Bridging the gap between genome-wide association studies and network medicine with GNExT

Lis Arend⬡,[1,2,∗] **Fabian Woller**⬡,[2,3] **Bastienne Rehor,**[1] **David Emmert**⬡,[2] **Johannes Frasnelli**⬡,[4] **Christian Fuchsberger**⬡,[2] **David B. Blumenthal**⬡[3] **and Markus List**⬡[1,5,∗]

[1]Data Science in Systems Biology, TUM School of Life Sciences, Technical University of Munich, Freising, Germany

[2]Institute for Biomedicine, Eurac Research, Bolzano, Italy

[3]Biomedical Network Science Lab, Department Artificial Intelligence in Biomedical Engineering, Friedrich-Alexander-Universität Erlangen-Nürnberg, Erlangen, Germany

[4]Department of Anatomy, Université du Québec à Trois-Rivières, Trois-Rivières, Canada

[5]Munich Data Science Institute, Technical University of Munich, Garching, Germany

∗Corresponding authors. lis.arend@tum.de, markus.list@tum.de

Abstract

Motivation: A growing volume of large-scale genome-wide association study (GWAS) datasets offers unprecedented power to uncover the genetic determinants of complex traits, but existing web-based platforms for GWAS data exploration provide limited support for interpreting these findings within broader biological systems. Systems medicine is particularly well-suited to fill this gap, as its network-oriented view of molecular interactions enables the integration of genetic signals into coherent network modules, thereby opening opportunities for disease mechanism mining and drug repurposing.

Results: We introduce GNExT (GWAS network exploration tool), a web-based platform that moves beyond the variant-level effect and significance exploration provided by existing solutions. By including MAGMA and Drugst.One, GNExT allows its users to study genetic variants on the network level down to the identification of potential drug repurposing candidates. Moreover, GNExT advances over the current state of the art by offering a highly standardized Nextflow pipeline for data import and preprocessing, allowing researchers to easily deploy their study results on a web interface. We demonstrate the utility of GNExT using a genome-wide association meta-analysis of human olfactory identification, in which the framework translated isolated GWAS signals to potential pharmacological targets in human olfaction.

Availability and Implementation: The complete GNExT ecosystem, including the Nextflow preprocessing pipeline, the backend service, and frontend interface, is publicly available on GitHub (`https://github.com/dyhealthnet/gnext_nf_pipeline`, `https://github.com/dyhealthnet/gnext_platform`). The public instance of the GNExT platform on olfaction is available under `http://olfaction.gnext.gm.eurac.edu`.

Contact: lis.arend@tum.de, markus.list@tum.de

Keywords genome-wide association studies, network medicine, Nextflow, web interface, data exploration

1. Introduction

Advances in genotyping, together with widespread electronic health records and large population-based cohorts, have transformed the scale of genetic studies (Linder *et al.*, 2021). Modern datasets now comprise genetic profiles spanning millions of loci for millions of individuals across thousands of traits, enabling genome-wide association studies (GWAS) of unprecedented breadth (Uffelmann *et al.*, 2021). These high-dimensional GWAS resources underpin diverse downstream analyses and have substantially advanced our understanding of the genetic architecture and biological mechanisms of human traits and diseases (Uffelmann *et al.*, 2021; Visscher *et al.*, 2017; Sollis *et al.*, 2023).

Despite the growth of GWAS data, standard single variant association tests fail to fully capture the genetic architecture of complex traits, which are highly polygenic and driven by numerous variants of small individual effects (Stringer *et al.*, 2011; de Leeuw *et al.*, 2015; Wang *et al.*, 2011; Park *et al.*, 2011). To control genome-wide false positives under this high-dimensional testing burden, very stringent significance thresholds are required, which severely reduce power to detect weak but genuine associations unless sample sizes reach several hundred thousand individuals (Wang *et al.*, 2011; Gibson, 2010; Park *et al.*, 2011). To address these limitations and capture the biological architecture underlying complex traits, gene-based approaches have gained prominence (Neale and Sham, 2004; Kontou and Bagos, 2024). By aggregating numerous

weak variant-level signals within genes or genomic regions into composite gene-level statistics, these methods reduce the multiple testing burden, capture combined genetic effects, and increase power to detect genuine disease contributors (Neale and Sham, 2004; de Leeuw *et al.*, 2015). While methods such as MAGMA (de Leeuw *et al.*, 2015), VEGAS (Liu *et al.*, 2010), and PascalX (Krefl *et al.*, 2023), enhance statistical power, they primarily identify associations without fully resolving the biological context underlying these signals. Bridging this gap requires combining association results with mechanistic insights, a task for which systems medicine and network-based approaches provide essential frameworks (Galindez *et al.*, 2023).

In systems medicine, increasing attention is being directed toward the observation that disease-associated genes are not randomly distributed in biological networks but cluster within specific regions of the human interactome. These genes, typically termed seed genes, and the interactions of their encoded proteins with neighboring molecules are used to expand the initial seed set and define disease modules that represent the underlying pathogenic mechanisms (Menche *et al.*, 2015; Galindez *et al.*, 2023). Beyond providing mechanistic insight, disease modules form the basis for network pharmacology strategies that systematically identify therapeutic targets by considering both seed and neighboring nodes (Nogales *et al.*, 2022). This enables the discovery of targets that can be modulated by existing drugs and thus supports drug repurposing, an attractive alternative to de novo drug development given its reduced cost and shorter timelines (Pushpakom *et al.*, 2019; Krishnamurthy *et al.*, 2022; Hopkins, 2008).

