Sex-Dependent Effects of Chronic Stress During Adolescence on Cognitive Bias and Functional Connectome in Young Adult Rats

1 Twain Dai[1,2*], Liz Jaeschke-Angi[1,2], Marissa Penrose-Menz[1,2], Tim Rosenow[3],

Jennifer Rodger[1,2]

[1]School of Biological Sciences, The University of Western Australia, Perth, WA, Australia

[2]Perron Institute for Neurological and Translational Sciences, The University of Western Australia, Perth, WA, Australia

[3]Centre for Microscopy, Characterisation & Analysis, Research Infrastructure Centres, The University of Western Australia, 6009 Western Australia, Australia

* Correspondence:

Twain Dai

twain.dai@research.uwa.edu.au

Six keywords: Resting-State Functional MRI, Chronic Restraint Stress, Judgement Bias, Open Field Test, Sex Differences

Word Counts: 4999

Abstract

Negative cognitive biases in depression are more pronounced in females than in males. This sex difference emerges during adolescence, a sensitive developmental stage when chronic stress exposure increases the risk of depression in adulthood. Neuroimaging studies in adult depression suggest that sex differences in negative bias are associated with aberrant functional connectivity. However, the neurobiology linking adolescent stress to sex-specific cognitive bias and resting-state network reorganization in adults remains poorly understood. The study aimed to investigate the longitudinal effects of chronic restraint stress (CRS) during adolescence on cognitive bias and functional connectome in emerging adulthood. 28 Wistar rats (sex-balanced; aged five weeks on arrival) were trained on a judgment bias task with distinct tactile cues signaling differential rewards. Bias was quantified from responses to ambiguous probe trials. Following training, animals were randomly and equally assigned to CRS or control groups (sex-balanced). Offline resting-state functional MRI scans were conducted at adolescent baseline (pre-CRS) and again in adulthood (post-CRS), followed by probe trials to assess neural and behavioral changes. Behaviors were analyzed using linear mixed-effects models, and network connectivity was assessed using Network-Based R-Statistics. Both behavioral and network analyses identified sex-dependent differences in change from baseline to post-CRS. Specifically, females showed a greater tendency toward negative bias than males. Network analysis suggested female-specific reorganization within a subnetwork, spanning brainstem, limbic, striatal, insula, parahippocampal, orbitofrontal, and retrosplenial regions. These results highlight the translational importance of considering sex and distributed network organization when modelling how adolescent stress may shape later vulnerability to depression.

Introduction

Adolescence is a critical developmental period during which the brain undergoes profound structural and functional refinements, particularly cortical-subcortical circuits that govern stress regulation and cognitive processing (1, 2). Recent large-scale connectomics studies reveal that different neural circuits undergo heterochronic refinement during the transition to adulthood (3, 4). While primary sensory-motor circuits reach functional stability in early childhood, higher-order affective and cognitive networks experience ongoing refinement throughout adolescence. This developmental mismatch highlights adolescence as a pivotal turning point where functional organization of the brain is uniquely sensitive to environmental adversities (5).

Chronic stress during adolescence is known to derail these developmental trajectories by disrupting the normative refinement of cortical-subcortical circuits, significantly increasing the risk of depression in adulthood (6). The resulting psychiatric vulnerability is behaviorally characterized by negative cognitive biases, a systematic tendency to prioritize the processing of aversive information over neutral or positive stimuli (7). These biases are not merely symptoms of affective dysfunction but are considered core factors that predict the onset and chronicity of depression (8, 9). This predictive relationship is notably stronger in females. Longitudinal behavioral evidence demonstrates that the interaction between negative cognitive biases and stress more robustly affects depression trajectories in females than in males during adolescence (10).

The sex difference in negative cognitive biases is mirrored by distinct neural signatures. Specifically, adolescent females with depression exhibit unique recruitment of the posterior cingulate and supramarginal gyrus when faced with negative distractors, a response absent in their male counterparts (11). Furthermore, the neural correlates of cognitive bias are divergent during memory processing: while adolescent females with depression exhibit amygdala hyperactivity during negative recall, male counterparts are characterized by reduced hippocampal engagement during positive recall (12). Although human neuroimaging studies in adult depression suggest that sex differences in negative cognitive bias are associated with aberrant functional connectivity, the neurobiology linking adolescent chronic stress to sex-dependent cognitive bias and resting-state network reorganization remains poorly understood. Gaining a deeper understanding of these mechanisms could inform novel, targeted strategies for preventing adult depression.

Although animal models cannot fully capture the heterogeneity and social complexity of adolescent adversity in humans, they offer unique advantages for examining the neurobiology of chronic stress under highly controlled conditions (13, 14). In rodents, chronic stress during

adolescence has been shown to induce detrimental effects on affective behaviors in adulthood

(15, 16). Similar to humans, a key manifestation is a shift toward negative bias, which can be

objectively quantified using translational paradigms such as the judgment bias task (JBT). By

measuring how animals interpret ambiguous cues to assess levels of optimism and pessimism,

the JBT has demonstrated that adolescent stress induces a negative bias, mirroring that in

human depression (17, 18). However, most preclinical research continues to prioritize males,

leaving the neural mechanisms driving female-specific cognitive biases following chronic stress

largely unexplored (19, 20). This knowledge gap is further magnified by a lack of longitudinal

data mapping the sex-specific developmental trajectory of neural network as it unfolds from

adolescent stress exposure through to adulthood (15).

Therefore, this study employed a longitudinal design to examine cognitive bias and whole-brain

resting-state functional architecture within the same cohort of males and females, before and

after adolescent chronic stress. The study aimed to determine whether this stress exposure was

associated with sex-dependent alterations in cognitive bias and resting-state network

organization in emerging adulthood.

Methods

Ethics Statement

Ethical approvals for the animal experiment (2023/ET000552) were obtained from the animal ethics committee of the University of Western Australia (UWA).

Animals and Study Design

28 Wistar rats (14 females/14 males; aged 5 weeks and weighing 110 ~ 200 g on arrival) were sourced from the Ozgene Animal Resources Centre, Western Australia. All rats were housed under a reverse 12-hour light-dark cycle with ad libitum food and water, in a temperature-controlled room located at UWA's Animal Care Unit (Nedlands, WA). Rats aged 55 ~ 70 days were categorized as adolescents, whereas those aged 70 ~ 150 days were considered emerging adults (21). While the oestrous cycle was not explicitly tracked, the longitudinal nature of behavioral task and the focus on resting-state connectome reorganization likely mitigated the influence of acute hormonal fluctuations on primary outcomes. All animals (housed in pairs) were acclimatized to the new environment and handled for nine days after their arrival and prior to experiments.

Figure 1. Study Design and Timeline. All animals were habituated to the new environment for nine days after their arrival and prior to experiments. All rats were trained on JBT, for up to nine days. Behavioral tests and MRI scan were conducted before and after 14 consecutive days of CRS. Investigators were not blinded to group allocation during CRS administration or behavioral testing, because knowledge of group assignment was required to deliver the appropriate procedures.

