

A Complete Public Domain Family Genomics Dataset

Manuel Corpas, Mike Cariaso, Alain Coletta, David Weiss, Andrew Harrison, Federico Moran, Huanming Yang

Abstract

BACKGROUND: The availability of open access genomic data is essential for the personal genomics field. Public genomic data allow comparative analyses, testing of new tools and genotype-phenotype association studies. Personal genomics data of unrelated individuals are available in the public domain, notably the Personal Genome Project; however, to date genomics family data and metadata are severely lacking, mainly due to cost, privacy concerns or restricted access to Next Generation Sequencing (NGS) technology. Family data have a lot to offer as they allow the study of heritability, something which is impossible to do just by using unrelated individuals.

FINDINGS: A whole family from Southern Spain decided to genotype, sequence and analyse their personal genomes making them publicly available under a Creative Commons 0 license (CC0; commonly denominated as public domain). These data include a) five 23andMe SNP chip genotype bed files, b) four raw exomes with their assorted bam files and VCF files, c) a metagenomic raw sequencing data file and d) derived data of likely phenotypes using SNPedia-derived tools.

CONCLUSIONS: To our knowledge this is the first CC0 released set of genomic, phenotypic and metagenomic data for a whole family. This dataset is also unique in that it was obtained through direct-to-consumer genetic tests. Hence any ordinary citizen with enough budget and samples should be able to reproduce this experiment. We envisage this dataset to be a useful resource for a variety of applications in the personal genomics field as a) negative control data for trait association discovery, b) testing data for development of new software and c) sample data for heritability studies. We encourage prospective users to share with us derived results so that they can be added to our existing collection.

Keywords: Whole Exome Sequencing, Genotypes, Phenotypes, Variation, Personal Genomics, Family Analysis

Data Description

As the price of genome sequencing and all other assorted sequencing techniques become more affordable, this technology is expected to be adopted among the general population in the future. Next Generation Sequencing (NGS) experiments are data rich, but the interpretation of personal genomes remains a challenge as knowledge is spread across many different resources, many of them closed source or with no metadata with which to perform automatic computational analyses. This lack of open source software and tools is in part due to absence of data models with which to test and develop new applications. Family models readily available for development, testing and controls are particularly necessary for the personal genomics field as they are more useful than those of individual genomes, providing the context that affords the research of trait heritability [1].

In this data article, we provide the complete genomics dataset of five blood-related family members that performed a series of direct-to-consumer (DTC) personal genomics tests. To our knowledge this is the first dataset released to the public for a whole family using only DTC means. The genotype data were obtained from 23andMe while all the sequencing data from the BGI. The dataset is summarised in *Table 1*. We call members of this family Mother, Father, Aunt, Daughter and Son. The data presented here include both raw and processed data. The rationale for including raw data, even though most users will not require them, is that current processing tools (e.g. genome aligners; variant call software) may provide different results depending on the type of configuration used. Whenever a type of data can be deduced from another, we have left the former out; e.g. bam indexes can be deduced from bam files so we have not included bam indexes. Also in the case of bam files, files belonging to the same sample have been merged into one. Due to these data having been obtained using DTC means, sometimes the source or platform utilised for the same test may be slightly different between individuals because of a different version of the platform used (indicated in Table 1). We have minimised the impact of platform versions by using the same pipelines and methods for alignment and variant calling.

Type	Provider	Source	Platform	Format	Year Obtained	# Files	Individual(s)	Approx. Size (MB)
Genotype	23andMe	Saliva	SNP chip v2	bed.zip	2009	1	Son	5
Genotype	23andMe	Saliva	SNP chip v3	bed.zip	2011	1	Mother	8
Genotype	23andMe	Saliva	SNP chip v3	bed.zip	2011	1	Father	8
Genotype	23andMe	Saliva	SNP chip v3	bed.zip	2011	1	Daughter	8
Genotype	23andMe	Saliva	SNP chip v3	bed.zip	2011	1	Aunt	8
Annotation	SNPedia	SNP chip v2	Phenotypes	txt	2012	1	Son	0.3
Annotation	SNPedia	SNP chip v3	Phenotypes	txt	2012	1	Mother	0.4
Annotation	SNPedia	SNP chip v3	Phenotypes	txt	2012	1	Father	0.4
Annotation	SNPedia	SNP chip v3	Phenotypes	txt	2012	1	Daughter	0.4
Annotation	SNPedia	SNP chip v3	Phenotypes	txt	2012	1	Aunt	0.4
Whole Exome Sequencing	BGI	Saliva	SE Illumina HiSeq 2000	fastq.gz	2011	4	Son	2400
Whole Exome Sequencing	BGI	Saliva	PE Illumina HiSeq 2000	fastq.gz	2013	2	Mother	2000
Whole Exome Sequencing	BGI	Saliva	PE Illumina HiSeq 2000	fastq.gz	2013	2	Father	2000
Whole Exome Sequencing	BGI	Saliva	PE Illumina HiSeq 2000	fastq.gz	2013	2	Sister	2400
Alignment	InSilicoDB	SE Illumina HiSeq 2000	BWA	bam	2013	1	Son	2900
Alignment	InSilicoDB	PE Illumina HiSeq 2000	BWA	bam	2013	1	Mother	1800
Alignment	InSilicoDB	PE Illumina HiSeq 2000	BWA	bam	2013	1	Father	1600
Alignment	InSilicoDB	PE Illumina HiSeq 2000	BWA	bam	2013	1	Daughter	2200
Annotation	InSilicoDB	Bam from Son, Mother, Father, Daughter	GATK	VCF	2013	1	Son, Mother, Father, Daughter	124.6
Metagenomics	BGI	Fecal frozen sample	PE Illumina HiSeq 2000	fastq.gz	2013	2	Son	1000
TOTAL						27		18463.5

Table 1: Summary of the data presented in this article, belonging to a family of 5 individuals from Southern Europe. The dataset includes a total of 27 files and 18,463 GB of data.