In parallel, the landscape of post-GWAS analyses has rapidly evolved to accommodate the increasing complexity and diversity of genetic research needs (Kontou and Bagos, 2024). Broadly, existing resources fall into three classes. First, online post-GWAS analysis platforms such as FUMA (Watanabe *et al.*, 2017), postGWAS (Wang *et al.*, 2024), GWAShug (Cao *et al.*, 2025), and CTG-VL (Cuellar-Partida *et al.*, 2019), allow users to upload, explore, and interpret summary statistics on a per-trait basis. FUMA, specifically, is a web-based interface enabling the comprehensive functional mapping and annotation of GWAS results by integrating multiple biological databases and supporting users in annotating, prioritizing, visualizing, and interpreting associations for individual phenotypes. Notably, platforms such as FUMA and CTG-VL also offer gene-based association testing with MAGMA and fastBAT (Bakshi *et al.*, 2016), respectively. Several of these platforms also host curated analyses in which post-GWAS results are made publicly accessible. Nevertheless, they are designed for post-GWAS analyses on a single trait at a time, reflecting their emphasis on detailed, phenotype-specific investigation. The second category encompasses global or study-specific software tools primarily designed for GWAS data dissemination. While the GWAS Catalog (Sollis *et al.*, 2023) and openGWAS (Elsworth *et al.*, 2020) serve as central repositories for curated GWAS results, the GWAS Atlas (Liu *et al.*, 2023) extends functionality by enabling interactive exploration of curated GWAS and post-GWAS results. Study-specific portals, including the GWAS Explorer (Machiela *et al.*, 2023) and the collection of Human Genetics Amplifier (HuGeAMP) knowledge portals, such as the Cardiovascular Disease Knowledge Portal (Costanzo *et al.*, 2023), are designed to focus on specific cohorts or disease areas. The third category consists of frameworks like PheWeb (Gagliano Taliun *et al.*, 2020) and its successor PheWeb 2 (Bellavance *et al.*, 2026), which enable the exploration of large-scale, population-specific GWAS datasets through interactive visualization, navigation, and publication of GWAS and PheWAS results. A distinct feature of PheWeb is its support for deploying independent instances, allowing researchers to operate the platform on their own datasets without the need to redesign study-specific frameworks. This capability has already supported several large-scale Biobank and population genetics resources (Bycroft *et al.*, 2018; Kurki *et al.*, 2023; Zawistowski *et al.*, 2023; Nagai *et al.*, 2017). While PheWeb 2 extends the original PheWeb by functionalities enabling exploration and comparison of stratified GWAS summary-level data, further downstream post-GWAS analyses, such as gene-based tests, are not integrated.

Despite the breadth of available tools operating on GWAS summary statistics, most platforms remain centered on trait-specific visualization and exploration, offering limited integration of systems-level concepts. As a result, these tools provide limited insight into how genetic associations converge into coherent biological processes, and do not support the translation of these mechanistic insights into therapeutic hypothesis generation, an area where network medicine offers substantial advantages.

Building on principles of interactive, scalable, and decentralized deployment, we introduce GNExT (GWAS Network Exploration Tool), a framework that extends this paradigm by integrating GWAS summary statistics with network medicine methodologies (Figure 1). A dedicated Nextflow-based preprocessing pipeline, which processes GWAS data together with user-defined configuration parameters, substantially streamlines and automates data preparation, thereby enabling data extensions beyond existing deployments. Variant-level association statistics are aggregated into gene-level metrics using MAGMA, and all generated output files are consolidated into a directory that serves as the primary input for deploying the GNExT platform. The GNExT platform can be deployed via a docker-compose stack and provides variant-, gene-, and trait-centric pages for exploratory analyses. Potential risk genes can be identified by applying significance thresholds to gene-level P-values. These genes serve as seed nodes for detecting disease modules in protein-protein interaction networks through the Drugst.One interface (Maier *et al.*, 2024). GNExT thus facilitates the elucidation of candidate disease mechanisms and the prioritization of potential therapeutic targets directly from GWAS summary statistics.

2. Results

2.1. Nextflow preprocessing pipeline allows for efficient and flexible data preparation

Our Nextflow-based pre-processing pipeline (Di Tommaso *et al.* (2017)) provides a modular and extensible workflow architecture that orchestrates and parallelizes workflow execution and allows resuming computations after an error without repeating previously completed, computationally intensive steps. Likewise, it supports incremental dataset expansion by enabling the

Figure 1 Overview of the GNExT framework. Input data consists of GWAS summary statistics together with study-specific configuration parameters. A Nextflow-driven preprocessing pipeline transforms and organizes the data in a scalable way, ultimately generating the primary input directory required for platform deployment. GNExT extends its functionalities beyond data exploration by linking GWAS results to network medicine through gene-based analyses and an integrated, interactive mechanism mining interface. Created with BioRender.com.

addition of new traits while reusing previously generated trait-specific results, avoiding redundant computation. The pipeline is widely usable as it supports local execution through Conda, containerized environments via Docker or Singularity, and distributed execution using SLURM-based HPC systems, ensuring flexibility, scalability, and reproducibility for large-scale genomic analyses.

The Nextflow pipeline prepares GWAS summary data for integration into the GNExT platform, including data harmonization, variant annotation using the Ensembl Variant Effect Predictor (VEP) (McLaren *et al.* (2016)), and data processing into Lightning Memory-Mapped Database (LMDB) structures to enable efficient downstream querying (Figure 2). In addition, analogous to PheWeb, the pipeline generates preprocessed inputs for standard visualizations and tabular outputs, including Manhattan, Q-Q, and PheWAS plots as well as a Top Hits table. An important step in the pipeline is the use of MAGMA for variant-gene-mapping, which allows us to identify candidate risk genes for downstream network medicine analysis, such as module discovery and drug repurposing (see Methods for details).

2.2. GNExT application connects GWAS with network medicine in a decentralized manner

The GNExT backend and frontend receive the output of the Nextflow pre-processing pipeline along with a set of configuration parameters (see Methods section).