Animals were randomly and equally assigned to two sex-balanced groups: control (7 females/7 males) and CRS groups (7 females/7 males). On Day 9 after their arrival, animals began single-housed and mild food restrictions overnight, with food pellets removed at 4 pm. From Day 10 onward, animals were trained on the JBT (*Figure 1*). Animals remained under mild food restrictions, with two hours of free access to food each day until pre-probe was completed.

Animals were excluded from further experiments if they did not successfully complete JBT

training within nine days. Once an animal was verified to have learned the task through the pre-

probe test, it underwent a pre-OFT, which habituates rats to the testing arena. Following the

pre-OFT, animals received no intervention until Day 21. Baseline MRI scan, OFT and JBT probe

for all animals were conducted on Day 21, 22 and 23, respectively. From Day 24 onwards,

animals in the CRS group underwent CRS for 14 consecutive days. The second MRI scan, OFT

and JBT probe for all rats were carried out on Day 38, 39, and 40, respectively. For the baseline

and post-CRS probes, food was removed overnight starting at 4 pm on Day 22 and 39,

respectively.

Judgment Bias Task

Task Design

The JBT in the present study was Go/Go task and adapted from Brydges and Hall (22). The

task required animal's responses to different tactile stimuli, olfactory and spatial cues, which

were associated with different reward values. The task involved two reward values (large and

small), two distinct tactile stimuli (coarse and fine), two distinct olfactory cues (cinnamon and

coriander) and two distinct spatial cues (left and right), resulting in eight task combinations

(Supplementary Table 1). The assignment of task combinations was randomized equally across

animals in each sex group and remained consistent for each animal throughout the experiment.

The olfactory and location cues were employed to accelerate memory formation in learning the

association between tactile stimuli and reward values.

Apparatus

The JBT apparatus (*Figure 2A*) was custom-built from acrylic in tint gray (The Plastic People,

Osborne Park, WA). It consisted of three compartments: one start compartment (50 cm length ×

50 cm width × 60 cm height) and two reward compartments (30 cm length × 15 cm width × 60

cm height). One removable barrier separated the start compartment from the reward

compartments. A sandpaper (tactile stimuli) was attached to the center of the start compartment

using Velcro tapes when required. One ceramic foraging bowl containing cheerios (rewards)

was placed at the end of each reward compartment. One small muslin bag containing coriander

or cinnamon was hung on each sidewall of each reward compartment.

Procedures and Bias Measures

The JBT consisted of four phases: spatial and olfactory learning, tactile learning, training, and

probe phases (*Figure 2*).

During the spatial and olfactory learning phase, animals were trained to learn the assigned

association among reward values, spatial and olfactory cues. Different amounts of cheerios

(rewards) were placed in two reward compartments. Animals were free to retrieve rewards from

both compartments. This phase was considered complete when animals retrieved rewards from

each compartment 5 times. Animals then proceeded to tactile learning the following day.

During the tactile learning phase, animals were further trained to learn the assigned association

between reward values and tactile stimuli, in addition to the spatial and olfactory cues. Animals

had free access to both reward compartments for a total of 30 trials. In each trial, only one

compartment was baited, and a coarse (P60) or fine (P1500) sandpaper was secured to the

floor in the center of the start compartment. The high and low reward trials were randomly

assigned, with the rule that no more than two trials in a row were assigned the same size of

reward. When animals completed 90% (27/30) of the trials within an hour, they proceeded to the

training phase the following day.

The training phase was identical to the tactile learning phase, except that animals were only

allowed to enter one reward compartment of their choice and prevented from entering the other

reward compartment by placing an acrylic barrier following their choice. In addition, in one in

every five trials, no reward was initially placed in either reward compartment; but animals were

awarded a corresponding size of reward immediately after making the right choice, while no

reward was given for an incorrect choice. This controlled any olfactory cues from cheerios that

might guide decision-making and also served as reinforcement to enhance learning and reduce

extinction. The training phase consisted of up to three training sessions in a day, with each

training session containing 16 trials. Training continued until animals achieved 75% (12/16)

accuracy in a training session.

All male rats (N = 14) completed the learning and training phases within four days, except for

one that spent five days. Conversely, female rats (N = 12) took longer, with durations ranging

from four to nine days. Two female rats did not complete the learning and training phases within

nine days and were excluded from further experiments (Final N = 26; 12 females/14 males).

Following the completion of training, animals underwent one pre-probe, one baseline probe, and

one post-CRS probe session. The pre-probe was conducted the day after training was

completed to verify that animals had learned assigned task combination. Baseline and post-

CRS probes were carried out on Day 23 and 40, respectively. The probe phase was similar to

the training phase, except that there were a total of 30 trials and one in every five trials was

unrewarded. In these six unrewarded trials, three intermediate and ambiguous grades of

sandpaper (P240, P320, P400) were randomly assigned and each presented twice as probes.

The random presentation of three ambiguous tactile stimuli aimed to ensure that the animals did

not learn that there were no rewards in ambiguous trials and always chose a reward

compartment. All 26 animals completed 30 trials in the pre-probe, baseline and post-CRS probe

sessions. Additionally, no refresher training was conducted during the CRS period.

Figure 2. Learning, Training and Probe Protocols. The graph shows the procedure from a rat assigned task combination 1 (*Supplementary Table 1*). **A).** In the spatial and olfactory learning phase, the rat was trained to learn that high reward (three cheerios) was associated with cinnamon scent and left compartment, and low reward (half cheerio) was associated with coriander scent and right compartment; **B).** In the tactile learning phase, the same rat was further trained to learn that fine sandpaper indicated low reward and coarse sandpaper indicated high reward. In the training phase, an acrylic barrier was lowered to block movement between compartments as soon as the rat had made its choice; **C).** In the probe trials, one in every five trials was unrewarded. In these six unrewarded trials, three intermediate and ambiguous grades of sandpaper (P240, P320, P400) were randomly assigned and each presented twice as probes.

Bias was assessed using response data from six ambiguous trials in the probe session that met

the completion criteria, on an integer scale ranging from -3 to 3. A score of 3 represented an

extremely positive bias, characterized by six high-reward responses; a score of 0 suggested a

neutral bias, characterized by equal high-reward and low reward responses; whereas a score of

-3 indicated an extremely negative bias, characterized by six low-reward responses. The overall

response accuracy across 24 trials presenting distinct stimuli and the response accuracy

associated with high and low reward outcomes were calculated. Additionally, the total time

taken to complete a probe session (probe duration) was recorded.

Chronic Restraint Stress

From Day 24 to Day 37, CRS was performed on a bench situated in one corner of a spacious

animal holding room, with rats in the intervention group (6 females/7 males) placed in a

transparent plastic tube facing the wall to mitigate visual distractions (23). Each session started

between 12:00 and 13:00 PM, lasting for three hours daily to reduce the influence of circadian

rhythm. Following each session, animals were immediately returned to their home cages. The

control group (6 females/7 males) stayed in their home cages without undergoing CRS.

