HTSeq – A Python framework to work with high-throughput sequencing data
Simon Anders, Paul Theodor Pyl and Wolfgang Huber
Genome Biology Unit, European Molecular Biology Laboratory, 69111 Heidelberg, Germany
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ABSTRACT
Motivation: A large choice of tools exists for many standard tasks in the analysis of high-throughput sequencing (HTS) data. However, once a project deviates from standard work flows, custom scripts are needed.

Results: We present HTSeq, a Python library to facilitate the rapid development of such scripts. HTSeq offers parsers for many common data formats in HTS projects, as well as classes to represent data such as genomic coordinates, sequences, sequencing reads, alignments, gene model information, variant calls, and provides data structures that allow for querying via genomic coordinates. We also present htsq-count, a tool developed with HTSeq that preprocesses RNA-Seq data for differential expression analysis by counting the overlap of reads with genes.

Availability: HTSeq is released as open-source software under the GNU General Public Licence and available from http://www-huber.embli.de/HTSeq or from the Python Package Index https://pypi.python.org/pypi/HTSeq.

Contact: sander@fs.fum.de

1 INTRODUCTION
The rapid technological advance in high-throughput sequencing (HTS) has led to the development of many new kinds of assays, each of which requires the development of a suitable bioinformatical analysis pipeline. For the recurring “big tasks” in a typical pipeline, such as alignment and assembly, the bioinformatics practitioner can choose from a range of standard tools. For more specialised tasks, and in order to interface between existing tools, customised scripts often need to be written.

Here we present HTSeq, a Python library to facilitate the rapid development of scripts for processing and analysis of high-throughput sequencing (HTS) data. HTSeq includes parsers for common file formats for a variety of types of input data and is suitable as a general platform for a diverse range of tasks. A core component of HTSeq is a container class that simplifies working with data associated with genomic coordinates, i.e., values attributed to genomic positions (e.g., read coverage) or to genomic intervals (e.g., genomic features such as exons or genes). Two stand-alone applications developed with HTSeq are distributed with the package, namely htsq-qa for read quality assessment and htsq-count for preprocessing RNA-Seq alignments for differential expression calling.

Most of the features described in the following sections have been available since the initial release of the HTSeq package in 2010. Since then, the package and especially the htsq-count script have found considerable use in the research community. The present article provides a description of the package and also reports on recent improvements.

HTSeq comes with extensive documentation, including a tutorial that demonstrates the use of the core classes of HTSeq and discusses several important use cases in detail. The documentation, as well as HTSeq’s design, is geared towards allowing users with only moderate Python knowledge to create their own scripts, while shielding more advanced internals from the user.

2 COMPONENTS AND DESIGN OF HTSEQ
2.1 Parser and record objects
HTSeq provides parsers for reference sequences (FASTA), short reads (FASTQ), short-read alignments (the SAM/BAM format and some legacy formats), and for genomic feature, annotation and score data (GFF/GTF, VCF, BED, Wiggle).

Each parser is provided as a class whose objects are tied to a file name or open file or stream and work as iterator generators, i.e., they may be used in the head of a for loop and will yield a sequence of record objects that are taken up by the loop variable. These record objects are instances of suitable classes to represent the data records. Where appropriate, different parsers will yield the same type of record objects. For example, the record class SequenceWithQualities is used whenever sequencing read with base-call qualities needs to be presented, and hence yielded by the FastqParser class and also present as a field in the SAM_Alignment objects yielded by SAM_Reader or BAM_Reader parser objects (Figure 1). Specific classes (GenomicPosition and GenomicInterval) are used to represent genomic coordinates or intervals, and these are guaranteed to always follow a fixed convention (namely, following Python conventions, zero-based, with intervals being half-open) and parser classes take care to apply appropriate conversion when the input format uses different convention. The reverse is true for functions to write files.

To offer good performance, large parts of HTSeq, are written in Cython (Behnel et al., 2011), a tool to translate Python code augmented with type information to C. While the code for reading and writing all text-based formats, including text SAM files, is written in Python/Cython and hence has no external dependencies, the classes BAM_Reader and BAM_Writer wrap around functionality from PySam (Heger et al.) and are available only if that package has been installed.

The SAM_Alignment class offers functionality to facilitate dealing with complex, e.g. gapped, alignments (Figure 1b), with multiple alignments and with paired-end data. The latter is challenging...
3 DOCUMENTATION AND CODING STRATEGIES

HTSeq comes with extensive documentation to guide developers. Care has been taken to expect only moderate experience with Python from the reader. A “Tour” offers an overview over the classes and principles of HTSeq by demonstrating their use in simple examples. Then, two common use cases are discussed in detail to show how HTSeq can be applied to complex tasks.

The first use case is that of aggregate coverage profiles: Given ChiP-seq data, e.g. from histone marks, we want to calculate the profile of these marks with respect to specific positions in the genome, such as transcription start sites (TSSs), by aligning coverage data in windows centred on the TSSs and averaging over the TSSs of all genes or a subset thereof. In this use case, one needs to integrate information from two position-specific data sources, namely a list of TSSs obtained from annotation data and the aligned reads. Hence, one may either iterate through the reads first, store this information in a GenomicArray and then use position-specific access to it when iterating through the list of TSSs, or, first store the TSSs in a GenomicArray and use this afterwards when iterating through the reads. In either case, one data set is kept in memory in a form allowing for fast random access, while the other is iterated through with only summary information being kept. These approaches are prototypical for scripts built on HTSeq and hence explained and demonstrated in detail in the documentation (Section “A detailed use case: TSS plots”).