With GNExT, users can query variant-, trait-, or gene-level result pages, analogous to PheWeb (Supplementary Figure 1A). In addition, a dedicated Top Hits page summarizes the most significant variants identified across all traits represented within the platform, offering a concise overview of key associations. The variant-specific page provides detailed Ensembl VEP annotations, allele frequency comparisons with 1000 Genomes (1000 Genomes Project Consortium *et al.*, 2015) and gnomAD (Chen *et al.*, 2024), cross-references to external databases, and nearest-gene assignments (Supplementary Figure 1B). An

integrated LocusZoom-based PheWAS view (Boughton *et al.*, 2021), accompanied by a summary table, presents all trait associations for the variant with interactive filtering options. Within the interface, selecting a gene symbol redirects to the gene-specific page, which lists the most significant proximal variant for each trait and provides a trait-specific LocusZoom plot displaying regional association signals, LD patterns, and gene annotations (Supplementary Figure 1C).

Moreover, the trait-specific page visualizes all association signals from a single GWAS through interactive Manhattan and Q–Q plots and includes an extended Top Hits table with newly introduced retrieval options that filter results by P-value, genomic window, or chromosomal region. Gene-level associations from MAGMA are displayed analogously, and significant genes (default Bonferroni-corrected P-value < 0.05) can be selected as seeds and transferred to the network medicine interface for downstream analyses (Supplementary Figure 2A).

The network medicine-specific page allows users to select a predefined seed gene list and transfer the seed genes to the integrated Drugst.One interface (Supplementary Figure 2B). Drugst.One provides tools for disease module refinement, drug repurposing, and additional analyses such as functional enrichment (see Methods). Users may choose among multiple biological network sources, including seven protein-protein, five drug-protein, three protein-disease, and three drug-disease interaction databases, with NeDRExxx (Sadegh *et al.* (2021)) selected by default as this knowledge graph integrates more than twenty different data sources on drug, gene, protein, and other interactions. Users can also navigate to the gene-specific pages to inspect GWAS signals for seed genes or newly identified module genes.

2.3. Linking olfactory GWAS signals to systems-level mechanisms with GNExT

To demonstrate how GNExT bridges the gap between statistical associations and mechanistic understanding, we applied the framework to a recent GWAS meta-analysis of human olfactory

Figure 2 Overview of the Nextflow-based preprocessing pipeline. The pipeline comprises two main stages. First, GWAS data are harmonized, and a reference VCF containing all unique variants is assembled for downstream annotation. Two LMDB databases are generated to store variant-rsID and variant-gene mappings. The workflow also produces JSON files for Manhattan and Q-Q plots, a Top Hits table, and per-chromosome data files for efficient variant-level retrieval. If enabled by user configuration, MAGMA input data are prepared, and gene-level association statistics are computed. All outputs generated by the preprocessing pipeline are consolidated into a deployment directory, which forms the essential data backbone for launching a GNExT instance.

identification (Förster *et al.*, 2025). While GWAS successfully identifies genetic loci associated with traits, interpreting these findings in the context of biological systems remains a challenge. Here, we utilized GNExT to move beyond isolated variant lists and reconstruct the signaling pathways and potential drug interactions underlying olfactory function.

We utilized summary statistics from the "score_all" phenotype ($n = 21,495$), a global measure of olfactory ability derived from the Sniffin' Sticks identification test. Initial exploration via GNExT confirmed the strong, expected signal on chromosome 11 (designated as locus 8 in Förster *et al.* (2025)) (Figure 3A). To resolve these variant-level signals into

interpretable biological units, we employed MAGMA within the Nextflow preprocessing pipeline, identifying 33 significant seed genes (Bonferroni-corrected P-value < 0.05), predominantly in olfactory receptor (OR) gene families OR5 (17 genes), OR8 (11 genes), and OR9 (2 genes), with additional associations in OR10AG1, OR9G4, and TRIM51 (Supplementary Figure 2A and Supplementary Table 1). However, examining receptors in isolation offers limited insight into the downstream signaling machinery that translates chemical binding into neuronal perception.

To reconstruct the functional context of these receptors, we transferred these 33 (seed) genes into the Drugst.One network medicine environment within GNExT. Three of the 33 genes were not present in the underlying NeDReX database (Sadegh *et al.*, 2021) and therefore could not be incorporated (Supplementary Figure 2B and Supplementary Table 1). We hypothesized that while ORs drive the genetic signal, they must physically interact with downstream signal transducers to effect perception. Using the "Quick drug search" option, triggering the sequential execution of the Multi-Steiner tree module detection and closeness centrality algorithm for drug prioritization, we identified a coherent network module comprising 13 out of the 30 integrated seed genes and 20 additional connector genes, i.e., proteins not significant in the GWAS itself but essential for maintaining network integrity (Figure 3B). Functional enrichment of the network module with g.Profiler (Raudvere *et al.*, 2019) highlighted the significant enrichment in molecular function (MF) and biological processes (BP) terms related to olfaction, including "Odorant binding", "Olfactor receptor activity", and "Detection of chemical stimulus involved in sensory perception of smell" (Figure 3C). Consistent with expectations, all seed genes except TRIM51 contributed to the olfaction-associated enriched terms (Figure 3D). In turn, connector genes such as FLT3, AURKB, PRKAA1, or FFAR2 contributed to terms associated with olfactory transduction downstream from odorant binding, such as "G-Protein coupled receptor signaling pathways", "molecular transducer activity" or "signal transduction". This aligns with the known biology of olfactory perception, where ORs (GPCRs) rely on downstream cascades involving adenylate cyclases and kinase activity to transmit signals to the olfactory bulb.

