Genomic selection for seed yield enhances flax breeding efficiency

Frank M. You[1*], Chunfang Zheng[1], John Joseph Zagariah Daniel[1], Pingchuan Li[1], Kenneth Jackle[2], Megan House[2], Bunyamin Tar'an[2], Sylvie Cloutier[1]

[1] Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, 960 Carling Avenue, Ottawa, ON, K1A 0C6, Canada

[2] Crop Development Centre, Department of Plant Sciences, University of Saskatchewan, 51 Campus Drive, Saskatoon, SK, S7N 5A8, Canada

***** Correspondence: Frank M. You, Email: frank.you@agr.gc.ca; Tel.: +1-613-314-2596.

Abstract

Genomic selection (GS) is a promising strategy to improve breeding efficiency for complex traits such as seed yield by enabling early selection and reducing reliance on extensive field testing. However, practical deployment of GS remains challenging due to limited training populations sizes and reduced prediction accuracies when models are applied to true breeding germplasm. In this study, we evaluated GS for flax (*Linum usitatissimum* L.) seed yield under realistic breeding scenarios, with a focus on across-population prediction (APP) and breeding decision support rather than model benchmarking. Using historical germplasm collections and a newly developed breeding-oriented population as training sets, GS performance was assessed across multiple independent test populations representing contemporary breeding lines evaluated in replicated yield trials. APP accuracies reached r = 0.84 when training and test populations were genetically aligned, supporting routine breeding deployment. Training population composition emerged as a key determinant of prediction success, with breeding-oriented populations consistently outperforming broad germplasm collections for predicting true breeding lines. Check-based selection analyses showed that GS reliably reproduced phenotypic advancement decisions while eliminating 61–91% of low-performing lines, resulting in 48–78% reduction in field evaluation costs for a typical cohort of 300 lines. Marker subsampling analyses further indicated that moderate-density genotyping-by-sequencing panels (~2,500–3,000 SNPs) are sufficient to achieve stable prediction accuracies. Overall, these results demonstrate that GS for seed yield in flax is ready for routine integration into breeding programs, offering a practical pathway to reduce costs, accelerate breeding cycles, and enhance selection efficiency.

Keywords: flax; genomic selection; seed yield; across-population prediction; training population design; breeding efficiency; genotyping-by-sequencing; check-based selection

Introduction

Flax (*Linum usitatissimum* L.) is a dual-purpose crop cultivated for both oilseed (linseed) and fiber production, with distinct breeding objectives for each end use (Duk et al. 2021; Soto-Cerda et al. 2013; Povkhova et al. 2021; Praczyk and Wielgusz 2021). In linseed breeding programs, seed yield is the single most important selection target, followed by oil content and quality. As in all crops, genetic improvement for seed yield is challenging due to its complex genetic architecture, environmental sensitivity, and genotype-by-environment interactions. Consequently, phenotypic selection for YLD relies heavily on multi-year, multi-location field trials, making yield improvement both time-consuming and costly.

Genomic selection (GS), which uses genome-wide markers to predict genomic estimated breeding values (GEBVs), has emerged as a promising strategy to accelerate genetic gain for complex traits such as yield (Meuwissen et al. 2001; Olatoye et al. 2019; Beyene et al. 2019; Kumar et al. 2024). By enabling early-generation selection and reducing dependence on extensive field testing, GS has the potential to shorten breeding cycles and improve resource efficiency. In flax, GS has been explored for seed yield, agronomic traits, seed quality, and disease resistance (Hoque et al. 2024; Khan et al. 2023; Lan et al. 2020; He et al. 2018; You et al. 2022). Despite these advances, routine deployment of GS for seed yield in flax breeding programs remains limited, largely due to the small size of the training populations, shifting breeding pools, and uncertainties about the abilities of the prediction models to perform when applied to breeding germplasm unrelated to the training set(s).

A key limitation of many GS studies is their reliance on within-population cross-validation (CV) to assess prediction accuracy (PA). Because CV evaluates prediction within the same population used for model training, it often inflates PA. In practical breeding, however, GS models must predict untested breeding lines evaluated across different years and environments, a scenario more appropriately captured by across-population prediction (APP).Studies in cereals and legumes have shown that prediction accuracies decline sharply under APP (Lozada et al. 2019; Juliana et al. 2018; Vitale et al. 2025), highlighting the need to evaluate GS strategies

under conditions that reflect actual breeding deployments. Rather than benchmarking model architectures under idealized conditions, this study focuses on evaluating GS performance under deployment-oriented scenarios that mirror real breeding decision scenarios.

Beyond model choice, emerging evidence suggests that training population composition and genetic relevance to breeding targets are dominant factors governing APP performance. Training sets based on broad germplasm collections provide extensive diversity but often include genetic variation that is weakly informative for predicting elite breeding germplasm. In contrast, breeding-oriented training sets composed of recent selections and parental lines better represent the genetic architecture of future candidates, thereby improving prediction robustness. However, systematic evaluations of training set designs, marker types, and practical decision outcomes for seed yield prediction in flax are still lacking.

The objectives of this study were therefore to evaluate GS for flax seed yield under actual breeding scenarios, with an emphasis on APP and breeding decision support rather than model benchmarking. Specifically, we aimed to (i) assess how training set design influences PA across independent breeding populations, (ii) compare GS performance of multiple marker types under APP conditions, (iii) determine the minimum marker density required for reliable prediction of breeding lines, and (iv) evaluate the practical utility of GS for check-based selection decisions, and in terms of cost reduction and breeding cycle acceleration.

By integrating historical germplasm collections with newly developed breeding populations and evaluating GS performance across multiple independent test datasets, this study provides a deployment-oriented assessment of GS for seed yield in flax. Our results offer practical guidance for training population construction, genotyping strategy, and GS integration into routine flax breeding pipelines.

Materials and methods

Training populations

A core collection was originally assembled to capture the genetic diversity of the global flax germplasm (Diederichsen et al. 2013). The initial collection comprised 381 accessions from 38 countries, including fiber and linseed landraces, breeding lines, and cultivars (You et al. 2017). To better represent modern breeding germplasm, 10 breeding lines and cultivars from Canadian

breeding programs were incorporated, expanding the core collection to 391 accessions. The final set included 298 linseed and 93 fiber accessions, of which 378 lines had complete phenotypic datasets. Two GS training sets based on the core collection were defined: CORE287, consisting of 287 linseed accessions, and CORE378, comprising 378 accessions representing both linseed and fiber flax.

To better represent contemporary breeding germplasm and increase genetic diversity and relatedness to current breeding materials, a breeding-oriented training population, designated BP296, was developed. BP296 contains two germplasm components. The first component included 45 varieties and breeding lines that have been used as parents in recent years in the Crop Development Centre (CDC, University of Saskatchewan, Canada) flax breeding program. This set included cultivars released within the past 15 years, which were largely absent from the original core collection, as well as germplasm recently introduced from foreign breeding programs. Because these accessions have been extensively used as parental lines, they provide strong genetic connectivity to contemporary breeding populations. These 45 parental lines were not phenotyped in the same historical trials as the CORE populations and therefore could not be directly merged with the core collection. To generate a unified dataset, 254 breeding lines derived from recent breeding populations were selected in 2023. These 254 lines and the 45 parental accessions were evaluated together in the same field experiments conducted in 2025, ensuring directly comparable phenotypic data. After quality control, 296 of 299 lines with complete genotypic and phenotypic data were retained, forming the BP296 dataset, which was used as both a training and a test dataset in GS analyses.

Test populations

In addition to BP296, three independent test populations were used for APP. The first test population comprised a combined set of 260 lines (255 inbred lines plus five of the six parents (excluding the reference cultivar CDC Bethune) from three biparental populations (BM, EV, and SU) with distinct genetic backgrounds (You et al. 2018a). The BM population comprised 91 F_6-derived recombinant inbred lines (RILs) from a cross between the high-yielding cultivars CDC Bethune and Macbeth. The EV population consisted of 88 F_6-derived RILs from a cross between the low linolenic acid breeding line E1747 and the French fiber cultivar Viking. The SU population contained 76 F_1-derived doubled haploid (DH) lines from a cross between the yellow-

seeded, low linolenic acid breeding line SP2047 and the high linolenic acid breeding line UGG5-5.

The second and third test populations consisted of yellow-seeded (YS) and brown-seeded (BS) breeding lines, targeting the newer yellow-seeded and the traditional, brown-seeded market classes, respectively. These lines were selected from the CDC's flax breeding program in 2023 and 2024. A total of 33 YS lines and 57 BS lines, together with their respective parent and check cultivars, were evaluated in four independent yield trials conducted across three or four locations during 2023–2024 using randomized complete block designs (RCBD) with three replications per location (**Table 1**). For GS analyses, the YS lines, associated parents and check cultivars were designated YS38, and the similar BS set was designated BS61.

Table 1 Summary of yield trials used for genomic prediction of seed yield.

Yield trial	**Year**	**Code**[†]	**No. of test lines**	**Location**	**Check/parent cultivars**[‡]
Yellow seed secondary	2024	2024YSS (YS38)	28	U of S Campus, Kernen, Melfort	**AAC Bright (2017)**, CDC Dorado (2016), CDC Melyn (2018), and Double-Low
Yellow seed secondary and tertiary	2023-2024	2023–24YST (YS38)	5	U of S Campus, Kernen, Melfort, Avonlea (2024 only)	**AAC Bright (2017)**, CDC Dorado (2015), and NuLin VT50 (2009)
Brown seed secondary	2024	2024BSS (BS61)	37	U of S Campus, Kernen, Melfort	CDC Kernen (2021), CDC Esme (2023), **CDC Glas (2012)**
Brown seed secondary and tertiary	2023-2024	2023–24BST (BS61)	20	U of S Campus, Kernen, Melfort, Avonlea (2024 only)	CDC Kernen (2021), CDC Esme (2023), **CDC Glas (2012)**, and CDC Rowland (2018)
Preliminary	2025	BP296	299	U of S Campus	**CDC Glas (2012), AAC Bright (2017),** CDC Bethune (1998), CDC Kernen (2021), CDC Plava (2015)

[†] Codes indicate the experiment, with test population codes shown in parentheses. [‡] National check cultivars (in bold) are AAC Bright for yellow-seeded, and CDC Glass for brown-seeded

trials. CDC Rowland is an internal check. Numbers in parentheses denote years of cultivar release. U of S: University of Saskatchewan.

Sequencing and single nucleotide polymorphism (SNP) markers

All individuals of the flax core collection (CORE378), the combined biparental population (BMEVSU260) and the 45 varieties and breeding lines of BP296 were genotyped by whole-genome sequencing (WGS) on Illumina platforms using the 100- (PE100) or 150-bp (PE150) paired end read mode, as previously described (You et al. 2018a; He et al. 2023). The 99 breeding lines and check/parent cultivars from the CDC flax breeding program (YS38 and BS61) and the 254 breeding lines of the BP296 set were genotyped using genotyping-by-sequencing (GBS) with PE150 reads using the *Pst*I–*Msp*I enzyme protocol (Elshire et al. 2011).

All lines were grown in root trainers (72 plants per tray) in a growth chamber under a 20 h light/4 h dark cycle (24°C/18°C). When seedlings were 5-8 cm tall, approximately 50–80 mg of leaf tissue was collected from the shoot tip into Qiagen 96-well collection tubes pre-cooled on dry ice. Samples were lyophilized for 48-72 h in a FreeZone 6 L benchtop freeze dryer (Labconco, Kansas City, MO, USA) and stored at −20°C until DNA extraction.

