Mount Fuji's stubby peak: the genotypic density of additive landscapes near maximal fitness

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Abstract

Additive fitness landscapes—also called Mount Fuji landscapes—are the simplest and most widely used models of sequence-function relationships. As such, they play essential roles across multiple areas of biology, including evolutionary theory, quantitative genetics, gene regulation, and protein science. One of the most basic properties of any fitness landscape is its *genotypic density*—the number of sequences near a given fitness value. Understanding this density is especially important near fitness peaks, as it quantifies the supply of high-fitness genotypes. Here I study the genotypic density of additive landscapes near fitness peaks. Although this density is well known to be approximately Gaussian near the middle of the fitness range, its behavior near maximal fitness has not been reported. I begin by deriving a saddle-point approximation that accurately describes the genotypic density of additive landscapes over virtually the entire fitness range. I then show that the log density follows a power law near maximal fitness, with an exponent determined by how much the best allele at each position outperforms its nearest competitor. This power-law behavior holds over a substantial fraction of fitness values, besting the Gaussian approximation on both simulated and empirical landscapes across roughly a quarter to a third of the fitness range. Under certain conditions this behavior also extends to globally epistatic landscapes (defined as nonlinear functions over one or more additive traits), though with a reduced range of validity. These findings advance our understanding of one of the most fundamental models of sequence-function relationships. In particular, they reveal that the uppermost reaches of Mount Fuji landscapes, rather than being sharply peaked, are actually quite stubby.

1 Introduction

Fitness landscapes—quantitative mappings from biological sequences to fitness values[1]—have long been a key organizing concept in evolutionary biology [1, 2, 3]. The simplest and most widely used model for such landscapes is the *additive landscape*, in which each position in a sequence contributes independently to fitness. Additive landscapes are used across multiple areas of biology, both in theoretical efforts and in empirical studies of sequence-function relationships. Indeed, many experimentally measured fitness landscapes either show approximately additive behavior or are well-described by nonlinear functions of one or more additive traits [4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15]. A thorough understanding of the quantitative properties of additive landscapes is therefore warranted.

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[1]I use the term *fitness* to refer to any scalar function of sequence, regardless of whether that function represents evolutionary pressure, biological activity, or some biophysical quantity.

One of the simplest questions one can ask about a fitness landscape is how many sequences have fitness near a given value. I refer to this quantity as *genotypic density.* In the case of additive landscapes, the Central Limit Theorem (CLT) suggests that the genotypic density is approximately Gaussian in the bulk of the fitness distribution. However, this bulk approximation is qualitatively wrong near the maximal achievable fitness. Although the importance of genotypic density has been recognized across several areas of the literature, the quantitative form of the near-peak density of additive fitness landscapes has not been reported.

In the fitness landscape literature, additive landscapes are sometimes known as *Mount Fuji* landscapes, a moniker chosen to emphasize their simple topographic structure: a single peak that is evolutionarily accessible from any starting sequence [16, 17, 18]. Although the Gaussian behavior of genotypic density in the bulk was realized early on in this context [16], the behavior near maximal fitness appears not to have been investigated. This is perhaps because additive landscapes have primarily been considered as null models against which to assess more complex epistatic models (including NK, Rough Mount Fuji, and House of Cards models) [19, 20, 21], rather than as objects of primary scientific interest. The near-peak genotypic density nevertheless has evolutionary implications: it quantifies the supply of high-fitness genotypes accessible by mutation, and therefore shapes both adaptive and purifying selection.

In quantitative genetics, additive models have long provided a theoretical foundation for the study of plant and animal breeding [22, 23]. Of particular importance is the infinitesimal model, which assumes that a trait is determined by the sum of effects across a large number of loci [24]. Both theory and data indicate that additive genetic contributions account for most of the heritable variation in complex traits, even when epistasis is present at individual loci [25]. The distribution of trait values in a population under selection has been studied extensively, including using concepts and methods from statistical physics [26, 27]. In particular, the departure of population trait distributions from Gaussianity has been explored [28]. Genotypic density itself, however, does not appear to have received a similar treatment.

In the bioinformatic study of gene regulation, additive landscapes are commonly used to model the binding of transcription factors (TFs) to DNA. Additive models such as position weight matrices are routinely used to scan genomes for putative TF binding sites [29, 30, 31, 32, 33, 34, 35], and large databases of such models are widely used [36, 37, 38, 39]. Assessing the statistical significance of high-scoring sites requires accurate estimates of the upper tail of the genotypic density, and a few studies have developed methods for doing so [40, 41, 42]. Of particular note is Madsen et al. [42], who developed an estimate of p-values for PWMs using a saddle-point approximation similar to the one I use below [43]. However, neither Madsen et al. [42] nor other work in this area has described the mathematical form of the near-peak density.

Beginning with foundational studies by Berg and von Hippel [44, 45], researchers at the interface of statistical physics and population genetics have used additive models for TF binding energy to study the evolution of TF binding sites [46, 47, 48, 32, 49]. Genotypic density plays an important role in this literature. When evolution is modeled in the weak-mutation limit, the equilibrium distribution of binding site sequences takes the form of a canonical ensemble from statistical physics, in which fitness values are weighted by an exponentially tilted form of the genotypic density [50, 51, 52, 53]. The original work of Berg and von Hippel derives the exact genotypic density for an idealized additive model in which every mutation away from the highest-fitness sequence produces the same fitness deficit. To treat more general additive models, Sengupta et al. [47] derived a saddle-point estimate of genotypic density (the same estimate I consider below), but used it only to recover the Gaussian approximation suggested by the CLT.

Here I study the genotypic density of additive landscapes near maximal fitness. I first derive a saddle-point approximation to this density and show that it is remarkably accurate across nearly

the entire fitness range. I then investigate the near-peak behavior of this approximation and find that, quite generally, the log density follows a power law. This means that, as fitness decreases from its maximum, the number of available genotypes initially expands very rapidly, then grows more and more slowly until finally it matches up with the predictions of the Gaussian approximation. The landscape is therefore not sharply peaked at its maximum but instead is broad and gently rounded—more like the top of a rolling hill than the summit of a mountain. This motivates my use of the term "stubby."

I validate these findings on simulated landscapes and on two empirical landscapes: one from a massively parallel reporter assay on a transcriptional promoter [5], and one from a deep mutational scanning experiment on a protein domain [54]. In all cases these landscapes exhibit stubby peaks of the predicted mathematical form. I then show that globally epistatic landscapes defined on one or more traits often exhibit similarly stubby peaks, though with caveats. The derivations of these results are provided below. Additional mathematical details are given in Supplemental Information (SI).

2 Results

2.1 Preliminaries

Consider sequences of length L comprised of characters from an alphabet of size C (e.g. $C = 4$ for DNA or RNA, $C = 20$ for proteins). Each sequence x is represented by a one-hot encoded vector $x = \{x_{lc}\}$, where $x_{lc} = 1$ if character c occurs at position l and $x_{lc} = 0$ otherwise. Additive fitness landscapes have the form

$$f(x) = \theta_0 + \sum_{l,c} \theta_{lc}\, x_{lc}, \tag{1}$$

where θ_{lc} quantifies the effect of character c at position l and θ_0 is an overall fitness baseline. In what follows I denote the maximal fitness effect at position l by $\theta_l^{\max} = \max_c \theta_{lc}$, the optimal character by $c_l^{\max}$, and the fitness-maximizing sequence by $x_{\max}$. The maximum achievable fitness is $F_{\max} = f(x_{\max})$. $F_{\min}$ denotes minimum achievable fitness.