Open Field Test and Behavioral Automation

OFT (24) was conducted in an open arena (50 cm length × 50 cm width × 60 cm height) to

evaluate anxiety-like behaviors, with each animal tested individually. Animals were positioned

initially in the center zone and then allowed to freely explore the arena for ten minutes. The

apparatus was wiped with 70% ethanol to remove olfactory cues after each test session. Each

session was recorded using a Samsung A70 mobile phone and saved in mp4 format for further

processing and analysis. A total of 52 recordings were acquired at baseline and post-CRS. All

original recordings were edited to be ten minutes in duration to remove irrelevant (non-test

related) footage. The edited videos were then imported into DeepLabCut (25) to track animal

movement by labeling seven body points (*Figure 3A*) and four corners of the testing arena

(*Figure 3B*). Main steps involved in the behavioral labelling were: 1). Manually label 11 tracking

points of 300 randomly selected video frames (20 frames × 15 videos); 2). Train a deep neural

network using labelled frames; 3). Extract the coordinates of all tracking based on the trained

network. For each recording, DeepLabCut automatically generated one individual CSV file

containing the coordinates of 11 tracking points for all video frames, along with labelling

confidence (ranging from 0 ~ 100%) for each coordinate.

All DeepLabCut coordinate files derived from OFT recordings were imported into R 4.4.0. and

processed with DLCAnalyzer (26) and in-house R scripts (*Supplementary Scripts*). For each

video, the arena outline was first determined by the median coordinates of each corner point

using DLCAnalyzer. Also, rat body points with a likelihood below 95% or located outside the

testing arena were replaced through data interpolation. Then, the whole arena was virtually

divided into a series of 10 x 10 cm blocks based on the arena outline using the in-house script.

The outer zone consisted of 16 blocks (in blue, *Figure 3B*), while the inner zone contained 9

blocks (in beige, *Figure 3B*). The percentage of time spent in the outer zone during the first five

minutes, second five minutes, and the total ten minutes was computed as a classical measure

of anxiety-like behavior.

Figure 3. Labels Used to Train the Deep Neural Network. A). Rat with 7 body points;
B). OFT outline with 4 corners. Beige denotes inner zone. Blue denotes the outer zone.

MRI Acquisition and Batch Processing

Rats were scanned using Bruker Biospec 94/30 US/R pre-clinical MRI machine (Bruker BioSpin

GmbH, Germany) operating at 9.4 T (400MHz; H-1). Animals were anesthetized with isoflurane

and medetomidine during each MRI session; the anesthesia protocol was described and

discussed thoroughly in previous work (23, 27). *Supplementary Table 2* summaries the

parameters of two scanning sequences: T2w sMRI and rs-fMRI. Both raw images for each scan

session were compiled in one ParaVision 6.3.4 package in the format of PvDatasets.

A total of 23 animals completed both baseline and post-CRS scans. One control and one CRS

female rat did not complete their baseline scans due to irregular respiratory rate, and one

control male's baseline package was corrupted due to technical issues. The batch processing of

MRI data followed the same workflow as previous studies (27, 28). The workflow mainly

included data organization to the Brain Imaging Data Structure (29), bias field correction, skull

stripping, denoising, atlas registration to the Waxholm Space atlas (RRID:SCR_017124; 30),

and functional connectivity computation. To compute the functional connectome of the whole

brain for each animal, an in-house functional parcellation was mapped onto processed

functional images (N = 46) to derive subject-specific time series for 13 functional networks (28).

Fisher's r-to-Z transformation (31) and partial correlation with Tikhonov regularization (default

penalty strength of 0.1; 32, 33) was applied to these time series, resulting in 78 ($\frac{13 \times 12}{2}$) network

connectivity/edges.

Statistical Analysis

Behavioral Measures

Paired odds ratio (OR) for transition to negative bias from baseline to post-CRS was defined as

the ratio of discordant transitions within each sex, using McNemar's matched-pairs framework

(34, 35). 95% confidence interval (CI) was computed on the log scale using the standard

asymptotic variance for discordant pairs and exponentiated to return to the odds ratio scale. The

sex difference in bias shifts was summarized as the ratio of paired odds ratios (ROR), with a

log-scale CI calculated by combining the log-scale variances from the sex-specific paired ORs

(35).

All behavioral measures were analyzed using Linear Mixed-Effects Models (LMMs) using the

lme4 package (36) in R. These measures included bias score, response accuracy, probe

duration, and the dwell time in the periphery of the OFT. To account for the longitudinal,

repeated-measures structure of the data and the dependency within individual animals (N = 26),

models were fitted with the formula: outcome ~ sex × group × timepoint + (1 | animal). This

included sex (male vs female), group (CRS vs control), and timepoint (baseline vs post-CRS) as

fixed factors and their three-way interaction, with a random intercept for animal to control for

repeated-measures variance. Model assumptions, including residual normality and

homoscedasticity, were rigorously assessed using the performance package (37). For

dependent variables violating these assumptions, a bootstrap method was applied to refit their

models (36, 38). Significant three-way interactions were decomposed by assessing simple two-

way interactions at each sex stratum, followed by simple main effects of timepoint, stratified by

sex and group. Post-hoc comparisons were conducted using the rstatix (39) R packages, with

Bonferroni corrections applied for multiple comparisons.

All measurements were presented as mean ± SD, unless otherwise specified. Effect sizes were

reported with partial eta squared ($\eta^2 p$) and partial omega squared ($\omega^2 p$). Since $\omega^2 p$ is widely

viewed as lesser biased to $\eta^2 p$, effect sizes were evaluated based on $\omega^2 p$ and classified into

four categories: < 0.02 as very small, 0.02 ~ 0.13 as small, 0.13 ~ 0.26 as medium, and ≥ 0.26

as large (40). Statistical significance was set as $p < 0.05$.

Network Connectivity

The network connectivity of the whole brain (78 edges) was analyzed using Network-Based R-

Statistics with mixed-effects models (NBR; 41) to account for the longitudinal, repeated-

measures structure of the data from 23 animals. NBR extends the original Network-Based

Statistics framework (42) by incorporating linear mixed-effects models and applying permutation

testing for family-wise error (FWE) control. This approach enables inference at the subnetwork

level rather than relying solely on edgewise inference. First, a linear mixed-effect model was

fitted to each edge using the same model structure as for the behavioral analysis in the previous

section. Edges with p values below the initial significance threshold ($p < 0.05$) were then

clustered into a subnetwork based on shared nodes. The term "subnetwork" refers specifically

to the connected set of edges identified by NBR and not to a subset of the 13 predefined

functional networks. Finally, the statistical significance of the subnetwork was evaluated against

an empirical null distribution of subnetwork size and strength generated through 5000

permutations. P values were corrected for FWE and calculated as the proportion of

permutations in which the maximal component statistic equaled or exceeded the observed

values. The identified subnetwork was visualized as a chord diagram, and the corresponding

null distributions of subnetwork size and strength were shown as density plots. Significant three-

way interactions were further decomposed by assessing simple two-way interactions at each

sex stratum, followed by simple main effects of timepoint within each sex-by-group stratum.

These follow-up analyses followed the same procedure described above, but were restricted to

the identified subnetwork.