For GBS, genomic DNA (gDNA) was extracted with the Qiagen DNeasy® kit (Qiagen, Toronto, ON, Canada) and quantified using the Quant-iT™ PicoGreen™ dsDNA assay (ThermoFisher Scientific, Waltham, MA, USA) following the manufacturers' instructions. The gDNA was normalized to 10 ng/µl, and 200 ng samples were submitted to the Université Laval's Institut de biologie intégrative et des systèmes (IBIS; Quebec, QC, Canada) for library preparation. GBS libraries were pooled and sequenced on the Illumina HiSeq 2500 platform to generate PE150 reads, as previously described in Cloutier et al. (2024).

For SNP discovery, all reads were aligned to the CDC Bethune v3.0 reference genome (NCBI accession JAZBJT000000000, You et al., in preparation) using bwa v0.7.19 (Li and Durbin 2009). Single nucleotide polymorphisms (SNPs) were called with SAMtools v1.22.1 (Li et al. 2009) and filtered using the following threshold criteria: read depth > 3, mapping quality (MAPQ) on the Phred scale > 20, minor allele frequency (MAF) ≥ 0.05, call rate ≥ 80%, heterozygosity ≤ 20% (per SNP), and exclusion of multi-allelic sites. The workflow was implemented in the improved AGSNP pipeline (Kumar et al. 2012; You et al. 2012). After quality control using the above criteria, SNPs in all training and test datasets were imputed using Beagle

v5.5 with default parameters (Browning et al. 2021). The distribution of filtered SNPs was visualized using the CMplot R package (v4.5.1) with a 100-Kb window size (Yin et al. 2021).

Haplotype and PC marker construction

In addition to SNP, haplotype block (HAP) and principal component (PC) markers were generated. HAPs were estimated with the rtm-gwas-snpldb tool in RTM-GWAS v2020.0 (He et al. 2017), which applies the Haploview "Gabriel" confidence-interval algorithm (Gabriel et al. 2002; Barrett et al. 2005). Pairwise linkage disequilibrium (LD) was measured using D' with 95% confidence intervals. SNP pairs were classified as strong LD, inconclusive, or strong recombination, and HAPs were defined when ≥95% of informative pairs were in strong LD (default thresholds: CI(D') lower ≥ 0.70, upper ≥ 0.98). PC markers were obtained by principal component analysis (PCA) of the SNP dataset when the first *N* PCs explaining 95% of total variance were retained for prediction.

Yield data analysis

The core collection (378 accessions) was phenotyped for seed yield over four years (2009–2012) at two locations: the Morden Research and Development Centre Farm, Morden, Manitoba, and the Kernen Crop Research Farm, near Saskatoon, Saskatchewan, Canada (You et al. 2017). The experimental design was a type-2 modified augmented design (MAD2) (Lin and Poushinsky 1985). Yields were adjusted for soil heterogeneity using the MAD pipeline (You et al. 2013). Best linear unbiased predictions (BLUPs) across years and locations were obtained using *lme4* v1.1-37 (Bates et al. 2015), with genotypes, years and locations treated as random effects.

The BM, EV and SU populations were phenotyped at the same locations. BM was evaluated for four years (2009–2012), and EV and SU for three years (2010–2012), also using MAD2 designs. Phenotypic adjustments and BLUP estimation followed the same procedure as the core collection (You et al. 2013).

The 90 selected breeding lines and nine checks/parents making out the test populations were evaluated in independent yield trials in 2023–2024 at three (secondary trials) or four (tertiary trials) locations (Table 1). Linear mixed models were fitted using test lines (genotypes) as fixed effects, and blocks (replications), trials, locations, years, and lines × locations as random

effects. The yield estimates of lines measured through the mixed model were considered observed ("true") breeding values.

Field evaluation of the BP296 population was conducted using a structured multi-experiment design to control field heterogeneity and improve phenotypic reliability. The 296 lines were divided into eight subsets, each containing 36–37 lines. Each subset was evaluated as an independent experiment using RCBD with two replications. Five common check cultivars were included in every experiment, resulting in 41–42 entries per experiment and a total of eight parallel experiments. To further assess data robustness and minimize potential seed source effects, a second, independent set of eight experiments was conducted in a separate field using the same experimental design and line allocation. The only difference between the two experimental sets was the seed source: one set used seeds increased in a greenhouse at AAFC (Ottawa, Canada), and the other used seeds provided by the CDC which were advanced in New Zealand during the winter of 2024. For both seed sources, seed increases were derived from a single plant per line to ensure genetic uniformity.

Phenotypic data from the two experimental sets were first analyzed separately using a two-stage multi-environment trial (MET) approach (Piepho et al. 2012; Piepho et al. 2008). In the first stage, adjusted least squared means were obtained for each line within individual experiments using mixed linear models accounting for block effects. In the second stage, line effects were jointly analyzed across experiments to obtain overall seed yield estimates. Seed yield estimates derived from the two experimental sets were highly correlated (Pearson's r = 0.85), indicating strong consistency between seed sources. Consequently, data from both sets were combined, and a final two-stage MET analysis was performed to generate consolidated BLUPs for seed yield, which were used in subsequent GS analyses.

Genomic selection models and prediction accuracy evaluation

Sixteen GS models implemented in the MultiGS-P pipeline (You et al. 2026) were evaluated. These included four linear and regularized models (R_RRBLUP, R_GBLUP, Elastic Net, and BRR), three machine learning (ML) models (Random Forest Regression (RFR), XGBoost and LightGBM), six deep learning (DL) models, two hybrid model that integrate RR-BLUP with deep networks (DeepResBLUP and DeepBLUP), as well as a stacking-based ensemble learner (EnsembleGS). In EnsembleGS, predictions from R_RRBLUP, Elastic Net, BRR, LightGBM,

and MLPGS were combined through model stacking. Default hyperparameters, which were carefully tuned using the flax datasets and provided by MultiGS-P (You et al. 2026), were applied to all ML and DL models. For each model, SNP, HAP, and PC marker types were evaluated.

PA was assessed as Pearson correlation between GEBVs and observed phenotypic values. Two evaluation strategies were employed: (i) five-fold CV within training populations, repeated across 10 training-test partitions (yielding 50 total CV replicates, as each partition generated five folds used in turn as the test set), to assess model and marker-representation performances, and (ii) APP, in which three training populations were used to predict four independent breeding line datasets.

Evaluation of prediction accuracies across marker numbers in across-population prediction

To determine the number of SNP markers required for reliable prediction of new breeding lines, simulations were conducted using randomly sampled SNP subsets. For the BMEVSU260 test dataset, 33,596 SNPs shared between the test and training populations were available, from which subsets ranging from 500 to 10,000 SNPs (in increments of 500) were generated, with ten random replications per subset. The rationale for capping the SNPs at 10,000 was two-fold: (i) preliminary analyses had demonstrated that PA reached a plateau when marker density exceeded that number, and (ii) the aim was to determine the minimum number of markers required for practical APP. For the BP296 test population, approximately 6,000 SNPs were common between the test and training populations, and subsets ranging from 500 to 5,500 SNPs (in increments of 500) were similarly generated, with ten replications per subset.

The representative GS models R_RRBLUP, LightGBM, and DeepBLUP, implemented in MultiGS-P, were evaluated using SNP markers. PAs were then compared across incremental marker numbers and models to assess the trade-off between marker density and APP performance.

Population genetic diversity and differentiation analysis

To access genetic diversity and relatedness of training and test populations, nucleotide diversity within populations and genetic differentiation between populations were quantified using harmonized SNP markers. For each population, SNP-based nucleotide diversity (π) was

calculated as the mean pairwise nucleotide difference per SNP. Prior to analysis, SNPs were harmonized across all VCF files of training and test datasets to retain only shared loci with consistent reference and alternate alleles.

Pairwise population genetic differentiation was estimated using the Weir–Cockerham FST statistic (Weir and Cockerham 1984) based on shared SNPs between population pairs. Genotype data from each pair of populations were combined, and subpopulation assignments were used to calculate FST components following the original Weir–Cockerham formula. All diversity and differentiation statistics were computed using a custom Python pipeline built on the scikit-allel framework, which is implemented in the MultiGS toolkiet (https://github.com/AAFC-ORDC-Crop-Bioinfomatics/MultiG).

Results

Training and test datasets

A total of six flax populations were included in this study, comprising three training populations and four test populations that collectively represent both historical germplasm collections and contemporary breeding germplasm (Table 2).

Table 2 Summary of training and test populations used in flax genomic selection.

Population type	**Population description**	**Code**	**Lines with trait data**	**Total filtered SNPs** †	**Common SNPs used for prediction**‡	**Nucleotide diversity (π)**
Training	Core collection	CORE378	378	1,699,363	5,604	0.3991
	Linseed subset of the core collection	CORE287	287	1,696,929	5,604	0.4007
Training/Test	Breeding population (253 breeding lines + 43 parents)	BP296	253	11,235	5,604	0.3838
			43	1,218,703	5,604	
Test	Combined biparental population	BMEVSU260	260	43,268	33,596	0.3727
	Yellow-seeded breeding lines	YS38	38	11,714	5,604	0.3549

Population type	Population description	Code	Lines with trait data	Total filtered SNPs †	Common SNPs used for prediction‡	Nucleotide diversity (π)
	Brown-seeded breeding lines	BS61	61	11,714	5,604	0.2891

† Number of SNPs after filtering using the following criteria: SNP call rate ≥ 0.8, heterozygous rate ≤ 0.2, and minor allele frequency ≥ 0.05. YS and BS were analyzed together as a single group for SNP discovery but evaluated separately for prediction accuracy. ‡ The value 33,596 denotes SNPs shared between BMEVSU260 and the CORE378 and CORE287 datasets, where 5,604 denotes SNPs shared across all training datasets and the YS38 and BS61 test datasets.

Two historical training populations were derived from a global flax core collection: CORE378 includes both linseed and fiber morphotypes, while CORE287 contains the linseed-only subset. All accessions in the CORE populations were genotyped using whole-genome shotgun sequencing (WGS) (You et al. 2018a; He et al. 2023). Following SNP discovery and filtering, 1,699,363 high-quality SNPs were retained in CORE378 and 1,696,929 SNPs in CORE287 (Fig. S1A). These populations exhibited high nucleotide diversity (π = 0.3991–0.4007), consistent with their broad and diverse germplasm origins.

A third training population, BP296, was developed to better represent modern breeding-oriented germplasm. BP296 consisted of 296 lines, including 253 advanced breeding lines and 43 parental or germplasm accessions that have been recently released or are actively used in flax breeding programs. The 253 breeding lines were genotyped using a GBS platform, generating an average sequence depth of 2.5 Gb per line and a mean coverage of 5.16× (Table S1). The 43 parental or germplasm accessions were genotyped using WGS, and SNPs common to both datasets were retained. After harmonization, 5,604 SNPs were shared across all BP296 lines (Fig. S1C). Despite its breeding-focused composition, BP296 displayed nucleotide diversity (π = 0.3838) comparable to that observed in the CORE populations. Owing to its genetic relevance to contemporary breeding germplasm, BP296 was evaluated both as an independent training population and, in some analyses, as a test population. When BP296 was used as a test population in APP analyses, the training data consisted exclusively of the CORE287 or CORE378 populations, no data from BP296 was used in model training for these specific evaluations.

