Parental age at reproduction accelerates offspring pace of life in *Gryllus bimaculatus*.

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Open research statement

Data and code are not yet provided but will be made openly and freely available on request during the review process. Data will be uploaded to the Dryad data repository and code will be uploaded to Zenodo open repository on acceptance of the manuscript.

Abstract

Across invertebrates, it is widely known that older parents produce offspring with abbreviated lifespans (i.e., the Lansing effect). Yet the ecological and evolutionary ramifications of such nongenetic parental age effects remain unclear. Two demographic processes could culminate in a Lansing effect, with different fitness implications: (i) Later-conceived offspring may display greater mortality from conception, constraining fitness from birth. (ii) Alternatively, they may exhibit an increased age-specific mortality (i.e., accelerated senescence), with negative effects that manifest after maturity. If parental age accelerates offspring senescence without affecting initial mortality rate, then later-conceived offspring may maintain fitness by shifting to a faster pace-of-life. We exposed *Gryllus bimaculatus* parents to one of three experimental temperatures to generate environmental variation in parent mortality. Using a longitudinal within-parent study, we tested whether parental age influenced offspring longevity via increased initial mortality or accelerated senescence, and whether these effects depended on the parents' thermal environment. Contrary to our predictions, we found no evidence that parental age reduced offspring longevity. Instead, parental age mediated offspring pace-of-life. Irrespective of the parents' thermal environment, the earliest- and latest-conceived offspring were demographically distinct. Later-conceived offspring exhibited accelerated development, a smaller adult mass, and improved reproductive success. Unexpectedly, they also showed extended adult lifespans. This increased adult lifespan did not arise from reduced initial mortality, but from a delayed onset and slower progression of actuarial senescence. The slowed senescence in later-conceived offspring is unlikely to reflect a direct parental age effect, potentially arising indirectly through causal effects on adult size. Our results challenge the universality of the Lansing effect and contribute to a growing recognition that parental age has important life-history consequences. Accelerated life-histories in later-conceived offspring may be adaptive, enabling these individuals to compete over dwindling seasonal resources or overcome seasonal time constraints. Determining whether these demographic parental age effects accumulate across generations, or persist under natural environments, will be essential for establishing their ecological consequences and proximate role in the evolution of ageing.

Introduction

Classical theories of ageing have implicitly assumed that senescence, the progressive increase to mortality and reduction in reproduction and somatic maintenance with increasing chronological age, is an exclusively within-individual phenomenon (Hamilton, 1966; Abrams, 1991; Monaghan et al. 2008; Nussey et al. 2013). Here, senescent damage is restricted to the soma, while the germline remains protected, or passes through rejuvenating mechanisms, that prevent the unregulated accumulation of senescent damage across generations (Kirkwood & Holliday, 1979; Rose, 1984; Kern et al. 2001; Monaghan & Metcalfe, 2019). Yet growing evidence suggests senescence can be transmitted intergenerationally to have penetrant detrimental effects on offspring quality (i.e., the Lansing effect or parental effect senescence; Lansing, 1947; Monaghan et al. 2020). It is crucial to understand the importance of these nongenetic parental age effects in both the evolution of senescence and species life-histories, as their adaptive role has previously been understated (Moorad & Nussey, 2016; Van Den Heuvel et al. 2016; Barks & Laird, 2020; Hernández et al. 2020).

Although evidence for negative effects of parental age at reproduction on offspring traits, hereafter 'parental effect senescence, continues to accumulate from both captive and natural populations (see: Kern et al. 2001; Priest et al. 2002; Schroeder et al. 2015; Lippens et al. 2017; Noguera et al. 2018; Monoghan et al. 2020; Reichert et al. 2020; Noguera et al. 2021; Angell et al. 2022), there is limited agreement over its universality and the underlying mechanisms involved (King, 1983; Anderson et al. 2022; Ivimey-Cook et al. 2023). Some of the earliest evidence for parental effect senescence comes from Lansing (1947), who demonstrated that old-aged lines of the

parthenogenic rotifer *Philodina citrina* produced offspring with abbreviated lifespans

(i.e., the Lansing effect), with this effect propagating across generations. More

recently, a meta-analysis investigating the general tendency for the Lansing effect by

Ivemy-Cook et al. (2023) demonstrated that a one-unit increase in maternal age was

associated with a 17 to 22% reduction in the offspring's adult lifespan, on average.

Alongside adult longevity, detrimental parental age effects have also been

demonstrated on embryo viability (Bloch Qazi et al. 2017; Noguera, 2021), juvenile

survival (Kern et al. 2001; Reid et al. 2010; Torres et al. 2011; Ivimey-Cook & Moorad,

2020), development and growth rate (Fox & Dingle, 1994; Singh & Omkar, 2009; Lind

et al. 2015), size at birth and maturity (Opit & Throne, 2007; Vega-Trejo et al. 2018),

and reproductive performance (Hercus & Hoffmann, 2000; Schroeder et al. 2015; Van

Daalen et al. 2022). However, there is a growing uncertainty over the ubiquity of this

phenomenon; studies frequently find conflicting results, even between laboratory

strains (Priest et al. 2002; Anderson et al. 2022), including neutral (Ivimey-Cook &

Moorad, 2018; Travers et al. 2021) or positive parental age effects on offspring

longevity (an "inverse Lansing effect"; Marshall et al. 2010; Anderson et al. 2022).

This lack of consistency raises challenges in identifying the physiological pathways,

biological factors, and environments that are important for parental effect

senescence (Priest et al. 2002; Monaghan et al. 2020).

Despite an extensive history of research, numerous issues plague the current study

designs, which could perturb, or spuriously create, the observed parental age effects.

First, most studies remain cross-sectional (see: Priest et al. 2002; Plaistow et al.

2015; Bloch Qazi et al. 2017; Bock et al. 2019; Wylde et al. 2019; Angell et al. 2022),

and cannot control for the differential survival of parental phenotypes (i.e., selective

disappearance: Nussey et al. 2008; Reid et al. 2010; Ivimey-Cook & Moorad, 2018;

Marasco et al. 2025). Second, studies into parental age effects are often

correlational, and there is a growing necessity for experimental manipulations of

parent longevity (both genetic and environmental) to tease apart the causal effects of

parental senescence on offspring performance (Kern et al. 2001; Bock et al. 2019).

Previous work has demonstrated that parental effect senescence can exhibit

genotypic heterogeneity, differing between strains artificially selected for early- or

late-life performance (Kern et al. 2001; Priest et al. 2002; Anderson et al. 2022).

However, there exists limited research on whether environmental variation in parent

mortality could also modulate the expression of parental age effects, which could

have important implications for understanding the intergenerational impacts of health

and lifestyle. Evidence suggests that adverse parental age effects may be context-

dependent, being influenced by the parents' diet and population density, though the

proximate role of parents' thermal environment has received little attention, despite

the known importance of temperature in modulating the lifespan of ectothermic

organisms (Fox & Dingle, 1994; Plaistow & Benton, 2009; Gribble et al. 2014; Burraco

et al. 2020; Morimoto, 2022). The role of temperature in dictating longevity in

ectothermic insects is complex, with chronic exposure to warm temperatures either

raising baseline mortality (i.e., temperature induced frailty) or accelerating

senescence, which may have different implications for offspring fitness (Mair et al.

2003; Burraco et al. 2020; Ma et al. 2021. Third, few have considered how the

variability of offspring phenotypes changes with parental age. It could be that

parental effect senescence not only governs the offspring's average trait value, but

also generates greater variation in offspring phenotypes, as has been similarly

observed for other parental environmental stressors (e.g., increasing temperature; De

Jong, 1995; O'Dea et al. 2016). Any change in the variability of offspring phenotypes

may be adaptive (i.e., bet-hedging; Crean & Marshall, 2009) or reflect physiological

constraints (Debat & David, 2001; Olsson & Uller, 2002), such as the senescent

instability of the parents' reproductive systems (Monaghan & Metcalfe, 2019). Finally,

despite the evolutionary importance of parental age effects in understanding

senescence, most studies have tested for their isolated effects on offspring longevity –

the end point of senescence (Promislow et al. 2022). However, this precludes any

effects of parental age on offspring age-specific mortality (i.e., *how* the offspring

senesce) and reproductive performance. Assessing the joint effects of parental age on

offspring mortality and reproduction is a necessary step in establishing the fitness,

demographic, and evolutionary consequences of parental effect senescence

(Kirkpatrick & Lande, 1989; Nussey et al., 2013; Bloch Qazi et al. 2017; Monaghan &

Metcalfe, 2019).

Monaghan et al. (2020) identified that increasing parental age could reduce offspring

longevity through two non-exclusive demographic processes. First, offspring from old-

age parents may be conceived with an increased frailty. Here, their mortality is

higher from conception as they initiate life with either the senescent damage (e.g.,

shortened telomeres, increased point mutation rates, abnormal DNA methylation

patterns, heightened mitochondrial dysfunction) accumulated by their aged parents

(Díaz & Esponda, 2004; Kong et al. 2012; Monaghan & Metcalfe, 2019; Dutt & Laird,

2024), or experience age-dependent reductions in either pre- or post-natal parental

provisioning (Bouwhuis et al. 2010; Lemaître & Gaillard, 2017; Muller et al. 2017;

Ivimey-Cook & Moorad, 2018). Here, this increased frailty persists, but does not

necessarily worsen, as the offspring age. Alternatively, offspring from old-aged

parents may exhibit increased age-specific mortality (i.e., accelerated senescence),

with the negative effects of their accumulated senescent damage only manifesting

after maturity, being buffered against during the developmental period (Monaghan et

al. 2020). Despite the breadth of studies demonstrating parental effect senescence,

few have assessed its demographic underpinnings. Indeed, those that have reported

conflicting results, with either increased frailty (Dutt & Laird, 2024), accelerated

senescence (Plaistow et al. 2015) or both (Bock et al. 2019), being implicated in

reducing offspring longevity. Although both demographic processes reduce survival,

this may not necessarily culminate in reducing offspring fitness. Rather, the two

processes can have different consequences for offspring life-histories. If offspring

exhibit increased frailty, then their fitness is likely constrained from birth.

Conversely, if parental age accelerates offspring senescence, but has no effect on

frailty, then offspring may undertake a phenotypically plastic switch to a fast pace-of-

life strategy. In this latter scenario, offspring may accelerate development to reach

reproductive maturity earlier in life, increasing investment in their early life fitness

contributions prior to their hastened onset of senescence (King, 1983; Bouwhuis et al.

2010; Plaistow et al. 2015; Monaghan et al. 2020). Thus, although increasing parental

age may have no effect on offspring fitness, the offspring produced at the extreme

ends of the parent's life could be demographically distinct (Van Noordwijk & De Jong,

1986; Travers et al. 2021; Anderson et al. 2022).

Using the two-spotted cricket (*Gryllus bimaculatus*), and an experimental,

longitudinal within-individual study design, we investigated how parental age affected

offspring mortality trajectories and life-history traits. *G.bimaculatus* has previously

been shown to display negative effects of parental age on offspring traits, including

longevity. However, this was with a cross-sectional study in which group housed

offspring of old and young parents were compared, rather than through any direct

impact of *within* parent senescence on individually housed offspring unaffected by

group competition effects (Noguera, 2021). In this study, we first addressed a key

conceptual issue: does experimentally induced environmental heterogeneity in parent

mortality mediate the expression of parental effect senescence? To this end, we

manipulated the temperature parents experienced over their adult lifespan, creating

variation in parent longevity. We then investigated whether parental age effects

manifested differently depending on the parents' temperature treatment, predicting

that any adverse parental age effects would be exacerbated under a warm

environment. Then, we aimed to disentangle which demographic process (frailty or

actuarial senescence) was implicated in reducing offspring longevity. Here, we

confine our inferences of senescence to the offspring's adult, rather than total,

lifespan. Adult eclosion marks the initiation point from which senescence progresses

in *Gryllus spp.*, when the individual has shifted resources from growth to reproduction

(Rodríguez-Muñoz et al. 2019). After determining how parental effect senescence

arises, we asked whether the following life-history traits were affected by increasing

parental age, in line with an expected switch to a fast pace-of-life strategy: post-

natal development time, adult size, and offspring reproductive performance. Finally,

for these listed traits, not only did we estimate the effect of parental age on their

average value, but we also aimed to establish how their variability changed over the

parents' lifespan.

Methods

2.1 Laboratory conditions

We conducted the study outlined below from September 2023 to September 2024,

using an outbred (<8 generation descendants from the wild) laboratory-reared

population of *G.bimaculatus*, located at the Scottish Centre for Ecology and the

Natural Environment (SCENE), University of Glasgow. All animals were individually

housed in plastic containers (75H × 125W × 185Lmm for the parent generation,

75H × 108W × 108Lmm for the offspring generation), containing *ad-libitum* rabbit

pellets (Supreme Petfoods Science Selective Adult Rabbit food) a cotton-soaked water

tube, and an egg carton for shelter. We individually housed individuals to remove any

effects that intra-specific competition may have on life-history traits (Simmons,

1987). Individuals were kept in incubators set to a 14:10 hour day: night cycle with a

relative humidity (RH) of ~50%. We continuously monitored temperature and relative

humidity through the experimental period with loggers (Elitech RC 5+, RS-191A

Temperature Loggers + DOOMAY mini hygrometer). We rotated all individuals amongst

the incubators weekly, ensuring animals were alternated between incubators and

shelve locations.

2.2 Creating the parent generation

Colony animals (12 pairs) were mated opportunistically, avoiding pairing related animals, to minimise inbreeding. Eggs were collected ad-hoc from the oviposition substrate (damp cotton) before being plated and incubated in a climate room for nine days at 28.5°C (14:10 hour day: night cycle; RH of 40-60%), after which we initiated daily hatch checks. Once hatched, the parent generation was housed individually in incubators (set temperature 25.0-26.5°C; *equally experienced by all offspring*). From January 2024, we monitored late-stage instars daily to record the exact date of adult eclosion. As adults can be damaged shortly after eclosing, we waited at least 24 hours after observed emergence to measure their mass using a VWR SMG4i precision balance (readability ±0.01mg). We photographed all individuals (Panasonic Lumix DC-FZ82 camera + Raynox DCR-250 macro lens (focal length: 60mm) + set to *"autofocus"* mode) to measure pronotum width/length (mm). To prevent individuals from moving during photography, we placed them in a petri dish filled with cotton (100mm x 15mm), with a scale attached. We measured pronotum width and length (in mm) from the images with the line tool in IMAGEJ (Schneider et al. 2012). Images were analysed by a single observer (Mark D. Pitt; MDP) to eliminate any between-observer measurement bias. Pronotum width and length are assumed to be more fixed indicators of structural size than adult mass, as daily variation in hydration, satiation, and oviposition can cause mass to have low within-individual repeatability (Gershman et al. 2022). However, we observed reasonably tight correlations between all three measures of adult body size (pronotum width ~ pronotum length, Pearson's correlation coefficient r [95% CI] = 0.856 [0.760, 0.916]; pronotum width ~ adult body mass,

Pearson's correlation coefficient *r* [95% CI] = 0.758 [0.610, 0.855]; pronotum length ~

adult body mass, Pearson's correlation coefficient *r* [95% CI] = 0.808 [0.685, 0.886],

using a subset of photographs, N = 51 animals). Therefore, we selected adult body

mass as our only proxy of adult size, assuming this measurement captured relevant

biological variation. Survival for the parent generation was monitored weekly, at

which point we also undertook husbandry (e.g. replaced food, water, and cleaned

housing).

242   2.3 Parent mating design

We paired parent animals pseudo-randomly, such that inbreeding was minimised, and,

where possible, members of a pair were age matched. Most parents were mated in

their second week of adult life, with an average adult age of 1.4 ± 0.67 weeks (mean

± SD) at first mating. This allowed time for the oocytes in the females to mature,

which usually occurs in the initial 10 days post adult eclosion (Lorenz, 2007). During

the staged matings, the parent pair were placed into a plastic container

(75H × 125W × 185Lmm) and allowed to interact for 15 minutes. Pairs were watched

continuously, and mating was assumed to be successful after visual confirmation of

spermatophore transfer. We then immediately assigned pairs to be continuously

maintained under one of the three temperature treatments (25.5°C, 28.0°C, or

30.5°C). Previous experimental work has revealed that the survival of *Gryllus spp.* is

sensitive to experimental temperature manipulations, both constant and fluctuating,

over their adult lifespan (Centeno Filho et al. 2023). Additionally, we have

demonstrated that the strength of reproductive senescence is temperature dependent

in *G.bimaculatus*, with females exposed to the highest temperatures exhibiting the

steepest age-dependent reductions to fecundity and hatching success (*in prep*). Our

temperature manipulations successfully created experimental variation in parent

longevity, with the Kaplan-Meier curve demonstrating that parents experiencing the

warmer temperature treatments exhibited reduced survival relative those maintained

at 25.5°C (*Appendix S1: Section S1*).

All parent animals were housed individually between mating attempts. We mated

pairs weekly, with offspring being collected and raised from the parents first eight

mating attempts. At the last mating from which offspring were housed, parents had

an average adult age of 8.52 ± 0.80 weeks (mean ± SD). The parents were repeatedly

mated with the same partner to reduce the likelihood of stored sperm senescence

confounding our results (Ribou & Reinhardt, 2012; Monaghan et al. 2020). We did not

directly observe any subsequent mating attempts after the first mating attempt.

