Improvement in Orchard Nutrient Status and Productivity Through Integrated Nutrient Management in Mango (*Mangifera indica* L.) Under Medium-Density Planting

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Graphical abstract

Abstract

Balanced nutrient management is essential for sustaining productivity and soil health in perennial fruit crops. A two-year field study (2022-2024) was conducted to evaluate the effects of integrating soil and foliar micronutrient applications with reduced recommended doses of fertilizers (RDF) on vegetative growth, nutrient status, yield, fruit quality, and economic performance of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting. Nine nutrient modules combining 25-100% RDF with soil- and foliar-applied Fe, Zn, B, Cu, and Ca were compared using ANOVA, PCA, and PLS-SEM. Integrated treatments significantly enhanced canopy height, spread, and volume, with the 75% RDF + two foliar micronutrient sprays producing the greatest vegetative growth. Yield attributes, fruit size, retention, and shelf life improved markedly under micronutrient-enriched treatments. The highest fruit yield and benefit-cost ratio was obtained with 75% RDF + two foliar sprays, outperforming the conventional 100% RDF. Soil pH and EC were

moderated, and organic carbon improved slightly under integrated nutrient management. Soil and leaf macro- and micronutrient concentrations were consistently higher with combined soil-foliar approaches. PCA clustered high-performing nutrient modules with yield and canopy variables, while PLS-SEM identified plant nutrient status as the strongest predictor of fruit yield. Overall, integrating micronutrient supplementation with 75% RDF improved nutrient-use efficiency, productivity, and economic returns while reducing dependence on full-dose fertilizers. The study highlights 75% RDF + foliar micronutrient sprays as an effective and sustainable nutrient management strategy for enhancing yield and orchard performance in mango.

Keywords: Mango; Micronutrients; Foliar spray; Integrated nutrient management; PCA; PLS-SEM; Nutrient-use efficiency; Canopy growth; Yield; Soil fertility

1. Introduction

Mango (*Mangifera indica* L.) is a major perennial fruit crop of the tropics and subtropics and the most widely cultivated species of the family Anacardiaceae. Its domestication in the Indian subcontinent dates back more than four millennia, reflecting its long-standing association with human societies (Kaur 2017; Pandey et al. 2025). Believed to have originated in the Indo-Burma region, mango exhibits broad adaptability to diverse agroclimatic conditions and thrives across a range of soil types, temperatures, and rainfall regimes (Singh et al. 2016; Nasron et al. 2021). Cytologically, M. indica is diploid (2n = 40), and the genus Mangifera comprises over 26 related species distributed mainly in Southeast Asia, several of which bear edible fruits (Yang et al. 2022; Hussain et al. 2021). Recognized globally as the "King of Fruits," mango is valued for its distinctive aroma, rich biochemical composition, and balanced flavor (Maldonado-Celis et al. 2019; Peng et al. 2022). Mango fruits are nutritionally essential, providing vitamin A ($\approx$4800 IU 100 g^{-1}), vitamin C, B-complex vitamins, malic acid, proteins, sugars, fats, and important minerals such as calcium and iron (Lebaka et al. 2021; Hassan et al. 2022, Lingwan & Yadav 2025). India cultivates mango on nearly 2.4 million hectares, producing about 22.7 million tonnes annually, with an average productivity of 9.5 t ha^{-1} (NHB 2024-2025). However, productivity remains below potential due to suboptimal orchard management and soil fertility constraints.

Traditional orchards established with seedling rootstocks and wide spacing have low tree density and limited productivity. Increasing land pressure, urbanization, and resource constraints have accelerated the adoption of medium- and high-density planting systems, which improve land-use efficiency and yield potential (Hahn et al. 2022; Lingwan et al., 2023). Concurrent advances in canopy management, fertigation, and irrigation scheduling have further supported the expansion of these intensive systems (Tripathi et al. 2020; Singh et al. 2001). These approaches enhance productivity per unit land and water and provide viable options for sustaining fruit production under changing climatic and resource scenarios (Dalvi et al. 2010; Singh et al. 2012, Lingwan et al., 2023). Under intensified planting, efficient nutrient management becomes critical for maintaining orchard productivity and long-term soil health. Perennial fruit trees continuously remove substantial quantities of nutrients through vegetative growth and fruit harvest, necessitating regular replenishment to prevent soil fertility decline (Lather et al. 2021; Rezende et al. 2024, Koley et al.,2025). Although inorganic fertilizers are widely used due to their rapid availability, imbalanced or excessive application, particularly of NPK, can reduce nutrient-use efficiency and induce soil nutrient imbalances (Xin et al. 2025; Chen et al. 2022). Long-term fertilizer use can alter soil chemical, physical, and biological properties; nitrogen may cause acidification, while phosphorus can undergo fixation, limiting the availability of associated nutrients (Wang et al. 2022; Khan et al. 2025, Yadav et al.,2019a). Over time, disparities between nutrient addition and removal can result in depletion of secondary and micronutrients (Jones et al. 2013, Yadav et al., 2019b), thereby reducing soil health and increasing environmental risks (Vitousek et al. 2009, Jain et al.,2025). Most long-term nutrient studies have focused on annual crops, while perennial orchard systems characterized by deeper root systems and internal nutrient recycling remain comparatively understudied (Jayasinghege et al. 2024, Lingwan et al.,2025). In mango, although several investigations have reported the effects of NPK and micronutrient supplementation, these studies are generally short-term, emphasize yield alone, and often overlook the interactions among soil fertility, nutrient dynamics, and economic performance (Ravikiran 2018; Srivastava and Hu 2019; Ahmad et al. 2018; Chaudhari and Singh 2019, Shagun et al., 2024). Research on nutrient behavior in medium-density orchards is particularly limited, and advanced multivariate approaches capable of unraveling complex soil-plant-yield relationships remain largely unexplored. To address these gaps, the present study evaluates the long-term effects of integrating reduced recommended fertilizer doses (RDF) with soil and foliar micronutrient

applications in a medium-density mango orchard. The study examines impacts on soil fertility, plant nutrient status, yield, and economic returns. Principal Component Analysis (PCA) and Partial Least Squares Structural Equation Modeling (PLS-SEM) were employed to analyze the causal relationships between nutrient pools and productivity. We hypothesize that integrating reduced RDF with targeted micronutrient supplementation will enhance nutrient-use efficiency, improve soil fertility, and increase yield and profitability, and that multivariate modeling will reveal positive causal pathways linking optimized nutrient pools with improved productivity in mango orchards.

2. Materials and Methods

2.1 Orchard site and experimental period

The study was conducted during 2023-2024 at the Horticulture Research Centre, Patharchatta, GBPUA&T, Pantnagar, Uttarakhand ($\approx$29.5°N, 79.3°E; 244 m a.s.l.; **Fig.1**). The experiment used a medium-density mango (*Mangifera indica* L. cv. Dashehari) orchard of uniform 13-year-old trees planted at 5×5 m spacing (400 trees ha^{-1}; **Fig.2**).

2.2 Soil and climate

The site lies in the Tarai agro-ecological zone with a sub-humid subtropical climate. The experimental soil is silty clay loam (Patharchatta Series II, Mollisol). Baseline soil physical-chemical properties (pH, EC, organic carbon, available N, P, K, and extractable micronutrients) were determined before the start of the experiment (**Table 1 for initial status**).

2.3 Experimental design and treatments

The experiment was laid out in a randomized block design (RBD) with 10 treatments and three replications. Treatments evaluated combinations of full, reduced (75%, 50%, 25%) recommended fertilizer dose (RDF = 1000 g N + 750 g P_2O_5 + 1000 g K_2O + 50 kg FYM tree^{-1} year^{-1}) and soil-applied micronutrients (Fe, Cu, Zn each 100 g tree^{-1} and B 50 g tree^{-1}), together with one or two foliar sprays of micronutrients. In brief treatment details:

- T_1 (Control): 100% RDF applied in the basin after harvest.

- T_2-T_4: 75%, 50%, 25% RDF, respectively, + soil micronutrients (Fe, Cu, Zn @100 g each + B @50 g tree⁻¹) + one foliar spray (Fe, Ca, Zn each 0.50% + B- 0.10%) (just before flowering and marble stages as a single event).

- T_5-T_7: 75%, 50%, 25% RDF, respectively, + soil micronutrients (as above) + two foliar sprays (same foliar composition; applied just before flowering and at marble stage).

- T_8-T_{10}: 75%, 50%, 25% RDF, respectively, without soil micronutrients but with two foliar sprays (just before flowering and at marble stage).