In addition to BP296, three independent test populations were used for APP. The first test population, BMEVSU260, comprised 260 inbred lines (255 recombinant inbred or doubled haploid lines plus five parental lines) derived from three biparental populations (BM, EV, and SU) with distinct genetic backgrounds [19]. These lines were genotyped using WGS, generating an average of more than 28 million PE100 reads per sample and a mean genome coverage of 11.41× (Table S1). Of these, 94.47% of reads aligned to the CDC Bethune v3.0 reference genome, yielding 43,268 high-quality SNPs (You et al. 2018b), of which 33,596 overlapped with the CORE training populations (Fig. S1B; Table 2).

The second and third test populations consisted of breeding lines selected from the Crop Development Centre (CDC; University of Saskatchewan) flax breeding program in 2023 and 2024. These included 33 yellow-seeded (YS38) and 57 brown-seeded (BS61) lines, together with their respective parent and check cultivars. These lines were evaluated in replicated yield trials conducted across three or four locations over one or two years (Table 1). Genotyping was performed using GBS, producing an average of 6.0 million PE150 reads per sample, with 97.70% of reads mapping to the CDC Bethune v3.0 reference genome (Table S1). This resulted in 11,714 high-quality SNPs, of which 5,604 were shared with the training populations (Fig. S1C; Table 2).

Due to differences in genotyping platforms, the number of SNPs shared between training and test populations varied. BMEVSU260 shared 33,596 SNPs with the CORE populations but only ~300 SNPs with GBS-based datasets, whereas YS38, BS61, and BP296 shared 5,604 SNPs, making them more directly comparable for APP analyses.

Pairwise population genetic differentiation, estimated using Weir–Cockerham FST (Weir and Cockerham 1984) (Table 3), revealed clear contrasts among populations. The CORE populations showed moderate to strong differentiation from the breeding test populations (FST = 0.2458–0.3739), reflecting their broader and more historical germplasm composition. In contrast, BP296 exhibited substantially lower differentiation from YS38 (FST = 0.0899) and BS61 (FST = 0.1323) than either CORE287 or CORE378, indicating closer genetic proximity to contemporary breeding germplasm. BMEVSU260 showed moderate differentiation from the CORE populations (FST ~0.24-0.26), consistent with its biparental origin. Collectively, these results demonstrate that BP296 occupies an intermediate genetic position between historical germplasm collections and modern breeding populations, supporting its use as a biologically-relevant training population for genomic prediction in applied flax breeding.

Table 3 Pairwise population genetic differentiation (FST) among training and test populations

Population	CORE287	CORE378	YS38	BS61
CORE378	0.2458			
YS38	0.2574	0.2712		
BS61	0.3488	0.3739	0.2036	
BP296	0.2379	0.2548	0.0899	0.1323

Cross-validation based prediction accuracy for seed yield

CV was first used to evaluate the prediction capacity of different training populations, genomic selection models, and marker representations under ideal within-population conditions. PA, measured as the Pearson correlation between predicted and observed YLD, was assessed using a five-fold CV scheme with 10 replicates (50 runs in total) for each combination of training population, model, and marker type (Fig. 1; Table S2).

Across the three training populations, CV-based PAs were generally high, with mean values typically ranging from 0.75 to 0.85 for most linear, ML, and DL models when using SNP or HAP markers (Table S2). The highest mean PA observed across all CV analyses was 0.84, achieved by RR-BLUP and BRR models using BP296 or CORE287 as the training population. In contrast, prediction accuracies based on PC representations were consistently lower and exhibited substantially higher variability across models, particularly for DL architectures (Fig. 1B; Table S2).

When comparing training populations, BP296 showed CV performance comparable to or slightly exceeding that of CORE287 and CORE378 across most models and marker types. Notably, BP296 maintained high PAs despite being composed primarily of recent breeding lines rather than broad germplasm accessions, indicating that its genetic diversity is sufficient to support effective within-population prediction. CORE287 generally outperformed CORE378 across models, whereas CORE378 exhibited a modest reduction in PA for several models, consistent with its mixed linseed and fiber flax composition.

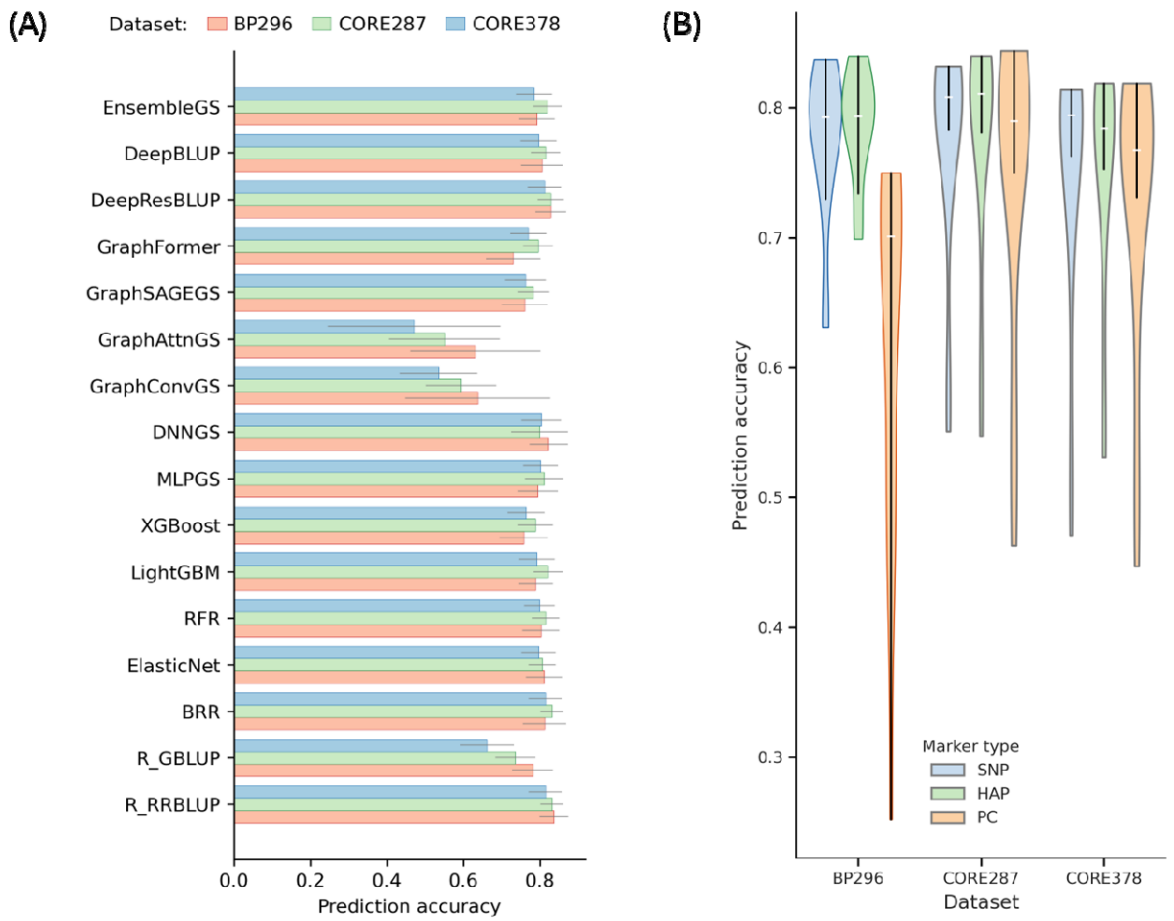

Fig. 1 Cross-validation (CV)–based prediction accuracies (PAs) for seed yield. (A) Bar chart showing the mean PA for each model using SNP markers, with error bars indicating the standard deviation. (B) Violin plots summarizing the distribution of PAs across 16 models and all three marker types. PA was evaluated using a five-fold CV scheme with 10 random training-test partitions, resulting in 50 total replicates for each training dataset.

Model-specific trends were largely consistent across training populations. Linear mixed models (RR-BLUP, GBLUP and BRR) performed robustly, achieving high and stable PAs with both SNP and HAP markers. Tree-based ML models (RFR, XGBoost and LightGBM) and shallow DL models (MLPGS and DNNGS) performed comparably, with no systematic advantage over linear baselines under CV conditions. Among graph-based DL models, GraphSAGEGS and GraphFormer consistently outperformed GraphConvGS and GraphAttnGS, although their overall performance remained slightly below that of the best linear and hybrid models. Hybrid approaches integrating RR-BLUP with deep architectures (DeepBLUP and DeepResBLUP) consistently achieved prediction accuracies comparable to, and in some cases

marginally higher than, linear mixed models, while exhibiting moderate variability across training populations.

The distribution of CV-based PAs across all 16 models further highlighted differences among marker types (Fig. 1B). SNP and HAP markers produced similarly high and stable PA distributions across training populations, whereas PC-based markers resulted in broader and lower PA distributions, particularly for BP296. These results indicate that dimensionality-reduced PC features are less effective at capturing the genetic signal underlying seed yield under CV conditions, especially when training populations consist of heterogeneous breeding germplasm.

Overall, CV-based analyses demonstrated that multiple model classes and marker representations can achieve high PA under ideal within-population conditions, and that BP296 performs at least as well as historical CORE collections as a training population. However, because CV relies on random partitions within the same population, the results primarily reflect model capacity rather than predictive robustness in real-life breeding scenarios. Consequently, APP analyses were conducted to evaluate model performance under conditions that emulate practical breeding deployment.

Across-population prediction accuracy for seed yield

Four independent test datasets—BMEVSU260, YS38, BS61, and BP296—were predicted using three marker types (SNP, HAP and PC), with models trained on the historical core collections (CORE378 and its linseed-only subset CORE287) and, where applicable, the breeding-oriented training population BP296.

Across all test datasets, APP-based PAs were generally lower and more variable than those observed under cross-validation (Figure 2, Tables S2-S4), reflecting the increased difficulty of predicting across genetically and temporally distinct populations. Nevertheless, PAs frequently exceeded 0.60 and, in some cases, reached 0.80 or higher, demonstrating the feasibility of genomic prediction for seed yield under actual breeding schemes.

Training population choice had a pronounced impact on APP performance. In particular, BP296 consistently outperformed the historical CORE collections when used as the training population for predicting contemporary breeding lines. For the YS38 test set, BP296 achieved the highest PA for most models, with maximum values reaching 0.85 with GBLUP, substantially exceeding the PA values obtained using CORE287 or CORE378 (Fig. 2B). Similar trends were

observed for BS61, where BP296 generally provided higher or comparable PAs relative to CORE-based training, despite the greater variability observed across models for this training set (Fig. 2C).