Instead, pairs were housed together for three hours, before being separated. For four

pairs, we observed that one member of the pair had died after the staged mating,

though we were unable to identify if this was a natural death or cannibalism. Not all

parents produced hatchlings from all eight mating attempts, with some pairs ceasing

to produce viable eggs earlier than others. Due to capacity constraints, parents were

removed from their temperature treatment to a climate room (28.5°C; 14:10 hour

day: night cycle; RH of 40-60%) following three consecutive mating attempts that

resulted in no viable hatchlings (*Appendix S1: Section S1*). Our final sample size

consisted of 78 pairs (156 animals), with these representing animals that produced

viable eggs from their first mating attempt onwards. The sample sizes for each

temperature treatment were as follows: 27 pairs at 25.5°C, 23 pairs at 28°C, and 28

pairs at 30.5°C.

2.4 Creating the offspring generation

After each staged mating, we allowed females 15 hours to lay, providing them with a

petri dish (60mm x 13mm) of sand (Children's Play Sand), covered with a damp cotton

pad for laying. As laying effort may vary daily, we provided females with sand for

three days consecutively. Where possible, we only collected and incubated eggs from

the first and third batches of eggs. All eggs were photographed before being

incubated on a petri dish (100mmx 15mm; filled with damp cotton) in the climate

room. Eggs were inspected daily and, if hatchlings were observed, they were

photographed (Panasonic Lumix DC-FZ82 camera, focal length: 35-40mm; *for another*

study) as a group before being housed individually (*details below*). We defined a

successful hatching as any nymph that had completely removed itself from its egg

casing. Hatching was asynchronous, and if there were still unhatched eggs present,

we returned the remaining eggs to the climate room. Hatching checks continued daily

until all eggs on a dish had hatched or up to 14-days after first incubation, after which

remaining eggs were assumed to be non-viable.

From each mating attempt, we individually housed up to three offspring per parent

pair. Offspring were inspected weekly for any adult emergence or mortality. As per

the parent generation, we waited two days after adult emergence to measure adult

mass. Overall, our sample size consists of 1317 individually housed offspring from 78

parent pairs, while complete life-history data was collected from 1309 offspring. Eight

offspring had uncertainty surrounding their adult emergence dates and were excluded

from analyses on offspring development time, adult lifespan, and adult mass. A

further two animals had missing mass measurements and were thus removed from any

size analysis.

2.5 Offspring reproductive performance

Once the offspring generation reached adulthood, we mated a subset of individuals to

determine how parental age affected offspring reproductive success. We collected

and incubated offspring eggs from a single mating, at which offspring had a mean

adult age of 4.60 ± 1.44 weeks (mean $\pm$ SD), assuming this window represented an

intermediate adult age (e.g., neither young nor old). To increase the diversity of

offspring that could be paired, we binned parental age into two-levels, producing a

subset of individuals from either "early" or "late" aged parents. Offspring from the

"early" cohort descended from the first two parental mating attempts, while the

"late" cohort of offspring descended from parental mating attempts five through to

eight. We mated the offspring within the levels of parental age and parental

temperature. As per the parent generation, we paired offspring opportunistically,

minimising inbreeding.

We performed staged matings, egg collection, photography, and incubation, and

hatching checks as per the parent generation. We counted eggs and hatchlings using

the cell count tool in IMAGEJ, with these counts being conducted by a single observer

(MDP). For ease of analysis, we pooled measures of fecundity and hatching success

across all egg batches. Our sample size consists of eggs laid by 83 offspring pairs

descending from 45 parent pairs ("early" = 47 offspring descending from 32 parent

pairs, "late" = 36 offspring descending from 26 parent pairs). Meanwhile, a single pair

was removed from the hatching success analysis due to unreliable count data (many

nymphs had died at the time of the hatchling counts), leaving 82 egg batches for the

offspring hatching success analysis.

2.6 Data Analysis

2.6.1. General procedure for modelling life-history traits in brms

Model workflow

We undertook all statistical analysis and data visualisation in *RStudio*

v.2025.05.1+513, using *R v.4.3.3* (R Core Team, 2024). We used the package *brms*

v.2.21.0 (Bürkner, 2017), implementing *rstan* v.2.32.2 (Gelman et al. 2015; Stan

Development Team, 2024) as a backend, to create Bayesian generalised linear mixed

models (GLMMs). We follow the location-scale approach to regression modelling,

where both first-order (i.e., the mean/location (μ)) and second-order (i.e., the

variance/scale (σ or ϕ)) traits are estimated (O'Dea et al. 2022; Nakagawa et al.

2025). This location-scale modelling approach permits heteroskedasticity, where

differences in error variance exist across the predictors (Barreda & Silbert, 2023).

Additionally, in models containing a zero-inflation component (z), we also directly

estimated z to identify whether any predictors were associated with zero outcomes in

the response.

We included both fixed (β) and random (u) terms in the location submodel, while only

fixed effects were included in the scale and zero-inflation components. This single-

hierarchical framework ensured that the distributional parameters remained

identifiable (Nakagawa et al. 2025). Where possible, we included the parents' age as

a random slope, allowing the effect of parental age to differ between parent pairs.

We performed model simplification within the scale and zero-inflation component by

removing predictors exhibiting no effect or those that were weakly identifiable to

reduce the risk of model overparametrisation (Barreda & Silbert, 2023). We always

retained parental age in the scale and zero-inflation components, being central to our

underlying question (i.e., whether parental age affects the *variability* of offspring

traits). In the location component, we did not perform model simplification beyond

the removal of non-meaningful interactions (including quadratic terms) to ease the

interpretation of single-effect predictors. All continuous covariates were z-

transformed (mean = 0, standard deviation (SD) = 1), to facilitate prior specification.

For each fixed effect, we show the posterior mean estimates alongside the 95%

credible intervals (CI's) from the *brms* output. Additionally, we assessed the existence

and importance of the fixed effects with both the probability of direction (pd) and

Region of Practical Equivalence (ROPE), as estimated from *bayestestR v.0.17.0*

(Makowski et al. 2019). We used *pd* to describe the proportion of posterior estimates

in the same direction as the posterior mean, to assess whether the effect existed. We

assumed fixed effects with a pd ≥ 98% were real (i.e., differed from zero; Makowski et

al. 2019). However, the pd only indicates directionality, not the practical importance

nor magnitude of effect, which is better described by the ROPE statistic. ROPE

defines an area around the null value effectively equivalent to zero, and of negligible

interest (Kruschke, 2018). In this study, we set ROPE to the equivalent of $\pm 0.1 \times SD_y$

(Kruschke, 2018). This range was consistently applied across model families, with

appropriate transformation to the log-link scale when necessary. Meanwhile for

models with the logit link (i.e., Bernoulli & beta-binomial) we set ROPE to the default

(±0.18). We inferred that parameter posterior distributions with > 95% of estimates in

ROPE were negligible, while those with <5% within the ROPE had a substantial effect

(Makowski et al. 2019; Schwaferts & Augustin, 2020). We interpreted cases between

these thresholds with caution, weighing the magnitude of the estimate (ROPE) with

the directional evidence (pd). We provide all pd and ROPE statistics, together with

model summaries, in the supplementary results (*Appendix S1: Table S3 – S20*). We did

not apply ROPE to the scale or zero-inflation components, as defining areas of

negligible effect for these parameters lacked interpretability.

For post-hoc inferences, we used the package *emmeans v.1.10.1* (Lenth, 2024) to

report posterior medians and 95% HDI's (Highest density intervals) on the response

scale. Additionally, as a visual measure of effect size, we then converted these post-

hoc medians to a standardised mean difference (SMD), by dividing the pairwise

difference by the raw standard deviation of the response. Using the package

bayesplot v.1.11.1 (Gabry & Mahr, 2025), we also plotted posterior distributions for

fixed-effect predictors, plotting the posterior medians ± 95% CI's. We used the

package *ggplot2 v.4.0.0* (Wickham, 2016) to visualise effect sizes, the raw data, and

model predictions for the trait mean and variance. To visualize the entire distribution

of offspring traits (i.e., variance), we plotted the probability mass for discrete

outcomes ($p(x)$), the probability density for continuous outcomes ($f(x)$), and the

cumulative incidence for time-to-event outcomes (*F(x)*). The plotted predictions were

based on the median estimate ± 95% CI across posterior draws.

Parental age: within-subject centring and selective disappearance

We used within-subject centring to control for between-individual heterogeneity in

parental age (Van De Pol & Wright, 2009). First, we described the between-parent

effect with the *parents' average age at reproduction ($\overline{age}$)*, calculated as the average

age of the mother across mating attempts resulting in successfully hatched eggs.

Second, the within-parent effect was captured by the *parents' delta age at*

reproduction (Δage), calculated as the difference between the parents' average age

and the mother's age at the current mating attempt. Here, we concluded that the

effect of *Δage* on each offspring trait represented the true within-individual parental

age effect. Across all models, a unit increase in *Δage* corresponds to a single standard

deviation increase in the parents' age at reproduction, or 2.06 weeks on the original

scale. We could not include the father's age at reproduction as an additional

covariate, as this was tightly correlated with maternal age (Pearson's correlation

coefficient *r* [95% CI] = 0.98 [0.97, 0.98]).

As a test for selective disappearance, we calculated the difference between the

slopes of these two parental age terms (using the *hypothesis ()* function from *brms*). If

the slopes for $\overline{age}$ and *Δage* matched (i.e., did not differ from zero), then we assumed

the between-individual parental age effect equaled the within-individual effect (Van

De Pol & Wright, 2009). If the Posterior probability (*Post.prob*) for the Δslopes

between the two terms had an estimate <0.98, we inferred there was uncertain

evidence for selective disappearance. The parents' delta age was initially included as

a quadratic, before the quadratic term was removed to ease interpretation of the

linear slope and simplify tests of selective disappearance (Fay et al. 2022), as its

inclusion did not improve Out-of-sample predictive performance (*details below*). We

aimed to include the parents' cumulative number of successful mating attempts (i.e.,

mating attempts from which eggs hatched, and offspring were subsequently housed)

as an additional covariate, to tease apart any variation in offspring phenotypes

explained by accumulated reproductive effort. However, this term was tightly

correlated with *Δage* owing to our longitudinal study design (cumulative number of

mating attempts ~ *Δage*, Pearson's correlation coefficient r [95% CI] = 0.94 [0.92,

0.95]; *Appendix S1: Figure S2*) and was not included in our modelling framework.

Model settings

Unless otherwise stated, we used the following settings for our fitted models: four

Hamiltonian Monte Carlo (HMC) parallel chains, 5000 iterations each (2500 warmup

chains which were discarded), and a thinning of one. This resulted in 10,000 post-

warmup posterior samples per parameter. We followed standard practice and

monitored model convergence and the effective sample sizes as recommended by

Gelman & Shalizi (2013). We used the potential scale reduction statistic ($\hat{R}$) supplied

by the model output to compare between and within-chain estimates for the model

parameters. We assumed that chains had sufficiently mixed when $\hat{R}$ <1.01 (Gelman &

Rubin, 1992; Vehtari et al. 2021). Model convergence was said to be achieved when

the effective sample sizes (both bulk and tail) for model parameters were >1,000

(Gelman & Rubin, 1992; Vats & Knudson, 2021).

Model diagnostics

To assess goodness-of-fit, we inspected the Leave-One-Out Probability Integral

Transform (LOO-PIT) plots from *bayesplot*, ensuring the LOO-PIT integrals were

uniformly distributed (Vehtari et al. 2017; Säilynoja et al. 2022; Nguyen et al. 2025).

Additionally, we used the *ppc_loo_intervals()* function to determine how well our

model predicted each observation. We also visualised how well the empirical

distribution of the data (*y*) fitted to the distributions of simulated data (*yrep*) from

the posterior predictive distribution, using the *pp_check()* function (Gabry et al. 2019;

Winter & Bürkner, 2021). Additionally, we checked for any multicollinearity between

predictors (VIF) using the package *car v.3.1.2 (*Fox & Weisberg, 2019*)*.

Model predictive performance

To assess the Out-of-sample predictive performance of each model, we used the

expected log pointwise predictive density (*elpd*), calculated with Pareto smoothed

importance sampling Leave-One-Out cross-validation (PSIS-LOO; Vehtari et al. 2015,

2017). For a series of models fitted to the same data, a higher *elpd* value indicates

greater predictive performance (Vehtari et al. 2016). We assumed that models had a

similar predictive ability if the difference in between their *elpd* (*Δelpd*) was < four

(Sivula et al. 2020). When the predictive ability of the compared models was

comparable, we selected the simplest model (with fewest predictors/no interactions).

Choice of prior distribution

For each trait, we conducted prior sensitivity analysis by fitting models with

progressively tighter priors. These ranged from the default flat priors provided by

brms to normally distributed constraining priors that assumed no effect of the fixed

effects on the response (Gelman et al., 2008, 2017). Unless otherwise stated, we

fitted our final models with weakly informative priors, using $\beta_1 \sim normal$ *(mean = 0, SD*

= 1 SD_y) priors for fixed effects, accounting for the scale of the response and the

model's link function (Lemoine et al. 2016; Lemoine, 2019). This weakly informative

prior assumes a one standard deviation increase in the predictor leads to an

approximately one standard deviation change in the response, and rarely any larger

scale changes (Møller & Jennions, 2002; Lemoine, 2019). For a detailed overview of

our prior specification process, including exact prior values for each model parameter

and their justification, see *Appendix S1: Section S3.*

2.6.2 General procedure for survival analyses

Choice of mortality distribution

We used Bayesian survival trajectory analysis (*BaSTA v.2.0.1*; Colchero et al. 2012) to

build parametric survival models explaining variation in offspring mortality. *BaSTA*

uses a Monte Carlo Markov chains (MCMC) algorithm, describing mortality, the

instantaneous risk of death given survival to age x, as $\mu(|\theta)$, where x is age and θ are

the model parameters (Colchero & Clark, 2012). We fitted all possible mortality

distributions parameterised by *BaSTA* (exponential, Gompertz, Weibull, and logistic)

which we extended with the Makeham (*c;* Makeham, 1867) and bathtub (Siler, 1979)

terms. The predictive performance of all the models was compared using their

deviance information criterian (DIC) value. DIC describes the predictive power of each

model by approximating the expected predictive deviance and penalises for model

complexity, with a lower DIC indicating that the model has improved predictive

ability (Spiegelhalter et al. 2002; Millar, 2009). We selected the model with the

lowest DIC, with a model said to be the best-performing when its ΔDIC < 3 from

alternative candidate models. If models had comparable DIC values, we selected the

simplest model (with the fewest predictors/no interactions). Out of all mortality

distributions, we found the Weibull-Makeham model had the best predictive

performance (*Appendix S1: Tables S9 and S21*).

Weibull Makeham model parameters

The Weibull-Makeham model (Pinder et al. 1978) assumes that mortality risk changes

as a power function of age, describing mortality as:

$$\mu_b(x|b0, b1) = c + b0b1^{b0}x^{b0-1}.$$

Under the three-parameter Weibull-Makeham model, *b0* represents the shape

parameter, *b1* is the scale parameter, and *c* is the Makeham parameter, while x

denotes the individual's age.

The shape parameter (*b0*) describes the pattern mortality follows through time. When

b0 equals one, mortality is held constant, while when *b0* equals two, mortality risk

increases linearly with age, and when *b0* is greater than two, mortality exhibits an

accelerating increase over time. Meanwhile, *b1* determines the characteristic scale of

failure times (i.e., when 63.2% of individuals have died, which we interpret as the age

of onset for senescence) of the offspring (Gómez et al. 2023). *BaSTA* inverses the

scale parameter (*1/b1*), with larger values of *b1* corresponding to an earlier

characteristic failure time. Taken together, *b0* and *b1* describe age-specific mortality;

b1 scales the relationship between the age of the individual (x) and the shape of the

curve (*b0*), setting the inflection point for when the mortality increase occurs.

Finally, *c* is the age-independent baseline mortality risk (i.e., background mortality).

Here, we estimated the effect of our predictors on each mortality parameter. In

BaSTA, both *c* and *b1* have units in years, while *b0* is unitless.

Covariate structure and model settings

We only included categorical predictors in our models (using the *"fused"* covariate

structure) for which *BaSTA* provides Kullback-Leibler discrepancy calibration values

(KLDC; Kullback & Leibler, 1951). Between a pair of predictors, KLDC provides a

measure of the distance between their posterior distributions for each parameter. A

value of 0.5 indicates that posterior distributions overlap completely, while a KLDC of

one indicates that the posterior distributions exhibit no overlap (McCulloch, 1989).

When KLDC >0.8, we concluded that there was a substantial difference between the

compared predictors in their effect on the mortality parameter, assuming this

indicated a genuine biological effect (Karabatsos, 2006; Rodríguez-Muñoz et al. 2019).

Our final posterior distributions were based on four MCMC chains with 80,000

iterations (burn-in of 1001), and a thinning interval of 50, resulting in n = 6320 post-

burn-in posterior samples per parameter. We assessed the convergence of the

parameter estimates using $\hat{R}$. Despite reducing the length of the burn-in sequence,

the simulations in all our models rapidly converged, mixing well when stabilising

towards the target distribution ($\hat{R}$ <1.002), and serial autocorrelation was low (<5%).

We assessed model fit visually by overlaying the parametric survival curves over the

Kaplan-Meier plots of observed survival (*Appendix S1: Figures S10 and S18*). For each

parameter, we report the posterior mean along with the 95% CI's. We used *ggplot2* to

visualise parameter posterior distributions alongside the offspring's cumulative

survival curves and instantaneous hazard rates (i.e., mortality). In the final plots, we

show the full CI's for the posterior distributions for each mortality parameter, while

model predictions for the survival and hazard curves reflect the median ± 95% CI

estimates across posterior draws.

