Context-dependent ATP7 Interactions with Parkinson's Disease-associated Genes Modulate

Copper Homeostasis Phenotypes

Brooke M. Allen[1*], Nadia Gonzalez[2-3*], Erica Werner[4], Victor Faundez[4], and Alysia D. Vrailas-

Mortimer[1-3, #]

1. Illinois State University, School of Biological Sciences, Normal, IL, USA

2. Oregon State University, Department of Biochemistry and Biophysics, Corvallis, OR, USA

3. Oregon State University, Linus Pauling Institute, Corvallis, OR, USA

4. Emory University, Department of Cell Biology, Atlanta, GA, USA

*Denotes equal contribution

Running Title: LRRK2 and Copper Homeostasis

Key words/phrases: LRRK2, ATP7, Copper Homeostasis, Parkinson's disease

Corresponding author:

Alysia D. Vrailas-Mortimer

Oregon State University

Department of Biochemistry and Biophysics

Corvallis, OR 97331

Phone: 541-737-9551

e-mail: Alysia.Mortimer@oregonstate.edu

ORCID: 0000-0001-5927-096X

Brooke Allen ORCID: 0009-0002-6028-3340

Nadia Gonzalez ORCID: 0009-0001-3889-6573

Erica Werner ORCID: 0000-0002-8183-16001

Victor Faundez ORCID: 0000-0002-2114-5271

Summary Statement

We performed a genetic interaction screen to explore the relationship between copper

homeostasis and Parkinson's disease and other neurodegeneration associated genes.

Abstract

Copper is an essential micronutrient required by enzymes that catalyze oxygen-dependent

reactions, but toxic in excess. Mutations in the ATP7A and ATP7B copper transporters cause

neuropsychiatric symptoms and neurodegeneration by mechanisms that remain to be elucidated.

We previously reported that the ATP7A biochemical interactome is enriched in Parkinson's

disease (PD) and neurodegeneration associated genes, yet the functional outcomes of these

interactions are unknown. Using *Drosophila,* we tested genetic interactions between ATP7

mutants that alter copper levels and a subset of these PD and neurodegeneration causative genes

and found sex differences with some candidate genes enhancing ATP7 deleterious phenotypes in

both sexes, while others were sex specific. Most notably, we found that Lrrk2 (Lrrk), the most

commonly mutated gene in familial forms of PD, protects against ATP7 dysfunction in

epidermal epithelial cells with a stronger effect in males than females. However, in dopaminergic

neurons Lrrk plays a role in intracellular copper induced toxicity in females but not males,

supporting context dependent interactions between ATP7A and PD-associated genes to protect

against disruptions in copper homeostasis.

Introduction

Copper is an essential micronutrient that acts as a structural component and catalytic cofactor for

key cellular processes, such as antioxidant defense, mitochondrial respiration, energy and iron

metabolism, and neurotransmitter production[1-3]. Disruptions in copper homeostasis can cause

oxidative damage and impaired function of critical cuproenzymes, resulting in cellular

dysfunction and cell death[4-6]. Therefore, copper homeostasis is tightly regulated within the cell

by the copper transporters ATP7A and B which pump cytoplasmic copper into the lumen of the

Golgi apparatus for metalation of secreted cuproenzymes. Excess levels of cytoplasmic copper,

cause ATP7A/B translocation to the plasma membrane, pumping excess copper out of the cell[7-

[10]. Thus, deficiencies in ATP7B leads to copper accumulation at the cellular level and at the

organism level, resulting in the neurodegenerative disorder Wilson's disease[5,6]. Whereas

ATP7A mutations can cause Menkes disease, a multi-system disorder that includes widespread

neurodegeneration caused by decreased copper in the brain due to defective dietary copper

absorption[10,11]. Similarly, in the fruit fly, *Drosophila melanogaster*, genetic manipulation of the

sole homologue *ATP7* causes a copper depletion phenotype affecting neuronal function[12] and

structure[13] and induces early mortality[14]. We have found that ATP7A interacts with proteins

associated with neurodegeneration and neurodevelopmental disorders[13], and ATP7A mutant

fibroblasts have altered expression of proteins associated with Parkinson's disease (PD) or

neurodegeneration, including elevated expression of the early onset neurodegenerative disease

protein, UCHL1[14]. Furthermore, we confirmed this link between ATP7 and UCHL1 through

genetic interactions in *Drosophila*[14].

PD is the second most common neurodegenerative disease and is characterized by loss of

dopaminergic neurons in the substantia nigra, impaired mitochondria, and increased oxidative

stress[15,16]. Familial forms of PD account for only 5-15% of cases, suggesting a strong

environmental component for this disease[15–17]. However, low brain copper levels can lead to

dysfunction and neurodegeneration as well, and copper levels are reduced in mice treated with

MPTP, a potent inducer of PD-like phenotypes[18,19]. Additionally, key mitochondrial proteins

such as Cytochrome c oxidase and superoxide dismutase (SOD1) are cuproenzymes, and ATP7A

mutant fibroblasts have impaired mitochondria[14,20]. Furthermore, both elevated and reduced

levels of copper can induce oxidative stress through different mechanisms[21,22], suggesting that

both increased and decreased levels of intracellular copper may contribute to PD. This is

supported by findings that administering $CuSO_4$ or Cu(II)(atsm) improve PD-like phenotypes in

mouse models[23–25], while other studies found improvement with treatment with a copper

chelator[26,27]. Furthermore, copper interacts with proteins mutated in familial forms of PD. DJ-1

can act as a copper chaperone to protect against metal toxicity[28,29], while copper binding to

alpha-synuclein promotes its aggregation[30,31]. In addition, loss of ATP7A exacerbated motor

phenotypes in a mouse alpha-synuclein (*SCNA*) model[32], and in *C. elegans*, PARKIN and

LRRK2 PD-like phenotypes were enhanced by environmental exposure to copper[33]. These data

suggest a strong connection between copper homeostasis and PD, however the mechanism

remains poorly understood.

To further explore this link between copper homeostasis and PD, we tested the functional

consequences of PD- or neurodegeneration associated genes that we identified as ATP7A/B

interactors in our previous proteomics screens[13,14] genetically interact with ATP7 using the

Drosophila model system, which we have shown recapitulates many of the phenotypes of

ATP7A/B human cell culture models[13,14,20,34]. We used two paradigms previously validated that

allow us to differentiate roles of copper homeostasis in non-neuronal cells (the epidermal

epithelium) and neuronal cells (dopaminergic neurons) in response to increased cellular copper

by ATP7 knockdown (RNAi) or copper depletion by overexpression of ATP7 in a cell

autonomous manner[13,20,34–37]. We find that inhibition of specific candidate genes enhances ATP7

knockdown phenotypes in epidermal epithelial cells in a sex dependent manner with some genes

showing interactions with both sexes while others only interacting in a sex-specific manner. We

focused on Lrrk, the fly homologue of Lrrk2 which is the most commonly mutated gene in

familial forms of PD and found that it genetically interacts with ATP7 in both males and females

in the epidermal epithelium but only in female dopaminergic neurons. These data suggest that

disruptions in copper homeostasis may broadly affect genes involved in PD/neurodegeneration

and that these interactions may contribute to disease pathology.

Results

Coessentiality network analysis of ATP7A interactome

We previously found that loss of ATP7A, a model of increased cellular copper in human cells

results in elevated levels of the neurodegeneration associated gene UCHL1 and that UCHL1

inhibition rescues ATP7 phenotypes in human cell culture and flies[38], suggesting that other

neurodegeneration genes in the ATP7A interactome could modulate copper-dependent

phenotypes as well. We first tested potential genetic interactions between ATP7A and the

interactome PD/neurodegeneration genes by performing a coessentiality network analysis in

silico (Figure S1). We assessed genome-scale fitness correlations between ATP7A with

ARPC1A, ARPC1B, GIGYF2, HIP1, LRKK2, VPS13C and VPS35 mining the Dependency

Map (DepMap) datasets[39,40]. Using the FIREWORKS tool (Fitness Interaction Rank-

Extrapolated netWORKs)[41], we identified that correlated and anticorrelated genes reveal

pathways and regulatory mechanisms connecting genes of interest with ATP7A positively (red

edges) and negatively (blue edges) interacting with Parkinson's and neurodegeneration genes.