To explore whether the identified subnetwork could be represented as a scalar outcome, the

mean connectivity across edges within the subnetwork was computed and analyzed using a

follow-up linear mixed-effects model. This scalar summary did not reproduce the significant

three-way interaction, suggesting that a simple average of edge values did not adequately

capture the subnetwork pattern identified by NBR.

Scripts and workspace for statistical analysis can be found in *Supplementary Script* and

Workspace.

Results

Bias Measures

Following CRS, female rats showed higher odds of transitioning towards a negative bias (paired OR = 3.67, 95% CI: 0.60 ~ 22.30; exact P = 0.22), whereas males showed no directional change (paired OR = 1.00, 95% CI: 0.10 ~ 9.61; exact P = 1.00). The female-to-male ROR was 3.67 (95% CI: 0.20 ~ 66.31), with wide confidence intervals reflecting limited discordant transitions.

For the bias, the LMM revealed a significant three-way interaction with a medium effect size among sex, group, and timepoint ($F_{1,22}$ = 4.95, p = 0.037, η^2p = 0.18, ω^2p = 0.14). The model also identified a significant main effect of sex, with a medium effect size ($F_{1,22}$ = 4.87, p = 0.038, η^2p = 0.18, ω^2p = 0.14). Decomposition via simple two-way interaction analyses showed a significant interaction between group and timepoint with a strong effect size for female rats ($F_{1,10}$ = 9.20, p = 0.013, η^2p = 0.48, ω^2p = 0.41), but not in male rats ($F_{1,12}$ = 0.00, p = 1.000, η^2p = 0.00, ω^2p = 0.00; *Figure 4A*). Subsequent simple main effects of timepoint within each sex and group stratum were not significant (*Figure 4B*).

Figure 4. Plots Showing Findings from LMM for Bias. A). Interaction plot showing the interaction between group and timepoint was significant for female rats, but not for male rats; **B).** Violin plots showing no significant differences in the bias at baseline and post-CRS, for each sex-by-group stratum. Each paired set of observations is connected by a line.

Other Behavioral Measures

For response accuracy, probe duration and OFT measures, LMM models did not identify significant three-way interactions among sex, group, and time (*Supplementary Spreadsheet*). However, significant main effects of sex were identified for the probe duration and OFT measures, but not for response accuracy. For instance, there was no significant three-way

319 interaction ($F_{1,22}$ = 0.94, p = 0.34, η²p = 0.04, ω²p = 0.00), but a significant main effect of sex
320 ($F_{1,22}$ = 6.47, p = 0.02, η²p = 0.23, ω²p = 0.19) for probe duration.

Network Connectivity

NBR analysis identified a nine-edge subnetwork associated with the sex × group × timepoint
interaction (*Figure 5A)*. These nine edges spanned subcortical, cerebellar, brainstem, and
cortical networks, with multiple edges involving the ventral striatum–cortical and spatial attention
networks (Table 1). This subnetwork was significant by size (p_{FWE} = 0.04; *Figure 5B*), but not by
strength (p_{FWE} = 0.11; *Figure 5C*). Further decomposition revealed five edges within the
subnetwork that were associated with a significant interaction between group and timepoint in
female rats (p_{FWE} = 0.02). There five edges (edge 21, 26, 35, 50 and 76) were interconnected
through two primary networks: auditory thalamus–brainstem and lateral–temporal cortical
networks (Table 1). Subsequent simple main effects of timepoint within each sex and group
stratum were not significant for the subnetwork.

Table 1. Edges Comprising the Subnetwork Identified by the NBR Analysis

	Edges		t values
ID*	Network 1	Network 2	
21	*Hypothalamus–Thalamus*	*Auditory Thalamus–Brainstem*	0.33
22	Vermis–Left Dorsal cortex of inferior colliculi Network	(Para)Flocculus–Brainstem	0.24
26	*Lateral–Temporal Cortical Network*	*(Para)Flocculus–Brainstem*	-0.99
31	Left Cerebellum–Auditory Processing Network	Ventral Striatum–Cortical Network	0.19
35	*Auditory Thalamus–Brainstem*	*Ventral Striatum–Cortical Network*	-0.73
50	*Lateral–Temporal Cortical Network*	*Salience–orbitofrontal Network*	-0.33
67	Vermis–Left Dorsal cortex of inferior colliculi Network	Spatial Attention Network	0.36
75	Ventral Striatum–Cortical Network	Spatial Attention Network	-0.27
76	*Corpus Striatum–Cortical Network*	*Spatial Attention Network*	-0.07

*Note: * Edge ID indicates the original ordering of each edge in the upper triangle of the full connectivity matrix (13 × 13). Details of these networks can be found in Dai et al. (28).*

Figure 5. Plots Showing Findings from NBR Analysis. A). Chord diagram showing the nine-edge subnetwork identified by NBR analysis. Ribbon color indicates the sign and magnitude of the edge-level test statistic; **B).** Density plot showing null distribution of maximal subnetwork size over 5000 permutations. The dashed line indicates the significant size, corrected for FWE (p = 0.04); **C).** Density plot showing null distribution of maximal subnetwork strength over 5000 permutations. The dashed line indicates the observed subnetwork strength, which did not survive FWE corrections (p = 0.11).

Discussion

The present study aimed to determine whether chronic stress during adolescence was associated with sex-dependent alterations in cognitive bias and functional connectome in emerging adulthood. A major innovation of the work lies in the longitudinal integration of a translationally relevant judgment bias assay with resting-state connectomes, which allows behavioral and network trajectories to be examined in parallel within the same cohort. The findings indicate that the change in bias and resting-state reorganization from baseline to post-CRS were sex-dependent. Specifically, females showed a greater tendency toward negative bias than males, and network analysis identified a subnetwork spanning brainstem, limbic, striatal, insular, parahippocampal, orbitofrontal, and retrosplenial regions. Together, these results highlight the translational importance of considering sex and distributed network organization when modelling how adolescent stress may shape later vulnerability to depression.

Negative Bias: Longitudinal Effects of Adolescent CRS in Adult Females and Males

The primary behavioral finding of this study was that the directional change in ambiguity appraisal from baseline to post-CRS differed between females and males, as indicated by a significant three-way interaction and a subsequent two-way interaction in females. However, the absence of significant simple main effects within the female CRS group calls for a cautious interpretation. These data do not suggest a uniform transition toward negative bias in females after chronic stress. Rather, the significant interactions indicate that the female CRS and control groups differed in how ambiguity appraisal changed from baseline to post-CRS, whereas no comparable pattern was evident in males. This interpretation is consistent with the paired transition analysis suggesting a greater tendency toward negative bias in females than in males. Importantly, because neither task performance nor anxiety-like behavior showed significant interactions or simple main effects of time, this tendency toward negative bias is less likely to reflect a broad behavioral disruption and more likely to reflect a selective change in ambiguity appraisal. This result is also unlikely to be explained by normative maturation alone, given that both control and CRS groups were assessed in early adolescence and again in young adulthood.