Prior sensitivity analysis

To assess the sensitivity of the mortality parameters to our choice of prior structure,

we conducted the analysis using three prior distributions (default, weakly-

informative, and moderately-informative), before selecting normally distributed

weakly informative priors for covariates, as recommended by Lemoine (2019).

Specifically, we set the prior for *c* and b0 to the weakly-informative *BaSTA* defaults

(c~normal [0,1], *b0*~normal [1.5, 1]). For *b1,* we adapted the default prior (normal [0,

1]) to a scale that was appropriate for the short lifespan of *G.bimaculatus*. Given

BaSTA was developed for long-lived wild vertebrates, the default *b1* prior is centred

incorrectly for our study species. To prevent inappropriate prior artifacts from

affecting posterior estimates, we scaled the *b1* prior to the raw population mean for

total or adult lifespan (*details below*). All priors were truncated to be ≥ 0, given the

Weibull model's parameterisation. We also evaluated tighter moderately informative

priors, with the same mean, but ½ the SD of the weakly informative priors.

2.6.3. Statistical models

Approach to modelling offspring longevity

We initially fitted a bathtub term to the Weibull model to capture the steep early-life

mortality of our study population. However, inspection of the Kaplan-Meier curve

against the parametric survival curve indicated the bathtub shape was too shallow to

estimate early-life mortality (*Appendix S1*: *Figure S3*). Therefore, we used a mixed

modelling approach to describe offspring survival. We first modelled the offspring's

probability of early-life survival (from birth to week four of life; used as a proxy of

frailty) using a GLMM. Here, the response represented whether an individual offspring

survived their first four weeks of life (i.e., probability of survival: survived = 1, died =

0). We also modelled offspring juvenile survival (from week four post-hatching until

adulthood), where the response represented whether an individual offspring survived

the remainder of the juvenile period (survived =1, died = 0).

Then, conditional on the offspring surviving their first four weeks, we modelled their

total lifespan and mortality using a dual Weibull modelling approach. First, we

modelled mean offspring longevity using a GLMM, with the *brms* Weibull model

allowing for the inclusion of multiple covariates, random terms, and tests of selective

disappearance. However, under *brms,* we could not place covariates on the shape

parameter (k; *analogous to b0*) as this quickly inflated model complexity, with the

model failing to converge as indicated by low ESS values (<100) and $\hat{R}$ values > 1.5.

Here, we only estimated the effect of the model covariates on the trait mean (μ).

Subsequently, we then used the Weibull-Makeham model parameterised by *BaSTA* to

567   describe variation in the offspring's baseline (*c*) and age-specific mortality (*b1* and

568   *b0*).

GLMMS on offspring traits

Models on each offspring trait used different likelihoods and subsets of available data

(*Table 1*), but the covariate structure remained comparable across GLMMs on the

following: early-life survival, juvenile survival, total lifespan, development time,

adult mass, and adult lifespan. Here, we included, as fixed-effects: *Δage*, $\overline{age}$, and

the parents' temperature treatment (*Temp*: three-level factor; "25.5", "28", and

"30.5°C"). For development time, adult mass, and adult lifespan, we also included

offspring sex (two-level factor: "Male", "Female") as a fixed effect. Meanwhile, for

adult mass, the mother's adult mass (grams) was additionally fitted as a fixed effect.

Our global model on offspring total lifespan did not include offspring sex, to allow for

the inclusion of unsexed nymphs that died before adulthood. Therefore, we

conducted a subset analysis on the sexed adults, enabling offspring sex to predict

variation in their total lifespan. In all models, we included parent pair ID as a random

intercept and fitted *Δage* as a random slope.

Where possible, in the location component, we tested support for the three-way

interaction between *offspring sex x Temp x Δage*, including all lower-order

interactions between these terms. We then fitted two additional models including

either the two-way interaction between *Temp x Δage* or *offspring sex x Δage*, to test

whether parental age effects differed among temperature treatments or between

offspring sexes. For the scale component, to prevent model overparmaterisation, we

only tested the two-way interactions between *Temp x Δage* or *offspring sex x Δage*.

All interactions were removed from our final inference, as they did not improve

predictive performance. However, we always report estimates for the *Temp x Δage*

interaction from the location component, as this was central to answering whether

parental age effects exhibited any environmental heterogeneity.

For the GLMMs on offspring fecundity and hatching success, we only included the

following fixed-effects: the parents' age category (two-level factor: "Early-aged",

"Late-aged") and the parents' temperature treatment. Additionally, we tested

support for the two-way interaction between these two-terms, before we dropped the

interaction from the final model to ease the interpretation of single effect predictors.

Here, we never included interactive terms in the scale component to retain model

identifiability, as these models only used a small subset of individuals. In likelihoods

containing a variance or zero-inflation component (σ, ϕ, or z), we initially placed

both *Δage* (or the parents' age category for reproductive traits) and *Temp* as fixed

effects. However, *Temp* was subsequently removed from all scale and zero-inflation

models, as its effect on trait variance was weakly estimated.

Offspring mortality models

We used Weibull-Makeham models specified by *BaSTA* to describe variation in

offspring mortality, across either the offsprings total lifespan (total mortality;

estimated for offspring that survived their first four weeks of life) and adult lifespan

(adult mortality; from adult eclosion until death; *Table 1*). We first included the

isolated effects of parental age (three-level factor: "Early-aged", "Middle-aged", or

"Late-aged") or *Temp* as single-effect predictors on each mortality parameter (c, $b1$,

and *b0*), before assessing their joint effect (two-way *parental age* x *parental*

temperature interaction). To improve model convergence, parental age was binned

into a three-level factor ("Early-aged": parent mating attempts one & two, "Middle-

aged": mating attempts three to five, and "Late-aged": mating attempts six to eight).

For the subset of sexed offspring, we also tested support for the interaction between

the parents' age x offspring sex. We then compared each candidate model to the null

model, which contained no predictors. If the DIC of the null model was lower than the

alternative candidate models with covariates, we concluded that the predictors did

not explain variation in offspring mortality.

We used a weakly-informative prior structure, placing the default priors on *b0* and *c*,

while we scaled the *b1* prior. For total lifespan, (raw mean ± SD lifespan of 18.31 ±

3.97 weeks), this conversion resulted in a truncated *b1* ~ half-normal (2.57, 0.5) prior,

permitting characteristic lifespans of between 10 and 40 weeks. Meanwhile, for adult

lifespan, (raw mean ± SD adult lifespan = 9.53 ± 3.16 weeks), we used a truncated

b1~half-normal (4.94, 1) prior, permitting characteristic adult lifespans of between

five and 24 weeks. Prior sensitivity analysis confirmed these priors remained weakly

informative and did not change our posterior inferences when compared to the

default prior structure in *BaSTA* (*Appendix S1: Table S9* and *Table S21*).

Table 1. Error structures, parameters, link-functions, and sample sizes used by models on each offspring trait.

Offspring trait (response)	Likelihood & Package	Parameters & link	n	Corresponding Appendices
Early-life survival (*survived =yes/no*)	Bernoulli (*brms*)	μ (logit)	1317 offspring 78 parent pairs	*Section S4* *Model selection*: Table S2 *Estimates*: Table S3
Juvenile survival (*survived =yes/no*)	Bernoulli (*brms*)	μ (logit)	987 offspring 77 parent pairs	*Section S5* *Model selection*: Table S4 *Estimates*: Table S5
Total lifespan (*weeks*)	Weibull (*brms*)	μ (log) k (identity)	987 offspring 77 parent pairs sexed subset: 947	*Section S6.1* *Model selection*: Table S6 *Estimates*: Table S7, S8
Total mortality (*c & b1 = years b0 = unitless*)	Weibull-Makeham (*BaSTA*)	c, $b0$, $b1$ (identity)	987 offspring 77 parent pairs sexed subset: 947	*Section S6.2* *Model selection*: Table S9 *Estimates*: Table S10
Development time (*weeks*)	Lognormal (*brms*)	μ (log) σ (log)	939 offspring 77 parent pairs	*Section S7* *Model selection*: Table S11 *Estimates*: Table S12
Adult mass (*grams*)	Gaussian (*brms*)	μ (identity) σ (log)	937 offspring 77 parent pairs	*Section S8* *Model selection*: Table S13 *Estimates*: Table S14
Fecundity (*No. eggs laid*)	Negative binomial (*brms*)	μ (log) ϕ (log)	83 offspring 45 parent pairs	*Section S9* *Model selection*: Table S15 *Estimates*: Table S16
Hatching success (*No. hatched eggs per trial*)	Zero-inflated beta-binomial (*brms*)	μ (logit) ϕ (log) z (logit)	82 offspring 45 parent pairs	*Section S10* *Model selection*: Table S17 *Estimates*: Table S18
Adult lifespan (*weeks*)	Weibull (*brms*)	μ (log) k (identity)	938 offspring 77 parent pairs	*Section S11.1* *Model selection*: Table S19 *Estimates*: Table S20
Adult mortality (*c & b1 = years b0 = unitless*)	Weibull-Makeham (*BaSTA*)	c, $b0$, $b1$ (identity)	939 offspring 77 parent pairs	*Section S11.2* *Model selection*: Table S21 *Estimates*: Table S22

Note: Sample sizes describe the number of offspring from our starting pool (n = 1317

individuals) included in each model. Corresponding supplements refer to appendix

sections detailing the model selection process (e.g., choice of likelihood, evidence

for interactions) and parameter estimates.

Abbreviations: μ = location parameter; k = shape parameter (Weibull model in brms);

c = Makeham term; $b0$, $b1$ = age-specific mortality parameters; σ = residual standard

deviation; ϕ = dispersion parameter (Negative binomial/Zero-inflated beta-

binomial); z = zero-inflation parameter.

3. Results

We report estimates for fixed effects on either the mean trait outcome (location component (μ); β_1), on trait variability (scale component (σ/ϕ); γ_1), and on the probability of zero outcomes (zero-inflation component (z); δ_1). We also provide the probability of direction (pd), which indicates the direction of the effect, and the percentage of posterior estimates in the region of practical equivalence (ROPE), indicating the practical magnitude of the effect.

3.1 Offspring early-life and juvenile survival

We found no evidence that parental age affected offspring early-life survival ($\beta_{\Delta age}$ = -0.01, 95% CI [-0.18, 0.15], pd = 54.95%, 100% in ROPE; *Figure 1B, Appendix S1: Figure S4*) and uncertain evidence for a slight positive effect on juvenile survival ($\beta_{\Delta age}$ = 0.30, 95% CI [-0.04, 0.65], pd = 95.62%, 23.44% in ROPE; *Figure 1B, Appendix S1: Figure S5*). Additionally, there was limited evidence for an interactive effect of parental age and temperature on either early-life survival ($\beta_{\Delta age:Temp\ (28.0°C)}$ = 0.07, 95% CI [-0.33, 0.47], pd = 62.47, 63.53% in ROPE; $\beta_{\Delta age:Temp\ (30.5°C)}$ = 0.01, 95% CI [-0.39, 0.40], pd = 52.81%, , 66.53% in ROPE) or juvenile survival ($\beta_{\Delta age:Temp\ (28.0°C)}$ = 0.03, 95% CI [-0.72, 0.80], pd = 52.22%, 38.11% in ROPE; $\beta_{\Delta age:Temp\ (30.5°C)}$ = -0.30, 95% CI [-1.06, 0.44], pd = 78.03%, 30.20% in ROPE).

Figure 1. A) Overview of measured offspring traits. B) Standardised mean

difference (i.e., effect size) of the parents' *Δage* on each offspring trait. Points

represent the standardised mean difference and error bars show the 95% credible

intervals. Samples sizes are shown as the number of offspring/number of parent pairs.

Pairwise comparisons are based on the predicted difference in the average trait value

(μ) between early-conceived (-1.96 SDs from the mean parental age) and late-

conceived (+1.96 SDs from the mean parental age) offspring; a difference in parental

age of ~8 weeks. For fecundity and hatching success, pairwise comparisons are based

on the mean difference between the two categorical parental age groups ("early-

aged" versus "late-aged"). All effects were standardised by the standard deviation of

the response. For traits modelled on the log-odds scale (i.e., hatching success), the

credible intervals are shown on the response scale and appear wider due to the non-

linear back-transformation. Effects are strong when the credible intervals are further

away from zero, while intervals overlapping zero show greater uncertainty.

3.2 Offspring total lifespan and mortality

There was uncertain evidence for a positive parental age effect on the offspring's

total lifespan ($\beta_{\Delta age}$ = 0.02, 95% CI [0.00, 0.03]; *Appendix S1: Figure S6*). Although this

increase was well supported (pd = 98.84%), the effect was mostly negligible (96.19% in

ROPE; 0.03 log-weeks per SD increase in parental age). Post-hoc comparisons revealed

little difference in total lifespan, with the latest-conceived offspring (+1.96 SDs from

mean parental age) surviving for a median 18.80 weeks (95% HDI [18.30, 19.40

weeks]), compared with 17.70 weeks (95% HDI [17.20, 18.30 weeks]) for the earliest-

conceived offspring (-1.96 SDs from mean parental age; *Figure 1B*). There was limited

evidence this parental age effect differed across temperature treatments ($\beta_{\Delta age:Temp}$

$_{(28.0°C)}$ = -0.03, 95% CI [-0.06, 0.01], pd = 93.60%, 51.39% in ROPE; $\beta_{\Delta age:Temp\ (30.5°C)}$ = -

0.03, 95% CI [-0.06, 0.00], pd = 96.47%, 43.83% in ROPE). These findings were further

supported by our mortality models, which found only weak positive effects of parental

age on both offspring baseline (*c*) and age-specific mortality (*b0* and *b1*; *Appendix S1:*

Section S6.2).

3.3 Offspring development time

Offspring development time decreased with increasing parental age ($\beta_{\Delta age}$ = -0.04, 95%

CI [-0.05, -0.03]; *Figure 2A, Appendix S1: Figure S11*), with this effect being strongly

supported (pd = 100% and 0% in ROPE; ±0.01 log weeks per SD increase in parental

age). The latest-conceived offspring (+1.96 SD parental age) eclosed at 8.43 weeks on

average (95% HDI [8.23, 8.64 weeks]) compared with 9.81 weeks (95% HDI [9.58, 10.07

weeks]) for the earliest-conceived offspring (-1.96 SD parental age; *Figure 1B, Figure*

2B). This parental age effect was likely independent of the parents' temperature

treatment ($\beta_{\Delta age:Temp\ (28.0°C)}$ = -0.01, 95% CI [-0.04, 0.02], pd = 77.67%, 44.35% in ROPE;

$\beta_{\Delta age:Temp\ (30.5°C)}$ = 0.00, 95% CI [-0.03, 0.02], pd = 57.75%, 56.34% in ROPE).

Figure 2. A) The effect of parent's age on offspring's mean development time. On

the x-axis is the parents' age at reproduction (mean-centred) and on the y-axis is

offspring development time (weeks). Small dots are raw data while large dots are raw

means ± SE. The lines and ribbons represent the posterior mean ± 95% CI. The colours

represent the parental age effect: red dashed line = between-parent, blue solid line =

within-parent. **B) The effect of the parents' age at reproduction on the cumulative**

incidence of offspring adult emergence. The x-axis depicts offspring age (weeks),

while the y-axis displays the cumulative incidence of adult emergence, *F(x).* Lines and

710 ribbons are the posterior predictions for *F(x)*. The colours represent within-parent

711 deviations in age relative to the parent's mean age: red = "early-aged" (-1.96 SDs

712 from the mean parental age), green = "middle-aged" (the mean parental age), blue =

713 "late-aged" (+1.96 SDs from the mean parental age). The dashed lines represent the

714 posterior median estimate for development time per parental age category. N = 939

715 offspring from 77 parent pairs.

3.4 Offspring adult mass

717 Not only did offspring from older parents accelerate development; they may have

718 done so at the expense of adult mass ($\beta_{\Delta age}$ = -0.02, 95% CI [-0.04, -0.01]; *Figure 3A*,

719 *Appendix S1: Figure S12*). Although consistently negative (pd = 99.99%), the strength

720 of this effect was uncertain (24.66% in ROPE; ±0.02g per SD increase in parental age).

721 Post-hoc comparisons revealed the latest-conceived offspring (+1.96 SD parental age)

722 were the smallest at 0.71 g (95% HDI [0.69, 0.74 g]), while earliest-conceived

723 offspring (-1.96 SD parental age) weighed, on average, 0.80 g (95% HDI [0.77, 0.83 g]),

724 a mean difference of 11.77% (95% HDI [5.60, 18.01%]; *Figure 1B*). We found limited

725 evidence that this parental age effect depended on the parents' temperature

726 treatment ($\beta_{\Delta age:Temp\ (28.0°C)}$ = -0.02, 95% CI [-0.05, 0.01], pd = 78.03%, 55.54% in ROPE;

727 $\beta_{\Delta age:Temp\ (30.5°C)}$ = -0.02, 95% CI [-0.05, 0.00], pd = 57.27%, 33.87% in ROPE).

