

1 **Type 2 diabetes promotes cell centrosome amplification and the role**
2 **of AKT-ROS-dependent signalling of ROCK1 and 14-3-3 σ**

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19 **Running title:**

20 Centrosome amplification in type 2 Diabetes and its mechanisms

21

22

23 **Abstract**

24 Type2 diabetes is associated with oxidative stress which can cause cell centrosome
25 amplification. The study investigated centrosome amplification in type 2 diabetes and
26 the underlying mechanisms. We found that centrosome amplification was increased in
27 the peripheral blood mononuclear cells (PBMC) from the type 2 diabetic patients,
28 which correlated with the levels of fasting blood glucose and HbA1c. High glucose,
29 insulin and palmitic acid, alone or in combinations, induced ROS production and
30 centrosome amplification. Together, they increased AKT activation as well as the
31 expression, binding and centrosome translation of ROCK1 and 14-3-3 σ . Results from
32 further analyses showed that AKT-ROS-dependent upregulations of expression,
33 binding and centrosome translocation of ROCK1 and 14-3-3 σ was the molecular
34 pathway underlying the centrosome amplification induced by high glucose, insulin
35 and palmitic acid. Moreover, the increases in AKT activation and ROS production as
36 well as expression, binding and centrosome distribution of ROCK1 and 14-3-3 σ were
37 confirmed in the PBMC from the patients with type 2 diabetes. In conclusion, our
38 results show that type 2 diabetes promotes cell centrosome amplification, and suggest
39 that the diabetic pathophysiological factors-activated AKT-ROS-dependent signalling
40 of ROCK1 and 14-3-3 σ is the underlying molecular mechanism.

41

42 **Key words:** Centrosome amplification; Type 2 diabetes; AKT; ROS; ROCK1;
43 14-3-3 σ ; Pathophysiological factors.

44

45 INTRODUCTION

46 Type 2 diabetes is a common non-communicable disease. At certain stage of the
47 disease development, patients present pathophysiological features such as
48 hyperglycemia, hyperinsulinemia and increased plasma level of free fatty acids. At
49 latter stage, patients may develop insulin deficiency, but continue to have
50 hyperglycemia and increased plasma level of free fatty acids. These
51 pathophysiological factors are able to trigger the production of reactive oxygen
52 species (ROS) (Inoguchi et al., 2000; May and de Haen, 1979) and DNA damage (Lee
53 and Chan, 2015) *in vitro*, which may explain why oxidative stress and DNA damage
54 are increased *in vivo* in patients with type 2 diabetes (Lee and Chan, 2015; Orié et al.,
55 1999).

56
57 Cell centrosome amplification refers to a cell with more than two centrosomes. There
58 is evidence that oxidative stress and DNA damage can cause centrosome
59 amplification (Chae et al., 2005; Dodson et al., 2004). At the molecular level, many
60 signal mediators for cell centrosome amplification have been identified, which
61 include those inside or outside the centrosomes. Overexpression of PLK4 (Basto et al.,
62 2008) or aurora A (Castellanos et al., 2008), both are centrosome proteins, can cause
63 centrosome amplification. Wang and co-workers show that Chk2 is located in
64 centrosome, which controls centrosome amplification induced by sub-toxic
65 concentrations of hydroxyurea (Wang et al., 2015). Overexpression of cyclin E
66 (Bagheri-Yarmand et al., 2010) which can be in centrosome was also to promote
67 centrosome amplification. Ras (Zen et al., 2010) which is a protein outside the
68 centrosomes has been shown to cause centrosome amplification. Interestingly,
69 BRCA1 promotes centrosome amplification when is translocated to centrosome (Zou
70 et al., 2014), which shows a dynamic communication between centrosome and its
71 surrounding environment for centrosome amplification. At the cellular and organismal
72 levels, centrosome amplification regulates cell division, movement and intracellular
73 transport (Anderhub et al., 2012), and increases cancer invasion potential (Godinho et

74 al., 2014). While robust centrosome amplification results in multipolar division follow
75 by mitosis catastrophe (Pannu et al., 2012), there is evidence that moderate
76 centrosome amplification can induce genome instability (Ganem et al., 2009) and
77 tumorigenesis (Levine et al., 2017).

78

79 Since type 2 diabetes is associated with oxidative stress (Inoguchi et al., 2000; May
80 and de Haen, 1979; Lee and Chan, 2015; Orié et al., 1999) which can cause
81 centrosome amplification (Chae et al., 2005; Dodson et al., 2004), in the present study,
82 we investigate cell centrosome amplification in type 2 diabetes and the underlying
83 mechanisms.

84

85 **RESULTS**

86 **Cell centrosome amplification is increased in patients with type 2 diabetes, which** 87 **is in correlation with fasting blood glucose and HbA1c**

88 To investigate whether centrosome amplification is associated with type 2 diabetes,
89 first of all, we compared the extent of centrosome amplification in PBMCs from 32
90 patients with type 2 diabetes and 12 healthy subjects. The clinical characteristics of
91 the volunteers were summarized in Table 1. Compared with the healthy subjects, the
92 diabetic patients had higher body mass index, fasting blood glucose, HbA1c and
93 triglyceride. Moreover, we compared the level of centrosome amplification between
94 the healthy subject and diabetic patients. The images of centrosome and centrosome
95 amplification were shown in Fig. 1A. The patients had a greater degree of cell
96 centrosome amplification than the healthy subjects (8.7 % v.s.3.4%; $p<0.01$) (Fig. 1B);
97 there was a 2.6-fold increase in centrosome amplification in PBMCs from the patients.
98 When the diabetic patients were divided into two groups according to their fasting
99 blood glucose level less or greater than 10mM, there was a step-wise increase in
100 centrosome amplification ($p<0.01$; Fig. 1C). Indeed, in all the volunteers, correlation
101 analysis showed that the extent of cell centrosome amplification was correlated with
102 the fasting blood glucose level ($R^2 = 0.7$; $p<0.01$; Fig. 1D) and HbA1c ($R^2=0.7$;

103 p<0.01; Fig. 1E). Similarly, in the patients alone, the level of centrosome
104 amplification was also correlated with fasting blood glucose ($R^2 = 0.5$; p <0.01; Fig.
105 1F) and HbA1c ($R^2=0.4$; p<0.01; Fig. 1G).

106

107 Suggested Table 1 here

108 Suggested Figure 1 here

109

110 **High glucose, insulin and palmitic acid, alone or in combinations, induce**
111 **centrosome amplification via ROS production**

112 We then investigated whether the pathophysiological factors in type 2 diabetes could
113 induce centrosome amplification via specific molecular pathway using colon cancer
114 cells as an experimental model. Some results were verified in normal human breast
115 epithelial cells wherever it was considered to be meaningful, which was to verify
116 findings from experiments using cancer cells in non-cancerous cell model. Palmitic
117 acid, the most common saturated free fatty acid, was used to represent free fatty acids.
118 We tested ROS pathway to start with. Fluorescent spectrophotometry was used to
119 quantify the level of ROS when experiments involved more than three samples.
120 Otherwise, flow cytometry analysis was employed. The results showed that high
121 glucose, insulin and palmitic acid, alone or in combinations, significantly induced
122 ROS production (Fig. 2A) and centrosome amplification (Figs. 2C) in the cancer cells.
123 High glucose, insulin and palmitic acid individually induced centrosome amplification
124 in concentration-dependent manners (figures not shown). They, together, also
125 triggered ROS production (Fig. 2B) and centrosome amplification (Fig. 2D) in the
126 normal human breast epithelial cells. Antioxidant N-acetylcysteine (NAC) was able to
127 inhibit the ROS production and the centrosome amplification (Figs. 2A -2D). The
128 potential of high glucose, insulin and palmitic acid in inducing centrosome
129 amplification was three factors > two factors > single factor (all at p<0.05; Fig. 2C).

130

131 Suggested Figure 2 here

132

133 **Functional transcriptomic analysis identifies AKT, ROCK1 and 14-3-3 σ as signal**
134 **mediators for the centrosome amplification**

135 To further identify the molecular signals for the centrosome amplification, we
136 performed a functional transcriptomic analysis between the control samples and those
137 treated with high glucose, insulin and palmitic acid. The differentially expressed
138 genes were summarized in the supplementary data sheet (Table S1). In total, 729
139 genes were differentially expressed, with 508 upregulated and 221 downregulated.
140 Results of bioinformatic annotation were included in supplementary figures S1-S5.
141 Functional enrichment using GO analysis assigned the genes to 37 terms in three
142 categories of molecular component, molecular function and biological process (Fig.
143 S1). The top terms were molecular function, cellular component, biological process
144 and cell, which had 9%, 9%, 8.4% and 8.3% of the genes, respectively. KEGG
145 annotation identified 37 pathways which were related to different functional groups
146 (Fig. S2). The top pathways were signal transduction, infectious diseases, cancer,
147 immune system and endocrine system, which had 15.6%, 11.6%, 11.1%, 7.4% and 7.0%
148 of the genes, respectively. Notably, AKT (Fig.S3), ROCK (Fig. S4) and 14-3-3 σ
149 (Fig.S5) pathways were activated, which were chosen for further functional analysis.
150 Western blot analyses confirmed that high glucose, insulin and palmitic acid activated
151 AKT, which was inhibited by AKT inhibitor Ly294002 or siRNA (Fig. 3A). Similarly,
152 ROCK1 (Fig. 3B) and 14-3-3 σ (Fig. 3C) protein levels were upregulated, which were
153 inhibited their specific siRNA (Figs. 3B and 3C). We then performed functional
154 analyses to examine whether AKT, ROCK1 and 14-3-3 σ mediated the centrosome
155 amplification. Indeed, inhibition of AKT using chemical inhibitor or siRNA could
156 inhibit the centrosome amplification (Fig. 3D). Similarly, siRNA for ROCK1 (Fig. 3E)
157 and siRNA for 14-3-3 σ (Fig. 3F) were also able to inhibit the centrosome
158 amplification. Moreover, AKR chemical inhibitor, AKT siRNA, ROCK1 siRNA and
159 14-3-3 σ siRNA individually were all able to inhibit the centrosome in the normal
160 human breast epithelial cells (Figs. 3G and 3H).

161

162 Suggested Figure 3 here

163

164 **AKT-ROS-dependent signaling of ROCK1 and 14-3-3 σ is the molecular pathway**
165 **for the centrosome amplification**

166 We next delineated the signal transduction pathway for the centrosome amplification
167 in the colon cancer cells. The logic was that inhibition of an upstream signal would
168 inhibit downstream one(s), while inhibition of a downstream signal would not affect
169 the upstream one(s). We observed that inhibition of AKT using chemical inhibitor
170 (Fig. 4A) orsiRNA (Fig. 4B) was able to inhibit ROS production as well as the
171 upregulations of ROCK1 (Figs. 4C) and 14-3-3 σ (Fig. 4D). NAC was able to inhibit
172 the upregulation of ROCK1 (Fig. 4E) and 14-3-3 σ (Fig. 4F), but did not affect the
173 activation of AKT (figure not shown). ROCK1 or 14-3-3 σ siRNA did not affect AKT
174 activation or ROS production (figures not shown).

175

176 Suggested Figure 4 here

177

178 **High glucose, insulin and palmitic acid increase the binding between ROCK1**
179 **and 14-3-3 σ as well as their centrosome translocation**

180 Next, we investigated the relationships amongst ROCK1, 14-3-3 σ and centrosome in
181 the cancer cells by using ROCK1 and 14-3-3 σ antibodies to pull down their binding
182 partners, as well as to analyse the centrosome translocation of the two proteins. In
183 pull-down samples using ROCK1 antibody, 14-3-3 σ protein was detected, which was
184 increased in the cells treated with high glucose, insulin and palmitic acid (Fig. 5A).
185 Similarly, high glucose, insulin and palmitic acid increased the pull down ofROCK1
186 using 14-3-3 σ antibody (Fig. 5A). AKT inhibitor and NAC were able to inhibit the
187 increase in co-precipitation of ROCK1 and14-3-3 σ from the cells treated by high
188 glucose, insulin and palmitic acid (Figs. 5A). These observations on the Western blot
189 analysis images were confirmed when the images were quantified (Figs. 5B and 5C.
190 As shown in confocal images in Figs. 5D and 5E respectively, ROCK1 and 14-3-3 σ
191 were localized in the centrosome in all the treated samples. In the cells with more than
192 one centrosome, only half of the centrosomes had ROCK1, with another half of

193 centrosomes free of ROCK1 (Fig. 5D). The fluorescent intensity of ROCK1 staining,
194 which indicates the level of ROCK1 in the centrosomes, was higher in the treated
195 cells compared to that in the control cells, which was inhibited by AKT inhibitor or
196 NAC but not 14-3-3 σ siRNA (Fig. 5F). Comparing with that only 1% of the control
197 cells had 14-3-3 σ in the centrosomes, approximately 20% of the cells treated with
198 high glucose, insulin and palmitic acid had 14-3-3 σ in the centrosomes, which was
199 inhibited by AKT inhibitor, NAC or ROCK1 siRNA (all at $p < 0.01$; Figs. 5E and 5G).

200

201 Suggested Figure 5 here

202

203 **The increases in AKT activation and ROS production as well as expression,**
204 **binding and centrosome distribution of ROCK1 and 14-3-3 σ *in vitro* are verified**
205 **in the PBMC from the volunteers**

206 Finally, we investigated whether the molecular signalling events found *in vitro*
207 occurred *in vivo* in patients with type 2 diabetes. We compared these molecular events
208 between the PBMC from healthy volunteers and those from type 2 diabetic patients.
209 The results showed that PBMC from the diabetic patients had increased level of AKT
210 activation (Fig. 6A) and ROS production (Fig. 6B). The PBMC from the patients also
211 had increased expression (Fig. 6A), binding (Figs. 6C) and centrosome distribution of
212 ROCK1 and 14-3-3 σ (Figs. 6D and 6E).

213

214 Suggested Figure 6 here

215

216 **DISCUSSION**

217 Our results showed that patients with type 2 diabetes had 2.6-fold increase in cell
218 centrosome amplification *in vivo* in PBMC, which correlated with poor glycemic
219 control (Fig. 1). Pathophysiological factors in type 2 diabetes, i.e., high glucose,
220 insulin and palmitic acid, could trigger centrosome amplification (Figs. 2C and 3D).
221 AKT-ROS-dependent upregulation of expression, binding and centrosome

222 translocation of ROCK1 and 14-3-3 σ was the underlying molecular pathway (Figs.
223 3-6).

