Apparent failure and cryptic success of disease control via intermediate host population
reduction: Density dependence in copepods has implications for Guinea worm disease
transmission

Authors: **Nadia Raytselis[1]**, Maya Risin[1], Layla Steinbock[1], Logan T. Story[1], Jane Miller[1],
Ta'Nyia Heard[1], Kevin Xie[1], Ella Rintala[1], Inesha Gupta[1], Stephanie Gutierrez[1],
Alexander Strauss[2,3,4], David J. Civitello[1]

1. Department of Biology, Emory University, Atlanta, GA, 30322, USA
2. Odum School of Ecology, University of Georgia, Athens, GA, 30602, USA
3. Center for the Ecology of Infectious Diseases, University of Georgia, Athens, GA, 30602
4. River Basin Center, University of Georgia, Athens, GA, 30602

Abstract

Pest control often causes high mortality, reducing population density but enabling recovery when

survivors are freed from intraspecific competition and density dependent demographic rates.

Understanding these demographic responses is critical for effective management, especially

when life stages differ in the harm they cause. We evaluated mortality-based management of

intermediate copepod hosts to control Guinea worm (GW) transmission, combining

mathematical models with a population dynamics experiment designed to mimic current

pesticide control methods across a range of intensities. Despite initial reductions in density,

copepods recovered so rapidly that control appeared to fail, enabled by extremely fast maturation

rates in low-density populations. However, model simulations of GW transmission showed that

although total and stage-specific densities rebounded, infectious adult copepods —the proximate

cause of infections—were strongly suppressed, indicating cryptic success. Ultimately, this study

highlights the importance of understanding mechanisms of density dependence when designing

and optimizing pest control interventions.

Introduction

Elucidating the mechanisms that regulate populations is a fundamental goal of population ecology. Negative feedbacks on population growth emerge because of competition for space (Sebens 1982) and resources (Svanbäck & Bolnick 2007), degradation of the environment (Calizza *et al.* 2017), or other mechanisms that reduce individual performance as density rises (Lu & Wang 2025; Stockseth *et al.* 2025). Reductions in birth rates (Gotelli 2008) and increases in death rates are ubiquitous inhibitors of population growth (Hixon & Jones 2005). In populations with stage structure, negative density dependence can also arise through maturation rates or cannibalism (Park *et al.* 1965; Dennis *et al.* 2001; Walker *et al.* 2021). Despite the reality that all populations experience negative density dependence (lest they increase infinitely), it remains challenging to identify which form(s) operate most strongly for any species and few studies attempt to do so (Kaspersson *et al.* 2013).

The forms of density dependence are particularly relevant for pest control, because interventions are designed to reduce pest density. This reduction is often followed by rapid recovery, driven by relaxed density dependence (Karatayev *et al.* 2015) or potential acquisition of resistance (Cao *et al.* 2025), limiting benefits and necessitating repeated interventions (McIntire & Juliano 2018). The ensuing dynamics of repeated pest suppression and recovery can be highly sensitive to density-dependent vital rates (Smith 1997). For example, strong negative density dependence can cause compensation or overcompensation, the phenomena in which control efforts actually increase the density or biomass of some or all life stages (Karatayev *et al.* 2015; de Roos 2020; Grosholz *et al.* 2021). Even when interventions successfully reduce density, populations can rebound faster than expected if the remaining individuals are released from density-dependent reductions in maturation or birth rates (Karatayev *et al.* 2015). Such responses

may be strongest in stage-structured populations, where removal of more competitive classes, or

reductions in density overall, elicit stage-specific compensatory responses (Karatayev *et al.*

2015). Thus, delineating the form(s) and magnitude of density dependence can improve

management of the ecological issues that pests cause, such as disease transmission, crop losses,

and biodiversity harm.

Here we combine an experiment manipulating pest control intensity with mechanistic

modeling to identify which density-regulating mechanisms affect copepods, the intermediate

hosts of the Guinea worm (GW) parasite, *Dracunculus medinensis*. Copepods exhibit stage-

structured populations (van den Bosch & Gabriel 1994) and experience repeated control

interventions due to their role in GW transmission (Gindola *et al.* 2022). Guinea worm disease

(GWD) is a globally impactful infectious disease, causing disability and negative economic

impacts in endemic areas (Pellegrino *et al.* 2022). Cyclopoid copepods are obligate intermediate

hosts that become infected through consumption of GW larvae and facilitate transmission to

humans and other definitive mammalian hosts (Simonetti *et al.* 2023). GWD has been targeted

for eradication for several decades (World Health Organization 1986), with few cases remaining

across five endemic countries (World Health Organization 2025). In GW-endemic regions,

application of the pesticide Abate™ (active compound: temephos) is a major component of

eradication efforts (Gindola *et al.* 2022). Abate™ can cause >99% immobilization of copepods

in laboratory studies (Grunert *et al.* 2022), although copepod species vary widely in their

sensitivity (Hanazato *et al.* 1989; Leboulanger *et al.* 2011). In Abate™-treated sites, copepod

populations often exhibit immediate reductions in density, but routinely recover, necessitating

application at 28-day intervals to support eradication (Gindola *et al.* 2022). The density-

dependent demographic responses of these intermediate hosts – along with implications for

disease control – are unknown.

Copepod stage structure has implications for both their population dynamics and GW

transmission. Neonatal copepods (nauplii) are too small to consume GW larvae, but larger

juveniles (copepodites) and adults can consume parasites and become infected (McCullough

1983; Bimi 2007). Once ingested, parasites molt twice over 10-16 days within the copepod

before maturing into third stage larvae, the only stage infectious to definitive hosts that imbibe

infected copepods (Gonzalez Engelhard *et al.* 2021). Additionally, adults can cannibalize nauplii

(van den Bosch & Santer 1993). Finally, public health officials have shared anecdotally that

adults and copepodites may be more sensitive to Abate™ than nauplii. Critically, the

consequences of these stage-structured interactions for disease management are unknown.