Throughout this paper I refer to two empirical additive landscapes (Fig. 1). The first describes the transcriptional activity of variants of the *Escherichia coli lac* promoter ($L = 75$, $C = 4$), measured using a massively parallel reporter assay (MPRA) called Sort-Seq [5]. The second describes the binding affinity of variants of the B1 domain of protein G (GB1; $L = 55$, $C = 20$) to IgG, as measured using a deep mutational scanning (DMS) experiment based on RNA display [54]. Additive models were inferred for both data sets using MAVE-NN [55], which accounts for nonlinear experimental effects when determining the values of model parameters.

2.2 Defining the genotypic density

The focus of this paper is the quantitative form of the genotypic density, $\rho(F)$, which is defined so that $\rho(F)\, dF$ is the number of sequences within an infinitesimal interval $[F, F + dF]$. Formally, this density is written in terms of Dirac delta functions:

$$\rho(F) = \sum_x \delta(F - f(x)). \tag{2}$$

Below I develop various smooth mathematical approximations to $\rho(F)$. The reader might reasonably question what it means to approximate $\rho(F)$ with a smooth function, since no nonzero smooth function will provide a good approximation in the standard squared-error sense.

Figure 1: Additive effects for the two empirical landscapes analyzed in this paper. (a) Additive effects of the *lac* promoter landscape of Kinney et al. [5] ($L = 75$, $C = 4$), visualized as a sequence logo [56]. (b) Additive effects of the GB1 landscape of Olson et al. [54] ($L = 55$, $C = 20$), visualized as a heatmap. The parameters of both models were inferred from data using MAVE-NN [55].

Perhaps the most intuitive reason for considering smoothed approximations to $\rho(F)$ is that, in reality, fitness values are never measured with infinite precision. One way to account for this is to replace the delta function in Eq. 2 with a Gaussian distribution that has a standard deviation comparable to the uncertainty. Doing this yields a kernel density estimate that is smooth by construction. In practice, the form of this estimate is almost completely independent of the width of the Gaussian as long as the uncertainty scale is (a) much less than the fitness range $F_{\max} - F_{\min}$, and (b) much greater than the gaps between the extremal fitness values ($F_{\max}$ and $F_{\min}$) and their nearest neighbors. This condition is satisfied for a wide range of uncertainties, and almost always holds in practice.

2.3 The bulk approximation

In an additive landscape, the fitness of each sequence is a sum of L independent contributions. The CLT therefore suggests that these values are approximately normally distributed, and thus that $\rho(F)$ can be approximated by

$$\rho_{\text{bulk}}(F) = \frac{C^L}{\sqrt{2\pi\sigma_{\text{bulk}}^2}} \exp\left[-\frac{(F - F_{\text{mean}})^2}{2\sigma_{\text{bulk}}^2}\right], \tag{3}$$

where F_{mean} and σ_{bulk}^2 are the mean and variance of the fitness values observed for randomly generated sequences. The CLT, however, only guarantees accuracy within a few standard deviations of the mean. Indeed, Fig. 2 shows that this Gaussian form accurately describes the central mass of $\rho(F)$, but breaks down dramatically near $F_{\max}$. Specifically, $\rho_{\text{bulk}}(F)$ provides increasingly inflated estimates of $\rho(F)$ as F approaches $F_{\max}$ from below, and predicts nonzero density at all fitness values greater than $F_{\max}$, where the true density vanishes. The breakdown worsens as sequences grow longer. Assuming the distribution of θ_{lc} values remains fixed, the width of the fitness distribution scales as $\sigma_{\text{bulk}} \propto \sqrt{L}$, whereas $F_{\max} - F_{\text{mean}}$ grows linearly with L. The fitness peak therefore recedes to ever more extreme quantiles as L increases, placing it in a regime where the CLT provides no useful information. To characterize $\rho(F)$ across the full fitness range—and in particular near $F_{\max}$—a fundamentally different approach is required.

2.4 The tilted distribution and free fitness

To derive an improved estimate of $\rho(F)$, we first introduce a family of *exponentially tilted* distributions over sequence space:

$$p_\beta(x) = e^{\beta f(x) - \Phi(\beta)}, \tag{4}$$

where β is a scalar that parameterizes the family, and

$$\Phi(\beta) = \log \sum_x e^{\beta f(x)} \tag{5}$$

is a function that serves to normalize the distribution. By varying β from $-\infty$ to ∞ we can concentrate this distribution over sequences having specific fitness values. In particular, $\beta = 0$ recovers the uniform distribution on sequence space (which has fitness values concentrated near F_{mean}), while sufficiently large β concentrates the distribution near F_{max}.

Distributions of the form in Eq. 4 arise naturally in both population genetics and statistical physics. In the weak-mutation limit of Wright-Fisher processes with reversible mutation rates, $p_\beta(x)$ describes the stationary distribution over genotypes, where β is related to the effective population size via $\beta = 2N_e$ [50, 51, 53, 46, 49]. If instead we interpret x as the microstate of a physical system and $-f(x)$ as the corresponding energy, then $p_\beta(x)$ describes the distribution over microstates that arises when the system is at temperature $1/\beta$. In such contexts p_β is called the *canonical ensemble*.

The normalizing term $\Phi(\beta)$ encodes important information about the tilted distribution. Indeed, this quantity has previously been termed *free fitness* [57, 46, 53, 27], as it is analogous to negative the free energy in physical systems. One of the key properties is that $\Phi(\beta)$ is a cumulant-generating function for the marginal distribution of fitness values

$$p_\beta(F) = \sum_x p_\beta(x)\delta(F - f(x)). \tag{6}$$

In other words, the mean of $p_\beta(F)$ is $\mu_\beta = \Phi'(\beta)$, the variance is $\sigma_\beta^2 = \Phi''(\beta)$, and so on. This is true regardless of the functional form of $f(x)$.

In the specific case where $f(x)$ is additive (Eq. 1), free fitness can be written in closed form:

$$\Phi(\beta) = \beta\theta_0 + \sum_l \log \sum_c e^{\beta\theta_{lc}}. \tag{7}$$

Expressions for the mean and variance are given in SI Sec. S1. It is readily shown that the higher-order cumulants become irrelevant as L grows, and thus that $p_\beta(F)$ becomes well-approximated by the Gaussian distribution

$$p_\beta(F) = \frac{1}{\sqrt{2\pi\sigma_\beta^2}} \exp\left[-\frac{(F - \mu_\beta)^2}{2\sigma_\beta^2}\right]. \tag{8}$$

2.5 The saddle-point approximation

We now return to the problem of estimating the genotypic density $\rho(F)$ in Eq. 2. Since the weight for each sequence in the tilted distribution depends on x only through $f(x)$, it can be taken outside the sum in Eq. 6 to give

$$p_\beta(F) = e^{\beta F - \Phi(\beta)}\rho(F). \tag{9}$$

We now have two distinct expressions for $p_\beta(F)$, Eq. 8 and Eq. 9. Equating these and solving for $\rho(F)$ yields the candidate approximation for genotypic density

$$\rho_\beta(F) = \frac{e^{\Phi(\beta)-\beta F}}{\sqrt{2\pi\sigma_\beta^2}} \exp\left[-\frac{(F-\mu_\beta)^2}{2\sigma_\beta^2}\right].$$ (10)

We are free to choose β however we like, but we expect that $\rho_\beta(F)$ will be a good approximation only when F is within a few standard deviations σ_β of the mean μ_β. Indeed, Eq. 10 reduces to the bulk approximation Eq. 3 if we set $\beta = 0$, and the entire motivation for this paper is that $\rho_{\text{bulk}}(F)$ is a poor estimate of the true density $\rho(F)$ when F is near F_{max}.