(Note: Full treatment details are summarized in Table X: Supplementary materials)

2.4 Orchard management and fertilizer application

Basins (radius: 1.5 m from trunk) were prepared around each tree. Farmyard manure was applied in October. P, K, and micronutrient basal doses were incorporated into the basins in November. Nitrogen was applied in two equal splits: first immediately after flowering and the second at the mustard-to-pea stage of fruit development. Foliar sprays (10 L water tree⁻¹ per spray) were prepared by dissolving micronutrient salts and applied with a tractor sprayer at the timings (just before flowering and at marble stage). Standard cultural practices (irrigation, weeding, pruning, pest and disease control) were followed uniformly across treatments. Conceptual experimental workflow for the integrated nutrient management study in medium-density mango orchards is presented in **(Fig.3),** and the methods and timings of nutrient application are illustrated in (**Fig. 4).**

2.5 Sampling and laboratory analysis

2.5.1 Soil sampling

Soil samples were collected from each replication after harvest in both years from the 0-30 cm depth using a post-hole auger. Samples were air-dried, sieved (<2 mm), and stored for analysis. Soil pH and electrical conductivity (EC) were determined in 1:2.5 soil: water extracts. Organic carbon was estimated by Walkley-Black wet oxidation. Available N was measured by the alkaline permanganate method; available P by Olsen's bicarbonate extraction and colorimetry; available K by ammonium acetate extraction and flame photometry. DTPA extraction (pH 7.3) was used for available Fe, Zn, and Cu, and concentrations were measured by atomic absorption

spectrophotometry (AAS) (Pant et al., 2023). Boron was estimated by the hot CaCl□ extractable method and quantified spectrophotometrically. An overview of soil sample collection (0-30 cm depth) is presented in **Fig. 5.**

2.5.2 Plant sampling

Mature (4-5-month-old) fully expanded leaves were collected after harvest from each tree for nutrient analysis **Fig. 5**. Leaves were washed, oven-dried at 65 ± 1 °C to constant weight, ground, and sieved (<2 mm). Plant tissue digestion was performed using di-acid (HNO□:HClO□) digestion. Total N was determined by the Kjeldahl method, P by colorimetry (vanadate–molybdate), K by flame photometry, and Fe, Zn, Cu, and Ca by AAS. Boron in plant tissue was determined after dry-ashing by the Azomethine-H method.

2.6 Growth, yield, quality, and benefit: cost ratio measurements

Tree height (from first branch to apex), canopy spread (N-S and E-W), canopy volume (Westwood & Roberts, 1970), and their percentage increases were recorded at the start and end of the experimental period. Fruit set, fruit drop, and fruit retention were recorded on tagged panicles. At harvest, fruit number per tree and mean fruit weight (mean of 20-25 fruits per tree) were used to calculate yield per tree (kg tree□¹) and extrapolated to t ha□¹. Yield efficiency was calculated as yield per unit canopy volume (kg m□³). Fruit dimensions (length, width) and fruit weight were measured on representative samples using digital calipers and a balance. Shelf life was assessed at ambient temperature (25 ± 2 °C) by recording days until acceptable edible ripeness. The B: C ratio was determined to assess the economic efficiency of each treatment, showing the returns per rupee invested.

2.7 Multivariate analyses and statistics

Data for two consecutive years were pooled after testing for homogeneity of error variance. Treatment effects were tested by analysis of variance (ANOVA) appropriate for an RBD; when significant, means were separated using Tukey's HSD at $P \leq 0.05$. The Persian correlation coefficient was calculated to quantify the strength and direction of associations among soil, plant, and yield variables, providing a robust non-parametric measure suitable for non-normal or heterogeneous datasets. Principal Component Analysis (PCA) was used to identify major patterns among soil, plant, and yield variables (all variables standardized before analysis) (Shree et al.,

178   2019; Lingwan et al., 2022). Partial Least Squares Structural Equation Modeling (PLS-SEM) was

179   performed to evaluate causal pathways among latent constructs representing Plant Growth, Plant

180   Nutrients, Soil Nutrients, and Fruit Yield. PCA was run in OriginPro; PLS-SEM and path

181   modelling were implemented in SmartPLS (version 4.1.1.5). Model quality was assessed using

182   standard metrics (R^2, composite reliability, AVE, discriminant validity, VIF, and SRMR).

3. Results

3.1. Vegetative growth

Canopy growth parameters of mango cv. Dashehari responded significantly to nutrient

management treatments during 2022-2023 and 2023-2024 **(Table 2).** Canopy height, spread and

volume varied markedly across treatments. The treatment receiving 75% RDF with two foliar

micronutrient sprays recorded the maximum canopy height, whereas the 100% RDF treatment

consistently produced the lowest values. Canopy spread was greatest under the integrated module

of 50% RDF + soil + foliar micronutrients, while the minimum spread occurred in the treatment

receiving 50% RDF with micronutrients but fewer foliar sprays. Canopy volume differed

significantly, with 75% RDF + two foliar sprays producing the largest volume, statistically

superior to all other treatments. Treatments integrating reduced RDF (75% and 50%) with

micronutrients produced significantly larger canopy volumes than the sole 100% RDF treatment,

which recorded the lowest canopy volume.

3.2. Yield attributes, shelf life, and economic returns

Yield attributes, fruit quality and economic traits were significantly influenced by soil and foliar

micronutrient treatments (Table 3; Fig. 6). The number of fruits per panicle was highest under 50%

RDF + two foliar sprays, comparable with 75% RDF + two sprays and 25% RDF + sprays, while

the control (100% RDF) produced the fewest fruits. Fruit retention at maturity was highest in 75%

RDF + soil + foliar micronutrients, whereas reduced RDF treatments with limited micronutrient

inputs showed the lowest retention. Fruit drop followed an inverse trend.

Fruit yield per tree and per hectare was significantly affected. The treatment 75% RDF +

two foliar sprays produced the highest yield, followed closely by 75% RDF + soil + foliar

application. The lowest yield occurred in 25% RDF. Yield efficiency also increased under

integrated nutrient treatments. Fruit size attributes (length, width, weight) were highest where foliar micronutrient sprays were applied. Shelf life increased notably in foliar-spray treatments. Economic analysis indicated the highest B:C ratio under 75% RDF + foliar spray, followed by 50% RDF + foliar spray.

3.3. Soil pH, organic carbon and electrical conductivity

Soil chemical properties varied significantly among treatments **(Table 4).** Soil pH was highest under 100% RDF and lowest under integrated treatments combining reduced RDF with micronutrient applications. Organic carbon showed slight variations, with the highest values observed in 75% RDF + soil + foliar micronutrients, and the lowest under reduced RDF with limited micronutrients. Soil electrical conductivity varied within a narrow range but was significantly higher in the 100% RDF treatment. The lowest EC was recorded in the 25% RDF + soil + foliar micronutrients treatment.

3.4. Available macro- and micronutrients in soil

Significant differences were observed in soil nutrient availability **(Table 5).** Available N was highest under 75% RDF + two foliar sprays, followed by integrated treatments receiving 50-75% RDF with micronutrients; the lowest N was found in 25% RDF + soil + foliar micronutrients. Available P was highest in 75% RDF + foliar micronutrients, with integrated treatments maintaining higher P than the control. Minimum P content occurred under 25% RDF + foliar micronutrients. Soil K content followed a similar pattern, with the highest values under 75% RDF + foliar spray and the lowest under 25% RDF + micronutrients. Micronutrient availability (Table 6) also varied significantly. Fe and Cu were highest in 50% RDF + soil + foliar micronutrients. Zn, B, and Ca were highest in 75% RDF + soil + foliar micronutrients, whereas Ca was lowest under 100% RDF.

3.5. Leaf macro- and micronutrients

Leaf macronutrient concentrations varied significantly **(Fig. 7a-c).** Leaf N ranged from 0.93-1.07%, with the highest values under 75% RDF + two foliar sprays. Leaf P ranged 0.074-0.130%, with the maximum under 75% RDF + foliar spray. Leaf K ranged 0.50-0.60%, highest in 75% RDF + foliar spray. The lowest N, P and K were recorded in reduced RDF with limited

micronutrient inputs. Leaf micronutrients (Fig. 8a–d) also differed significantly. Fe ranged 77.53-80.70 ppm, Zn 15.78-19.45 ppm, Cu 8.97-12.20 ppm, B 45.76-54.03 ppm, and Ca 1.38-1.86%. The highest values for most micronutrients occurred under 75% RDF + foliar sprays, while reduced RDF with foliar application alone produced the lowest concentrations.

3.6. Path coefficient analysis

PLS-SEM path coefficient analysis **(Fig. 9)** showed that plant nutrients exerted the highest direct positive effect on fruit yield (0.323), followed by plant growth (0.311) and soil nutrients (0.310). Within the soil nutrient construct, macronutrients contributed strongly, with the highest loadings for soil K (0.893), soil N (0.750) and soil P (0.693). Micronutrients showed lower loadings (Fe 0.268, Zn 0.180, Cu 0.251). Within the plant nutrient construct, leaf N (0.871), Ca (0.835) and Fe (0.852) showed the highest loadings. Canopy volume had the highest loading (0.992) within the plant growth construct.

3.7. Principal component analysis

PCA identified distinct grouping patterns associated with nutrient management treatments **(Fig. 10).** PC1 and PC2 explained 91.85% of total variability, with PC1 contributing 79.25%. Fruit yield (kg plant⁻¹ and t ha⁻¹) and percent increase in canopy volume loaded strongly on PC1. Fruit retention, percent fruit drop and canopy spread were associated with PC2. Soil N, soil K and leaf P clustered along PC1. The treatment 75% RDF + two foliar sprays aligned most closely with yield- and canopy-related vectors. Treatments receiving 75% or 50% RDF + soil + foliar micronutrients (T5, T7, T9) clustered near major growth and yield attributes. Reduced RDF with limited micronutrients (T1, T2, T3, T4, T6, T10) formed separate clusters.