224

225 Type 2 diabetes increases the risk of developing all-site cancer, with the exception of
226 prostate cancer (Giovannucci et al., 2010). Cancer patients with type 2 diabetes have
227 poorer prognosis than those without diabetes (Mills et al., 2013). However, it remains
228 unknown why and how type 2 diabetes favours cancer development. Centrosome
229 amplification can initiate in animal model (Levine et al., 2017) and increase cancer
230 cell invasion potential (Godinho et al., 2014). In the present study, we showed that
231 patients with type 2 diabetes had increased level of cell centrosome amplification *in*
232 *vivo* (Fig. 1). Thus, we speculate whether there is a link between the
233 diabetes-associated centrosome amplification (Fig. 1) and the increased cancer risk in
234 type 2 diabetes.

235

236 Our results (Figs. 3-6) support the findings that AKT (Na et al., 2015) and ROS (Chae
237 et al., 2005) can mediate centrosome amplification, and further place AKT upstream
238 of ROS. In an apoptosis model, activation of ROCK1 causes AKT inactivation (Zhang
239 et al., 2016), which is different from our results that placed ROCK1 downstream of
240 AKT activation (Figs. 4C). Our result (Fig. 4E) showed that ROS was upstream of
241 ROCK1, which is in agreement with the observations by Shen and Wang (Shen and
242 Wang, 2015). Oh and Jang showed that AKT can be upstream of 14-3-3 σ (Oh and
243 Jang, 2009), which is supported by our result (Figs. 4D) that AKT was upstream of
244 14-3-3 σ .

245

246 As shown in Fig. 4, inhibition of AKR inhibits all other signals. Inhibition of ROS
247 inhibits ROCK1 and 14-3-3 σ but not AKT. Inhibition of ROCK1 or 14-3-3 σ did not
248 affect any other signals. This suggests that AKT-ROS-dependent signalling of
249 ROCK1 and 14-3-3 σ is the pathway underlying the centrosome amplification by the
250 experimental treatment. Our results (Figs. 5D and 5E) showed that ROCK1, but not
251 14-3-3 σ , was a centrosome protein. ROCK1 was present in half of the centrosomes

252 when a cell had two or more centrosomes. Whether it is present only in the mother
253 centrosomes remains to be clarified. In most cases, 14-3-3 σ was present in the
254 centrosomes only in the treated cells (Fig. 5E). Knockdown of ROCK1 inhibited
255 14-3-3 σ translocation to centrosome, while knockdown of 14-3-3 σ did not affect the
256 translocation of ROCK1 to the centrosomes (Figs. 5F and 5G), which suggests that
257 ROCK1 transports 14-3-3 σ to centrosome or is involved in 14-3-3 σ translocation to
258 centrosome after treatment. Downregulation of either ROCK1 or 14-3-3 σ inhibited
259 the centrosome amplification (Figs. 3E and 3F), which suggests that the integrity of
260 ROCK1 and 14-3-3 σ complex is required for the centrosome amplification. These
261 data suggest that the pathophysiological factors of type 2 diabetes activate AKT-ROS
262 signalling which promotes the expression and binding of ROCK1 and 14-3-3 σ as well
263 as their translocation to centrosome to promote centrosome amplification.

264

265 High glucose, insulin and palmitic acid, alone or in combinations, did not affect the
266 cell viability under the experimental conditions, with the exception that high glucose
267 treatment increased cell viability (data not shown). In some cell line models,
268 centrosome amplification is associated with cell cycle arrest (Yih et al., 2006). In our
269 study, high glucose, insulin and palmitic acid, alone or in combination, did not disturb
270 the cell cycle (data not shown), which suggests that cell cycle arrest is not a
271 prerequisite for centrosome amplification under our experimental conditions. At the
272 concentrations of 10, 20 and 30 μ M, linoleic acid did not significantly affect the
273 centrosome amplification (figure not shown), suggesting that unsaturated free fatty
274 acids are unable to suppress the centrosome amplification.

275

276 In our study, 3.4% of the PBMC from the healthy subjects displayed centrosome
277 amplification (Fig. 1B), which agrees with the finding by Dementyeva and
278 do-workers that approximately 3% of the peripheral blood cells from healthy donors
279 display centrosome amplification (Dementyeva et al., 2010).

280

281 It is known that obesity increases the risk for cancer (Mazzarella, 2005). There is also

282 evidence that type 1 diabetes modestly increases the risk for cancer (Zendehdel et al.,
283 2003). Obesity is associated with increased plasma levels of insulin and free fatty
284 acids (Golay et al., 1986). Type 1 diabetes is associated with hyperglycaemia and
285 increased free fatty acid level. All these pathophysiological factors were able to cause
286 centrosome amplification (Figs. 2C-2D). Thus, we speculate whether centrosome
287 amplification could play a role the cancer development in obesity and type 1 diabetes.
288 Moreover, our results (Figs. 3B and 3C) showed that the diabetic pathophysiological
289 factors could upregulate ROCK1 and 14-3-3 σ which are associated with
290 neurodegeneration (Joo et al., 2015; Hu et al., 2016), upon which we further speculate
291 whether ROCK1 and/or 14-3-3 σ could be new clues for the development of
292 Alzheimer's disease which shows an increased risk in type 2 diabetes (Zhang et al.,
293 2016).

294

295 The study involved limited number of subjects. Further analysis with large cohorts is
296 required to confirm the findings. In the study, we used PBMC to demonstrate the
297 increase in centrosome amplification in type 2 diabetes. This raises a concern of
298 whether centrosome amplification occurs in other tissues prone to diabetes-related
299 cancer.

300

301 In conclusion, our results show that type 2 diabetes promotes cell centrosome
302 amplification, and suggest that activation of AKT-ROS-dependent upregulations in
303 expression, binding and centrosome translocation of ROCK1 and 14-3-3 σ by the
304 pathophysiological factors in type 2 diabetes is the underlying molecular mechanism.

305

306 **MATERIALS AND METHODS**

307 **Chemicals, antibodies and cells**

308 All chemicals were purchased from Sigma (St. Louis, MO, USA). Gama-tubulin
309 antibody (No. ab27074; mouse antibody) was purchased from Abcam (Cambridge,
310 UK). Rock1 antibody (No. 4035; rabbit antibody) was provided by cell signaling
311 technology (Boston, MA, USA). 14-3-3 σ antibody (No. PLA0201; rabbit antibody)
312 was purchased from Sigma (St. Louis, MO, USA). Other antibodies were provided by

313 Cell Signalling Technology (Boston, MA, USA). HCT116 colon cancer cells were
314 kindly provided by Dr. B. Vogelstein of the Johns Hopkins University School of
315 Medicine, who produced the cell line. Normal human breast epithelial cells
316 (PCS-600-010) and culture medium were purchased from the American Type Culture
317 Collection (ATCC, Manassas, VA, USA). The culture medium and reagents for the
318 colon cancer cells were purchased from Gibco (Beijing, China). The palmitic acid
319 stock was conjugated to fatty acid-free bovine albumin in a 3:1 molar ratio at 37 °C
320 for 1 hour before use. Anti-gamma tubulin antibody was used to detect centrosome by
321 innumofluorescent staining. ROCK1 siRNA treatment largely reduced the staining of
322 ROCK1. 14-3-3 was seen in the centrosome only after experimental treatment. These
323 observations showed that non-specific staining by the antibodies would not affect the
324 experimental observations.

325

326 **Clinical study**

327 Institutional approval and written informed consent were obtained from the medical
328 ethics committee and all participating subjects, respectively. All the volunteers were
329 consecutively recruited during 2014 and 2016 at the Shanxi Hospital of Integrative
330 Western and Chinese Medicine and Shanxi Medical University without any selection
331 bias. Type 2 diabetes was diagnosed according to the 1999 WHO criteria. All healthy
332 subjects were free hypertension. Each volunteer donated 5-ml blood sample. All the
333 clinical data were collected by clinical doctors responsible for the clinical study.

334

335 **Cell culture**

336 HCT116 cells were maintained in DMEM (low glucose, 5mM) supplemented with 50
337 U/ml penicillin, 50 g/ml streptomycin and 10% (v/v) foetal calf serum. Human
338 primary breast epithelial cells were cultured in basal medium (PCS-600-030) with a
339 growth kit (PCS-600-040, ATCC, Manassas, USA) according to ATCC instructions.
340 Epithelial cells from the second passage in our lab were used in the study. In cell
341 model studies, cells treated for 12 hours were used for ROS quantification. Cells
342 treated for 24 hours were harvested for co-immunoprecipitation assay. Those treated
343 for 30 hours were used for transcriptomic analysis. Cells treated for 48 hours were
344 used for quantification of centrosome number and protein distribution in centrosomes.
345 We performed time course assays and the time points were chosen, since these time
346 points produced the highest level of differences for the measurements between the

347 control and the treated samples.

348

349 **Quantification of ROS**

350 Changes in the intracellular ROS levels were determined using the
351 2,7'-dichlorofluorescein (DCFH-DA, Beyotime, Shanghai, China) probe on a
352 spectrofluorometer (SpectraMax M5, Molecular Devices, Silicon Valley USA). Cells
353 grown in a 24-well plate were treated, washed twice with PBS, and incubated with
354 DCFH-DA for 20 min at 37 °C. Then, the cells were harvested, washed once in PBS,
355 and transferred to a 96-well plate. Optical density values were obtained at an
356 excitation wavelength of 488 nm and an emission wavelength of 525 nm.

357 Alternatively, the ROS levels were quantified using a flow cytometer (FACSCalibur,
358 BD, New Jersey, USA) according to the manufacturer's instructions.

359

360 **Confocal microscopy**

361 A cover slip was placed in a well of a 6-well plate. Cells were plated at a density of
362 50,000 cells per well. Cells grown on the cover slips were fixed in cold methanol and
363 acetone (1:1; v:v) for 6 min at -20°C, followed by three washes with PBS (10 min
364 each time). Then, the cells were incubated with 0.1% Triton X-100 for 15 minutes and
365 3% BSA for 1 hour. The cells were incubated with a primary antibody in 3% BSA in
366 PBS overnight at 4 °C, washed twice with PBS, and incubated with a
367 FITC-conjugated secondary antibody in 3% BSA in 1×PBS for 1 hour at room
368 temperature in the dark. Finally, the cells were mounted with mounting medium.

369 Confocal microscopy was performed using the Zeiss LSM880 microscope
370 (Oberkochen, Germany) with a 1.4 NA oil-immersion lens, and image processing was
371 performed with Zen software (Oberkochen, Germany). One hundred cells were
372 counted for their centrosome numbers for the percentage of centrosome amplification.

373

374 To separate peripheral blood mononuclear cells (PBMC), we loaded a 5-ml blood
375 sample on the top of separation medium (LTS1077; TBD, Tianjin, China) in a
376 centrifuge tube and centrifuged at 250 g for 20 min. The cells on the top were
377 collected, washed twice in PBS, smeared onto a cover slip, and air dried for 24 hours
378 at room temperature.

379

380

381 **Transcriptomic profiling and bioinformatic annotation**

382 The cells were harvested after treatment and total RNA was extracted. Two cDNA
383 libraries (control or treatment with high glucose, insulin, and palmitic acid) were
384 constructed using the Illumina TruSeq RNA Sample Preparation Kit (Illumina, USA)
385 according to the manufacturer's instructions. After several steps of purification,
386 adapter addition, and cDNA length selection, the two libraries were sequenced
387 independently using an Illumina HiSeq™500 platform (Shanghai Personal
388 Biotechnology Co., Shanghai, China). Pathway assignments were generated using GO
389 (geneontology.org) and KEGG databases (www.kegg.jp).

390

391 **Western blot analysis**

392 The cells were lysed in RIPA buffer. Proteins were separated by polyacrylamide gel
393 electrophoresis and transferred onto PVDF membrane. After blocking for 1 hour at
394 room temperature with TBST containing 0.05% (v/v), Tween-20 and 5% (w/v)
395 non-fat milk, the membranes were incubated with primary antibodies overnight at
396 4 °C, followed by washes with TBST containing 0.05% Tween-20. The membranes
397 were then incubated with a horseradish peroxidase-conjugated secondary antibody for
398 1 hour at room temperature. ECL reagents (Thermo Biosciences, Massachusetts, USA)
399 were used to visualize the protein bands which were captured on X-ray film.

400

401 **Co-immunoprecipitation**

402 Cells were harvested under non-denaturing conditions, washed by ice-cold PBS for 3
403 times, lysed in 0.5 ml ice-cold cell lysis buffer and centrifuged. Supernatant was
404 collected to a new tube and incubated with 20 ul Protein G Plus /Protein A agarose
405 (Miliproe, IP05 USA) with gentle shaking for 2 hours at 4 oC. Protein G Plus /Protein
406 A agarose was then removed by centrifuge for 10 min at 4 oC, and supernatant was
407 incubated with primary antibody overnight at 4 oC with gentle shaking. After that,
408 30ul Protein G Plus /Protein A agarose were added and incubated under gentle
409 shaking for 4 hours at 4 oC. Finally Protein G Plus /Protein A agarose was collected
410 by centrifuge.

411

412 **Knockdown of protein level**

413 The pre-designed siRNA oligonucleotides (Songon Technology, Shanghai, China) for
414 Akt1, ROCK1 and 14-3-3σ were:

415 GAGUUUGAGUACCUGAAGCUGUU (sense) and
416 AACAGCUUCAGGUACUCAAACUC (antisense);
417 UGAUCUUGUAGCUCCCGCAUGUGUC (sense) and
418 ACUAGAACAUCGAGGGCGUACACAG (antisense); and
419 ACCTGCTCTCAGTAGCCTA (sense) and TAGGCTACTGAGAGCAGGT
420 (antisense), respectively. HCT116 cells (5×10^4 cells per well) were seeded in 6-well
421 plates and cultured for 24 hours, and then were transfected with 200 pM siRNA
422 oligonucleotides using Lipofectamine 2000 transfection reagent (Invitrogen,
423 California, USA), according to the manufacturer's instructions. The total AKT protein
424 level was evaluated by Western blot analysis 24 hours after transfection.

425

426 **Statistical analysis**

427 All the experiments including the transcriptomic profiling were performed in triplicate.
428 The data are expressed as the mean \pm SD. Student's t-test was performed to compare
429 the difference between two groups. Multi-group comparisons were performed using
430 one-way ANOVA analysis. Linear regression analysis was performed for correlations.
431 The statistical analysis software package SPSS21 was employed for the statistical
432 comparisons. A p value < 0.05 was considered statistically significant.