The CLT suggests that the approximation in Eq. 8 is most accurate when $\mu_\beta = F$. We therefore choose β to be a function of F defined implicitly by this requirement, i.e.,

$$\rho_{\text{saddle}}(F) = \frac{e^{\Phi(\beta)-\beta F}}{\sqrt{2\pi\sigma_\beta^2}} \quad \text{where } \beta \text{ is s.t. } \Phi'(\beta) = F.$$ (11)

This is known as the *saddle-point* approximation [58, 59, 43, 60]. The name comes from an alternative derivation in which one approximates the value of a contour integral in the complex plane by its value near a stationary point of the integrand (i.e., a saddle-point). The above derivation, however, more clearly explains why the saddle-point approximation works so well throughout the entire fitness range. See Reid [43] for further discussion of these two derivations.

2.6 Performance of the saddle-point approximation

To assess the performance of ρ_{saddle}, I compared its predictions to the true densities of two simulated landscapes and of the two empirical landscapes mentioned earlier. The results are shown in Fig. 2. In all cases ρ_{saddle} closely tracks ρ over the entire fitness range. By contrast, ρ_{bulk} progressively overestimates ρ as F approaches F_{max}, ultimately deviating by many orders of magnitude. This agreement holds equally for simulated and empirical landscapes over both DNA ($C = 4$) and protein ($C = 20$) alphabets.

Computing the true density ρ is complicated by the fact that all four landscapes in Fig. 2 are too large to exhaustively enumerate all fitness values. I therefore used the dynamic programming algorithm of Touzet and Varré [41] instead. This algorithm computes cumulative counts above a given threshold by iteratively refining upper and lower bounds using discretized fitness values. Running this algorithm to full precision across the entire fitness range is prohibitive. Restricting it to the upper 30% of the range and terminating refinement before full convergence, however, yields certified upper and lower bounds that are visually indistinguishable in Fig. 2.

2.7 Near-peak scaling for binary alphabets

Eq. 11 does not by itself reveal the mathematical form of genotypic density near maximal fitness. What we require is an explicit expression for $\rho(F)$ near F_{max}, one that is not obscured by the implicit dependence of β on F. To this end, we express the free fitness as $\Phi(\beta) = \beta F_{\text{max}} + LE(\beta)$, where

$$E(\beta) = \frac{1}{L}\sum_{l=1}^{L}\log\left(1+\sum_{c\neq c_l^{\text{max}}}e^{-\beta\delta_{lc}}\right), \quad \delta_{lc} = \theta_l^{\text{max}} - \theta_{lc}.$$ (12)

Figure 2: Accuracy of the saddle-point approximation on simulated and empirical landscapes. (a,b) Results for simulated landscapes having Gaussian fitness effects θ_{lc}. (c) Results for the *lac* promoter landscape of Kinney et al. [5]. (d) Results for the GB1 landscape of Olson et al. [54]. Each panel compares the saddle-point approximation ρ_{saddle} (orange solid line) to both the Gaussian approximation ρ_{bulk} (olive dashed line) and the true density ρ_{true} (black solid line) estimated by the algorithm of Touzet and Varré [41]. Vertical dotted lines indicate F_{max} and F_{mean}.

We call this the *per-site deficit contribution* to free fitness, and define the *per-site fitness deficit* as $\epsilon = (F_{\text{max}} - F)/L$. Substituting these into Eq. 11 and dropping the prefactor (which is sub-leading for large L) gives

$$\frac{1}{L} \log \rho_{\text{saddle}} \approx \beta\epsilon + E(\beta), \qquad (13)$$

where β is an implicit function of ϵ determined by $E'(\beta) = -\epsilon$. Since every $\delta_{lc} > 0$ in Eq. 12, $E(\beta) \to 0$ as $\beta \to \infty$. However, the near-peak behavior of ρ_{saddle} depends on precisely how $E(\beta)$ approaches this limit. We therefore seek to determine the leading-order behavior of $E(\beta)$ as $\beta \to \infty$.

To simplify this analysis, we first consider binary alphabets ($C = 2$). In this case, the deficit contribution becomes

$$E_{\text{bin}}(\beta) = \frac{1}{L} \sum_{l=1}^{L} \log\left(1 + e^{-\beta\Delta_l}\right), \qquad (14)$$

where Δ_l is the (positive) fitness gap between the two additive effects at position l. Replacing the empirical distribution of these gaps with a smooth density $p_{\text{gap}}(\Delta)$ gives

$$E_{\text{bin}}(\beta) \approx \int \log\left(1 + e^{-\beta\Delta}\right) p_{\text{gap}}(\Delta)\, d\Delta = \frac{1}{\beta} \int K(s)\, p_{\text{gap}}\left(\tfrac{s}{\beta}\right) ds, \qquad (15)$$

where we have substituted $s = \beta\Delta$ and defined $K(s) = \log(1 + e^{-s})$. It is helpful to think of $K(s)$ as a smoothing kernel: it equals $\log 2$ at the origin, has width of order 1, decays as e^{-s} for $s \gg 1$, and has area $\frac{\pi^2}{12}$ (see SI Sec. S2.1). Since larger β probes $p_{\text{gap}}(\frac{s}{\beta})$ at ever-smaller arguments, only the behavior of $p_{\text{gap}}(\Delta)$ near $\Delta = 0$ matters.

When p_{gap} is *regular* (by this I mean $0 < p_{\text{gap}}(0) < \infty$), Eq. 15 can be evaluated to leading order:

$$E_{\text{bin}}(\beta) \approx \frac{\pi^2 p_{\text{gap}}(0)}{12\beta} + O(\beta^{-2}). \qquad (16)$$

The saddle-point condition then gives $\epsilon = \frac{\pi^2 p_{\text{gap}}(0)}{12\beta^2}$, which inverts to $\beta = \sqrt{\frac{\pi^2 p_{\text{gap}}(0)}{12\epsilon}}$. Substituting Eq. 16 and this expression for β back into Eq. 13 yields our desired result:

$$\frac{1}{L}\log\rho \approx B\epsilon^{1/2}, \quad \text{where} \quad B = \sqrt{\frac{\pi^2 p_{\text{gap}}(0)}{3}}. \qquad (17)$$

It is tempting to assume that p_{gap} is always regular, and indeed it is if each θ_{lc} parameter is drawn i.i.d. from a sufficiently well-behaved distribution (see SI Sec. S3). Regularity is not guaranteed, however, when the magnitude of additive effects varies across positions. For example, if most positions in a sequence have little to no effect, the gap distribution may scale as Δ^γ near $\Delta = 0$ for some negative exponent γ.[2] This is an important case to consider because many empirical landscapes exhibit this type of behavior, and major deviations from the predicted exponent $\frac{1}{2}$ are often observed in such landscapes.

We therefore consider the more general case where $p_{\text{gap}}(\Delta) \sim c\Delta^\gamma$ near $\Delta = 0$, for some exponent $\gamma > -1$ and coefficient $c > 0$. Repeating the asymptotic analysis with this assumption yields a more general power law:

$$\frac{1}{L}\log\rho \approx B\,\epsilon^\alpha, \quad \text{where} \quad \alpha = \frac{1 + \gamma}{2 + \gamma}, \qquad (18)$$

and B is a constant that depends on γ and c; see SI Sec. S2 for a full derivation. Note that the exponent α increases monotonically with γ. For regular gap distributions ($\gamma = 0$), one recovers $\sqrt{\epsilon}$ scaling ($\alpha = \frac{1}{2}$). When the gap distribution is enriched for small gaps ($\gamma < 0$), as is expected when most positions have small fitness effects, the density falls off more quickly near F_{max} than $\sqrt{\epsilon}$ scaling predicts, producing a stubbier peak ($0 < \alpha < \frac{1}{2}$). Conversely, a deficit of small gaps ($\gamma > 0$) causes the density to fall off more slowly, producing a sharper peak ($\frac{1}{2} < \alpha < 1$).