To understand copepod demographic responses to episodic culling, and to predict

implications for disease control, we conducted a mesocosm experiment that mimicked the 28-

day treatment interval of Abate™ by physically removing copepods (either applied equally

across stages or biased toward larger individuals). We hypothesized that more intense

interventions (higher removal percentages) would cause greater reductions in population density,

but that relaxed density dependence might cause compensation or overcompensation between

interventions. Further, we speculated that these effects would be more pronounced in populations

where both size classes were removed. Finally, we hypothesized that even if copepods

rebounded rapidly ('apparent failure'), risk of parasite transmission might remain low ('cryptic

success') due to high adult turnover rate and the two-week period required for parasite

development from first stage to infectious third stage larvae.

Surprisingly, all interventions failed to reduce mean copepod density over the 14-week

intervention period. Copepods recovered extremely rapidly due to relaxed density dependence on

maturation rates. However, despite this apparent failure, model simulations indicate that

interventions may still reduce transmission risk by specifically suppressing the density of

infectious copepods. These results illustrate how density-dependent mechanisms interact with

transmission biology to drive cryptic success of control. Stage structure is a key feature of the

biology of many pest species, e.g., root-feeding crop pests (Johnson & Rasmann 2015) and

mosquito vectors of human diseases (Neale & Juliano 2019; Walker *et al.* 2021), and control

interventions often focus on specific life stages that are not always the proximate cause of harm,

e.g., larvicides to control pathogens transmitted by adult mosquitoes (Choi *et al.* 2019). Thus,

understanding the underlying mechanisms of stage-structured density dependence could enhance

our ability to address a diverse range of ecological challenges.

Methods

Copepod Maintenance and Propagation

We obtained *Eucyclops sp.* copepods isolated near Mandelia, Chad, an active focus of GW

transmission. We maintained copepods in 0.5x HHCOMBO artificial lake with 1X

micronutrients (hereafter COMBO; Kilham *et al.* 1998) in 1 L beakers at 26C with a 12:12

daylight cycle. We fed them 7.5 mg/L of powdered egg yolk (Judee's, Plain City, Ohio)

suspension every Monday, Wednesday, and Friday (M/W/F), and transferred copepods to new

media approximately every three weeks by sieving. Once densities reached ~500-1,000

copepods/L, we poured one 1 L culture into 42, 15 L propagation tanks with 4 L COMBO.

Mesocosm Setup, Sampling, and Experimental Design

On September 20th, 2024, we filled 60 15 L tanks with 5 L COMBO and added 1.5 mg/L

powdered egg yolk on Monday/Wednesday/Friday to establish microbial communities. On

October 2nd, 2024 (day 0), we introduced copepods to tanks by adding seven 1 L aliquots

collected randomly from the 42 propagation tanks after stirring to homogenize. We then added

COMBO to 15 L total and fed tanks 7.5 mg/L every Monday/Wednesday/Friday throughout the

experiment. We replaced water weekly to counteract evaporation (<1 L/tank/week).

139   We maintained the experiment for 126 days and sampled every Friday starting on day 2.

To sample, we homogenized tanks (stirring with a plastic paddle) and collected a 500 mL surface

grab sample with a beaker. We replaced this volume with COMBO. We intervened at 28-day

intervals (days 8, 36, and 64) to mimic the field use of Abate™ (Gindola *et al.* 2022). We

implemented mechanical removal, rather than Abate™, to enable consistent gradients of size-

biased and size-unbiased removal. We factorially crossed intervention type (equal vs. size-

biased) with the percent of the volume (i.e., copepod population) removed (0-90%: 0, 10, 20, 30,

40, 50, 60, 70, 80, or 90%). For size-biased removal, we sieved water through 100 μm nylon

screen, capturing adults, juveniles, and larger nauplii, and returned the water and smaller nauplii

to tanks. For size-unbiased (i.e., equal) removal, we replaced the volume removed with fresh

COMBO. In Supplemental S7, we provide methodological details and results on the accuracy of

interventions relative to nominal treatment levels. After the first intervention, we converted the

10% and 20% treatments to 95% and 98% for the following interventions, because low initial

intensities caused negligible effects, and higher removal rates better match the stated efficacy of

the Abate™ (Grunert *et al.* 2022). On weeks with interventions, we sampled on Thursday and

intervened on Friday. We "skipped" the intervention scheduled for day 92 to observe responses

to treatment lapse, as can occur in the field due to inaccessibility from flooding.

Copepod Counting

We quantified copepods by sieving the 500 mL samples with 35 μm mesh (which captures all stages) and preserving them in 10 mL ethanol. We gently resuspended the samples and examined successive 1 mL aliquots at 10-25x magnification on a dissecting microscope until counting ≥ 50 copepods or screening the entire sample. We counted copepods in three morphologically discernible categories (nauplii, juveniles and eggless adults, and adult females with eggs).

Statistical Analysis of Intervention Effects on Copepod Density

We conducted all analyses in the R Computing Language v4.3.2 (R Core Team 2023). We tested for differences in mean population density over the first 14 weeks, when treatments were applied "on schedule", using planned contrasts between all interventions versus controls (Dunnett's tests; package emmeans (Lenth 2025). This analysis used a Generalized Linear Model (package glmmTMB; Bolker 2017) with mean density as the response, removal proportion, removal type, and their interaction as predictors, and the Gamma error distribution with a log link function.

State Space Modeling (SSM) of Copepod Population Dynamics

We built a stochastic population dynamic model and simultaneously fit it to all time series of observations from the experiment using the Partially Observed Markov Process (POMP) framework and the panelPomp package (Carles Bretó 2025). We connected likelihood estimates generated by the function pfilter() to an adaptive Markov Chain Monte Carlo (MCMC) sampler in the adaptMCMC package to conduct Bayesian inference, with priors informed by auxiliary experiments (Supplemental S2). POMP models are ideal for population dynamics data (Newman *et al.* 2023) where state variables (the "true" abundances of nauplii, juveniles, and adults) are partially observed as empirical subsample counts of nauplii (N_{obs}), juveniles/adult

males/adult females with no eggs (JOA_{obs} = "juveniles and other adults"), and adult females

(AF_{obs}) (Carles Bretó 2025).

