TITLE: Increasing absolute prey community density protects aposematic models and their

imperfect Batesian mimics: Evidence from Neotropical *Adelpha* butterflies

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ABSTRACT: Batesian mimicry is a defensive adaptation where predators learn to avoid

aposematic prey and generalize their warning signals to phenotypically similar mimics. The

phenotypic accuracy needed for mimics to benefit from this adaptation depends on the relative

densities of models and mimics and the model's unpalatability. As aposematic models become

more unpalatable or more common relative to their mimics, warning signals become stronger,

allowing even poor mimics to benefit. However, few studies have disentangled the importance of

relative frequencies of models and mimics from *absolute* density of the prey community (both

models and mimics) in driving relaxed selection on imperfect mimics. Here, we test the

hypothesis that increasing model unpalatability and absolute prey community density accelerates

predator avoidance learning and enhances protection for imperfect mimics. Using replicas of the

model *Adelpha iphiclus* (Linnaeus), its imperfect mimic *Adelpha serpa* (Boisduval), and the

palatable control *Junonia evarete* (Cramer), we conducted field experiments that enhanced

model unpalatability and doubled absolute prey density while maintaining a constant ratio of

model, mimic, and control phenotypes. We found that enhanced model unpalatability and

increased absolute density significantly reduced predation on all species, highlighting absolute

community density as an underappreciated mechanism shaping selection on imperfect Batesian

mimics.

KEYWORDS: Aposematism, predator learning, signal detection theory, Müllerian mimicry,

frequency-dependent selection, optimal foraging theory

INTRODUCTION

Predator–prey interactions drive the evolution of diverse and complex anti-predator

defenses and associated predator counterstrategies (Ruxton et al., 2018; Skelhorn et al., 2016;

Speed & Turner, 1999). Within this broad coevolutionary framework, Batesian mimicry

represents a classic predator avoidance strategy in which a palatable organism gains protection

from its predators by resembling an unpalatable, or otherwise defended, model population

(Bates, 1862). Although this classic understanding of Batesian mimicry predicts strong selection

for close model–mimic resemblance, many Batesian mimics do not perfectly resemble their toxic

models, raising important questions about the selective forces and ecological conditions that

allow imperfect mimicry to persist (Penney et al., 2012; Ruxton et al., 2018; Sherratt, 2002).

How perfectly a mimic must resemble its model depends on the foraging conditions of its

predators (Sherratt & Peet-Paré, 2017). When predators have little incentive to discriminate

between models and mimics, selection for precise resemblance is relaxed and predators

generalize phenotypic traits more broadly (Edmunds, 2000; Kikuchi & Pfennig, 2013). Under

this relaxed selection, mimics can deviate from their model's appearance and still gain protection

from predation, resulting in imperfect mimicry—where model–mimic resemblance is incomplete

but functionally sufficient (Sherratt, 2011; Turner et al., 1984).

Theoretical, field, and laboratory studies show that the degree of predator

generalization—and thus the level of phenotypic similarity needed to garner protection from

predators—depends on factors such as the degree of model unpalatability, the relative

abundances of models and mimics, and the availability of alternative prey (Brower et al., 1968;

Kokko et al., 2003; Lindström et al., 2004; Pfennig et al., 2001; Ritland, 1994; Sherratt, 2002).

Other works, both theoretical and empirical, highlight how environmental heterogeneity and

predator and prey community complexity can also relax selection on imperfect mimics (Ihalainen

et al., 2012; Kaczmarek et al., 2020; Kikuchi et al., 2022; Lindström et al., 1997, 2004;

Luedeman et al., 1981; Malcolm, 1990; Sherratt & Peet-Paré, 2017). Despite these advances, key

questions remain about the population- and community-level processes that allow imperfect

mimics to persist in natural environments.

Aposematic prey—species that use bright colors or other conspicuous traits to warn

predators about their toxicity—often vary in the strength of their chemical defense across natural

environments (Brower et al., 1968; Ritland, 1995). This natural variation in chemical defense can

correspond to changes in the dynamics of mimetic interactions (Prudic et al., 2019). Empirical

studies show that mimics survive better when their models are highly unpalatable (Lindström et

al., 1997; Pike & Burman, 2023), and that strongly defended models can accelerate predator

avoidance learning (Skelhorn & Rowe, 2006). Theoretical work further demonstrates that

increasing unpalatability strengthens the model's warning signal and relaxes selection on

mimetic accuracy, allowing imperfect mimics to persist despite copying the model's phenotype

imprecisely (Owen & Owen, 1984; Sherratt, 2002, 2011; Speed & Turner, 1999; Turner et al.,

1984). In other words, when the cost of attacking a toxic model is sufficiently high, predators

generalize warning signals more broadly, enabling imperfect mimics to remain sufficiently

protected even without close resemblance to the model.

Just as greater model unpalatability enhances protection for imperfect mimics, increasing

the abundance of a toxic model *relative* to its mimic also strengthens the model's warning signal,

thereby relaxing selection on high-fidelity mimicry (Edmunds, 2000; Kikuchi & Pfennig, 2013;

Ruxton et al., 2018; Sherratt & Peet-Paré, 2017). When the toxic model is highly abundant

relative to the mimic, predators face an increased risk of encountering it. Therefore, as the toxic

model becomes more abundant relative to the mimic, predators experience stronger selection to

avoid it, which also increases protection for mimics (Getty, 1985; Pfennig et al., 2001; Sherratt,

2011; Turner et al., 1984). Consistent with these theoretical predictions, several empirical studies

show that mimics exhibit higher survival when their models have a higher relative abundance

(Chouteau et al., 2016; Finkbeiner et al., 2018; Iserbyt et al., 2011; Lindström et al., 1997;

Nelson et al., 2011; Pfennig et al., 2001). These effects of relative frequency are also visible in

patterns of geographic variation in mimetic fidelity. For example, phenotypic resemblance

between the eastern coral snake model and its Batesian mimic the scarlet kingsnake is higher

near the model's range edge, where the model is relatively rare, and lower in the center of the

model's range, where the model is relatively common (Harper & Pfennig, 2007). This pattern is

consistent with the idea that selection for precise mimicry intensifies when the model is

relatively rare, while poor mimics can persist when the model is relatively common because the

strength of the model's warning signal is heightened. On the other hand, when a palatable mimic

is relatively more common than its toxic model, this can create a parasitic effect, where the

model's warning signal is so diluted by the high relative frequency of its mimic that survival of

both models and mimics decreases (Aubier et al., 2017; Heerwig et al., 2023; Rowland, Mappes,

et al., 2010; Skelhorn et al., 2011).

It is therefore well established that the relative frequency of toxic models and their

palatable mimics plays a crucial role in determining when mimics are protected from predation.

However, surprisingly little empirical work has disentangled these effects of *relative* frequency

from those driven by *absolute* prey density. The role of absolute density in driving predator

responses is well understood in the context of aposematic prey, where the strength of a warning

signal increases with the rate at which predators encounter that signal, which increases as the

absolute abundance of the aposematic organism increases (Chouteau et al., 2016; Endler &

Mappes, 2004; Ioannou et al., 2008; Lotka & Volterra, 1925). As such, high absolute abundance

reduces predation on defended aposematic prey (Alatalo & Mappes, 1996; Briolat et al., 2019;

Finkbeiner et al., 2012; Kaczmarek et al., 2020; Kikuchi et al., 2021). Previous work has isolated

relative model–mimic frequencies from total prey density, demonstrating that increasing the

relative frequency of an unpalatable model, while holding total prey density constant, can

enhance protection for imperfect Batesian mimics—thereby highlighting relative frequency as an

independent mechanism driving mimic protection (Lindström et al., 1997). Yet it remains

unclear whether changes in the absolute density of a mimetic community (models and mimics),

with model–mimic ratios held constant, also affect the survival of aposematic and mimetic prey.