These in silico findings support the model that ATP7A genetic interactions of similar complexity

may also occur in mammals as we find in *Drosophila*.

ATP7 Genetically Interacts with Parkinson's Disease Genes

We next tested if these predicted interactions from our coessentiality network analysis are

biological relevant using our fly ATP7 model (Table 1). In addition, we also tested two other

interactome candidates SCYL1 (yata in flies), a neurodegeneration associated protein which had

low expression levels, and the PD gene Synj1, which is a component of the endocytic pathway

that was identified in the interactome (Table 1)[13,14]. Knockdown of ATP7 in the fly dorsal

epidermal epithelium using pnr-GAL4 increases intracellular copper resulting in a decrease in

pigmentation in a central stipe along the back of the fly in both females and males due to

increased levels of intracellular copper (Figures 1B and 2B) as we and others have previously

observed. In addition, these flies also display a widening of the gap between the midline bristles

of the thorax (Figures 1B and 2B) as compared to out-crossed control pnr-GAL4 flies (Figures

1A and 2A). We next tested each of the ATP7A interactome genes and find that knockdown of

Arpc1 results in disorganized and missing midline bristles in females and males (Figures S2B

and S3B). Inhibition of either Gyf or yata results in missing scutellar bristles and some loss of

thoracic bristles (Figures S2C, J-L) in females with more mild phenotypes in males (Figures

S3C, J-L). Both Synj and Vps35 knockdown also leads to mild loss of scutellar bristles in

females and males (Figures S2E, I and S3E, I). We also observe sex differences with inhibition

of Hip1 and Vps13 which have no phenotype in females (Figures S2D, G-H) and mild scutellar

bristle loss in males (Figures S3D, G-H).

We next tested for genetic interactions between ATP7 down-regulation and inhibition of

these interactome genes. We find that in females, inhibition of either Arpc1, Hip1, or Vps13

enhances the ATP7 RNAi phenotype, resulting in a caved in thorax (Figure 1C, E, H, and Table

S1). In addition, the ATP7 RNAi phenotype is also enhanced in females with inhibition of yata

(line 1) with mild inhibition leading to increased bristle loss (Figure 1K and Table S1) and strong

inhibition (line 2) leading to lethality. We also observe an enhancement in disorganization of the

midline thoracic bristles when Vps35 was inhibited (Figure 1J and Table S1). However, not all

of the interactome genes interacted with ATP7 in females with neither inhibition of Gyf nor Synj

affecting the ATP7 down-regulation phenotype (Figure 1D, F-G and Table S1).

As ATP7 is on the X chromosome, males may be more sensitive to genetic perturbations

than females, and we find that similar to females Arpc1, Hip1, and Vps13 inhibition enhances

the ATP7 thorax caving in phenotype (Figure 2C, E, H and Table S1), though the effect of Arpc1

is not as strong as in females (Figure 2C vs 1C and Table S1). In addition, we find that mild

inhibition of yata (line 1) leads to a more severe enhancement of the ATP7 down-regulation

phenotype in males as compared to females (Figures 2K vs 1K and Table S1). Though Vps35

inhibition enhanced the ATP7 knockdown phenotype in females, we find no interaction in males

(Figure 2J and Table S1). In addition, unlike in females, inhibition of Synj (line 1) also leads to

enhancement of the phenotype (Figure 2F and Table S1), whereas similar to females Gyf

inhibition does not interact with ATP7 down-regulation in males (Figure 2D and Table S1).

These data suggest that normal expression of PD/neurodegeneration genes play a role in

protecting cells from increased intracellular copper and that males are more sensitive to these

perturbations.

Inhibition of Lrrk enhances ATP7 down-regulation epidermal epithelium phenotypes

Lrrk2 is the most commonly mutated gene in familial forms of PD[42] and regulates vesicular

trafficking, autophagy, and lysosomal function within cells[17,43–45]. Therefore, we next tested if

the Lrrk2 Drosophila orthologue, Lrrk, also plays a role in copper homeostasis using three

different RNAi lines (Table S2). We find that in both females and males, knockdown of Lrrk

alone does not affect the epidermal epithelium (Figures 3A-D, 4A-D, and Table S1). However,

when combined with down-regulation of ATP7, inhibition of Lrrk robustly enhanced the ATP7

RNAi thorax phenotype, yet to a different extent across lines (Figures 3F-H, 4F-H, and Table

S1). Knockdown of Lrrk results in a more severe phenotype than in males than in females. For

line 1, some males present with necrosis of the scutellum (Figure 4F') and line 2 displayed a

more pronounced caving of the thorax (Figure 4G). In addition, we find that line 3 presented

with mild phenotypes in females (Figure 3H and Table S1), whereas males had a more

pronounced phenotype with increased thoracic caving (Figure 4H and Table S1) though these

phenotypes were milder than line 2 (Figure 4G and Table S1). These data suggest that Lrrk acts

to protect epidermal epithelial cells from high levels of intracellular copper and is a more potent

interactor in males.

Lrrk and ATP7 down-regulation do not interact to regulate viability

As inhibiting Lrrk leads to a worsening of the ATP7 down-regulation thorax phenotypes, we

tested if this alters organismal viability. Though down-regulation of ATP7 does not affect

viability in either females or males (Figure S4A, D and Table S3), Lrrk inhibition results in

reduced viability (Figure S4B-C, E-F and Table S3). Furthermore, inhibiting Lrrk in the ATP7

down-regulation background reduces viability as compared to ATP7 knockdown alone, though

this is not significantly different from Lrrk inhibition alone (Figure S4B-C, E-F and Table S3).

These data show that the epithelial damage induced by disruption of the Lrrk and ATP7

interaction spares animal viability.

Inhibition of Lrrk rescues ATP7 down-regulation dopaminergic neuronal phenotypes in a sex

dependent manner

We have previously found that altering copper homeostasis in dopaminergic neurons, which are

lost in PD, decreases survival to a copper challenge[13,14]. Therefore, we tested whether Lrrk

affects these outcomes. We find that in both females and males, inhibition of either ATP7 or Lrrk

alone or in combination does not affect survival on glucose only food (Table S4). Knockdown of

ATP7 decreases survival in females when exposed to copper, whereas knockdown of Lrrk did

not affect survival as compared to the GAL4 control line (Figure 5A-C, Table S5). However,

inhibiting Lrrk in the ATP7 knockdown background, rescued survival (Figure 5A-C, Table S5).

In contrast, males died faster when exposed to copper and had decreased survival when either

ATP7 or Lrrk is inhibited (Figure 5D-F, Table S5). Knocking down Lrrk in the ATP7 down-

regulation background did not significantly alter survival as compared to Lrrk or ATP7 down-

regulation (Figure 5D-F, Table S5). These data show that Lrrk contributes to the toxicity caused

by increased intracellular copper in females but not males.

Inhibition of Lrrk enhances ATP7 over-expression epidermal epithelium phenotypes

In humans, both increased intracellular copper and decreased intracellular copper lead to

neurological disorders, therefore, we tested if inhibition of Lrrk also interacts with decreased

copper levels modelled by over-expression of ATP7. As we and others have previously reported,

over-expression of ATP7 in the epidermal epithelium results in decreased pigmentation and

bristle loss in both males and females (Figures 3I and 4I and[9,20,34–38]). When we combine

inhibition of Lrrk with over-expression of ATP7, we find increased caving in of the thorax

(Figures 3J-L, 4J-L, and Table S1) and in some instances a darkening of the thorax (Figure 3L

and Figure 4K') that is indicative of underlying internal necrosis[46,47]. Therefore, inhibition of

Lrrk further enhances intracellular copper depletion thorax phenotypes in both females and

males. Taken together, our data suggest that Lrrk normally acts to protect against both

accumulation (Figures 3E-H, 4E-H, and Table S1) and depletion of intracellular copper (Figures

3I-L, 4I-L, and Table S1).