We implemented the model by specifying ordinary differential equations that describe

deterministic rates of change in the densities of nauplii, N, juveniles, J, and adults, A. In this

model (Equations 1-6), nauplii are born, develop into juveniles, die from background causes, and

are cannibalized by adults. Juveniles mature from nauplii, develop into adults, and die of

background causes. Adults mature from juveniles, birth and cannibalize nauplii, and die of

background causes. Critically, birth rate (b), stage-specific death rates (d_i, with $i = N, J, or A$),

and stage-specific maturation rates (m_i, with $i = N or J$) depend exponentially on density, with

each stage potentially contributing differentially to density dependence via their impact on the

competitive environment (i.e., c_J and c_N represent the competitive effects of juveniles and nauplii

relative to adults). We allowed adults to cannibalize nauplii following a linear functional

response with attack rate α. On intervention days, we incorporated removal as additional

mortality on relevant stage(s). During all interventions, we assumed juveniles and adults were

removed according to treatment. We assumed nauplii were removed proportionally in size-

unbiased removal treatments and used a prior informed by an auxiliary experiment that showed

size-biased sieving removed approximately 45% of nauplii (Supplemental S5).

$\frac{dN}{dt} = b(N,J,A)\frac{A}{2} - m_N(N,J,A)N - d_N(N,J,A)N - \alpha NA$ Eq. 1

$\frac{dJ}{dt} = m_N(N,J,A)N - m_J(N,J,A)J - d_J(N,J,A)J$ Eq. 2

$\frac{dA}{dt} = m_J(N,J,A)J - d_A(N,J,A)A$ Eq. 3

with density-dependent functions:

$b(N,J,A) = b_M e^{\left(-\frac{c_b}{V}(c_N N + c_J J + A)\right)}$ Eq. 4

$$201 \qquad m_i(N, J, A) = m_{i0} e^{\left(\frac{-c_m}{V}(C_N N + C_J J + A)\right)} \qquad\qquad \text{Eq. 5}$$

$$202 \qquad d_i(N, J, A) = d_{i0} e^{\left(\frac{c_d}{V}(C_N N + C_J J + A)\right)} \qquad\qquad \text{Eq. 6}$$

204   To simulate the dynamics of the latent (or "true and unobserved") state of the system we

205 assumed demographic stochasticity and implemented the model with a one-day time step.

206 Specifically, we tracked individuals transitioning through stages or dying using random

207 multinomial draws where the probability of three potential outcomes for any individual

208 (remaining in the same state, maturing to the next state, or dying) depends on the relative rates of

209 each process. We assumed births followed Poisson random draws depending on the total birth

210 rate. Our observations differentiate N_{obs}, AF_{obs}, and JOA_{obs}. We assumed that each observation

211 followed a negative binomial distribution with a common overdispersion parameter. For each

212 observation, we assumed the predicted count was determined by the product of the abundance of

213 individuals of that stage at time t, the proportion of the tank sampled (always 0.5/15 L = 3.3%),

214 and the proportion of the 10 mL subsample counted. To align true and observed categories, we

215 assumed a 50:50 sex ratio with female reproduction, with adult females releasing eggs every

216 third day and appearing eggless for one day in each three-day interval (personal observation

217 TDH). With this model specified, we used the pfilter() function in panelPomp to estimate model

218 likelihoods from the data. To maintain variance in log-likelihoods < 1 (Endo *et al.* 2019), we

219 used 20 particles per likelihood estimation, resulting in variances of approximately 0.5. We ran

220 32 MCMC chains, with dispersed initial parameter sets for 250,000 iterations each, permitting

221 adaptation during the first 50,000 iterations (burn-in), which we later discarded.

222 ***Model-derived estimates of density dependence***

We used model-derived functions of density dependence, observed densities, and stage structure

from the experiment to visualize how the competitive environment shaped demographic rates.

We thinned chains to every 1,000[th] iteration, and calculated per-capita birth, death, cannibalism,

and maturation rates across a density gradient from 0 to 6,000 individuals/L for each parameter

set (the density range observed in the experiment). We fixed stage structure along this density

gradient (5.5% adults, 39% juveniles, and 55.5% nauplii, 1:7:10 ratio), based on the average

proportions in the experiment. We then generated the mean and 95% CI interval of posterior

predictions for each demographic rate along the gradient.

Compartmental (ODE) stage structured model with interventions and infection

Large-scale experiments with viable GW larvae are currently infeasible. Therefore, we extended

our model to gain applied insights into disease control. The extended model (Eq. S1-S11 in

Supplemental S1) allows parasites to infect adult copepods. We implemented a distributed

developmental delay (Hurtado & Kirosingh 2019) for copepods transitioning from exposed to

infectious to definitive hosts (Supplemental S1). This approach uses the "linear chain trick"

(Hurtado & Kirosingh 2019) to implement a variable time lag, as we hypothesized this delay

could be important for differentiating successful population regulation from successful disease

control. Once an adult copepod consumes the parasite, it takes approximately 15 days to become

infectious (Lindquist & Cross 2017). Thus, we added a developmental delay of approximately 15

days by implementing a chain of 60 exposed stages that copepods transition through at a rate of

4.3 d^{-1}, yielding a 15.45 ± 1.82-day delay from exposure to infectiousness.

Estimates of the force of infection for copepods are unavailable. Therefore, we assumed a

5% infection prevalence among adults, consistent with the limited field data available (Lyons

1972; Steib & Mayer 1988; Garrett *et al.* 2020), corresponding with λ = 0.0034/day. Further,

while parasites are likely introduced in discrete pulses (a single mature female can introduce 1

million L1 larvae (Greenwood 2012)), we implemented a constant force of infection to avoid

confounding results with day of introduction. Lastly, we assumed infected adults cannot

reproduce (Moorthy 1938).

Using this model, we evaluated the consequences of episodic mortality events on stage-

specific and total abundance, with emphasis on the density of infectious copepods, the proximate

hazard to definitive hosts. We plotted the temporal dynamics for the 90% mortality scenario, a

previously used metric for Abate[TM] success (South Sudan Guinea Worm Eradication Program

2013). We plotted mean densities of all stages across the whole simulation period for scenarios

ranging from 0-90% mortality.