Moreover, although several studies emphasize the importance of considering absolute density in

experimental tests of mimicry, none have manipulated absolute prey density while maintaining

constant model–mimic ratios (Lindström et al., 2001; Rowland et al., 2007; Rowland,

Hoogesteger, et al., 2010). As a result, it is difficult to determine whether relaxed selection on

imperfect mimics arises from shifts in relative frequency, changes in absolute community

density, or a combination of the two.

Here, we investigate how model unpalatability and absolute prey density shape protection

for imperfect Batesian mimics in Neotropical *Adelpha* butterflies, a phylogenetically diverse

genus characterized by extensive convergent evolution of wing patterns and the formation of

multi-species mimicry rings composed of both palatable and unpalatable taxa (Ebel et al., 2015;

Mullen et al., 2011; Páez V. et al., 2025; Willmott, 2003). In *Adelpha*, unpalatable models

typically specialize on toxic *Rubiaceae* host plants and likely sequester defensive secondary

metabolites from these plants, whereas palatable mimics are generalists that feed on non-

Rubiaceae hosts (Finkbeiner et al., 2017; Kessler & Baldwin, 2002; Lopes et al., 2004; Prudic et

al., 2007; Soto-Sobenis et al., 2001).

A well-studied pair within this system is the toxic *Rubiaceae* specialist *Adelpha iphiclus*

(Linnaeus) and its near-identical Batesian mimic *Adelpha serpa* (Boisduval) (Finkbeiner et al.,

2017, 2018). Despite their striking phenotypic similarity, field experiments in Costa Rica showed

that while predators avoided *A. iphiclus*, *A. serpa* was not protected from predation, indicating

that predators were able to discriminate between the two and selectively forage only on the

edible mimic (Finkbeiner et al., 2017). This predator discrimination, despite near-identical wing

patterns, renders *A. serpa* an imperfect mimic. Subsequent work demonstrated that protection for

this imperfect mimic is governed by negative frequency dependent selection: while *A. serpa* is

not protected in Costa Rica, where the species occurs at a similar frequency to its model, *A.*

serpa is protected in Ecuador, where it is much rarer relative to its model (Finkbeiner et al.,

2018). This work highlights the central role of relative model–mimic frequencies in maintaining

mimicry in *Adelpha* and positions *Adelpha* as a powerful system for testing how absolute prey

density and model toxicity interact to shape selection on mimicry complexes.

Using field predation experiments in Costa Rica, we test how model unpalatability and

absolute prey density impact survival of the chemically defended model *Adelpha iphiclus*, its

palatable, Batesian mimic *Adelpha serpa*, and a palatable control *Junonia evarete* (Cramer).

Experiments were carried out using artificial butterfly facsimiles at the same Costa Rican field

site used by Finkbeiner et al. (2017). To isolate absolute density effects from relative frequency

effects, we doubled the absolute facsimile prey density while maintaining a constant 1:1:1 ratio

of model (*A. iphiclus*), mimic (*A. serpa*), and control (*J. evarete*) facsimiles. We hypothesized

that, at a fixed model–mimic ratio, increasing absolute prey density and model unpalatability

would accelerate predator avoidance learning (Hypothesis 1) and increase protection for the

imperfect mimic (Hypothesis 2). To test this, we designed an experiment with two levels of

absolute prey density (low vs. high) and two levels of model unpalatability (unaltered vs.

experimentally enhanced), enabling us to investigate important questions about how model

unpalatability and absolute prey density interact to relax selection on mimetic communities.

<u>METHODS</u>

Field site

To explore how changes in model unpalatability and absolute prey density impact

protection for mimetic butterflies, we conducted a field predation experiment at the Organization

for Tropical Studies' (OTS) La Selva Biological Reserve (10°25'28"N, -84°0'18"W) in

Sarapiquí, Costa Rica between May and June 2022. La Selva contains 1,600 hectares of old

growth and secondary growth tropical lowland forest bordering the Northern section of Braulio

Carrillo National Park. As such, this reserve offers unparalleled access to a diverse and abundant

community of insectivorous avian predators. Several large-scale field experiments using similar

artificial butterfly replicas have been completed at La Selva, demonstrating that this field site is

suitable for conducting predation experiments of this nature (Finkbeiner et al., 2012, 2017,

2018).

Butterfly facsimile production

Artificial butterfly facsimiles were used to measure predation across experimental

treatments. We used color-accurate facsimiles of *Adelpha iphiclus* (model), *Adelpha serpa*

(mimic), and *Junonia evarete* (control) designed by Finkbeiner et al. (2017). In Finkbeiner et al.

(2017), spectral reflectance measurements were taken and compared against real butterfly wings

for each species, revealing no differences in discriminability between facsimiles and real

butterfly wings. These facsimiles were successfully used to assess the effects of relative

frequency on predator learning within the same *Adelpha* mimicry system in the same field site

(La Selva). Therefore, no changes were made to facsimile design in the present study, and

additional spectral reflectance measurements were not taken.

Facsimiles were constructed following protocols outlined in Robinson et al. (2025).

Facsimile images were double-printed using an Epson Stylus Pro 4900 printer with

UltraChrome® High Dynamic Range ink on Grade 1 Whatman filter paper (Finkbeiner et al.,

2012, 2017; Robinson et al., 2025). Once printed, facsimiles were pasted onto a cardstock

backing (brand: Bazzil®; color: mocha divine) using Krylon® high-strength spray adhesive.

After 48 hours, facsimiles were cut out and sprayed with Krylon® Matte Finish, which provided

moisture resistance and protection from color damage. Black twist ties were attached to each

facsimile, which allowed them to be easily affixed to foliage and branches in the field. Finally,

abdomens were constructed out of black Newplast® plasticine and attached to each facsimile.

Plasticine abdomens are often used in field predation experiments, and have been shown to allow

for effective detection of predation events through impressions left in the clay when predators

attempt to bite the facsimiles (Kikuchi & Pfennig, 2010; Niskanen & Mappes, 2005).

Predation experimental design

Using paired absolute prey density (low vs. high) and model unpalatability (unaltered vs.

experimentally enhanced) treatments across 100 independent field sites at La Selva Biological

Reserve, we assessed how variation in absolute prey density and palatability of the chemically

defended model *A. iphiclus* impacts predation rates on all facsimile types (Figure 1). Three

different butterfly facsimiles were included in all treatments: (1) *Adelpha iphiclus*, the toxic

model, (2) *Adelpha serpa*, the palatable Batesian mimic, and (3) *Junonia evarete*, a common

palatable butterfly species that was used as a control. To isolate the effect of absolute prey

density on facsimile survival, we maintained a 1:1:1 ratio of *A. iphiclus*, *A. serpa,* and *J. evarete*

phenotypes in all field sites, and doubled absolute density of all prey types in high-density

treatments relative to the low-density treatments (Figure 1). For low-density treatments, 20

facsimiles of each butterfly species were placed in each field site (*i.e.,* 60 facsimiles total/site).