433

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436

437 **Competing interests**

438 The authors declare that there is no conflict of interest.

439

440 **Author contribution**

441 P Wang and YC Lu performed most of the experiments. J Wang and YF LI
442 contributed the clinical studies. L Wang provided the technologies. H Yu was involved
443 in grant applications and the study design. A Kong and J Chan contributed the ROS
444 data set and were involved in designing the study. SC Lee was the principal
445 investigator, who was in charge the whole project, from grant application and study
446 design to manuscript preparation.

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553

554 **Figure Legends**

555

556 **Fig. 1.**

557 **Cell centrosome amplification is increased in patients with type 2 diabetes, which**
558 **is in correlation with fasting blood glucose and HbA1c.** (A): image of centrosome
559 amplification; (B): cell centrosome amplification is increased in patients with type 2
560 diabetes; (C): evidence that centrosome amplification correlates with the fasting blood
561 glucose; (D) and (E): correlation analysis shows that the centrosome amplification
562 correlates with the fasting blood glucose and HbA1c in all the volunteers; (F) and (G):
563 in the diabetic patients alone, centrosome amplification also correlates with fasting
564 blood glucose and HbA1c. Student t-test was used to compare the means between two
565 groups. One way ANOVA analysis was used to compare multiple groups. Linear
566 regression was performed for correlations. *: $p < 0.05$; **: $p < 0.01$, compared with

567 that in the control group.

568

569 **Fig. 2.**

570 **High glucose, insulin and palmitic acid, alone or in combinations, induce**
571 **centrosome amplification via ROS production. (A):** high glucose, insulin and
572 palmitic acid, alone or in combinations, triggers ROS production in the cancer cells;
573 **(B):** the pathophysiological factors, together, trigger ROS production in normal
574 human breast epithelial cell; **(C):** high glucose, insulin and palmitic acid, alone or in
575 combinations, can promote centrosome amplification in the cancer cells; **(D):** high
576 glucose, insulin and palmitic acid can trigger centrosome amplification in the normal
577 human breast epithelial cells. **(A)-(D):** NAC inhibits the ROS production and the
578 centrosome amplification. Glu: glucose, 15 mM; Ins: insulin, 5 nM; Pal: palmitic acid,
579 150 μ M; NAC: 3 mM. One way ANOVA analysis was used to compare multiple
580 groups. *: $p < 0.05$; **: $p < 0.01$.

581

582 **Fig. 3.**

583 **Functional transcriptomic analysis identifies AKT, ROCK1 and 14-3-3 σ as signal**
584 **mediators for the centrosome amplification. (A), (B) and (C):**the diabetic
585 pathophysiological factors activate of AKT, ROCK1 and 14-3-3 σ , respectively; **(D):**
586 inhibition of AKT using chemical inhibitor or siRNA blocks the centrosome
587 amplification in the colon cancer cells; **(E):** ROCK1 siRNA inhibits the centrosome
588 amplification; **(F):** 14-3-3 σ siRNA inhibits the centrosome amplification in normal
589 human breast epithelial cells. **(G):** AKT inhibitor inhibits centrosome amplification;
590 **(H):** AKT siRNA, ROCK1 siRNA and 14-3-3 σ siRNA inhibit centrosome
591 amplification in normal human breast epithelial cells. One way ANOVA analysis was
592 used to compare multiple groups. **: $p < 0.01$, compared with that in the control group;
593 #: $p < 0.01$, compared with that in the samples treated with Glu, Ins and Pal. Glu:
594 glucose, 15 mM; Ins: insulin, 5 nM; Pal: palmitic acid, 150 μ M; Ly294002: 30 μ M.

595

596

597 **Fig. 4.**

598 **AKT-ROS-dependent signaling of ROCK1 and 14-3-3 σ is the molecular pathway**
599 **for the centrosome amplification. (A) and (B):** AKT chemical inhibitor and siRNA
600 inhibit ROS production, respectively; (C): AKT chemical inhibitor or siRNA inhibits
601 ROCK1 upregulation; (D): AKT chemical inhibitor or SiRNA inhibits 14-3-3 σ
602 upregulation; (E) and (F): NAC inhibits the upregulation of ROCK1 and 14-3-3 σ ,
603 respectively. Glu: glucose, 15 mM; Ins: insulin, 5 nM; Pal: palmitic acid, 150 μ M;
604 Ly294002, 30 μ M; NAC: 3 mM.

605

606 **Fig. 5.**

607 **High glucose, insulin and palmitic acid increase the binding between ROCK1**
608 **and 14-3-3 σ as well as their centrosome translocation. (A):** use of ROCK1 or
609 14-3-3 σ antibodies to pull down their binding partner, and the influences of Ly294002
610 and NAC; (B) and (C): quantification of the gel images of Western blot analysis
611 shown in Fig. 5A; (D): confocal image of ROCK1 localization in the centrosomes;
612 (E): confocal image of 14-3-3 σ localization in the centrosomes; (F): high glucose,
613 insulin and palmitic acid increases level of ROCK1 localization in the centrosomes,
614 which is inhibited by AKT inhibitor and NAC but not 14-3-3 σ siRNA; (G):
615 experimental treatment increases the localization of 14-3-3 σ to the centrosomes,
616 which is inhibited by AKT inhibitor, NAC or ROCK1 siRNA. One way ANOVA
617 analysis was used to compare multiple groups. **: $p < 0.01$, compared with that in the
618 control group; #: $p < 0.01$, compared with that in the samples treated with Glu, Ins
619 and Pal. Glu: glucose, 15 mM; Ins: insulin, 5 nM; Pal: palmitic acid, 150 μ M;
620 Ly294002, 30 μ M; NAC, 3 mM.

621

622 **Fig. 6.**

623 **The increases in AKT activation and ROS production as well as expression,**
624 **binding and centrosome distribution of ROCK1 and 14-3-3 σ are confirmed in**
625 **the PBMC from the patients with type 2 diabetes. (A):** AKT activation as well as
626 expressions of ROCK1 and 14-3-3 σ are increased in PBMC from the patients; (B):

627 ROS production is increased in the PBMC from the patients; **(C)**: binding between
628 ROCK1 and 14-3-3 σ is increased in PBMC from the diabetic patients; **(D)** and **(E)**:
629 centrosome distributions of ROCK1 and 14-3-3 σ are increased in the PBMC from the
630 type 2 diabetic patients, respectively. For Western blot analyses and
631 co-immunoprecipitation, five clinical samples were randomly selected and combined
632 for the experiments. ROS quantification was performed in triplicate using different
633 samples. One way ANOVA analysis was used to compare multiple groups. **: p <
634 0.01.

Table 1. Clinical and biochemical characteristics of the volunteers

	Healthy subject	Diabetic patient
n	12	32
Age (years)	57±2	55±2
Sex ratio (male:female)	1:1	1:1
Body mass index (kg/m ²)	19.8±2.7	26.7±5*
Systolic blood pressure (mmHg)	133.3±5.7	131.7±17.2
Diastolic blood pressure (mmHg)	76.6±11.5	83.3±10.3
Fasting blood glucose (mmol/l)	5.3±0.3	10.1±3.7*
HbA1c (%)	5.6±	8.5±2.2*
Total cholesterol (mmol/l)	2.8±1.4	4.9±1.4
Triglyceride (mmol/l)	0.66(0.2-1.1)	2.3(1.2-3.5)*