Results

Copepod population dynamics and mortality events

Regardless of removal intensity, we found no significant decreases in mean population density

over the first 14-weeks of the study for size-unbiased (light blue) or size-biased (dark blue)

removal (Fig. 1). This apparent failure was consistent for both the total population and when

broken down by stage (Fig. 1, diamond, triangle, circle; all Dunnett's test $P > 0.05$).

Figure 1. Mean observed copepod densities in the experiment over the 14 weeks in which the

interventions were applied on schedule. Points and error bars represent treatment means ± SEs

(log-transformed). Regardless of treatment type or removal proportion, there were no significant

reductions in total copepod density (squares), or stage-specific densities of egg-bearing adult

females (AF; diamonds), juveniles plus other adults (JOA; triangles), or nauplii (N; circles).

Similar dynamics were observed regardless of whether removal was selective for large-bodied

individuals (size-biased; dark blue) or equally applied across all stages (size-unbiased; light

blue).

While we found no effect on mean density, the interventions did reduce densities

immediately after application (e.g., 95 and 98% removal, Fig. 2, Supplemental S5). However,

populations then rapidly rebounded. Populations in controls started at ~500-1,000 copepods/L,

grew slowly for several weeks, and ended with a boom and bust (Fig. 2). In other cases, density

in culled tanks temporarily surpassed controls, indicating overcompensation. For example,

following the third intervention, densities in the 50% size-biased treatment were briefly higher

than controls. In some cases, population recovery occurred so rapidly that declines were

imperceptible one week later (e.g., after the second intervention with 80% removal). The best fit

of the model generally captured the dynamics in intervention tanks better than that of the control

treatment, which is partially expected, as the control treatment represented only 10% of the

experiment.

Size-biased Removal:

Relative to controls, the 30% and 60% size-biased treatments grew steadily throughout

the entire experiment (Fig. 2, Supplemental S5A). Populations quickly recovered from

interventions, and observed dynamics generally fell within 95% credible intervals of the fitted

model predictions. The 40%, 50%, and 70% removal treatments more closely resembled the

controls, with large fluctuations in density at the end of the experiment. The 80%, 90%, and 95%

removal treatments had multiple extreme booms and busts. Finally, 98% removal treatments

fluctuated with low levels of population growth. These shifts sometimes fell outside the credible

interval of posterior predictions.

Size-unbiased Removal:

In contrast with controls, 30%, 70% and 80% size-unbiased removal treatments

maintained relatively constant abundances across all eighteen weeks, mostly within the credible

interval (Fig. 2, Supplemental S5A). The 40% and 60% tanks resembled the controls. The 50%

treatment also resembled the control; however, instead of final booms and busts, populations

declined steadily. The 90%, 95%, and 98% treatments had sustained peaks and troughs,

encompassed by the credible interval for most of the experiment. The last few weeks had more

extreme booms and busts, not captured within the credible interval.

Figure 2. Observed copepod population dynamics (points and error bars represent means ± SE)

and best-fitting posterior predictions with 95% credible intervals (black lines and shaded grey

regions). Treatments that are representative of the major population dynamics are shown, with all

treatments can be found in the supplemental (Supplemental S5A). Dynamics in control tanks are

plotted in green, size-biased removal tanks are plotted in light blue, and size-unbiased removal

tanks are in dark blue. Dashed vertical lines represent interventions. In general, interventions

caused immediate reductions in copepod abundance, but populations rebounded rapidly. In some

case, e.g., size-biased 50% and size-unbiased 90%, these rebounds surpassed abundances

observed in controls. For the 98% treatments, the first intervention was 20% removal, explaining

the small dip in the model fit.

Evaluating the Functional Forms of Density Dependence

The strength of negative density dependence varied significantly across processes of birth,

maturation, background death, and cannibalism. Density dependence was greatest for maturation

rates, with a >18,000-fold reduction across the observed density gradient (Fig. 3). Density

dependence on background death rate and cannibalism were also substantial, with a combined

4.4-fold increase over the density range (Fig. 3). However, we estimated essentially no effect on

birth rates, with <2% decrease over the density range.

Figure 3. Representation of the sources of negative density dependence affecting copepod

populations subject to control interventions. Lines and shaded regions represent mean and 95%

credible intervals for posterior model predictions for the effects of copepod population density

(with fixed stage structure) on key vital rates: birth rate (green), total death rate of nauplii (dark

blue; incorporating background deaths and cannibalism via adults), and naupliar maturation rate

(light blue). The rug indicates the observations of population density in the experiment. Adult

birth rate was insensitive to competition, declining only 0.9% over the observed density gradient

(indicated by the rug along the x-axis). In contrast, death rate increased 4.4-fold over the density

gradient. Most importantly, maturation rate was extremely sensitive, declining >18,000 fold over

the gradient.

Transmission Scenario Simulation Results

Simulated GW infection scenarios exhibited rapid, stage-specific recovery of copepods

following interventions (Fig. 4). With 90% intervention intensity, nauplii and non-infectious

adults rapidly rebounded to densities that surpassed those observed for untreated populations

(Fig. 4C, D), and juveniles reached similar, though lower densities until the next intervention

(Fig. 4B). Size-unbiased removal generally reduced copepod populations more strongly, but

these populations also showed slightly greater overcompensation (Fig. 4A, C). This transient

overcompensation reiterated a central result from the experiment (Fig. 1): that mean copepod

density appeared unaffected by interventions.

Despite this apparent failure, simulated interventions successfully reduced the density of

infectious copepods (Fig. 4A). Without intervention, control populations began with low

densities of infectious hosts, but these grew steadily (>75 infected copepods L^{-1} by the end of the

100-day simulation). However, both size-unbiased and size-biased suppressed infectious

copepod density below 10 L^{-1} (Fig. 4A). Evaluating mean densities of stages across an

intervention intensity gradient (Fig. 5), total population density and non-infected adults

(susceptible + exposed classes) remained relatively constant at all intensities, while the density of

infectious adults steadily declined across the gradient. Thus, despite observing that copepod

populations rebound rapidly – enabled by intense density dependence on maturation rates – the

density of infectious hosts, the proximate source of mammalian infections, is highly sensitive to

intervention.