Broadly, the present findings are consistent with evidence that sex differences in cognitive vulnerability to depression become more apparent during adolescence (10). One plausible mechanism is the sex-specific programming of the HPA axis during this period (18). Specifically, repeated glucocorticoid signaling in females may alter stress reactivity, subsequently biasing the appraisal of ambiguous stimuli toward more negative interpretations. In this respect, the present

study extends work in adult models (43, 44). Although stress exposure in adulthood can acutely

induce negative bias (45), these effects are often transient (46). This raises the possibility that

stress during adulthood may be insufficient to recalibrate cognitive appraisal, whereas

adolescence represents a critical period for the programming of adult affective processing (18,

47). The current work also extends the scope of the latent vulnerability framework (48), aligning

with convergent human and developmental evidence suggesting that a negative interpretation

bias under uncertainty is a sex-dependent recalibration of affective systems, gated by the

intersection of sex and developmental timing (49, 50).

Network Connectivity: Longitudinal Effects of Adolescent CRS in Adult Females
and Males

The network analysis showed an interaction pattern similar to that observed for bias, and follow-

up decomposition localized the female-specific interaction to a five-edge subnetwork. Because

this subnetwork was significant by size rather than strength, the effects of adolescent CRS

appear to have differed between females and males at the level of widespread network

reorganization rather than connectivity magnitude. This finding is consistent with prior evidence

showing that stress can reorganize large-scale brain networks (51, 52) and broadly aligns with

current models of depression that increasingly emphasize abnormalities across interacting

large-scale networks rather than dysfunction within a single affective circuit (53, 54).

The two primary networks highlighted in the follow–up analysis point to resting-state networks

associated with sensory regulation (55), multisensory integration (56) and contextual

association (57, 58). Within the identified subnetwork, the thalamus likely mediates sensory

gating (55), whereas the para–hippocampal regions, including the perirhinal, postrhinal, and

entorhinal cortices, support contextual and associative processing (56). Furthermore, the insula

and sensory association cortices may provide the necessary architecture for multisensory and

interoceptive integration (57, 58). Beyond these two primary networks, the subnetwork also

extends to functional networks associated with physiological state (59), motivational salience

(60-62), and spatial attention (63). Specifically, the hypothalamus–thalamus network likely

regulates physiological state through reciprocal connections. The hypothalamus monitors

internal and external conditions to influence autonomic, endocrine, and behavioral responses,

whereas the thalamus contributes to sensory and emotional processing. The ventral striatum–

cortical, corpus striatum–cortical, and salience–orbitofrontal networks are likely engaged in

motivational salience. The corticostriatal circuits serve as a high-level integrative system that

links cortical sensory, emotional, and cognitive input to behavioral prioritization and action

selection (60). The salience network and orbitofrontal cortex form a coordinated system for the

prioritization and appraisal of environmental cues (61, 62). The dorsal hippocampus and

retrosplenial cortex constitute the core functional components of the spatial attention network

(28), which likely supports the selective allocation of attention across space, thereby guiding

adaptive behaviors (63).

This functional profile is relevant to stress because mounting evidence suggests that chronic

stress can alter brain organization beyond classical affective circuits, such as the prefrontal-

limbic circuit. For example, a longitudinal imaging study shows structural and functional changes

across the thalamus, hypothalamus, striatum, brainstem, auditory cortex, insula, and entorhinal

cortex as stress exposure unfolds over time (51). Importantly, sex differences in neural

response to adolescent stress have been reported, with the clearest evidence pointing to sex-

dependent alterations in the hippocampus (64, 65), amygdala (66, 67), and hypothalamus (68,

69). More limited but growing evidence also supports the involvement of insula-prelimbic

pathway (70), ventral striatum (71) and brainstem dopaminergic nuclei such as the ventral

tegmental area (72). However, direct evidence for sex-dependent effects of adolescent stress

remains limited for other regions within the subnetwork identified here, including the thalamus,

auditory cortex, and entorhinal cortex. Further animal and human studies are therefore needed

to determine whether these regions also contribute to the sex differences in neural response to

adolescent stress.

Overall Sex Differences in Bias, Training and Probe Duration

The significant main effect of sex on bias and probe duration, together with longer training

duration in female Wistar rats, is also reported in Sprague-Dawley rats (46, 73, 74). The

absence of sex differences in response accuracy makes it less likely that the overall sex

differences in bias, training and probe duration reflect poorer task learning or maintenance in

one sex. Instead, these findings suggest a broader sex difference in response strategies

associated with choice, learning, and response speed (75, 76).

Limitations and Future Direction

Several limitations of this study should be acknowledged.

First, given the modest sample size and the absence of a prior power analysis, the findings on

bias and resting-state networks require cautious interpretation. Although both measures showed

significant sex-dependent interactions, simple main effects of timepoint within each sex-by-

group stratum were not significant. This may reflect limited statistical power after stratification,

particularly for the MRI analyses. Longitudinal studies with larger sample sizes are warranted to

determine whether these sex-dependent effects of adolescent CRS represent reliable changes

in behavior and network organization.

Second, the study applied one CRS protocol without physiological measures of stress response.

Equivalent restraint schedules do not guarantee equivalent biological stress load across sexes

(16). In addition, adolescent stressors vary in intensity, predictability, and social context, with

potentially different behavioral and neural outcomes in adulthood. Future work should examine

whether the present findings generalize across different adolescent stressors, while

incorporating physiological indices of stress response, such as corticosterone and adrenal

measures.

Third, the anesthetized "resting" state during rs-fMRI captures stable network reorganization,

whereas the awake "deciding" brain reflects dynamic cognitive processing (77, 78). Task

performance is likely to depend on multiple processes, including sensory discrimination,

selective attention, memory for learned cue–reward contingencies, and the transformation of

ambiguous sensory information into action (79). Future investigations should employ

multiphoton imaging or electrophysiology to record real-time neural activity during the task (80,

81), directly assessing how circuit dynamics give rise to individual choices under ambiguity.

Conclusion

This study suggests that adolescent chronic restraint stress in rodents is associated with sex-

dependent alterations in cognitive bias and distributed resting-state networks. The findings

highlight the importance of sex and network organization in understanding how adolescent

stress may shape later vulnerability to depression.

Acknowledgments

The authors thank Lakshini Piyasiti, Neha Nair, Rebecca Goodyear, and Will Copping for assistance with chronic restraint stress, and UWA Animal Care Services for excellent animal care. The authors also acknowledge the facilities and scientific and technical assistance of the National Imaging Facility, a National Collaborative Research Infrastructure Strategy (NCRIS) capability, at the Centre for Microscopy, Characterisation and Analysis, UWA.

CRediT authorship contribution statement

Twain Dai: Writing - Original Draft, Writing - Review & Editing, Conceptualization, Methodology, Software, Formal analysis, Investigation, Resources, Data Curation, Visualization**,** Project administration; **Liz Jaeschke-Angi:** Investigation (MRI acquisition)**, Marissa Penrose-Menz:** Investigation (MRI acquisition)**, Tim Rosenow:** Investigation (MRI acquisition)**, Jennifer Rodger:** Conceptualization, Writing - Review & Editing, Supervision**.**

Funding

TD was supported by the Australian Government International Research Training Program scholarship and the Byron Kakulas Prestige scholarship. JR was supported by a program grant from the Stan Perron Charitable Foundation. The funders had no role in study design; data collection, analysis, or interpretation; manuscript preparation; or the decision to submit for publication.