Figure 3. A) The effect of parent's age on offspring adult mass. On the x-axis is the

parents' age at reproduction (mean-centred) and on the y-axis is offspring adult mass

(g). Small dots represent raw data, large dots are raw means ± SE. Lines and ribbons

represent model predictions for the posterior mean ± 95% CI. The colours reflect the

type of parental age effect: red dashed line = between-parent, blue solid line =

within-parent. B) The effect of the parents' age at reproduction on the distribution

of offspring adult mass (probability density). On the x-axis is offspring adult mass

(g), while the y-axis shows probability density, *f(x)*. Lines and ribbons represent

posterior predictions for *f(x)*. The colours represent within-parent deviations in age

relative to the parent's mean age: red = "early-aged" (-1.96 SDs from their mean

parental age), green = "middle-aged" (their mean parental age), blue = "late-aged"

(+1.96 SDs from their mean parental age). The dashed lines represent the posterior

741   median estimate for adult mass per parental age category. N = 937 offspring from 77

742   parent pairs.

3.5 Offspring reproductive success

3.5.1 Offspring fecundity

Increasing parental age may have improved offspring fecundity, although this effect

carried broad posterior uncertainty ($\beta_{parental\ age\ (late\text{-}aged)}$ = 0.36, 95% CI [-0.04, 0.75];

Figure 4A, Appendix S1: Figure S13). Even though the direction of the effect was

uncertain (pd = 95.79%, *Figure 1B*), the effect may be moderate (11.99% in ROPE;

±0.13 log-eggs). Post-hoc comparisons revealed that offspring from late-aged parents

laid a median 342 eggs (95% HDI [246, 449 eggs]), compared with the 243 eggs (95%

HDI [177, 317 eggs]) produced by offspring from early-aged parents. There was no

evidence for a differential effect of parental age between the parent's temperature

treatments ($\beta_{parental\ age\ (late\text{-}aged):\ Temp\ (28.0°C)}$ = 0.25, 95% CI [-0.42, 0.91], pd = 76.87%,

24.83% in ROPE; $\beta_{parental\ age\ (late\text{-}aged):\ Temp\ (30.5°C)}$ = -0.25, 95% CI [-0.93, 0.44], pd =

76.27%, 23.80% in ROPE).

Figure 4. A) The effect of parental age on the mean number of eggs laid by the offspring. On the x-axis is the parents' age category, and on the y-axis is number of eggs laid by the offspring. Large dots represent model predictions ± 95% CI for mean offspring fecundity. Small dots are raw data. **B) The effect of parental age on the distribution of offspring fecundity (probability mass, *p(x)*).** Grey bars represent the median probability mass that the number of eggs laid by the offspring falls within each bin (binwidth = 50-eggs). The vertical-coloured bars are the 95% CI's around the probability mass, coloured by parental age (red = early-aged, blue = late-aged). Dashed lines represent the posterior median for offspring fecundity in each parental age group. Eggs from n = 83 offspring descending from 45 parent pairs.

3.5.2 Offspring hatching success

Parental age was associated with higher offspring hatching success ($\beta_{parental\ age\ (late-aged)}$ = 0.74, 95% CI [0.17, 1.33]; *Appendix S1: Figure S14*). This increase received strong support (pd = 99.42% and 0.42% in ROPE; ±0.18), despite exhibiting broad posterior uncertainty. The eggs laid by the latest conceived offspring had a median hatching probability of 0.25 (95% HDI [0.18, 0.33]), compared with 0.14 (95% HDI [0.08, 0.21]) for earlier-conceived offspring. However, this estimate does not account for the zero-inflation component of the model (i.e., complete reproductive failure). Although parental age showed little association with complete hatching failure ($\delta_{parental\ age\ (late-aged)}$ = -0.06, 95% CI [-1.17, 1.23], pd = 56.44%), inclusion of this component reduced the predicted difference in median hatching success between the two groups. Here, the average hatching probability dropped to 0.20 (95% CI [0.12, 0.29]) for later-conceived offspring and to 0.11 (95% CI [0.05, 0.18]) for earlier-conceived offspring (*Figure 5A*). When combining hatching probability with the total number of eggs laid, later-conceived offspring may have hatched a greater number of eggs, with an average of 56.80 hatched eggs (95% CI [33.50, 84.10]), compared to 31.10 eggs (95% CI [15.90, 52.00 eggs]) hatched by earlier-conceived offspring (*Figure 5B*). Finally, the observed parental age effect was broadly consistent across the parent's temperature treatments ($\beta_{parental\ age\ (late-aged):\ Temp\ (28.0°C)}$ = 0.46, 95% CI [-0.60, 1.49], pd = 79.96%, 19.51% in ROPE; $\beta_{parental\ age\ (late-aged):\ Temp\ (30.5°C)}$ = -0.32, 95% CI [-0.70, 1.33], pd = 73.58%, 73.58% in ROPE).

Figure 5. A) The effect of parental age on the proportion of eggs hatched by the

offspring. On the x-axis is the parents' age, and on the y-axis is the proportion of

eggs that the offspring hatched (accounting for the zero-inflation component). Large

dots represent the posterior mean ± 95% CI for hatching success. Small dots are raw

data. **B) The effect of parental age on the number of eggs hatched by the**

offspring. On the x-axis is the parents' age category, while on the y-axis is the

number of eggs that hatched. Here, we present the posterior mean predictions for

number of eggs hatched, estimated as a combination of the total number of eggs laid

by offspring in each group and their hatching probability. **C) The effect of parental**

age on the distribution of hatching success (probability mass, *p(x)*). Grey bars

represent the median probability that the number of eggs hatched by the offspring

801 falls within each bin (binwidth = 20-eggs). The vertical-coloured bars are the 95% CI's

802 around the probability mass, coloured by parental age (red = early-aged, blue = late-

803 aged). Dashed lines represent the median number of eggs hatched by offspring in each

804 parental age group. Model predictions for f(x) are visualised at the population median

805 clutch size (240 eggs). Eggs from n = 82 offspring descending from 45 parent pairs.

3.6 Offspring adult lifespan and age-dependent mortality

Given that increasing parental age has limited effect on the offspring's total lifespan

(*Figure 1B*), but accelerates their development time (*Figure 2A*), we would expect

later-conceived offspring to exhibit prolonged adult lifespans, opposite of a Lansing

effect. We found that increasing parental age extended the offspring's mean adult

lifespan ($\beta_{\Delta age}$ = 0.06, 95% CI [0.04, 0.08]; *Appendix S1: Figure S15*), with this increase

receiving strong support (pd = 100% and 5.27% in ROPE; ±0.04 log-weeks per SD

increase in parental age). Post-hoc comparisons revealed that the latest-conceived

offspring (+1.96 SD parental age) displayed a median adult lifespan of 10.60

weeks (95% HDI [10.17, 11.15 weeks]) compared to 8.51 weeks (95% HDI [8.12, 8.90

weeks]) for the earliest-conceived offspring (-1.96 SD parental age; *Figure 1B*). This

positive parental age effect was likely independent of the parents' temperature

($\beta_{\Delta age:Temp\ (28.0°C)}$ = -0.03, 95% CI [-0.08, 0.02]; pd = 86.27%, 72.52% in ROPE; $\beta_{\Delta age:Temp}$

$_{(30.5°C)}$ = -0.04, 95% CI [-0.09, 0.01]; pd = 93.82%, 57.19% in ROPE). However, there was

some uncertain evidence that this inverse Lansing effect may be stronger in daughters

than in sons ($\beta_{\Delta age:\ offspring\ sex\ (male)}$ = -0.04, 95% CI [-0.11, 0.04]; pd = 97.84%, 53.19% in

ROPE).

For *BaSTA* models on the offspring's adult age-specific mortality rates, the best-

supported model included the two-way interaction between parental age and

offspring sex (DIC = -2628.82), with this performing better than the model with

isolated effects of parental age (DIC = -2617.67) and the null model (DIC = -2597.37).

However, evidence for this sex-specific inverse Lansing effect was weak, with the

predictive performance of this model likely benefitting from the added inclusion of

offspring sex *(Appendix S1: Figure S17)*. The null model had improved performance

over both the parental temperature model (DIC = -2592.04) and the model including

the two-way interaction between *parental age x parental temperature* (DIC = -

2595.79; *Appendix S1: Figure S16*). For simplicity, we report mortality estimates from

the model with isolated effects of parental age (*Table 2*). Offspring from early and

late-aged parents entered adulthood with a similar baseline mortality (*c*). However,

those from late-aged parents displayed both a later onset (*b1*) and slowed rate (*b0*) of

actuarial senescence over their adult lifespan (*Table 2; Figure 6*).

Figure 6. The effect of parental age on offspring adult survival and mortality.

Shown on the right is the cumulative adult survival probability (top), with the median

adult lifespan for offspring from each parental age group (dashed lines; as estimated

from *BaSTA*), and the instantaneous adult hazard rate (bottom) against the offspring's

adult age (weeks). Lines and ribbons represent model predictions for adult survival

and mortality ± 95% CI, estimated from the posterior draws. For visualisation, we

transformed the hazard from the units 1/year to 1/week. The panels on the left show

the full posterior distributions of *b1*, *b0*, and *c*. Here, *b1* takes units 1/year, *c* is in

years, while *b0* is unitless. The colours describe the different parental age groups: red

= "early-aged", green = "middle-aged", and blue = "late-aged". N = 939 offspring

from 77 parent pairs.

 Table 2. Posterior estimates for the effect of parental age on adult mortality.

Mortality parameters	Estimate ± SE	95% CI
Makeham parameter (c)		
Parental age: Early-Aged	0.34 ± 0.21	0.03 - 0.81
Parental age: Middle-Aged	0.30 ± 0.14	0.07 – 0.60
Parental age: Late-Aged	0.18 ± 0.14	0.01 – 0.53
Scale parameter (b1)		
Parental age: Early-Aged	4.85 ± 0.10	4.65 – 5.05
Parental age: Middle-Aged	4.73 ± 0.07	4.57 – 4.88
Parental age: Late-Aged	4.37 ± 0.09	4.20 – 4.54
Shape parameter (b0)		
Parental age: Early-Aged	4.05 ± 0.28	3.54 – 4.65
Parental age: Middle-Aged	3.76 ± 0.18	3.42– 4.14
Parental age: Late-Aged	3.68 ± 0.22	3.29 – 4.13
KLDC pairwise comparisons		
Parental age comparison	c	b1	b0
Early-aged vs. Late-aged	0.73	**1.00**	**0.85**
Middle-aged vs. Early-aged	0.59	**0.80**	**0.82**
Late-aged vs. Middle-aged	0.61	**0.99**	0.55

Note: Shown for each mortality parameter is the estimate (posterior mean), the standard
errors, and the 95% CI. We also show the KLDC values for each mortality parameter to
describe the distance in the posterior distributions between each parental age category,
with estimates > 0.80 indicating genuine biological effects highlighted in bold. N = 939
offspring from 77 parent pairs.

Abbreviations: KLDC = Kullback-Leibler discrepancy calibration values; SE = Standard error;
95% CI = 95% Credible intervals

3.6 Variability of offspring traits

We found limited evidence that parental age affected the variability of the following offspring traits: development time (σ: $\gamma_{\Delta age}$ = 0.01, 95% CI [-0.05, 0.06], pd = 60.74%; *Figure 2B*), adult mass (σ: $\gamma_{\Delta age}$ = -0.02, 95% CI [-0.07, 0.03], pd = 76.63%; *Figure 3B*), or fecundity (ϕ: $\gamma_{parental\ age\ (late-aged)}$ = 0.27, 95% CI [-0.34, 0.88], pd = 81.22%; *Figure 4B*). However, offspring from late-aged parents may have exhibited increased precision (ϕ: *i.e., reduced variability*) in their hatching success. Here, most of their hatchling counts clustered towards the posterior mean, although the 95% CI's overlapped zero ($\gamma_{parental\ age\ (late-aged)}$ = 1.04, 95% CI [-0.01, 2.23], pd = 97.44%; *Figure 5C, Appendix S1: Figure S14*).

3.7 Selective disappearance of parent phenotypes

We found evidence of selective disappearance for the following offspring traits: juvenile survival (Δslope $_{\overline{age}-\Delta age}$= -0.66, 95% CI [-1.06, -0.26], *Post.Prob* = 1.00), development time (Δslope $_{\overline{age}-\Delta age}$= 0.03, 95% CI [0.02, 0.05], *Post.Prob* = 1.00; *Figure 2A*), and adult lifespan (Δslope $_{\overline{age}-\Delta age}$= -0.04, 95% CI [-0.06, -0.01], *Post.Prob* = 0.99). Here, the slope for the between-individual age effect progressed in the opposite direction (e.g., juvenile survival; *Appendix S1: Table S5*) or was considerably flatter (e.g., development time and adult lifespan; *Appendix S1: Table S12 and Table S20*) than the within-individual age estimate. Thus, if we did not control between-individual heterogeneity in the parents mean age at reproduction, then this could have masked the within-parent trajectories we observed.

Discussion

Our results question the universality of parental effect senescence and contribute to the growing knowledge that later-conceived offspring exhibit accelerated life-histories (Bouwhuis et al. 2010; Plaistow et al. 2015; Kroeger et al. 2020; Travers et al. 2021; Anderson et al. 2022). Irrespective of their parents' thermal environment, which induced experimental variation in parental mortality rate, the earliest- and latest-conceived offspring appeared demographically distinct. Offspring from old parents accelerated development, eclosed with a smaller adult mass, potentially exhibited improved reproductive investment, and, crucially, displayed an extended adult lifespan, opposite of a Lansing effect. This inverse Lansing effect arose not from a reduced initial frailty, but from a delayed onset and slowed progression of actuarial senescence. Below, we first discuss the lack of temperature induced environmental heterogeneity in our parental age effects, before outlining the following alternate explanations for our findings: do our results indicate constraints imposed by parental senescence or an adaptive late-life shift in the parents' reproductive strategy?

Environmental heterogeneity and parental age effects

Although it was clear that increasing temperatures exacerbated parent mortality, we found no convincing evidence that the parents' thermal environment mediated our parental age effects. According to the metabolic theory of ecology (Brown *et al.*, 2004), warming temperatures accelerate metabolic processes in ectotherms, resulting in the excessive generation of damaging reactive oxygen species (ROS; Munch & Salinas, 2009; Burraco et al. 2020). Under chronic exposure to high temperatures, this

could have accelerated the parent's rate of actuarial senescence through increased

ROS damage to lipids, proteins, and telomeres (Mair et al. 2003; Conti, 2008; Flouris

& Piantoni, 2015; Keil et al. 2015; Friesen et al. 2022). Alternatively, temperature

may have directly increased the parents' baseline mortality, without accelerating

senescence (Bowler & Terblanche, 2008). In the first scenario, we would expect some

component of the senescent physiology of old parents to be transferred to the

offspring, inducing detrimental age effects (such as the Lansing effect). Here, we

should have found evidence for a temperature by parental age interaction on

offspring adult lifespan, which is not supported by our data. However, if the Lansing

effect is best understood as a manifestation of accelerated actuarial senescence

(Kern, 2001), and if increasing temperatures only directly affected the parents

baseline mortality, independently of senescence, then this could explain the lack of

any environmental variation in our parental age effects. Hence, we believe the most

parsimonious explanation for the lack of a temperature mediated parental age effect

is that temperature did not increase parental mortality through physiologically

accelerated ageing, but rather through a direct environmental effect on age-

independent baseline-mortality

Parental age as a constraint: Egg provisioning and resource allocation trade-offs

Our inverse Lansing effect may have arisen from late-life improvements to pre-natal

provisioning. Older mothers could have displayed a terminal investment strategy

(Clutton-Brock, 1984; Velando et al. 2006; Jehan et al. 2021) or improved in body

condition with age (Stahlschmidt et al. 2013). Under this scenario, older mothers

would provision their eggs with high concentrations of macro and micronutrients

(e.g., lipids, carbohydrates, proteins, vitamins), thereby improving offspring survival

(Clutton-Brock, 1984; Mousseau & Fox, 1998; Gilbert & Manica, 2010). However, this

explanation appears unlikely given the substantial evidence for reproductive

senescence across insects (Begon & Parker, 1986; Fox & Czesak, 2000; Rivero et al.

2001; Fox et al. 2003; Bloch Qazi et al. 2017; Muller et al. 2017; Noguera, 2021), and

from our own system *(in prep)*. Even in instances where maternal provisioning

increases with age, the Lansing effect often remains (Plaistow et al. 2015; Anderson

et al. 2022), suggesting any age-dependent improvements in maternal provisioning do

not necessarily alleviate, and may even exacerbate, offspring senescence.

It is entirely possible that the Lansing effect (or inverse thereof) can arise from the

offspring's response to age-dependent changes in maternal pre-natal provisioning,

rather than from the inheritance of their old parents' senescent phenotype (Monaghan

et al. 2020). In this scenario, parental provisioning modulates how offspring resolve

existing life-history trade-offs between the competing demands of growth,

reproduction, and somatic maintenance (Stearns, 2000; Anderson et al. 2022).

Previous research in *Daphnia pulex* found that later-conceived offspring exhibited a

truncated lifespan, yet this was correlated with an increased body size, accelerated

growth rate, and improved early-life fecundity (Plaistow et al. 2015). Similar parental

age effects on body size, development, and reproductive scheduling have been

demonstrated in *Caenorhabditis elegans* and *Daphnia magna* (Travers et al. 2021;

Anderson et al. 2022), being attributed to an offspring response to late-life

improvements in the mothers' egg quality. This could indicate that increased maternal

provisioning initiates an alternate offspring growth trajectory in the latest-conceived

offspring, accelerating their growth and increasing their early-life productivity at the

consequence of reducing survival (Boersma & Wit, 1997; Metcalfe, 2003; Plaistow et

al. 2015). Increased investment in growth or reproduction is well-known to

exacerbate senescence, being linked to reduced antioxidant defences (Blount et al.