Figure 4. Temporal dynamics from the modeled transmission scenarios incorporating 90%

mortality events at 28-day intervals. Dashed vertical lines represent interventions. Copepod

stages are shown in different colors. Stages are divided into infectious adults (I), juveniles (J),

nauplii (N), and non-infectious adults (NI = susceptible + exposed adults). Control tanks are

dashed lines, 90% size-unbiased removal are solid lines, and 90% size-biased removal are the

dotted lines. The force of infection parameter (λ =0.0034) corresponded with a ~5% infection

rate among adults(Lyons 1972; Steib & Mayer 1988; Garrett *et al.* 2020). Nauplii and non-

infected adults surpass densities of controls (green) with both forms of removal (size-biased

[dark blue] and size-unbiased [light blue]). Juveniles nearly recover to densities akin to controls.

Most importantly, infectious copepods are severely depressed by both removal types.

Figure 5: Mean total density of copepod populations (light blue), non-infectious adults (green),

and infectious adults (dark blue) in 120-day transmission scenarios modeled along a gradient of

intervention intensity for size-biased (solid line) and size-unbiased (dashed line) strategies. As in

the experiment, total density and the density of non-infectious adults (susceptible and exposed)

were largely insensitive to interventions, remaining relatively constant despite 90% population

reductions imposed every 28 days. In contrast, the mean density of infectious adults, the

proximate driver of mammalian hazard of infection, decreased almost 10-fold along the same

gradient. Thus, despite the rapid recovery of copepod populations and the appearance of failed

control, intervention can be highly effective at reducing the proximate driver of infection in

humans and other mammals.

Discussion

Uncovering the mechanisms regulating populations is a fundamental goal in population

ecology (Kaspersson *et al.* 2013; Karatayev *et al.* 2015). We combined a pest control experiment

and mechanistic modeling to partition the magnitude and importance of four general putative

mechanisms of density regulation. Then, we integrated these empirically-derived regulating

effects with a transmission model to evaluate how relaxation of density dependence via realistic

control methods shapes effectiveness of interventions for GW, a target for global parasite

eradication (Pellegrino *et al.* 2022).

Our experiment investigated how copepod populations – intermediate hosts of GWD –

responded to periodic culling. We designed treatments to mimic the treatment of water sources

with Abate™, an intervention commonly used to prevent GWD in endemic countries. We

hypothesized that more aggressive control efforts would better suppress copepod populations,

but that relaxed density dependence might lead to overcompensation. Strikingly, even when we

episodically removed up to 98% of the population, densities across were statistically

indistinguishable from controls (Fig. 1, Supplemental S3). Similar rebounds have been observed

in other copepod species, including *Mesocyclops leuckarti* which overshot pre-treatment

densities following application of Triphenyltin, a fungicide (Kulkarni *et al.* 2014). Our

interventions had failed to reduce copepod densities, an alarming prospect as other apparent pest

control failures have posed immense challenges for public health and conservation practitioners

(Rivero *et al.* 2010; Neale & Juliano 2019; Grosholz *et al.* 2021). For example, populations of

mosquitoes, sand flies, assassin bugs, and fleas, which are routinely treated to control diseases

such as malaria, leishmania, chagas, and plague, respectively, show evidence of insecticide

resistance, threatening vector control (Cao *et al.* 2025). Further, specific mosquito species

including *Aedes* and *Culex sp.*, vectors of diseases such as dengue, chikungunya, Zika virus

(*Aedes*) (Patterson *et al.* 2016), and West Nile Virus (*Culex*) (Hamer *et al.* 2008), show evidence

of culling-induced overcompensation, indicating that control strategies must be careful to not

impose mortality at levels that lead to overcompensatory releases from density dependence

(Neale & Juliano 2019). A striking example of a pest control program that backfired is the effort

to eliminate the invasive European green crab in an estuary off of California (Grosholz *et al.*

2021). Culling of green crabs was an effective strategy for years until the population density was

reduced below a critical threshold, triggering stage-specific overcompensation in recruitment

(Grosholz *et al.* 2021). The result was a boom in juvenile crabs, a shift in size structure, and

exacerbation of negative impacts (Grosholz *et al.* 2021). Together, these examples underscore

the challenges of pest control – especially in light of stage structure.

To attempt to further explain the surprisingly fast recovery we observed, we investigated

the forms of density dependence regulating *Eucyclops sp.* populations. Few studies have parsed

specific drivers of density dependence in natural populations. In one example, stocking large

brown trout reduced the growth rate of smaller individuals (Kaspersson *et al.* 2013). In another,

dynamics of Swedish vole populations were consistent with density dependence in recruitment

alone (Lambin *et al.* 2025). Stage-specific density dependence in maturation rates has been

documented in some ecological systems (Karatayev *et al.* 2015; Walker *et al.* 2021), including

other copepods (Kulkarni *et al.* 2014), allowing for population persistence under significant

mortality (Karatayev *et al.* 2015). We determined that density dependence in *Eucyclops sp.* was

driven predominantly by maturation rates, with a >18,000-fold reduction along the density

gradient, with a modest ~4.4-fold increase in total mortality, and no change in birth rate (Fig. 3).

Thus, when interventions occur, surviving adults maintain birth rates, but nauplii mature

extremely quickly. Consequently, higher proportions of nauplii and juveniles survive to

development, enabling rapid recovery and temporary overcompensation. (Figs. 2 and 4).

For the goal of GWD management, the failure to reduce mean copepod population

densities seems alarming. However, not all copepod life stages host GW, (McCullough 1983;

Bimi 2007) and there is a significant delay between copepod exposure to GW and infectiousness

to mammals (~15 days; Gonzalez Engelhard *et al.* 2021). Thus, we speculated that if population

rebounds were driven by overcompensation in young stages that cannot transmit GW, then

apparent failures to control the overall population might actually conceal successes. We

evaluated this possibility by extending our model to include GW infection of adults with a

distributed delay from the exposed to infectious class. We tracked changes in the density and

stage structure of copepod populations across a gradient of intervention intensity.