Competing Interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Reference

1. Uddin LQ, Castellanos FX, Menon V. Resting state functional brain connectivity in child and adolescent psychiatry: where are we now? Neuropsychopharmacology (New York, NY). 2024;50(1):196-200.

2. Selemon LD. A role for synaptic plasticity in the adolescent development of executive function. Translational psychiatry. 2013;3(3):e238-e.

3. Sun L, Zhao T, Liang X, Xia M, Li Q, Liao X, et al. Human lifespan changes in the brain's functional connectome. Nature neuroscience. 2025;28(4):891-901.

4. Feng G, Wang Y, Huang W, Chen H, Cheng J, Shu N. Spatial and temporal pattern of structure–function coupling of human brain connectome with development. eLife. 2024;13.

5. Sisk LM, Gee DG. Stress and adolescence: vulnerability and opportunity during a sensitive window of development. Current opinion in psychology. 2022;44:286-92.

6. Rakesh D, Allen NB, Whittle S. Longitudinal changes in within-salience network functional connectivity mediate the relationship between childhood abuse and neglect, and mental health during adolescence. Psychological medicine. 2023;53(4):1552-64.

7. Orchard F, Pass L, Reynolds S. Associations Between Interpretation Bias and Depression in Adolescents. Cognitive therapy and research. 2016;40(4):577-83.

8. Beevers CG, Mullarkey MC, Dainer-Best J, Stewart RA, Labrada J, Allen JJB, et al. Association Between Negative Cognitive Bias and Depression: A Symptom-Level Approach. Journal of abnormal psychology (1965). 2019;128(3):212-27.

9. Vos LMW, Nieto I, Amanvermez Y, Smeets T, Everaert J. Do cognitive biases prospectively predict anxiety and depression? A multi-level meta-analysis of longitudinal studies. Clinical psychology review. 2025;116:102552.

10. Mezulis AH, Funasaki KS, Charbonneau AM, Hyde JS. Gender Differences in the Cognitive Vulnerability-Stress Model of Depression in the Transition to Adolescence. Cognitive therapy and research. 2010;34(6):501-13.

11. Chuang J-Y, Hagan CC, Murray GK, Graham JME, Ooi C, Tait R, et al. Adolescent Major Depressive Disorder: Neuroimaging Evidence of Sex Difference during an Affective Go/No-Go Task. Frontiers in psychiatry. 2017;8:119.

12. Young KD, Bellgowan PS, Bodurka J, Drevets WC. Functional neuroimaging correlates of autobiographical memory deficits in subjects at risk for depression. Brain Sci. 2015;5(2):144-64.

13. Lagisz M, Zidar J, Nakagawa S, Neville V, Sorato E, Paul ES, et al. Optimism, pessimism and judgement bias in animals: A systematic review and meta-analysis. Neuroscience and biobehavioral reviews. 2020;118:3-17.

14. Cryan JF, Holmes A. The ascent of mouse: advances in modelling human depression and anxiety. Nature reviews Drug discovery. 2005;4(9):775-90.

15. Borsini A, Giacobbe J, Mandal G, Boldrini M. Acute and long-term effects of adolescence stress exposure on rodent adult hippocampal neurogenesis, cognition, and behaviour. Molecular psychiatry. 2023;28(10):4124-37.

16. McCormick CM, Green MR. From the stressed adolescent to the anxious and depressed adult: Investigations in rodent models. Neuroscience. 2013;249:242-57.

17. Aylward J, Hales C, Robinson E, Robinson OJ. Translating a rodent measure of negative bias into humans: the impact of induced anxiety and unmedicated mood and anxiety disorders. Psychological medicine. 2020;50(2):237-46.

18. Chaby LE, Cavigelli SA, White A, Wang K, Braithwaite VA. Long-term changes in cognitive bias and coping response as a result of chronic unpredictable stress during adolescence. Frontiers in human neuroscience. 2013;7:328.

19. Mengelkoch S, Slavich GM. Sex Differences in Stress Susceptibility as a Key Mechanism Underlying Depression Risk. Current psychiatry reports. 2024;26(4):157-65.

20. Karp NA, Berdoy M, Gray K, Hunt L, Jennings M, Kerton A, et al. The Sex Inclusive Research Framework to address sex bias in preclinical research proposals. Nature communications. 2025;16(1):3763-6.

21. Ghasemi A, Jeddi S, Kashfi K. The laboratory rat: Age and body weight matter. EXCLI journal. 2021;20:1431-45.

22. Brydges NM, Hall L. A shortened protocol for assessing cognitive bias in rats. Journal of Neuroscience Methods. 2017;286:1-5.