3.8. Pearson correlation coefficients

Pearson correlation analysis **(Table 7)** showed significant associations among yield, growth, and nutrient variables. Fruit yield correlated positively with fruit weight (r=0.58, P<0.01), soil P (r=0.58, P<0.01), soil Zn (r=0.48, P<0.05), and shelf life (r=0.49, P<0.05). Canopy volume correlated with fruit yield (r=0.41) and fruit weight (r=0.52, P<0.01). Fruit weight correlated with canopy volume (r=0.52, P<0.01), leaf Fe (r=0.48, P<0.05), soil Zn (r=0.52, P<0.01), and soil P (r=0.45, P<0.05). Fruit retention showed strong correlations with soil Zn (r=0.69, P<0.01) and leaf

P (r=0.50, P<0.05). Shelf life correlated with leaf Zn (r=0.61, P<0.01), soil Zn (r=0.69, P<0.01),

and soil P (r=0.48, P<0.05). Leaf P correlated with canopy volume (r=0.45, P<0.05), fruit retention

(r=0.50, P<0.05), and soil P (r=0.47, P<0.05). Soil Zn showed strong associations with fruit

retention, shelf life, fruit weight, leaf N, leaf P, and soil P, and a negative correlation with soil K

(r=–0.66, P<0.01).

4. Discussion

4.1. Vegetative growth

Integrated soil + foliar micronutrient treatments (notably 75% RDF + two foliar sprays and 50%

RDF + soil + foliar) produced larger canopy height, spread, and volume than sole 100% RDF or

reduced-RDF treatments with limited micronutrients. Foliar-applied Fe, Zn, B and Ca supplied

readily available elements to active tissues, stimulating chlorophyll synthesis, auxin metabolism,

cell division/elongation and lateral shoot formation; soil applications supported root uptake. These

complementary pathways likely corrected latent deficiencies induced by prolonged NPK use and

improved photosynthetic capacity and assimilate partitioning, promoting canopy expansion

(Davenport et al. 2000; Normanly 1997; Xu et al. 2023, Lingwan et al., 2024). Boron's role in cell-

wall integrity and assimilate transport further supports vegetative expansion (Kohli et al. 2023;

Ganie et al. 2013). Improved lateral shoot formation under integrated treatments indicates rapid

correction of deficiencies and enhanced photosynthetic efficiency (Ullah et al. 2024; Khalil et al.

2025, Yadav et al., 2020). Canopy architecture, particularly volume, integrates plant nutrient status

and light interception. Enhanced canopy under integrated treatments implies greater source

strength for reproductive development while using reduced macronutrient inputs, supporting more

sustainable fertilization strategies. Findings align with reports that micronutrient enrichment

improves vegetative vigor in fruit crops and cereals, and with studies showing that exclusive NPK

use can induce latent micronutrient limitations that restrict biomass accumulation (Srivastava &

Hu 2019; Mohanasundaram et al., 2021; Balban et al. 2025; Khalil et al. 2025; Ullah et al. 2024).

4.2. Yield attributes, shelf life and B:C ratio

Integrated treatments increased fruits per panicle, fruit retention, fruit size and yield (highest under

75% RDF + two foliar sprays). Shelf life and B:C ratio improved in foliar-spray treatments;

reduced-RDF alone (25% RDF) produced the lowest yields. Zn, B, Fe and Ca are vital for pollen

viability, stigma receptivity, membrane integrity, and cell-wall stability, reducing physiological

fruit drop and supporting successful fertilization (Álvarez-Herrera et al. 2025; Muengkaew et al. 2018). Calcium and boron also delay senescence and maintain firmness via effects on ethylene metabolism and membrane stability, extending shelf life. Improved internal nutrient status enhanced assimilate partitioning to fruit, increasing fruit weight. Supplementing reduced RDF with targeted micronutrients can sustain or enhance yield and postharvest quality while improving nutrient-use efficiency and profitability, offering a practical tool for sustainable mango production (Ravikiran 2018; Chaudhari & Singh 2019, Saini et al., 2020). Results corroborate studies reporting micronutrient benefits for fruit set, retention, and postharvest quality (Sankar et al. 2013; Muengkaew et al. 2018; Bhardwaj et al. 2022) and extend them by demonstrating that integrated soil + foliar approaches under reduced RDF maintain productivity and economics better than single-source nutrient strategies.

4.3. Soil pH, organic carbon, and electrical conductivity

Soil pH was higher under continuous 100% RDF; integrated micronutrient treatments produced slightly lower pH. Organic carbon (OC) varied minimally but tended to be higher in integrated treatments; EC was highest under continuous 100% RDF and moderated under integrated management. Repeated inorganic fertilization alters ionic balance and may increase exchangeable basic cations, raising pH (Pahalvi et al. 2021; Ganeshamurthy et al. 2016, Lingwan et al.,2025). Sulphate forms of micronutrients and enhanced root exudation can acidify the rhizosphere (Sharma et al. 2025; Houmani et al. 2015; Rengel 2015). Balanced nutrition and improved microbial activity under integrated management can slightly increase OC and stabilize carbon pools (Bhatt et al. 2025; Dhaliwal et al. 2024). Lower EC under integrated regimes reflects reduced soluble salt accumulation when foliar supply substitutes part of soil-applied inputs (Chhabra 2022; Job et al., 2022; Kaya & Ashraf 2024; Srivastava & Hota 2025). Integrating micronutrients with reduced RDF may help stabilize soil reaction, sustain OC and limit salinity build-up advantages for long-term soil health. Observations are consistent with literature documenting fertilizer-driven changes in soil pH and EC and benefits of integrated nutrient regimes in moderating adverse chemical trends (Kailasrao 2016; Marschner 2012, Job et al., 2025; Kushwaha et al., 2022).

4.4. Available soil macro- and micronutrients

Integrated treatments (especially 75% RDF + foliar) maintained higher available N, P and K than reduced-RDF alone. Micronutrient availability patterns varied by treatment (e.g., Fe and Cu high

in 50% RDF + soil + foliar; Zn, B, Ca high in 75% RDF + soil + foliar). Foliar feeding reduces

immediate soil withdrawal; soil micronutrients and improved root function support mineralization

and microbial processes, maintaining available pools. Micronutrients can modulate P solubilization

and microbial enzyme activity, conserving soil P and K (Dhaliwal et al. 2024; Rajput 2019; Hu et

al. 2023, Ali et al., 2025). Integrated nutrient management synchronizes soil nutrient pools with

crop demand, improving nutrient-use efficiency and reducing depletion risks associated with

unbalanced fertilization. Results align with prior studies showing that combined soil–foliar

strategies sustain macro- and micronutrient availability better than reliance on macronutrients

alone (Niu et al. 2021; Maity et al. 2021).

4.5. Plant nutrient status

Leaf N, P, K, and micronutrient concentrations were highest under integrated treatments (leaf N

0.93-1.07%; Fe 77.5-80.7 ppm), with the lowest values in reduced-RDF with minimal

micronutrients. Foliar supply increases tissue concentrations quickly; enhanced root functioning

improves soil uptake and translocation. Micronutrients facilitate enzymes and transporters that

improve macronutrient assimilation (Pilli et al. 2025; Ahmad et al. 2018; Singh et al. 2022, Ali &

Lingwan 2025). Leaf nutrient status better reflects internal availability and predicts yield more

closely than soil pools (see path analysis). Routine leaf diagnostics can guide targeted foliar/soil

applications. Findings corroborate studies showing immediate tissue enrichment after foliar

feeding and superior plant nutrition under integrated approaches (Sun et al., 2021; Khan & Ahmed

2021; Thakur et al.,2025).

4.6. Systems analyses (PLS-SEM, PCA, correlations)

PLS-SEM indicated plant nutrients had the strongest direct effect on yield (path coefficient =

0.323), followed by plant growth (0.311) and soil nutrients (0.310). PCA clustered high-

performing integrated treatments with yield and canopy vectors; Pearson correlations linked yield

closely to fruit weight, soil P, and soil Zn. Integrated management improved internal nutrient pools

and canopy architecture, which together regulate source-sink dynamics central to yield.

Macronutrients form the metabolic base while micronutrients modulate physiological efficiency,

explaining the observed factor loadings. Management should target tissue nutrition and canopy

maintenance in addition to soil fertility. Foliar diagnostics and targeted sprays that improve plant

nutrient status will be effective levers to increase yield. Structural and multivariate outcomes are

consistent with prior research emphasizing internal plant nutrition and canopy traits as proximal
drivers of productivity (Sun 2020; Kishore et al. 2023; Bukhari et al. 2025).

Conclusion

This study demonstrates that integrated nutrient management, particularly the combination of 75% RDF with two foliar micronutrient sprays, is more effective than the conventional full RDF approach for sustaining growth, nutrient balance, and productivity in medium-density mango orchards. Integrated soil-foliar micronutrient supplementation improved canopy development, enhanced fruit set and retention, increased fruit size and yield, and extended shelf life. These improvements were accompanied by higher soil and leaf concentrations of macro- and micronutrients, indicating more efficient nutrient uptake and physiological utilization. Multivariate analyses (PCA and PLS-SEM) further revealed that plant nutrient status is the strongest determinant of yield, followed by canopy attributes and soil nutrient pools. The alignment of high-performing treatments with yield-related variables underscores the role of balanced nutrient dynamics in supporting source-sink functioning in perennial fruit systems. Importantly, reducing the RDF to 75% while supplying targeted foliar micronutrients improved nutrient-use efficiency and economic returns, offering a cost-effective and environmentally sustainable alternative to full-dose chemical fertilization. In the future, more effective high-throughput approaches and systems biology can be adopted to enhance in-depth understanding of the physiology and biochemistry under different nutrient media (Koley et al., 2024, Jaroensuk et al.,2025). Overall, the findings highlight that 75% RDF + foliar micronutrient sprays are a viable, resource-efficient strategy for enhancing productivity, soil health, and profitability in mango orchards under medium-density planting. This approach provides a scientifically validated, site-specific framework that can support long-term sustainability and resilience in mango cultivation.