When we simulated GW transmission scenarios, we found strong support for stage-

specific recovery dynamics, as documented in other zooplankton systems (Nilsson *et al.* 2010;

Kulkarni *et al.* 2014). Specifically, the densities of nauplii, juveniles, and non-infectious adults

all recovered rapidly to meet or exceed pre-intervention densities following each successive

mortality event (Fig. 4), indicating compensation or overcompensation (Abrams 2009; Kulkarni

et al. 2014; de Roos 2020; Grosholz *et al.* 2021). Critical to GW transmission, the density of

infectious adults remained extremely low. Thus, despite the apparent failure of these

interventions to control copepod populations, they appear highly effective at specifically

controlling the life stage that is the proximate cause of GW infections. We attribute this effect to

the approximately15-day developmental period from first stage to infectious third stage GW

larvae (Lindquist & Cross 2017; Gonzalez Engelhard *et al.* 2021), which is long enough to

prevent accumulation of infectious copepods before each successive intervention (Fig 4., Fig. 5).

Similar lags of parasite development in intermediate hosts and vectors are present in other

systems. For example, in Anopheline mosquitoes, a 2-3 week incubation period separates vector

exposure to *Plasmodium* and infectiousness to humans (Crutcher JM 1996). Similarly,

Trypanosoma maturation in tsetse flies requires approximately three weeks (Telleria *et al.* 2014).

Thus, reducing the overall density of intermediate hosts or vectors may not be essential for

success if control can cause high turnover in the stage(s) responsible for transmission.

Our results raise the importance of field surveillance targeted at infected copepods. At

present, diagnosing infections in copepods from field is infeasible due to GW rarity and the scale

of geographic surveillance (Pellegrino *et al.* 2022). Therefore, current eradication programs rely

on monitoring the total density of copepods before and after Abate^{TM} applications. If copepod

communities in the field recover as rapidly as the *Eucyclops sp.* examined in our study, officials

might falsely conclude that control is ineffective, because population densities return to pre-

treatment levels every month. However, our experiment and model suggest that this observation

could conceal a cryptic success: reduction in the density of infectious copepods. Thus, we

recommend the development of tools to facilitate large-scale spatial and temporal diagnostics of

GW-infected copepods to evaluate the relevance of this counterintuitive finding.

We note some important caveats to translating these insights to the field. First, we

focused on a single copepod isolate. Greater understanding of copepod diversity in demographic

rates and sensitivity to Abate^{TM} would greatly benefit eradication programs. Second, we

implemented mechanical removal, which enabled a precise experimental design, but can differ

from chemical control. Experiments with Abate^{TM} are needed to evaluate chemical persistence,

delayed or sublethal effects, and the consequences of over- or underdosing. Lastly, our analysis

identified high importance for negative density dependence on maturation rates and mortality,

and no effect on birth rates. Generally, the model fit the data well for the first 70-80 days of the

experiment, albeit with some initial lags in population growth (Fig. 2). This imperfect fit could

be due to missing key processes in the model, such as an Allee effect, where mating failure

occurs at low population densities (seen for copepods (Kramer *et al.* 2008)). The inability of the

model to precisely recover empirical dynamics could also be due to nonconstant conditions in the

tanks (e.g., the accumulation of nutrients (Sahandi *et al.* 2023) and/or waste products or

differences in temperature or photoperiod over the course of the experiment). Nevertheless, the

POMP approach has major benefits, including representation of constraints in the observation

process and incorporation of process noise. Lastly, future studies of copepod control and

recovery could consider other functional forms for key demographic processes, including Type-

II or Type-III functional responses for cannibalism.

Chemical control of copepod populations represents a critical intervention for national

GW eradication programs (Gindola *et al.* 2022). This study highlights that nuance may be

needed to evaluate this intervention: population dynamics may suggest that control efforts are

failing or even backfiring, but direct analyses of infectious copepods, the proximate cause of

human cases and animal infections should be prioritized. Future laboratory studies should

expand to greater copepod diversity and directly evaluate pesticide formulations in use.

Ultimately, a deeper understanding of copepod control may improve the portfolio of GWD

interventions and contribute to the eradication of this devastating parasite.

485   More broadly, pest control is inherently designed to reduce population density (Ni *et al.*

486 2025), so understanding density dependence is critical to predicting responses (McIntire &

487 Juliano 2018). This work underscores the value of defining which life stages of pests are most

488 impactful, elucidating how stage-structured pest populations respond to interventions, and

489 developing a mechanistic understanding of density dependent mechanisms regulating

490 populations under control, especially in cases where focal populations experience drastic and

491 rapid shifts in density. Together, this information can be applied to anticipate and explain

492 potentially counterintuitive consequences of interventions. A thorough understanding of these

493 dynamics is critical for forecasting long-term population trajectories and their sensitivity to

494 density-dependent perturbations, paramount to predicting the influence of environmental and

495 ecological stressors on population dynamics.

497 **Acknowledgements**

498 This work was supported by funding from The Carter Center. We thank the Programme National

499 d'Eradication du Ver de Guinée - Tchad for collecting and providing the Chadian *Eucyclops*

500 *sp.* copepods used in our study, and the Yen lab at the Georgia Institute of Technology for

501 providing the culture to us. We thank The Carter Center and other Civitello lab members for

502 feedback on this manuscript. We also thank M. Hoogshagen and J. Raytselis for helping with

503 sampling over the holidays, and B. Lukubye for moral support.

Literature Cited

1.

Abrams, P.A. (2009). When does greater mortality increase population size? The long history and diverse mechanisms underlying the hydra effect. *Ecology Letters*, 12, 462-474.

2.

Bimi, L. (2007). Potential vector species of Guinea worm (Dracunculus medinensis) in Northern Ghana. *Vector Borne Zoonotic Dis*, 7, 324-329.

3.

Bolker, B. (2017). {glmmTMB} Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling.

4.

Calizza, E., Costantini, M.L., Careddu, G. & Rossi, L. (2017). Effect of habitat degradation on competition, carrying capacity, and species assemblage stability. *Ecology and Evolution*, 7, 5784-5796.

5.

Cao, L.J., Chen, J.C., Thia, J.A., Schmidt, T.L., Ffrench-Constant, R., Zhang, L.X. *et al.* (2025). Recurrent mutations drive the rapid evolution of pesticide resistance in the two-spotted spider mite Tetranychus urticae. *Elife*, 14.