2003, 2016), increased telomere attrition (Hall et al. 2004; Monaghan & Haussmann,

2006; Houben et al. 2008; Monaghan, 2014), and heightened oxidative stress

responses (Smith et al. 2016; Janssens & Stoks, 2020). Additionally, there may be

maintenance costs that scale with size, such as an increased physiological cost of

moulting (Hessen & Alstad Rukke, 2000). Yet there is a major caveat to interpreting

these studies relative to our findings; the species used display indeterministic growth,

with maternal age and adult size being tightly correlated (Bolanowski et al. 1981;

Glazier, 1992; Plaistow et al. 2015; Travers et al. 2021). Any improvements in pre-

natal provisioning could reflect the larger size of older mothers, rather than late-life

changes to provisioning.

Within our study we found an inverse pattern; later-conceived offspring exhibited

slowed senescence and a smaller adult mass. Could this reflect reductions in pre-natal

provisioning by older mothers? Anderson et al. (2022) showed that the Lansing effect

can operate in both directions between clones of *D.magna* depending on how

maternal provisioning changes with age, with the inverse Lansing effect fully

explained by reduced maternal lipid provisioning of the neonates. Brief periods of

dietary restriction have been widely demonstrated to extend lifespan, by slowing

metabolic processes, decelerating growth rates, and reducing body size (Masoro,

2005, 2006; Spindler, 2010; Bock et al. 2019). Thus, our inverse Lansing effect may

not arise from parental senescence *per se*; meaning we should expect to find similar

patterns in younger mothers if their egg provisioning was somehow experimentally

reduced (e.g., through caloric restriction; Gribble et al. 2014; Hibshman et al. 2016).

However, under this explanation, we would also expect later-conceived offspring to

display both reduced reproductive performance and prolonged development, with this

relationship being well established across insects (Zhang et al. 2019). For example,

reduced egg provisioning has been frequently demonstrated to delay maturity in

Callosobruchus maculatus (Fox, 1994; Fox et al. 2003; Lind et al. 2015). We found

that increasing parental age did not constrain offspring fecundity (there was even

limited evidence for an improvement in hatching success), and the later-conceived

offspring accelerated, rather than slowed, their development. Finally, and crucially,

this increased caloric restriction does not necessarily explain the ameliorated

senescence of later-conceived offspring. For example, in *Drosphilla melanogaster*,

dietary restriction does not slow the accumulation of senescent damage, instead

extending longevity by lowering the immediate risk of death (i.e., reduced baseline

mortality), with individuals carrying no memory of their past restricted feeding when

switched to a normal diet (Mair et al. 2003). Thus, it appears unlikely that reduced

pre-natal provisioning explains the accelerated life-histories and slowed senescence

we observed in latest-conceived offspring.

Alternatively, given we did not measure the offspring's initial size at hatching, or

subsequent growth rates, it remains possible that offspring from old parents exhibited

both slowed growth rates and an earlier adult emergence by eclosing at a smaller

body size (Vega-Trejo et al. 2018), a pattern supported by our data (*Figure 3*).

Furthermore, we were unable to tease apart the correlation between reproductive

history/experience and parental age, which covaried in our experimental design

(*Appendix S1: Figure S2*). Given the known importance of reproductive history in

influencing physiological condition and future reproductive investment (Bell, 1980;

Gershman, 2008; Stiver & Alonzo, 2009), it is possible that any deteriorations in pre-

natal provisioning reflect the cost of accumulated reproductive effort rather than an

effect of parental age *per se* (Monaghan et al. 2020). Finally, if mothers did

display late-life reductions in pre-natal provisioning, then our inverse Lansing effect

could have instead manifested from the selective disappearance of offspring

phenotypes *within* families (Fox et al. 2003). In this situation, later-conceived

offspring experience heightened mortality during embryogenesis, consequently

leaving a non-random subset of high-quality or long-lived offspring phenotypes in the

hatchling pool (*viability selection hypothesis*; Hadfield, 2008; Mojica & Kelly, 2010).

However, we believe viability selection is unlikely to explain our findings as the initial

mortality in the first four weeks after hatching was independent of parental age

(*Figure 1B*), suggesting no increased mortality rate in frail individuals conceived by

old parents.

Indirect parental age effects

It is plausible to speculate that the Lansing effect (and our inverse Lansing effect)

manifests indirectly, through effects of parental age on offspring adult size. This is

supported given the numerous studies finding that parental age is either positively or

negatively correlated with offspring size, yet so far these effects on size are

consistently in the opposite direction to those on adult lifespan (Plaistow et al. 2015;

Noguera, 2021; Anderson et al. 2022). Likewise, those finding that the offspring's

adult lifespan is independent of parental age also tend to find the joint absence of

any parental age effects on offspring adult size (Tregenza et al. 2026). This

relationship has previously been described by Gershman et al. 2022. This suggests

that parental age could either directly affect multiple offspring phenotypic traits,

including adult size, independently of each other (*Figure 7A*) or operate entirely

indirectly, entirely through a direct parental age effect on offspring development

time (*Figure 7B*).

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Figure 7. The non-mutually exclusive causal relationships between parental age,

offspring development time, and offspring adult lifespan (Cause → Effect). Direct

effects are shown as solid blue lines, while indirect effects are shown as grey dashed

lines. The sign describes the direction of the relationship: + = positive relationship, - =

negative relationship. **A) Direct parental age effects.** Under this scenario, parental

age directly affects all offspring phenotypic traits independently. There also exists a

trade-off between reproduction and adult lifespan. **B) Indirect parental age effects**

on offspring adult size and lifespan, arising from causal effects of parental age on

offspring development time. Here, any effect of parental age on offspring adult size

and adult lifespan operates entirely indirectly, through the direct parental age effect

on the time to eclosion. Diagrams adapted from Gershman et al. (2022).

The pathway to causality in our system may have progressed as follows: first, parental

age could directly accelerate offspring development times, with it being adaptive for

later-conceived offspring to exhibit an earlier adult emergence (e.g., due to seasonal

time constraints; *discussed below*). Second, this accelerated development limits the

time available for somatic growth, with the latest-produced offspring being smaller

on reaching adulthood. Indeed, this positive correlation between development time

and adult size has been well-established across insects (Mousseau & Dingle, 1991;

Roff, 2000). Finally, as later-conceived offspring may eclose with a smaller adult

mass, they could have incurred less damage from ROS during development, thereby

slowing senescence (*growth-lifespan trade-off*; Metcalfe & Monaghan, 2001; Metcalfe,

2003). Here, our inverse Lansing effect arises indirectly, through causal parental age

effects on offspring development time (Gershman et al. 2022). Alternatively, parental

age may have direct detrimental effects on offspring adult longevity, but this Lansing

effect is masked by both direct and indirect (i.e., through development time)

negative effects of parental age on offspring adult size. Although such a causal

pathway seems intuitive, the observed negative parental age effect on adult mass was

small. Approximately 20% of the posterior distribution fell within the region of

practical equivalence (ROPE). As such, a meaningful proportion of posterior estimates

corresponded to a negligible change in adult mass.

It is possible that, alongside their earlier adult emergence, later-conceived offspring

improved their early-life fecundity, as is being increasingly evidenced (Bouwhuis et al.

2010; Plaistow et al. 2015; Travers et al. 2021; Dutt & Laird, 2024), but such

improvements were partially offset by their smaller adult mass. Across invertebrates,

a smaller soma constrains reproduction, owing to how the ovarian tissues scale with

somatic size (Honěk & Honek, 1993; Berger et al. 2012; Sturm, 2016). Although we

found limited evidence for a positive parental age effect on offspring fecundity, eggs

laid by later-conceived offspring may have exhibited improved hatching success.

Crucially, we cannot infer whether the reproductive scheduling of later-conceived

offspring was shifted to increase early-life productivity, as we only measured

reproductive effort from a single mating attempt at an intermediate adult age of ~5

weeks, e.g. when the offspring were neither young nor old. At the very least,

increasing parental age did not appear to constrain offspring fecundity and may even

improve the offspring's *per-capita* reproductive investment.

Regardless of the causal pathway, our explanation here assumes a direct effect of

parental age on offspring development time and early-life fecundity. Therefore, is it

adaptive for later-conceived offspring to accelerate their life-histories?

Adaptive parental age effects: *shifts to a fast pace-of life*

Collectively, our results suggest that later-conceived offspring exhibit a fast pace-of-life strategy. This phenomenon appears taxonomically widespread, being observed in duckweed (*Lemna minor*; Dutt & Laird, 2024), black rockfish (*Sebastes melanops*; Berkeley et al. 2004), great tits (*Parus major*; Bouwhuis et al. 2010), yellow-bellied marmots (*Marmota flaviventer*; Kroger et al. 2020), Daphnia (*D. pulex* and *D.magna*; Plaistow et al. 2015; Anderson et al. 2022), Nematodes (*C.elegans*; Travers et al. 2021), crickets (*Gryllus vocalis*; Gershamn et al. 2022*)*, milkweed bugs (*Oncopeltus fasciatus*; Phelan & Frumhoff, 1991*)*, zig-zag ladybirds (*Menochilus sexmaculatus*; Singh et al. 2021), and even from Lansing's original findings in *Philodina citrina* (Lansing, 1947; King, 1983), yet despite this evidence, the predominant perception remains that parental age negatively affects offspring adult longevity. We argue that parental age effects on the pace-of-life could be adaptive in the special case of seasonal insects with a definite environmental end point to reproduction.

For example, it may be adaptive for old parents to produce offspring with accelerated life-histories if they are more prevalent late in the season (Phelan & Frumhoff, 1991; Zehnder et al. 2007; Gershman et al. 2022). In this circumstance, seasonal deteriorations in environmental conditions and increasing resource scarcity limits the time available for older parents to complete reproduction (*"time-stress"* hypothesis; Gotthard, 2008; Metcalfe et al. 2002). In response to these time-constraints, old parents may influence offspring phenotypes through one of two pathways; they may conceive offspring that undergo diapause (Bigelow, 1962; Mousseau & Roff, 1989; Mousseau & Dingle, 1991) or produce offspring with uninterrupted development, that

exhibit an earlier adult emergence and increased early-life productivity, with these

offspring seeing some fitness returns before the season deteriorates (Roff, 1980;

Phelan & Frumhoff, 1991; Zehnder et al. 2007). The latter strategy is common within

species which survive through the winter as adults rather than arresting development

and diapausing as nymphs. Uninterrupted development carries substantial costs; for

every directly-developing generation initiated before winter, the length of the season

suitable for growth and reproduction increasingly narrows (Nylin et al. 2009). Thus,

later-conceived offspring may experience strong selection to accelerate development

and redirect investment from growth to current reproduction, resulting in them

eclosing at a less-than-optimal size (Roff, 1980; Nylin et al. 1989; Abrams et al., 1996;

Pfenning et al. 2007; Larsdotter Mellström et al. 2010). Alternatively, later-conceived

offspring may encounter heightened intra-specific competition from their earlier-

conceived siblings, who express an age-related competitive advantage (Plaistow et al.

2007; Benton et al. 2008; Travers et al. 2021). The earlier emergence of offspring

from young parents creates a growth race among later-conceived offspring, who are

then forced to eclose as adults earlier in life (Stearns, 2000; Pásztor et al. 2022).

These time constraints on growth may be avoided in species that diapause as nymphs,

as they could synchronise their growth in the following year. For example, recent

evidence from a wild population of *G.campestris* suggests that increasing parental age

has no effect on the offspring's adult size and, subsequently, adult lifespan (Tregenza

et al. 2026). However, despite being a close relative of *G.bimaculatus* (hybrids

between the two remain fertile; Tyler et al. 2013), *G.campestris* is univoltine, with

the nymphs undergoing an obligate diapause at their penultimate instar (Martínez-

Viejo et al. 2024), and the later-conceived offspring here may not experience the

same time limitations on growth and reproduction as in our uninterrupted developing

multivoltine *G.bimaculatus*.

If development time is under strong selection, we would expect to see two patterns;

not just a reduction in mean development time but also reduced variability in

development time. However, despite strong effects of parental age on the average

development time, there was limited evidence that *variation* in offspring

development time changed with parental age. Rather, we found substantial variability

in development time regardless of parental age. This lack of canalisation could

suggest there is little selection for later-conceived offspring to accelerate

development, or, inversely, there may be strong selection on offspring development

time across all parental ages. It is also conceivable that selection actively maintains

variability in development time, irrespective of parental age, especially if the

seasonal environment varies unpredictably (Philippi & Seger, 1989; Simons &

Johnston, 2006; Crean & Marshall, 2009).

Scope for future research

Producing offspring with accelerated life-histories late in the season could be

adaptive given that *G.bimaculatus* is a multivoltine species, enabling another fully

functional generation to be realised before conditions deteriorate. From an

evolutionary perspective, an increase in the number of generations per year

facilitates rapid population growth and a greater potential for adaptation (Altermatt,

2010). What remains is in understanding whether these observed non-genetic effects

of parental age can accumulate over multiple generations to influence grandoffspring

phenotypes. Previous evidence for the transgenerational inheritance of parental age effects remains ambiguous, with some finding immediate parental age effects while, in others, the effects persist across multiple generations (Bloch Qazi et al. 2017; Wylde et al. 2019; Travers et al. 2021; Bleu et al. 2022). Evolutionary theory predicts that if parental age effects do accumulate, then they may alter selection gradients on late-life reproduction (Kern et al. 2001; Plaistow et al. 2007; Moorad & Nussey, 2016; Barks & Laird, 2020). Additionally, given our emphasis on the adaptive importance of parental age effects in life-history evolution, it would be necessary to perform a similar longitudinal study under a natural environment. Our study population was maintained under captive conditions, where no predators were present and food was provided *ad libitum,* so that two of the main drivers of life history variation were controlled for but not directly examined. The resolution of these life history trade-offs might work in different, or even opposite, ways when predators and food limitation are introduced into the system (Zajitschek et al. 2020; Promislow et al. 2022). Currently, little is known on the importance of parental age effects for wild insect populations, and the two studies that have investigated such effects are limited in how much they can conclude (Angell et al. 2022; Tregenza et al. 2026). Both these studies were unable to control potential sources of viability selection (including effects of parental age on early-life mortality) and only investigated parental age effects over a single generation. Establishing whether demographic parental age effects can accumulate across generations, or persist under natural environments, will be a necessary next step for establishing their ecological importance and role in the evolution of ageing rates.

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Conflict of Interest Statement

The authors declare no conflicts of interest.

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Author Contributions

Mark D. Pitt (MDP), Brendan O'Connor (BC), Timothy D. Sheen (TS), Tom Tregenza (TT), and Jelle J. Boonekamp (JJB) conceived and designed the study. MDP, BC, TDS, and JJB collected the data. MDP, BC, and JJB conducted the data analysis, while MDP and JJB wrote the first draft of the manuscript. All authors contributed extensively to revise the manuscript.

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Ethics Statement

All work involving housing, husbandry, and care of the animals involved in this study was performed at the highest standard possible. Animals were cleaned and fed

1186   weekly, and housed individually from conception, removing welfare concerns

1187   regarding overcrowding.

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1189   **Use of AI statement**

1190   Chat GPT-5.3 was used to correct syntax errors in the R-scripts and to help develop

1191   code for plotting the model outputs. The corresponding author MDP takes full

1192   responsibility over the code and has ensured it remains accurate. No AI was used in

1193   any written aspect of this manuscript.

References

Abrams, Peter A. 1991. 'The Fitness Costs of Senescence: The Evolutionary Importance of Events in Early Adult Life'. *Evolutionary Ecology* 5 (4): 343–60. https://doi.org/10.1007/BF02214152.

Abrams, Peter A., Olof Leimar, Soren Nylin, and Christer Wiklund. 1996. 'The Effect of Flexible Growth Rates on Optimal Sizes and Development Times in a Seasonal Environment'. *The American Naturalist* 147 (3): 381–95. https://doi.org/10.1086/285857.

Altermatt, Florian. 2010. 'Climatic Warming Increases Voltinism in European Butterflies and Moths'. *Proceedings of the Royal Society B: Biological Sciences* 277 (1685): 1281–87. https://doi.org/10.1098/rspb.2009.1910.

Anderson, Cora E., Morad C. Malek, Rachael A. Jonas-Closs, Yongmin Cho, Leonid Peshkin, Marc W. Kirschner, and Lev Y. Yompolsky. 2022. 'Inverse Lansing Effect: Maternal Age and Provisioning Affecting Daughters' Longevity and Male Offspring Production'. *The American Naturalist* 200 (5): 704–21. https://doi.org/10.1086/721148.

Angell, Christopher S., Rebecca Janacek, and Howard D. Rundle. 2022. 'Maternal and Paternal Age Effects on Male Antler Flies: A Field Experiment'. *The American Naturalist* 199 (3): 436–42. https://doi.org/10.1086/718236.

Barks, Patrick M., and Robert A. Laird. 2020. 'Parental Age Effects and the Evolution of Senescence'. *The American Naturalist* 195 (5): 886–98. https://doi.org/10.1086/708271.

Barreda, Santiago, and Noah Silbert. 2023. *Bayesian Multilevel Models for Repeated Measures Data: A Conceptual and Practical Introduction in R*. 1st edn. 250-60. Routledge. https://doi.org/10.4324/9781003285878.