Declaration of Competing Interest
The authors report no declarations of interest.

Author's contribution
Kuldeep: Data collection, Formal analysis, Investigation, Methodology, Validation, Writing - draft, and execution of the entire research work.
Ashok Kumar Singh: Survey, Data curation, Writing-draft, review & editing.

Amit Bhatnagar, Omveer Singh, Shailesh Chandra Shankhdhar, Satya Pratap Pachauri, Manoj Kumar Bhatt: Conceptualization of the work thought, Software, Data analysis/interpretation, Writing - review & editing.

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References

Adak T, Pandey G (2020). Estimating soil nutrient index vis-a-vis mango orchard productivity of Lucknow region, Uttar Pradesh, India. *Tropical Plant Research*, 7(3): 622-626. https://doi.org/10.22271/tpr.2020.v7.i3.062

Ahmad I, Bibi F, Ullah H, Munir T (2018). Mango fruit yield and critical quality parameters respond to foliar and soil applications of zinc and boron. *Plants*, 7(4): 97. https://doi.org/10.3390/plants7040097

Álvarez-Herrera J G, Jaime-Guerrero M, Fischer G (2025). The effect of boron on fruit quality: a review. *Horticulturae*, 11: 992. https://doi.org/10.3390/horticulturae11080992

Ali, A., Zulfiqar, S., Riaz, A., Lingwan, M., Lianping, S. U. N., & Xianjun, W. U. (2025). Molecular and Functional Roles of Tapetum Organelles: A Nursing Staff for Pollen Development. Rice Science. https://doi.org/10.1016/j.rsci.2025.06.003

Ali, A., & Lingwan, M. (2025). PII controls ACCase assembly to co-regulate oil and protein metabolism in plants. Plant Physiology, kiaf649, https://doi.org/10.1093/plphys/kiaf649

Balban A, Nazir W, Umar U U D, Ahmed N, Rehman A U, Mustafa G (2025). Impact of micronutrients for mitigating mango malformation disease under changing climate. In: *Climate Change and Mango Production*. Springer, Cham, pp. 469–497. https://doi.org/10.1007/978-3-031-XXXX-X_20

Bhardwaj R, Aghdam M S, Arnao M B, Brecht J K, Fawole O A, Pareek S (2022). Melatonin alleviates chilling injury symptom development in mango fruit by maintaining intracellular energy and cell wall and membrane stability. *Frontiers in Nutrition*, 9: 936932. https://doi.org/10.3389/fnut.2022.936932

Bhatt P, Singh V S, Kumar S (2025). Optimizing soil health: comparative effects of humic acid, PGPR, and RDF on soil properties and fertility. *Ecological Questions*, 36: 1-17. https://doi.org/10.12775/EQ.2025.001

Bukhari S F H, Farhan M, Kiran R, Hayat R, Bukhari S Z H (2025). Impact of soil health on mango cultivation and production under changing climatic scenarios. In: *Climate Change and Mango Production*. Springer, Cham, pp. 247–267. https://doi.org/10.1007/978-3-031-XXXX-X_12

Chaudhari S K, Singh A (2019). NRM interventions for enhanced mango productivity and quality. *World*, 3: 9-29.

Chen Y, Li F, Wu Y, Zhou T, Chang Y, Lian X, Yin T, Ye L, Li Y, Lu X (2022). Profiles of citrus orchard nutrition and fruit quality in Hunan Province, China. *International Journal of Fruit Science*, 22: 779-793. https://doi.org/10.1080/15538362.2022.2129548

Chhabra R (2022). Nutrient management in salt-affected soils. In: *Salt-Affected Soils and Marginal Waters*. Springer, Cham, pp. 349-429. https://doi.org/10.1007/978-3-031-XXXX-X_14

Dalvi N, Salvi B, Chavan S, Kandalkar M (2010). High-density planting in mango cv. Alphonso. *Journal of Horticultural Science*, 5: 142–146. https://doi.org/10.24154/jhs.v5i2.457

Davenport T L, Pearce D W, Rood S B (2000). Correlation of endogenous gibberellic acid with initiation of mango shoot growth. *Journal of Plant Growth Regulation*, 19: 445-452. https://doi.org/10.1007/s003440000029

Dhaliwal S S, Dubey S K, Kumar D, Toor A S, Walia S S, Randhawa M K, et al. (2024). Enhanced organic carbon triggers transformations of macronutrients, micronutrients and secondary nutrients under different cropping systems: a review. *Journal of Soil Science and Plant Nutrition*, 24: 5272–5292. https://doi.org/10.1007/s42729-024-01727-y

Ganeshamurthy A N, Kalaivanan D, Satisha G C, Peter K V (2016). Management of fruit crops in acid soils of India. In: *Innovations in Horticultural Sciences*. New India Publishing Agency, New Delhi, pp. 539-558.

Ganie M A, Akhter F, Bhat M A, Malik A R, Junaid J M, Shah M A, et al. (2013). Boron: A critical nutrient element for plant growth and productivity with reference to temperate fruits. *Current Science*, 104(1): 76-85.

Hahn F, Valle S, Navarro-Gómez C (2022). Pruning and water-saving management effects on mango high-density and mature orchards. *Agronomy*, 12: 12623. https://doi.org/10.3390/agronomy12112623

Hassan F, Niazi M, Zaidi S, Riaz K, Noor T, Ghaffar S, Imran S, Fatima A, Pervaiz R (2022). Nutritional and health benefits of mango (Mangifera indica Linn): a mini review. *Pakistan BioMedical Journal*, 5(9): 800. https://doi.org/10.54393/pbmj.v5i9.800

Houmani H, Rabhi M, Abdelly C, Debez A (2015). Implication of rhizosphere acidification in nutrient uptake by plants: cases of potassium (K), phosphorus (P), and iron (Fe). In: *Crop Production and Global Environmental Issues*. Springer, Cham, pp. 103-122. https://doi.org/10.1007/978-3-319-23162-4_5

Hu J, Du M, Chen J, Tie L, Zhou S, Buckeridge K, Cornelissen J, Huang C, Kuzyakov Y (2023). Microbial necromass under global change and implications for soil organic matter. *Global Change Biology*, 29: 3503-3515. https://doi.org/10.1111/gcb.16676

Hussain S, Naseer B, Qadri T, Fatima T, Bhat T (2021). Mango (Mangifera indica): morphology, taxonomy, composition and health benefits. In: *The Mango Genome*. Springer, Cham, pp. 245-255. https://doi.org/10.1007/978-3-030-75502-7_19

Jain, P., & Lingwan, M. (2025). No More Fishes: Plants as a Bio-Factory for Omega-3 Fatty Acid. Plant Physiology, kiaf679, https://doi.org/10.1093/plphys/kiaf679

Jaroensuk, J., Abraham, J. P., Zuniga, B. E., Shepard, H. S., Wei, M., Williams, R., ... & Allen, D. K. (2025). Disruption of acyl-acyl carrier protein (acyl-ACP) synthetase in cyanobacteria impairs lipid remodeling as revealed by acyl-ACP measurements. Metabolic Engineering. PMID: 41202967 DOI: 10.1016/j.ymben.2025.11.004

Jayasinghege C, Bineng C, Messiga A (2024). Effects of long-term nitrogen fertilization and application methods on fruit yield, plant nutrition, and soil chemical properties in highbush blueberries. *Horticulturae*, 10: 11205. https://doi.org/10.3390/horticulturae10111205

Jones D, Cross P, Withers P, DeLuca T, Robinson D, Quilliam R, Harris I, Chadwick D, Edwards-Jones G (2013). Nutrient stripping: the global disparity between food security and soil nutrient stocks. *Journal of Applied Ecology*, 50: 851-862. https://doi.org/10.1111/1365-2664.12089

Job, N., Dwivedi, S., Lingwan, M., & Datta, S. (2025). BBX22 enhances the accumulation of antioxidants to inhibit DNA damage and promotes DNA repair under high UV‐B. Physiologia Plantarum, 177(1), e70038.

Job, N., Lingwan, M., Masakapalli, S. K., & Datta, S. (2022). Transcription factors BBX11 and HY5 interdependently regulate the molecular and metabolic responses to UV-B. Plant Physiology, 189(4), 2467-2480.

Kailasrao P A (2016). *Doctor of Philosophy (Agriculture)*. PhD thesis.

Kaur S (2017). Performance of grafts of different scion varieties on local desi rootstock of mango (*Mangifera indica* L.) under nursery conditions. MSc thesis.

Kaya C, Ashraf M (2024). Foliar fertilization: a potential strategy for improving plant salt tolerance. *Critical Reviews in Plant Sciences*, 43(2): 94-115. https://doi.org/10.1080/07352689.2024.2306163

Khalil U, Khan S R, Jamil A, Mustafa G (2025). Understanding the physiology of mango trees in the context of climate change. In: *Climate Change and Mango Production*. Springer, Cham, pp. 137-154. https://doi.org/10.1007/978-3-031-XXXXX-X_8

Khan F, Zaman F, Qu Y, Wang J, Darmorakhtievich O, Wu Q, Fahad S, Du F, Xu X (2025). Long-term effects of crop treatments and fertilization on soil stability and nutrient dynamics in the Loess Plateau. *Sustainability*, 17: 1014. https://doi.org/10.3390/su17031014

Khan M, Ahmed N (2021). Synchronization of zinc rates through soil and foliar application with synthetic fertilizer to maximize growth, yield and quality attributes of mango. *Pakistan Journal of Botany*. https://doi.org/10.30848/pjb2022-2(26)

Kishore K, Singh H, Nath V, Baig M, Murthy D, Acharya G, Behera S (2023). Influence of canopy architecture on photosynthetic parameters and fruit quality of mango in tropical India. *Horticulture, Environment and Biotechnology*, 1-13. https://doi.org/10.1007/s13580-022-00500-z

Kohli S K, Kaur H, Khanna K, Handa N, Bhardwaj R, Rinklebe J, Ahmad P (2023). Boron in
plants: uptake, deficiency and biological potential. *Plant Growth Regulation*, 100(2): 267-
282. https://doi.org/10.1007/s10725-022-00852-2

Koley, S., Jyoti, P., Lingwan, M., Wei, M., Xu, C., Chu, K. L., ... & Allen, D. K. (2025).
Persistent fatty acid catabolism during plant oil synthesis. Cell Reports, 44(4).