In the extreme case where all gaps are bounded below by some $\Delta_0 > 0$, $\log\rho$ grows approximately linearly with ϵ (i.e., $\alpha = 1$), but is also tempered by a log-linear term in ϵ that reduces this growth away from $\epsilon = 0$. This is exemplified by the landscape proposed by [44], in which $F_{\text{max}} - f(x)$ is Δ_0 times the Hamming distance between x and x_{max}, and consequently $p_{\text{gap}}(\Delta) = \delta(\Delta - \Delta_0)$. See SI Sec. S4 for details.

Figure 3 validates these predictions for binary landscapes with four choices of gap distribution: regular ($\gamma = 0$), enriched for small gaps ($\gamma < 0$), depleted of small gaps ($\gamma > 0$), and fixed gap size (Hamming-distance landscape). In all cases, the near-peak approximation closely tracks ρ_{saddle} near F_{max}, while ρ_{bulk} progressively overestimates the density.

2.8 Near-peak scaling for general alphabets

The preceding analysis was restricted to binary alphabets, where each position contributes a single fitness gap Δ_l to $E(\beta)$. This allowed $E(\beta)$ to be evaluated in the large-L limit by replacing the

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[2]We require $\gamma > -1$ in order for p_{gap} to be normalizable.

Figure 3: Near-peak scaling for additive landscapes on binary sequences ($C = 2$, $L = 200$). Each panel compares the saddle-point approximation ρ_{saddle} (orange solid line) to the Gaussian approximation ρ_{bulk} (olive dashed line) and the near-peak approximation ρ_{peak} (blue dashed line). (a) Gaussian fitness effects ($\theta_{lc} \sim \text{N}(0, 1)$), producing a regular gap distribution with $\gamma = 0$ and $\alpha = 1/2$. (b) Left-skewed gap density ($p_{\text{gap}} \propto \Delta^{-1/2}$, $\gamma = -1/2$), enriched for small gaps and producing a stubbier peak ($\alpha = 1/3$). (c) Right-skewed gap density ($p_{\text{gap}} \propto \Delta$, $\gamma = 1$), depleted of small gaps and producing a sharper peak ($\alpha = 2/3$). (d) Hamming-distance landscape with uniform gap $\Delta = 1$ at all positions ($\alpha = 1$ with log-linear correction). Vertical dotted lines indicate F_{max} and F_{mean}.

sum over positions with an integral against the gap density $p_{\text{gap}}(\Delta)$. For alphabets with $C \geq 3$ characters, however, $E(\beta)$ in Eq. 12 includes contributions from multiple non-optimal characters at each position. The resulting multidimensional integral cannot be evaluated so easily.

Fortunately, we can bound $E(\beta)$ both above and below using the binary alphabet results from Sec. 2.7. Define Δ_l to be the gap between the additive effects of the optimal and runner-up characters at position l, and let $E_{\text{bin}}(\beta)$ be the deficit contribution obtained by restricting the alphabet to only these two characters. For the lower bound, this restriction removes terms from the sum inside the logarithm in Eq. 12, which can only decrease $E(\beta)$, giving $E_{\text{bin}}(\beta) \leq E(\beta)$. For the upper bound, we use $\Delta_l \leq \delta_{lc}$ for all $c \neq c_l^{\text{max}}$, together with the subadditivity of $\log(1 + x)$, to get

$$E(\beta) \leq \frac{1}{L} \sum_l \log\left[1 + (C-1)\, e^{-\beta \Delta_l}\right] \leq \frac{C-1}{L} \sum_l \log\left[1 + e^{-\beta \Delta_l}\right]. \tag{19}$$

We thus obtain the sandwich bound,

$$E_{\text{bin}}(\beta) \le E(\beta) \le (C-1)\, E_{\text{bin}}(\beta). \tag{20}$$

This implies that $E(\beta)$ exhibits the same asymptotic behavior in the $\beta \to \infty$ limit as its binary alphabet counterpart. In particular, when the runner-up gap distribution satisfies $p_{\text{gap}}(\Delta) \sim c\,\Delta^\gamma$ near the origin, the scaling exponent $\alpha = (1+\gamma)/(2+\gamma)$ carries over unchanged from the binary case, though the constant B may differ.

There is one additional consideration. If all C characters at a position make the same contribution to fitness ($\delta_{lc} = 0$ for all c), that position contributes a constant $\frac{1}{L}\log C$ to $E(\beta)$ independent of β. The cumulative effect of such positions is to add a constant to $\frac{1}{L}\log\rho$. Similar but smaller contributions arise at positions where some but not all characters are tied for maximal fitness. These considerations motivate the following scaling form for the near-peak log genotypic density of additive landscapes:

$$\frac{1}{L}\log\rho \;\approx\; A + B\,\epsilon^\alpha, \tag{21}$$

for constants A and B and exponent $0 \le \alpha \le 1$. In practice, I recommend fitting α, A, and B directly to each landscape of interest. As we will see below, this approach provides an accurate description of the near-peak density across a range of simulated and empirical landscapes.

Figure 4 tests this scaling form on the same four landscapes considered in Fig. 2: two simulated (i.i.d. Gaussian effects with $L = 100$, $C = 4$ and $L = 100$, $C = 20$) and two empirical (*lac* promoter and GB1 protein). For each landscape, I fit $L^{-1}\log\rho = A + B\,\epsilon^\alpha$ to the saddle-point density in the near-peak region, with A, B, and α as free parameters. The two simulated landscapes give $\alpha \approx 0.53$, close to $1/2$, consistent with the theoretical prediction for i.i.d. fitness effects. The two empirical landscapes give $\alpha \approx 0.40$, slightly below $1/2$, consistent with a mild enrichment of small gaps.

It is useful to ask over what fraction of the fitness range the near-peak scaling outperforms the Gaussian approximation. To quantify this, I define the *crossover fitness* F_{cross} as the fitness value at which ρ_{peak} becomes a closer approximation to ρ_{saddle} than ρ_{bulk} is. The *crossover ratio* $r = (F_{\text{max}} - F_{\text{cross}})/(F_{\text{max}} - F_{\text{mean}})$ measures the fraction of the fitness range (from F_{mean} to F_{max}) over which the near-peak scaling dominates. Because both $\log\rho_{\text{bulk}}$ and $\log\rho_{\text{peak}}$ scale linearly with L, the fitness at which they cross is determined by an L-independent equation, and so the crossover ratio r is independent of sequence length. Figure 4 marks F_{cross} for each of the four landscapes. Across these landscapes, r ranges from 0.26 to 0.34, indicating that the near-peak scaling governs roughly a quarter to a third of the fitness range.

2.9 Single-trait global epistasis models

The results above apply to purely additive landscapes. In many experimental settings, however, the observed fitness is better described not as a direct sum of per-position effects but rather as a nonlinear readout of an underlying additive trait. More precisely, the measured fitness takes the form $F = g(\phi)$, where $\phi(x) = \theta_0 + \sum_{l,c}\theta_{lc}\,x_{lc}$ is an additive trait and g is a nonlinear function. Such models are commonly referred to as global epistasis models [61, 10, 12, 62][3]. I now show that, when the fitness maximum occurs at an extremal value of the additive trait and the slope of g is nonzero there, the near-peak scaling exponent α is preserved under such a nonlinear transformation, with only the coefficients A and B modified.