Data are means ± SD, or medians (range). *p<0.05

Fig. 1A

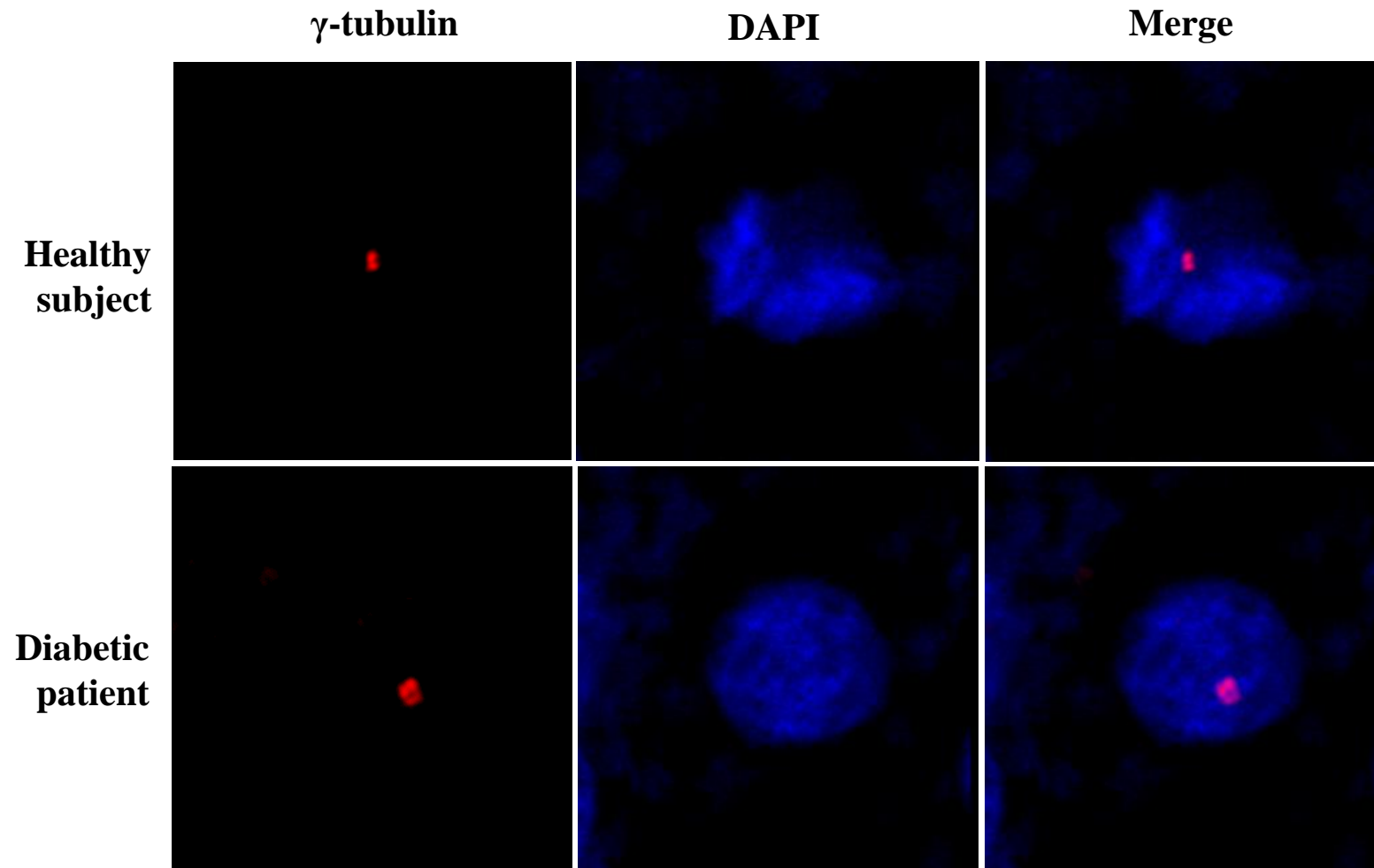


Fig. 1B

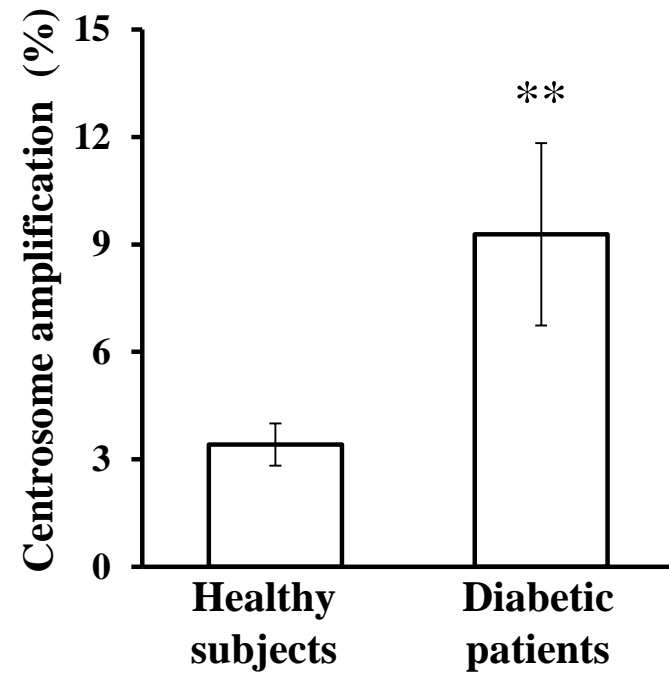


Fig. 1C

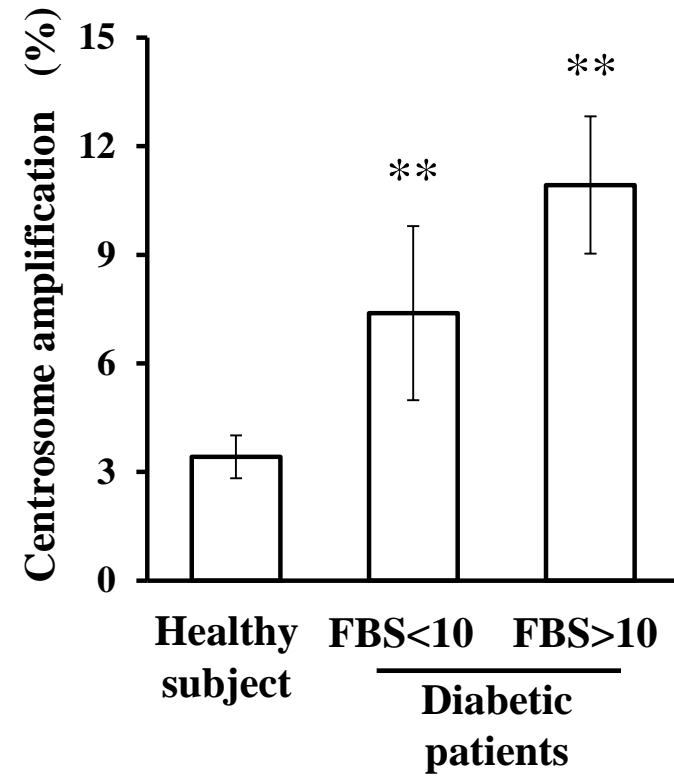


Fig. 1D

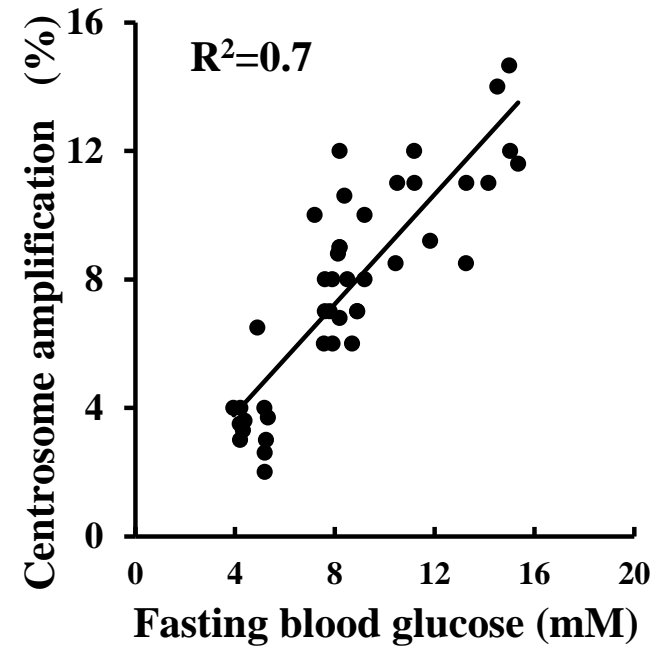


Fig. 1E

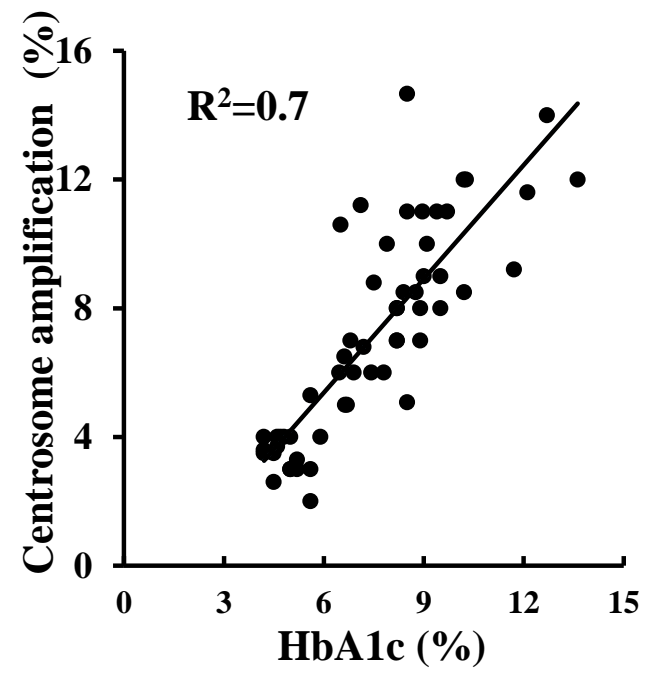


Fig. 1F

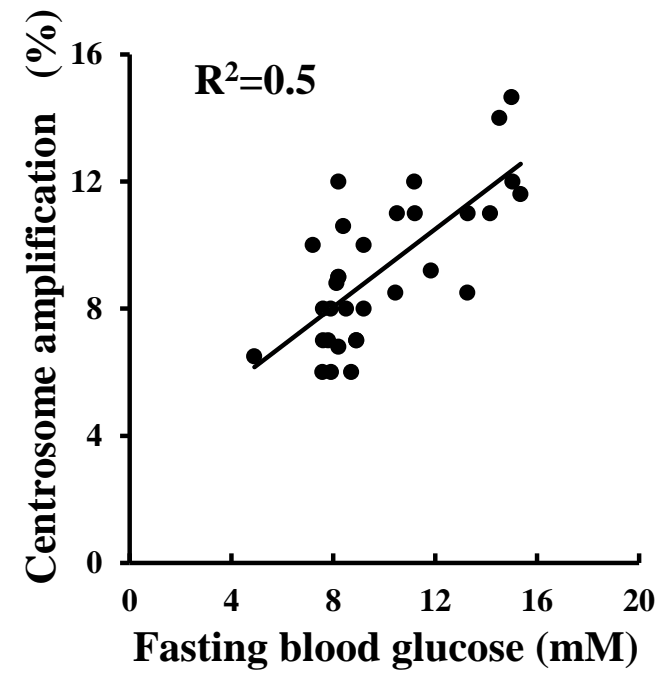


Fig. 1G

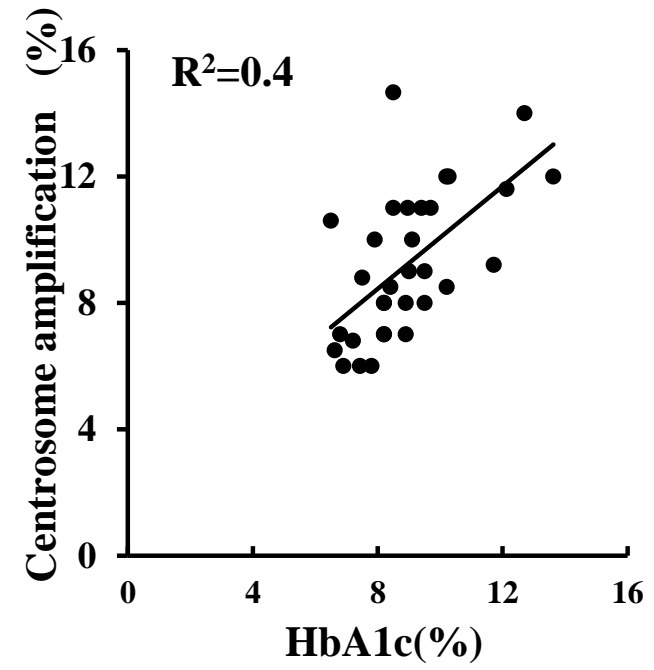


Fig. 2A

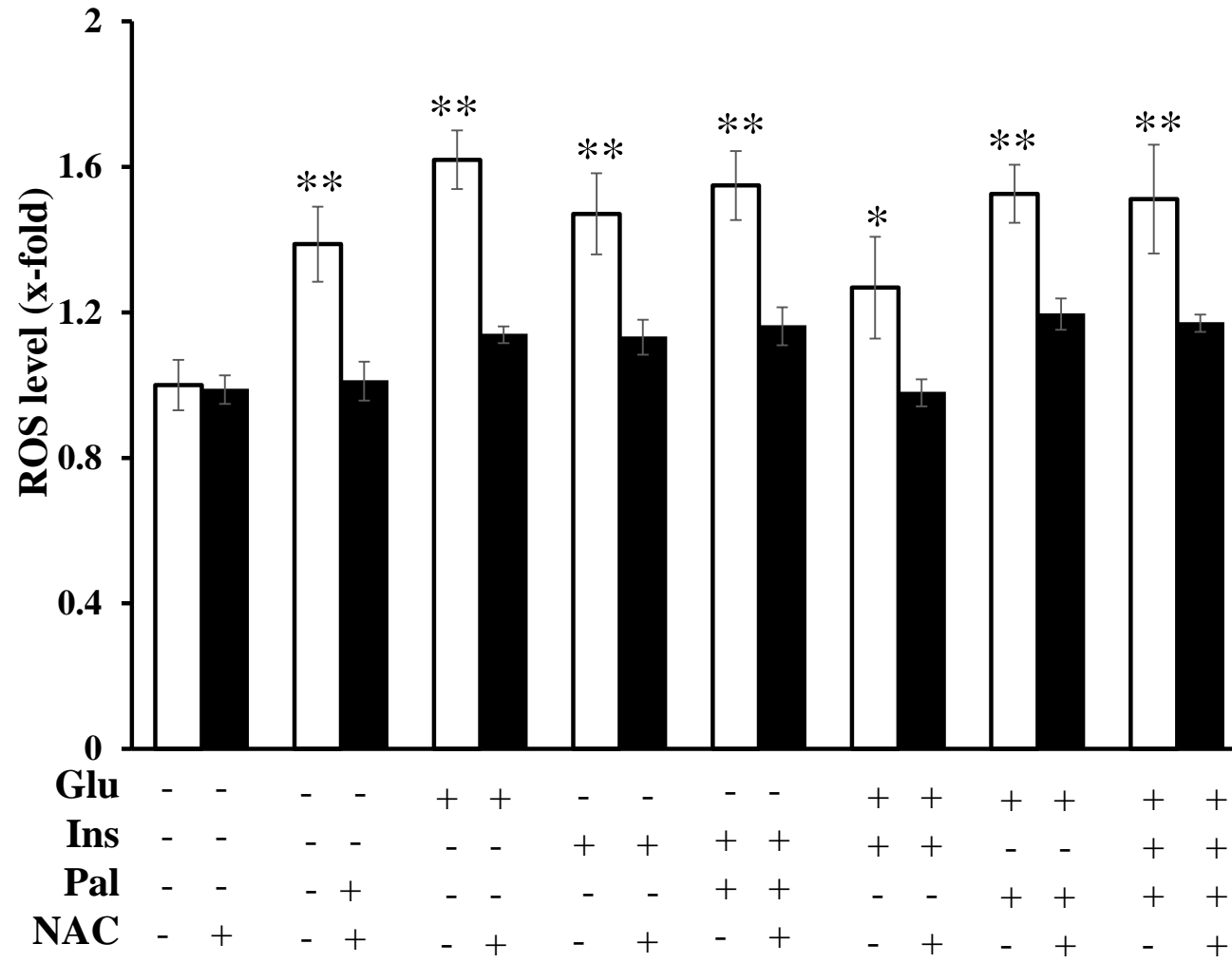