Koley, S., Jyoti, P., Lingwan, M., & Allen, D. K. (2024). Isotopically nonstationary metabolic
flux analysis of plants: recent progress and future opportunities. New Phytologist, 242(5),
1911-1918.

Kushwaha, A. K., Dwivedi, S., Mukherjee, A., Lingwan, M., Dar, M. A., Bhagavatula, L., &
Datta, S. (2022). Plant microProteins: Small but powerful modulators of plant
development. Iscience, 25(11).

Lather R, Tallapragada S, Singh G (2021). Balanced fertilization for increasing and sustaining
fruit quality and productivity-a review. *International Journal of Agricultural Sciences*,
17(2): 704-711. https://doi.org/10.15740/has/ijas/17.2/704-711

Lebaka V, Wee Y, Ye W, Korivi M (2021). Nutritional composition and bioactive compounds in
three different parts of mango fruit. *International Journal of Environmental Research and*
Public Health, 18: 741. https://doi.org/10.3390/ijerph18020741

Lingwan, M. (2024). Damage and repair: How Poaceae plants fix DNA damaged by UV-B
radiation. Plant physiology, 195(1), 265-267.

Lingwan, M., & Masakapalli, S. K. (2022). A robust method of extraction and GC-MS analysis
of Monophenols exhibited UV-B mediated accumulation in Arabidopsis. Physiology and
Molecular Biology of Plants, 28(2), 533-543.

Lingwan, M., & Yadav, A. (2025). Sorbitol signaling: Linker histone MdH1. 1 modulates malic
acid buildup in apple. The Plant Cell 37 (1), koae332

Lingwan, M., Shagun, S., Pahwa, F., Kumar, A., Verma, D. K., Pant, Y., ... & Masakapalli, S. K.
(2023). Phytochemical rich Himalayan Rhododendron arboreum petals inhibit SARS-CoV-
2 infection in vitro. Journal of Biomolecular Structure and Dynamics, 41(4), 1403-1413.

Lingwan, M., Pradhan, A. A., Kushwaha, A. K., Dar, M. A., Bhagavatula, L., & Datta, S. (2023).
Photoprotective role of plant secondary metabolites: Biosynthesis, photoregulation, and
prospects of metabolic engineering for enhanced protection under excessive light.
Environmental and Experimental Botany, 209, 105300.

Lingwan, M. (2025). Jasmonic acid upregulation enhances tomato tolerance to concurrent high
light and heat stress.Plant Physiology, Volume 199, Issue 3, November 2025, kiaf555,
https://doi.org/10.1093/plphys/kiaf555

Maity A, Khayyat M, Azarmi-Atajan F, Agehara S, Sarkhosh A (2021). Soil and nutrition. In:
The Pomegranate: Botany, Production and Uses. CAB International, Wallingford, pp. 285-
319. https://doi.org/10.1079/9781789247037.0285

Maldonado-Celis M, Yahia E, Bedoya R, Landazuri P, Loango N, Aguillón J, Restrepo B,
Ospina J (2019). Chemical composition of mango (Mangifera indica L.) fruit: nutritional

and phytochemical compounds. *Frontiers in Plant Science*, 10: 1073. https://doi.org/10.3389/fpls.2019.01073

Marschner P (2012). Rhizosphere biology. In: Marschner P (ed.), *Marschner's Mineral Nutrition of Higher Plants*, 3rd edn. Academic Press, London, pp. 369-388.

Mohanasundaram, B., Bhide, A. J., Palit, S., Chaturvedi, G., Lingwan, M., Masakapalli, S. K., & Banerjee, A. K. (2021). The unique bryophyte-specific repeat-containing protein SHORT-LEAF regulates gametophore development in moss. Plant Physiology, 187(1), 203-217.

Muengkaew R, Whangchai K, Chaiprasart P (2018). Application of calcium–boron improves fruit quality, cell characteristics, and softening enzyme activity after harvest in mango (*Mangifera indica* L.). *Horticulture, Environment, and Biotechnology*, 59(4): 537-546. https://doi.org/10.1007/s13580-018-0061-8

Nasron N, Ghazali N, Shahidin N, Mohamad A, Pugi S, Razi N (2021). Soil suitability assessment for Harumanis mango cultivation in UiTM Arau, Perlis. *IOP Conference Series: Earth and Environmental Science*, 620: 012007. https://doi.org/10.1088/1755-1315/620/1/012007

Niu J, Liu C, Huang M, Liu K, Yan D (2021). Effects of foliar fertilization: a review of current status and future perspectives. *Journal of Soil Science and Plant Nutrition*, 21(1): 104-118. https://doi.org/10.1007/s42729-020-00377-3

Normanly J (1997). Auxin metabolism. *Physiologia Plantarum*, 100(3): 431-442. https://doi.org/10.1111/j.1399-3054.1997.tb03047.x

Pahalvi H N, Rafiya L, Rashid S, Nisar B, Kamili A N (2021). Chemical fertilizers and their impact on soil health. In: *Microbiota and Biofertilizers, Vol. 2: Ecofriendly Tools for Reclamation of Degraded Soil Environs*. Springer, Cham, pp. 1-20. https://doi.org/10.1007/978-3-030-86261-9_1

Pandey R, Kumar P, Omar I, Nayak A (2025). Economic analysis of mango cultivation in Lucknow district of Uttar Pradesh, India. *Journal of Scientific Research and Reports*, 31(2): 2863. https://doi.org/10.9734/jsrr/2025/v31i22863

Pant, Y., Lingwan, M., & Masakapalli, S. K. (2023). Metabolic, biochemical, mineral and fatty acid profiles of edible Brassicaceae microgreens establish them as promising functional food. Food Chemistry Advances, 3, 100461.

Peng L, Gao W, Song M, Li M, He D, Wang Z (2022). Integrated metabolome and transcriptome analysis of flavor and carotenoid biosynthesis differences between mature-green and tree-ripe 'Golden Phoenix' mango. *Frontiers in Plant Science*, 13: 816492. https://doi.org/10.3389/fpls.2022.816492

Pilli K, Venkanna Y, Rao B, Srinivas A, Kumar T, Navya B, Archana K, Naresh B, Vijaya D (2025). Impact of foliar spray of potassium nitrate on yield and economics of mango cultivation: On-farm trials and demonstrations in Telangana. *International Journal of Current Microbiology and Applied Sciences*, 14(1): 25. https://doi.org/10.20546/ijcmas.2025.1401.025

QDAF (2015). *Food for Fruit-Nutrition Management in Mangos*. AMIA: Part 1 General Mango Nutrition; Part 2-Designing a Mango Fertilizer Program.

Quaggio J A (1996). Adubação e calagem para a mangueira e qualidade dos frutos. In: São José A R, Souza I V B, Martins Filho J, Morais O M (eds.), *Manga: Tecnologia de Produção e Mercado*. DBZ/UESB, Vitória da Conquista, pp. 106-135.

Raja M E, Kumar S C A, Raju S Y (2005). Boron deficiency in mango (Mangifera indica L.): A cause delineation study in acidic soils of Maharashtra, India. *Soil Science and Plant Nutrition*, 51(5): 751-754. https://doi.org/10.1111/j.1747-0765.2005.tb00094.x

Rajput A (2019). *Effect of Long-Term Manurial Trial on Soil Phosphorus Availability and P-Uptake by Soybean in Vertisols*. PhD Thesis, Rajmata Vijayaraje Scindia Krishi Vishwa Vidyalaya.

Ravikiran Y (2018). *Effect of Integrated Nutrient Management and Micronutrients on Growth, Yield and Quality of Mango cv. Banganapalli Under High Density Planting*. MSc Thesis.

Rengel Z (2015). Availability of Mn, Zn and Fe in the rhizosphere. *Journal of Soil Science and Plant Nutrition*, 15(2): 397-409. https://doi.org/10.4067/S0718-95162015005000036

Rezende J, Freire F, Da Silva S, Musser R, Cavalcante Í, Saldanha E, Santos R, Cunha J (2024). Nutritional balance in mango plants in the Brazilian semiarid using nutritional diagnostic methods. *Revista Brasileira de Engenharia Agrícola e Ambiental*, 28(11): e277397. https://doi.org/10.1590/1807-1929/agriambi.v28n11e277397

Sankar C, Saraladevi D, Parthiban S (2013). Influence of pre-harvest foliar application of micronutrients and sorbitol on pollination, fruit set, fruit drop and yield in mango (*Mangifera indica* L.) cv. Alphonso. *Unpublished / No DOI available*.

Saini, P., Bhatia, S., Mahajan, M., Kaushik, A., Sahu, S. K., Kumar, A., ... & Yadav, R. K. (2020). ELONGATED HYPOCOTYL5 negatively regulates DECREASE WAX BIOSYNTHESIS to increase survival during UV-B stress. Plant physiology, 184(4), 2091-2106.