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[3]Note that this definition of global epistasis is based on the structure of the underlying equations and is distinct from the phenomenological definition sometimes used in discussions of diminishing returns epistasis [6, 63]

Figure 4: Near-peak scaling for general alphabets, tested on the same four landscapes as Fig. 2. Each panel plots $\log \rho(F)$ versus F, comparing the saddle-point approximation ρ_{saddle} (orange solid line), the Gaussian approximation ρ_{bulk} (olive dashed line), and the near-peak scaling form ρ_{peak} (blue dashed line), obtained by fitting $L^{-1} \log \rho = A + B\,\epsilon^\alpha$ to each landscape. The vertical dotted line at F_{cross} marks the crossover fitness at which ρ_{peak} becomes a closer approximation to ρ_{saddle} than ρ_{bulk}, with the crossover ratio r labeled. Vertical dotted lines indicate F_{mean} and F_{max}. (a) Simulated, $L = 100$, $C = 4$. (b) Simulated, $L = 100$, $C = 20$. (c) *lac* promoter ($L = 75$, $C = 4$). (d) GB1 protein ($L = 55$, $C = 20$).

The key observation is that, sufficiently close to the fitness peak, g is well-approximated by its linearization about ϕ_{max}. Under this linearization, the fitness deficit at each sequence is simply a rescaled version of its trait deficit: $F_{\text{max}} - F \approx (\phi_{\text{max}} - \phi) g'_{\text{max}}$, where $g'_{\text{max}} = g'(\phi_{\text{max}})$. Because this rescaling is linear, it preserves the power-law form of the gap distribution near zero and therefore preserves the scaling exponent α.

To state this more precisely, recall that the near-peak scaling of the additive trait (Eq. 21) can be written as

$$\frac{1}{L} \log \rho_\phi \approx A_\phi + B_\phi\,\epsilon_\phi^\alpha, \tag{22}$$

where $\epsilon_\phi = (\phi_{\text{max}} - \phi)/L$ is the per-site trait deficit. Applying the linearization to transform from trait space to fitness space, and defining the per-site fitness deficit $\epsilon_F = (F_{\text{max}} - F)/L$, gives (see SI Sec. S5 for derivation):

$$\frac{1}{L} \log \rho_F \approx A_F + B_F\,\epsilon_F^\alpha, \tag{23}$$

Figure 5: Scaling under global epistasis. A simulated additive landscape ($L = 100$, $C = 4$, Gaussian θ_{lc}) is transformed by sigmoid nonlinearities $g(\phi)$ of varying steepness. (a) The log genotypic density $\log \rho_\phi$ in trait space versus ϕ, showing the saddle-point approximation (orange solid line) and the near-peak approximation (blue dashed line). Vertical dotted lines indicate $\phi_{\max}$ (red) and ϕ_{mean} (green). (b) The same quantities plotted against $\sqrt{\epsilon_\phi}$, showing the region of approximate linearity from which B_ϕ is extracted. (c) Three sigmoid nonlinearities with midpoint $\phi_{1/2} = \phi_{\max} - 4\sigma$ (gray dotted line) and varying span parameter s_ϕ, which controls steepness. (d) The transformed density $\log \rho_F$ versus $\sqrt{\epsilon_F}$ for each sigmoid (solid lines) compared to the theoretical prediction $A_F + B_F\sqrt{\epsilon_F}$ (dashed lines in matching colors), where A_F and B_F are computed analytically from A_ϕ, B_ϕ, and $g'_{\max}$ with no additional fitting (see SI Sec. S5).

where $A_F = A_\phi - \frac{1}{L} \log g'_{\max}$ and $B_F = (g'_{\max})^{-\alpha} B_\phi$. This approximation holds as long as the linearization of g remains accurate over the fitness range of interest. In practice, the flatter the nonlinearity is at $\phi_{\max}$, the more heavily it compresses the range over which near-peak scaling applies.

Figure 5 illustrates these predictions on a simulated landscape ($L = 100$, $C = 4$) subjected to sigmoid nonlinearities of varying steepness. The coefficients A_F and B_F are computed analytically from the scaling behavior of the additive trait ϕ and from the value of $g'_{\max}$, with no additional fitting. In all cases the predicted $\sqrt{\epsilon_F}$ scaling closely matches the exact transformed density, confirming that the near-peak exponent is preserved under global epistasis. As expected, sigmoids that are flatter at $\phi_{\max}$ (due to greater saturation) yield a larger B_F and a more compressed range of validity.

2.10 Multi-trait global epistasis models

The preceding section considered fitness landscapes defined by a nonlinear function of a single additive trait. More generally, however, fitness may depend on multiple additive traits simultaneously. I now show that the saddle-point machinery and the near-peak scaling results derived above extend naturally to this multi-trait setting. Consider K additive traits

$$\phi_k(x) = \theta_{0k} + \sum_{l,c} \theta_{lck} \, x_{lc}, \qquad k = 1, \ldots, K, \tag{24}$$

Taken together, these map each sequence x to a K-dimensional trait vector $\vec{\phi}(x) = (\phi_1, \ldots, \phi_K)$. To define a tilted distribution, we introduce a complementary vector of K inverse temperatures $\vec{\beta} = (\beta_1, \ldots, \beta_K)$. The resulting tilted distribution has the form $p_{\vec{\beta}}(x) \propto e^{\vec{\beta} \cdot \vec{\phi}(x)}$. The key observation is that the exponent $\vec{\beta} \cdot \vec{\phi}(x)$ is itself an additive function of x, with effective per-position parameters $\tilde{\theta}_{lc} = \sum_k \beta_k \, \theta_{lck}$. The tilted distribution therefore factorizes across positions exactly as in the single-trait case. The corresponding cumulant-generating function is

$$\Phi(\vec{\beta}) = \sum_l \log \sum_c \exp\left(\sum_k \beta_k \, \theta_{lck} \right). \tag{25}$$

Its gradient $\nabla \Phi$ gives the mean of each trait under the tilted distribution, and the entries of the Hessian matrix Φ'' give the covariances.

The saddle-point approximation generalizes directly [60, Sec. 6.5]. As in the single-trait case, we choose $\vec{\beta}$ so that the target trait vector $\vec{\phi}$ coincides with the mean of the tilted distribution. Because Φ is strictly convex, this condition uniquely determines $\vec{\beta}$. The resulting genotypic density, which is joint across K traits, is

$$\rho_{\text{saddle}}(\vec{\phi}) = \frac{e^{\Phi(\vec{\beta}) - \vec{\beta} \cdot \vec{\phi}}}{(2\pi)^{K/2} \, |\Phi''(\vec{\beta})|^{1/2}}, \quad \text{where} \quad \nabla \Phi(\vec{\beta}) = \vec{\phi}, \tag{26}$$

and where $|\Phi''|$ denotes the determinant of the Hessian. This reduces to Eq. 11 when $K = 1$.

The near-peak scaling derived for single-trait global epistasis (Sec. 2.9) also extends to the multi-trait setting. Suppose fitness is a nonlinear function of the trait vector, $F = g(\vec{\phi})$. The achievable trait vectors form a bounded region in K-dimensional trait space. If g attains its maximum F_{max} on the boundary of this region, and has nonzero gradient there, then g can be linearized in the vicinity of the maximum. This linearization projects the K traits onto a single effective additive trait, $\tilde{\phi} = \sum_k c_k \, \phi_k$, where $\vec{c} = \nabla g(\vec{\phi}_{\text{max}})$. The near-peak scaling results from the preceding sections then apply directly, with the exponent α determined by the gap distribution of $\tilde{\phi}$. As in the single-trait case, the range of fitness values over which this scaling holds may be reduced by the nonlinearity of g.