Begon, Michael, and Geoffrey A. Parker. 1986. 'Should Egg Size and Clutch Size Decrease with Age?' *Oikos* 47 (3): 293-302. https://doi.org/10.2307/3565440.

Bell, Graham. 1980. 'The Costs of Reproduction and Their Consequences'. *The American Naturalist* 116 (1): 45–76. https://doi.org/10.1086/283611.

Benton, T. G., J. J. H. St Clair, and S. J. Plaistow. 2008. 'Maternal Effects Mediated by Maternal Age: From Life Histories to Population Dynamics'. *Journal of Animal Ecology* 77 (5): 1038–46. https://doi.org/10.1111/j.1365-2656.2008.01434.x.

Berger, David, Martin Olofsson, Magne Friberg, Bengt Karlsson, Christer Wiklund, and Karl Gotthard. 2012. 'Intraspecific Variation in Body Size and the Rate of Reproduction in Female Insects – Adaptive Allometry or Biophysical Constraint?' *Journal of Animal Ecology* 81 (6): 1244-58. https://doi.org/10.1111/j.1365-2656.2012.02010.x.

Berkeley, Steven A., Colin Chapman, and Susan M. Sogard. 2004. 'MATERNAL AGE AS A
DETERMINANT OF LARVAL GROWTH AND SURVIVAL IN A MARINE FISH, *SEBASTES*
MELANOPS'. *Ecology* 85 (5): 1258–64. https://doi.org/10.1890/03-0706.

Bigelow, R. S. 1962. 'Factors Affecting Developmental Rates and Diapause in Field
Crickets'. *Evolution* 16 (4): 396-406. https://doi.org/10.2307/2406175.

Bleu, Josefa, Sandrine Meylan, Jean Clobert, and Manuel Massot. 2022.
'Grandmaternal Age at Reproduction Affects Grandoffspring Body Condition,
Reproduction and Survival in a Wild Population of Lizards'. *Functional Ecology* 36 (4):
826–36. https://doi.org/10.1111/1365-2435.13996.

Bloch Qazi, Margaret C., Paige B. Miller, Penny M. Poeschel, Mai H. Phan, Joseph L.
Thayer, and Christian L. Medrano. 2017. 'Transgenerational Effects of Maternal and
Grandmaternal Age on Offspring Viability and Performance in *Drosophila*
Melanogaster'. *Journal of Insect Physiology* 100 (July): 43–52.
https://doi.org/10.1016/j.jinsphys.2017.05.007.

Blount, Jonathan D., Neil B. Metcalfe, Kathryn E. Arnold, Peter F. Surai, Godefroy L.
Devevey, and Pat Monaghan. 2003. 'Neonatal Nutrition, Adult Antioxidant Defences
and Sexual Attractiveness in the Zebra Finch'. *Proceedings of the Royal Society of*
London. Series B: Biological Sciences 270 (1525): 1691–96.
https://doi.org/10.1098/rspb.2003.2411.

Blount, Jonathan D., Emma I. K. Vitikainen, Iain Stott, and Michael A. Cant. 2016.
'Oxidative Shielding and the Cost of Reproduction'. *Biological Reviews* 91 (2): 483–97.
https://doi.org/10.1111/brv.12179.

Bock, Martha J., George C. Jarvis, Emily L. Corey, Emily E. Stone, and Kristin E.
Gribble. 2019. 'Maternal Age Alters Offspring Lifespan, Fitness, and Lifespan
Extension under Caloric Restriction'. *Scientific Reports* 9 (1): 3138.
https://doi.org/10.1038/s41598-019-40011-z.

Boersma, Bart, and Jan Maarten Wit. 1997. 'Catch-up Growth'. *Endocrine Reviews* 18
(5): 646-61. https://doi.org/10.1210/edrv.18.5.0313.

Bolanowski, Mark A., Richard L. Russell, and Lewis A. Jacobson. 1981. 'Quantitative
Measures of Aging in the Nematode *Caenorhabditis Elegans*. I. Population and
Longitudinal Studies of Two Behavioral Parameters'. *Mechanisms of Ageing and*
Development 15 (3): 279–95. https://doi.org/10.1016/0047-6374(81)90136-6.

Bouwhuis, S., A. Charmantier, S. Verhulst, and B. C. Sheldon. 2010. 'Trans-
Generational Effects on Ageing in a Wild Bird Population'. *Journal of Evolutionary*
Biology 23 (3): 636–42. https://doi.org/10.1111/j.1420-9101.2009.01929.x.

Bowler, Ken, and John S. Terblanche. 2008. 'Insect Thermal Tolerance: What Is the
Role of Ontogeny, Ageing and Senescence?' *Biological Reviews* 83 (3): 339–55.
https://doi.org/10.1111/j.1469-185X.2008.00046.x.

Brown, James H., James F. Gillooly, Andrew P. Allen, Van M. Savage, and Geoffrey B.
West. 2004. 'TOWARD A METABOLIC THEORY OF ECOLOGY'. *Ecology* 85 (7): 1771–89.
https://doi.org/10.1890/03-9000.

Bürkner, Paul-Christian. 2017. '**Brms** : An *R* Package for Bayesian Multilevel Models
Using *Stan*'. *Journal of Statistical Software* 80 (1): 1-28.
https://doi.org/10.18637/jss.v080.i01.

Burraco, Pablo, Germán Orizaola, Pat Monaghan, and Neil B. Metcalfe. 2020. 'Climate
Change and Ageing in Ectotherms'. *Global Change Biology* 26 (10): 5371–81.
https://doi.org/10.1111/gcb.15305.

Centeno Filho, Boaventura Lobo, Guilherme Martins Limberger, Kathellen Pintado
Esteves, Duane Barros Da Fonseca, and Fábio Everton Maciel. 2023. 'Mortality,
Metabolic Rate, and Oviposition of *Gryllus* (*Gryllus*) *Assimilis* (Fabricius, 1775)
(Orthoptera: Gryllidae) Females under Constant and Fluctuating Warm
Temperatures'. *Journal of Thermal Biology* 114 (May): 103574.
https://doi.org/10.1016/j.jtherbio.2023.103574.

Clutton-Brock, T. H. 1984. 'Reproductive Effort and Terminal Investment in
Iteroparous Animals'. *The American Naturalist* 123 (2): 212–29.
https://doi.org/10.1086/284198.

Colchero, Fernando, and James S. Clark. 2012. 'Bayesian Inference on Age-specific
Survival for Censored and Truncated Data'. *Journal of Animal Ecology* 81 (1): 139–49.
https://doi.org/10.1111/j.1365-2656.2011.01898.x.

Colchero, Fernando, Owen R. Jones, and Maren Rebke. 2012. 'BaSTA: An R Package
for Bayesian Estimation of Age-specific Survival from Incomplete Mark–
Recapture/Recovery Data with Covariates'. *Methods in Ecology and Evolution* 3 (3):
466–70. https://doi.org/10.1111/j.2041-210X.2012.00186.x.

Conti, B. 2008. 'Considerations on Temperature, Longevity and Aging'. *Cellular and*
Molecular Life Sciences 65 (11): 1626–30. https://doi.org/10.1007/s00018-008-7536-1.

Crean, Angela J., and Dustin J. Marshall. 2009. 'Coping with Environmental
Uncertainty: Dynamic Bet Hedging as a Maternal Effect'. *Philosophical Transactions of*
the Royal Society B: Biological Sciences 364 (1520): 1087–96.
https://doi.org/10.1098/rstb.2008.0237.

De Jong, G. 1995. 'Phenotypic Plasticity as a Product of Selection in a Variable
Environment'. *The American Naturalist* 145 (4): 493–512.
https://doi.org/10.1086/285752.

Debat, Vincent, and Patrice David. 2001. 'Mapping Phenotypes: Canalization,
Plasticity and Developmental Stability'. *Trends in Ecology & Evolution* 16 (10): 555–
61. https://doi.org/10.1016/S0169-5347(01)02266-2.

Díaz, Hugo, and Pedro Esponda. 2004. 'Ageing-Induced Changes in the Cortical
Granules of Mouse Eggs'. *Zygote* 12 (2): 95–103.
https://doi.org/10.1017/S0967199404002680.

Dutt, Priyanka, and Robert A. Laird. 2024 'A Demographic Assessment of the Lansing
Effect in Duckweed (*Lemna Turionifera* Landolt)'. *Biology Letters* 20 (10): 20240271.
https://doi.org/10.1098/rsbl.2024.0271.

Fay, Rémi, Julien Martin, and Floriane Plard. 2022. 'Distinguishing Within- from
Between-individual Effects: How to Use the Within-individual Centring Method for
Quadratic Patterns'. *Journal of Animal Ecology* 91 (1): 8–19.
https://doi.org/10.1111/1365-2656.13606.

Flouris, Andreas D., and Carla Piantoni. 2015. 'Links between Thermoregulation and
Aging in Endotherms and Ectotherms'. *Temperature* 2 (1): 73–85.
https://doi.org/10.4161/23328940.2014.989793.

Fox, C. W., M. L. Bush, and W. G. Wallin. 2003. 'Maternal Age Affects Offspring
Lifespan of the Seed Beetle, *Callosobruchus Maculatus*'. *Functional Ecology* 17 (6):
811–20. https://doi.org/10.1111/j.1365-2435.2003.00799.x.

Fox, C. W., and H. Dingle. 1994. 'Dietary Mediation of Maternal Age Effects on
Offspring Performance in a Seed Beetle (Coleoptera: Bruchidae)'. *Functional Ecology*
8 (5): 600-606. https://doi.org/10.2307/2389921.

Fox, Charles W. 1994. 'The Influence of Egg Size on Offspring Performance in the Seed
Beetle, *Callosobruchus maculatu*s'. *Oikos* 71 (2): 321-325.
https://doi.org/10.2307/3546280.

Fox, Charles W., and Mary Ellen Czesak. 2000. 'Evolutionary Ecology of Progeny Size
in Arthropods.' *Annual Review of Entomology* 45 (1): 341–69.
https://doi.org/10.1146/annurev.ento.45.1.341.

Fox, John, and Sanford Weisberg. *An R companion to applied regression*. Sage
publications, 2018.

Friesen, Christopher R., Erik Wapstra, and Mats Olsson. 2022. 'Of Telomeres and
Temperature: Measuring Thermal Effects on Telomeres in Ectothermic Animals'.
Molecular Ecology 31 (23): 6069–86. https://doi.org/10.1111/mec.16154.

Gabry, Jonah, Daniel Simpson, Aki Vehtari, Michael Betancourt, and Andrew Gelman.
2019. 'Visualization in Bayesian Workflow'. *Journal of the Royal Statistical Society
Series A: Statistics in Society* 182 (2): 389-402. https://doi.org/10.1111/rssa.12378.

Gabry, Jonah, Tristan Mahr. 'Bayesplot: Plotting for Bayesian models'. *R package*
version 1.11.1. https://mc-stan.org/bayesplot/.

Gelman, Andrew, Aleks Jakulin, Maria Grazia Pittau, and Yu-Sung Su. 2008. 'A Weakly
Informative Default Prior Distribution for Logistic and Other Regression Models'. *The*
Annals of Applied Statistics 2 (4): 1360-83. https://doi.org/10.1214/08-AOAS191.

Gelman, Andrew, Daniel Lee, and Jiqiang Guo. 2015. 'Stan: A Probabilistic
Programming Language for Bayesian Inference and Optimization'. *Journal of*
Educational and Behavioral Statistics 40 (5): 530–43.
https://doi.org/10.3102/1076998615606113.

Gelman, Andrew, and Donald B. Rubin. 1992. 'Inference from Iterative Simulation
Using Multiple Sequences'. *Statistical Science* 7 (4): 457-472.
https://doi.org/10.1214/ss/1177011136.

Gelman, Andrew, and Cosma Rohilla Shalizi. 2013. 'Philosophy and the Practice of
Bayesian Statistics'. *British Journal of Mathematical and Statistical Psychology* 66 (1):
8–38. https://doi.org/10.1111/j.2044-8317.2011.02037.x.

Gelman, Andrew, Daniel Simpson, and Michael Betancourt. 2017. 'The Prior Can Often
Only Be Understood in the Context of the Likelihood'. *Entropy* 19 (10): 555.
https://doi.org/10.3390/e19100555.

Gershman, Susan N. 2008. 'Sex-Specific Differences in Immunological Costs of Multiple
Mating in *Gryllus Vocalis* Field Crickets'. *Behavioral Ecology* 19 (4): 810–15.
https://doi.org/10.1093/beheco/arn040.

Gershman, Susan N., Owen G. Miller, and Ian M. Hamilton. 2022. 'Causes and
Consequences of Variation in Development Time in a Field Cricket'. *Journal of*
Evolutionary Biology 35 (2): 299–310. https://doi.org/10.1111/jeb.13971.

Gilbert, James D. J., and Andrea Manica. 2010. 'Parental Care Trade-Offs and Life-
History Relationships in Insects'. *The American Naturalist* 176 (2): 212–26.
https://doi.org/10.1086/653661.

Glazier, Douglas S. 1992. 'Effects of Food, Genotype, and Maternal Size and Age on
Offspring Investment in *Daphnia magna*'. *Ecology* 73 (3): 910–26.
https://doi.org/10.2307/1940168.

Gómez, Yolanda M., Diego I. Gallardo, Carolina Marchant, Luis Sánchez, and Marcelo
Bourguignon. 2023. 'An In-Depth Review of the Weibull Model with a Focus on Various
Parameterizations'. *Mathematics* 12 (1): 56. https://doi.org/10.3390/math12010056.

Gotthard, Karl. 2008. 'Adaptive Growth Decisions in Butterflies'. *BioScience* 58 (3):
222–30. https://doi.org/10.1641/B580308.

Gribble, Kristin E., George Jarvis, Martha Bock, and David B. Mark Welch. 2014.
'Maternal Caloric Restriction Partially Rescues the Deleterious Effects of Advanced
Maternal Age on Offspring'. *Aging Cell* 13 (4): 623–30.
https://doi.org/10.1111/acel.12217.

Hadfield, Jarrod D. 2008. 'Estimating Evolutionary Parameters When Viability
Selection Is Operating'. *Proceedings of the Royal Society B: Biological Sciences* 275
(1635): 723–34. https://doi.org/10.1098/rspb.2007.1013.

Hall, Margaret E., Lubna Nasir, Francis Daunt, et al. 2004. 'Telomere Loss in Relation
to Age and Early Environment in Long-Lived Birds'. *Proceedings of the Royal Society
of London. Series B: Biological Sciences* 271 (1548): 1571–76.
https://doi.org/10.1098/rspb.2004.2768.

Hamilton, W. D. 1966. 'The Moulding of Senescence by Natural Selection'. *Journal of
Theoretical Biology* 12 (1): 12–45. https://doi.org/10.1016/0022-5193(66)90184-6.

Hercus, Miriam J., and Ary A. Hoffmann. 2000. 'Maternal and Grandmaternal Age
Influence Offspring Fitness in Drosophila.' *Proceedings of The Royal Society B:
Biological Sciences* 267 (1457): 2105–10. https://doi.org/10.1098/rspb.2000.1256.

Hernández, Christina M., Silke F. Van Daalen, Hal Caswell, Michael G. Neubert, and
Kristin E. Gribble. 2020. 'A Demographic and Evolutionary Analysis of Maternal Effect
Senescence'. *Proceedings of the National Academy of Sciences* 117 (28): 16431–37.
https://doi.org/10.1073/pnas.1919988117.

Hessen, Dag O., and Nina Alstad Rukke. 2000. 'The Costs of Moulting in *Daphnia;
Mineral Regulation of Carbon Budgets'. *Freshwater Biology* 45 (2): 169–78.
https://doi.org/10.1046/j.1365-2427.2000.00670.x.

Hibshman, Jonathan D., Anthony Hung, and L. Ryan Baugh. 2016. 'Maternal Diet and
Insulin-Like Signaling Control Intergenerational Plasticity of Progeny Size and
Starvation Resistance'. *PLOS Genetics* 12 (10): e1006396.
https://doi.org/10.1371/journal.pgen.1006396.

Honěk, Alois, and Alois Honek. 1993. 'Intraspecific Variation in Body Size and
Fecundity in Insects: A General Relationship'. *Oikos* 66 (3): 483-92.
https://doi.org/10.2307/3544943.

Houben, Joyce M. J., Harald J. J. Moonen, Frederik J. Van Schooten, and Geja J.
Hageman. 2008. 'Telomere Length Assessment: Biomarker of Chronic Oxidative
Stress?' *Free Radical Biology and Medicine* 44 (3): 235–46.
https://doi.org/10.1016/j.freeradbiomed.2007.10.001.

Ivimey-Cook, Edward, and Jacob Moorad. 2018. 'Disentangling Pre- and Postnatal
Maternal Age Effects on Offspring Performance in an Insect with Elaborate Maternal
Care'. *The American Naturalist* 192 (5): 564–76. https://doi.org/10.1086/699654.

Ivimey-Cook, Edward, and Jacob Moorad. 2020. 'The Diversity of Maternal-Age Effects
upon Pre-Adult Survival across Animal Species'. *Proceedings of the Royal Society B:*
Biological Sciences 287 (1932): 20200972. https://doi.org/10.1098/rspb.2020.0972.

Ivimey-Cook, Edward R., Sarema Shorr, and Jacob A. Moorad. 2023. 'The Distribution
of the Lansing Effect across Animal Species'. *Evolution* 77 (2): 608–15.
https://doi.org/10.1093/evolut/qpac045.