Inhibition of Lrrk reduces viability of ATP7 over-expression

We next tested if Lrrk interacts with copper depletion for organismal viability and find that

similar to ATP7 down-regulation, over-expression of ATP7 does not affect viability (Figure 6

and Table S6). However, inhibition of Lrrk in the ATP7 over-expression background reduces

viability in females and males (Figure 6B-C, E-F and Table S6) but only leads to a more severe

reduction as compared to Lrrk inhibition alone in females (Figure 6A-C and Table S6). These

data indicate that in females, Lrrk acts to protect epidermal epithelial cells from copper depletion

but not accumulation toxicity during development to promote survival to adulthood.

Inhibition of Lrrk in dopaminergic neurons reduces survival of ATP7 over-expression in

response to environmental copper exposure

As copper depletion in the brain leads to neurological disorders, we next tested if Lrrk interacts

with ATP7 over-expression in dopaminergic neurons. In females, over-expression of ATP7 leads

to decreased survival, which is not affected by inhibition of Lrrk (Figure 7A-C, Table S7).

However, in males ATP7 over-expression had no effect on survival in response to copper

exposure though inhibition of Lrrk resulted in decreased survival (Figure 7D-F, Table S7). In

addition, Lrrk inhibition in the ATP7 over-expression background also led to decreased survival

to levels similar to Lrrk inhibition alone (Figure 7D-F, Table S7), suggesting that inhibition of

Lrrk does not sensitize the cell to copper depletion. Therefore, Lrrk does not interact with ATP7

over-expression in dopaminergic neurons in response to environmental copper exposure.

Discussion

Copper homeostasis and PD

Copper dysregulation and PD share many key features including increased oxidative stress,

protein aggregation, and mitochondrial damage[21,22]. In addition, PD has been associated with

disruptions in copper homeostasis and copper-associated toxicity[21] with PD associated genes

intersecting with signaling pathways that regulate metal homeostasis and counteract oxidative

stress[44]. We have previously performed proteomic analyses to identify interactors of the copper

transporter ATP7A and identified a number of genes associated with PD and/or

neurodegeneration[13,14]. We further confirmed a genetic interaction between ATP7A and one of

these candidate genes, UCHL1, which is elevated in ATP7A mutant fibroblasts and inhibition of

UCHL1 rescued ATP7A phenotypes[14] and may be a potential target for the development of new

therapeutics. Here, we have further explored how wide-spread this relationship is by testing for

genetic interactions between ATP7 and other candidate genes from the ATP7A interactome[13,14].

We find that a subset (7 out of the 8 genes tested) genetically interact with ATP7 and

play a role in protecting epidermal epithelial cells from increased intracellular copper. Five of

these genes (Lrrk, Hip1, Synj, Vps13, and Vps35) are associated with PD, while Arpc1 (ARPC1)

interacts with known PD genes[48–51]. Two common themes emerge about the functions of those

genes that interacted with ATP7: 1) a role in mitochondrial function/health (Vps13, Vps35, and

Arpc1)[44,49,50,52–56] and 2) a role in endocytosis and Golgi trafficking (Vps13, Vps35, Arpc1, yata

and Hip1)[4,54,57–60]. In mammalian cell culture, VPS13C plays a role in ER-late

endosomes/lysosome communication[57] and localizes to the outer mitochondrial membrane[53]

where it may also mediate communication between the mitochondria and the ER/endosomal

system. Vps13 (VPS13C in mammals) also leads to mitochondrial dysfunction and damage and

plays a role in parkin/PINK1 mediated mitophagy[53]. Similarly, a mouse model of VPS35

recapitulated many PD phenotypes including mitochondrial dysfunction and damage[54] and

impaired mitophagy in SH-SY5Y cells[55]. Interestingly both VPS13C and VPS35 play a role in

iron homeostasis[61,62], along with our findings, these data suggest that these genes may play a role

in responding to metal dyshomeostasis. Arpc1 regulates actin polymerization and plays a role in

a variety of cellular processes including the endosomal trafficking[59]. Upon mitochondrial

damage, actin filaments are assembled by Arp2/3 around the damaged mitochondria and

inhibition of Arp2/3 impacts mitochondrial fusion[63] and parkin mediated mitophagy[56]. In

addition Arp2/3 is required for the function of the retromer complex, which includes VPS35[51],

suggesting that the nexus between mitochondrial and endosomal-lysosomal processes may be a

key target for the copper induced toxicity and PD.

Though HIP1 and yata have no known mitochondrial functions, they also play a role in

endocytosis and endosomal trafficking. HIP1 regulates clathrin-mediated endocytosis[60] and

trafficking of oxidative stress responsive cell receptors[64], and Hip1 loss of function has been

implicated in Huntington's disease, a neurodegenerative disorder in which oxidative stress and

mitochondrial dysfunction are prominent pathogenic features[65], suggesting that HIP1-dependent

trafficking may have downstream effects on mitochondrial homeostasis. yata (SCYL1 in

mammals) is associated with several neurodegenerative diseases and is an accessory factor for

the components of coatomer I (COPI) complex to regulate trafficking between the Golgi and

ER[58]. Inhibition of yata in both mice and flies leads to neurodegeneration[66,67] and interacts with

other proteins associated with neurodegenerative diseases such as APPL (Amyloid Precursor

Protein-like) in flies[66] and TDP-43 in mice[68]. We have previously found that ATP7 interacts with

the conserved oligomeric Golgi (COG) complex, a Golgi apparatus vesicular tether[20]. This

connection between intracellular trafficking and copper homeostasis suggests that when these

trafficking genes are disrupted it could exacerbate copper toxicity by impairing copper

sequestration, exocytosis, or recycling or metal binding proteins.

Sex differences in response to copper dysregulation

PD and other neurological disorders can present with skewed sex ratios. For example, men are

1.5 times more likely to develop PD than women[69,70]. In addition, as ATP7A is a sex-linked

gene, Menkes disease primarily affects males[71]. Similarly, we find sex-dependent interactions

among these genes with Synj interacting with ATP7 in males but not females. We also observed

differences in severity of phenotypes across sexes as Arpc1 has milder phenotypes in males than

females while yata and Lrrk have more severe phenotypes in males than in females. These data

suggest that males and females may have different protective mechanisms against copper
dysregulation that intersect with some PD-associated genes but not others.
312
313 *Lrrk and ATP7 interactions are context dependent*
314 Lrrk is a kinase that regulates diverse intracellular processes, such as: mitophagy, innate
315 immunity, vesicular transport, and autophagy[17,44,45], pathways that are frequently disrupted
316 during metal-induced stress, including copper toxicity[20,72–76].
317
We find that the interaction between Lrrk and ATP7 are dependent on the context. For example,
we find that Lrrk protects against both increased and depleted levels of intracellular copper in
epidermal epithelial cells to maintain thoracic structure in both sexes. However, in response to
increased copper, males are more severely affected when Lrrk is inhibited. Furthermore, only in
females and only during intracellular copper depletion in the epidermal epithelium does Lrrk act
to promote organismal viability. These interactions are also cell type specific as we find that
inhibition of Lrrk in females, but not males, improved survival in response to increased levels of
intracellular copper (ATP7 down-regulation with copper exposure) but had no effect on survival
in response to depletion of intracellular copper (ATP7 over-expression with copper exposure).
These data suggest that Lrrk contributes to intracellular copper toxicity, and since it plays a role
in stress response pathways[17], Lrrk2 may help buffer cells against copper-induced toxicity.
Additionally, as the pathogenic G2019S mutation in Lrrk is a gain of function mutation[77,78],
disruptions in copper homeostasis could lead to an earlier age of onset or more severe
symptoms/faster disease progression.
332
Through this study, we have confirmed that the ATP7A interactome identified from
human cell culture and patient samples[13,38], represent biologically significant interactions that
contribute to how cells respond to disruptions in cellular homeostasis. In addition, we also find
that there is an intimate and common connection between copper homeostasis and genetic forms
of PD, which may also provide new insights into how environmental exposure to copper and
other heavy metals contribute to sporadic forms of PD.
339
340 **Materials and Methods**

Coessentiality Analysis

Coessentiality Analysis was performed with the FIREWORKS engine[41] using ATP7A with ARPC1A, ARPC1B, GIGYF2, HIP1, LRKK2, VPS13C and VPS35 genes as entries in the Pan-Cancer dataset as Context. The engine was run with the top 5 primary nodes and the top five secondary nodes positively and negatively correlated.