23. Seewoo BJ, Hennessy LA, Feindel KW, Etherington SJ, Croarkin PE, Rodger J. Validation of Chronic
Restraint Stress Model in Young Adult Rats for the Study of Depression Using Longitudinal
Multimodal MR Imaging. eNeuro. 2020;7(4):1-22.
24. Seibenhener ML, Wooten MC. Use of the Open Field Maze to Measure Locomotor and Anxiety-like
Behavior in Mice. Journal of visualized experiments. 2015(96):e52434-e.
25. Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, et al. DeepLabCut: markerless
pose estimation of user-defined body parts with deep learning. Nature Neuroscience.
2018;21(9):1281-9.
26. Sturman O, von Ziegler L, Schläppi C, Akyol F, Privitera M, Slominski D, et al. Deep learning-based
behavioral analysis reaches human accuracy and is capable of outperforming commercial
solutions. Neuropsychopharmacology. 2020;45(11):1942-52.
27. Dai T, Seewoo BJ, Hennessy LA, Bolland SJ, Rosenow T, Rodger J. Identifying reproducible resting
state networks and functional connectivity alterations following chronic restraint stress in
anaesthetized rats. Frontiers in Neuroscience. 2023;17:1151525.
28. Dai T, Algar SD, Small M, Zalesky A, Rodger J. Identifying Neuroimaging Biomarkers of Resilience
and Vulnerability to Chronic Stress in An Animal Model: An Exploratory Analysis. bioRxiv.
2025:2025.08.05.667822.
29. Gorgolewski KJ, Auer T, Calhoun VD, Craddock RC, Das S, Duff EP, et al. The brain imaging data
structure, a format for organizing and describing outputs of neuroimaging experiments. Scientific
Data. 2016;3(1):160044.
30. Papp EA, Leergaard TB, Calabrese E, Johnson GA, Bjaalie JG. Waxholm Space atlas of the Sprague
Dawley rat brain. Neuroimage. 2014;97:374-86.
31. Smith SM, Miller KL, Salimi-Khorshidi G, Webster M, Beckmann CF, Nichols TE, et al. Network
modelling methods for FMRI. Neuroimage. 2011;54(2):875-91.
32. Liégeois R, Santos A, Matta V, Van De Ville D, Sayed AH. Revisiting correlation-based functional
connectivity and its relationship with structural connectivity. Network neuroscience (Cambridge,
Mass). 2020;4(4):1235-51.
33. Suri S, Topiwala A, Filippini N, Zsoldos E, Mahmood A, Sexton CE, et al. Distinct resting-state
functional connections associated with episodic and visuospatial memory in older adults.
NeuroImage (Orlando, Fla). 2017;159:122-30.
34. McNemar Q. Note on the sampling error of the difference between correlated proportions or
percentages. Psychometrika. 1947;12(2):153-7.
35. Cleophas TJ, Zwinderman AH. McNemar's Odds Ratios. In: Cleophas TJ, Zwinderman AH, editors.
Clinical Data Analysis on a Pocket Calculator: Understanding the Scientific Methods of Statistical
Reasoning and Hypothesis Testing. Cham: Springer International Publishing; 2016. p. 275-8.
36. Douglas B, Martin M, Ben B, Steve W. Fitting Linear Mixed-Effects Models Using lme4. Journal of
statistical software. 2015;67(1):1-48.
37. Lüdecke D, Ben-Shachar M, Patil I, Waggoner P, Makowski D. performance: An R Package for
Assessment, Comparison and Testing of Statistical Models. Journal of open source software.
2021;6(60):3139.
38. Loughin TM. Bootstrap Methods and Their Application. Alexandria: American Statistical Association;
1999. p. 334-5.
39. Kassambara A. rstatix: Pipe-friendly framework for basic statistical tests. CRAN: Contributed
Packages. 2019.
40. Cohen J. A Power Primer. Psychological bulletin. 1992;112(1):155-9.
41. Gracia-Tabuenca Z, Alcauter S. NBR: Network-based R-statistics for (unbalanced) longitudinal
samples. 1.1 ed. Cold Spring Harbor: Cold Spring Harbor Laboratory Press; 2020.
42. Zalesky A, Fornito A, Bullmore ET. Network-based statistic: Identifying differences in brain networks.
NeuroImage (Orlando, Fla). 2010;53(4):1197-207.
43. Rygula R, Papciak J, Popik P. Trait Pessimism Predicts Vulnerability to Stress-Induced Anhedonia in
Rats. Neuropsychopharmacology (New York, NY). 2013;38(11):2188-96.
44. Papciak J, Popik P, Fuchs E, Rygula R. Chronic psychosocial stress makes rats more 'pessimistic' in
the ambiguous-cue interpretation paradigm. Behavioural brain research. 2013;256:305-10.
45. Enkel T, Gholizadeh D, von Bohlen und Halbach O, Sanchis-Segura C, Hurlemann R, Spanagel R, et
al. Ambiguous-Cue Interpretation is Biased Under Stress- and Depression-Like States in Rats.
Neuropsychopharmacology (New York, NY). 2010;35(4):1008-15.

46. Aliphon B, Dai T, Moretti J, Penrose-Menz M, Mulders WHAM, Blache D, et al. A repeated measures
cognitive affective bias test in rats: comparison with forced swim test. Psychopharmacology.
2022.
47. Avital A, Richter-Levin G. Exposure to juvenile stress exacerbates the behavioural consequences of
exposure to stress in the adult rat. The international journal of neuropsychopharmacology.
2005;8(2):163-73.
48. McCrory EJ, Viding E. The theory of latent vulnerability: Reconceptualizing the link between childhood
maltreatment and psychiatric disorder. Development and psychopathology. 2015;27(2):493-505.
49. Nweze T, Ezenwa M, Ajaelu C, Hanson JL, Okoye C. Cognitive variations following exposure to
childhood adversity: evidence from a pre-registered, longitudinal study. EClinicalMedicine.
2023;56:101784-.
50. Ho TC, Buthmann J, Chahal R, Miller JG, Gotlib IH. Exploring sex differences in trajectories of
pubertal development and mental health following early adversity. Psychoneuroendocrinology.
2024;161:106944-.
51. Magalhães R, Barrière DA, Novais A, Marques F, Marques P, Cerqueira J, et al. The dynamics of
stress: a longitudinal MRI study of rat brain structure and connectome. Molecular psychiatry.
2018;23(10):1998-2006.
52. Hermans EJ, Henckens MJAG, Joëls M, Fernández G. Dynamic adaptation of large-scale brain
networks in response to acute stressors. Trends in neurosciences (Regular ed). 2014;37(6):304-
14.
53. Kaiser RH, Andrews-Hanna JR, Wager TD, Pizzagalli DA. Large-Scale Network Dysfunction in Major
Depressive Disorder: A Meta-analysis of Resting-State Functional Connectivity. JAMA Psychiatry.
2015;72(6):603-11.
54. Zhang Z, Zhang Y, Wang H, Lei M, Jiang Y, Xiong D, et al. Resting-state network alterations in
depression: a comprehensive meta-analysis of functional connectivity. Psychological medicine.
2025;55:e63.
55. Shine JM, Lewis LD, Garrett DD, Hwang K. The impact of the human thalamus on brain-wide
information processing. Nature reviews Neuroscience. 2023;24(7):416-30.
56. Choi I, Demir I, Oh S, Lee S-H. Multisensory integration in the mammalian brain: diversity and
flexibility in health and disease. Philosophical transactions of the Royal Society of London Series
B Biological sciences. 2023;378(1886):20220338.
57. Peng X, Burwell RD. Beyond the hippocampus: The role of parahippocampal-prefrontal
communication in context-modulated behavior. Neurobiology of learning and memory.
2021;185:107520-.
58. Doan TP, Lagartos-Donate MJ, Nilssen ES, Ohara S, Witter MP. Convergent Projections from
Perirhinal and Postrhinal Cortices Suggest a Multisensory Nature of Lateral, but Not Medial,
Entorhinal Cortex. Cell reports (Cambridge). 2019;29(3):617-27.e7.
59. Goel M, Mittal A, Jain VR, Bharadwaj A, Modi S, Ahuja G, et al. Integrative Functions of the
Hypothalamus: Linking Cognition, Emotion and Physiology for Well-being and Adaptability.
Annals of Neurosciences. 2025;32(2):128-42.
60. Shipp S. The functional logic of corticostriatal connections. Brain Structure and Function.
2017;222(2):669-706.
61. Rolls ET, Cheng W, Feng J. The orbitofrontal cortex: reward, emotion and depression. Brain
communications. 2020;2(2):fcaa196-fcaa.
62. Schimmelpfennig J, Topczewski J, Zajkowski W, Jankowiak-Siuda K. The role of the salience network
in cognitive and affective deficits. Frontiers in human neuroscience. 2023;17:1133367-.
63. Fiebelkorn IC, Kastner S. Functional Specialization in the Attention Network. Annual review of
psychology. 2020;71(1):221-49.
64. Brydges NM, Moon A, Rule L, Watkin H, Thomas KL, Hall J. Sex specific effects of pre-pubertal stress
on hippocampal neurogenesis and behaviour. Translational psychiatry. 2018;8(1):271-12.
65. Hyer MM, Shaw GA, Goswamee P, Dyer SK, Burns CM, Soriano E, et al. Chronic adolescent stress
causes sustained impairment of cognitive flexibility and hippocampal synaptic strength in female
rats. Neurobiology of stress. 2021;14:100303.
66. Yohn NL, Blendy JA. Adolescent Chronic Unpredictable Stress Exposure Is a Sensitive Window for
Long-Term Changes in Adult Behavior in Mice. Neuropsychopharmacology (New York, NY).
2017;42(8):1670-8.