Finally, we asked whether this systems-level view could generate actionable pharmacological insights. By querying the expanded network for drug-gene interactions using harmonic centrality, we identified 39 drugs targeting the connector genes, specifically FLT3, AURKB, PRKAA1, or FFAR2 (Figure 3). In our network, FLT2, AURKB, and PRKAA1 were connected to OR8U1, OR9G1, and OR5M3, respectively. Of these, only OR5M3 has been deorphanized and responds to furaneol, which gives different foods a caramel-note (Haag *et al.*, 2021). The drugs connected to FLT2, AURKB, and PRKAA1 belong to the group of tyrosine kinase inhibitors (TKI), which target tyrosine kinases that govern cell growth and division and are used in the treatment of various types of cancer (Su *et al.*, 2023; Yu *et al.*, 2023; Ou *et al.*, 2024). Crucially, this network-derived finding offers a mechanistic hypothesis for a known clinical phenomenon: TKIs are associated with olfactory dysfunction such as anosmia, hyposmia, and parosmia (Debbaneh *et al.*, 2023; Fu *et al.*, 2025; van Elst *et al.*, 2022; Barbieri *et al.*, 2023). Furthermore, tyrosine kinase levels in the olfactory

Figure 3 Results of the olfaction use case. (A) Manhattan plot from the GNExT olfaction instance, revealing a strong genome-wide significant association signal on chromosome 11. (B) Network module inferred using a Multi-Steiner Tree algorithm, consisting of MAGMA-identified significant genes (seed genes) and interacting genes required to connect the seed genes (connector genes). Drug prioritization was performed using harmonic centrality. (C) Functional enrichment analysis of the network module genes using g.Profiler, with significantly enriched terms ranked by intersection size, bars colored according to the $-log_{10}$ adjusted P-value, and the corresponding adjusted P-values displayed within the bars. (D) Mapping of seed and connector genes to the enriched functional terms identified in (C).

bulb play a role in odor memory consolidation, which may be a potential pathway by which these drugs affect olfactory perception (Tong *et al.*, 2018). Through this workflow, GNExT successfully translated raw GWAS summary statistics into a plausible mechanistic hypothesis, linking genetic variants to a specific class of drugs known to perturb the phenotype.

3. Discussion

The GNExT ecosystem provides researchers with a streamlined framework for deploying a web-based platform that enables comprehensive exploration and visualization of GWAS summary statistics, while incorporating network medicine methodologies. Specifically, through the application of MAGMA and Drugst.One, GNExT is the first web-based platform that connects GWAS data and network medicine. To enable an interactive interface, minimize deployment effort, and thereby attract a broader user base, we developed a Nextflow-based pipeline for data preparation and preprocessing. Through Nextflow's built-in workflow orchestration and caching mechanisms, all pipeline components are executed efficiently and reproducibly, enabling both streamlined deployment and efficient extension of existing platforms.

By applying GNExT to a GWAS meta-analysis of human olfactory identification, we demonstrate that the framework facilitates the interpretation of variant-level association signals by contextualizing them into signaling pathways and prioritizing putative drug-target interactions relevant to olfactory function.

However, the GNExT ecosystem also comes with certain limitations: For the execution of the gene-based testing that ensures the application of network medicine approaches, we selected MAGMA as a state-of-the-art tool and focused on the simplest variant-to-gene mapping strategy. However, small genomic windows fail to capture long-range regulatory interactions, which can span hundreds of kilobases and are increasingly recognized as critical contributors to gene regulation (Groenewoud *et al.*, 2022). In contrast, expanding flanking regions introduces substantial noise by assigning variants to unrelated genes, thereby diluting true biological signals. Integrating experimentally derived regulatory interaction datasets, such as eQTL or Hi-C datasets, can mitigate these issues (Groenewoud *et al.*, 2022). Consequently, enabling more flexible, user-defined variant-to-gene mapping strategies represents an important avenue for enhancing future versions of GNExT.

Moreover, the current implementation supports the selection of only a single reference population across all traits. For large-scale studies, enabling trait-specific reference population selection would provide greater flexibility.

Finally, the exploration and visualization of stratified GWAS results, as supported in PheWeb 2 (**?**), are not yet available with GNExT. The ability to compare network medicine outcomes across stratified analyses would be of particular interest and represents a promising direction for future development.

4. Methods

4.1. Platform input

The primary input to GNExT's preprocessing pipeline is a CSV phenotype file specifying the traits to be analyzed, including the file paths to the corresponding GWAS summary statistics. GWAS files based on either GRCh37 or GRCh38 can be used, since reference files are provided for both genome assemblies. The preprocessing pipeline additionally requires study-specific and computational configurations: the study-specific configurations define parameters such as the output

directory, the structure of the GWAS summary statistics (via column indices), VEP annotation settings, and MAGMA parameters; the computational configurations specify memory and CPU allocations for each workflow component.

For the deployment of the GNExT platform after preprocessing, only a study-specific .env file must be defined. This file specifies absolute paths to the phenotype file, output directory of the preprocessing pipeline, the Typesense volume, and core service parameters such as the Typesense host and port. It also controls feature flags (e.g., activation of MAGMA components), theming parameters, study identifiers, and citation metadata. Optional predefined example traits, variants, and genes can be included to guide users during initial exploration. These configuration options ensure that the platform can be adapted to diverse research contexts while preserving a consistent and user-friendly interface. Full descriptions of all configuration parameters for both preprocessing and deployment are available in the corresponding GitHub repositories.

4.2. Harmonization of GWAS data formats

To accommodate diverse input formats, the first stage of the preprocessing pipeline harmonizes GWAS summary statistics. This step uses the Python package ZORP (`https://github.com/statgen/zorp`), which parses the GWAS data and abstracts format-specific differences. ZORP also exports the harmonized data as tab-separated, gzipped files, while variant sorting and indexing are handled in bash. The resulting harmonized dataset provides a standardized foundation for all downstream processing steps, ensuring independence from the original file structure.