Total facsimile density was doubled in high-density treatments, with 40 facsimiles of each

species in each field site (*i.e.,* 120 facsimiles total/site).

Low- and high-density treatments were paired with unaltered and experimentally

enhanced model unpalatability treatments to evaluate how model unpalatability mediates the

effect of prey density on predator foraging decisions (Figure 1). In the enhanced unpalatability

treatments, facsimiles of *A. iphiclus* were sprayed with a 2% Bitrex solution (denatonium

benzoate) to enhance this species' naturally occurring chemical defense. Bitrex is an extremely

bitter compound that has been shown to effectively drive avoidance learning in avian predators

(Curley et al., 2015; McLellan et al., 2023; Siddall & Marples, 2008; Winsor et al., 2020). Bitrex

was reapplied to *A. iphiclus* facsimiles daily in all enhanced unpalatability field sites. Palatability

was not experimentally manipulated in treatments with unaltered model unpalatability.

213   Each treatment (low-density, no Bitrex; low-density, Bitrex; high-density, no Bitrex; and

high-density, Bitrex) was replicated in 25 independent field sites at La Selva Biological Reserve,

resulting in 100 total field sites used in this experiment. Therefore, each low-density treatment

involved 500 facsimiles per species and 1,500 facsimiles total, while each high-density treatment

involved 1,000 facsimiles per species and 3,000 facsimiles total. Field sites were separated by at

least 250 meters, which aligns with estimated avian home range sizes in the Neotropics (Samuel

et al., 1985; Wang et al., 2007). This design minimizes the possibility that a small group of

predators were responsible for all facsimile attacks, and enables us to sample across independent

predator communities (Finkbeiner et al., 2012; Kristiansen et al., 2018). Within each site,

facsimiles were attached to foliage at least 1 meter high with dorsal wings facing upward. This

placement resembles perching behavior and ensures that facsimiles were visible to avian

predators. All facsimiles were checked daily for evidence of avian predation, which is often

characterized by triangular beak-shaped or deep puncture marks (Figure 2). Suspected predation

events were photographed, and facsimiles were reset daily by smoothing or replacing plasticine

abdomens. A maximum of one predation event was recorded for each facsimile on each day of

our experiment, and all facsimiles were removed after four days. All photographs were evaluated

by a team of researchers and categorized independently as "avian predation" or "non-avian

predation".

Statistical analysis

To test the hypothesis that increased absolute prey density and enhanced model unpalatability accelerate predator avoidance learning (H1), we fitted a mixed-effects Cox proportional hazards model using the *survival* and *coxme* packages (T. Therneau, 2015; T. M. Therneau, 2001). This time-to-event modeling approach allows the rate at which predation events occur to vary across levels of a response variable. Sites that experienced no predation events during the four-day experiment were excluded from all analyses. The rationale for this exclusion is that the absence of attacks across all facsimile types at these sites likely reflects a lack of insectivorous predator activity rather than treatment-specific behavioral responses, and therefore their inclusion could bias estimates of attack rates. Our initial model included species, absolute density, model unpalatability, and all two-way and three-way interactions as predictor variables to test whether the interactive effects of density and unpalatability differed across species. Following model selection, the final model included only the fixed effects of species, absolute density, and model unpalatability, with field site included as a random effect.

To test the hypothesis that increased absolute prey density and enhanced model unpalatability reduce overall predation on aposematic models and their Batesian mimics (H2), we fitted a generalized linear mixed-effects model (GLMM) with a binomial distribution using the *glmmTMB* package (Brooks et al., 2017). This analysis used the same subset of data as the survival analysis (*i.e.,* excluding sites with no recorded predator activity). Following the same model selection process outlined for the Cox proportional hazards model, the final GLMM included species, absolute density, and model unpalatability as fixed effects, and field site as a random effect.

Model selection for both analyses was based on comparisons of Akaike's Information

Criterion (AIC) values and likelihood ratio tests conducted using the *anova* function. Model

comparisons are presented in Table S1 (Cox proportional hazards model) and Table S2

(GLMM). Model assumptions for the GLMM were evaluated using the *DHARMa* package

(Hartig, 2022), which indicated no significant deviations from expectations under a binomial

distribution. For both models, we conducted pairwise contrasts of species using the *emmeans*

package (Lenth, 2023), and applied a Holm p-value correction to account for multiple

comparisons. All analyses for both models were performed with R statistical software (R Core

Team, 2022).

RESULTS

Across 100 independent field sites, 86 experienced at least one avian predation event (25

in the high-density, no Bitrex treatment; 22 in the high-density, Bitrex treatment; 19 in the low-

density, no Bitrex treatment; and 20 in the low-density, Bitrex treatment). All subsequent

analyses are based on these 86 field sites, with 14 field sites excluded.

Considering our first hypothesis regarding the rate of learning, we found that absolute

density, model unpalatability, and species all affected the rate at which predators learned to avoid

warning signals (Figure 3; Table 1). Specifically, our mixed-effects Cox proportional hazards

model showed that avian predators learn to avoid facsimiles significantly faster under high

absolute density conditions ($\chi^2 = 5.5503$, p = 0.01848) and under enhanced model unpalatability

conditions ($\chi^2 = 14.1156$, p = 0.00017). We also found a significant main effect of species ($\chi^2 =$

9.8344, p = 0.00732), and our analysis of pairwise contrasts demonstrated that predators learn to

avoid *A. iphiclus* (model) significantly faster than *A. serpa* (mimic; p = 0.0193) and *J. evarete*

(control; p = 0.0090).

Considering our second hypothesis regarding overall predation rate, we found that

absolute density, model unpalatability, and species all affected the overall predation rate.

Specifically, our generalized linear mixed effect model revealed significant main effects of

absolute density (χ^2 = 5.4996, p = 0.0190212), model unpalatability (χ^2 = 14.1333, p =

0.0001703), and species (χ^2 = 9.9402, p = 0.0069426) on overall predation rates (Figure 4; Table

2). We found that all facsimile types were attacked significantly less under high absolute density

conditions and conditions of enhanced model unpalatability (Figure 4). We also found that

facsimiles of *A. iphiclus* experience significantly less predation than facsimiles of *A. serpa* and *J.*

evarete (Table 2).

<u>DISCUSSION</u>

We hypothesized that increasing absolute prey density and enhancing model

unpalatability would i) accelerate predator avoidance learning and ii) reduce overall predation

pressure on the *Adelpha* mimicry complex. Consistent with this prediction, higher absolute prey

density, while holding model–mimic relative frequencies constant, increased the rate at which

predators learned to avoid our facsimiles and reduced overall predation on the facsimile prey

community. Similarly, enhanced model unpalatability accelerated predator learning and lowered

predation rates. The combination of high absolute density and enhanced model unpalatability

produced the lowest overall predation on the mimicry complex, suggesting that greater absolute

density together with stronger chemical defense most effectively amplified the model's warning

signal.