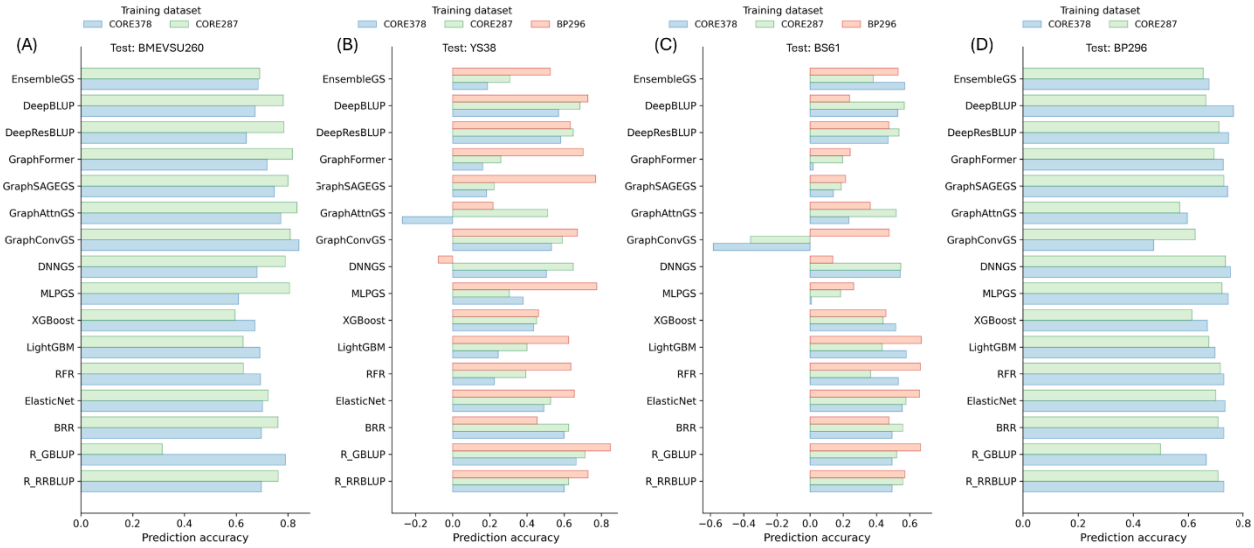

Fig. 2 Prediction accuracies (PAs) of seed yield obtained using 15 genomic selection models with SNP markers across different training populations and test datasets: (A) BMEVSU260, (B) YS38, (C) BS61, and (D) BP296.

In contrast, for the BMEVSU260 biparental population, CORE287 consistently outperformed CORE378 across nearly all models, with maximum PAs reaching 0.84 for several graph-based and hybrid models (Figs 2A and 3A). This result is consistent with the close genetic relationship between BMEVSU260 and the linseed-only CORE287 population (Table 2). BP296 was not evaluated as a training population for BMEVSU260 due to limited marker overlap resulting from differences in genotyping platforms.

When BP296 was treated as test population, CORE378 achieved slightly higher prediction accuracies (0.01–0.03) than CORE287 across models (Fig. 2D), despite marginally higher genetic differentiation between CORE378 and BP296 (FST = 0.2548) compared with CORE287 and BP296 (FST = 0.2349). This pattern likely reflects the broader allelic diversity and genetic coverage of CORE378, which includes both linseed and fiber germplasm and therefore more fully represents the genetic space occupied by BP296. In contrast, CORE287 contains only linseed accessions and captures a narrower allelic spectrum. These results indicate

that, when the test population lies largely within the diversity range of the training population, broader genetic coverage can outweigh small differences in genetic relatedness in determining PA using APP.

Across all APP analyses, no single model class consistently dominated all test datasets, and relative model rankings differed substantially from those observed under CV. Linear mixed models (RR-BLUP, GBLUP, BRR) and regularized regression models (ElasticNet) showed robust and stable performances across most scenarios, frequently ranking among the top-performing methods. Tree-based ML models (Random Forest, XGBoost, LightGBM) exhibited moderate to high PA, although their performance was more sensitive to training population choice.

DL and hybrid models displayed a wide range of performances under APP. Hybrid models integrating RR-BLUP with deep architectures (DeepBLUP and DeepResBLUP) consistently achieved high PAs across multiple test datasets, particularly when training and test populations were genetically compatible. Graph-based DL models showed strong performance for BMEVSU260, with GraphConvGS and GraphAttnGS achieving some of the highest PAs (> 0.83), but their performance was less stable for YS38 and BS61, occasionally yielding low or even negative correlations when training–test divergence was large. These results indicate that model complexity alone does not guarantee superior APP performance, and that robustness across populations remains a critical consideration.

The heritability of seed yield in the flax core collection was estimated at 0.41 (You et al. 2017), which roughly corresponds to a phenotypic selection accuracy of 0.64 ($=\sqrt{0.41}$) (Falconer and Mackay 1996). As such, the GS PA for YS and BS lines and 254 lines in BP296 exceeds the expected accuracy of phenotypic selection.

Marker type influenced APP performance, although its effect was generally secondary to training population choice. Across test datasets, SNP- and HAP-based markers consistently outperformed PC-based markers, particularly for BP296-trained models predicting YS38 and BS61. PCs frequently resulted in reduced or unstable PAs, especially for graph-based DL models. Nevertheless, the relative performance of SNPs and HAPs was broadly comparable across models, suggesting that moderate-density GBS markers are sufficient for effective APP of seed yield when appropriate training populations are used.

Taken together, APP analyses demonstrate that training population composition is a primary determinant of PA for seed yield in flax, often outweighing differences among models or marker types. The breeding-oriented population BP296 consistently provided superior or comparable prediction performance for contemporary breeding lines, whereas historical CORE collections performed best when predicting genetically related materials. These results highlight the importance of aligning training population design with breeding objectives and confirm the merit of APP-based evaluation for evaluating GS strategies intended for practical deployment.

Check-based selection decision of breeding lines

Because breeding decisions are ultimately threshold-based rather than correlation-based, we further evaluated GS performance using check-based advancement decisions. To evaluate the practical utility of GS for breeding decision-making, we examined GS-based predictions for the YS38, BS61 and BP296 test datasets using check cultivars as benchmarks for advancement decisions. In Canadian flax breeding programs, AAC Bright is the main check cultivar for yellow-seeded flax, whereas CDC Glas and CDC Rowland (advanced trials only) serve as standard checks for brown-seeded flax. For each test dataset, the training population and model combination that achieved the highest APP accuracy was used for CK-based selection analyses.

For the YS38 dataset, predictions generated using BP296 as the training population and the GBLUP model with HAPs achieved the highest PA (r = 0.84). Using AAC Bright as the benchmark check, GS accurately identified all four breeding lines that outperformed the check cultivar for observed seed yield, together with four additional promising candidates that were not selected based solely on phenotypic performance (Fig. 3B). Importantly, GS consistently excluded five lines from the 2023–2024 YS trial that underperformed the check cultivar in both observed and predicted yields (green dots in Fig. 3B). If GS-based selection had been used, the number of lines advanced to subsequent trials would have been reduced from 33 to 8, representing a 75% reduction in field-testing requirements, while retaining all superior-yielding candidates identified by phenotypic selection.

For the BS61 dataset, the highest prediction accuracy was achieved using BP296 as the training population with the LightGBM model and haplotype markers (r = 0.67). Using CDC Glas as the primary benchmark check (Fig. 3C), GS identified four of the seven lines exceeding the check based on observed yields (57%), along with one additional high-GEBV candidate. The

three GS-missed superior lines ranked third, fifth, and seventh phenotypically, indicating low risk of excluding elite candidate lines. Under this GS-assisted decision framework, only five lines would be advanced, representing a 91% reduction in trial size (from 57 to 5).

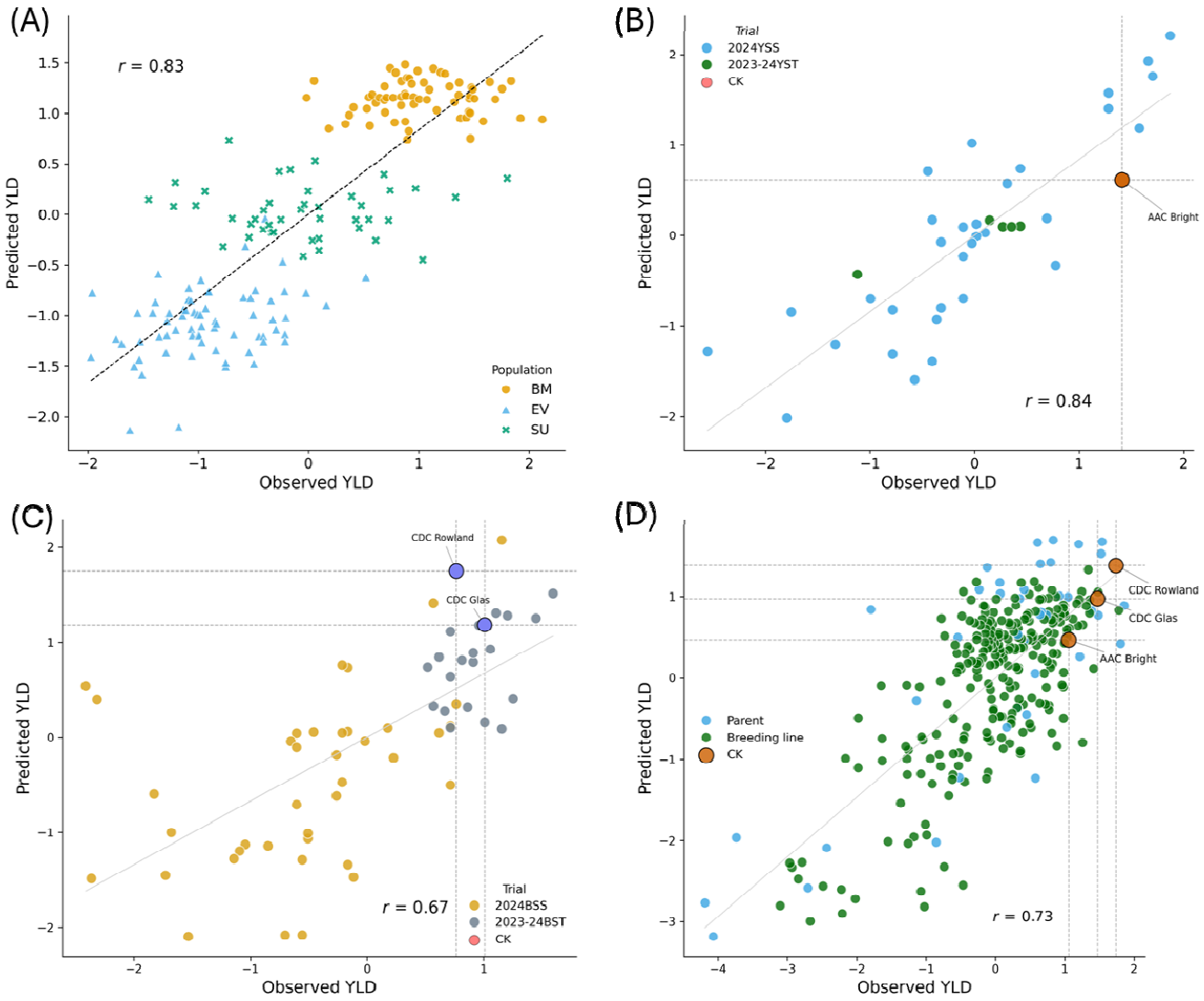

Fig. 3 Across-population prediction (APP) of seed yield (YLD) using four test datasets illustrating the ability of genomic selection to reproduce check-based advancement decisions while filtering effectively low-performing lines. (A) Scatter plot of observed versus predicted seed yield for 260 combined biparental inbred lines (BMEVSU260) using the MLPGS model with 7,871 haplotype (HAP) markers. (B) Scatter plot of observed versus predicted seed yield for the YS38 dataset using the GBLUP model with 1,945 HAP markers. (C) Scatter plot of observed versus predicted seed yield for the BS61 dataset using the LightGBM model with 1,933 HAP markers. (D) Scatter plot of observed versus predicted seed yield for the BP296 dataset using the deep learning model DNNGS with 5,604 SNP markers. “CK” denotes check cultivars. Vertical and horizontal reference lines indicate thresholds for check-based selection under genomic and

phenotypic selection. Observed and predicted see yield values were standardized to a mean of 0 and a standard deviation of 1.