Data available encompass genotypes, annotations, alignments, whole exome sequencing and metagenomics sequencing. Processed data files containing predicted phenotypes, bam files and VCF files [2] are based on the raw data provided (SNP chip, Illumina HiSeq). Although data describing observed phenotypic traits are not included at present, our intention is to provide them in the near future. All variations are SNP-based and no Copy Number Variation (CNV) annotations are provided at this point. Whenever possible, data have been kept compressed to economise data storage. Years in which the data were obtained have also been included to roughly show the timeline of family tests. The five 23andMe SNP chip genotypes are in bed format, mapped to the human reference NCBI36. These datasets were taken in the summer of 2009 (Son; version 2 of 23andMe; ~0.5M SNPs) and January 2011 (Mother, Father, Daughter, Aunt; 23andMe SNP chip version 3, ~1M SNPs per individual). Whole exome sequencing from Son, Mother, Father, Sister was done from saliva samples using Illumina HiSeq 2000 with an expected coverage of ~30x. The Son's exome was carried out first (January

2012), using the Agilent 44M Single End (SE) capture chip. The other three exomes (Mother, Father, Daughter) were sequenced in January 2013 using the Agilent 44M Paired End (PE) capture chip each. The metagenomics sequencing has been performed for Son in 2013 from a fecal sample using Illumina shotgun sequencing. The processing of phenotypes was performed using Promethease, a tool that takes SNPedia [3] genotype-phenotype associations. Promethease requires 23andMe bed files as input and outputs text files with SNP ids and their annotation. Processing of exome data was carried out with InSilicoDB [4] a web-based storage hub. For all of the exomes, InSilicoDB produced bam files with BWA [5]. Once bam files were created, they were merged into one per individual. InSilicoDB generated variant calls using the Genome Analysis Toolkit (GATK) 'best practice variant detection method' pipeline [6]. Variant discovery was performed using joint variant calling, outputting one VCF file for all four processed exomes. All pipelines and data analyses in InSilicoDB are available at <https://insilicodb.org/app/browse/?q=ISDB11122>. All raw data are available in figShare at <http://figshare.com/articles/Corpasome/693052> from which they can be cited.

Availability and requirements

Project name: Corpasome

Project home page: <http://figshare.com/articles/Corpasome/693052>

Operating system: Platform independent

License: Creative Commons 0

Any restrictions to use by non-academics: None

Availability of supporting data

Processed data are available at

<https://insilicodb.org/app/browse/?q=ISDB11122>. Raw data are available at <http://figshare.com/articles/Corpasome/693052>.

Non-financial competing interest

The individuals of this family are the parents, sister and aunt of Manuel Corpas. Individuals from the whole family have provided their informed consent to make these data freely accessible.

Acknowledgements

We are grateful to Fiona Nielsen for useful comments on the manuscript.

References

1. Glusman G, Cariaso M, Jimenez R, Swan D, Greshake B, Bhak J, Logan DW, Corpas M: **Low budget analysis of Direct-To-Consumer genomic testing familial data.** *F1000Res* 2012.

2. Danecek P, Auton A, Abecasis G, Albers CA, Banks E, DePristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, McVean G, Durbin R, 1000 Genomes Project Analysis Group: **The variant call format and VCFtools.** *Bioinformatics* 2011, **27**:2156–2158.
3. Cariaso M, Lennon G: **SNPedia: a wiki supporting personal genome annotation, interpretation and analysis.** *Nucleic Acids Research* 2012, **40**:D1308–12.
4. Coletta A, Molter C, Duqué R, Steenhoff D, Taminau J, de Schaetzen V, Meganck S, Lazar C, Venet D, Detours V, Nowé A, Bersini H, Weiss Solís DY: **InSilico DB genomic datasets hub: an efficient starting point for analyzing genome-wide studies in GenePattern, Integrative Genomics Viewer, and R/Bioconductor.** *Genome Biol.* 2012, **13**:R104.
5. Li H, Durbin R: **Fast and accurate short read alignment with Burrows-Wheeler transform.** *Bioinformatics* 2009, **25**:1754–1760.
6. DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, Philippakis AA, del Angel G, Rivas MA, Hanna M, McKenna A, Fennell TJ, Kernytsky AM, Sivachenko AY, Cibulskis K, Gabriel SB, Altshuler D, Daly MJ: **A framework for variation discovery and genotyping using next-generation DNA sequencing data.** *Nat. Genet.* 2011, **43**:491–498.