Across all treatments, predators learned to avoid *A. iphiclus* (model) faster than *A. serpa*

(mimic) and *J. evarete* (control), and the defended model experienced the lowest predation

overall. However, we did not find evidence of a significant interaction between species, absolute

density, and model unpalatability, suggesting that all three species (model, mimic, and control)

benefitted equally from these enhanced warning signals. These findings align with previous work

showing that greater model unpalatability can accelerate predator learning and relax selection on

imperfect mimics (Ihalainen et al., 2007; Lindström et al., 1997; Pike & Burman, 2023). The

comparable benefit observed for the palatable control further suggests that the protective effects

of heightened warning signals may extend to particularly poor mimics, which aligns with

previous work demonstrating that protection for poor mimics is greater in complex environments

with high signal diversity (Beatty et al., 2004; Ihalainen et al., 2012; Sherratt & Peet-Paré, 2017).

Although treatment effects did not differ significantly among species, predators

consistently learned to avoid the chemically defended model more quickly than either the mimic

or the control, and the model maintained the lowest predation rates across all treatments. Thus,

while strengthening the model's warning signal enhanced protection for imperfect mimics,

aposematic prey still derived the greatest benefit. This pattern is consistent with the expectation

that stronger warning signals elicit stronger avoidance responses (Skowron Volponi et al., 2025),

and previous work demonstrating that increasing absolute density increases protection for

aposematic prey (Briolat et al., 2019; Jeschke & Tollrian, 2000). However, our results do not

align with studies suggesting that Batesian mimicry can be parasitic (Aubier et al., 2017;

Heerwig et al., 2023). In contrast, we found that increasing absolute prey density while

maintaining constant relative frequencies reduced predation on both the aposematic model and

its imperfect mimic. This result suggests that mimicry may only become parasitic when mimics

are relatively more abundant than their models and that increasing the absolute density of the

mimicry complex may permit increased mimetic abundance without generating parasitic effects

(Rowland, Mappes, et al., 2010). To our knowledge, this study represents the first field-based

predation experiment to isolate absolute density effects on predator foraging by increasing

absolute prey density while maintaining a constant 1:1:1 ratio of models, mimics, and controls.

Negative frequency dependent selection shapes protection for imperfect mimics, where

decreasing the abundance of a mimic relative to its defended model increases protection for the

mimic (Pfennig et al., 2001; Ruxton et al., 2018). This pattern has been documented in several

mimicry systems, including the *Adelpha* mimicry complex (Chouteau et al., 2016; Finkbeiner et

al., 2018; Shine et al., 2022). Previous work has shown that in Costa Rica, avian predators can

distinguish between the toxic model *A. iphiclus* and the imperfect mimic *A. serpa* despite their

striking phenotypic similarity, likely because both species occur at similar population densities

(Finkbeiner et al., 2017, 2018). In contrast, protection for *A. serpa* increases in Ecuador, where

this Batesian mimic is much rarer relative to its model, underscoring the role of relative

abundance in driving relaxed selection on imperfect mimics (Finkbeiner et al., 2018). Negative

frequency dependent selection is thought to operate by increasing the rate at which predators

encounter the model relative to the mimic, thereby strengthening the model's warning signal

(Ioannou et al., 2008; Lindström et al., 1997; Luedeman et al., 1981). Our results suggest that

increasing the absolute density of a mimetic community, and thereby increasing predator

encounter rates with all prey types equally (Endler & Mappes, 2004; Ioannou et al., 2008; Mols

et al., 2004), can similarly strengthen the model's warning signal. This finding indicates that a

higher absolute abundance of mimics may not dilute the model's warning signal when relative

frequencies are maintained. As such, we highlight an underappreciated role for the absolute

density of a mimicry complex, independent of model–mimic relative frequencies, in shaping

relaxed selection on imperfect mimicry.

Although our experiment did not directly account for alternative prey availability or

predator hunger, both factors likely influenced predator decision-making and represent important

limitations of this study. When alternative prey are abundant, the immediate payoff of sampling

imperfect mimics is low, reducing the incentive for predators to invest in learning fine-scale

distinctions between aposematic models and their mimics and favoring broad generalization of

warning signals (Kokko et al., 2003; Lindström et al., 2004; Sherratt & Peet-Paré, 2017). La

Selva supports a highly diverse insect community that likely provides ample alternative food

sources for insectivorous birds (MacDade et al., 1994). While we did not directly measure the

density of alternative prey at La Selva, this ecological context suggests that predators in our field

sites could afford to generalize avoidance of the model (Dill, 1975; Sherratt, 2003), driving

reduced predation on *A. iphiclus,* its mimic *A. serpa*, and the palatable control *J. evarete* in our

experiment when that model's warning signal was strengthened. Together, these patterns are

consistent with an information-acquisition mechanism underlying relaxed selection on imperfect

Batesian mimics. Although avian predators are capable of fine-scale discrimination between

similar phenotypes—including *A. iphiclus* and *A. serpa* (Finkbeiner et al., 2017; Langham, 2004;

Llaurens et al., 2014) they are expected to use this information only when the benefits outweigh

the costs (Sherratt & Peet-Paré, 2017). The observed reduced predation on the palatable control

when the model's warning signal was strengthened suggests that increasing the costs associated

sampling facsimiles can promote global avoidance, with predators rejecting all facsimiles in

favor of alternative food sources.

Predator motivation to distinguish between models and mimics is also likely influenced by

hunger levels and the relative nutritional value of mimetic prey compared to alternative food

sources (Halpin et al., 2014; Kaczmarek et al., 2020; Kokko et al., 2003; Luedeman et al., 1981).

Predators have been shown to forage on toxic prey or learn to differentiate toxic prey from

phenotypically similar mimics when hunger is extreme, when defended prey offer high

nutritional rewards, or when alternative food sources are scarce (Barnett et al., 2007; Halpin et

al., 2014; Kokko et al., 2003; Lindström et al., 2004; Skelhorn & Rowe, 2007; Veselý et al.,

2016). While our findings provide novel insight into absolute density as a mechanism underlying

imperfect mimicry, our experimental design did not allow us to assess how variation in predator

hunger or prey nutritional value modulates these dynamics. Our experiment did not provide

predators with a food reward, which may have contributed to the extreme generalization we

observed. Indeed, if our facsimiles had provided nutritional value to predators, predators may

have been sufficiently incentivized to differentiate between the different phenotypes, and this is

an important area for future work.

While our observed attack rates were lower than expected based on previous work

conducted in the tropics (Finkbeiner et al., 2018; Seymoure & Aiello, 2015), they are comparable

to several other experiments that used artificial models to assess avian predation (Kristiansen et

al., 2018; Palmer et al., 2018; Robinson et al., 2025). We consider that these relatively low attack

rates reflect our conservative predation scoring criteria and the use of a negative stimulus in the

absence of a food reward. We also observed substantial variation in predator activity among field

sites; for example, all sites in the high-density, no Bitrex treatment experienced at least one

predation event, whereas predator activity was detected at only 19 of 25 field sites in the low-

density, no Bitrex treatment. Although filtering our data to remove sites that experienced no

predation and including site as a random effect in our model helps account for this heterogeneity,

some uncertainty is inherent to field-based predation experiments. Our results highlight the

potential for absolute density of mimetic communities to impact protection for imperfect

Batesian mimics and provide a critical foundation for future work to leverage controlled

experiments that could further disentangle the mechanisms underlying absolute density effects

on predator foraging decisions.