3 Discussion

The core finding of this work is that the genotypic density of additive fitness landscapes follows an exponentiated power law near maximal fitness (see Eq. 21). The exponent of the power law, α, determines the shape of this density near the peak: α is constrained to lie between 0 and 1, with larger α producing sharper peaks and smaller α producing stubbier ones. The value of α depends on how the fitness gaps between optimal and runner-up characters at each position are

distributed near zero, and thus varies from landscape to landscape. Notably, α is independent of both sequence length and alphabet size. In idealized landscapes, where the additive effects are drawn from a single distribution, one obtains $\alpha = \frac{1}{2}$ regardless of the specific distribution. In more realistic situations, however, the scale of additive effects can vary substantially from position to position, causing α to deviate from $\frac{1}{2}$. In particular, one should expect to find $\alpha < \frac{1}{2}$ (increased stubbiness) on most empirical landscapes because positions with small-to-negligible effects typically dominate over positions with large effects.

These results, to my knowledge, provide the first quantitative description of near-peak abundance of genotypes for any class of fitness landscapes.[4] Indeed, the Gaussian approximation suggested by the CLT fundamentally breaks down near the fitness maximum of additive landscapes. If taken literally, it predicts that many sequences lie far above $F_{\max}$. If the Gaussian approximation is instead truncated at maximal fitness, it still predicts a vast overabundance of high-fitness sequences, often yielding densities that are off by many orders of magnitude. By contrast, the exponentiated power law that I describe is highly accurate in this regime. It reveals that, as fitness is decreased from its maximum, the number of available genotypes initially expands at a near-infinite rate, then grows increasingly slowly until it settles down to the growth rate predicted by the Gaussian approximation. The result is a broad and gently rounded (i.e., stubby) summit.

The power law given in Eq. 21 comes with several caveats. First, it applies only to additive and globally epistatic landscapes. The consequences of specific epistatic interactions on the near-peak density have yet to be investigated. Second, the derivation assumes that sequence length L is large. Anecdotally, however, this scaling behavior is often observed even for landscapes describing TF binding motifs ($L \sim 10$). Third, the derivation assumes that the distribution of position-specific fitness gaps behaves as a power law near zero. That said, Eq. 21 appears to provide a good empirical fit to many additive landscapes, even when α, A, and B must be fit empirically rather than derived from the gap distribution. Finally, the scaling behavior extends to globally epistatic landscapes only when the nonlinearity attains its maximum on the boundary of achievable trait values and does so with nonzero gradient. And even when these conditions are satisfied, the behavior is often valid over a substantially reduced range of fitness values.

Looking forward, the mathematical methods used in this work might prove useful for addressing other questions about fitness landscapes. The distribution of fitness effects (DFE) is of central importance in evolutionary genetics [64, 65], and these methods could illuminate how the DFE behaves near peak fitness. Related questions, such as the distribution of Hamming distances among sequences at a given fitness level, could also be addressed. Finally, the multi-trait saddle-point approximation in Eq. 26 naturally suggests applications to Fisher's geometric model [66, 67]. It would be particularly interesting, in the spirit of Hwang et al. [68], to see if similar techniques could shed light on how the dramatic variation in genotypic density that occurs across the fitness range affects the predictions of Fisher's model.

4   Acknowledgments

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[4]That is, apart from the highly idealized Hamming-distance landscape studied by Berg and von Hippel [44].

5 Data availability

All the code used to carry out this analysis and generate the figures is publicly available at `https://github.com/jbkinney/26_stubby`.

References

[1] Sewall Wright. The roles of mutation, inbreeding, crossbreeding, and selection in evolution. *Proceedings of the Sixth International Congress of Genetics*, 1:356–366, 1932.

[2] John Maynard Smith. Natural selection and the concept of a protein space. *Nature*, 225: 563–564, 1970.

[3] Inês Fragata, Alexandre Blanckaert, Marco António Dias Louro, David A. Liberles, and Claudia Bank. Evolution in the light of fitness landscape theory. *Trends in Ecology & Evolution*, 34(1):69–82, 2019. doi: 10.1016/j.tree.2018.10.009.

[4] Maurício Carneiro and Daniel L. Hartl. Adaptive landscapes and protein evolution. *Proceedings of the National Academy of Sciences*, 107(suppl. 1):1747–1751, 2010. doi: 10.1073/pnas. 0906192106.

[5] Justin B. Kinney, Anand Murugan, Curtis G. Callan, and Edward C. Cox. Using deep sequencing to characterize the biophysical mechanism of a transcriptional regulatory sequence. *Proceedings of the National Academy of Sciences*, 107(20):9158–9163, 05 2010. ISSN 0027-8424. doi: 10.1073/pnas.1004290107.

[6] Sergey Kryazhimskiy, Daniel P. Rice, Elizabeth R. Jerison, and Michael M. Desai. Global epistasis makes adaptation predictable despite sequence-level stochasticity. *Science*, 344(6191): 1519–1522, 2014. doi: 10.1126/science.1250939.

[7] Michael A. Stiffler, Doeke R. Hekstra, and Rama Ranganathan. Evolvability as a function of purifying selection in TEM-1 β-lactamase. *Cell*, 160(5):882–892, 2015. doi: 10.1016/j.cell. 2015.01.035.

[8] Karen S. Sarkisyan, Dmitry A. Bolotin, Margarita V. Meer, Dinara R. Usmanova, Alexander S. Mishin, George V. Sharonov, Dmitry N. Ivankov, Nina G. Bozhanova, Mikhail S. Baranov, Onuralp Soylemez, Natalya S. Bogatyreva, Peter K. Vlasov, Evgeny S. Egorov, Maria D. Logacheva, Alexey S. Kondrashov, Dmitry M. Chudakov, Ekaterina V. Putintseva, Ilgar Z. Mamedov, Dan S. Tawfik, Konstantin A. Lukyanov, and Fyodor A. Kondrashov. Local fitness landscape of the green fluorescent protein. *Nature*, 533(7603):397–401, 2016. doi: 10.1038/ nature17995.

[9] Rhys M Adams, Thierry Mora, Aleksandra M Walczak, and Justin B Kinney. Measuring the sequence-affinity landscape of antibodies with massively parallel titration curves. *eLife*, 5: e23156, 12 2016. doi: 10.7554/elife.23156.

[10] Zachary R Sailer and Michael J Harms. Detecting high-order epistasis in nonlinear genotype-phenotype maps. *Genetics*, 205(3):1079 – 1088, 2017. ISSN 0016-6731. doi: 10.1534/genetics. 116.195214.

[11] Jakub Otwinowski. Biophysical inference of epistasis and the effects of mutations on protein stability and function. *Mol Biol Evol*, 35(10):2345 – 2354, 10 2018. ISSN 0737-4038. doi: 10.1093/molbev/msy141.

[12] Jakub Otwinowski, David M McCandlish, and Joshua B Plotkin. Inferring the shape of global epistasis. *Proc Natl Acad Sci USA*, 08 2018. ISSN 0027-8424. doi: 10.1073/pnas.1804015115.

[13] Pablo Baeza-Centurion, Belén Miñana, Jörn M. Schmiedel, Juan Valcárcel, and Ben Lehner. Combinatorial genetics reveals a scaling law for the effects of mutations on splicing. *Cell*, 176 (3):549–563.e23, 2019. doi: 10.1016/j.cell.2018.12.010.