4.3. Variant annotation using Ensembl VEP

Prior to variant annotation, the pipeline generates batch-specific VCF files for subsets of traits, which are subsequently merged into a single reference VCF containing all unique variants. This strategy remains efficient even when processing thousands of traits and ensures that each variant is annotated only once rather than repeatedly across individual GWAS files. For annotation, users may either supply a path to an existing local VEP cache or allow the pipeline to download it automatically. Cache retrieval and annotation rely on the nf-core modules `ensemblvep_download` and `ensemblvep_vep` (Ewels *et al.*, 2020). We then populate two LMDB databases that function as on-demand lookup tables for the GNExT instances. The first LMDB database maps genetic variants to their corresponding rsIDs using the annotated VCF file. This avoids the need to append rsID annotations directly to every GWAS file, a process that can take several hours for large trait collections. A second LMDB database stores variant-to-gene mappings, recording for each variant the genes it overlaps or those located within a user-defined window. To ensure consistency, we use the gene coordinates of protein-coding genes from the same Ensembl release as VEP. For GRCh37 and GRCh38, gene annotation data is available for the two most recent Ensembl releases (114 and

115), and a script is provided to generate this data for other releases.

4.4. Features for result exploration

The generation of Manhattan and Q-Q plots within the GNExT framework builds upon the implementation provided by PheWeb, which produces the required visualization data in JSON format prior to platform deployment. Top genetic associations are extracted, whereas non-significant variants are aggregated into bins. This binning strategy substantially reduces the volume of data, compressing millions of variant-level values into manageable aggregates, thereby improving data transfer efficiency during API calls. The Top Hits table is generated by collecting all non-binned variants across traits, ranking them by P-value, and truncating the list according to a user-defined threshold. The resulting data are exported in JSON format for integration into the platform. Significance thresholds for Manhattan plots and criteria for defining Top Hits are fully configurable. Additionally, the pipeline produces BGZF-compressed and accompanying TBI-indexed files for each chromosome and each metric (P-value, allele frequency, effect size, and standard error). These indexed files support efficient variant-level data retrieval over all traits, required for PheWAS plots and variant-specific summary tables.

4.5. Gene-based association analysis

Gene-based association analysis is performed using MAGMA, aggregating single-variant summary statistics into gene-level P-values. We employ MAGMA's default SNP-wise model, which uses the sum of squared SNP Z-statistics as the gene-level test statistic. To account for linkage disequilibrium (LD) among genetic variants, while avoiding the need for study-specific individual-level genotype data, LD structure is typically estimated using the 1000 Genomes Phase 3 reference dataset (1000 Genomes Project Consortium et al., 2015). In this study, we generated harmonized and consistent reference data files for GRCh37 and GRCh38, which are deposited in a Zenodo repository.

MAGMA requires a gene annotation file specifying gene identifiers, chromosomal start and end coordinates, and strand information. Here, we use the same annotation source as employed for the LMDB variant-to-gene database. MAGMA subsequently performs an analogous internal positional mapping of variants to genes, assigning each variant to a gene when it falls within the gene boundaries or a predefined upstream or downstream window. Window parameters are held constant across the entire pipeline.

4.6. Disease module mining and drug repurposing with Drugst.One

GNExT integrates the Drugst.One interface, a plug-and-play web platform that converts systems biology methods into interactive tools for network analysis, disease mechanism mining, and drug repurposing. The interface consists of three components: a central network visualization window, a right-hand menu for direct network manipulation, and a left-hand panel system for node selection, task configuration and initiation, and result inspection. Analytical tasks operate on user-defined subnetworks, with results presented either directly in the Drugst.One interface or linking to external web pages. In GNExT, users can select a set of seed genes from the MAGMA results and transfer them into the integrated Drugst.One interface within the GNExT platform for the following downstream network-based analyses (for further details, we refer to the original Drugst.One and Drugst.One DREAM publications (Maier et al., 2024; Spindler et al., 2025):

- Network integration: The selected seed proteins are mapped into a user-defined protein-protein interaction network, such as NeDRex, BioGRID, IID, IntAct, STRING, and APID, to investigate their interconnections. NeDRex is used by default, as it integrates all of these underlying databases into a unified interaction resource.
- Enrichment analysis: Gene set and pathway enrichment for any selected set of nodes can be performed using g:Profiler (Raudvere et al., 2019), while functional coherence assessment is available through DIGEST (Adamowicz et al., 2022). Both downstream analyses are accessible from the "Analysis" panel and automatically redirect to the corresponding web interfaces with the results preloaded.
- Disease module mining: The module mining functionality enables the detection of disease-associated protein modules within larger interactomes using dedicated disease module mining algorithms or centrality measures (Maier et al., 2024). This analysis can be employed using the "Module detection" functionality within the "Analysis" panel. By default, a Multi-Steiner Tree approach is executed directly through the "Connect genes" panel.
- Drug prioritization: Seed and connector proteins can subsequently be evaluated as potential drug targets by querying established drug-protein interaction databases and ranking candidate compounds accordingly. This analysis is performed via the "Drug ranking" functionality in the "Analysis" panel, where users may select centrality-based metrics or TrustRank as the prioritization method.

As applied in the olfaction use case, the "Quick drug search" option triggers the sequential execution of the Multi-Steiner tree module detection and harmonic centrality algorithm for drug prioritization (Maier et al., 2024).

4.7. Platform implementation

The preprocessing pipeline is implemented in Nextflow with support for local execution via Conda, containerized environments via Docker or Singularity, and distributed execution using SLURM-based HPC systems. The processed data are made accessible through a Django-based backend integrated with a Vue.js frontend. A Typesense search engine powers the platform's autocomplete and query functionalities, enabling rapid and efficient retrieval of millions of genetic variants in real time. The entire system is containerized with Docker to ensure straightforward deployment, reproducibility, and portability across diverse computational environments.