Drosophila Strains and Husbandry

The following strains were obtained from the Bloomington *Drosophila* Stock Center, Bloomington, IL: w1118 (#5905), Ddc-Gal4 (#7009), pnr-Gal4 (#3039), UAS-Lrrk-RNAi-1 (#32457), UAS-Lrrk-RNAi-2 (#39019), and UAS-Lrrk-RNAi-3 (#35249). The UAS-ATP7-RNAi (#108159) stock was obtained from the Vienna *Drosophila* Resource Center. The UAS-ATP7-wt (overexpression) was a gift from Richard Burke, Monash University, Australia. See Table S2 for a full list of genotypes. All stocks were reared on standard Molasses Food (Genesee Scientific) at 25°C in 12hr:12hr light:dark cycle.

Imaging

ATP7A interactome candidate genes were knocked down in epidermal epithelium cells in either an ATP7 loss of function or overexpression background using the pnr-Gal4 driver. 1-week-old flies were imaged using a LEICA S6D stereomicroscope with a Leica EC4 Digital Camera. Z-stacks were then aligned and max projections generated using Zerene Stacker.

Viability Assay

Three biological replicates of 8-10 virgin females were crossed with 4-5 males for each genotype. The flies were transferred into new vials every three days resulting in three technical replicates for each cross. Once the offspring began eclosing, they were tracked for nine days, accounting for the number of males and females for each expected genotype were recorded. All crosses were reared on standard Molasses Food (Genesee Scientific) at 25°C in 12hr:12hr light:dark cycle.

Copper Toxicity Assay

Copper feedings were performed on 1-week-old virgin females and males reared separately.

ATP7 was either knockdown or overexpressed alone or in combination with Lrrk knockdown in

the dopaminergic neurons using the Ddc-Gal4 driver. Flies were starved for 3 hours and

transferred to empty vials with Whatman glass microfiber filter discs saturated in either 1 mM

$CuSO_4$ + 5% glucose solution or 5% glucose solution. Survival was assayed for every 12-24

hours for five days. Experiments were performed with a minimum of 10 replicates per genotype

per sex with ~10 flies per replicate at 25°C in 12hr:12hr light:dark cycle.

Statistical Analysis

The results of the copper toxicity assay were tested for statistical significance testing survival

and proportion data. Survival data was tested using Kaplan-Meier survival analysis, log-rank

tests, and pairwise comparisons to assess the significance between genotypes and survival time.

Proportion data for copper survival and viability were tested by creating a generalized linear

model and an analysis of deviance table was used to fit variance using chi-squared tests for

significance, per-time contrasts were conducted with post emmeans testing with tukey

adjustment. All statistical analyses were performed in R.

Acknowledgements

We would also like to acknowledge the Bloomington Drosophila Stock Center (NIH

P40OD018537), and the Vienna Drosophila Resource Center for providing fly stocks used in this

study. Funding for this project included start-up funds from Illinois State University and Oregon

State University, and R15AR070505 to AVM, and 1RF1AG060285 to VF. EW is supported by

NIH grant ES034796 and NG is supported by 1T32GM160704-01. The content is solely the

responsibility of the authors and does not necessarily represent the official views of the National

Institutes of Health. The funders had no role in study design, data collection and analysis,

decision to publish, or preparation of the manuscript.

Author Contributions

BA and NG performed experiments and data analysis. EW, VF, and AVM made intellectual

contributions to experimental design, data analysis and interpretation, and provided funding. All

authors were involved in reading and editing the manuscript.

Conflict of Interest Statement

The authors have no conflicts of interest.

Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon

reasonable request.

References

1. Henriksen, C., and Arnesen, E.K. (2023). Copper – a scoping review for Nordic Nutrition
    Recommendations 2023. Food Nutr. Res. *67*, 10.29219/fnr.v67.10322.
    https://doi.org/10.29219/fnr.v67.10322.

2. An, Y., Li, S., Huang, X., Chen, X., Shan, H., and Zhang, M. (2022). The Role of Copper
    Homeostasis in Brain Disease. Int. J. Mol. Sci. *23*, 13850.
    https://doi.org/10.3390/ijms232213850.

3. EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA) (2015). Scientific Opinion
    on Dietary Reference Values for copper. EFSA J. *13*, 4253.
    https://doi.org/10.2903/j.efsa.2015.4253.

4. Lutsenko, S., Bhattacharjee, A., and Hubbard, A.L. (2010). Copper handling machinery of
    the brain. Met. Integr. Biometal Sci. *2*, 596–608. https://doi.org/10.1039/c0mt00006j.

5. Lutsenko, S., Barnes, N.L., Bartee, M.Y., and Dmitriev, O.Y. (2007). Function and
    Regulation of Human Copper-Transporting ATPases. Physiol. Rev. *87*, 1011–1046.
    https://doi.org/10.1152/physrev.00004.2006.

6. Chen, L., Min, J., and Wang, F. (2022). Copper homeostasis and cuproptosis in health and
    disease. Signal Transduct. Target. Ther. *7*, 378. https://doi.org/10.1038/s41392-022-01229-y.

7. Lane, A.R., Roberts, B.R., Fahrni, C.J., and Faundez, V. (2025). A primer on copper biology
    in the brain. Neurobiol. Dis. *212*, 106974. https://doi.org/10.1016/j.nbd.2025.106974.

8. Zlatic, S., Comstra, H.S., Gokhale, A., Petris, M.J., and Faundez, V. (2015). Molecular Basis
    of Neurodegeneration and Neurodevelopmental Defects in Menkes Disease. Neurobiol. Dis.
    81, 154–161. https://doi.org/10.1016/j.nbd.2014.12.024.

9. Hartwig, C., Zlatic, S.A., Wallin, M., Vrailas-Mortimer, A., Fahrni, C.J., and Faundez, V.
    (2019). Trafficking mechanisms of P-type ATPase copper transporters. Curr. Opin. Cell
    Biol. *59*, 24–33. https://doi.org/10.1016/j.ceb.2019.02.009.

10. Kaler, S.G. (2011). ATP7A-related copper transport diseases—emerging concepts and future
trends. Nat. Rev. Neurol. *7*, 15–29. https://doi.org/10.1038/nrneurol.2010.180.

11. Tümer, Z., and Møller, L.B. (2010). Menkes disease. Eur. J. Hum. Genet. *18*, 511–518.
https://doi.org/10.1038/ejhg.2009.187.

12. Hwang, J.E.C., de Bruyne, M., Warr, C.G., and Burke, R. (2014). Copper overload and
deficiency both adversely affect the central nervous system of Drosophila. Met. Integr.
Biometal Sci. *6*, 2223–2229. https://doi.org/10.1039/c4mt00140k.

13. Comstra, H.S., McArthy, J., Rudin-Rush, S., Hartwig, C., Gokhale, A., Zlatic, S.A.,
Blackburn, J.B., Werner, E., Petris, M., D'Souza, P., et al. (2017). The interactome of the
copper transporter ATP7A belongs to a network of neurodevelopmental and
neurodegeneration factors. eLife *6*, e24722. https://doi.org/10.7554/eLife.24722.

14. Zlatic, S.A., Vrailas-Mortimer, A., Gokhale, A., Carey, L.J., Scott, E., Burch, R., McCall,
M.M., Rudin-Rush, S., Davis, J.B., Hartwig, C., et al. (2018). Rare Disease Mechanisms
Identified by Genealogical Proteomics of Copper Homeostasis Mutant Pedigrees. Cell Syst.
6, 368-380.e6. https://doi.org/10.1016/j.cels.2018.01.008.