[14] Tyler N. Starr, Allison J. Greaney, Sarah K. Hilton, Daniel Ellis, Katharine H.D. Crawford, Adam S. Dingens, Mary Jane Navarro, John E. Bowen, M. Alejandra Tortorici, Alexandra C. Walls, Neil P. King, David Veesler, and Jesse D. Bloom. Deep mutational scanning of SARS-CoV-2 receptor binding domain reveals constraints on folding and ACE2 binding. *Cell*, 182 (5):1295–1310.e20, 2020. ISSN 0092-8674. doi: 10.1016/j.cell.2020.08.012.

[15] Andre J. Faure, Aina Martí-Aranda, Cristina Hidalgo-Carcedo, Antoni Beltran, Jörn M. Schmiedel, and Ben Lehner. The genetic architecture of protein stability. *Nature*, 634(8035): 995–1003, 2024. doi: 10.1038/s41586-024-07966-0.

[16] Takuyo Aita and Yuzuru Husimi. Fitness spectrum among random mutants on mt. fuji-type fitness landscape. *Journal of Theoretical Biology*, 182(4):469–485, 1996. ISSN 0022-5193. doi: 10.1006/jtbi.1996.0189.

[17] Takuyo Aita and Yuzuru Husimi. Adaptive walks by the fittest among finite random mutants on a mt. fuji-type fitness landscape II. effect of small non-additivity. *Journal of Mathematical Biology*, 41(3):207–231, 2000. ISSN 0303-6812. doi: 10.1007/s002850000046.

[18] Takuyo Aita, Shunichi Morinaga, and Yuzuru Husimi. Thermodynamical interpretation of evolutionary dynamics on a fitness landscape in a evolution reactor, I. *Bulletin of Mathematical Biology*, 66:1371–1403, 2004. doi: 10.1016/j.bulm.2004.01.004.

[19] Ivan G. Szendro, Martijn F. Schenk, Jasper Franke, Joachim Krug, and J. Arjan G. M. de Visser. Quantitative analyses of empirical fitness landscapes. *Journal of Statistical Mechanics: Theory and Experiment*, 2013:P01005, 2013. doi: 10.1088/1742-5468/2013/01/P01005.

[20] Johannes Neidhart, Ivan G. Szendro, and Joachim Krug. Adaptation in tunably rugged fitness landscapes: the rough Mount Fuji model. *Genetics*, 198:699–721, 2014. doi: 10.1534/genetics. 114.167668.

[21] Sakshi Pahujani and Joachim Krug. Complexity and accessibility of random landscapes. *SciPost Physics Lecture Notes*, 108, 2025. doi: 10.21468/SciPostPhysLectNotes.108.

[22] Bruce Walsh and Michael Lynch. *Evolution and Selection of Quantitative Traits*. Oxford University Press, 2018. ISBN 978-0-19-883087-0. doi: 10.1093/oso/9780198830870.001.0001.

[23] Nicholas H. Barton and Peter D. Keightley. Understanding quantitative genetic variation. *Nature Reviews Genetics*, 3:11–21, 2002. doi: 10.1038/nrg700.

[24] N. H. Barton, A. M. Etheridge, and A. Véber. The infinitesimal model: Definition, derivation, and implications. *Theoretical Population Biology*, 118:50–73, 2017. doi: 10.1016/j.tpb.2017. 06.001.

[25] William G. Hill, Michael E. Goddard, and Peter M. Visscher. Data and theory point to mainly additive genetic variance for complex traits. *PLoS Genetics*, 4(2):e1000008, 2008. doi: 10.1371/journal.pgen.1000008.

[26] N. H. Barton and J. B. Coe. On the application of statistical physics to evolutionary biology. *Journal of Theoretical Biology*, 259:317–324, 2009. doi: 10.1016/j.jtbi.2009.03.019.

[27] N. H. Barton and H. P. de Vladar. Statistical mechanics and the evolution of polygenic quantitative traits. *Genetics*, 181:997–1011, 2009. doi: 10.1534/genetics.108.099309.

[28] Michael Turelli and N. H. Barton. Genetic and statistical analyses of strong selection on polygenic traits: what, me normal? *Genetics*, 138:913–941, 1994. doi: 10.1093/genetics/138.3.913.

[29] Gary D Stormo. DNA binding sites: representation and discovery. *Bioinformatics*, 16(1):16 – 23, 01 2000. ISSN 1367-4803. doi: 10.1093/bioinformatics/16.1.16. Read (date unknown) Read all the way through.

[30] G Stormo and D Fields. Specificity, free energy and information content in protein-DNA interactions. *Trends Biochem Sci*, 23(3):109 – 113, 1998.

[31] B Foat, A Morozov, and H Bussemaker. Statistical mechanical modeling of genome-wide transcription factor occupancy data by MatrixREDUCE. *Bioinformatics*, 22(14):e141 – 9, 07 2006. ISSN 1367-4803. doi: 10.1093/bioinformatics/btl223.

[32] Marko Djordjevic, Anirvan M. Sengupta, and Boris I. Shraiman. A biophysical approach to transcription factor binding site discovery. *Genome Research*, 13(11):2381–2390, 2003. doi: 10.1101/gr.1271603.

[33] Justin B. Kinney, Gašper Tkačik, and Curtis G. Callan. Precise physical models of protein–DNA interaction from high-throughput data. *Proceedings of the National Academy of Sciences*, 104(2):501–506, 01 2007. ISSN 0027-8424. doi: 10.1073/pnas.0609908104. Wrote.

[34] O Elemento, N Slonim, and S Tavazoie. A universal framework for regulatory element discovery across all genomes and data types. *Mol Cell*, 28(2):337 – 350, 10 2007. ISSN 1097-2765. doi: 10.1016/j.molcel.2007.09.027.

[35] Charles E Grant, Timothy L Bailey, and William Stafford Noble. FIMO: scanning for occurrences of a given motif. *Bioinformatics*, 27(7):1017 – 1018, 04 2011. ISSN 1367-4803. doi: 10.1093/bioinformatics/btr064.

[36] Damla Ovek Baydar, Ieva Rauluseviciute, Dina R Aronsen, Romain Blanc-Mathieu, Ine Bonthuis, Herman de Beukelaer, Katalin Ferenc, Alice Jegou, Vipin Kumar, Roza Berhanu Lemma, Jérémy Lucas, Mathis Pochon, Chang M Yun, Vivekanandan Ramalingam, Salil Sanjay Deshpande, Aman Patel, Georgi K Marinov, Austin T Wang, Alejandro Aguirre, Jaime A Castro-Mondragon, Damir Baranasic, Jeanne Chèneby, Sveinung Gundersen, Morten Johansen, Aziz Khan, Marieke L Kuijjer, Eivind Hovig, Boris Lenhard, Albin Sandelin, Klaas Vandepoele, Wyeth W Wasserman, François Parcy, Anshul Kundaje, and Anthony Mathelier. JASPAR 2026: expansion of transcription factor binding profiles and integration of deep learning models. *Nucleic Acids Research*, 54(D1):D184–D193, 2025. ISSN 0305-1048. doi: 10.1093/nar/gkaf1209.

[37] Ilya E Vorontsov, Irina A Eliseeva, Arsenii Zinkevich, Mikhail Nikonov, Sergey Abramov, Alexandr Boytsov, Vasily Kamenets, Alexandra Kasianova, Semyon Kolmykov, Ivan S Yevshin, Alexander Favorov, Yulia A Medvedeva, Arttu Jolma, Fedor Kolpakov, Vsevolod J Makeev, and Ivan V Kulakovskiy. HOCOMOCO in 2024: a rebuild of the curated collection of binding models for human and mouse transcription factors. *Nucleic Acids Research*, 52(D1):D154–D163, 2024. ISSN 0305-1048. doi: 10.1093/nar/gkad1077.