Fig. 2B

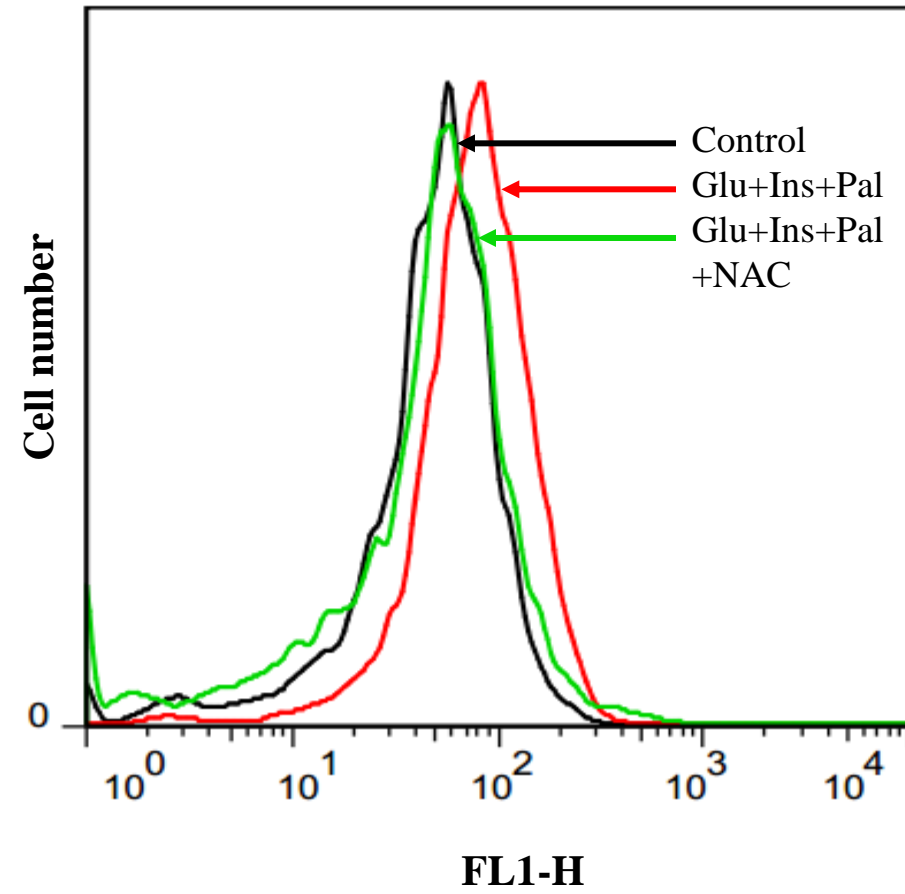


Fig. 2C

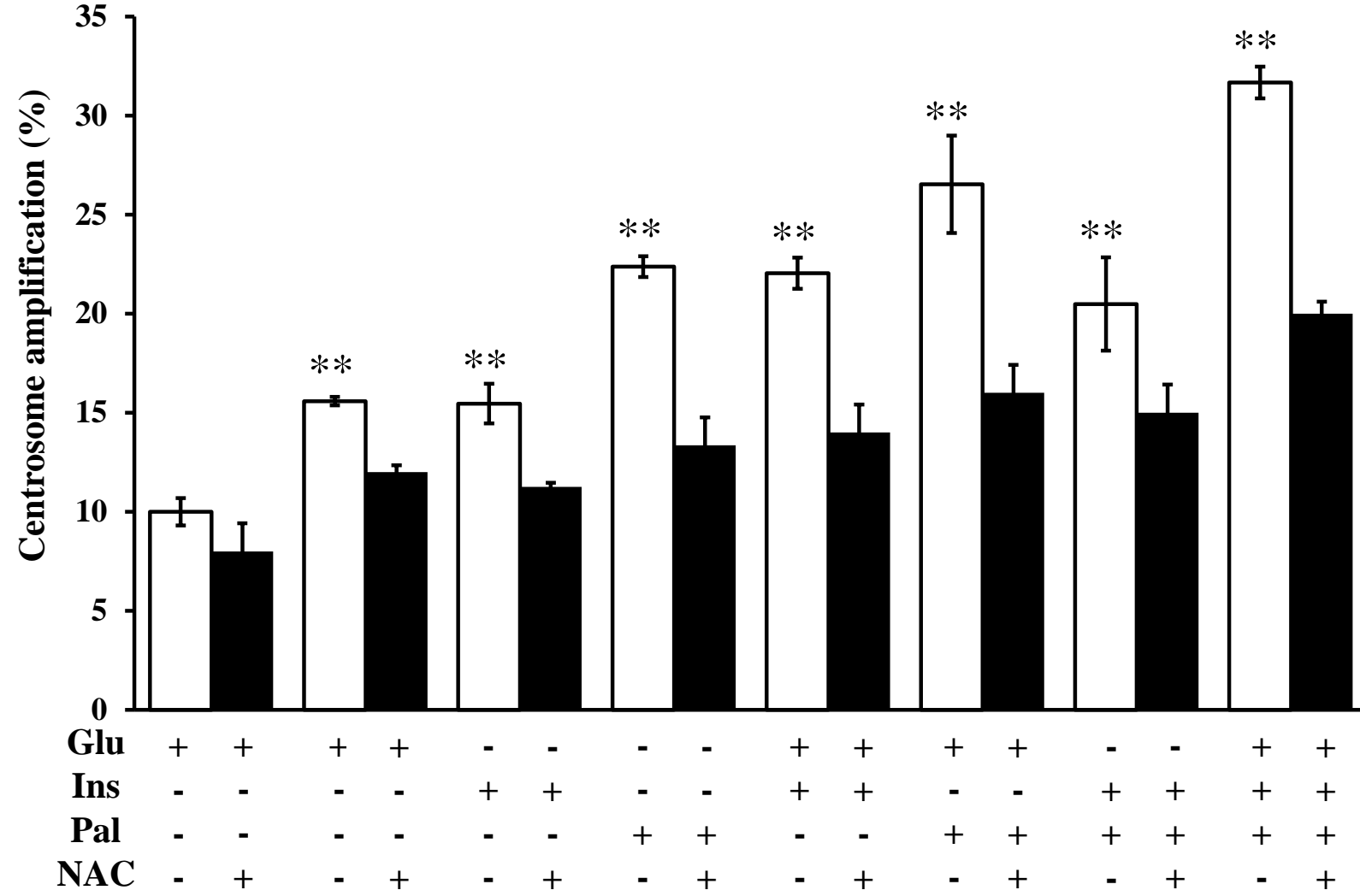


Fig. 2D

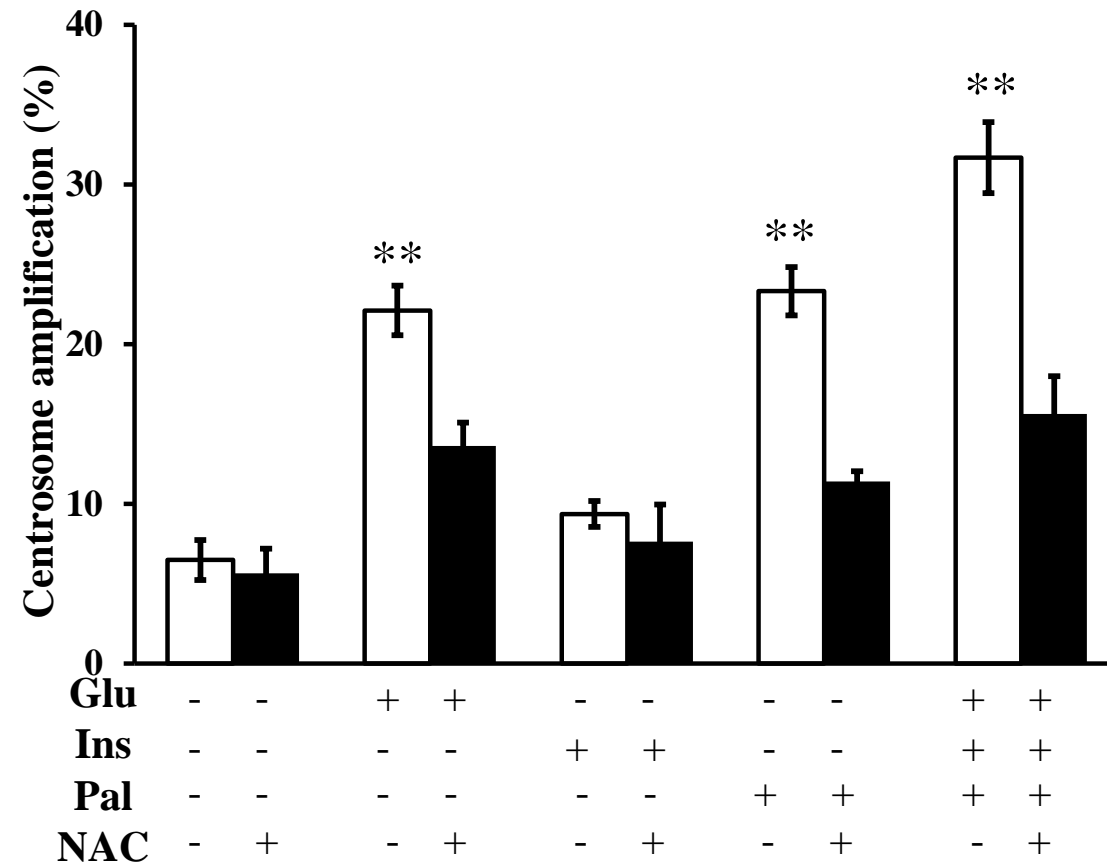


Fig. 3A

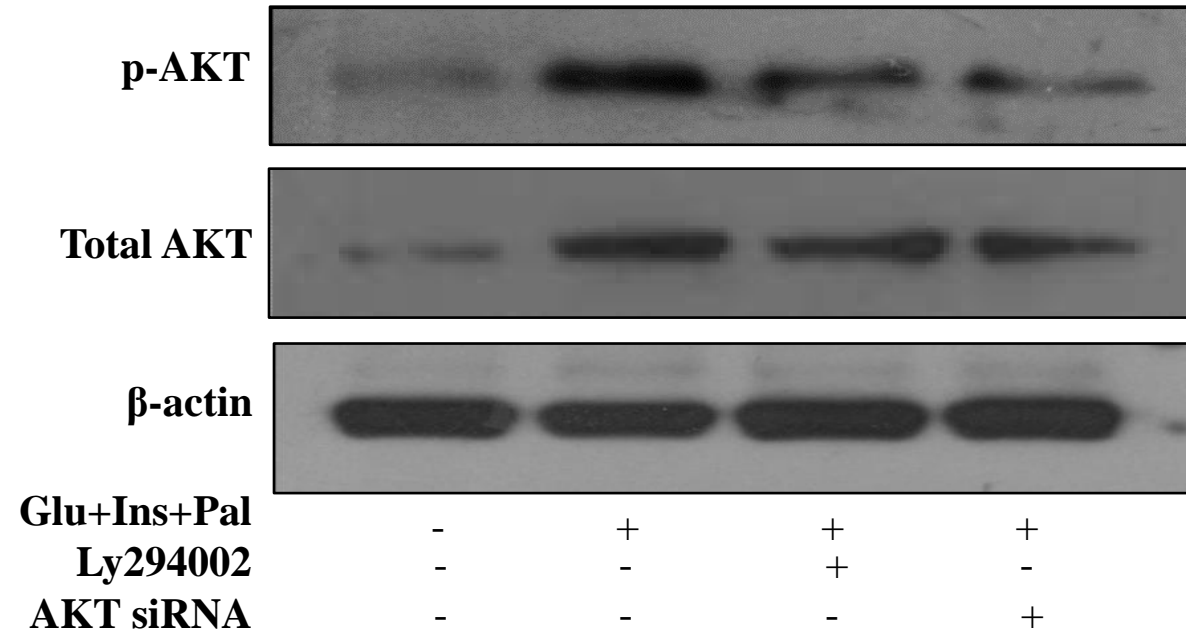


Fig. 3B

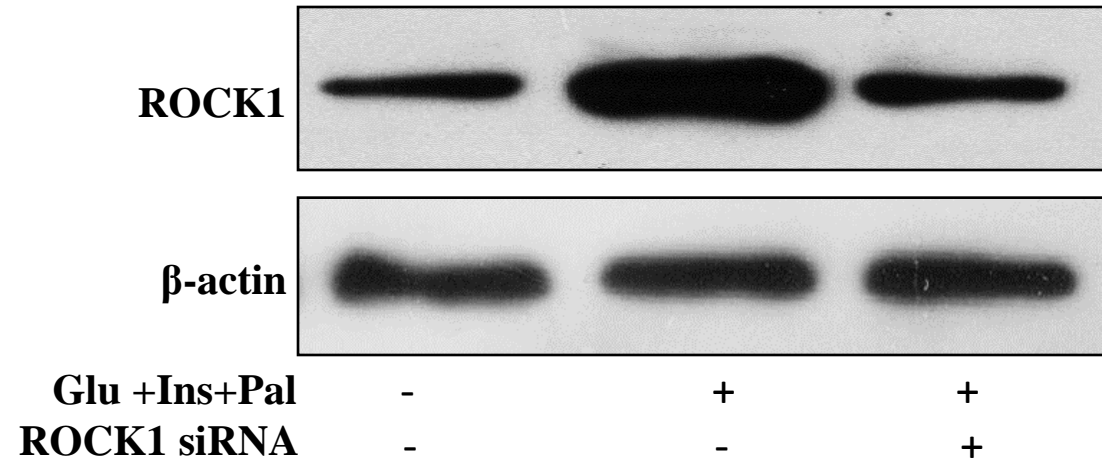


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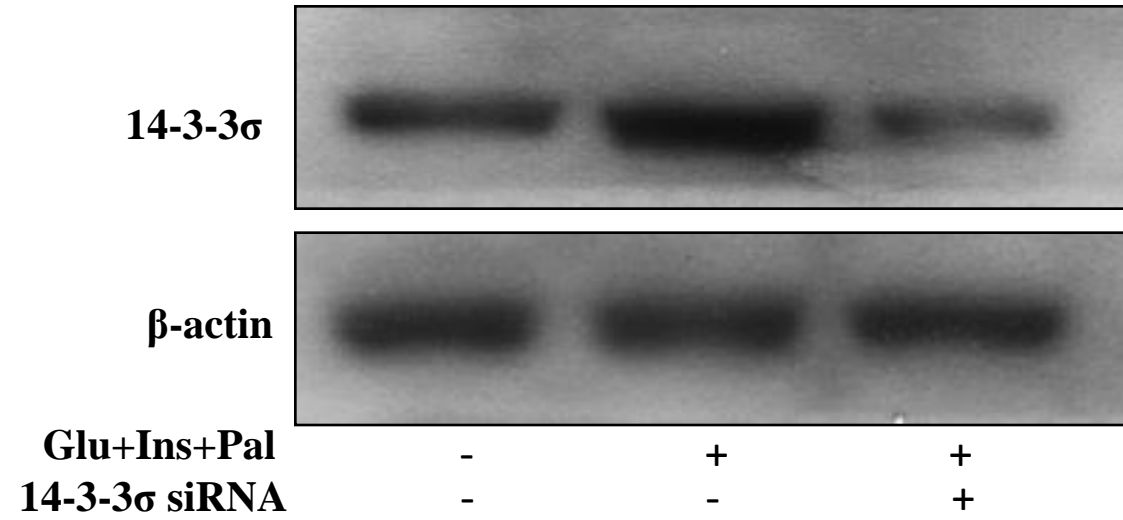


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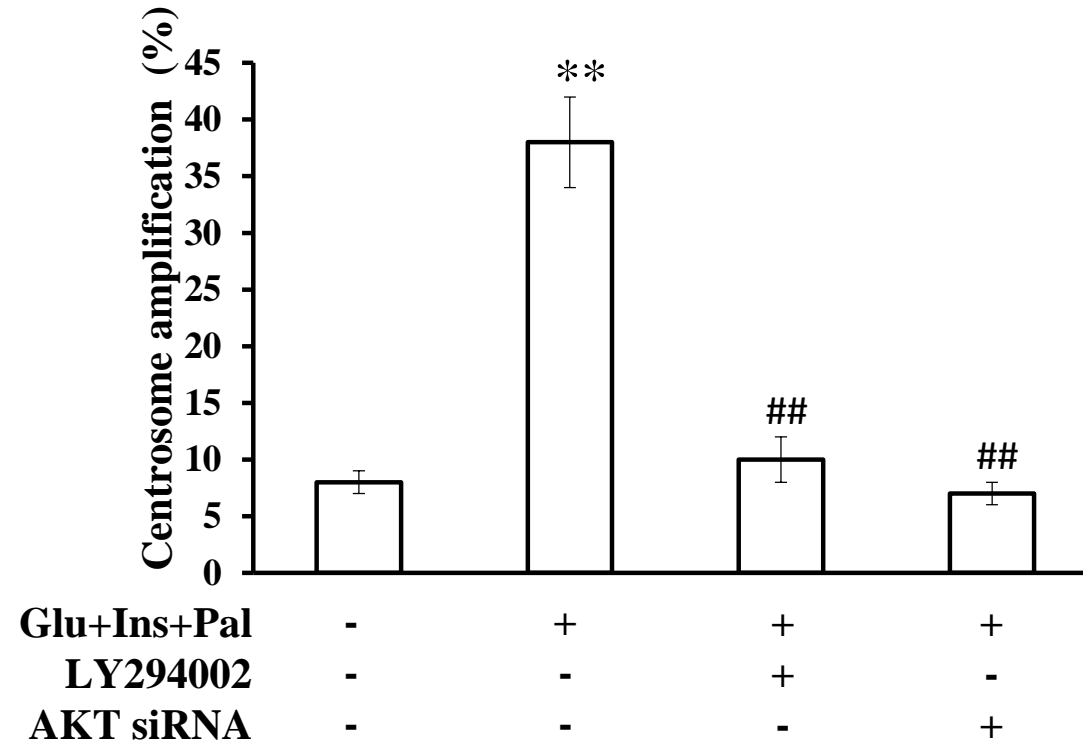