6.

Carles Bretó, J.W., Aaron A. King, and Edward L. Ionides (2025). panelPomp: Analysis of Panel Data via Partially Observed Markov Processes in R. *The R Journal*.

7.

Choi, L., Majambere, S. & Wilson, A.L. (2019). Larviciding to prevent malaria transmission. *Cochrane Database Syst Rev*, 8, Cd012736.

8.

Crutcher JM, H.S. (1996). *Malaria*, Medical Microbiology.

9.

de Roos, A. (2020). The impact of population structure on population and community dynamics. pp. 53-73.

10.

Dennis, B., Desharnais, R.A., Cushing, J.M., Henson, S.M. & Costantino, R.F. (2001). Estimating Chaos and Complex Dynamics in Insect Populations *Ecological Monographs*, 71, 277-303.

11.

Endo, A., van Leeuwen, E. & Baguelin, M. (2019). Introduction to particle Markov-chain Monte Carlo for disease dynamics modellers. *Epidemics*, 29, 100363.

12.

Garrett, K.B., Box, E.K., Cleveland, C.A., Majewska, A.A. & Yabsley, M.J. (2020). Dogs and the classic route of Guinea Worm transmission: an evaluation of copepod ingestion. *Sci Rep*, 10, 1430.

13.

Gindola, Y., Getahun, D., Mohammed, K., Kamau, E., Camara, B., Wossen, M. *et al.* (2022). Abate application practices in the Guinea worm endemic region of Gambella, Ethiopia: identification of elimination gaps. *The Journal of Infection in Developing Countries*, 16, 20S-25S.

14.

Gonzalez Engelhard, C.A., Hodgkins, A.P., Pearl, E.E., Spears, P.K., Rychtář, J. & Taylor, D. (2021). A mathematical model of Guinea worm disease in Chad with fish as intermediate transport hosts. *Journal of Theoretical Biology*, 521, 110683.

15.

Gotelli, N. (2008). *A Primer of Ecology* Fourth edn.

16.

Greenwood, D. (2012). 63 - Helminths: Intestinal worm infections; filariasis; schistosomiasis; hydatid disease. In: *Medical Microbiology (Eighteenth Edition)* (eds. Greenwood, D, Barer, M, Slack, R & Irving, W). Churchill Livingstone Edinburgh, pp. 655-666.

17.

Grosholz, E., Ashton, G., Bradley, M., Brown, C., Ceballos-Osuna, L., Chang, A. *et al.* (2021). Stage-specific overcompensation, the hydra effect, and the failure to eradicate an invasive predator. *Proceedings of the National Academy of Sciences*, 118, e2003955118.

18.

Grunert, R., Box, E., Garrett, K., Yabsley, M. & Cleveland, C. (2022). Effects of Temephos (Abate®), Spinosad (Natular®), and Diflubenzuron on the Survival of Cyclopoid Copepods. *Am J Trop Med Hyg*, 106, 818-822.

19.

Hamer, G.L., Kitron, U.D., Brawn, J.D., Loss, S.R., Ruiz, M.O., Goldberg, T.L. *et al.* (2008). Culex pipiens (Diptera: Culicidae): A Bridge Vector of West Nile Virus to Humans. *Journal of Medical Entomology*, 45, 125-128.

20.

Hanazato, T., Iwakuma, T., Yasuno, M. & Sakamoto, M. (1989). Effects of temephos on zooplankton communities in enclosures in a shallow eutrophic lake. *Environmental Pollution*, 59, 305-314.

21.

Hixon, M.A. & Jones, G.P. (2005). Competition, Predation, and Density-Dependent Mortality in Demersal Marine Fishes *Ecology*, 86, 2847-2859.

22.

Hurtado, P.J. & Kirosingh, A.S. (2019). Generalizations of the 'Linear Chain Trick': incorporating more flexible dwell time distributions into mean field ODE models. *J Math Biol*, 79, 1831-1883.

586   23.

587   Johnson, S.N. & Rasmann, S. (2015). Root-feeding insects and their interactions with organisms
588   in the rhizosphere. *Annu Rev Entomol*, 60, 517-535.
589   24.

590   Karatayev, V.A., Kraft, C.E. & Zipkin, E.F. (2015). Racing through life: maturation rate
591   plasticity regulates overcompensation and increases persistence. *Ecosphere*, 6, art203.
592   25.

593   Kaspersson, R., Sundström, F., Bohlin, T. & Johnsson, J.I. (2013). Modes of Competition:
594   Adding and Removing Brown Trout in the Wild to Understand the Mechanisms of
595   Density-Dependence. *PLOS ONE*, 8, e62517.
596   26.

597   Kilham, S., Kreeger, D., Lynn, S., Goulden, C. & Herrera, L. (1998). COMBO: A defined
598   freshwater culture medium for algae and zooplankton. *Hydrobiologia*, 377, 147-159.
599   27.

600   Kramer, A.M., Sarnelle, O. & Knapp, R.A. (2008). Allee Effect Limits Colonization Success of
601   Sexually Reproducing Zooplankton. *Ecology*, 89, 2760-2769.
602   28.

603   Kulkarni, D., Hommen, U., Schäffer, A. & Preuss, T.G. (2014). Ecological interactions affecting
604   population-level responses to chemical stress in Mesocyclops leuckarti. *Chemosphere*,
605   112, 340-347.
606   29.

607   Lambin, X., Begon, M., Burthe, S.J., Graham, I.M., MacKinnon, J.L., Telfer, S. *et al.* (2025).
608   Density-dependent recruitment but not survival drives cyclic dynamics in a field vole
609   population. *Proceedings of the National Academy of Sciences*, 122, e2509516122.
610   30.

611   Leboulanger, C., Schwartz, C., Somville, P., Diallo, A.O. & Pagano, M. (2011). Sensitivity of
612   Two Mesocyclops (Crustacea, Copepoda, Cyclopidae), from Tropical and Temperate
613   Origins, to the Herbicides, Diuron and Paraquat, and the Insecticides, Temephos and
614   Fenitrothion. *Bulletin of Environmental Contamination and Toxicology*, 87, 487-493.
615   31.