For the BP296 dataset, analyses focused on the 253 new breeding lines, excluding parental and germplasm accessions. Using CORE287 as the training population, the DNNGS model with SNP markers achieved the highest PA (r = 0.73). When AAC Bright was used as the benchmark check, phenotypic selection identified 15 lines superior to the check based on predicted breeding values. Applying GS-based selection, 95/253 lines would be advanced to the next stage of yield testing, representing a 62% reduction. Among these, 13/15 lines (87%) selected based on their phenotypic performance would be retained, including the two highest-yielding lines across all replicated field trials. GS-based selection can substantially reduce population size for advanced testing while maintaining a low risk of excluding elite candidate lines.

Across all three test datasets, GS effectively reproduced check-based phenotypic advancement decisions while consistently filtering out low-performing entries. By enabling earlier selection, reducing the number of lines prior to extensive multi-environment testing, and preserving high-performing candidates, GS has the potential to enhance breeding efficiency and accelerate cultivar development in flax.

Number of markers required for prediction of breeding lines

To assess the minimum marker density required for effective genomic prediction of breeding lines, across-population prediction analyses were conducted using random subsets of SNPs for BMEVSU260 and BP296, with CORE287 as the training population. Three models representing linear, machine-learning, and hybrid approaches (R_RRBLUP, LightGBM, and DeepBLUP) were evaluated using increasing numbers of randomly sampled SNPs shared between training and test populations (Fig. 4).

Across both test populations, PA increased with marker density up to 2,000–2,500 SNPs, after which PA gains plateaued or only marginally increased. For the BMEVSU260 biparental population, R_RRBLUP and DeepBLUP reached stable prediction accuracies of approximately 0.74–0.76 once marker numbers reached or exceeded 2,500 SNPs, with little improvement observed when marker density increased to 10,000 SNPs (Fig. 4A). LightGBM showed lower overall performance and greater variability, but similarly failed to benefit from additional

markers beyond this threshold. For the BP296 breeding population, all three models exhibited a consistent upward trend up to ~2,500 SNPs, achieving prediction accuracies of approximately 0.68–0.72, followed by minimal improvement or increased variability at higher marker densities (Fig. 4B).

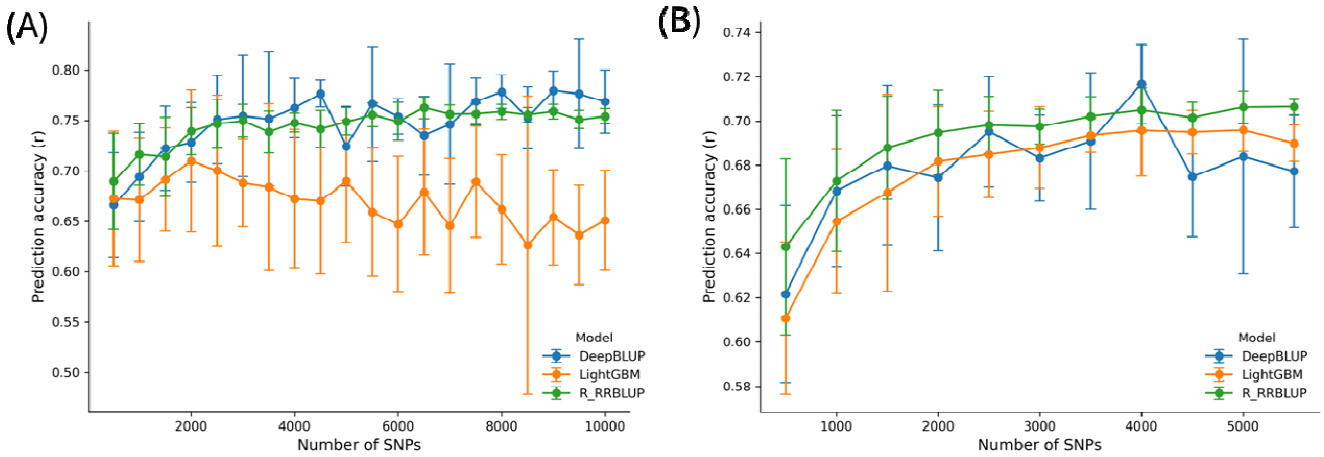

Fig. 4 Prediction accuracies (PAs) of seed yield with increasing numbers of SNP markers. (A) BMEVSU260 test dataset, (B) BP296 test dataset. The CORE287 population was used as the training set, and predictions were generated using SNP-based R_RRBLUP (green), LightGBM (orange) and DeepBLUP (blue) models. SNP marker subsets of 500 to 10,000 (in 500 increments), were randomly sampled from the SNP set common to both training and test populations (x-axis). Ten independent random subsets were generated per SNP bins, and error bars represent standard deviation among replicates.

These results indicate that approximately 2,500 genome-wide SNP markers are sufficient to capture most of the predictive signal for seed yield in flax across breeding populations under APP conditions. Increasing marker density beyond this level provided diminishing returns for prediction accuracy across all evaluated models, suggesting that moderate-density GBS marker panels are adequate for routine genomic prediction of breeding lines.

Discussion

Training population design for genomic prediction of seed yield

Training population composition emerged as a primary determinant of genomic prediction performance for seed yield in this study, particularly under APP scenarios that reflect actual breeding deployment. While historical core collections provide broad genetic diversity, our

results demonstrate that genetic relevance to contemporary breeding germplasm outweighs population size or diversity alone for predicting the performance of new breeding lines.

The breeding-oriented population BP296 consistently outperformed historical CORE collections when predicting modern yellow- and brown-seeded breeding lines (YS38 and BS61), despite its lower marker density and narrower genetic diversity than CORE287 and CORE378. In contrast, CORE collections performed best when predicting genetically related biparental materials but showed reduced accuracy when applied to elite breeding lines. These patterns indicate that broad germplasm diversity can dilute predictive signal when the training population is misaligned with breeding targets.

Notably, these contrasts were largely obscured under cross-validation but became evident under APP, highlighting the limitations of CV for evaluating deployment-ready GS strategies. Collectively, these results support the use of breeding-aligned, dynamically updated training populations as a foundation for effective genomic selection of complex traits such as seed yield.

Genomic selection models and marker types

Across all across-population prediction (APP) scenarios, classical linear mixed models remained among the most robust and consistently performing approaches for predicting seed yield in flax. Regularized linear models and Bayesian implementations showed stable performance across training–test combinations, highlighting their continued suitability for routine breeding applications. In contrast, more complex machine learning and deep learning models exhibited greater sensitivity to training population composition, with performance varying substantially across test populations. Hybrid models that integrate RR-BLUP with deep architectures achieved competitive results in several scenarios, but did not consistently outperform linear baselines, indicating that increased model complexity alone does not guarantee improved robustness under APP.

Marker types influenced prediction performance, although its effect was secondary to training population design. Across both cross-validation and APP analyses, SNP- and haplotype-based markers consistently outperformed PC markers, particularly when predicting contemporary breeding germplasm. PC markers frequently resulted in reduced or unstable prediction performance, especially for non-linear models, likely because dimensionality reduction obscures local LD patterns and additive genetic effects that are critical for yield

prediction (Weber et al. 2023; Lin et al. 2024; Crossa et al. 2017; Du et al. 2018). By contrast, SNPs and haplotypes captured genome-wide LD more directly and provided broadly comparable performance across models. Haplotype-based markers achieved prediction accuracies that were broadly comparable to those obtained with SNP markers, accounting for approximately half of the top-performing model–population combinations, while substantially reducing marker dimensionality.

Together, these results indicate that robust genomic prediction of seed yield in flax depends more strongly on the genetic relevance of the training population and the use of genome-wide marker types than on the choice of increasingly complex model architectures. Well-regularized linear models combined with moderate-density SNP or haplotype markers therefore remain reliable and practical choices for genomic selection in applied flax breeding programs.

Genotyping-by-sequencing (GBS) markers

High-density SNP arrays are widely used in crops such as wheat, maize, and soybean (Lozada et al. 2019; Polzer et al. 2025; Li et al. 2024); however, GBS remains a primary genotyping platform for GS in many breeding programs due to its flexibility, scalability, and cost-effectiveness. In actual GS pipelines, training populations are often genotyped with high marker density (e.g., whole-genome sequencing or deep GBS), whereas test populations are genotyped at lower depth to reduce costs. This imbalance reduces the number of high-quality SNPs shared between training and test sets, which can directly influence APP performance.

Consistent with this expectation, APP accuracy in our study was strongly associated with the number and genomic distribution of SNPs shared between training and test populations, and not strictly with the marker density *per se*. The combined biparental population BMEVSU260, which shared more than 35,000 SNPs with the training population, achieved PAs of up to r ~ 0.83, comparable to within-population cross-validation results. In contrast, breeding populations genotyped using standard GBS protocols yielded approximately 12,000 SNPs, of which ~7,000 were shared with the training set, resulting in prediction accuracies up to r ~ 0.79 across models and marker types. These results indicate that once shared markers adequately capture genome-wide linkage disequilibrium, additional markers provide diminishing returns for PA.

This conclusion was further supported by marker subsampling simulations. Across two representative test populations (BMEVSU260 and BP296) and three model classes, PA increased rapidly with marker number up to approximately 2,000–2,500 SNPs, beyond which gains plateaued (Fig. 4). Below this threshold, accuracies declined markedly, particularly for breeding populations with greater genetic heterogeneity. Together, these results suggest that moderate-density GBS panels (~2,500–3,000 well-distributed SNPs) are sufficient to achieve stable and reliable PAs for seed yield in flax under APP conditions.

From a breeding perspective, these findings support the use of cost-efficient GBS strategies that prioritize an adequate number of shared markers over a larger number of markers. Such designs enable reliable genomic prediction while minimizing genotyping costs and computation time, thereby facilitating routine GS implementation for early-stage selection and optimizing resource allocation in flax breeding programs.

Genomic selection integration into flax breeding program

APP remains one of the major challenges for implementing GS in practical breeding programs. While within-population CV often yields high prediction accuracies (You et al. 2022; Lan et al. 2020; He et al. 2019), its utility for predicting new breeding lines is limited. CV-based accuracies can be inflated due to shared genetic structure, close relatedness between training and test sets, and the ability of dense marker coverage to capture existing LD patterns. As a result, CV performance frequently overestimates the robustness of GS models which does not translate when applied to novel breeding germplasm.

Consistent with this limitation, relatively few studies have reported APP performance, and those that have generally observed lower PAs. For example, in wheat, PAs of grain yield ranged from -0.14 to 0.43 when two biparental populations were predicted using a diverse panel of soft red winter wheat accessions as the training set (Lozada et al. 2019). In CIMMYT's elite yield nurseries, PAs declined from 0.35–0.44 within nurseries to 0.05–0.15 across nurseries, largely reflecting the genetic relatedness among lines (Juliana et al. 2018). Nevertheless, even under these conditions, GS at moderate selection intensity successfully identified a large proportion of top-performing lines (Juliana et al. 2018). Further improvements in APP have been achieved by expanding training populations across years and environments, with prediction accuracies increasing from 0.11 to 0.23 when five years of historical data were used for training

(Vitale et al. 2025). Similar moderate APP accuracies have been reported for yield and quality traits in soybean and maize (Polzer et al. 2025; Dzievit et al. 2021).