Fig. 3E

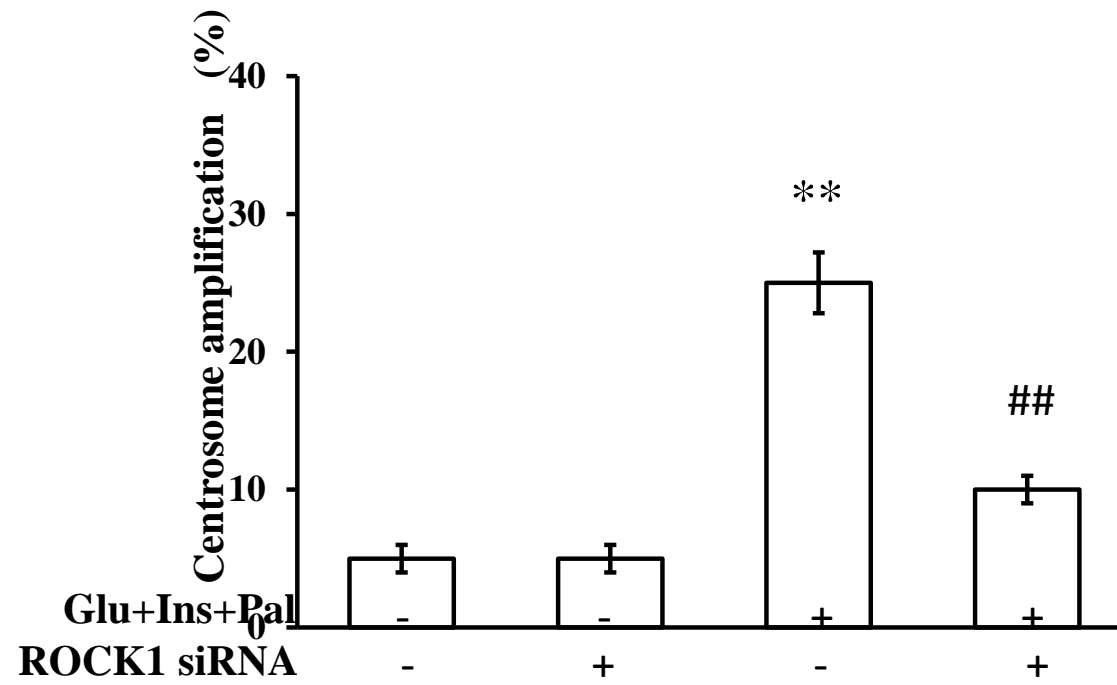


Fig. 3F

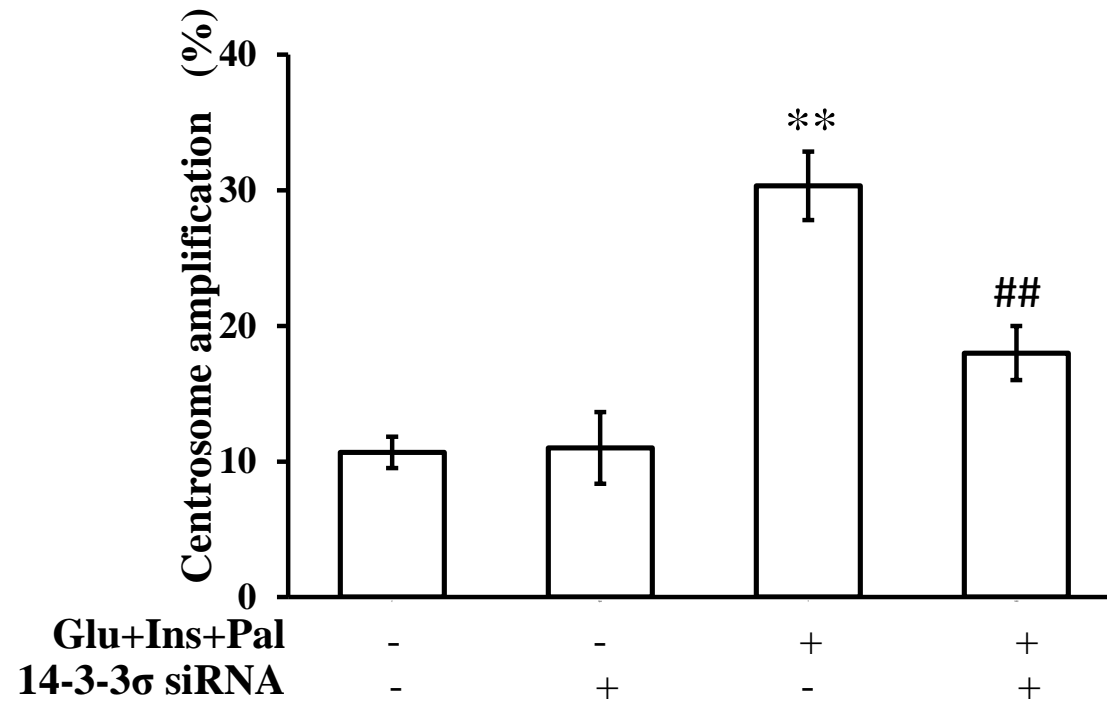


Fig. 3G

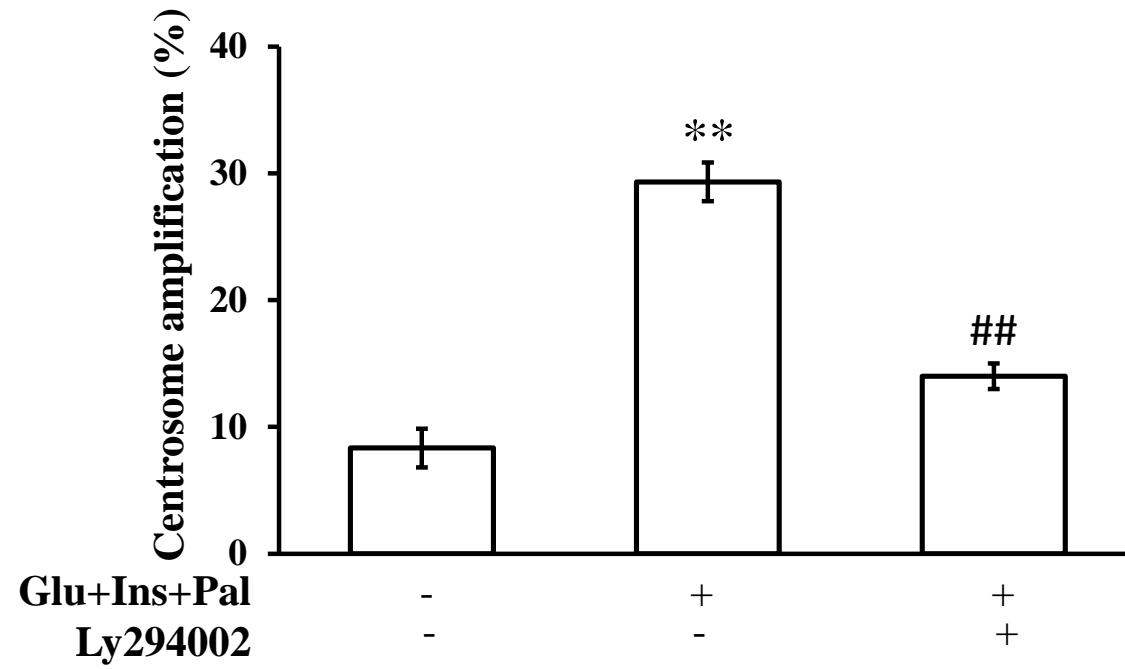


Fig. 3H

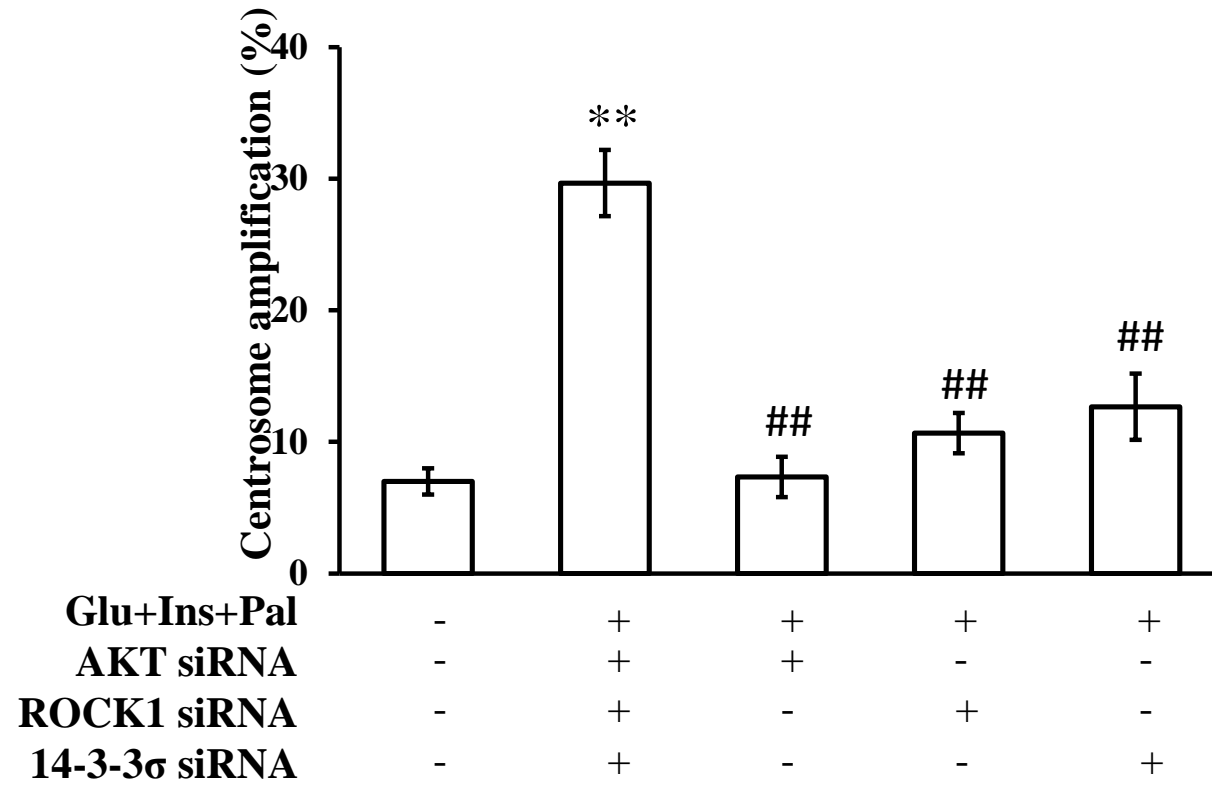


Fig. 4A

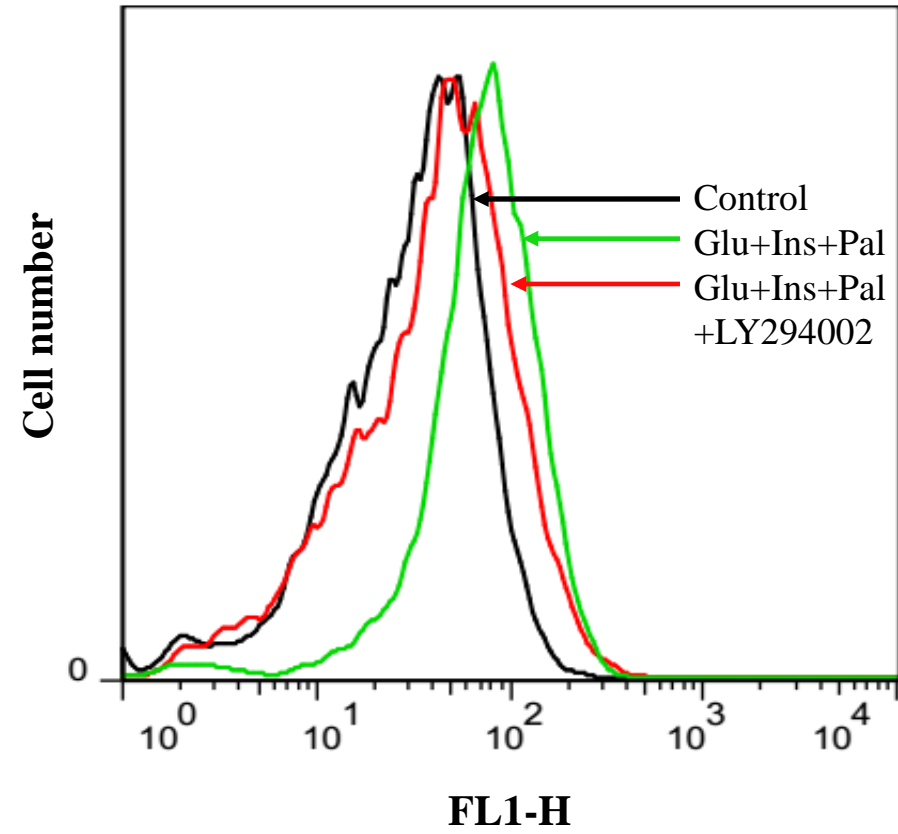


Fig. 4B

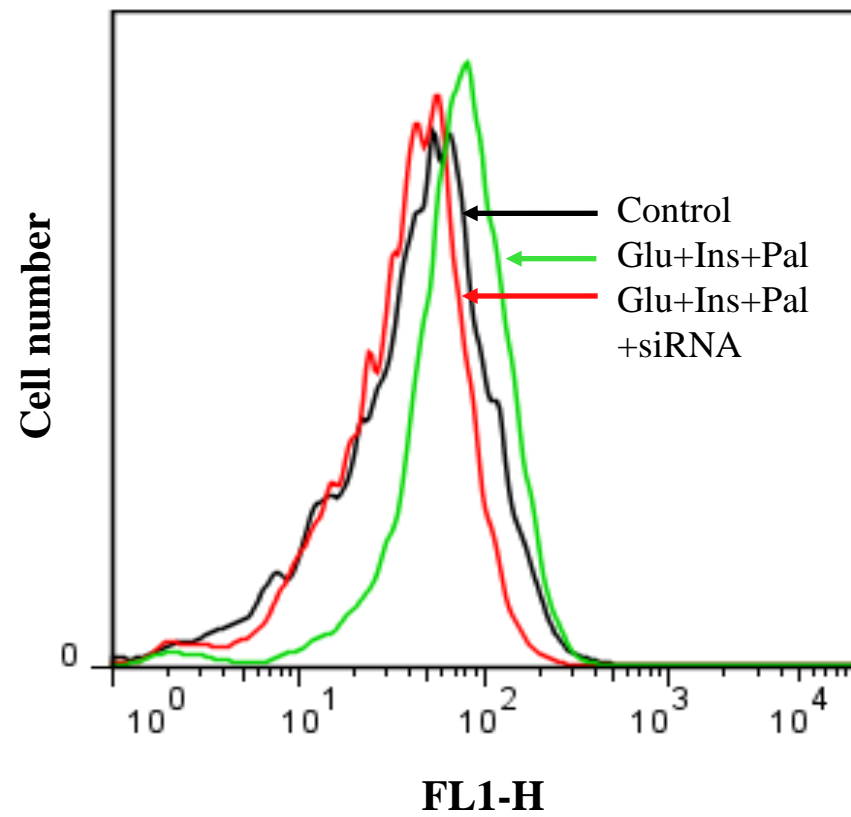