15. Tysnes, O.-B., and Storstein, A. (2017). Epidemiology of Parkinson's disease. J. Neural
Transm. *124*, 901–905. https://doi.org/10.1007/s00702-017-1686-y.

16. Pringsheim, T., Jette, N., Frolkis, A., and Steeves, T.D.L. (2014). The prevalence of
Parkinson's disease: A systematic review and meta-analysis. Mov. Disord. *29*, 1583–1590.
https://doi.org/10.1002/mds.25945.

17. Klein, C., and Westenberger, A. (2012). Genetics of Parkinson's Disease. Cold Spring Harb.
Perspect. Med. *2*, a008888. https://doi.org/10.1101/cshperspect.a008888.

18. Alvarez-Vega, R., Rios, C., Rojas, P., and Kravsov, J. (1992). MPTP treatment decreases
striatal copper content in mice. Proc. West. Pharmacol. Soc. *35*, 25–27.

19. Matusch, A., Depboylu, C., Palm, C., Wu, B., Höglinger, G.U., Schäfer, M.K.-H., and
Becker, J.S. (2010). Cerebral bioimaging of Cu, Fe, Zn, and Mn in the MPTP mouse model
of Parkinson's disease using laser ablation inductively coupled plasma mass spectrometry
(LA-ICP-MS). J. Am. Soc. Mass Spectrom. *21*, 161–171.
https://doi.org/10.1016/j.jasms.2009.09.022.

20. Hartwig, C., Méndez, G.M., Bhattacharjee, S., Vrailas-Mortimer, A.D., Zlatic, S.A.,
Freeman, A.A.H., Gokhale, A., Concilli, M., Werner, E., Sapp Savas, C., et al. (2021).
Golgi-Dependent Copper Homeostasis Sustains Synaptic Development and Mitochondrial
Content. J. Neurosci. Off. J. Soc. Neurosci. *41*, 215–233.
https://doi.org/10.1523/JNEUROSCI.1284-20.2020.

21. Bisaglia, M., and Bubacco, L. (2020). Copper Ions and Parkinson's Disease: Why Is
Homeostasis So Relevant? Biomolecules *10*, 195. https://doi.org/10.3390/biom10020195.

22. Davies, K.M., Bohic, S., Carmona, A., Ortega, R., Cottam, V., Hare, D.J., Finberg, J.P.M.,
Reyes, S., Halliday, G.M., Mercer, J.F.B., et al. (2014). Copper pathology in vulnerable
brain regions in Parkinson's disease. Neurobiol. Aging *35*, 858–866.
https://doi.org/10.1016/j.neurobiolaging.2013.09.034.

23. Hung, L.W., Villemagne, V.L., Cheng, L., Sherratt, N.A., Ayton, S., White, A.R., Crouch,
P.J., Lim, S., Leong, S.L., Wilkins, S., et al. (2012). The hypoxia imaging agent CuII(atsm)
is neuroprotective and improves motor and cognitive functions in multiple animal models of
Parkinson's disease. J. Exp. Med. *209*, 837–854. https://doi.org/10.1084/jem.20112285.

24. Cheng, Y., Wang, G., Yang, X., Wang, Y., Li, D., Zhao, Y., and Zhang, F. (2025).
Artesunate alleviates Parkinson's disease by targeting astrocyte MT2A to attenuate
dopamine neuronal cuproptosis. Pharmacol. Res. *219*, 107895.
https://doi.org/10.1016/j.phrs.2025.107895.

25. Alcaraz-Zubeldia, M., Rojas, P., Boll, C., and Ríos, C. (2001). Neuroprotective Effect of
Acute and Chronic Administration of Copper (II) Sulfate against MPP+ Neurotoxicity in
Mice. Neurochem. Res. *26*, 59–64. https://doi.org/10.1023/A:1007680616056.

26. Aguirre, P., García-Beltrán, O., Tapia, V., Muñoz, Y., Cassels, B.K., and Núñez, M.T.
(2017). Neuroprotective Effect of a New 7,8-Dihydroxycoumarin-Based Fe2+/Cu2+
Chelator in Cell and Animal Models of Parkinson's Disease. ACS Chem. Neurosci. *8*, 178–
185. https://doi.org/10.1021/acschemneuro.6b00309.

27. Leuci, R., Brunetti, L., Tufarelli, V., Cerini, M., Paparella, M., Puvača, N., and Piemontese,
L. (2024). Role of copper chelating agents: between old applications and new perspectives in
neuroscience. Neural Regen. Res. *20*, 751–762. https://doi.org/10.4103/NRR.NRR-D-24-
00140.

28. Björkblom, B., Adilbayeva, A., Maple-Grødem, J., Piston, D., Ökvist, M., Xu, X.M., Brede,
C., Larsen, J.P., and Møller, S.G. (2013). Parkinson disease protein DJ-1 binds metals and
protects against metal-induced cytotoxicity. J. Biol. Chem. *288*, 22809–22820.
https://doi.org/10.1074/jbc.M113.482091.

29. Girotto, S., Cendron, L., Bisaglia, M., Tessari, I., Mammi, S., Zanotti, G., and Bubacco, L.
(2014). DJ-1 is a copper chaperone acting on SOD1 activation. J. Biol. Chem. *289*, 10887–
10899. https://doi.org/10.1074/jbc.M113.535112.

30. Paik, S.R., Shin, H.J., and Lee, J.H. (2000). Metal-catalyzed oxidation of alpha-synuclein in
the presence of Copper(II) and hydrogen peroxide. Arch. Biochem. Biophys. *378*, 269–277.
https://doi.org/10.1006/abbi.2000.1822.

31. Walke, G., Kumar, R., and Wittung-Stafshede, P. (2024). Copper ion incorporation in α-
synuclein amyloids. Protein Sci. Publ. Protein Soc. *33*, e4956.
https://doi.org/10.1002/pro.4956.

32. Casey, M., Zou, D., Reijo Pera, R.A., Hensley-McBain, T., and Cabin, D.E. (2025). Alpha-
synuclein null mutation exacerbates the phenotype of a model of Menkes disease in female
mice. Front. Neurosci. *19*, 1613171. https://doi.org/10.3389/fnins.2025.1613171.

33. Weishaupt, A.-K., Ruecker, L., Meiners, T., Schwerdtle, T., Silva Avila, D., Aschner, M.,
and Bornhorst, J. (2024). Copper-mediated neurotoxicity and genetic vulnerability in the
background of neurodegenerative diseases in C. elegans. Toxicol. Sci. Off. J. Soc. Toxicol.
201, 254–262. https://doi.org/10.1093/toxsci/kfae092.

34. Lane, A.R., Scher, N.E., Bhattacharjee, S., Zlatic, S.A., Roberts, A.M., Gokhale, A.,
Singleton, K.S., Duong, D.M., McKenna, M., Liu, W.L., et al. (2025). Adaptive protein
synthesis in genetic models of copper deficiency and childhood neurodegeneration. Mol.
Biol. Cell *36*, ar33. https://doi.org/10.1091/mbc.E24-11-0512.

35. Hwang, J.E.C., de Bruyne, M., Warr, C.G., and Burke, R. (2014). Copper overload and
deficiency both adversely affect the central nervous system of Drosophila†. Metallomics *6*,
2223–2229. https://doi.org/10.1039/c4mt00140k.

36. Norgate, M., Lee, E., Southon, A., Farlow, A., Batterham, P., Camakaris, J., and Burke, R.
(2006). Essential roles in development and pigmentation for the Drosophila copper
transporter DmATP7. Mol. Biol. Cell *17*, 475–484. https://doi.org/10.1091/mbc.e05-06-
0492.

37. Burke, R., Commons, E., and Camakaris, J. (2008). Expression and localisation of the
essential copper transporter DmATP7 in Drosophila neuronal and intestinal tissues. Int. J.
Biochem. Cell Biol. *40*, 1850–1860. https://doi.org/10.1016/j.biocel.2008.01.021.