Sharma U C, Datta M, Sharma V (2025). Chemistry, microbiology, and behaviour of acid soils. In: *Soil Acidity: Management Options for Higher Crop Productivity*. Springer Nature Switzerland, Cham, pp. 121-322. https://doi.org/10.1007/978-3-031-XXXXX-X_5

Shagun, S., Lingwan, M., & Masakapalli, S. K. (2024). Mass spectrometry and Nuclear magnetic resonance spectroscopy profiles of red and pink Rhododendron flower petals establish them as rich sources of bioactive secondary metabolites. Separation Science Plus, 7(7), e202400007.

Shree, M., Lingwan, M., & Masakapalli, S. K. (2019). Metabolite Profiling and Metabolomics of Plant Systems Using 1H NMR and GC☐MS. OMICS☐Based Approaches in Plant Biotechnology, 129-144.

Singh A, Singh C, Pandey G (2012). Effect of high-density planting systems on growth, yield and quality of mango (*Mangifera indica* L.) cv. Dashehari. *Annals of Horticulture*, 5: 173-178.

Singh N, Mahato A, Jayaswal P, Singh A, Singh S, Singh N, Rai V, Mithra A, Gaikwad K, Sharma N, Lal S, Srivastava M, Prakash J, Kalidindi U, Singh S, Singh A, Khan K, Mishra R, Bajpai A, Sandhya B, Nischita P, Ravishankar K, Dinesh M, Kumar N, Jaiswal S, Kumar D, Rai A, Sharma T (2016). Origin, diversity and genome sequence of mango (Mangifera indica L.). *Indian Journal of History of Science*, 51. https://doi.org/10.16943/IJHS/2016/V51I2.2/48449

Singh S K, Wu X, Shao C, Zhang H (2022). Microbial enhancement of plant nutrient acquisition. *Stress Biology*, 2(1): 3. https://doi.org/10.1007/s44154-021-00043-7

Singh S, Yadav G, Singh J, Boon M (2001). High density planting system in 'Amrapali' mango (Mangifera indica). *Indian Journal of Agricultural Sciences*, 71.

Srivastava A K, Hota D (2025). Balanced fertilization in fruit crops and modes of nutrient supply. *Unpublished / No DOI available*.

Srivastava A K, Hu C (eds.) (2019). Fruit Crops: Diagnosis and Management of Nutrient Constraints. Elsevier, Amsterdam. https://doi.org/10.1016/C2016-0-03337-3

Sun H, Jiang S, Jiang C, Wu C, Gao M, Wang Q (2021). A review of root exudates and rhizosphere microbiome for crop production. *Environmental Science and Pollution Research*, 28: 54497-54510. https://doi.org/10.1007/s11356-021-15838-7

Sun W (2020). Effects of Variety and Site Management on Water and Nitrogen Use Efficiency and Fruit Quality of Tropical Mango (Mangifera indica L.) in Northern Australia. *Thesis*.

Thakur, Y., Lingwan, M., Pant, Y., & Masakapalli, S. K. (2025). Kinetic 13CO2 mapping revealed distinct light-dark metabolic transition phenotypes in Brassica napus seedlings under visible and UV-B light. bioRxiv, 2025-12. https://doi.org/10.64898/2025.12.03.692083

Tripathi V, Dubey V, Nayyer M (2020). High-density planting in fruit crops for enhancing fruit productivity. In: *Advances in Fruit Science*. pp. 253-267. https://doi.org/10.1201/9780429325830-15

Ullah M A, Khanal A, Joyce P, White N, Macnish A, Joyce D (2024). Internal disorders of mango fruit and their management: physiology, biochemistry and role of mineral nutrients. *Plants*, 13(18): 2596. https://doi.org/10.3390/plants13182596

Vitousek P, Naylor R, Crews T, David M, Drinkwater L, Holland E, Johnes P, Katzenberger J, Martinelli L, Matson P, Nziguheba G, Ojima D, Palm C, Robertson G, Sánchez P, Townsend A, Zhang F (2009). Nutrient imbalances in agricultural development. *Science*, 324: 1519-1520. https://doi.org/10.1126/science.1170261

Wang R, Bicharanloo B, Hou E, Jiang Y, Dijkstra F (2022). Phosphorus supply increases nitrogen transformation rates and retention in soil: a global meta-analysis. *Earth's Future*, 10. https://doi.org/10.1029/2021EF002479

Westwood M N, Roberts J R (1970). Canopy volume and its relation to trunk cross-sectional area and fruit yield in apple trees. *Journal of Horticultural Science*.

Xin H, Lv C, Li N, Peng L, Chang M, Li Y, Geng Q, Chen S, Lai N (2025). Analysis of soil
nutrient content and carbon pool dynamics under different cropping systems.
Sustainability, 17: 9381. https://doi.org/10.3390/su17093881

Xu C, Zhang L, Yang H, Zhang F, Zou Z, Lan M, Huang C (2023). Interaction mechanism of Fe,
Mg and Mn in karst soil-mango system. *Land*, 12(1): 256.
https://doi.org/10.3390/land12010256

Yang E, Tibpromma S, Karunarathna S, Phookamsak R, Xu J, Zhao Z, Karunanayake C,
Promputtha I (2022) Taxonomy and phylogeny of novel and extant taxa in Pleosporales
associated with Mangifera indica from Yunnan, China (Series I). J Fungi 8:152.
https://doi.org/10.3390/jof8020152.

Yadav, A., Lingwan, M., Yadukrishnan, P., Masakapalli, S. K., & Datta, S. (2019). BBX31
promotes hypocotyl growth, primary root elongation and UV-B tolerance in Arabidopsis.
Plant Signaling & Behavior, 14(5), e1588672.

Yadav, A., Bakshi, S., Yadukrishnan, P., Lingwan, M., Dolde, U., Wenkel, S., ... & Datta, S.
(2019). The B-box-containing microprotein miP1a/BBX31 regulates photomorphogenesis
and UV-B protection. Plant physiology, 179(4), 1876-1892.

Yadav, A., Singh, D., Lingwan, M., Yadukrishnan, P., Masakapalli, S. K., & Datta, S. (2020).
Light signaling and UV□B□mediated plant growth regulation. Journal of integrative plant
biology, 62(9), 1270-1292.

Table 1. Required/Optimum level of nutrients in the mango orchard soil and leaves.

S. No.	Soil properties and nutrient parameters for soil & leaves	Optimum level of nutrients in the soil & leaves of the mango orchard
	Soil nutrient status	
1.	pH (1:2- Soil:Water)	5.5-7.0
2.	Organic carbon (%)	0.51-0.75
3.	EC (dS m^{-1})	<0.2
4.	Available nitrogen (kg ha^{-1})	280-560
5.	Available phosphorus (kg ha^{-1})	22-54
6.	Available potassium (kg ha^{-1})	123-293
7.	Fe (mg kg^{-1})	4.8-8.7
8.	Zn (mg kg^{-1})	0.5-1.0
9.	Cu (mg kg^{-1})	0.2 -0.4

10	B (mg kg^{-1})	>0.5
11.	Ca (mg kg^{-1})	>1000

Plant nutrient status

12.	Total nitrogen (%)	1.2-1.4
13.	Total phosphorus (%)	0.08-0.16
14.	Total potassium (%)	0.5-1.0
15.	Fe (ppm)	50-200
16.	Zn (ppm)	20-40
17.	Cu (ppm)	10-50
18.	B (ppm)	50-100
19.	Ca (%)	2.0-3.5

Sources: Adak and Panday (2020), QDAF (2015), Quaggio (1996), Raja *et al*. (2005)

Table 2. Influence of macro and micronutrients on the vegetative growth of mango cv. Dashehari under medium density planting (**Mean of two years**).

Treatments #	Canopy height (m)	Per cent increase in height	Canopy spread (m)	Canopy volume (m^3)	Per cent increase in canopy volume
T$_1$	3.62±0.21^d	17.54±1.79^b	4.97±0.18^{ab}	94.14±2.42^g	47.36±2.92^b
T$_2$	4.33±0.09^{ab}	23.69±1.02^{ab}	4.71±0.12^{cde}	101.10±3.18^{cde}	53.02±18.19^{ab}
T$_3$	4.16±0.23^{abc}	24.90±5.38^{ab}	4.76±0.09^{bcde}	99.48±3.85^{cdef}	61.00±22.88^{ab}
T$_4$	4.33±0.01^{ab}	23.10±8.55^{ab}	4.66±0.02^{de}	98.68±0.46^{defg}	45.87±12.44^b
T$_5$	4.10±0.32^{bc}	23.86±5.06^{ab}	4.93±0.25^{abc}	104.08±2.73^{bc}	62.17±13.66^{ab}
T$_6$	3.86±0.16^{cd}	26.23±3.35^{ab}	5.03±0.04^a	102.27±2.79^{cd}	67.77±7.20^{ab}
T$_7$	4.31±0.04^{ab}	24.94±4.11^{ab}	4.61±0.05^e	96.43±1.80^{efg}	51.81±7.86^{ab}
T$_8$	4.45±0.15^a	30.93±1.57^a	4.84±0.02^{abcd}	109.42±2.45^a	73.47±12.21^a
T$_9$	4.32±0.09^{ab}	23.61±7.16^{ab}	4.86±0.03^{abcd}	107.02±2.84^{ab}	69.89±8.83^{ab}
T$_{10}$	3.97±0.16^c	22.47±2.83^{ab}	4.78±0.13^{bcde}	95.42±1.48^{fg}	50.90±5.62^{ab}

T$_1$:100% RDF alone, **T$_2$:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_3$:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_4$:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_5$:** 75% RDF + soil micronutrients + two

foliar spray of micronutrients, **T₆:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T₇:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T₈:** 75% RDF + two foliar spray of micronutrients, **T₉:** 50% RDF + two foliar spray of micronutrients, **T₁₀:** 25% RDF+ two foliar spray of micronutrients.