Fig. 4C

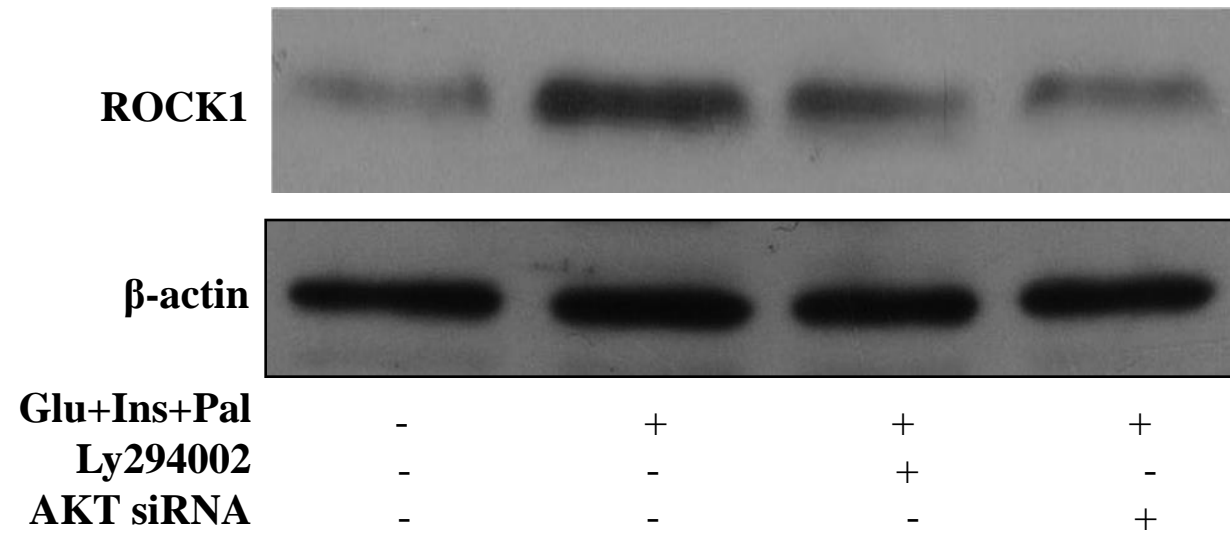


Fig. 4D

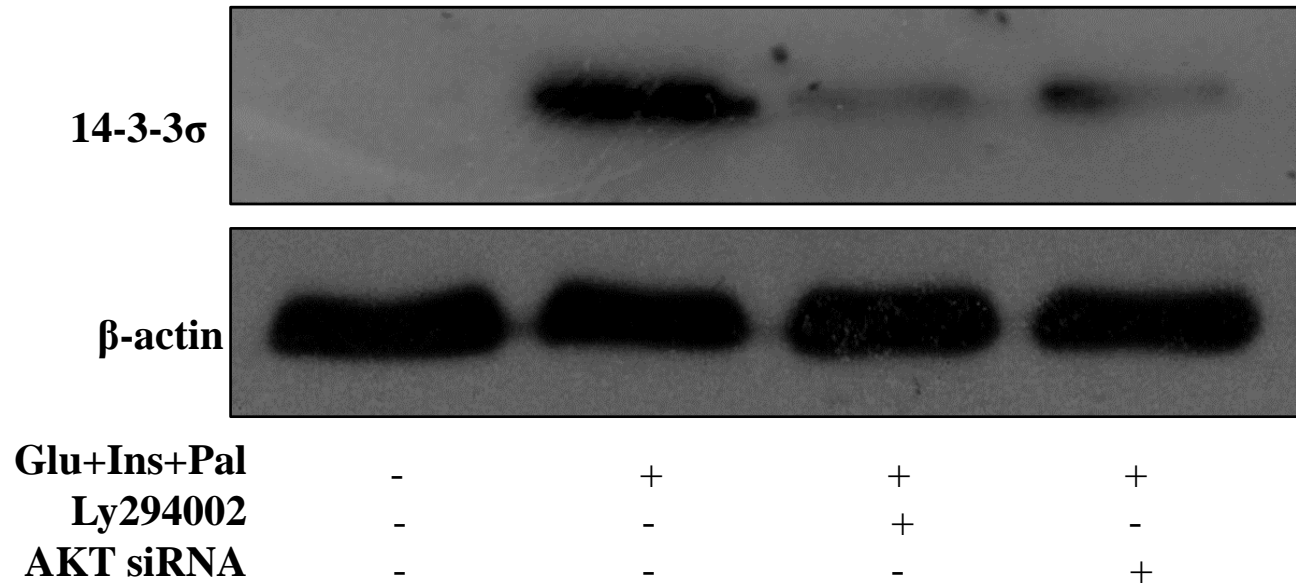


Fig. 4E

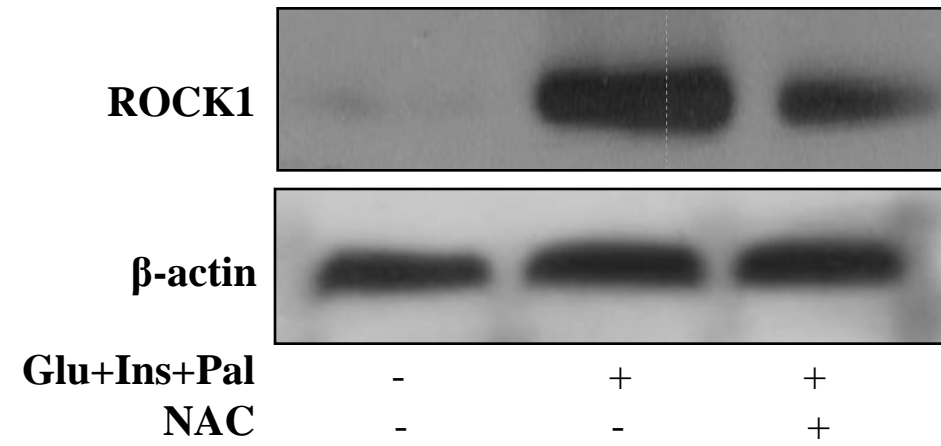


Fig. 4F

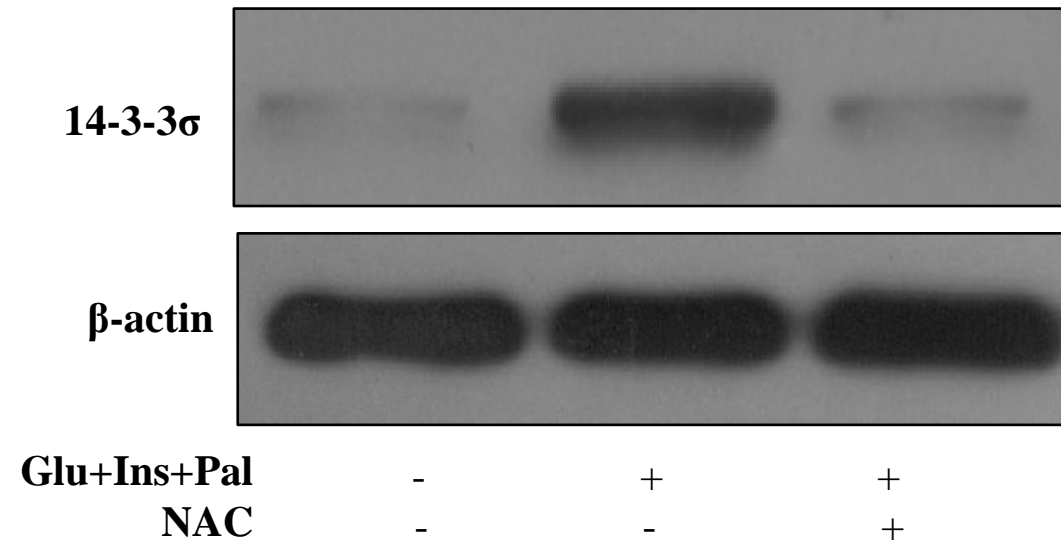


Fig. 5A

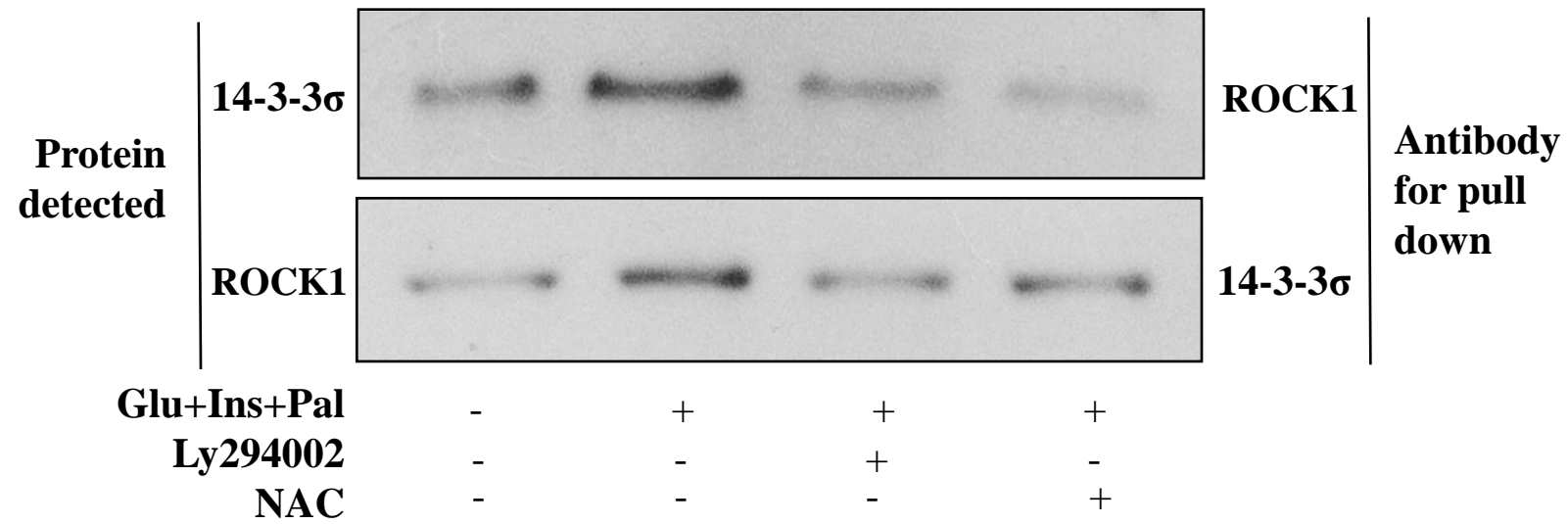


Fig. 5B