616   Lenth, R.V. (2025). emmeans: Estimated Marginal Means, aka Least-Squares Means.
617   32.

618   Lindquist, H.D.A. & Cross, J.H. (2017). 195 - Helminths. In: *Infectious Diseases (Fourth
619   Edition)* (eds. Cohen, J, Powderly, WG & Opal, SM). Elsevier, pp. 1763-1779.e1761.
620   33.

621   Lu, A. & Wang, L. (2025). Hygiene conditions explain larval density-dependent performance in
622   Plutella xylostella with sufficient food. *Journal of Asia-Pacific Entomology*, 28, 102444.
623   34.

624   Lyons, G.R. (1972). Guineaworm infection in the Wa district of north-western Ghana. *Bull
625   World Health Organ*, 47, 601-610.

35.

McCullough, F.S. (1983). Cyclopoid copepods: their role in the transmission and control of
dracunculiasis. (ed. Press, NA). World Health Organization, Geneva Ecology and
Control of Vectors Unit, Division of Vector Biology and Control.
36.

McIntire, K.M. & Juliano, S.A. (2018). How can mortality increase population size? A test of
two mechanistic hypotheses. *Ecology*, 99, 1660-1670.
37.

Moorthy, V.N. (1938). Observations on the development of Dracunculus medinensis larvae in
Cyclops. *American Journal of Hygiene*.
38.

Neale, Z.R. & Juliano, S.A. (2019). Finding the sweet spot: What levels of larval mortality lead
to compensation or overcompensation in adult production? *Ecosphere*, 10, e02855.
39.

Newman, K., King, R., Elvira, V., de Valpine, P., McCrea, Rachel S. & Morgan, B.J.T. (2023).
State-space models for ecological time-series data: Practical model-fitting. *Methods in
Ecology and Evolution*, 14, 26-42.
40.

Ni, J., Wang, J., Zhang, W., Chen, E., Gao, Y., Sun, J. *et al.* (2025). Sustainable control and
integrated management through a one health approach to mitigate vector-borne disease.
One Health, 20, 101018.
41.

Nilsson, K.A., Persson, L. & Van Kooten, T. (2010). Complete compensation in Daphnia
fecundity and stage-specific biomass in response to size-independent mortality. *Journal
of Animal Ecology*, 79, 871-878.
42.

Park, T., Mertz, D.B., Grodzinski, W. & Prus, T. (1965). Cannibalistic Predation in Populations
of Flour Beetles. *Physiological Zoology*, 38, 289-321.
43.

Patterson, J., Sammon, M. & Garg, M. (2016). Dengue, Zika and Chikungunya: Emerging
Arboviruses in the New World. *West J Emerg Med*, 17, 671-679.
44.

Pellegrino, C., Patti, G., Camporeale, M., Belati, A., Novara, R., Papagni, R. *et al.* (2022).
Guinea Worm Disease: A Neglected Diseases on the Verge of Eradication. *Trop Med
Infect Dis*, 7.
45.

R Core Team (2023). R: A Language and Environment for Statistical Computing.
46.

Rivero, A., Vézilier, J., Weill, M., Read, A.F. & Gandon, S. (2010). Insecticide Control of Vector-Borne Diseases: When Is Insecticide Resistance a Problem? *PLOS Pathogens*, 6, e1001000.

47.

Sahandi, J., Sorgeloos, P., Xiao, H., Mu, F., Wang, X., Qi, Z. *et al.* (2023). The use of microbes in culture of harpacticoid copepod (Tigriopus japonicus Mori) for suppression of Vibrio and enhancement of population growth, enzyme activity and microbial longevity. *Aquaculture*, 563, 739008.

48.

Sebens, K.P. (1982). Competition for Space: Growth Rate, Reproductive Output, and Escape in Size. *The American Naturalist*, 120, 189-197.

49.

Simonetti, O., Zerbato, V., Maurel, C., Cosimi, L., Babich, S., Cavalli, F. *et al.* (2023). The current state of knowledge on dracunculiasis: a narrative review of a rare neglected disease. *Infez Med*, 31, 500-508.

50.

Smith, G.C. (1997). An analysis of the form of density dependence in a simulation model of a seasonal breeder undergoing control. *Ecological Modelling*, 95, 181-189.

51.

South Sudan Guinea Worm Eradication Program (2013). Management and Technical Guidelines. (ed. Sudan, MoH-RoS).

52.

Steib, K. & Mayer, P. (1988). Epidemiology and vectors of Dracunculus medinensis in northwest Burkina Faso, West Africa. *Annals of Tropical Medicine & Parasitology*, 82, 189-199.

53.

Stockseth, L., Neale, Z. & Rudolf, V.H.W. (2025). Strengthening of negative density dependence mediates population decline at high temperatures. *Ecology*, 106, e70030.

54.

Svanbäck, R. & Bolnick, D.I. (2007). Intraspecific competition drives increased resource use diversity within a natural population. *Proceedings of the Royal Society B: Biological Sciences*, 274, 839-844.

55.

Telleria, E.L., Benoit, J.B., Zhao, X., Savage, A.F., Regmi, S., e Silva, T.L.A. *et al.* (2014). Insights into the Trypanosome-Host Interactions Revealed through Transcriptomic Analysis of Parasitized Tsetse Fly Salivary Glands. *PLOS Neglected Tropical Diseases*, 8, e2649.

56.

van den Bosch, F. & Gabriel, W. (1994). A model of growth and development in copepods. *Limnology and Oceanography*, 39, 1528-1542.

57.

van den Bosch, F. & Santer, B. (1993). Cannibalism in Cyclops abyssorum. *Oikos*, 67, 19-28.
58.

Walker, M., Robert, M.A. & Childs, L.M. (2021). The importance of density dependence in
juvenile mosquito development and survival: A model-based investigation. *Ecological*
Modelling, 440, 109357.
59.

World Health Organization (1986). Decisions and list of resolutions. World Health Assembly.
60.

World Health Organization (2025). Dracunculiasis (Guinea-worm disease).

713