Overall, our results align with previous work demonstrating that increased unpalatability

of defended models enhances protection for aposematic prey and their imperfect mimics, and

further suggest that, in complex natural environments with abundant alternative prey, the degree

of mimetic perfection needed to benefit from enhanced warning signals may be broader than

previously appreciated. Our findings also indicate that absolute density effects can strengthen

warning signals in nature, driving stronger predator avoidance responses through mechanisms

analogous to those associated with variation in model unpalatability and model–mimic relative

frequencies (Endler & Mappes, 2004; Pfennig et al., 2001). Considered within the broader

mimicry literature, this work underscores the importance of distinguishing between relative

frequency and absolute density when testing hypotheses about relaxed selection in the context of

mimetic prey. However, more work is needed to clarify how factors such as alternative prey

availability, predator hunger, and the nutritional value of unpalatable prey modulate the effects

of absolute prey density on protection for imperfect Batesian mimics.

DATA AVAILABILITY: Data from the field predation experiments and code files for all

analyses can be found at https://github.com/butterfliesrcool/Absolute_Density_Project.

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 FIGURES & TABLES

Figure 1. Schematic representation of paired density (low vs. high) and unpalatability (unaltered vs. enhanced) experimental treatments. To isolate absolute density effects on predator foraging, the relative frequency of model and mimic and control facsimiles was held constant at a 1:1:1 ratio across treatments and absolute prey density was doubled in the high-density treatments.

Alt Text: Graphical representation of paired experimental design with unaltered vs. enhanced unpalatability treatments and low vs. high density treatments where absolute community density is doubled in high density treatments

431
Figure 2. (A) Approximate size of *A. iphiclus* facsimiles. Wingspan was consistent across
facsimile types. Images show evidence of avian predation on facsimiles in the field for *A. iphiclus*
(B), *A. serpa* (C), and *J. evarete* (D).

Alt Text: Panels A through D show images of facsimiles. Panel A illustrates that facsimiles are
approximately 5 cm in height, while panels B through D show images of *A. phiclus*, *A. serpa*,
and *J. evarete* facsimiles, respectively. All images show evidence of avian predation, which is
identified through triangular beak-shaped marks or deep puncture marks in clay absomens.

440
441 **Figure 3.** Kaplan-Meier survival curves comparing the fixed effect of (A) species, (B) absolute
442 density, and (C) model unpalatability on facsimile survival, demonstrating the speed of predator
443 avoidance learning across predictor variables. Asterisk indicates significant difference (p < 0.05)
444 in survival probabilities based on mixed-effects Cox proportional hazards model and pairwise
445 comparisons across species.

Figure 4. Estimated fixed effects of species, density, and unpalatability on facsimile attack rates based on results of generalized linear mixed model with binomial distribution. Color indicates unpalatability condition (orange = no added unpalatability, blue = unpalatability of *A. iphiclus* facsimiles was enhanced with a 2% Bitrex solution). Error bars represent 95% confidence intervals.

465 **Table 1.** Results of best fit mixed-effects Cox proportional hazards regression model evaluating
466 effect of species, absolute prey density, and model unpalatability on facsimile attack rates over
467 time as a proxy for predator learning speed. Significant p-values are shown in bold (* p < 0.05, **
468 p <0.01, *** p < 0.001)

fixed effect	DF	χ^2	hazard ratio (95% CI)	p-value
species	2	9.8344		**0.00732 ****
A. iphiclus (reference)				
A. serpa			1.46 (1.1, 1.95)	
J. evarete			1.54 (1.16, 2.05)	
absolute prey density	1	5.5503	0.74 (0.57, 0.95)	**0.01848 ***
model unpalatability	1	14.1156	0.62 (0.48, 0.79)	**0.00017 *****

469
470
471 **Table 2.** Results of best fit generalized linear mixed model with binomial distribution testing fixed
472 effect of species, model unpalatability, and absolute prey density on facsimile attack rates. Field
473 site was included as a random effect. Results of pairwise contrasts of species are shown; p-values
474 for all pairwise comparisons were adjusted using the Holm method for multiple comparisons.
475 Significant p-values are shown in bold (* p < 0.05, ** p <0.01, *** p < 0.001)

fixed effects	χ^2	df	p-value	
species	9.9402	2	**0.0069426 ****	
model unpalatability	14.1333	1	**0.0001703 *****	
absolute prey density	5.4996	1	**0.0190212 ***	
random effects	**variance**	**standard deviation**		
field site	0.05342	0.2311		
pairwise contrast	**estimate**	**standard error**	**z-ratio**	**p-value**
A. iphiclus – A. serpa	-0.3834	0.147	-2.601	**0.0186***
A. iphiclus – J. evarete	-0.4353	0.146	-2.984	**0.0085****
A. serpa – J. evarete	-0.0519	0.131	-0.395	0.6926

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REFERENCES

Alatalo, R. V., & Mappes, J. (1996). Tracking the evolution of warning signals. *Nature*, *382*(6593), 708–710. https://doi.org/10.1038/382708a0

Aubier, T. G., Joron, M., & Sherratt, T. N. (2017). Mimicry among Unequally Defended Prey Should Be Mutualistic When Predators Sample Optimally. *The American Naturalist*, *189*(3), 267–282. https://doi.org/10.1086/690121

Barnett, C., Bateson, M., & Rowe, C. (2007). State-dependent decision making: Educated predators strategically trade off the costs and benefits of consuming aposematic prey. *Behavioral Ecology*, *18*(4), 645–651. https://doi.org/10.1093/beheco/arm027

Bates, H. W. (1862). XLVIII.—*Contributions to an insect fauna of the Amazon valley* C OLEOPTERA: L ONGICORNES. *Annals and Magazine of Natural History*, *9*(54), 446–458. https://doi.org/10.1080/00222936208681261

Beatty, C. D., Beirinckx, K., & Sherratt, T. N. (2004). The evolution of müllerian mimicry in multispecies communities. *Nature*, *431*(7004), 63–66. https://doi.org/10.1038/nature02818

Briolat, E. S., Burdfield-Steel, E. R., Paul, S. C., Rönkä, K. H., Seymoure, B. M., Stankowich, T., & Stuckert, A. M. M. (2019). Diversity in warning coloration: Selective paradox or the norm? *Biological Reviews*, *94*(2), 388–414. https://doi.org/10.1111/brv.12460

Brooks, M., E., Kristensen, K., Benthem, K., J. ,van, Magnusson, A., Berg, C., W., Nielsen, A., Skaug, H., J., Mächler, M., & Bolker, B., M. (2017). glmmTMB Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling. *The R Journal*, *9*(2), 378. https://doi.org/10.32614/RJ-2017-066

Brower, L. P., Ryerson, W. N., Coppinger, L. L., & Glazier, S. C. (1968). Ecological Chemistry

and the Palatability Spectrum. *Science*, *161*(3848), 1349–1350.

https://doi.org/10.1126/science.161.3848.1349

Chouteau, M., Arias, M., & Joron, M. (2016). Warning signals are under positive frequency-

dependent selection in nature. *Proceedings of the National Academy of Sciences*, *113*(8),

2164–2169. https://doi.org/10.1073/pnas.1519216113

Curley, E. A. M., Rowley, H. E., & Speed, M. P. (2015). A field demonstration of the costs and

benefits of group living to edible and defended prey. *Biology Letters*, *11*(6), 20150152.

https://doi.org/10.1098/rsbl.2015.0152

Dill, L. M. (1975). Calculated risk-taking by predators as a factor in Batesian mimicry. *Canadian*

