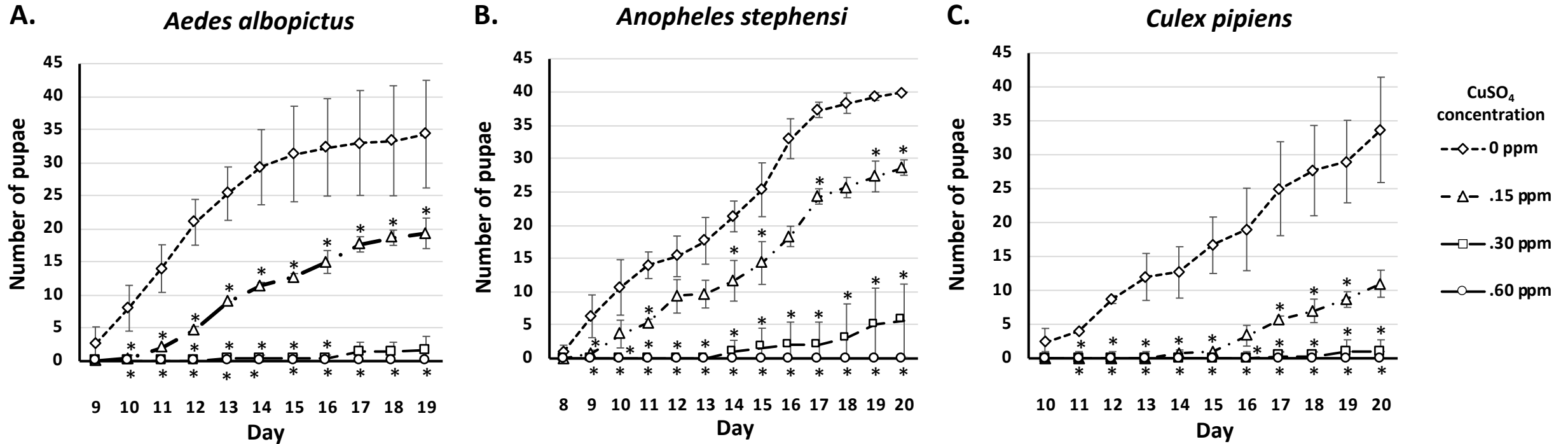


Figure 2. The number of emerging pupae of *A. albopictus*, *A. stephensi*, and *C. pipiens* on exposure to 0.15, 0.30, and 0.60 ppm of CuSO₄. *significantly different from control, p<0.05 (Mann-Whitney U test)

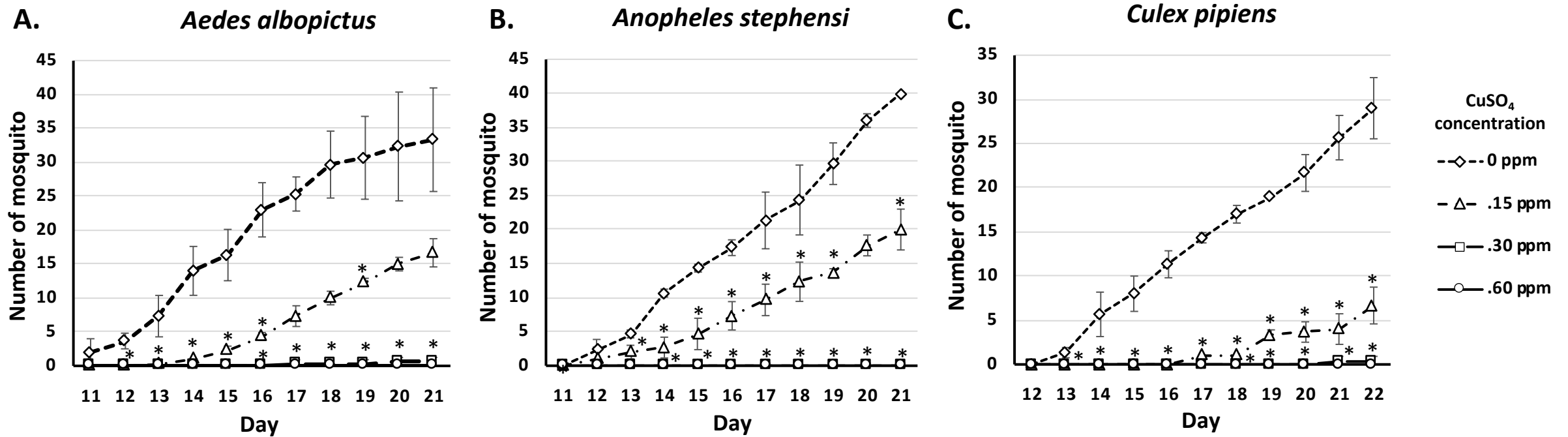


Figure 3. The number of emerging *A. albopictus*, *A. stephensi*, and *C. pipiens* mosquito on exposure to 0.15, 0.30, and 0.60 ppm of CuSO₄. *significantly different from control, p<0.05 (Mann-Whitney U test)