Janssens, Lizanne, and Robby Stoks. 2020. 'Oxidative Stress Mediates Rapid
Compensatory Growth and Its Costs'. *Functional Ecology* 34 (10): 2087–97.
https://doi.org/10.1111/1365-2435.13616.

Jehan, Charly, Camille Sabarly, Thierry Rigaud, and Yannick Moret. 2021. 'Late-life
Reproduction in an Insect: Terminal Investment, Reproductive Restraint or
Senescence'. *Journal of Animal Ecology* 90 (1): 282–97.
https://doi.org/10.1111/1365-2656.13367.

Karabatsos, George. 2006. 'Bayesian Nonparametric Model Selection and Model
Testing'. *Journal of Mathematical Psychology* 50 (2): 123–48.
https://doi.org/10.1016/j.jmp.2005.07.003.

Keil, Gerald, Elizabeth Cummings, and João Pedro De Magalhães. 2015. 'Being Cool:
How Body Temperature Influences Ageing and Longevity'. *Biogerontology* 16 (4): 383–
97. https://doi.org/10.1007/s10522-015-9571-2.

Kern, Stefanie, Martin Ackermann, Martin Ackermann, Stephen C. Stearns, and
Tadeusz J. Kawecki. 2001. 'Decline in Offspring Viability as a Manifestation of Aging in
Drosophila melanogaster'. *Evolution* 55 (9): 1822–31. https://doi.org/10.1111/j.0014-
3820.2001.tb00831.x.

King, Charles E. 1983. 'A Re-Examination of the Lansing Effect'. In *Biology of Rotifers*,
edited by Birger Pejler, R. Starkweather, and Th. Nogrady. Springer Netherlands.
https://doi.org/10.1007/978-94-009-7287-2_15.

Kirkpatrick, Mark, and Russell Lande. 1989. 'THE EVOLUTION OF MATERNAL
CHARACTERS'. *Evolution* 43 (3): 485–503. https://doi.org/10.1111/j.1558-
5646.1989.tb04247.xsing

Kirkwood, T. B. L., and Robin Holliday. 1979. 'The Evolution of Ageing and Longevity'.
Proceedings of the Royal Society of London. Series B. Biological Sciences 205 (1161):
531–46. https://doi.org/10.1098/rspb.1979.0083.

Kong, Augustine, Michael L. Frigge, Gisli Masson, et al. 2012. 'Rate of de Novo
Mutations and the Importance of Father's Age to Disease Risk'. *Nature* 488 (7412):
471–75. https://doi.org/10.1038/nature11396.

Kroeger, Svenja B., Daniel T. Blumstein, Kenneth B. Armitage, Jane M. Reid, and
Julien G. A. Martin. 2020. 'Older Mothers Produce More Successful Daughters'.

Proceedings of the National Academy of Sciences 117 (9): 4809–14.
https://doi.org/10.1073/pnas.1908551117.

Kruschke, John K. 2018. 'Rejecting or Accepting Parameter Values in Bayesian Estimation'. *Advances in Methods and Practices in Psychological Science* 1 (2): 270–80. https://doi.org/10.1177/2515245918771304.

Kullback, Solomon and Richard A. Leibler. 1951. 'On information and sufficiency'. *The annals of mathematical statistics*, 22(1): 79-86. https://www.jstor.org/stable/2236703

Lansing, A. I. 1947. 'A Transmissible, Cumulative, and Reversible Factor in Aging.' *The Journals of Gerontology* 2 (3): 228–39. https://doi.org/10.1093/geronj/2.3.228.

Larsdotter Mellström, Helena, Magne Friberg, Anna-Karin Borg-Karlson, Rushana Murtazina, Mikael Palm, and Christer Wiklund. 2010. 'Seasonal Polyphenism in Life History Traits: Time Costs of Direct Development in a Butterfly'. *Behavioral Ecology and Sociobiology* 64 (9): 1377–83. https://doi.org/10.1007/s00265-010-0952-x.

Lemaître, Jean-François, and Jean-Michel Gaillard. 2017. 'Reproductive Senescence: New Perspectives in the Wild'. *Biological Reviews* 92 (4): 2182–99. https://doi.org/10.1111/brv.12328.

Lemoine, Nathan P. 2019. 'Moving beyond Noninformative Priors: Why and How to Choose Weakly Informative Priors in Bayesian Analyses'. *Oikos* 128 (7): 912–28. https://doi.org/10.1111/oik.05985.

Lemoine, Nathan P., Ava Hoffman, Andrew J. Felton, Lauren Baur, Francis Chaves, Jesse Gray, Giang Yu, Melinda D. Smith. 2016. 'Underappreciated Problems of Low Replication in Ecological Field Studies'. *Ecology* 97 (10): 2554–61. https://doi.org/10.1002/ecy.1506.

Lenth, Russell. 2024. emmeans: Estimated Marginal Means, aka Least-Squares Means. *R package version 1.10.1*. https://CRAN.R-project.org/package=emmeans.

Lind, Martin I., Elena C. Berg, Ghazal Alavioon, and Alexei A. Maklakov. 2015. 'Evolution of Differential Maternal Age Effects on Male and Female Offspring Development and Longevity'. *Functional Ecology* 29 (1): 104–10. https://doi.org/10.1111/1365-2435.12308.

Lippens, Cédric, Bruno Faivre, Clothilde Lechenault, and Gabriele Sorci. 2017. 'Aging parasites produce offspring with poor fitness prospects'. *Biology Letters* 13 (2): 20160888. https://doi.org/10.1098/rsbl.2016.0888.

Lorenz, Matthias W. 2007. 'Oogenesis-Flight Syndrome in Crickets: Age-Dependent Egg Production, Flight Performance, and Biochemical Composition of the Flight Muscles in

Adult Female *Gryllus bimaculatus*'. *Journal of Insect Physiology* 53 (8): 819–32. https://doi.org/10.1016/j.jinsphys.2007.03.011.

Ma, Chun-Sen, Gang Ma, and Sylvain Pincebourde. 2021. 'Survive a Warming Climate: Insect Responses to Extreme High Temperatures'. *Annual Review of Entomology* 66 (1): 163–84. https://doi.org/10.1146/annurev-ento-041520-074454.

Magara, Henlay J. O., Chrysantus M. Tanga, Brian L. Fisher, Abdelmutalab G.A. Azrag, Saliou Niassy, James P. Egonyu, Sylvain Hugel et al. 2024. 'Impact of Temperature on the Bionomics and Geographical Range Margins of the Two-Spotted Field Cricket *Gryllus bimaculatus* in the World: Implications for Its Mass Farming'. *PLOS ONE* 19 (4): e0300438. https://doi.org/10.1371/journal.pone.0300438.

Mair, William, Patrick Goymer, Scott D. Pletcher, and Linda Partridge. 2003. 'Demography of Dietary Restriction and Death in *Drosophila*'. *Science* 301 (5640): 1731–33. https://doi.org/10.1126/science.1086016.

Makeham, William Matthew. 1867. 'On the Law of Mortality'. *Journal of the Institute of Actuaries (1866)* 13 (6): 325–58. https://doi.org/10.1017/S2046166600003238.

Makowski, Dominique, Mattan Ben-Shachar, and Daniel Lüdecke. 2019. 'bayestestR: Describing Effects and Their Uncertainty, Existence and Significance within the Bayesian Framework'. *Journal of Open Source Software* 4 (40): 1541. https://doi.org/10.21105/joss.01541.

Makowski, Dominique, Mattan S. Ben-Shachar, S. H. Annabel Chen, and Daniel Lüdecke. 2019. 'Indices of Effect Existence and Significance in the Bayesian Framework'. *Frontiers in Psychology* 10 (December): 2767. https://doi.org/10.3389/fpsyg.2019.02767.

Marasco, Valeria, Winnie Boner, Kate Griffiths, Shirley Raveh, and Pat Monaghan. 2025. 'Hidden Causes of Variation in Offspring Reproductive Value: Negative Effects of Maternal Breeding Age on Offspring Telomere Length Persist Undiminished Across Multiple Generations'. *Ecology Letters* 28 (1): e70043. https://doi.org/10.1111/ele.70043.

Marshall, Dustin J., Selina S. Heppell, Stephan B. Munch, and Robert R. Warner. 'The relationship between maternal phenotype and offspring quality: do older mothers really produce the best offspring?'. *Ecology* 91 (10): 2862-2873. https://doi.org/10.1890/09-0156.1.

Martínez-Viejo, David, Rolando Rodríguez-Muñoz, and Alfredo F. Ojanguren. 2024. 'Altitudinal Variation in Reproductive Investment among *Gryllus campestris* Populations'. *PLOS ONE* 19 (10): e0312048. https://doi.org/10.1371/journal.pone.0312048.

Masoro, Edward J. 2005. 'Overview of Caloric Restriction and Ageing'. *Mechanisms of*
Ageing and Development 126 (9): 913–22.
https://doi.org/10.1016/j.mad.2005.03.012.

Masoro, Edward J. 2006. 'Dietary Restriction-Induced Life Extension: A Broadly Based
Biological Phenomenon'. *Biogerontology* 7 (3): 153–55.
https://doi.org/10.1007/s10522-006-9015-0.

McCulloch, Robert E. 1989. 'Local Model Influence'. *Journal of the American*
Statistical Association 84 (406): 473–78.
https://doi.org/10.1080/01621459.1989.10478793.

Metcalfe, N. 2003. 'Growth versus Lifespan: Perspectives from Evolutionary Ecology'.
Experimental Gerontology 38 (9): 935–40. https://doi.org/10.1016/S0531-
5565(03)00159-1.

Metcalfe, Neil B., Colin D. Bull, and Marc Mangel. 2002. 'Seasonal Variation in Catch-
up Growth Reveals State-Dependent Somatic Allocations in Salmon'. *Evolutionary*
Ecology Research 4 (6): 871 - 881.

Metcalfe, Neil B., and Pat Monaghan. 2001. 'Compensation for a Bad Start: Grow Now,
Pay Later?' *Trends in Ecology & Evolution* 16 (5): 254–60.
https://doi.org/10.1016/S0169-5347(01)02124-3.

Millar, Russell B. 2009. 'Comparison of Hierarchical Bayesian Models for Overdispersed
Count Data Using DIC and Bayes' Factors'. *Biometrics* 65 (3): 962–69.
https://doi.org/10.1111/j.1541-0420.2008.01162.x.

Mojica, Julius P., and John K. Kelly. 2010. 'Viability Selection Prior to Trait Expression
Is an Essential Component of Natural Selection'. *Proceedings of the Royal Society B:*
Biological Sciences 277 (1696): 2945–50. https://doi.org/10.1098/rspb.2010.0568.

Møller, Anders, and Michael D. Jennions. 2002. 'How Much Variance Can Be Explained
by Ecologists and Evolutionary Biologists?' *Oecologia* 132 (4): 492–500.
https://doi.org/10.1007/s00442-002-0952-2.

Monaghan, P., A. Charmantier, D. H. Nussey, and R. E. Ricklefs. 2008. 'The
Evolutionary Ecology of Senescence'. *Functional Ecology* 22 (3): 371–78.
https://doi.org/10.1111/j.1365-2435.2008.01418.x.

Monaghan, Pat. 2014. 'Organismal Stress, Telomeres and Life Histories'. *Journal of*
Experimental Biology 217 (1): 57–66. https://doi.org/10.1242/jeb.090043.

Monaghan, Pat, and Mark F. Haussmann. 2006. 'Do Telomere Dynamics Link Lifestyle
and Lifespan?' *Trends in Ecology & Evolution* 21 (1): 47–53.
https://doi.org/10.1016/j.tree.2005.11.007.

Monaghan, Pat, Alexei A. Maklakov, and Neil B. Metcalfe. 2020. 'Intergenerational Transfer of Ageing: Parental Age and Offspring Lifespan'. *Trends in Ecology & Evolution* 35 (10): 927–37. https://doi.org/10.1016/j.tree.2020.07.005.

Monaghan, Pat, and Neil B. Metcalfe. 2019. 'The Deteriorating Soma and the Indispensable Germline: Gamete Senescence and Offspring Fitness'. *Proceedings of the Royal Society B: Biological Sciences* 286 (1917): 20192187. https://doi.org/10.1098/rspb.2019.2187.

Moorad, Jacob A., and Daniel H. Nussey. 2016. 'Evolution of Maternal Effect Senescence'. *Proceedings of the National Academy of Sciences* 113 (2): 362–67. https://doi.org/10.1073/pnas.1520494113.

Morimoto, Juliano. 2022. 'Parental Ecological History Can Differentially Modulate Parental Age Effects on Offspring Physiological Traits in *Drosophila*'. *Current Zoology* 68 (4): 391–99. https://doi.org/10.1093/cz/zoab081.

Mousseau, T A, and H. Dingle. 1991. 'Maternal Effects in Insect Life Histories'. *Annual Review of Entomology* 36 (1): 511–34. https://doi.org/10.1146/annurev.en.36.010191.002455.

Mousseau, T., and Charles W. Fox. 1998. 'The Adaptive Significance of Maternal Effects'. *Trends in Ecology & Evolution* 13 (10): 403–7. https://doi.org/10.1016/S0169-5347(98)01472-4.

Mousseau, Timothy A., and Derek A. Roff. 1989. 'ADAPTATION TO SEASONALITY IN A CRICKET: PATTERNS OF PHENOTYPIC AND GENOTYPIC VARIATION IN BODY SIZE AND DIAPAUSE EXPRESSION ALONG A CLINE IN SEASON LENGTH'. *Evolution* 43 (7): 1483–96. https://doi.org/10.1111/j.1558-5646.1989.tb02598.x.

Muller, Doriane, David Giron, Emmanuel Desouhant, Benjamin Rey, Jérôme Casas, Nicolas Lefrique, Bertanne Visser. 2017. 'Maternal Age Affects Offspring Nutrient Dynamics'. *Journal of Insect Physiology* 101 (August): 123–31. https://doi.org/10.1016/j.jinsphys.2017.07.011.

Munch, Stephan B., and Santiago Salinas. 2009. 'Latitudinal Variation in Lifespan within Species Is Explained by the Metabolic Theory of Ecology'. *Proceedings of the National Academy of Sciences* 106 (33): 13860–64. https://doi.org/10.1073/pnas.0900300106.

Nakagawa, Shinichi, Santiago Ortega, Elena Gazzea, Malgorzata Lagisz, Anna Lenz, Erick Lundgren, Ayumi Mizuno. 2025. 'Location–Scale Models in Ecology and Evolution: Heteroscedasticity in Continuous, Count and Proportion Data'. *Methods in Ecology and Evolution* 17 (2): 554-566. https://doi.org/10.1111/2041-210x.70203.

Nguyen, Alan B. H., Marco Bonici, Glen McGee, and Will J. Percival. 2025. 'LOO-PIT: A
Sensitive Posterior Test'. *Journal of Cosmology and Astroparticle Physics* 2025 (01):
008. https://doi.org/10.1088/1475-7516/2025/01/008.

Noguera, José Carlos. 2021. 'Heterogenous Effects of Father and Mother Age on
Offspring Development'. *Behavioral Ecology* 32 (2): 349-58.
https://doi.org/10.1093/beheco/araa153.

Noguera, José C., Neil B. Metcalfe, and Pat Monaghan. 2018. 'Experimental
demonstration that offspring fathered by old males have shorter telomeres and
reduced lifespans'. *Proceedings of the Royal Society B: Biological Sciences* 285 (1874):
20180268. https://doi.org/10.1098/rspb.2018.0268.

Nussey, D. H., T. Coulson, M. Festa-Bianchet, and J. -M. Gaillard. 2008. 'Measuring
Senescence in Wild Animal Populations: Towards a Longitudinal Approach'. *Functional*
Ecology 22 (3): 393–406. https://doi.org/10.1111/j.1365-2435.2008.01408.x.

Nussey, Daniel H., Hannah Froy, Jean-François Lemaitre, Jean-Michel Gaillard, and
Steve N. Austad. 2013. 'Senescence in Natural Populations of Animals: Widespread
Evidence and Its Implications for Bio-Gerontology'. *Ageing Research Reviews* 12 (1):
214–25. https://doi.org/10.1016/j.arr.2012.07.004.

Nylin, Sören, Georg H. Nygren, Lina Söderlind, and Constantí Stefanescu. 2009.
'Geographical Variation in Host Plant Utilization in the Comma Butterfly: The Roles of
Time Constraints and Plant Phenology'. *Evolutionary Ecology* 23 (5): 807–25.
https://doi.org/10.1007/s10682-008-9274-0.

Nylin, Sören, Per-Olof Wickman, and Christer Wiklund. 1989. 'Seasonal Plasticity in
Growth and Development of the Speckled Wood Butterfly, *Pararge aegeria*
(Satyrinae)'. *Biological Journal of the Linnean Society* 38 (2): 155–71.
https://doi.org/10.1111/j.1095-8312.1989.tb01571.x.

O'Dea, Rose E., Daniel W. A. Noble, Sheri L. Johnson, Daniel Hesselson, and Shinichi
Nakagawa. 2016. 'The Role of Non-Genetic Inheritance in Evolutionary Rescue:
Epigenetic Buffering, Heritable Bet Hedging and Epigenetic Traps'. *Environmental*
Epigenetics 2 (1): dvv014. https://doi.org/10.1093/eep/dvv014.