38. Zlatic, S.A., Vrailas-Mortimer, A., Gokhale, A., Carey, L.J., Scott, E., Burch, R., McCall,
M.M., Rudin-Rush, S., Davis, J.B., Hartwig, C., et al. (2018). Rare Disease Mechanisms
Identified by Genealogical Proteomics of Copper Homeostasis Mutant Pedigrees. Cell Syst.
6, 368-380.e6. https://doi.org/10.1016/j.cels.2018.01.008.

39. Arafeh, R., Shibue, T., Dempster, J.M., Hahn, W.C., and Vazquez, F. (2025). The present
and future of the Cancer Dependency Map. Nat. Rev. Cancer *25*, 59–73.
https://doi.org/10.1038/s41568-024-00763-x.

40. Tsherniak, A., Vazquez, F., Montgomery, P.G., Weir, B.A., Kryukov, G., Cowley, G.S., Gill,
S., Harrington, W.F., Pantel, S., Krill-Burger, J.M., et al. (2017). Defining a Cancer
Dependency Map. Cell *170*, 564-576.e16. https://doi.org/10.1016/j.cell.2017.06.010.

41. Amici, D.R., Jackson, J.M., Truica, M.I., Smith, R.S., Abdulkadir, S.A., and Mendillo, M.L.
(2021). FIREWORKS: a bottom-up approach to integrative coessentiality network analysis.
Life Sci. Alliance *4*. https://doi.org/10.26508/lsa.202000882.

42. Ravinther, A.I., Dewadas, H.D., Tong, S.R., Foo, C.N., Lin, Y.-E., Chien, C.-T., and Lim,
Y.M. (2022). Molecular Pathways Involved in LRRK2-Linked Parkinson's Disease: A
Systematic Review. Int. J. Mol. Sci. *23*, 11744. https://doi.org/10.3390/ijms231911744.

43. Taymans, J.-M., Fell, M., Greenamyre, T., Hirst, W.D., Mamais, A., Padmanabhan, S., Peter,
I., Rideout, H., and Thaler, A. (2023). Perspective on the current state of the LRRK2 field.
Npj Park. Dis. *9*, 104. https://doi.org/10.1038/s41531-023-00544-7.

44. Trinh, J., and Farrer, M. (2013). Advances in the genetics of Parkinson disease. Nat. Rev.
Neurol. *9*, 445–454. https://doi.org/10.1038/nrneurol.2013.132.

45. Sanna, G., Del Giudice, M.G., Crosio, C., and Iaccarino, C. (2012). LRRK2 and vesicle
trafficking. Biochem. Soc. Trans. *40*, 1117–1122. https://doi.org/10.1042/BST20120117.

46. Zhang, B., Binks, T., and Burke, R. (2020). The E3 ubiquitin ligase Slimb/β-TrCP is
required for normal copper homeostasis in *Drosophila*. Biochim. Biophys. Acta BBA - Mol.
Cell Res. *1867*, 118768. https://doi.org/10.1016/j.bbamcr.2020.118768.

47. Green, C., Levashina, E., McKimmie, C., Dafforn, T., Reichhart, J.M., and Gubb, D. (2000).
The necrotic gene in Drosophila corresponds to one of a cluster of three serpin transcripts
mapping at 43A1.2. Genetics *156*, 1117–1127. https://doi.org/10.1093/genetics/156.3.1117.

48. Karpenko, M.N., Muruzheva, Z.M., Ilyechova, E.Y., Babich, P.S., and Puchkova, L.V.
(2023). Abnormalities in Copper Status Associated with an Elevated Risk of Parkinson's
Phenotype Development. Antioxid. Basel Switz. *12*, 1654.
https://doi.org/10.3390/antiox12091654.

49. Kinnart, I., Manders, L., Heyninck, T., Imberechts, D., Praschberger, R., Schoovaerts, N.,
Verfaillie, C., Verstreken, P., and Vandenberghe, W. (2024). Elevated α-synuclein levels
inhibit mitophagic flux. NPJ Park. Dis. *10*, 80. https://doi.org/10.1038/s41531-024-00696-0.

50. Sarkar, S., Olsen, A.L., Sygnecka, K., Lohr, K.M., and Feany, M.B. (2021). α-synuclein
impairs autophagosome maturation through abnormal actin stabilization. PLoS Genet. *17*,
e1009359. https://doi.org/10.1371/journal.pgen.1009359.

51. Petyuk, V.A., Yu, L., Olson, H.M., Yu, F., Clair, G., Qian, W.-J., Shulman, J.M., and
Bennett, D.A. (2021). Proteomic Profiling of the Substantia Nigra to Identify Determinants
of Lewy Body Pathology and Dopaminergic Neuronal Loss. J. Proteome Res. *20*, 2266–
2282. https://doi.org/10.1021/acs.jproteome.0c00747.

52. Vidyadhara, D.J., Lee, J.E., and Chandra, S.S. (2019). Role of the Endolysosomal System in
Parkinson's disease. J. Neurochem. *150*, 487–506. https://doi.org/10.1111/jnc.14820.

53. Lesage, S., Drouet, V., Majounie, E., Deramecourt, V., Jacoupy, M., Nicolas, A., Cormier-
Dequaire, F., Hassoun, S.M., Pujol, C., Ciura, S., et al. (2016). Loss of VPS13C Function in
Autosomal-Recessive Parkinsonism Causes Mitochondrial Dysfunction and Increases
PINK1/Parkin-Dependent Mitophagy. Am. J. Hum. Genet. *98*, 500–513.
https://doi.org/10.1016/j.ajhg.2016.01.014.

54. Niu, M., Zhao, F., Bondelid, K., Siedlak, S.L., Torres, S., Fujioka, H., Wang, W., Liu, J., and
Zhu, X. (2021). VPS35 D620N knockin mice recapitulate cardinal features of Parkinson's
disease. Aging Cell *20*, e13347. https://doi.org/10.1111/acel.13347.

55. Ma, K.Y., Fokkens, M.R., Reggiori, F., Mari, M., and Verbeek, D.S. (2021). Parkinson's
disease-associated VPS35 mutant reduces mitochondrial membrane potential and impairs
PINK1/Parkin-mediated mitophagy. Transl. Neurodegener. *10*, 19.
https://doi.org/10.1186/s40035-021-00243-4.

56. Fung, T.S., Ghosh, A., Zavala, M.R., Nichtova, Z., Shukal, D., Tigano, M., Csordas, G.,
Higgs, H.N., and Chakrabarti, R. (2025). Peri-mitochondrial actin filaments inhibit Parkin
assembly by disrupting ER-mitochondria contacts. EMBO Rep., 1–32.
https://doi.org/10.1038/s44319-025-00561-y.

57. Kumar, N., Leonzino, M., Hancock-Cerutti, W., Horenkamp, F.A., Li, P., Lees, J.A.,
Wheeler, H., Reinisch, K.M., and De Camilli, P. (2018). VPS13A and VPS13C are lipid
transport proteins differentially localized at ER contact sites. J. Cell Biol. *217*, 3625–3639.
https://doi.org/10.1083/jcb.201807019.

58. Burman, J.L., Bourbonniere, L., Philie, J., Stroh, T., Dejgaard, S.Y., Presley, J.F., and
McPherson, P.S. (2008). Scyl1, mutated in a recessive form of spinocerebellar
neurodegeneration, regulates COPI-mediated retrograde traffic. J. Biol. Chem. *283*, 22774–
22786. https://doi.org/10.1074/jbc.M801869200.

59. Taunton, J., Rowning, B.A., Coughlin, M.L., Wu, M., Moon, R.T., Mitchison, T.J., and
Larabell, C.A. (2000). Actin-dependent propulsion of endosomes and lysosomes by
recruitment of N-WASP. J. Cell Biol. *148*, 519–530. https://doi.org/10.1083/jcb.148.3.519.

60. Buenaventura, T., Kanda, N., Douzenis, P.C., Jones, B., Bloom, S.R., Chabosseau, P.,
Corrêa, I.R., Bosco, D., Piemonti, L., Marchetti, P., et al. (2018). A Targeted RNAi Screen
Identifies Endocytic Trafficking Factors That Control GLP-1 Receptor Signaling in
Pancreatic β-Cells. Diabetes *67*, 385–399. https://doi.org/10.2337/db17-0639.