[38] E. Wingender, X. Chen, R. Hehl, H. Karas, I. Liebich, V. Matys, T. Meinhardt, M. Prüß, I. Reuter, and F. Schacherer. TRANSFAC: an integrated system for gene expression regulation. *Nucleic Acids Research*, 28(1):316–319, 2000. ISSN 0305-1048. doi: 10.1093/nar/28.1.316.

[39] M Pachkov, I Erb, N Molina, and E van Nimwegen. SwissRegulon: a database of genome-wide annotations of regulatory sites. *Nucl Acids Res*, 35(Database issue):D127 – 31, 2007. ISSN 0305-1048. doi: 10.1093/nar/gkl857.

[40] Rodger Staden. Methods for calculating the probabilities of finding patterns in sequences. *Computer Applications in the Biosciences*, 5(2):89–96, 1989. doi: 10.1093/bioinformatics/5.2.89.

[41] Hélène Touzet and Jean-Stéphane Varré. Efficient and accurate P-value computation for Position Weight Matrices. *Algorithms for Molecular Biology*, 2:15, 2007. doi: 10.1186/1748-7188-2-15.

[42] Tobias Madsen, Asger Hobolth, Jens Ledet Jensen, and Jakob Skou Pedersen. Significance evaluation in factor graphs. *BMC Bioinformatics*, 18:199, 2017. doi: 10.1186/s12859-017-1614-z.

[43] Nancy Reid. Saddlepoint methods and statistical inference. *Statistical Science*, 3(2):213–238, 1988. doi: 10.1214/ss/1177012906.

[44] Otto G. Berg and Peter H. von Hippel. Selection of DNA binding sites by regulatory proteins. Statistical-mechanical theory and application to operators and promoters. *Journal of Molecular Biology*, 193:723–750, 1987.

[45] Otto G. Berg and Peter H. von Hippel. Selection of DNA binding sites by regulatory proteins. II†. The binding specificity of cyclic AMP receptor protein to recognition sites. *Journal of Molecular Biology*, 200:709–723, 1988. doi: 10.1016/0022-2836(88)90482-2.

[46] Guy Sella and Aaron E. Hirsh. The application of statistical physics to evolutionary biology. *Proceedings of the National Academy of Sciences*, 102(27):9541–9546, 2005. doi: 10.1073/pnas.0501865102.

[47] Anirvan M. Sengupta, Marko Djordjevic, and Boris I. Shraiman. Specificity and robustness in transcription control networks. *Proceedings of the National Academy of Sciences*, 99(4):2072–2077, 2002. doi: 10.1073/pnas.022388499.

[48] Ulrich Gerland, J. David Moroz, and Terence Hwa. Physical constraints and functional characteristics of transcription factor–DNA interaction. *Proceedings of the National Academy of Sciences*, 99(19):12015–12020, 2002. doi: 10.1073/pnas.192693599.

[49] Richard A. Neher and Boris I. Shraiman. Statistical genetics and evolution of quantitative traits. *Reviews of Modern Physics*, 83:1283–1300, 2011. doi: 10.1103/RevModPhys.83.1283.

[50] Johannes Berg, Stana Willmann, and Michael Lässig. Adaptive evolution of transcription factor binding sites. *BMC Evolutionary Biology*, 4:42, 2004. doi: 10.1186/1471-2148-4-42.

[51] V Mustonen and M Lassig. Evolutionary population genetics of promoters: predicting binding sites and functional phylogenies. *Proc Natl Acad Sci USA*, 102(44):15936 – 15941, 10 2005. ISSN 0027-8424. doi: 10.1073/pnas.0505537102.

[52] Ville Mustonen, Justin Kinney, Curtis G. Callan, and Michael Lässig. Energy-dependent fitness: A quantitative model for the evolution of yeast transcription factor binding sites. *Proceedings of the National Academy of Sciences*, 105(34):12376–12381, 08 2008. ISSN 0027-8424. doi: 10.1073/pnas.0805909105.

[53] Michael Lässig. From biophysics to evolutionary genetics: statistical aspects of gene regulation. *BMC Bioinformatics*, 8(Suppl 6):S7, 2007. doi: 10.1186/1471-2105-8-S6-S7.

[54] C Anders Olson, Nicholas C Wu, and Ren Sun. A comprehensive biophysical description of pairwise epistasis throughout an entire protein domain. *Current biology : CB*, 24(22):2643 – 2651, 11 2014. ISSN 0960-9822. doi: 10.1016/j.cub.2014.09.072.

[55] Ammar Tareen, Mahdi Kooshkbaghi, Anna Posfai, William T. Ireland, David M. McCandlish, and Justin B. Kinney. MAVE-NN: learning genotype-phenotype maps from multiplex assays of variant effect. *Genome Biology*, 23:98, 2022. doi: 10.1186/s13059-022-02661-7.

[56] Ammar Tareen and Justin B Kinney. Logomaker: beautiful sequence logos in python. *Bioinformatics*, 36(7):2272–2274, 2019. ISSN 1367-4803. doi: 10.1093/bioinformatics/btz921.

[57] Yoh Iwasa. Free fitness that always increases in evolution. *Journal of Theoretical Biology*, 135 (3):265–281, 1988. doi: 10.1016/S0022-5193(88)80243-1.

[58] Charles G. Darwin and Ralph H. Fowler. On the partition of energy. *Philosophical Magazine Series 6*, 44(261):450–479, 1922. doi: 10.1080/14786440908565189.

[59] Henry E. Daniels. Saddlepoint approximations in statistics. *The Annals of Mathematical Statistics*, 25(4):631–650, 1954. doi: 10.1214/aoms/1177728652.

[60] O.E. Barndorff-Nielsen and D.R. Cox. *Asymptotic Techniques for Use in Statistics*. Monographs on Statistics and Applied Probability. Springer US, 1989. ISBN 9780412314001. URL https://books.google.com/books?id=UQ9yIrZpMToC.

[61] Tyler N. Starr and Joseph W. Thornton. Epistasis in protein evolution. *Protein Science*, 25: 1204–1218, 2016. doi: 10.1002/pro.2897.

[62] Júlia Domingo, Pablo Baeza-Centurion, and Ben Lehner. The causes and consequences of genetic interactions (epistasis). *Annual Review of Genomics and Human Genetics*, 20:433–460, 2019. doi: 10.1146/annurev-genom-083118-014857.

[63] Gautam Reddy and Michael M. Desai. Global epistasis emerges from a generic model of a complex trait. *eLife*, 10:e64740, 2021. doi: 10.7554/eLife.64740.

[64] John H. Gillespie. A simple stochastic gene substitution model. *Theoretical Population Biology*, 23:202–215, 1983.

[65] Adam Eyre-Walker and Peter D. Keightley. The distribution of fitness effects of new mutations. *Nature Reviews Genetics*, 8(8):610–618, 2007. ISSN 1471-0056. doi: 10.1038/nrg2146.

[66] R. A. Fisher. *The Genetical Theory of Natural Selection*. Clarendon Press, Oxford, 1930.

[67] H. Allen Orr. The population genetics of adaptation: the distribution of factors fixed during adaptive evolution. *Evolution*, 52:935–949, 1998. doi: 10.1111/j.1558-5646.1998.tb01823.x.

[68] Sungmin Hwang, Su-Chan Park, and Joachim Krug. Genotypic complexity of Fisher's geometric model. *Genetics*, 206:1049–1079, 2017. doi: 10.1534/genetics.116.199497.