The second use case discussed in detail is that of counting for each gene in a genome how many RNA-Seq reads map to it. In this context, the HTSeq class CigarOperation is demonstrated, which represents complex alignments in a convenient form (Figure 1b). This section of the documentation also explains HTSeq’s facilities to handle multiple alignments and paired-end data.
The remainder of the documentation provides references for all classes and functions provided by HTSeq, including those classes not used in the highlighted use cases of the tutorial part, such as the facilities to deal with variant-call format (VCF) files.

4 HTSEQ-COUNT

We distribute two stand-alone scripts with HTSeq, which can be used from the shell command line, without any Python knowledge, and also illustrate potential applications of the HTSeq framework.

The script htseq-qa is a simple tool for initial quality assessment of sequencing runs. It produces plots which summarise the nucleotide compositions of the positions in the read and the base-call qualities.

The script htseq-count is a tool for RNA-Seq data analysis: Given a SAM/BAM file and a GTF or GFF file with gene models, it counts for each gene how many aligned reads overlap its exons. These counts can then be used for gene-level differential expression analyses using methods such as DESeq2 (Love et al., 2014) or edgeR (Robinson et al., 2010). As the script is specified particularly for differential expression analysis, only reads mapping unambiguously to a single gene are counted while reads aligned to multiple positions or overlapping with more than one gene are discarded. To see why this is desirable, consider two genes with some sequence similarity, one of which is differentially expressed while the other one is not. A read that maps to both genes equally well should be discarded, because if it were counted for both genes, the extra reads from the differentially expressed gene may cause the other gene to be wrongly called differentially expressed, too. Another design choice made with the downstream application of differential expression testing in mind is to count fragments, not reads, in case of paired-end data. This is because the two mates originating from the same fragment provide only evidence for one cDNA fragment and should hence be counted only once.

As the htseq-count script has found widespread use over the last three years, we note that we recently replaced it with an overhauled version, which now allows processing paired-end data without the need to sort the SAM/BAM file by read name first. See the documentation for a list of all changes to the original version.

5 DISCUSSION

HTSeq aims to fill the gap between performant but monolithic tools optimised for specialised tasks and the need to write data processing code for HTS application entirely from scratch. For a number of the smaller tasks covered by HTSeq, good stand-alone solutions exist, e.g. FastQC (Andrews et al., 2011) for quality assessment or Trimmomatic (Bolger et al., 2014) for trimming of reads. If the specific approaches chosen by the developers of these tools are suitable for a user’s application, they are easier to use. However, the need to write customised code will inevitably arise in many projects, and then, HTSeq aims to offer advantages over more narrow programming libraries that focus on specific file formats, e.g. Pysam (Heger et al.) and Picard (Wysoker et al.) for SAM/BAM files, by integrating parsers for many common file formats and fixing conventions for data interchange between them. For R developers, similar functionality is now available within the Bioconductor project (Gentleman et al., 2004) with packages like Rsamtools (Morgan et al.) and GenomicRanges (Lawrence et al., 2013). Within Python, HTSeq complements Biopython (Cock et al., 2009), which provides similar functionality for more “classic” bioinformatics tasks such as sequence analysis and phylogenetic analyses but offers little support for HTS tasks.

While most uses of HTSeq will be the development of custom scripts for one specific analysis task in one experiment, it can also be useful for writing more general tools. The htseq-count script, for example, prepares a count table for differential expression analysis, a seemingly easy task, which, however, becomes complicated when ambiguous cases have to be treated correctly. Despite being written in Python, htseq-count offers decent performance: Tested on a standard laptop computer, htseq-count (version 0.6.1) processed about 1.2 million reads (0.6M read pairs) per minute, using about 250 MB of RAM to hold the human gene annotation in memory. When the file was sorted by position rather than read-name, so that mate pairs were not in adjacent record, processing time increased to a bit less than twice as much, and, for a SAM file of 26 GB, less than 450 MB of additional space in RAM were needed for the buffer holding reads with outstanding mates.

When HTSeq was first released in 2010, htseq-count was the first comprehensive solution for this task, and has since then been widely used. Recently, further tools for this task have become available, including the summarizeOverlap function in the GenomicRanges Bioconductor package (Lawrence et al., 2013) and the stand-alone tool featureCount (Liao et al., 2014), which achieves very fast run times due to being implemented in C. In a recent benchmark, Fonseca et al. (2014) compared htseq-count with these other counting tools and judged the accuracy of htseq-count favourably. Nevertheless, neither htseq-count nor the other tools offer much flexibility to deal with special cases, which is why the HTSeq documentation (section “Counting reads”) discusses in detail how users can write their own scripts for this important use case.

Interval queries are a recurring task in HTS analysis problems, and several libraries now offer solutions for different programming languages, including BEDtools (Quinlan and Hall, 2010; Dale et al., 2011) and IRanges/GenomicRanges (Lawrence et al., 2013). Typically, these methods take two lists of intervals and report overlaps between them. HTSeq uses a different paradigm, namely that one list of intervals is read in and stored in a GenomicArrayOfSets object, and then the other intervals are queried one by one, in a loop. This explicit looping can be more intuitive; one example is the read counting problem discussed above, where split reads, gapped alignments, ambiguous mappings etc. cause much need for treatment of special cases that is addressed by branching statements within the inner loop.

In conclusion, HTSeq offers a comprehensive solution to facilitate a wide range of programming tasks in the context of high-throughput sequencing data analysis.

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REFERENCES