Journal of Zoology, *53*(11), 1614–1621. https://doi.org/10.1139/z75-195

Ebel, E. R., DaCosta, J. M., Sorenson, M. D., Hill, R. I., Briscoe, A. D., Willmott, K. R., &

Mullen, S. P. (2015). Rapid diversification associated with ecological specialization in

Neotropical *Adelpha* butterflies. *Molecular Ecology*, *24*(10), 2392–2405.

https://doi.org/10.1111/mec.13168

Edmunds, M. (2000). Why are there good and poor mimics? *Biological Journal of the Linnean*

Society, *70*(3), 459–466. https://doi.org/10.1111/j.1095-8312.2000.tb01234.x

Endler, J. A., & Mappes, J. (2004). Predator Mixes and the Conspicuousness of Aposematic

Signals. *The American Naturalist*, *163*(4), 532–547. https://doi.org/10.1086/382662

Finkbeiner, S. D., Briscoe, A. D., & Mullen, S. P. (2017). Complex dynamics underlie the

evolution of imperfect wing pattern convergence in butterflies. *Evolution*, *71*(4), 949–

959. https://doi.org/10.1111/evo.13165

Finkbeiner, S. D., Briscoe, A. D., & Reed, R. D. (2012). The benefit of being a social butterfly:

Communal roosting deters predation. *Proceedings of the Royal Society B: Biological*

Sciences, *279*(1739), 2769–2776. https://doi.org/10.1098/rspb.2012.0203

Finkbeiner, S. D., Salazar, P. A., Nogales, S., Rush, C. E., Briscoe, A. D., Hill, R. I., Kronforst,

M. R., Willmott, K. R., & Mullen, S. P. (2018). Frequency dependence shapes the

adaptive landscape of imperfect Batesian mimicry. *Proceedings of the Royal Society B:*

Biological Sciences, *285*(1876), 20172786. https://doi.org/10.1098/rspb.2017.2786

Getty, T. (1985). Discriminability and the Sigmoid Functional Response: How Optimal Foragers

Could Stabilize Model-Mimic Complexes. *The American Naturalist*, *125*(2), 239–256.

https://doi.org/10.1086/284339

Halpin, C. G., Skelhorn, J., & Rowe, C. (2014). Increased predation of nutrient-enriched

aposematic prey. *Proceedings of the Royal Society B: Biological Sciences*, *281*(1781),

20133255. https://doi.org/10.1098/rspb.2013.3255

Harper, G. R., & Pfennig, D. W. (2007). Mimicry on the edge: Why do mimics vary in

resemblance to their model in different parts of their geographical range? *Proceedings of*

the Royal Society B: Biological Sciences, *274*(1621), 1955–1961.

https://doi.org/10.1098/rspb.2007.0558

Hartig, F. (2022). *DHARMa: Residual Diagnostics for Hierarchical (Multi-Level / Mixed)*

Regression Models (Version R package version 0.4.6) [Computer software].

https://CRAN.R-project.org/package=DHARMa

Heerwig, O. T., Jain-Schlaepfer, S. M. R., Sherratt, T. N., & Kikuchi, D. W. (2023). Effects of

predator associative learning and innate aversion on mimicry complexes. *Evolutionary*

Ecology, *37*(4), 709–720. https://doi.org/10.1007/s10682-023-10238-4

Ihalainen, E., Lindström, L., & Mappes, J. (2007). Investigating Müllerian mimicry: Predator

learning and variation in prey defences. *Journal of Evolutionary Biology*, *20*(2), 780–791.

https://doi.org/10.1111/j.1420-9101.2006.01234.x

Ihalainen, E., Rowland, H. M., Speed, M. P., Ruxton, G. D., & Mappes, J. (2012). Prey

community structure affects how predators select for Müllerian mimicry. *Proceedings of*

the Royal Society B: Biological Sciences, *279*(1736), 2099–2105.

https://doi.org/10.1098/rspb.2011.2360

Ioannou, C. C., Ruxton, G. D., & Krause, J. (2008). Search rate, attack probability, and the

relationship between prey density and prey encounter rate. *Behavioral Ecology*, *19*(4),

842–846. https://doi.org/10.1093/beheco/arn038

Iserbyt, A., Bots, J., Van Dongen, S., Ting, J. J., Van Gossum, H., & Sherratt, T. N. (2011).

Frequency-dependent variation in mimetic fidelity in an intraspecific mimicry system.

Proceedings of the Royal Society B: Biological Sciences, *278*(1721), 3116–3122.

https://doi.org/10.1098/rspb.2011.0126

Jeschke, J. M., & Tollrian, R. (2000). Density-dependent effects of prey defences. *Oecologia*,

123(3), 391–396. https://doi.org/10.1007/s004420051026

Kaczmarek, J. M., Kaczmarski, M., Mazurkiewicz, J., & Kloskowski, J. (2020). Numbers,

neighbors, and hungry predators: What makes chemically defended aposematic prey

susceptible to predation? *Ecology and Evolution*, *10*(24), 13705–13716.

https://doi.org/10.1002/ece3.6956

Kessler, A., & Baldwin, I. T. (2002). Plant responses to insect herbivory: The emerging

molecular analysis. *Annual Review of Plant Biology*, *53*(1), 299–328.

https://doi.org/10.1146/annurev.arplant.53.100301.135207

Kikuchi, D. W., Barfield, M., Herberstein, M. E., Mappes, J., & Holt, R. D. (2022). The Effect of

Predator Population Dynamics on Batesian Mimicry Complexes. *The American*

Naturalist, *199*(3), 406–419. https://doi.org/10.1086/718162

Kikuchi, D. W., Herberstein, M. E., Barfield, M., Holt, R. D., & Mappes, J. (2021). Why aren't

warning signals everywhere? On the prevalence of aposematism and mimicry in

communities. *Biological Reviews*, *96*(6), 2446–2460. https://doi.org/10.1111/brv.12760

Kikuchi, D. W., & Pfennig, D. W. (2010). High-model abundance may permit the gradual

evolution of Batesian mimicry: An experimental test. *Proceedings of the Royal Society B:*

Biological Sciences, *277*(1684), 1041–1048. https://doi.org/10.1098/rspb.2009.2000

Kikuchi, D. W., & Pfennig, D. W. (2013). Imperfect Mimicry and the Limits of Natural

Selection. *The Quarterly Review of Biology*, *88*(4), 297–315.

https://doi.org/10.1086/673758

Kokko, H., Mappes, J., & Lindström, L. (2003). Alternative prey can change model–mimic

dynamics between parasitism and mutualism. *Ecology Letters*, *6*(12), 1068–1076.

https://doi.org/10.1046/j.1461-0248.2003.00532.x

Kristiansen, E. B., Finkbeiner, S. D., Hill, R. I., Prusa, L., & Mullen, S. P. (2018). Testing the

adaptive hypothesis of Batesian mimicry among hybridizing North American admiral

butterflies. *Evolution*, *72*(7), 1436–1448. https://doi.org/10.1111/evo.13488

Langham, G. M. (2004). SPECIALIZED AVIAN PREDATORS REPEATEDLY ATTACK

NOVEL COLOR MORPHS OF HELICONIUS BUTTERFLIES. *Evolution*, *58*(12),

2783–2787. https://doi.org/10.1111/j.0014-3820.2004.tb01629.x

Lenth, R. (2023). *emmeans: Estimated Marginal Means, aka Least-Squares Means* (Version R