Against this backdrop, the APP results obtained in this study demonstrate the operational potential of GS for flax breeding. When training and test populations were genetically aligned, GS achieved prediction accuracies well within—or exceeding—the range considered useful for breeding decisions. For example, predicting the biparental population BMEVSU260 using CORE287 as the training population yielded accuracies as high as r = 0.83 with haplotype-based MLPGS (Fig. 3A). Similarly, using BP296 as the training population to predict YS38, haplotype-based GBLUP achieved r = 0.84 (Fig. 3B). For BS61, prediction accuracy reached r = 0.67 using HAP markers and LightGBM (Fig. 3C), while prediction of BP296 using CORE287, SNP markers and DNNGS achieved r = 0.73. These values exceed the commonly cited thresholds of practical utility for complex traits in cereals and legumes (r ~ 0.3–0.6) (Crossa et al. 2017; Juliana et al. 2020). Notably, the relatively high APP accuracies observed here contrast with many previous reports for yield traits and appear to be driven primarily by training population alignment rather than by increased model complexity.

Importantly, GS in this study was not evaluated solely as a prediction tool, but as a decision-support mechanism. GS-based selection consistently reproduced check-based advancement decisions while substantially reducing the number of lines requiring multi-location and multi-year field evaluation. By acting as an early-stage filter, GS enabled reductions of 60–90% in trial population size without increased risk of discarding elite candidates, thereby lowering costs and accelerating breeding cycles.

Based on these results, three practical principles emerge for integrating GS into flax breeding programs: (1) Training populations should be genetically and environmentally aligned with breeding targets, rather than relying exclusively on broad germplasm collections; (2) Training sets should be regularly updated with new breeding germplasm to maintain relevance and mitigate genetic drift; and (3) GS should be implemented as a gatekeeper in early selection stages, enabling substantial reductions in trial size (~60–90%) once prediction accuracies reach r ~ 0.6–0.8, thereby focusing the program’s costly phenotyping resources on the most promising candidates.

Together, these findings indicate that GS for seed yield in flax is sufficiently robust for routine integration into breeding pipelines, particularly when used to complement—rather than replace—phenotypic evaluation.

Cost of genomic selection

In flax breeding, field evaluation for seed yield represents a major financial and logistical investment. In the CDC flax breeding program, the cost per field plot—from seeding through harvest and data collection—is approximately $75 CAD, resulting in $150 per line for two replications at a single location in preliminary yield trials. Additional routine evaluations further increase costs, including oil profile determination by gas chromatography (FAME; ~$10 per sample) and disease resistance screening (e.g., Fusarium wilt; ~$12.50 per row). With two replications per line, the total cost per line is therefore approximately $195 CAD, and advancing 300 lines to preliminary trials requires an annual investment of approximately $58,500 CAD.

In contrast, the per-sample cost of genotyping-by-sequencing (GBS) is substantially lower. DNA preparation and library construction together cost approximately $17.10 per sample, while sequencing costs scale linearly with read depth and sample multiplexing on the NovaSeq platform. Across scenarios ranging from 192 to 768 samples per run, the per-sample sequencing costs remain linear. For example, using 768 samples on the NovaSeq 3200M PE150 platform, the per-sample cost can be approximated as $y = 1.3594x + 17.121$, where y is the cost per sample (CAD) and x is the number of million PE150 reads. Under this model, a sequencing depth of 6M PE150 reads per sample—corresponding to approximately 6,000 common SNPs—results in a per-sample cost of ~$25.27 CAD.

Importantly, results from marker number simulations (Fig. 4) indicate that ~2,500 genome-wide SNPs are sufficient to achieve stable and near-maximal PA for seed yield across breeding populations. The ~6,000 SNPs generated at this sequencing depth therefore exceed the minimum marker requirement, providing a conservative and robust genotyping strategy. Under this scenario, the total GBS cost for 300 lines is approximately $7,581 CAD. Based on check-based selection analyses, reducing the number of lines advanced to preliminary and subsequent yield trials by 61–91% would decrease the initial pool of 300 lines to 27–117 lines. Under this strategy, the combined cost of genotyping, yield trials, disease screening, and FAME analysis is

estimated at $12,846–$30,396 CAD, corresponding to 48–78% cost savings relative to phenotypic selection alone.

Overall, these results demonstrate that GS can substantially reduce the cost and scale of field evaluations conducted at early breeding stages (e.g., preliminary yield trials following initial line development), while retaining high-performing candidate lines. In addition to direct cost savings, earlier GS-based selection enables advancement of selected lines to contra-season nurseries (e.g., winter nurseries in New Zealand), further reducing breeding cycle time. By enabling earlier selection decisions, smaller field trials, and more efficient resource allocation, GS offers a practical and cost-effective strategy for accelerating flax breeding without increasing the risk of discarding elite germplasm.

While the cost analysis presented here reflects the initial integration of GS, maintaining PA over successive breeding cycles requires periodic updating of the training population with newly phenotyped breeding lines to mitigate model decay. This model upkeep incurs recurrent costs associated with genotyping and limited phenotyping; however, these activities are typically integrated into routine breeding operations and represent a small fraction of the extensive multi-location field-testing costs that are reduced by GS. Although ongoing investment in training population renewal is necessary, the net economic advantage of GS-assisted breeding remains strongly positive.

Conclusion

This study demonstrates that genomic selection for seed yield in flax can be effectively deployed under realistic breeding scenarios when training populations are genetically aligned with breeding targets. Across independent test populations, training population composition emerged as the primary determinant of prediction accuracy, often outweighing differences among models or marker representations. Genomic selection consistently reproduced check-based advancement decisions while substantially reducing the number of lines requiring extensive field evaluation, indicating its value as an early-stage decision-support tool rather than a replacement for phenotypic selection. Moderate-density GBS marker panels were sufficient to achieve stable prediction accuracy, enabling cost-efficient implementation. Together, these results provide practical guidance for integrating genomic selection into flax breeding programs and highlight the importance of breeding-oriented training population design for achieving robust prediction of

complex traits. When applied as a complement to phenotypic evaluation, genomic selection offers a viable pathway to improve breeding efficiency, reduce costs, and accelerate cultivar development.

Supplementary Information

The online version contains supplementary material available at xxxx.

Acknowledgements

We thank Dr. Liqian He for reviewing the draft manuscript and for providing valuable comments and suggestions, and Tara Edwards for DNA preparation.

Author contribution statement

FY contributed to conceptualization, funding acquisition, investigation, methodology, data analysis, and original draft preparation. SC contributed to conceptualization, funding acquisition, DNA preparation for SNP genotyping, data curation, and manuscript review and editing. BT contributed to conceptualization, funding acquisition, data analysis, and manuscript review and editing. JD contributed to software development. CZ, JD, and PL contributed to formal analysis, visualization, data curation, and manuscript review. KJ, MH and BT performed field trials for breeding lines and manuscript review. All authors read and approved the final manuscript.

Funding

This Sustainable Canadian Agricultural Partnership AgriScience Cluster (SCAP-ASC) project Activity 5A (SCAP-ACS-05) was funded by Agriculture and Agri-Food Canada (AAFC) and the Diverse Field Crop Cluster (DFCC) managed by Ag-West Bio Inc. The core collection research originated from the Total Utilization Flax GENomics (TUFGEN) project funded by Genome Canada and affiliated stakeholders.

Declarations

Conflict of interest

The authors declare they have no conflict of interest.

References

Barrett, J.C., B. Fry, J. Maller, and M.J. Daly, 2005 Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21 (2):263-265.

Bates, D., M. Mächler, B. Bolker, and S. Walker, 2015 Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* 67:1–48.

Beyene, Y., M. Gowda, M. Olsen, K.R. Robbins, P. Perez-Rodriguez *et al.*, 2019 Empirical comparison of tropical maize hybrids selected through genomic and phenotypic selections. *Front Plant Sci* 10:1502.

Browning, B.L., X. Tian, Y. Zhou, and S.R. Browning, 2021 Fast two-stage phasing of large-scale sequence data. *Am J Hum Genet* 108 (10):1880-1890.

Cloutier, S., T. Edwards, C. Zheng, H.M. Booker, T. Islam *et al.*, 2024 Fine-mapping of a major locus for Fusarium wilt resistance in flax (*Linum usitatissimum* L.). *Theor Appl Genet* 137 (1):27.

Crossa, J., P. Perez-Rodriguez, J. Cuevas, O. Montesinos-Lopez, D. Jarquin *et al.*, 2017 Genomic selection in plant breeding: methods, models, and perspectives. *Trends Plant Sci* 22 (11):961-975.

Diederichsen, A., P.M. Kusters, D. Kessler, Z. Bainas, and R.K. Gugel, 2013 Assembling a core collection from the flax world collection maintained by Plant Gene Resources of Canada. *Genet Resour Crop Evol* 60:1479–1485.

Du, C., J. Wei, S. Wang, and Z. Jia, 2018 Genomic selection using principal component regression. *Heredity (Edinb)* 121 (1):12-23.

Duk, M., A. Kanapin, T. Rozhmina, M. Bankin, S. Surkova *et al.*, 2021 The genetic landscape of fiber flax. *Front Plant Sci* 12:764612.

Dzievit, M.J., T. Guo, X. Li, and J. Yu, 2021 Comprehensive analytical and empirical evaluation of genomic prediction across diverse accessions in maize. *Plant Genome* 14 (3):e20160.

Elshire, R.J., J.C. Glaubitz, Q. Sun, J.A. Poland, K. Kawamoto *et al.*, 2011 A robust, simple genotyping-by-sequencing (GBS) approach for high diversity species. *PLoS One* 6 (5):e19379.

Falconer, D.S., and T.F.C. Mackay, 1996 *Introduction to Quantitative Genetics*. Longman.

Gabriel, S.B., S.F. Schaffner, H. Nguyen, J.M. Moore, J. Roy *et al.*, 2002 The structure of haplotype blocks in the human genome. *Science* 296 (5576):2225-2229.

He, L., Y. Sui, Y. Che, H. Wang, K.Y. Rashid *et al.*, 2023 Genome-wide association studies using multi-models and multi-SNP datasets provide new insights into pasmo resistance in flax. *Front Plant Sci* 14:1229457.

He, L., J. Xiao, K.Y. Rashid, G. Jia, P. Li *et al.*, 2019 Evaluation of genomic prediction for pasmo resistance in flax. *Int J Mol Sci* 20 (2):359.

He, L., J. Xiao, K.Y. Rashid, Z. Yao, P. Li *et al.*, 2018 Genome-wide association studies for pasmo resistance in flax (*Linum usitatissimum* L.). *Front Plant Sci* 9:1982.

He, S., J.C. Reif, and Y. Xu, 2017 RTM-GWAS: A restricted two-stage multi-locus genome-wide association analysis method for dissecting complex traits. *Hereditas* 154:8.

Hoque, A., J.V. Anderson, and M. Rahman, 2024 Genomic prediction for agronomic traits in a diverse Flax (*Linum usitatissimum* L.) germplasm collection. *Sci Rep* 14 (1):3196.

Juliana, P., R.P. Singh, H.J. Braun, J. Huerta-Espino, L. Crespo-Herrera *et al.*, 2020 Genomic selection for grain yield in the CIMMYT wheat breeding program-status and perspectives. *Front Plant Sci* 11:564183.

Juliana, P., R.P. Singh, J. Poland, S. Mondal, J. Crossa *et al.*, 2018 Prospects and challenges of applied genomic selection-A new paradigm in breeding for grain yield in bread wheat. *Plant Genome* 11 (3):180017.

Khan, K., H. Shazadee, S. Cloutie, and F.M. You, 2023 Genomics assisted breeding strategy in Flax, pp. 253-272 in *The Flax Genome, Compendium of Plant Genomes*, edited by F.M. You and B. Fofana.