O'Dea, Rose E., Daniel W. A. Noble, and Shinichi Nakagawa. 2022. 'Unifying Individual
Differences in Personality, Predictability and Plasticity: A Practical Guide'. *Methods*
in Ecology and Evolution 13 (2): 278-93. https://doi.org/10.1111/2041-210X.13755.

Olsson, M., and T. Uller. 2002. 'Developmental Stability and Genetic Architecture: A
Comparison within and across Thermal Regimes in Tadpoles'. *Journal of Evolutionary*
Biology 15 (4): 625-33. https://doi.org/10.1046/j.1420-9101.2002.00417.x.

Opit, G. P., and J. E. Throne. 2007. 'Influence of Maternal Age on the Fitness of
Progeny in the Rice Weevil, *Sitophilus oryzae* (Coleoptera: Curculionidae)'.

Environmental Entomology 36 (1): 83–89. https://doi.org/10.1603/0046-225X(2007)36%255B83:IOMAOT%255D2.0.CO;2.

Pásztor, Kata, Ádám Kőrösi, Ádám Gór, Viktor Szigeti, Flóra Vajna, and János Kis. 2022. 'Phenotypic Senescence in a Natural Insect Population'. *Ecology and Evolution* 12 (12): e9668. https://doi.org/10.1002/ece3.9668.

Pfenning, B., H. J. Poethke, and T. Hovestadt. 2007. 'Dealing with Time Constraints on Development: The Effect of Food Availability'. *Ecological Entomology* 32 (3): 273–78. https://doi.org/10.1111/j.1365-2311.2007.00867.x.

Phelan, John P., and Peter C. Frumhoff. 1991. 'Differences in the Effects of Parental Age on Offspring Life History between Tropical and Temperate Populations of Milkweed Bugs (*Oncopeltus Spp.*)'. *Evolutionary Ecology* 5 (2): 160–72. https://doi.org/10.1007/BF02270832.

Philippi, Tom, and Jon Seger. 1989. 'Hedging One's Evolutionary Bets, Revisited'. *Trends in Ecology & Evolution* 4 (2): 41–44. https://doi.org/10.1016/0169-5347(89)90138-9.

Pinder, John E., James G. Wiener, and Michael H. Smith. 1978. 'The Weibull Distribution: A New Method of Summarizing Survivorship Data'. *Ecology* 59 (1): 175–79. https://doi.org/10.2307/1936645.

Plaistow, S.J, and T. G. Benton. 2009. 'The Influence of Context-Dependent Maternal Effects on Population Dynamics: An Experimental Test'. *Philosophical Transactions of the Royal Society B: Biological Sciences* 364 (1520): 1049–58. https://doi.org/10.1098/rstb.2008.0251.

Plaistow, Stewart J., Christopher Shirley, Helene Collin, Stephen J. Cornell, and Ewan D. Harney. 2015. 'Offspring Provisioning Explains Clone-Specific Maternal Age Effects on Life History and Life Span in the Water Flea, *Daphnia pulex*'. *The American Naturalist* 186 (3): 376–89. https://doi.org/10.1086/682277.

Plaistow, Stewart J., James J. H. St Clair, Jane Grant, Jane Grant, and Tim G. Benton. 2007. 'How to Put All Your Eggs in One Basket: Empirical Patterns of Offspring Provisioning throughout a Mother's Lifetime'. *The American Naturalist* 170 (4): 520–29. https://doi.org/10.1086/521238.

Priest, Nicholas K., Benjamin Mackowiak, and Daniel E. L. Promislow. 2002. 'The Role of Parental Age Effects on the Evolution of Ageing'. *Evolution* 56 (5): 927–35. https://doi.org/10.1111/j.0014-3820.2002.tb01405.x.

Promislow, Daniel E. L., Thomas Flatt, and Russell Bonduriansky. 2022. 'The Biology of Aging in Insects: From *Drosophila* to Other Insects and Back'. *Annual Review of Entomology* 67 (1): 83–103. https://doi.org/10.1146/annurev-ento-061621-064341.

R Core Team. 2024. R: A Language and Environment for Statistical Computing. *R*
Foundation for Statistical Computing, Vienna, Austria. *R version 4.3.3.*
https://www.R-project.org/.

Reichert, Sophie, Vérane Berger, John Jackson, Simon N. Chapman, Win Htut, Khyne
U. Mar, and Virpi Lummaa. 'Maternal age at birth shapes offspring life-history
trajectory across generations in long-lived Asian elephants'. *Journal of animal*
ecology 89 (4): 996-1007. https://doi.org/10.1111/1365-2656.13049.

Reid, Jane M., Eric M. Bignal, Sue Bignal, Davy I. McCracken, Maria I. Bogdanova, and
Pat Monaghan. 2010. 'Parent Age, Lifespan and Offspring Survival: Structured
Variation in Life History in a Wild Population'. *Journal of Animal Ecology* 79 (4): 851–
62. https://doi.org/10.1111/j.1365-2656.2010.01669.x.

Ribou, Anne-Cécile, and Klaus Reinhardt. 2012. 'Reduced Metabolic Rate and Oxygen
Radicals Production in Stored Insect Sperm'. *Proceedings of the Royal Society B:*
Biological Sciences 279 (1736): 2196–203. https://doi.org/10.1098/rspb.2011.2422.

Rivero, A., D. Giron, and J. Casas. 2001. 'Lifetime Allocation of Juvenile and Adult
Nutritional Resources to Egg Production in a Holometabolous Insect'. *Proceedings of*
the Royal Society of London. Series B: Biological Sciences 268 (1473): 1231–37.
https://doi.org/10.1098/rspb.2001.1645.

Rodríguez-Muñoz, Rolando, Jelle J. Boonekamp, David Fisher, Paul Hopwood, and Tom
Tregenza. 2019. 'Slower Senescence in a Wild Insect Population in Years with a More
Female-Biased Sex Ratio'. *Proceedings of the Royal Society B: Biological Sciences* 286
(1900): 20190286. https://doi.org/10.1098/rspb.2019.0286.

Rodríguez-Muñoz, Rolando, Jelle J. Boonekamp, Xing P. Liu, Ian Skicko, Sophie
Haugland Pedersen, David N. Fisher, Paul Hopwood, Tom Tregenza. 2019. 'Comparing
Individual and Population Measures of Senescence across 10 Years in a Wild Insect
Population: INDIVIDUAL AND POPULATION SENESCENCE'. *Evolution* 73 (2): 293–302.
https://doi.org/10.1111/evo.13674.

Roff. 2000. 'Trade-Offs between Growth and Reproduction: An Analysis of the
Quantitative Genetic Evidence'. *Journal of Evolutionary Biology* 13 (3): 434-45.
https://doi.org/10.1046/j.1420-9101.2000.00186.x.

Roff, Derek. 1980. 'Optimizing Development Time in a Seasonal Environment:
The ?Ups and Downs? Of Clinal Variation'. *Oecologia* 45 (2): 202–8.
https://doi.org/10.1007/BF00346461.

Rose, Michael R. 1984. 'Laboratory Evolution of Postponed Senescence in *Drosophila*
melanogaster'. *Evolution* 38 (5): 1004. https://doi.org/10.2307/2408434.

Säilynoja, Teemu, Paul-Christian Bürkner, and Aki Vehtari. 2022. 'Graphical Test for
Discrete Uniformity and Its Applications in Goodness-of-Fit Evaluation and Multiple

Sample Comparison'. *Statistics and Computing* 32 (2): 32.
https://doi.org/10.1007/s11222-022-10090-6.

Schneider, Caroline A., Wayne S. Rasband, and Kevin W. Eliceiri. 2012. 'NIH Image to
ImageJ: 25 Years of Image Analysis'. *Nature Methods* 9 (7): 671–75.
https://doi.org/10.1038/nmeth.2089.

Schroeder, Julia, Shinichi Nakagawa, Mark Rees, Maria-Elena Mannarelli, and Terry
Burke. 2015. 'Reduced Fitness in Progeny from Old Parents in a Natural Population'.
Proceedings of the National Academy of Sciences 112 (13): 4021–25.
https://doi.org/10.1073/pnas.1422715112.

Schwaferts, Patrick, and Thomas Augustin. 2020. *Bayesian Decisions Using Regions of*
Practical Equivalence (ROPE): Foundations. Department of Statistics: Technical
Reports. 325. https://doi.org/10.5282/UBM/EPUB.74222.

Siler, William. 1979. 'A Competing-Risk Model for Animal Mortality'. *Ecology* 60 (4):
750–57. https://doi.org/10.2307/1936612.

Simmons, L. W. 1987. 'Competition Between Larvae of the Field Cricket, *Gryllus*
bimaculatus (Orthoptera: Gryllidae) and Its Effects on Some Life-History Components
of Fitness'. *The Journal of Animal Ecology* 56 (3): 1015-1027.
https://doi.org/10.2307/4963.

Simons, Andrew M., and Mark O. Johnston. 2006. 'ENVIRONMENTAL AND GENETIC
SOURCES OF DIVERSIFICATION IN THE TIMING OF SEED GERMINATION: IMPLICATIONS
FOR THE EVOLUTION OF BET HEDGING'. *Evolution* 60 (11): 2280-92.

Singh, K. and Omkar. 2009. 'Effect of Parental Ageing on Offspring Developmental and
Survival Attributes in an Aphidophagous Ladybird, *Cheilomenes sexmaculata*'. *Journal*
of Applied Entomology 133 (7): 500–504. https://doi.org/10.1111/j.1439-
0418.2008.01356.x.

Smith, Shona M., Ruedi G. Nager, and David Costantini. 2016. 'Meta-analysis Indicates
That Oxidative Stress Is Both a Constraint on and a Cost of Growth'. *Ecology and*
Evolution 6 (9): 2833–42. https://doi.org/10.1002/ece3.2080.

Spiegelhalter, David J., Nicola G. Best, Bradley P. Carlin, and Angelika Van Der Linde.
2002. 'Bayesian Measures of Model Complexity and Fit'. *Journal of the Royal*
Statistical Society Series B: Statistical Methodology 64 (4): 583–639.
https://doi.org/10.1111/1467-9868.00353.

Spindler, Stephen R. 2010. 'Caloric Restriction: From Soup to Nuts'. *Ageing Research*
Reviews 9 (3): 324–53. https://doi.org/10.1016/j.arr.2009.10.003.

Stahlschmidt, Zachary R., Njal Rollinson, Madison Acker, and Shelley A. Adamo. 2013.
'Are All Eggs Created Equal? Food Availability and the Fitness Trade-off between

Reproduction and Immunity'. *Functional Ecology* 27 (3): 800–806.
https://doi.org/10.1111/1365-2435.12071.

Stan Developpement Team. 2024. *RStan: the R interface to Stan. R package version 2.32.6.* https://mc-stan.org/.

Stearns, Stephen C. 2000. 'Life History Evolution: Successes, Limitations, and Prospects'. *Naturwissenschaften* 87 (11): 476–86.
https://doi.org/10.1007/s001140050763.

Stiver, Kelly A., and Suzanne H. Alonzo. 2009. 'Parental and Mating Effort: Is There Necessarily a Trade-Off?: (Invited Review)'. *Ethology* 115 (12): 1101–26.
https://doi.org/10.1111/j.1439-0310.2009.01707.x.

Sturm, Robert. 2016. 'Relationship between Body Size and Reproductive Capacity in Females of the Black Field Cricket (Orthoptera, Gryllidae)'. Austrain Journal of Forest Science. 48.

Torres, Roxana, Hugh Drummond, and Alberto Velando. 2011. 'Parental Age and Lifespan Influence Offspring Recruitment: A Long-Term Study in a Seabird'. *PLoS ONE* 6 (11): e27245. https://doi.org/10.1371/journal.pone.0027245.

Travers, Laura M., Hanne Carlsson, Martin I. Lind, and Alexei A. Maklakov. 2021. 'Beneficial Cumulative Effects of Old Parental Age on Offspring Fitness'. *Proceedings of the Royal Society B: Biological Sciences* 288 (1960): 20211843.
https://doi.org/10.1098/rspb.2021.1843.

Tregenza, Tom, Rolando Rodríguez-Muñoz, Alfredo F. Ojanguren, Paul Hopwood, and Jelle J. Boonekamp. 2026. 'Impacts of Parental Age and Inbreeding on Fitness in a Wild Insect'. *PLOS Biology* 24 (3): e3003713.
https://doi.org/10.1371/journal.pbio.3003713.

Tyler, Frances, Xavier A. Harrison, Amanda Bretman, Thor Veen, Rolando Rodríguez-Muñoz, and Tom Tregenza. 2013. 'Multiple Post-mating Barriers to Hybridization in Field Crickets'. *Molecular Ecology* 22 (6): 1640–49.
https://doi.org/10.1111/mec.12187.

Van Daalen, Silke F., Christina M. Hernández, Hal Caswell, Michael G. Neubert, and Kristin E. Gribble. 2022. 'The Contributions of Maternal Age Heterogeneity to Variance in Lifetime Reproductive Output'. *The American Naturalist* 199 (5): 603–16.
https://doi.org/10.1086/718716.

Van De Pol, Martijn, and Jonathan Wright. 2009. 'A Simple Method for Distinguishing Within- versus between-Subject Effects Using Mixed Models'. *Animal Behaviour* 77 (3): 753–58. https://doi.org/10.1016/j.anbehav.2008.11.006.

Van Den Heuvel, Joost, Sinead English, and Tobias Uller. 2016. 'Disposable Soma Theory and the Evolution of Maternal Effects on Ageing'. *PLOS ONE* 11 (1): e0145544. https://doi.org/10.1371/journal.pone.0145544.

Van Noordwijk, A. J., and G. De Jong. 1986. 'Acquisition and Allocation of Resources: Their Influence on Variation in Life History Tactics'. *The American Naturalist* 128 (1): 137–42. https://doi.org/10.1086/284547.

Vats, Dootika, and Christina Knudson. 2021. 'Revisiting the Gelman–Rubin Diagnostic'. *Statistical Science* 36 (4): 518-29. https://doi.org/10.1214/20-STS812.

Vega-Trejo, Regina, Loeske E. B. Kruuk, Michael D. Jennions, and Megan L. Head. 2018. 'What Happens to Offspring When Parents Are Inbred, Old or Had a Poor Start in Life? Evidence for Sex-specific Parental Effects'. *Journal of Evolutionary Biology* 31 (8): 1138–51. https://doi.org/10.1111/jeb.13292.

Vehtari, Aki, Andrew Gelman, and Jonah Gabry. 2017. 'Practical Bayesian Model Evaluation Using Leave-One-out Cross-Validation and WAIC'. *Statistics and Computing* 27 (5): 1413–32. https://doi.org/10.1007/s11222-016-9696-4.

Vehtari, Aki, Andrew Gelman, Daniel Simpson, Bob Carpenter, and Paul-Christian Bürkner. 2021. 'Rank-Normalization, Folding, and Localization: An Improved R̂ for Assessing Convergence of MCMC (with Discussion)'. *Bayesian Analysis* 16 (2): 667-718. https://doi.org/10.1214/20-BA1221.

Vehtari, Aki, Tommi Mononen, Ville Tolvanen, Tuomas Sivula, and Ole Winther. 2016. 'Bayesian Leave-One-Out Cross-Validation Approximations for Gaussian Latent Variable Models'. *Journal of Machine Learning Research* 17 (103): 1–38.

Vehtari, Aki, Daniel Simpson, Andrew Gelman, Yuling Yao, and Jonah Gabry. 2015. *Pareto Smoothed Importance Sampling*. Version 9. https://doi.org/10.48550/ARXIV.1507.02646.

Velando, Alberto, Hugh Drummond, and Roxana Torres. 2006. 'Senescent Birds Redouble Reproductive Effort When Ill: Confirmation of the Terminal Investment Hypothesis'. *Proceedings of the Royal Society B: Biological Sciences* 273 (1593): 1443–48. https://doi.org/10.1098/rspb.2006.3480.

Wickham, Hadley H. 2016. ggplot2: Elegant Graphics for Data Analysis. Springer-Verlag New York.

Winter, Bodo, and Paul-Christian Bürkner. 2021. 'Poisson Regression for Linguists: A Tutorial Introduction to Modelling Count Data with Brms'. *Language and Linguistics Compass* 15 (11). e12439. https://doi.org/10.1111/lnc3.12439.

Wylde, Zachariah, Foteini Spagopoulou, Amy K. Hooper, Alexei A. Maklakov, and Russell Bonduriansky. 2019. 'Parental Breeding Age Effects on Descendants' Longevity

Interact over 2 Generations in Matrilines and Patrilines'. *PLOS Biology* 17 (11):
e3000556. https://doi.org/10.1371/journal.pbio.3000556.

Zajitschek, Felix, Susanne Zajitschek, and Russell Bonduriansky. 2020. 'Senescence in
Wild Insects: Key Questions and Challenges'. *Functional Ecology* 34 (1): 26–37.
https://doi.org/10.1111/1365-2435.13399.

Zehnder, Caralyn B., Melissa A. Parris, and Mark D. Hunter. 2007. 'Effects of Maternal
Age and Environment on Offspring Vital Rates in the Oleander Aphid (Hemiptera:
Aphididae)'. *Environmental Entomology* 36 (4): 910–17.
https://doi.org/10.1093/ee/36.4.910.

Zhang, Dao-Wei, Zhong-Jiu Xiao, Bo-Ping Zeng, Kun Li, and Yan-Long Tang. 2019.
'Insect Behavior and Physiological Adaptation Mechanisms Under Starvation Stress'.
Frontiers in Physiology 10 (March): 163. https://doi.org/10.3389/fphys.2019.00163.