61. Xue, Y., Schmollinger, S., Attar, N., Campos, O.A., Vogelauer, M., Carey, M.F., Merchant,
S.S., and Kurdistani, S.K. (2017). Endoplasmic reticulum-mitochondria junction is required
for iron homeostasis. J. Biol. Chem. *292*, 13197–13204.
https://doi.org/10.1074/jbc.M117.784249.

62. Bórquez, D.A., Castro, F., Núñez, M.T., and Urrutia, P.J. (2022). New Players in Neuronal
Iron Homeostasis: Insights from CRISPRi Studies. Antioxid. Basel Switz. *11*, 1807.
https://doi.org/10.3390/antiox11091807.

63. Gatti, P., Schiavon, C., Cicero, J., Manor, U., and Germain, M. (2025). Mitochondria- and
ER-associated actin are required for mitochondrial fusion. Nat. Commun. *16*, 451.
https://doi.org/10.1038/s41467-024-55758-x.

64. Metzler, M., Li, B., Gan, L., Georgiou, J., Gutekunst, C.-A., Wang, Y., Torre, E., Devon,
R.S., Oh, R., Legendre-Guillemin, V., et al. (2003). Disruption of the endocytic protein HIP1
results in neurological deficits and decreased AMPA receptor trafficking. EMBO J. *22*,
3254–3266. https://doi.org/10.1093/emboj/cdg334.

65. Bhattacharyya, N.P., Banerjee, M., and Majumder, P. (2008). Huntington's disease: roles of
huntingtin-interacting protein 1 (HIP-1) and its molecular partner HIPPI in the regulation of
apoptosis and transcription. FEBS J. *275*, 4271–4279. https://doi.org/10.1111/j.1742-
4658.2008.06563.x.

66. Sone, M., Uchida, A., Komatsu, A., Suzuki, E., Ibuki, I., Asada, M., Shiwaku, H., Tamura,
T., Hoshino, M., Okazawa, H., et al. (2009). Loss of yata, a novel gene regulating the
subcellular localization of APPL, induces deterioration of neural tissues and lifespan
shortening. PloS One *4*, e4466. https://doi.org/10.1371/journal.pone.0004466.

67. Schmidt, W.M., Kraus, C., Höger, H., Hochmeister, S., Oberndorfer, F., Branka, M.,
Bingemann, S., Lassmann, H., Müller, M., Macedo-Souza, L.I., et al. (2007). Mutation in the
Scyl1 gene encoding amino-terminal kinase-like protein causes a recessive form of
spinocerebellar neurodegeneration. EMBO Rep. *8*, 691–697.
https://doi.org/10.1038/sj.embor.7401001.

68. Pelletier, S., Gingras, S., Howell, S., Vogel, P., and Ihle, J.N. (2012). An early onset
progressive motor neuron disorder in Scyl1-deficient mice is associated with mislocalization
of TDP-43. J. Neurosci. Off. J. Soc. Neurosci. *32*, 16560–16573.
https://doi.org/10.1523/JNEUROSCI.1787-12.2012.

69. Cerri, S., Mus, L., and Blandini, F. Parkinson's Disease in Women and Men: What's the
Difference? J. Park. Dis. *9*, 501–515. https://doi.org/10.3233/JPD-191683.

70. Moisan, F., Kab, S., Mohamed, F., Canonico, M., Guern, M.L., Quintin, C., Carcaillon, L.,
Nicolau, J., Duport, N., Singh-Manoux, A., et al. (2016). Parkinson disease male-to-female
ratios increase with age: French nationwide study and meta-analysis. J. Neurol. Neurosurg.
Psychiatry *87*, 952–957. https://doi.org/10.1136/jnnp-2015-312283.

71. Tümer, Z., and Møller, L.B. (2010). Menkes disease. Eur. J. Hum. Genet. *18*, 511–518.
https://doi.org/10.1038/ejhg.2009.187.

72. Aschner, M., Skalny, A.V., Lu, R., Martins, A.C., Tizabi, Y., Nekhoroshev, S.V.,
Santamaria, A., Sinitskiy, A.I., and Tinkov, A.A. (2024). Mitochondrial pathways of copper
neurotoxicity: focus on mitochondrial dynamics and mitophagy. Front. Mol. Neurosci. *17*,
1504802. https://doi.org/10.3389/fnmol.2024.1504802.

73. Lu, J., Liu, X., Li, X., Li, H., Shi, L., Xia, X., He, B.-L., Meyer, T.F., Li, X., Sun, H., et al.
(2024). Copper regulates the host innate immune response against bacterial infection via
activation of ALPK1 kinase. Proc. Natl. Acad. Sci. U. S. A. *121*, e2311630121.
https://doi.org/10.1073/pnas.2311630121.

74. O'Sullivan, E., O'Shea, D., Devereux, M., Kavanagh, K., and Howe, O. (2026). Autophagy-
dependent cell death and immune responses induced by heteroleptic copper(II),
manganese(II), and silver(I) complexes containing dicarboxylate and 1,10-phenanthroline
ligands: novel insights from in vitro and in vivo models. Biometals Int. J. Role Met. Ions
Biol. Biochem. Med. *39*, 337–358. https://doi.org/10.1007/s10534-025-00766-7.

75. Duncan, C., Bica, L., Crouch, P.J., Caragounis, A., Lidgerwood, G.E., Parker, S.J.,
Meyerowitz, J., Volitakis, I., Liddell, J.R., Raghupathi, R., et al. (2013). Copper modulates
the large dense core vesicle secretory pathway in PC12 cells. Met. Integr. Biometal Sci. *5*,
700–714. https://doi.org/10.1039/c3mt20231c.

76. Seo, Y., Cho, Y.-S., Huh, Y.-D., and Park, H. (2016). Copper Ion from Cu2O Crystal
Induces AMPK-Mediated Autophagy via Superoxide in Endothelial Cells. Mol. Cells *39*,
195–203. https://doi.org/10.14348/molcells.2016.2198.

77. Lee, B.D., Shin, J.-H., VanKampen, J., Petrucelli, L., West, A.B., Ko, H.S., Lee, Y.-I.,
Maguire-Zeiss, K.A., Bowers, W.J., Federoff, H.J., et al. (2010). Inhibitors of leucine-rich
repeat kinase-2 protect against models of Parkinson's disease. Nat. Med. *16*, 998–1000.
https://doi.org/10.1038/nm.2199.

78. Liu, Z., Wang, X., Yu, Y., Li, X., Wang, T., Jiang, H., Ren, Q., Jiao, Y., Sawa, A., Moran,
T., et al. (2008). A Drosophila model for LRRK2-linked parkinsonism. Proc. Natl. Acad. Sci.
U. S. A. *105*, 2693–2698. https://doi.org/10.1073/pnas.0708452105.

Figure Legends

Figure 1. ATP7 Interacts with Parkinson's Disease/Neurodegeneration Genes in the

Epidermal Epithelium in Females. Images of inhibition of ATP7 alone **A)** or in combination

with inhibition of different ATP7 interactome homologues using the pnr-GAL4 in females **B-L).**

Yellow arrows indicate widening of the thoracic midline bristles, red arrows indicate areas of

thoracic caving in, black arrows show scutellar bristle defects, and yellow arrowhead indicates

bristle disorganization.

Figure 2. ATP7 Interacts with Parkinson's Disease/Neurodegeneration Genes in the

Epidermal Epithelium in Males. Images of inhibition of ATP7 alone **A)** or in combination with

inhibition of different ATP7 interactome homologues using the pnr-GAL4 in males **B-L).**

Yellow arrows indicate areas of thoracic caving in, and red arrows indicate areas of thoracic

caving in, black arrows show scutellar bristle defects, and yellow arrowhead indicates missing

and disorganized thoracic bristles.