4.8. Generation of reference genome data

MAGMA requires an external reference panel in PLINK format (.bed, .fam, .bim) (Purcell *et al.*, 2007) to model LD among genetic variants when analyzing GWAS summary statistics. Many studies rely on the 1000 Genomes Project reference data (1000 Genomes Project Consortium *et al.*, 2015), which is distributed in the appropriate format for genome assembly GRCh37 both within the MAGMA package and via FUMA. Since our workflow additionally supports inputs aligned to genome assembly GRCh38 and to ensure consistency across genome builds, we therefore generated all reference datasets uniformly for both GRCh37 and GRCh38.

Due to the immense file sizes of the GRCh38 build, all corresponding VCF files were first cleaned by restricting the "FORMAT" field to only genotypes, and by completely removing all entries in the "INFO" field. Following the FUMA preprocessing strategy, multi-allelic variants were decomposed into bi-allelic records using bcftools. For GRCh37, copy number variants were removed. For both genome builds, all chromosome-specific VCF files were then concatenated with bcftools. The resulting VCF was converted to PLINK binary format using PLINK2, assigning unique variant identifiers based on genomic coordinates (chr:pos:ref:alt) rather than rsIDs to ensure compatibility across different dbSNP (Sherry *et al.*, 2001) releases. Finally, the resulting genotype data were stratified into five super-populations according to the provided panel file. The processed reference population files were deposited on Zenodo.

4.9. Application to GWAS olfaction data

To advance understanding of the genetic basis of human olfactory abilities, Förster *et al.* (2025) performed a fixed-effect GWAS meta-analysis on odor identification using Sniffin' Sticks across four cohorts totaling up to $21,495$ individuals of European ancestry, including overall and sex-stratified models for 13 traits. A global identification score (range 0 to 12) summed correct identifications. The corresponding GWAS meta-analysis results (14 GB) were downloaded, preprocessed with the Nextflow pipeline, and integrated into a GNExT platform (`http://olfaction.gnext.gm.eurac.edu`), comprising 39 traits, $23,592,897$ variants, and $19,414$ genes. Gene definitions were obtained from Ensembl release 114 (GRCh38), and gene-level statistics were computed using the default 10kb up- and downstream window size in conjunction with the newly generated reference genome data aligned to GRCh38.

For computational efficiency within the preprocessing pipeline, phenotypes were processed in batches of five. Ensembl VEP annotation represented the most time-consuming preprocessing step in terms of CPU usage (Supplementary Figure 3). Additional stages, including GWAS harmonization, Manhattan and Q-Q JSON file generation, chromosome-specific BGZ file construction, and MAGMA analyses, each exceeded one hour of cumulative CPU time. Nevertheless, the overall wall-clock runtime of the pipeline was markedly reduced through the systematic use of batching and parallelization, with 69 jobs executed in total. The complete workflow finished in approximately 11 hours, leveraging SLURM-based parallelization and the configuration parameters provided in the corresponding

GitHub repository. Finally, we assessed the peak resident set size (RSS), representing the maximum physical RAM occupied during execution, for the Nextflow preprocessing pipeline for both use cases. Unexpectedly, the mean peak RSS varied substantially across pipeline steps (Supplementary Figure 4). MAGMA input generation showed the highest memory consumption, followed by the harmonization of GWAS files. During MAGMA input generation, all reference variants were loaded into RAM to intersect with the GWAS variants. This memory-intensive design represents a potential target for optimization in future pipeline versions.

5. Data and code availability

Summary statistics from the study by Förster *et al.* (2025) were downloaded from `https://www.health-atlas.de/assays/88` (accession code 8VK7H50F6P-6). Data of the 1,000 Genomes Project for GRCh37 and GRCh38 were downloaded from `http://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/` and `http://ftp.1000genomes.ebi.ac.uk/vol1/ftp/data_collections/1000G_2504_high_coverage/working/20201028_3202_raw_GT_with_annot/`, respectively. The panel file specifying the samples super-populations was downloaded from `https://ftp.1000genomes.ebi.ac.uk/vol1/ftp/release/20130502/integrated_call_samples_v3.20130502.ALL.panel`. Gene annotation (GTF) files specifying gene coordinates were obtained from the corresponding Ensembl releases.

The source code of GNExT's Nextflow-based preprocessing pipeline is available at `https://github.com/DyHealthNet/gnext_nf_pipeline`, the platform source code at `https://github.com/DyHealthNet/gnext_platform`. All reference files, including 1,000 Genomes PLINK files and the gene annotation files, are available on Zenodo (`https://doi.org/10.5281/zenodo.17940903`). The scripts to generate these reference files are available at `https://github.com/DyHealthNet/gnext_reference_data`. The GNExT instance for the olfaction study is available at `http://olfaction.gnext.gm.eurac.edu`. The scripts and configuration file that were used to set up GNExT is available at the respective GitHub repositories.

6. Supplements

Supplementary data is available online.

Competing interests

M.L. consults for mbiomics GmbH. All other authors declare no competing interest.

Author contributions statement

L.A. implemented the preprocessing pipeline and developed the platform together with support from B.R. F.W. generated the reference datasets. L.A, F.W., and D.E. contributed to the olfaction use case, and J.F. assisted in interpreting the olfaction results. C.F., D.B.B., and M.L. obtained funding and supervised the project. All authors provided critical feedback and discussion, assisted in interpreting the results, writing the manuscript, and improving the web service.

Funding

The work was supported by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation – 516188180). D.E. and C.F. were supported by the Autonomous Province of Bolzano-Bozen under the funding scheme "Joint Projects South Tyrol–Germany 2024". J.F. is supported by FRQS (chercheur boursier sénior 352197) and NSERC (RGPIN-2022-04813).

Acknowledgements

We thank Lisa Spindler and Andreas Maier for their continued support in integrating Drugst.One, and Daniele Di Domizio for his technical assistance with the deployment process.

References

1000 Genomes Project Consortium, Auton, A., Brooks, L. D. *et al.* (2015). A global reference for human genetic variation. *Nature*, **526**, 68–74.