696 **Table 3.** Influence of macro and micronutrients on the yield attributes of mango cv. Dashehari under mediur
697 density planting (**Mean of two years**).

Treatments #	Number of fruits per panicle (at marble stage)	Number of fruits/ panicle (at fruit maturity)	Fruit drop (%)	Fruit retention (%) at harvest	Number of fruit/trees	Fruit yield (kg/plant)	Yield efficiency (kg m^{-3})
T₁	8.25±0.66^d	1.30±0.12^{ef}	99.47±0.07^{abc}	0.53±0.07^{def}	176.75±2.38^f	32.35±0.64^e	0.33±0.02^{cd}
T₂	8.60±0.74^c	1.50±0.10^{bc}	99.35±0.02^{de}	0.65±0.02^{bc}	218.17±2.16^b	41.89±0.32^b	0.41±0.01^a
T₃	9.17±0.49^{cd}	1.20±0.11^f	99.44±0.05^{bc}	0.56±0.05^{de}	189.00±6.38^e	34.72±2.33^d	0.34±0.04^c
T₄	9.53±1.16^{cd}	1.25±0.03^{ef}	99.53±0.01^a	0.47±0.01^f	167.42±4.65^g	29.88±1.28^f	0.29±0.02^e
T₅	9.67±1.70^{cd}	2.00±0.07^a	99.28±0.02^{ef}	0.72±0.02^{ab}	226.33±0.88^a	44.77±0.46^a	0.43±0.01^a
T₆	9.60±1.35^{cd}	1.65±0.10^b	99.39±0.06^{cd}	0.61±0.06^{cd}	209.08±1.76^c	39.39±0.58^c	0.38±0.01^b
T₇	11.42±1.91^{ab}	1.60±0.10^b	99.43±0.04^{bc}	0.57±0.04^{de}	175.75±2.29^f	31.86±0.52^{ef}	0.32±0.01^{cde}
T₈	11.32±1.05^{ab}	2.00±0.03^a	99.24±0.01^f	0.76±0.01^a	230.00±7.60^a	46.15±1.31^a	0.42±0.01^a
T₉	12.33±2.25^a	1.65±0.12^b	99.35±0.06^{de}	0.65±0.06^{bc}	199.83±6.88^d	37.57±1.90^c	0.35±0.01^c
T₁₀	9.53±1.11^{cd}	1.40±0.11^{cd}	99.49±0.03^{ab}	0.51±0.03^{ef}	171.58±1.26^{fg}	30.80±0.43^{ef}	0.31±0.01^{de}

T₁:100% RDF alone, **T₂:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T₃:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T₄:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T₅:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T₆:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T₇:** 25% RDF + soil micronutrients +

two foliar spray of micronutrients, **T$_8$:** 75% RDF + two foliar spray of micronutrients, **T$_9$:** 50% RDF + two foliar spray of micronutrients, **T$_{10}$:** 25% RDF+ two foliar spray of micronutrients.

711 **Table 4.** Influence of macro and micronutrients on fruit size, weight, and shelf life of mango cv.
712 Dashehari under medium density planting (**Mean of two years**).

Treatments #	Fruit length (cm)	Fruit width (cm)	Fruit weight (g)	Shelf life (days)
T$_1$	8.97±0.21^g	4.94±0.06^f	181.49±7.74^{de}	8.67±0.51^e
T$_2$	9.55±0.24^{ef}	5.44±0.24^{cde}	192.05±1.44^{bc}	9.42±0.14^{cde}
T$_3$	9.30±0.20^f	5.35±0.28^{de}	182.79±8.32^{de}	9.50±0.51^{bcde}
T$_4$	9.90±0.19^{de}	5.34±0.17^{de}	176.41±3.77^e	9.21±0.45^{de}
T$_5$	11.17±0.21^a	6.36±0.27^a	197.69±1.94^{ab}	10.42±0.19^{abc}
T$_6$	10.46±0.18^b	5.70±0.04^{bc}	188.40±2.30^{cd}	10.50±0.56^{ab}
T$_7$	10.41±0.09^{bc}	5.60±0.06^{cd}	179.23±2.37^e	11.08±0.73^a
T$_8$	10.11±0.28^{cd}	5.62±0.07^{cd}	200.17±0.84^a	11.17±0.57^a
T$_9$	10.87±0.16^a	5.96±0.08^b	187.93±3.39^{cd}	11.42±0.62^a
T$_{10}$	9.67±0.14^e	5.18±0.21^{ef}	177.33±1.82^e	9.83±0.90^{cde}

T$_1$:100% RDF alone, **T$_2$:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_3$:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_4$:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_5$:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_6$:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_7$:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_8$:** 75% RDF + two foliar spray of micronutrients, **T$_9$:** 50% RDF + two foliar spray of micronutrients, **T$_{10}$:** 25% RDF+ two foliar spray of micronutrients.

Table 5. Influence of macro and micronutrients on soil properties (pH, organic carbon, EC (dS m^{-1}), available nitrogen, phosphorus, and potassium) in a mango cv. Dashehari under medium density planting (**Mean of two years**).

Treatments #	pH	Organic carbon (%)	EC (dS m^{-1})	Available Nitrogen (kg ha^{-1})	Available Phosphorus (kg ha^{-1})	Available Potassium (kg ha^{-1})
T$_1$	7.47±0.03^a	0.83±0.01^{abcd}	0.33±0.02^a	202.34±1.04^{ab}	21.75±0.14^b	146.06±2.07^{abc}
T$_2$	7.30±0.04^{cde}	0.82±0.02^d	0.26±0.03^{cde}	200.78±1.23^b	21.00±0.13^c	144.24±1.07^{cde}
T$_3$	7.24±0.04^e	0.84±0.01^{abc}	0.30±0.01^b	201.62±1.08^{ab}	21.62±0.14^b	145.09±1.66^{bcd}
T$_4$	7.27±0.01^{de}	0.83±0.01^{cd}	0.24±0.02^e	197.48±0.02^c	21.05±0.19^c	143.73±1.63^{cde}
T$_5$	7.28±0.05^{de}	0.85±0.01^a	0.29±0.00^{bc}	200.91±0.84^b	21.76±0.25^{ab}	147.22±1.52^{ab}
T$_6$	7.37±0.02^b	0.83±0.01^{abcd}	0.29±0.01^b	202.55±0.72^{ab}	21.04±0.15^c	144.47±1.48^{cde}
T$_7$	7.37±0.03^b	0.83±0.01^{bcd}	0.29±0.01^b	195.96±1.34^c	21.76±0.07^{ab}	141.81±0.98^e
T$_8$	7.35±0.04^{bc}	0.83±0.00^{bcd}	0.27±0.02^{bcd}	203.25±1.28^a	22.02±0.01^a	147.87±1.22^a
T$_9$	7.31±0.04^{cd}	0.85±0.01^{ab}	0.28±0.02^{bcd}	202.00±1.35^{ab}	21.79±0.13^{ab}	146.30±0.50^{abc}
T$_{10}$	7.27±0.03^{de}	0.82±0.01^d	0.26±0.01^{de}	197.23±0.50^c	20.95±0.06^c	142.93±1.66^{de}

T$_1$:100% RDF alone, **T$_2$:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_3$:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_4$:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T$_5$:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_6$:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_7$:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T$_8$:** 75% RDF + two foliar spray of micronutrients, **T$_9$:** 50% RDF + two foliar spray of micronutrients, **T$_{10}$:** 25% RDF+ two foliar spray of micronutrients.

743 **Table 6.** Influence of macro and micronutrients on Fe, Zn, Cu, B & Ca content of soil in mango
744   cv. Dashehari under medium density planting (**Mean of two years**).

Treatments #	Fe (mg kg^{-1})	Zn (mg kg^{-1})	Cu (mg kg^{-1})	B (mg kg^{-1})	Ca (mg kg^{-1})
T_1	5.51 ± 0.06^b	0.54 ± 0.03^d	0.18 ± 0.01^{ef}	0.49 ± 0.02^{cd}	888.00 ± 2.65^h
T_2	5.57 ± 0.06^b	0.69 ± 0.03^{bc}	0.17 ± 0.01^f	0.51 ± 0.02^{bc}	925.67 ± 0.58^f
T_3	6.11 ± 0.27^a	0.68 ± 0.03^{bc}	0.20 ± 0.01^{bcd}	0.51 ± 0.01^{bc}	1011.00 ± 1.73^c
T_4	5.65 ± 0.26^b	0.69 ± 0.04^{ab}	0.19 ± 0.01^{de}	0.51 ± 0.02^{bc}	926.00 ± 1.00^f
T_5	5.40 ± 0.20^b	0.78 ± 0.06^a	0.22 ± 0.01^{ab}	0.54 ± 0.02^a	1045.67 ± 2.08^a
T_6	6.15 ± 0.09^a	0.77 ± 0.05^{ab}	0.24 ± 0.02^a	0.52 ± 0.01^{ab}	968.33 ± 0.58^e
T_7	6.09 ± 0.26^a	0.76 ± 0.03^{ab}	0.21 ± 0.01^b	0.52 ± 0.02^{ab}	1019.67 ± 0.58^b
T_8	5.49 ± 0.26^b	0.61 ± 0.06^{cd}	0.18 ± 0.01^{ef}	0.48 ± 0.02^{de}	1005.33 ± 3.21^d
T_9	5.69 ± 0.19^b	0.58 ± 0.06^d	0.21 ± 0.01^{bc}	0.47 ± 0.02^{de}	1010.67 ± 1.53^c
T_{10}	5.62 ± 0.07^b	0.56 ± 0.05^d	0.19 ± 0.01^{cde}	0.46 ± 0.02^e	905.67 ± 4.73^g

T_1:100% RDF alone, T_2: 75% RDF + soil micronutrients + one foliar spray of micronutrients, T_3: 50% RDF + soil micronutrients + one foliar spray of micronutrients, T_4: 25% RDF + soil micronutrients + one foliar spray of micronutrients, T_5: 75% RDF + soil micronutrients + two foliar spray of micronutrients, T_6: 50% RDF + soil micronutrients + two foliar spray of micronutrients, T_7: 25% RDF + soil micronutrients + two foliar spray of micronutrients, T_8: 75% RDF + two foliar spray of micronutrients, T_9: 50% RDF + two foliar spray of micronutrients, T_{10}: 25% RDF+ two foliar spray of micronutrients.