Kumar, R., S.P. Das, B.U. Choudhury, A. Kumar, N.R. Prakash *et al.*, 2024 Advances in genomic tools for plant breeding: harnessing DNA molecular markers, genomic selection, and genome editing. *Biol Res* 57 (1):80.

Kumar, S., F.M. You, and S. Cloutier, 2012 Genome wide SNP discovery in flax through next generation sequencing of reduced representation libraries. *BMC Genomics* 13:684.

Lan, S., C. Zheng, K. Hauck, M. McCausland, S.D. Duguid *et al.*, 2020 Genomic prediction accuracy of seven breeding selection traits improved by QTL identification in flax. *Int J Mol Sci* 21 (5):1577.

Li, C., Q. Yang, B. Liu, X. Shi, Z. Liu *et al.*, 2024 Ability of genomic prediction to bi-parent-derived breeding population using public data for soybean oil and protein content. *Plants (Basel)* 13 (9):1260.

Li, H., and R. Durbin, 2009 Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25 (14):1754-1760.

Li, H., B. Handsaker, A. Wysoker, T. Fennell, J. Ruan *et al.*, 2009 The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25 (16):2078-2079.

Lin, C.S., and G. Poushinsky, 1985 A modified augmented design (type 2) for rectangular plots. *Can J Plant Sci* 65:743–749.

Lin, Y.C., M. Mayer, D. Valle Torres, T. Pook, A.C. Holker *et al.*, 2024 Genomic prediction within and across maize landrace derived populations using haplotypes. *Front Plant Sci* 15:1351466.

Lozada, D.N., R.E. Mason, J.M. Sarinelli, and G. Brown-Guedira, 2019 Accuracy of genomic selection for grain yield and agronomic traits in soft red winter wheat. *BMC Genet* 20 (1):82.

Meuwissen, T.H., B.J. Hayes, and M.E. Goddard, 2001 Prediction of total genetic value using genome-wide dense marker maps. *Genetics* 157 (4):1819-1829.

Olatoye, M.O., L.V. Clark, J. Wang, X. Yang, T. Yamada *et al.*, 2019 Evaluation of genomic selection and marker-assisted selection in Miscanthus and energycane. *Mol Breed* 39 (12):171.

Piepho, H.-P., J. Möhring, A.E. Melchinger, and A. Büchse, 2008 BLUP for phenotypic selection in plant breeding and variety testing. *Euphytica* 161:209-228.

Piepho, H.P., J. Mohring, T. Schulz-Streeck, and J.O. Ogutu, 2012 A stage-wise approach for the analysis of multi-environment trials. *Biom J* 54 (6):844-860.

Polzer, C., H.J. Auinger, M. Teran-Pineda, A.C. Holker, M. Mayer *et al.*, 2025 Rapid cycling genomic selection in maize landraces. *Theor Appl Genet* 138 (4):75.

Povkhova, L.V., N.V. Melnikova, T.A. Rozhmina, R.O. Novakovskiy, E.N. Pushkova *et al.*, 2021 Genes associated with the flax plant yype (oil or fiber) identified based on genome and transcriptome sequencing data. *Plants (Basel)* 10 (12):2016.

Praczyk, M., and K. Wielgusz, 2021 Agronomic assessment of fibrous flax and linseed advanced breeding lines as potential new varieties. *Agronomy* 11 (10):1917.

Soto-Cerda, B.J., A. Diederichsen, R. Ragupathy, and S. Cloutier, 2013 Genetic characterization of a core collection of flax (*Linum usitatissimum* L.) suitable for association mapping studies and evidence of divergent selection between fiber and linseed types. *BMC Plant Biol* 13:78.

Vitale, P., O. Montesinos-Lopez, G. Gerard, G. Velu, Z. Tadesse *et al.*, 2025 Improving wheat grain yield genomic prediction accuracy using historical data. *G3 (Bethesda)* 15 (4).

Weber, S.E., M. Frisch, R.J. Snowdon, and K.P. Voss-Fels, 2023 Haplotype blocks for genomic prediction: a comparative evaluation in multiple crop datasets. *Front Plant Sci* 14:1217589.

Weir, B.S., and C.C. Cockerham, 1984 Estimating F-Statistics for the Analysis of Population Structure. *Evolution* 38 (6):1358-1370.

Yin, L., H. Zhang, Z. Tang, J. Xu, D. Yin *et al.*, 2021 rMVP: A Memory-efficient, Visualization-enhanced, and Parallel-accelerated Tool for Genome-wide Association Study. *Genom Proteom Bioinform* 19 (4):619-628.

You, F.M., K.R. Deal, J. Wang, M.T. Britton, J.N. Fass *et al.*, 2012 Genome-wide SNP discovery in walnut with an AGSNP pipeline updated for SNP discovery in allogamous organisms. *BMC Genomics* 13:354.

You, F.M., S.D. Duguid, D. Thambugala, and S. Cloutier, 2013 Statistical analysis and field evaluation of the type 2 modified augmented design (MAD) in phenotyping of flax (*Linum usitatissimum*) germplasms in multiple environments. *Austr J Crop Sci*:1789-1800.

You, F.M., G. Jia, J. Xiao, S.D. Duguid, K.Y. Rashid *et al.*, 2017 Genetic variability of 27 traits in a core collection of flax (*Linum usitatissimum* L.). *Front Plant Sci* 8:1636.

You, F.M., K.Y. Rashid, C. Zheng, N. Khan, P. Li *et al.*, 2022 Insights into the genetic architecture and genomic prediction of powdery mildew resistance in flax (*Linum usitatissimum* L.). *Int J Mol Sci* 23 (9):4960.

You, F.M., J. Xiao, P. Li, Z. Yao, G. Jia *et al.*, 2018a Genome-wide association study and selection signatures detect genomic regions associated with seed yield and oil quality in flax. *Int J Mol Sci* 19 (8):2303.

You, F.M., J. Xiao, P. Li, Z. Yao, G. Jia *et al.*, 2018b Chromosome-scale pseudomolecules refined by optical, physical and genetic maps in flax. *Plant J* 95 (2):371-384.

You, F.M., C. Zheng, J.J. Zagariah Daniel, P. Li, B. Tar'an *et al.*, 2026 MultiGS: A comprehensive and user-friendly genomic prediction platform Integrating statistical, machine learning, and deep learning models for breeders. *Crop Breed Genet Genom* 8 (1):e260004.

Supplementary Tables and Figures

Table S1. Summary statistics of Illumina paired-end (PE) reads generated for the test populations.

Population	No. of samples	Method	No. of PE reads per sample	% mapped to reference genome	Total size per sample (Mb)	Coverage (X)
Combined biparental population (BMEVSU260)	260	WGS PE100	28,155,405	94.47	5,631	11.17
Selected breeding lines and check cultivars in 2024 (BS38 + YS61)	99	GBS PE150	6,063,057	97.70	1,758	3.49
Selected cultivars and breeding lines (subset of BP296)	45	WGS PE150	33,579,835	99.33	10,083	20.00
Selected breeding lines (subset of BP296)	253	GBS PE150	8,484,494	99.82	2,545	5.05

WGS: whole-genome sequencing. Coverage (X) was calculated as the total read length divided by the genome size estimate of CDC Bethune v3.0 of 504 Mb. One PE read is equal to two single reads.

Table S2. Prediction accuracies (PAs) and their standard deviations for seed yield (YLD) under five-fold cross-validation (CV) using BP296, CORE287 and CORE378.

Model	BP296				CORE287				CORE378			
	HAP	PC	SNP	Max	HAP	PC	SNP	Max	HAP	PC	SNP	Max
R_RRBLUP	0.840 ± 0.038	0.750 ± 0.145	0.837 ± 0.038	**0.840 ± 0.038**	0.840 ± 0.029	0.834 ± 0.030	0.832 ± 0.030	**0.840 ± 0.029**	0.819 ± 0.042	0.815 ± 0.042	0.814 ± 0.043	**0.819 ± 0.042**
R_GBLUP	0.787 ± 0.052	0.463 ± 0.169	0.781 ± 0.054	**0.787 ± 0.052**	0.738 ± 0.052	0.528 ± 0.090	0.736 ± 0.052	**0.738 ± 0.052**	0.663 ± 0.070	0.624 ± 0.110	0.662 ± 0.070	**0.663 ± 0.070**
BRR	0.839 ± 0.039	0.721 ± 0.149	0.812 ± 0.056	**0.839 ± 0.039**	0.840 ± 0.029	0.834 ± 0.030	0.832 ± 0.030	**0.840 ± 0.029**	0.818 ± 0.042	0.814 ± 0.042	0.814 ± 0.043	**0.818 ± 0.042**
ElasticNet	0.814 ± 0.042	0.744 ± 0.146	0.811 ± 0.047	**0.814 ± 0.042**	0.803 ± 0.034	0.844 ± 0.028	0.806 ± 0.035	**0.844 ± 0.028**	0.789 ± 0.047	0.819 ± 0.041	0.797 ± 0.045	**0.819 ± 0.041**
RFR	0.799 ± 0.052	0.681 ± 0.246	0.803 ± 0.048	**0.803 ± 0.048**	0.812 ± 0.036	0.797 ± 0.039	0.815 ± 0.036	**0.815 ± 0.036**	0.791 ± 0.043	0.776 ± 0.044	0.799 ± 0.040	**0.799 ± 0.040**
LightGBM	0.793 ± 0.054	0.482 ± 0.161	0.789 ± 0.045	**0.793 ± 0.054**	0.810 ± 0.040	0.772 ± 0.052	0.822 ± 0.038	**0.822 ± 0.038**	0.779 ± 0.045	0.751 ± 0.049	0.792 ± 0.047	**0.792 ± 0.047**
XGBoost	0.779 ± 0.051	0.615 ± 0.243	0.758 ± 0.063	**0.779 ± 0.051**	0.781 ± 0.049	0.775 ± 0.053	0.788 ± 0.047	**0.788 ± 0.047**	0.753 ± 0.048	0.759 ± 0.043	0.764 ± 0.048	**0.764 ± 0.048**
MLPGS	0.790 ± 0.048	0.693 ± 0.181	0.794 ± 0.053	**0.794 ± 0.053**	0.825 ± 0.037	0.804 ± 0.043	0.811 ± 0.050	**0.825 ± 0.037**	0.803 ± 0.050	0.798 ± 0.044	0.802 ± 0.045	**0.803 ± 0.050**
DNNGS	0.808 ± 0.063	0.507 ± 0.347	0.823 ± 0.050	**0.823 ± 0.050**	0.815 ± 0.051	0.806 ± 0.041	0.799 ± 0.074	**0.815 ± 0.051**	0.810 ± 0.046	0.776 ± 0.083	0.804 ± 0.052	**0.810 ± 0.046**
GraphConvGS	0.703 ± 0.122	0.252 ± 0.267	0.638 ± 0.189	**0.703 ± 0.122**	0.613 ± 0.055	0.594 ± 0.144	0.594 ± 0.092	**0.613 ± 0.055**	0.557 ± 0.082	0.521 ± 0.129	0.535 ± 0.102	**0.557 ± 0.082**
GraphAttnGS	0.699 ± 0.081	0.476 ± 0.244	0.631 ± 0.170	**0.699 ± 0.081**	0.547 ± 0.153	0.463 ± 0.255	0.551 ± 0.145	**0.551 ± 0.145**	0.531 ± 0.105	0.447 ± 0.183	0.471 ± 0.226	**0.531 ± 0.105**
GraphSAGEGS	0.765 ± 0.065	0.724 ± 0.146	0.760 ± 0.060	**0.765 ± 0.065**	0.807 ± 0.039	0.750 ± 0.055	0.783 ± 0.042	**0.807 ± 0.039**	0.772 ± 0.047	0.731 ± 0.055	0.763 ± 0.053	**0.772 ± 0.047**
GraphFormer	0.734 ± 0.077	0.728 ± 0.151	0.730 ± 0.071	**0.734 ± 0.077**	0.817 ± 0.035	0.783 ± 0.039	0.796 ± 0.039	**0.817 ± 0.035**	0.780 ± 0.048	0.758 ± 0.045	0.770 ± 0.047	**0.780 ± 0.048**
DeepResBLUP	0.829 ± 0.042	0.737 ± 0.148	0.828 ± 0.040	**0.829 ± 0.042**	0.836 ± 0.032	0.813 ± 0.036	0.829 ± 0.034	**0.836 ± 0.032**	0.813 ± 0.043	0.791 ± 0.043	0.812 ± 0.044	**0.813 ± 0.043**
DeepBLUP	0.821 ± 0.050	0.726 ± 0.156	0.805 ± 0.054	**0.821 ± 0.050**	0.828 ± 0.030	0.826 ± 0.031	0.815 ± 0.038	**0.828 ± 0.030**	0.805 ± 0.044	0.803 ± 0.043	0.797 ± 0.047	**0.805 ± 0.044**
EnsembleGS	0.794 ± 0.055	0.709 ± 0.156	0.792 ± 0.047	**0.794 ± 0.055**	0.807 ± 0.039	0.771 ± 0.051	0.819 ± 0.038	**0.819 ± 0.038**	0.775 ± 0.042	0.747 ± 0.048	0.785 ± 0.047	**0.785 ± 0.047**