Adamowicz, K., Maier, A., Baumbach, J. *et al.* (2022). Online in silico validation of disease and gene sets, clusterings or subnetworks with DIGEST. *Brief. Bioinform.*, **23**, bbac247.

Bakshi, A., Zhu, Z., Vinkhuyzen, A. A. E. *et al.* (2016). Fast set-based association analysis using summary data from GWAS identifies novel gene loci for human complex traits. *Sci. Rep.*, **6**, 32894.

Barbieri, M. A., Sorbara, E. E., Russo, G. *et al.* (2023). Neuropsychiatric adverse drug reactions with tyrosine kinase inhibitors in gastrointestinal stromal tumors: An analysis from the european spontaneous adverse event reporting system. **15**, 1851.

Bellavance, J., Xiao, H., Chang, L. *et al.* (2026). Exploring and visualizing stratified GWAS results with PheWeb2. doi: 1–3.

Boughton, A. P., Welch, R. P., Flickinger, M. *et al.* (2021). LocusZoom.js: interactive and embeddable visualization of genetic association study results. *Bioinformatics*, **37**, 3017–3018.

Bycroft, C., Freeman, C., Petkova, D. *et al.* (2018). The UK biobank resource with deep phenotyping and genomic data. *Nature*, **562**, 203–209.

Cao, C., Tian, M., Li, Z. *et al.* (2025). GWAShug: a comprehensive platform for decoding the shared genetic basis between complex traits based on summary statistics. *Nucleic Acids Res.*, **53**, D1006–D1015.

Chen, S., Francioli, L. C., Goodrich, J. K. *et al.* (2024). A genomic mutational constraint map using variation in 76,156 human genomes. **625**, 92–100.

Costanzo, M. C., Roselli, C., Brandes, M. *et al.* (2023). Cardiovascular disease knowledge portal: A community resource for cardiovascular disease research. **16**, e004181.

Cuellar-Partida, G., Lundberg, M., Kho, P. F. *et al.* (2019). Complex-traits genetics virtual lab: A community-driven web platform for post-GWAS analyses. *bioRxiv*, doi: 10.1101/518027.

de Leeuw, C. A., Mooij, J. M., Heskes, T. *et al.* (2015). MAGMA: generalized gene-set analysis of GWAS data. *PLoS Comput. Biol.*, **11**, e1004219.

Debbaneh, P., McKinnon, L., Haidari, M. *et al.* (2023). Drug-induced olfactory and gustatory dysfunction: Analysis of FDA adverse events reporting system. **50**, 558–564.

Di Tommaso, P., Chatzou, M., Floden, E. W. *et al.* (2017). Nextflow enables reproducible computational workflows. *Nat. Biotechnol.*, **35**, 316–319.

Elsworth, B., Lyon, M., Alexander, T. *et al.* (2020). The MRC IEU OpenGWAS data infrastructure. *bioRxiv*, doi: 10.1101/2020.08.10.244293.

Ewels, P. A., Peltzer, A., Fillinger, S. *et al.* (2020). The nf-core framework for community-curated bioinformatics pipelines. *Nat. Biotechnol.*, **38**, 276–278.

Fu, Z.-H., Zhao, C., Wang, Y. *et al.* (2025). Pharmacovigilance imbalance analysis of VEGFR-TKI-related taste and smell disorders. **15**, 3118.

Förster, F., Emmert, D., Horn, K. *et al.* (2025). Genome-wide association meta-analysis of human olfactory identification discovers sex-specific and sex-differential genetic variants. *Nat. Commun.*, **16**, 5434.

Gagliano Taliun, S. A., VandeHaar, P., Boughton, A. P. *et al.* (2020). Exploring and visualizing large-scale genetic associations by using PheWeb. *Nat. Genet.*, **52**, 550–552.

Galindez, G., Sadegh, S., Baumbach, J. *et al.* (2023). Network-based approaches for modeling disease regulation and progression. *Comput. Struct. Biotechnol. J.*, **21**, 780–795.

Gibson, G. (2010). Hints of hidden heritability in GWAS. *Nat. Genet.*, **42**, 558–560.

Groenewoud, D., Shye, A. and Elkon, R. (2022). Incorporating regulatory interactions into gene-set analyses for GWAS data: A controlled analysis with the MAGMA tool. **18**, e1009908.

Haag, F., Hoffmann, S. and Krautwurst, D. (2021). Key food furanones furaneol and sotolone specifically activate distinct odorant receptors. **69**, 10999–11005.

Hopkins, A. L. (2008). Network pharmacology: the next paradigm in drug discovery. *Nat. Chem. Biol.*, **4**, 682–690.

Kontou, P. I. and Bagos, P. G. (2024). The goldmine of GWAS summary statistics: a systematic review of methods and tools. *BioData Min.*, **17**, 31.

Krefl, D., Brandulas Cammarata, A. and Bergmann, S. (2023). PascalX: a python library for GWAS gene and pathway enrichment tests. *Bioinformatics*, **39**, btad296.

Krishnamurthy, N., Grimshaw, A. A., Axson, S. A. *et al.* (2022). Drug repurposing: a systematic review on root causes, barriers and facilitators. *BMC Health Serv. Res.*, **22**, 970.

Kurki, M. I., Karjalainen, J., Palta, P. *et al.* (2023). FinnGen provides genetic insights from a well-phenotyped isolated population. *Nature*, **613**(7944), 508–518.

Linder, J. E., Bastarache, L., Hughey, J. J. *et al.* (2021). The role of electronic health records in advancing genomic medicine. *Annu. Rev. Genomics Hum. Genet.*, **22**, 219–238.

Liu, J. Z., McRae, A. F., Nyholt, D. R. *et al.* (2010). A versatile gene-based test for genome-wide association studies. *Am. J. Hum. Genet.*, **87**, 139–145.