591   package version 1.8.9) [Computer software]. https://CRAN.R-

592   project.org/package=emmeans

593   Lindström, L., Alatalo, R. V., Lyytinen, A., & Mappes, J. (2001). Strong antiapostatic selection

594   against novel rare aposematic prey. *Proceedings of the National Academy of Sciences*,

595   *98*(16), 9181–9184. https://doi.org/10.1073/pnas.161071598

596   Lindström, L., Alatalo, R. V., Lyytinen, A., & Mappes, J. (2004). THE EFFECT OF

597   ALTERNATIVE PREY ON THE DYNAMICS OF IMPERFECT BATESIAN AND

598   MÜLLERIAN MIMICRIES. *Evolution*, *58*(6), 1294–1302.

599   https://doi.org/10.1111/j.0014-3820.2004.tb01708.x

600   Lindström, L., Alatalo, R. V., & Mappes, J. (1997). Imperfect Batesian mimicry—The effects of

601   the frequency and the distastefulness of the model. *Proceedings of the Royal Society of*

602   *London. Series B: Biological Sciences*, *264*(1379), 149–153.

603   https://doi.org/10.1098/rspb.1997.0022

604   Llaurens, V., Joron, M., & Théry, M. (2014). Cryptic differences in colour among Müllerian

605   mimics: How can the visual capacities of predators and prey shape the evolution of wing

606   colours? *Journal of Evolutionary Biology*, *27*(3), 531–540.

607   https://doi.org/10.1111/jeb.12317

608   Lopes, S., Von Poser, G. L., Kerber, V. A., Farias, F. M., Konrath, E. L., Moreno, P., Sobral, M.

609   E., Zuanazzi, J. A. S., & Henriques, A. T. (2004). Taxonomic significance of alkaloids

610   and iridoid glucosides in the tribe Psychotrieae (Rubiaceae). *Biochemical Systematics*

611   *and Ecology*, *32*(12), 1187–1195. https://doi.org/10.1016/j.bse.2004.04.015

612   Lotka, A. J., & Volterra, V. (1925). Predator-prey model. *Elements of Physical Biology*.

Luedeman, J. K., McMorris, F. R., & Warner, D. D. (1981). Predators Encountering a Model-Mimic System with Alternative Prey. *The American Naturalist*, *117*(6), 1040–1048. https://doi.org/10.1086/283794

MacDade, L. A., Bawa, K. S., Hespenheide, H. A., & Hartshorn, G. S. (Eds.). (1994). *La Selva: Ecology and natural history of a neotropical rain forest*. University of Chicago Press.

Malcolm, S. B. (1990). Mimicry: Status of a classical evolutionary paradigm. *Trends in Ecology & Evolution*, *5*(2), 57–62. https://doi.org/10.1016/0169-5347(90)90049-J

McLellan, C. F., Cuthill, I. C., & Montgomery, S. H. (2023). Warning Coloration, Body Size, and the Evolution of Gregarious Behavior in Butterfly Larvae. *The American Naturalist*, *202*(1), 64–77. https://doi.org/10.1086/724818

Mols, C. M. M., Van Oers, K., Witjes, L. M. A., Lessells, C. M., Drent, P. J., & Visser, M. E. (2004). Central assumptions of predator–prey models fail in a semi–natural experimental system. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, *271*(suppl_3). https://doi.org/10.1098/rsbl.2003.0110

Mullen, S. P., Savage, W. K., Wahlberg, N., & Willmott, K. R. (2011). Rapid diversification and not clade age explains high diversity in neotropical *Adelpha* butterflies. *Proceedings of the Royal Society B: Biological Sciences*, *278*(1713), 1777–1785. https://doi.org/10.1098/rspb.2010.2140

Nelson, D. W. M., Crossland, M. R., & Shine, R. (2011). Foraging responses of predators to novel toxic prey: Effects of predator learning and relative prey abundance. *Oikos*, *120*(1), 152–158. https://doi.org/10.1111/j.1600-0706.2010.18736.x

Niskanen, M., & Mappes, J. (2005). Significance of the dorsal zigzag pattern of *Vipera latastei*

gaditana against avian predators. *Journal of Animal Ecology*, *74*(6), 1091–1101.

https://doi.org/10.1111/j.1365-2656.2005.01008.x

Owen, R. E., & Owen, A. R. G. (1984). Mathematical paradigms for mimicry: Recurrent

sampling. *Journal of Theoretical Biology*, *109*(2), 217–247.

https://doi.org/10.1016/S0022-5193(84)80004-1

Páez V., E., Kergoat, G. J., Chazot, N., Benmesbah, M., Briscoe, A. D., Finkbeiner, S. D.,

Freitas, A. V. L., Guralnick, R. P., Hill, R. I., Kronforst, M. R., Magaldi, L. M., Mullen,

S. P., Nakamura, I., Owens, H. L., Wahlberg, N., Woodbury, M., Elias, M., & Willmott,

K. R. (2025). Phylogeny, systematics and evolution of mimicry patterns in Neotropical

limenitidine butterflies. *Systematic Entomology*, e70007.

https://doi.org/10.1111/syen.70007

Palmer, D. H., Tan, Y. Q., Finkbeiner, S. D., Briscoe, A. D., Monteiro, A., & Kronforst, M. R.

(2018). Experimental field tests of Batesian mimicry in the swallowtail butterfly *Papilio*

polytes. *Ecology and Evolution*, *8*(15), 7657–7666. https://doi.org/10.1002/ece3.4207

Penney, H. D., Hassall, C., Skevington, J. H., Abbott, K. R., & Sherratt, T. N. (2012). A

comparative analysis of the evolution of imperfect mimicry. *Nature*, *483*(7390), 461–464.

https://doi.org/10.1038/nature10961

Pfennig, D. W., Harcombe, W. R., & Pfennig, K. S. (2001). Frequency-dependent Batesian

mimicry. *Nature*, *410*(6826), 323–323. https://doi.org/10.1038/35066628

Pike, T. W., & Burman, O. H. P. (2023). Model aversiveness and the evolution of imperfect

Batesian mimics. *Behavioral Ecology*, *34*(5), 907–912.

https://doi.org/10.1093/beheco/arad063

Prudic, K. L., Skemp, A. K., & Papaj, D. R. (2007). Aposematic coloration, luminance contrast,

and the benefits of conspicuousness. *Behavioral Ecology*, *18*(1), 41–46.

https://doi.org/10.1093/beheco/arl046

Prudic, K. L., Timmermann, B. N., Papaj, D. R., Ritland, D. B., & Oliver, J. C. (2019). Mimicry

in viceroy butterflies is dependent on abundance of the model queen butterfly.