Table S3. Prediction accuracies (PAs) of four test populations for seed yield (YLD) under across-population prediction (APP) using **CORE287** as the training population.

Model	Marker type	Test population			
		BP296	YS38	BS61	BMEVSU260
R_RRBLUP	PC	0.707	0.647	0.543	0.772
	HAP	0.717	0.585	0.595	0.794
	SNP	0.707	0.624	0.558	0.762
R_GBLUP	PC	0.586	0.711	0.521	0.310
	HAP	0.522	0.681	0.592	0.694
	SNP	0.499	0.711	0.521	0.314
BRR	PC	0.707	0.647	0.542	0.772
	HAP	0.717	0.584	0.595	0.794
	SNP	0.707	0.624	0.558	0.761
ElasticNet	PC	0.710	0.668	0.559	0.774
	HAP	0.690	0.571	0.616	0.722
	SNP	0.699	0.526	0.578	0.723
RFR	PC	0.680	0.658	0.370	0.738
	HAP	0.717	0.470	0.574	0.681
	SNP	0.716	0.393	0.363	0.627
LightGBM	PC	0.731	0.586	0.299	0.763
	HAP	0.673	0.528	0.598	0.621
	SNP	0.674	0.400	0.434	0.626
XGBoost	PC	0.696	0.542	0.249	0.679
	HAP	0.692	0.536	0.466	0.647
	SNP	0.612	0.451	0.440	0.595
MLPGS	PC	0.691	0.352	0.251	0.781
	HAP	0.730	0.267	0.095	0.824
	SNP	0.721	0.305	0.183	0.806
DNNGS	PC	0.693	0.698	0.517	0.777
	HAP	0.709	0.590	0.546	0.808
	SNP	0.734	0.648	0.546	0.789
GraphConvGS	PC	0.496	0.656	-0.372	0.820
	HAP	0.648	0.538	-0.151	0.835
	SNP	0.624	0.589	-0.359	0.808
GraphAttnGS	PC	0.508	0.710	-0.423	0.685
	HAP	0.572	0.411	-0.108	0.788
	SNP	0.568	0.511	0.518	0.834
GraphSAGEGS	PC	0.646	0.303	-0.005	0.779
	HAP	0.719	0.128	0.284	0.829
	SNP	0.728	0.223	0.188	0.800
GraphFormer	PC	0.697	0.306	0.168	0.754
	HAP	0.718	0.034	0.171	0.822
	SNP	0.692	0.259	0.196	0.817

Model	Marker type	Test population			
		BP296	**YS38**	**BS61**	**BMEVSU260**
DeepResBLUP	PC	0.655	0.574	0.347	0.781
	HAP	0.696	0.598	0.598	0.789
	SNP	0.710	0.648	0.535	0.784
DeepBLUP	PC	0.673	0.650	0.585	0.752
	HAP	0.731	0.552	0.484	0.798
	SNP	0.663	0.684	0.567	0.781
EnsembleGS	PC	0.728	0.546	0.236	0.776
	HAP	0.702	0.450	0.553	0.742
	SNP	0.653	0.307	0.380	0.690
	PC mean	**0.663**	**0.578**	**0.274**	**0.732**
	HAP mean	**0.685**	**0.470**	**0.407**	**0.762**
	SNP mean	**0.669**	**0.494**	**0.388**	**0.720**
	PC max	**0.731**	**0.711**	**0.585**	**0.820**
	HAP max	**0.731**	**0.681**	**0.616**	**0.835**
	SNP max	**0.734**	**0.711**	**0.578**	**0.834**

Table S4. Prediction accuracies (PAs) of four test populations for seed yield (YLD) under across-population prediction (APP) using **CORE378** as the training population.

Model	Marker type	Test population			
		BP296	YS38	BS61	BMEVSU260
R_RRBLUP	PC	0.720	0.598	0.451	0.703
	HAP	0.737	0.446	0.500	0.704
	SNP	0.728	0.600	0.493	0.697
R_GBLUP	PC	0.664	0.664	0.494	0.789
	HAP	0.673	0.608	0.565	0.765
	SNP	0.664	0.664	0.494	0.790
BRR	PC	0.720	0.597	0.451	0.703
	HAP	0.737	0.446	0.500	0.704
	SNP	0.728	0.599	0.493	0.697
ElasticNet	PC	0.726	0.626	0.444	0.749
	HAP	0.737	0.485	0.588	0.708
	SNP	0.732	0.490	0.555	0.701
RFR	PC	0.740	0.554	0.435	0.612
	HAP	0.716	0.247	0.553	0.673
	SNP	0.728	0.224	0.531	0.694
LightGBM	PC	0.717	0.595	0.500	0.652
	HAP	0.716	0.231	0.535	0.673
	SNP	0.696	0.245	0.579	0.691
XGBoost	PC	0.700	0.202	0.438	0.686
	HAP	0.675	0.214	0.553	0.586
	SNP	0.668	0.435	0.516	0.672
MLPGS	PC	0.735	0.320	0.267	0.645
	HAP	0.732	0.271	0.181	0.596
	SNP	0.744	0.380	0.009	0.609
DNNGS	PC	0.707	0.623	0.572	0.715
	HAP	0.767	0.515	0.481	0.758
	SNP	0.752	0.504	0.544	0.680
GraphConvGS	PC	0.533	0.289	-0.655	0.658
	HAP	0.422	0.528	-0.350	0.824
	SNP	0.473	0.531	-0.583	0.842
GraphAttnGS	PC	0.587	0.245	-0.268	0.662
	HAP	0.561	0.005	-0.276	0.833
	SNP	0.596	-0.271	0.233	0.772
GraphSAGEGS	PC	0.685	0.338	0.243	0.698
	HAP	0.742	0.091	0.145	0.743
	SNP	0.742	0.181	0.140	0.747
GraphFormer	PC	0.687	0.325	0.304	0.406
	HAP	0.735	0.118	0.047	0.735
	SNP	0.726	0.161	0.018	0.719

Model	Marker type	Test population			
		BP296	**YS38**	**BS61**	**BMEVSU260**
DeepResBLUP	PC	0.731	0.565	0.525	0.653
	HAP	0.713	0.446	0.375	0.713
	SNP	0.745	0.580	0.470	0.639
DeepBLUP	PC	0.665	0.552	0.436	0.696
	HAP	0.737	0.407	0.514	0.687
	SNP	0.763	0.570	0.528	0.673
EnsembleGS	PC	0.711	0.575	0.496	0.637
	HAP	0.706	0.190	0.527	0.650
	SNP	0.675	0.187	0.570	0.684
	PC mean	**0.689**	**0.479**	**0.321**	**0.667**
	HAP mean	**0.694**	**0.328**	**0.340**	**0.710**
	SNP mean	**0.698**	**0.380**	**0.349**	**0.707**
	PC max	**0.740**	**0.664**	**0.572**	**0.789**
	HAP max	**0.767**	**0.608**	**0.588**	**0.833**
	SNP max	**0.763**	**0.664**	**0.579**	**0.842**

Table S5. Prediction accuracies (PAs) of two test populations for seed yield (YLD) under across-population prediction (APP) using **BP296** as the training population.

Model	Marker type	YS38	BS61
R_RRBLUP	PC	0.726	0.570
	HAP	0.731	0.508
	SNP	0.728	0.571
R_GBLUP	PC	0.439	0.503
	HAP	0.841	0.652
	SNP	0.848	0.665
BRR	PC	0.725	0.568
	HAP	0.730	0.504
	SNP	0.453	0.475
ElasticNet	PC	0.717	0.610
	HAP	0.674	0.666
	SNP	0.654	0.659
RFR	PC	0.424	0.125
	HAP	0.555	0.564
	SNP	0.635	0.664
LightGBM	PC	0.595	0.076
	HAP	0.755	0.561
	SNP	0.623	0.670
XGBoost	PC	0.513	0.290
	HAP	0.383	0.450
	SNP	0.462	0.457
MLPGS	PC	0.745	0.643
	HAP	0.711	0.153
	SNP	0.775	0.262
DNNGS	PC	0.740	0.588
	HAP	0.273	0.668
	SNP	-0.076	0.136
GraphConvGS	PC	0.669	0.379
	HAP	0.688	-0.373
	SNP	0.670	0.477
GraphAttnGS	PC	-0.237	0.511
	HAP	-0.560	0.546
	SNP	0.217	0.362
GraphSAGEGS	PC	0.720	0.589
	HAP	0.726	0.272
	SNP	0.769	0.214
GraphFormer	PC	0.729	0.319
	HAP	0.774	0.380
	SNP	0.701	0.242
DeepResBLUP	PC	0.798	0.539

Model	Marker type	YS38	BS61
DeepBLUP	HAP	0.614	0.386
	SNP	0.633	0.474
	PC	0.719	0.574
EnsembleGS	HAP	0.674	0.340
	SNP	0.727	0.237
	PC	0.720	0.453
	HAP	0.790	0.500
	SNP	0.525	0.530
	PC mean	**0.609**	**0.459**
	HAP mean	**0.585**	**0.424**
	SNP mean	**0.584**	**0.443**
	PC max	**0.798**	**0.643**
	HAP max	**0.841**	**0.668**
	SNP max	**0.848**	**0.670**

Figure S1. SNP density along chromosomes (A) in the core collection of 378 accessions (CORE378) comprising ~1.7M SNPs, (B) 33,596 SNPs shared between CORE378 and the 260 test lines from the combined biparental population BMEVSU260, and (C) 5,604 SNPs shared by CORE378, CORE287, YS38, BS61 and BP296. Plots were generated with a 100-Kb window using the CMplot R package (Yin et al. 2021).