Liu, X., Tian, D., Li, C. *et al.* (2023). GWAS atlas: an updated knowledgebase integrating more curated associations in plants and animals. *Nucleic Acids Res.*, **51**, D969–D976.

Machiela, M. J., Huang, W.-Y., Wong, W. *et al.* (2023). GWAS explorer: an open-source tool to explore, visualize, and access GWAS summary statistics in the PLCO atlas. *Sci. Data*, **10**, 25.

Maier, A., Hartung, M., Abovsky, M. *et al.* (2024). Drugst.one - a plug-and-play solution for online systems medicine and network-based drug repurposing. *Nucleic Acids Res.*, **52**, W481–W488.

McLaren, W., Gil, L., Hunt, S. E. *et al.* (2016). The Ensembl Variant Effect Predictor. *Genome Biol.*, **17**, 122.

Menche, J., Sharma, A., Kitsak, M. *et al.* (2015). Disease networks. uncovering disease-disease relationships through the incomplete interactome. *Science*, **347**, 1257601.

Nagai, A., Hirata, M., Kamatani, Y. *et al.* (2017). Overview of the BioBank japan project: Study design and profile. *J. Epidemiol.*, **27**, S2–S8.

Neale, B. M. and Sham, P. C. (2004). The future of association studies: gene-based analysis and replication. *Am. J. Hum. Genet.*, **75**, 353–362.

Nogales, C., Mamdouh, Z. M., List, M. *et al.* (2022). Network pharmacology: curing causal mechanisms instead of treating symptoms. **43**, 136–150.

Ou, S.-H., Kilvert, H., Candlish, J. *et al.* (2024). Systematic review and network meta-analysis of lorlatinib with comparison to other anaplastic lymphoma kinase (ALK) tyrosine kinase inhibitors (TKIs) as first-line treatment for ALK-positive advanced non-smallcell lung cancer (NSCLC). **197**, 107968.

Park, J.-H., Gail, M. H., Weinberg, C. R. *et al.* (2011). Distribution of allele frequencies and effect sizes and their interrelationships for common genetic susceptibility variants. *Proc. Natl. Acad. Sci. U. S. A.*, **108**, 18026–18031.

Purcell, S., Neale, B., Todd-Brown, K. *et al.* (2007). PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.*, **81**, 559–575.

Pushpakom, S., Iorio, F., Eyers, P. A. *et al.* (2019). Drug repurposing: progress, challenges and recommendations. *Nat. Rev. Drug Discov.*, **18**(1), 41–58.

Raudvere, U., Kolberg, L., Kuzmin, I. *et al.* (2019). g:profiler: a web server for functional enrichment analysis and conversions of gene lists (2019 update). *Nucleic Acids Res.*, **47**, W191–W198.

Sadegh, S., Skelton, J., Anastasi, E. *et al.* (2021). Network medicine for disease module identification and drug repurposing with the NeDRex platform. *Nat. Commun.*, **12**, 6848.

Sherry, S. T., Ward, M. H., Kholodov, M. *et al.* (2001). dbSNP: the NCBI database of genetic variation. *Nucleic Acids Res.*, **29**, 308–311.

Sollis, E., Mosaku, A., Abid, A. *et al.* (2023). The NHGRI-EBI GWAS catalog: knowledgebase and deposition resource. *Nucleic Acids Res.*, **51**, D977–D985.

Spindler, L. M., Kersting, J., Manz, Q. *et al.* (2025). Drugst.one DREAM - drug repurposing through expert annotation and modification.

Stringer, S., Wray, N. R., Kahn, R. S. *et al.* (2011). Underestimated effect sizes in GWAS: fundamental limitations of single SNP analysis for dichotomous phenotypes. *PLoS One*, **6**, e27964.

Su, J., Lu, J., Zhang, J. *et al.* (2023). A meta-analysis of the efficacy and toxicity of tyrosine kinase inhibitors in treating patients with different types of thyroid cancer: how to choose drugs appropriately? **35**, 132–144.

Tong, M. T., Kim, T.-Y. P. and Cleland, T. A. (2018). Kinase activity in the olfactory bulb is required for odor memory consolidation. **25**, 198–205.

Uffelmann, E., Huang, Q. Q., Munung, N. S. *et al.* (2021). Genome-wide association studies. *Nat. Rev. Methods Primers*, **1**, 59.

van Elst, J. M., IJzerman, N. S., Mathijssen, R. H. J. *et al.* (2022). Taste, smell and mouthfeel disturbances in patients with gastrointestinal stromal tumors treated with tyrosine-kinase inhibitors. **30**, 2307–2315.

Visscher, P. M., Wray, N. R., Zhang, Q. *et al.* (2017). 10 years of GWAS discovery: Biology, function, and translation. *Am. J. Hum. Genet.*, **101**, 5–22.

Wang, L., Jia, P., Wolfinger, R. D. *et al.* (2011). Gene set analysis of genome-wide association studies: methodological issues and perspectives. *Genomics*, **98**, 1–8.

Wang, T., Yan, Z., Zhang, Y. *et al.* (2024). postGWAS: A web server for deciphering the causality post the genome-wide association studies. *Comput. Biol. Med.*, **171**, 108108.

Watanabe, K., Taskesen, E., van Bochoven, A. *et al.* (2017). Functional mapping and annotation of genetic associations with FUMA. *Nat. Commun.*, **8**, 1826.

Yu, Y., Huang, K., Lin, Y. *et al.* (2023). Tyrosine kinase inhibitors in HER2-positive breast cancer brain metastases: A systematic review and meta-analysis. **12**, 15090–15100.

Zawistowski, M., Fritsche, L. G., Pandit, A. *et al.* (2023). The Michigan Genomics Initiative: A biobank linking genotypes and electronic clinical records in Michigan Medicine patients. *Cell Genom.*, **3**, 100257.