Communications Biology, *2*(1), 68. https://doi.org/10.1038/s42003-019-0303-z

R Core Team. (2022). *R: A Language and Environment for Statistical Computing. R Foundation*

for Statistical Computing [Computer software]. https://www.R-project.org/

Ritland, D. B. (1994). Variation in Palatability of Queen Butterflies (Danaus Gilippus) and

Implications Regarding Mimicry. *Ecology*, *75*(3), 732–746.

https://doi.org/10.2307/1941731

Ritland, D. B. (1995). Comparative unpalatability of mimetic viceroy butterflies (Limenitis

archippus) from four south-eastern United States populations. *Oecologia*, *103*(3), 327–

336. https://doi.org/10.1007/bf00328621

Robinson, A. E., Novick, I., Herrmann, J., DeFelice, L., Engel, A., Famin, D., Fetherston, C.,

Frintu, B., Meyersiek, J., Mishi, M., Nguyễn, T. G. H., Buston, P. M., Sherratt, T. N., &

Mullen, S. P. (2025). Is temporal synchrony necessary for effective Batesian mimicry?

Proceedings of the Royal Society B: Biological Sciences, *292*(2039), 20241737.

https://doi.org/10.1098/rspb.2024.1737

Rowland, H. M., Hoogesteger, T., Ruxton, G. D., Speed, M. P., & Mappes, J. (2010). A tale of 2

signals: Signal mimicry between aposematic species enhances predator avoidance

learning. *Behavioral Ecology*, *21*(4), 851–860. https://doi.org/10.1093/beheco/arq071

Rowland, H. M., Ihalainen, E., Lindström, L., Mappes, J., & Speed, M. P. (2007). Co-mimics have a mutualistic relationship despite unequal defences. *Nature*, *448*(7149), 64–67. https://doi.org/10.1038/nature05899

Rowland, H. M., Mappes, J., Ruxton, G. D., & Speed, M. P. (2010). Mimicry between unequally defended prey can be parasitic: Evidence for quasi-Batesian mimicry. *Ecology Letters*, *13*(12), 1494–1502. https://doi.org/10.1111/j.1461-0248.2010.01539.x

Ruxton, G. D., Allen, W. L., Sherratt, T. N., & Speed, M. P. (2018). *Avoiding attack: The evolutionary ecology of crypsis, aposematism, and mimicry* (Second edition). Oxford University Press.

Samuel, M. D., Pierce, D. J., & Garton, E. O. (1985). Identifying Areas of Concentrated Use within the Home Range. *The Journal of Animal Ecology*, *54*(3), 711. https://doi.org/10.2307/4373

Seymoure, B. M., & Aiello, A. (2015). Keeping the band together: Evidence for false boundary disruptive coloration in a butterfly. *Journal of Evolutionary Biology*, *28*(9), 1618–1624. https://doi.org/10.1111/jeb.12681

Sherratt, T. N. (2002). The evolution of imperfect mimicry. *Behavioral Ecology*, *13*(6), 821–826. https://doi.org/10.1093/beheco/13.6.821

Sherratt, T. N. (2003). State-dependent risk-taking by predators in systems with defended prey. *Oikos*, *103*(1), 93–100. https://doi.org/10.1034/j.1600-0706.2003.12576.x

Sherratt, T. N. (2011). THE OPTIMAL SAMPLING STRATEGY FOR UNFAMILIAR PREY: THE OPTIMAL SAMPLING STRATEGY FOR UNFAMILIAR PREY. *Evolution*, *65*(7), 2014–2025. https://doi.org/10.1111/j.1558-5646.2011.01274.x

Sherratt, T. N., & Peet-Paré, C. A. (2017). The perfection of mimicry: An information approach. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *372*(1724), 20160340. https://doi.org/10.1098/rstb.2016.0340

Shine, R., Brown, G. P., & Goiran, C. (2022). Frequency-dependent Batesian mimicry maintains colour polymorphism in a sea snake population. *Scientific Reports*, *12*(1), 4680. https://doi.org/10.1038/s41598-022-08639-6

Siddall, E. C., & Marples, N. M. (2008). Better to be bimodal: The interaction of color and odor on learning and memory. *Behavioral Ecology*, *19*(2), 425–432. https://doi.org/10.1093/beheco/arm155

Skelhorn, J., Halpin, C. G., & Rowe, C. (2016). Learning about aposematic prey. *Behavioral Ecology*, *27*(4), 955–964. https://doi.org/10.1093/beheco/arw009

Skelhorn, J., & Rowe, C. (2006). Prey palatability influences predator learning and memory. *Animal Behaviour*, *71*(5), 1111–1118. https://doi.org/10.1016/j.anbehav.2005.08.011

Skelhorn, J., & Rowe, C. (2007). Predators' Toxin Burdens Influence Their Strategic Decisions to Eat Toxic Prey. *Current Biology*, *17*(17), 1479–1483. https://doi.org/10.1016/j.cub.2007.07.064

Skelhorn, J., Rowland, H. M., Delf, J., Speed, M. P., & Ruxton, G. D. (2011). Density-dependent predation influences the evolution and behavior of masquerading prey. *Proceedings of the National Academy of Sciences*, *108*(16), 6532–6536. https://doi.org/10.1073/pnas.1014629108

Skowron Volponi, M., Dapporto, L., Casacci, L. P., Barbero, F., Chiesa, E., & Volponi, P. (2025). How effective are insect aposematism and Batesian mimicry in deterring a wild avian predator? *Ecology*, *106*(6), e70135. https://doi.org/10.1002/ecy.70135

Soto-Sobenis, A., Castillo, B., Delgado, A., González, A., & Montenegro, R. (2001). Alkaloid

Screening of Herbarium Samples of Rubiaceae from Panama. *Pharmaceutical Biology*,

39(3), 161–169. https://doi.org/10.1076/phbi.39.3.161.5925

Speed, M. P., & Turner, J. R. G. (1999). Learning and memory in mimicry: II. Do we understand

the mimicry spectrum? *Biological Journal of the Linnean Society*, *67*(3), 281–312.

https://doi.org/10.1111/j.1095-8312.1999.tb01935.x

Therneau, T. (2015). *Mixed effects Cox models* [Computer software]. CRAN repository.

https://CRAN.R-project.org/package=coxme

Therneau, T. M. (2001). *survival: Survival Analysis* (p. 3.8-3) [Dataset].

https://doi.org/10.32614/CRAN.package.survival

Turner, J. R. G., Kearney, E. P., & Exton, L. S. (1984). Mimicry and the Monte Carlo predator:

The palatability spectrum, and the origins of mimicry. *Biological Journal of the Linnean*

Society, *23*(2–3), 247–268. https://doi.org/10.1111/j.1095-8312.1984.tb00143.x

Veselý, P., Ernestová, B., Nedvěd, O., & Fuchs, R. (2016). Do predator energy demands or

previous exposure influence protection by aposematic coloration of prey? *Current*

Zoology, zow057. https://doi.org/10.1093/cz/zow057

Wang, N., Zhang, Y., & Zheng, G. (2007). Home ranges and habitat vegetation characters in

breeding season of Narcissus Flycatcher and Yellow-rumped Flycatcher. *Frontiers of*

Biology in China, *2*(3), 345–350. https://doi.org/10.1007/s11515-007-0051-1

Willmott, K. R. (2003). Cladistic analysis of the Neotropical butterfly genus *Adelpha*

(Lepidoptera: Nymphalidae), with comments on the subtribal classification of

Limenitidini. *Systematic Entomology*, *28*(3), 279–322. https://doi.org/10.1046/j.1365-

3113.2003.00209.x

Winsor, A. M., Ihle, M., & Taylor, L. A. (2020). Methods for independently manipulating

palatability and color in small insect prey. *PLOS ONE*, *15*(4), e0231205.

https://doi.org/10.1371/journal.pone.0231205

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