67. Lovelock DF, Deak T. Acute stress imposed during adolescence yields heightened anxiety in Sprague
Dawley rats that persists into adulthood: Sex differences and potential involvement of the Medial
Amygdala. Brain research. 2019;1723:146392-.
68. Wulsin AC, Wick-Carlson D, Packard BA, Morano R, Herman JP. Adolescent chronic stress causes
hypothalamo–pituitary–adrenocortical hypo-responsiveness and depression-like behavior in adult
female rats. Psychoneuroendocrinology. 2016;65:109-17.
69. Rowson SA, Pleil KE. Influences of Stress and Sex on the Paraventricular Thalamus: Implications for
Motivated Behavior. Frontiers in behavioral neuroscience. 2021;15:636203.
70. Kin K, Francis-Oliveira J, Kano S-i, Niwa M. Adolescent stress impairs postpartum social behavior via
anterior insula-prelimbic pathway in mice. Nature communications. 2023;14(1):2975-15.
71. Warren BL, Sial OK, Alcantara LF, Greenwood MA, Brewer JS, Rozofsky JP, et al. Altered Gene
Expression and Spine Density in Nucleus Accumbens of Adolescent and Adult Male Mice
Exposed to Emotional and Physical Stress. Developmental neuroscience. 2014;36(3-4):250-60.
72. Zhu X, Grace AA. Sex- and exposure age-dependent effects of adolescent stress on ventral
tegmental area dopamine system and its afferent regulators. Molecular psychiatry.
2023;28(2):611-24.
73. Zhang Q, Li M, Wang Z, Chen F. Sex differences in learning and performing the Go/NoGo tasks.
Biology of sex differences. 2023;14(1):25-16.
74. Barker TH, Bobrovskaya L, Howarth GS, Whittaker AL. Female rats display fewer optimistic
responses in a judgment bias test in the absence of a physiological stress response. Physiology
& behavior. 2017;173:124-31.
75. Palmer JA, White SR, Chavez Lopez K, Laubach M. The Role of the Rat Prefrontal Cortex and Sex
Differences in Decision-Making. The Journal of neuroscience. 2024;44(46):e0550242024.
76. Chen CS, Ebitz RB, Bindas SR, Redish AD, Hayden BY, Grissom NM. Divergent Strategies for
Learning in Males and Females. Current biology. 2021;31(1):39-50.e4.
77. Tauber JM, Brincat SL, Stephen EP, Donoghue JA, Kozachkov L, Brown EN, et al. Propofol-mediated
Unconsciousness Disrupts Progression of Sensory Signals through the Cortical Hierarchy.
Journal of cognitive neuroscience. 2024;36(2):394-413.
78. Bonhomme V, Staquet C, Montupil J, Defresne A, Kirsch M, Martial C, et al. General Anesthesia: A
Probe to Explore Consciousness. Frontiers in systems neuroscience. 2019;13:36.
79. Levy I, Schiller D. Uncertainty in learning and decision-making: Introduction to the special issue.
Cognitive, affective, & behavioral neuroscience. 2023;23(3):459-64.
80. Vickers ED, McCormick DA. Pan-cortical 2-photon mesoscopic imaging and neurobehavioral
alignment in awake, behaving mice. eLife. 2024;13.
81. Steinmetz NA, Aydin C, Lebedeva A, Okun M, Pachitariu M, Bauza M, et al. Neuropixels 2.0: A
miniaturized high-density probe for stable, long-term brain recordings. Science (American
Association for the Advancement of Science). 2021;372(6539).
679

Figure legends and Tables

Figure 1. Study Design and Timeline. All animals were habituated to the new environment for nine days after their arrival and prior to experiments. All rats were trained on JBT, for up to nine days. Behavioral tests and MRI scan were conducted before and after 14 consecutive days of CRS. Investigators were not blinded to group allocation during CRS administration or behavioral testing, because knowledge of group assignment was required to deliver the appropriate procedures.

Figure 2. Learning, Training and Probe Protocols. The graph shows the procedure from a rat assigned task combination 1 (*Supplementary Table 1*). **A).** In the spatial and olfactory learning phase, the rat was trained to learn that high reward (three cheerios) was associated with cinnamon scent and left compartment, and low reward (half cheerio) was associated with coriander scent and right compartment; **B).** In the tactile learning phase, the same rat was further trained to learn that fine sandpaper indicated low reward and coarse sandpaper indicated high reward. In the training phase, an acrylic barrier was lowered to block movement between compartments as soon as the rat had made its choice; **C).** In the probe trials, one in every five trials was unrewarded. In these six unrewarded trials, three intermediate and ambiguous grades of sandpaper (P240, P320, P400) were randomly assigned and each presented twice as probes.

Figure 3. Labels Used to Train the Deep Neural Network. A). Rat with 7 body points; **B).** OFT outline with 4 corners. Beige denotes inner zone. Blue denotes the outer zone.

Figure 4. Plots Showing Findings from LMM for Bias. A). Interaction plot showing the interaction between group and timepoint was significant for female rats, but not for male rats; **B).** Violin plots showing no significant differences in the bias at baseline and post-CRS, for each sex-by-group stratum. Each paired set of observations is connected by a line.

Figure 5. Plots Showing Findings from NBR Analysis. A). Chord diagram showing the nine-edge subnetwork identified by NBR analysis. Ribbon color indicates the sign and magnitude of the edge-level test statistic; **B).** Density plot showing null distribution of maximal subnetwork size over 5000 permutations. The dashed line indicates the significant size, corrected for FWE (p = 0.04)**; C).** Density plot showing null distribution of maximal subnetwork strength over 5000 permutations. The dashed line indicates the observed subnetwork strength, which did not survive FWE corrections (p = 0.11).

Table 1. Edges Comprising the Subnetwork Identified by the NBR Analysis

	Edges		t values
ID*	Network 1	Network 2	
21	*Hypothalamus–Thalamus*	*Auditory Thalamus–Brainstem*	0.33
22	Vermis–Left Dorsal cortex of inferior colliculi Network	(Para)Flocculus–Brainstem	0.24
26	*Lateral–Temporal Cortical Network*	*(Para)Flocculus–Brainstem*	-0.99
31	Left Cerebellum–Auditory Processing Network	Ventral Striatum–Cortical Network	0.19
35	*Auditory Thalamus–Brainstem*	*Ventral Striatum–Cortical Network*	-0.73
50	*Lateral–Temporal Cortical Network*	*Salience–orbitofrontal Network*	-0.33
67	Vermis–Left Dorsal cortex of inferior colliculi Network	Spatial Attention Network	0.36
75	Ventral Striatum–Cortical Network	Spatial Attention Network	-0.27
76	*Corpus Striatum–Cortical Network*	*Spatial Attention Network*	-0.07

*Note: * Edge ID indicates the original ordering of each edge in the upper triangle of the full connectivity matrix (13 × 13). Details of these networks can be found in Dai et al. (28).*