Figure 3. ATP7 Interacts with Lrrk in the Epidermal Epithelium in Females. Images of control **A)** and different Lrrk RNAi lines **B-D)** using the pnr-GAL4 in females. ATP7 down-regulation alone **E)** or in combination with inhibition of Lrrk **F-H)**. ATP7 over-expression **I)** alone or in combination with inhibition of Lrrk **J-L)**. Yellow arrows indicate widening of the bristles along the midline of the thorax. Red arrows indicate thoracic caving in, and yellow arrowheads indicate bristle loss.

Figure 4. ATP7 Interacts with Lrrk in the Epidermal Epithelium in Males. Images of control **A)** and different Lrrk RNAi lines **B-D)** using the pnr-GAL4 in males. ATP7 down-regulation alone **E)** or in combination with inhibition of Lrrk **F-H)**. ATP7 over-expression **I)** alone or in combination with inhibition of Lrrk **J-L)**. Red arrows indicate thoracic caving in, and black arrowheads indicate necrotic tissue.

Figure 5. Lrrk Interacts with ATP7 Down-regulation and Copper Exposure in Dopaminergic Neurons. Female **A-C)** and male **D-F)** flies exposed to copper with either inhibition of ATP7 alone or in combination with Lrrk inhibition using the Ddc-GAL4.

Figure 6. Lrrk Interacts with ATP7 Over-expression and Viability. The normalized ratio of female **A-C)** and male **D-F)** flies that were expected to reach adulthood (viability) with either down-regulation of ATP7 alone or in combination with Lrrk inhibition using the Ddc-GAL4. For Lrrk RNAi line 1 females, statistical results $\chi^2 = 10.93$, df $= 3$, p-value $= 0.01211$. For Lrrk RNAi line 2 females, statistical results $\chi^2 = 120.44$, df $= 3$, p-value $< 2.2e-16$. For Lrrk RNAi line 3 females, statistical results $\chi^2 = 41.642$, df $= 3$, p-value $= 4.778e-09$. For Lrrk RNAi line 1 males, statistical results $\chi^2 = 12.034$, df $= 3$, p-value $= 0.007268$. For Lrrk RNAi line 2 males, statistical results $\chi^2 = 141.32$, df $= 3$, p-value $< 2.2e-16$. For Lrrk RNAi line 3 females, statistical results $\chi^2 = 31.117$, df $= 3$, p-value $= 8.03e-07$.

Figure 7. Lrrk Interacts with ATP7 Over-expression and Copper Exposure in Dopaminergic Neurons. Female **A-C)** and male **D-F)** flies exposed to copper with either over-expression of ATP7 alone or in combination with Lrrk inhibition using the Ddc-GAL4.

Figure S1. Coessentiality Network Analysis of ATP7A.

Coessentiality network analysis showing genome-scale fitness correlations of ATP7A and Parkinson's and neurodegeneration genes as inputs. Nodes correspond to first and second level interaction nodes. Network of correlated (red edges) and anticorrelated genes (blue edges) was built with the FIREWORKS tool.

Figure S2. Inhibition of Lrrk in the Epidermal Epithelium in Females. Images of inhibition of ATP7 interactome genes using the pnr-GAL4 in females. Yellow arrows indicate missing and disorganization thoracic midline bristles. Black arrows show loss of scutellar bristles.

Figure S3. Inhibition of Lrrk in the Epidermal Epithelium in Males. Images of inhibition of ATP7 interactome genes using the pnr-GAL4 in females. Yellow arrows indicate missing and disorganization thoracic midline bristles. Black arrows show loss of scutellar bristles.

Figure S4. Lrrk Interacts with ATP7 Down-regulation and Viability. The normalized ratio of female **A-C) and** male **D-F)** flies that were expected to reach adulthood (viability) with either down-regulation of ATP7 alone or in combination with Lrrk inhibition using the Ddc-GAL4. For Lrrk RNAi line 1 females, statistical results $\chi^2 = 7.1566$, df = 3, p-value = 0.06707. For Lrrk RNAi line 2 females, statistical results $\chi^2 = 112.05$, df = 3, p-value < 2.2e-16. For Lrrk RNAi line 3 females, statistical results $\chi^2 = 47.872$, df = 3, p-value = 2.267e-10. For Lrrk RNAi line 1 males, statistical results $\chi^2 = 6.052$, df = 3, p-value = 0.1091. For Lrrk RNAi line 2 males, statistical results $\chi^2 = 97.556$, df = 3, p-value < 2.2e-16. For Lrrk RNAi line 3 females, statistical results $\chi^2 = 64.697$, df = 3, p-value = 5.823e-14.

Table 1. *Drosophila* Homologues of ATP7A Interactome Genes
746

Table S1. Summary of Interactions Between ATP7 Down-regulation and Interactome Genes in the Epidermal Epithelium
749

Table S2. *Drosophila* Genotypes
751

Table S3. Statistical Comparisons of Lrrk Inhibition and ATP7 Down-regulation Viability

Table S4. Statistical Comparisons of Glucose Treated Lrrk and ATP7 Down-regulation or
Over-expression Survival

Table S5. Statistical Comparisons of Copper Treated Lrrk and ATP7 Down-regulation
Survival

Table S6. Statistical Comparisons of Lrrk Inhibition and ATP7 Over-expression Viability

Table S7. Statistical Comparisons of Copper Treated Lrrk and ATP7 Over-expression
Survival

Females

Wild-type · ATP7 RNAi · ATP7 RNAi Arpc1 RNAi · ATP7 RNAi Gyf RNAi · ATP7 RNAi Hip1 RNAi · ATP7 RNAi Synj RNAi -1

ATP7 RNAi Synj RNAi -2 · ATP7 RNAi Vps13 RNAi-1 · ATP7 RNAi Vps13 RNAi-2 · ATP7 RNAi Vps35 RNAi · ATP7 RNAi yata RNAi -1 · ATP7 RNAi yata RNAi -3

Figure 1

Males

Figure 2

Females

Wild-Type	LRRK RNAi -1	LRRK RNAi -2	LRRK RNAi -3
A	B	C	D

ATP7 RNAi	ATP7 RNAi LRRK RNAi -1	ATP7 RNAi LRRK RNAi -2	ATP7 RNAi LRRK RNAi -3
E	F	G	H

ATP7 OE	ATP7 OE LRRK RNAi -1	ATP7 OE LRRK RNAi -2	ATP7 OE LRRK RNAi -3
I	J	K	L

Figure 3

Males

Wild-Type | LRRK RNAi -1 | LRRK RNAi -2 | LRRK RNAi -3

A | B | C | D

ATP7 RNAi | ATP7 RNAi LRRK RNAi -1 | ATP7 RNAi LRRK RNAi -1 | ATP7 RNAi LRRK RNAi -2 | ATP7 RNAi LRRK RNAi -3

E | F | F' | G | H

ATP7 OE | ATP7 OE LRRK RNAi -1 | ATP7 OE LRRK RNAi -2 | ATP7 OE LRRK RNAi -2 | ATP7 OE LRRK RNAi -3

I | J | K | K' | L

Figure 4

Figure 5

Figure 6

Figure 7

Drosophila Candidate Gene	Drosophila Gene Abbreviation	Human Homolog Gene Name	Human Homolog Abbreviation	OMIM Number	PARK Number
Actin-related protein 2/3 complex, subunit 1	Arpc1	Actin-Related Protein 2/3 Complex	ARPC1	604223	N/A
Gigyf	Gyf	GRB10 interaction GYF protein 2	GIGYF2	607688	N/A
Huntingtin interacting protein 1	Hip1	Huntingtin interacting protein 1	HIP1	601767	PARK11
Leucine-rich repeat kinase	Lrrk	Leucine rich repeat kinase 2	Lrrk2	607060	PARK8
Synaptojanin	Synj	Synaptojanin 1	Synj1	615530	PARK20
Vacuolar protein sorting 13	Vps13	Vacuolar protein sorting 13C	VPS13C	616840	PARK23
Vacuolar protein sorting 35	Vps35	VPS35 retromer complex component	VPS35	601501	PARK17
yata	yata	SCYL like pseudokinase 1	SCYL1	616719	N/A

Table 1. *Drosophila* Homologues of ATP7A Interactome Genes