748   **Table 7.** Pearson correlation coefficients among fruit yield, growth, soil, and plant nutrient variables in medium-density mango orchards.

Variable	Fruit yield (kg/plant)	Canopy volume	Fruit weight	Fruit retention (%)	Shelf life (days)	Leaf N	Leaf P	Leaf K	Leaf Fe	Leaf Zn	Soil N	Soil P	Soil K	Soil Fe	Soil Zn
Fruit yield (kg/plant)	1.000	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Canopy volume (m³)	0.41	1.000	-	-	-	-	-	-	-	-	-	-	-	-	-
Fruit weight (g)	0.58**	0.52**	1.000	-	-	-	-	-	-	-	-	-	-	-	-
Fruit retention (%)	0.37	0.35	0.36	1.000	-	-	-	-	-	-	-	-	-	-	-
Shelf life (days)	0.49*	0.33	0.42	-0.38	1.000	-	-	-	-	-	-	-	-	-	-
Leaf N (%)	0.26	0.31	0.30	0.25	0.32	1.000	-	-	-	-	-	-	-	-	-
Leaf P (%)	0.49*	0.45*	0.40	0.50*	0.40	0.22	1.000	-	-	-	-	-	-	-	-
Leaf K (%)	0.29	0.33	0.31	0.23	0.42	0.31	0.27	1.000	-	-	-	-	-	-	-
Leaf Fe (mg/kg)	0.37	0.39	0.48*	0.20	0.35	0.17	0.34	0.19	1.000	-	-	-	-	-	-
Leaf Zn (mg/kg)	0.18	0.28	0.30	0.00	0.61**	0.42	0.28	0.17	0.24	1.000	-	-	-	-	-
Soil N (kg/ha)	0.14	0.31	0.28	–0.02	0.17	0.27	0.36	-0.00	0.51*	-0.10	1.000	-	-	-	-
Soil P (kg/ha)	0.58**	0.47*	0.45*	0.04	0.48*	0.35	0.47*	0.04	0.49*	0.05	0.49*	1.000	-	-	-
Soil K (kg/ha)	0.26	0.33	0.30	–0.10	0.42	0.21	0.25	–0.11	0.58**	-0.10	0.58**	0.48*	1.000	-	-
Soil Fe (mg/kg)	0.20	0.22	0.21	0.26	0.26	0.14	0.24	0.05	0.41	0.26	0.20	0.18	-0.46	1.000	-
Soil Zn (mg/kg)	0.48*	0.39	0.52**	0.69**	0.69**	0.53*	0.68**	0.15	0.43	0.69**	0.26	0.69**	–0.66**	-0.46	1.000

749   **Note:** * and ** represent significance at $P < 0.05$ and $P < 0.01$, respectively.

Figure 1. Experimental Research Unit at HRC, Pattarchatta, U.S. Nagar, Uttarakhand (India).

Figure 2. Planting layout of mango (*Mangifera indica* L.) cv. Dashehari under INM practices in medium-density planting. T₁:100% RDF alone, **T₂:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T₃:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T₄:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T₅:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T₆:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T₇:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T₈:** 75% RDF + two foliar spray of micronutrients, **T₉:** 50% RDF + two foliar spray of micronutrients, **T₁₀:** 25% RDF+ two foliar spray of micronutrients.

Figure 3. Conceptual experimental workflow for the integrated nutrient management study in medium-density mango orchards.

Figure 4. Overview of the doses of fertilizer, time, and application methods (*Mangifera indica* L.) cv. Dashehari for mango Crops under medium density planting. T_1:100% RDF alone, T_2: 75% RDF + soil micronutrients + one foliar spray of micronutrients, T_3: 50% RDF + soil micronutrients + one foliar spray of micronutrients, T_4: 25% RDF + soil micronutrients + one foliar spray of micronutrients, T_5: 75% RDF + soil micronutrients + two foliar spray of micronutrients, T_6: 50% RDF + soil micronutrients + two foliar spray of micronutrients, T_7: 25% RDF + soil micronutrients + two foliar spray of micronutrients, T_8: 75% RDF + two foliar spray of micronutrients, T_9: 50% RDF + two foliar spray of micronutrients, T_{10}: 25% RDF+ two foliar spray of micronutrients.

Figure 5. Overview of soil sample collection (0-30 cm depth) and plant sample collection from 4-5-month-old mature mango leaves.

Figure 6. **Influence of INM practices on yield and B:C Ratio of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting (Mean of two years). T_1:**100% RDF alone, **T_2:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T_3:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T_4:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T_5:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T_6:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T_7:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T_8:** 75% RDF + two foliar spray of micronutrients, **T_9:** 50% RDF + two foliar spray of micronutrients, **T_{10}:** 25% RDF+ two foliar spray of micronutrients.

(a) Total Nitrogen in leaves (%)

(b) Total Phosphorus in leaves (%)

(c) Total Potassium in leaves (%)

Figure 7. Influence of INM practices on leaf macronutrient (a) nitrogen, (b) phosphorus, and (c) Potassium content of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting (Mean of two years). T_1:100% RDF alone, T_2: 75% RDF + soil micronutrients + one foliar spray of micronutrients, T_3: 50% RDF + soil micronutrients + one foliar spray of micronutrients, T_4: 25% RDF + soil micronutrients + one foliar spray of micronutrients, T_5: 75% RDF + soil micronutrients + two foliar spray of micronutrients, T_6: 50% RDF + soil micronutrients + two foliar spray of micronutrients, T_7: 25% RDF + soil micronutrients + two foliar spray of micronutrients, T_8: 75% RDF + two foliar spray of micronutrients, T_9: 50% RDF + two foliar spray of micronutrients, T_{10}: 25% RDF+ two foliar spray of micronutrients.

| (a) Total iron and Zinc in leaves (ppm) | (b) Total Copper in leaves (ppm) |

| (c) Total Boron in leaves (ppm) | (d) Total Calcium in leaves (ppm) |

Figure 8. Influence of INM practices on leaf micronutrient (a) iron & zinc (b) copper (c) boron (d) calcium content of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting (Mean of two years). T_1:100% RDF alone, T_2: 75% RDF + soil micronutrients + one foliar spray of micronutrients, T_3: 50% RDF + soil micronutrients + one foliar spray of micronutrients, T_4: 25% RDF + soil micronutrients + one foliar spray of micronutrients, T_5: 75% RDF + soil micronutrients + two foliar spray of micronutrients, T_6: 50% RDF + soil micronutrients + two foliar spray of micronutrients, T_7: 25% RDF + soil micronutrients + two foliar spray of micronutrients, T_8: 75% RDF + two foliar spray of micronutrients, T_9: 50% RDF + two foliar spray of micronutrients, T_{10}: 25% RDF+ two foliar spray of micronutrients.

Figure 9. Path diagram showing direct and indirect effects of soil nutrients, plant nutrients, and plant growth on fruit yield of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting (Mean of two years). T_1:100% RDF alone, **T_2:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T_3:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T_4:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T_5:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T_6:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T_7:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T_8:** 75% RDF + two foliar spray of micronutrients, **T_9:** 50% RDF + two foliar spray of micronutrients, **T_{10}:** 25% RDF+ two foliar spray of micronutrients.

Figure 10. PCA biplot showing the relationship of INM treatments with soil nutrients, leaf nutrient status, growth, yield, and fruiting attributes of mango (*Mangifera indica* L.) cv. Dashehari under medium-density planting. T₁:100% RDF alone, **T₂:** 75% RDF + soil micronutrients + one foliar spray of micronutrients, **T₃:** 50% RDF + soil micronutrients + one foliar spray of micronutrients, **T₄:** 25% RDF + soil micronutrients + one foliar spray of micronutrients, **T₅:** 75% RDF + soil micronutrients + two foliar spray of micronutrients, **T₆:** 50% RDF + soil micronutrients + two foliar spray of micronutrients, **T₇:** 25% RDF + soil micronutrients + two foliar spray of micronutrients, **T₈:** 75% RDF + two foliar spray of micronutrients, **T₉:** 50% RDF + two foliar spray of micronutrients, **T₁₀:** 25% RDF+ two foliar spray of micronutrients.

Graphical abstract