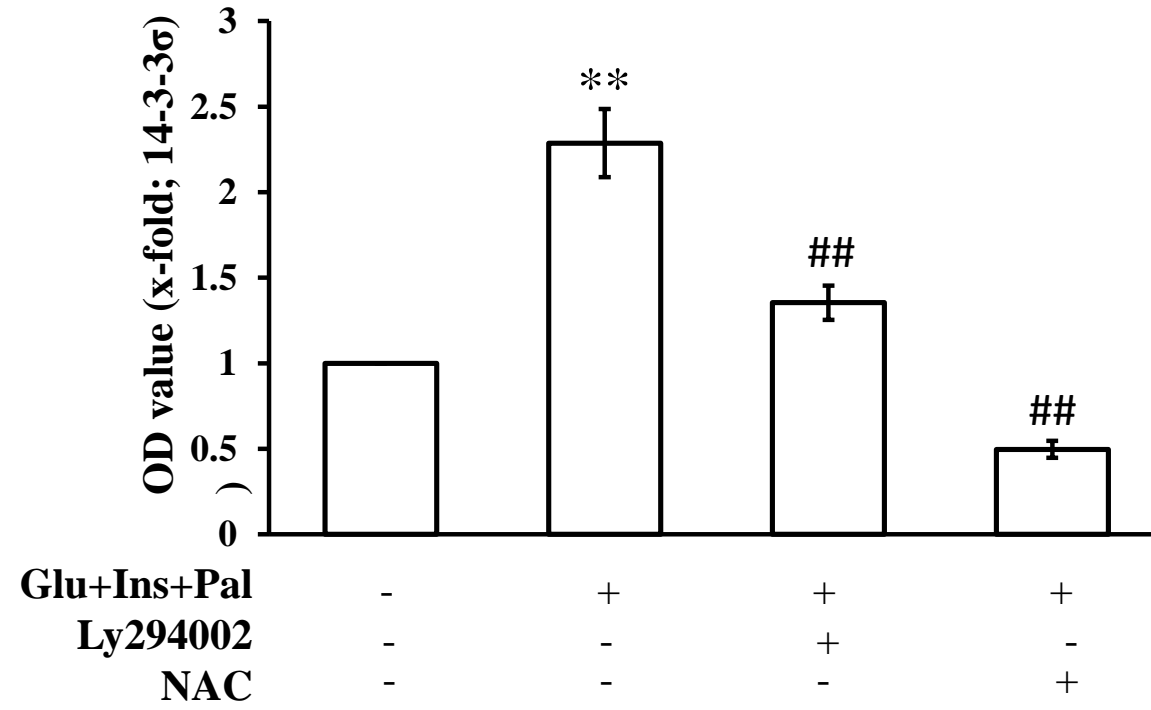


Fig. 5C

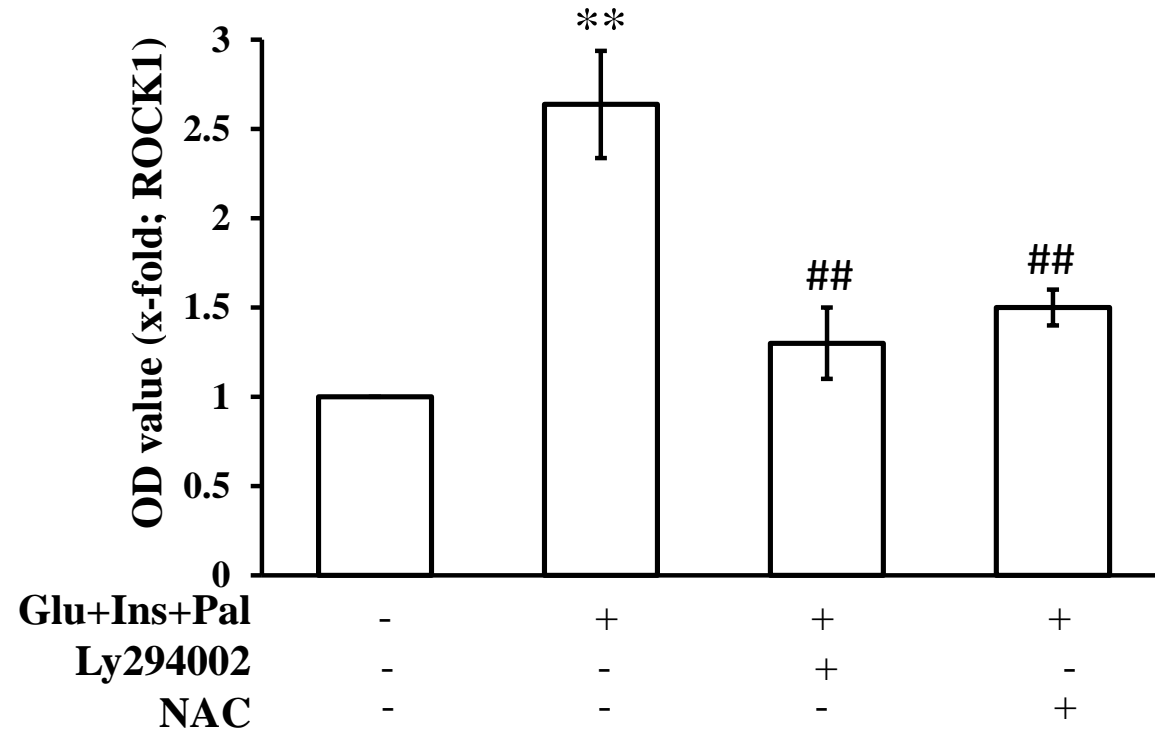


Fig. 5D

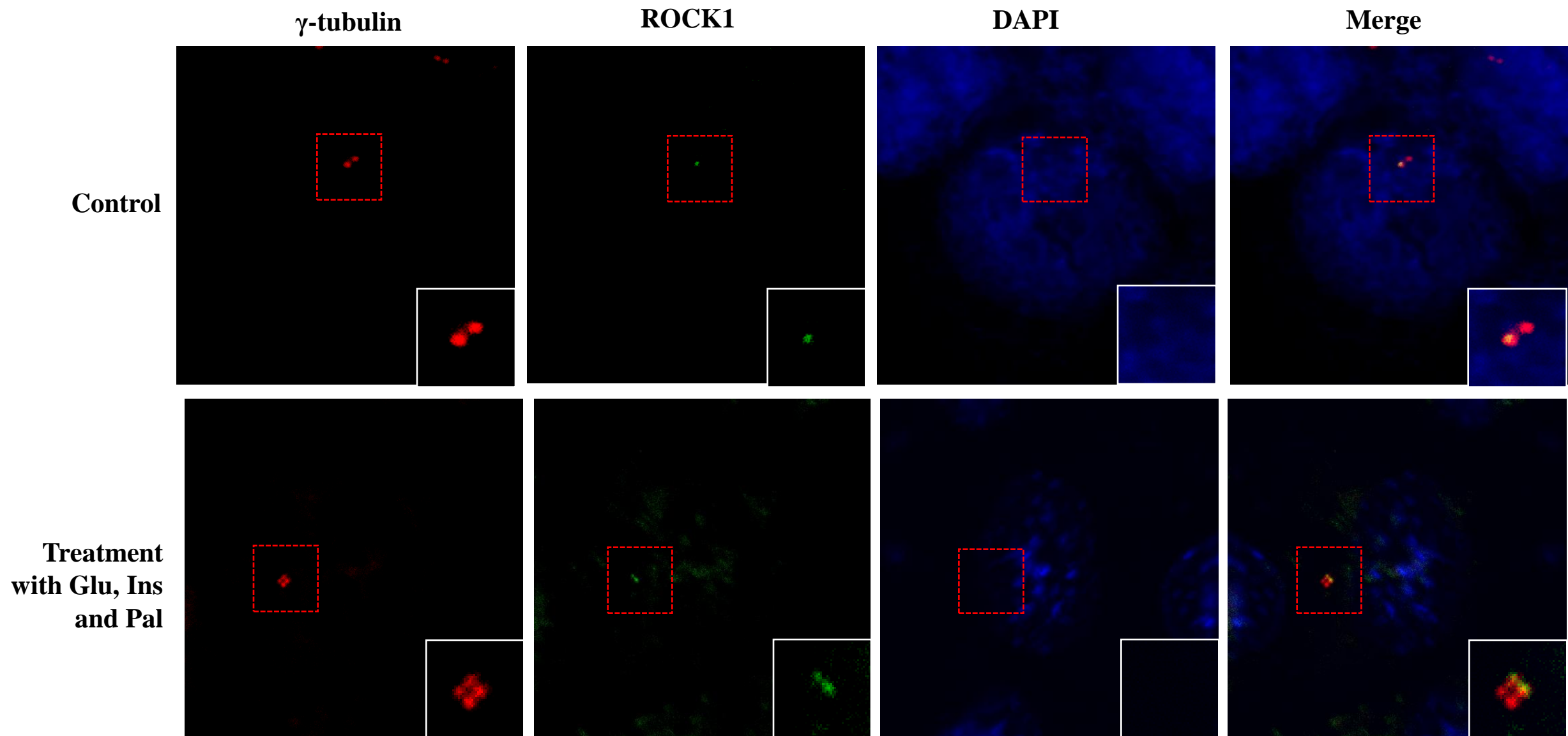


Fig. 5E

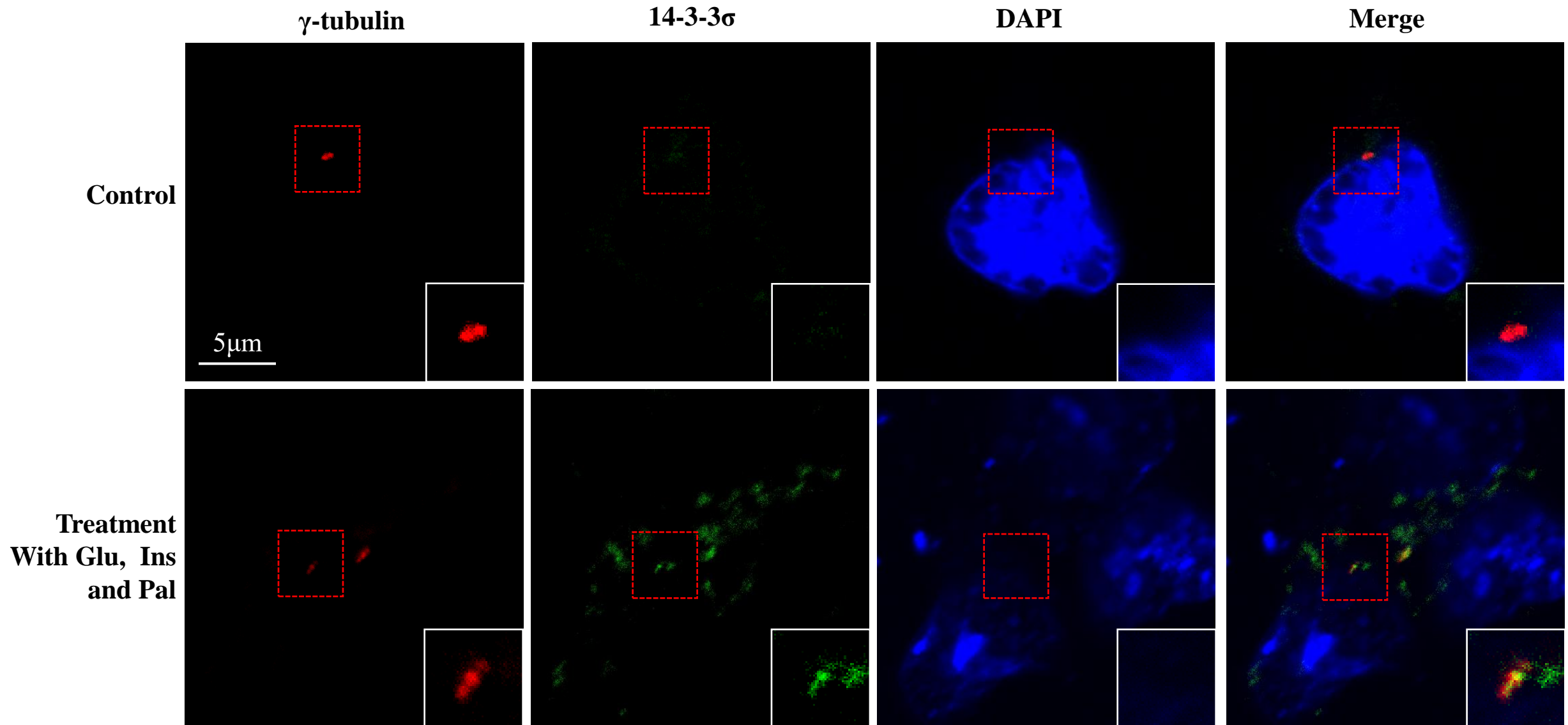


Fig. 5F

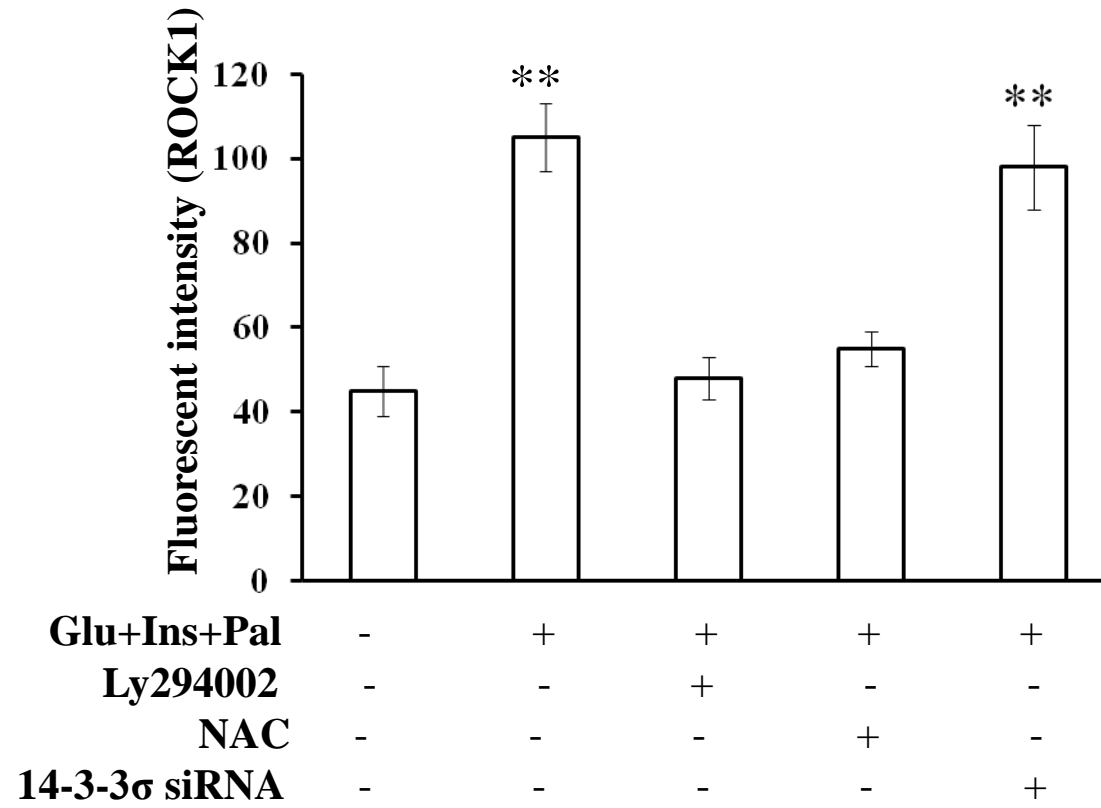


Fig. 5G

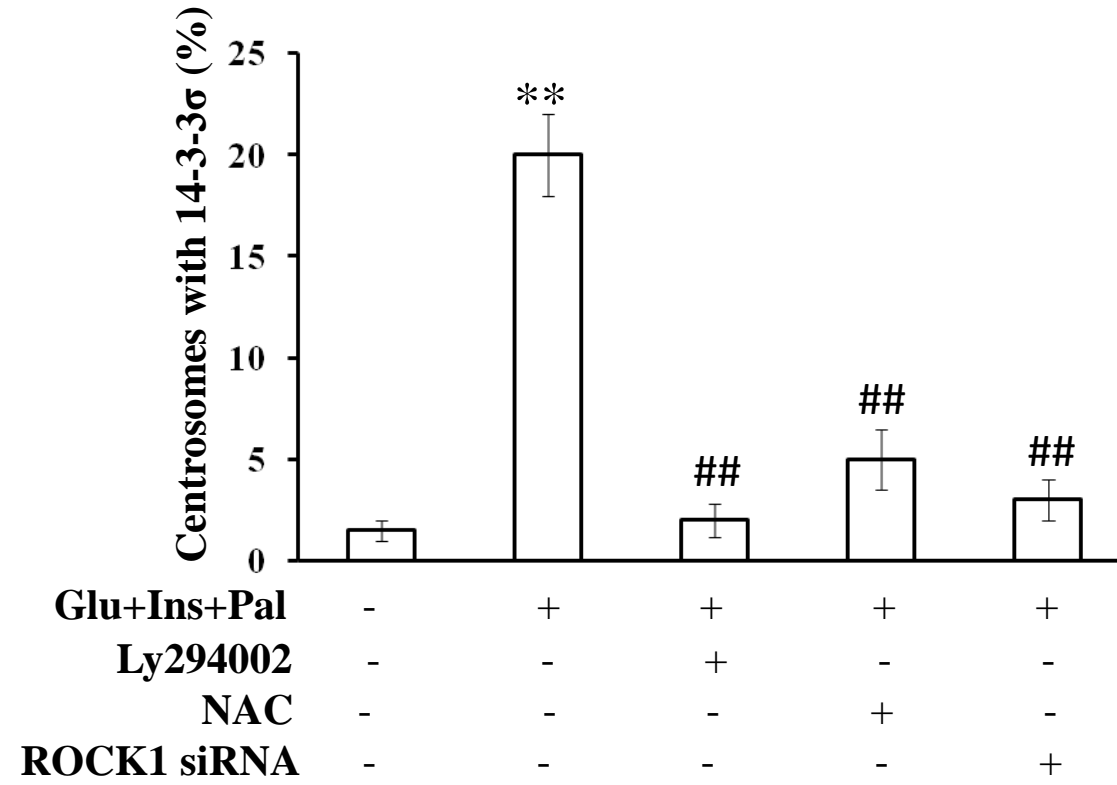


Fig. 6A

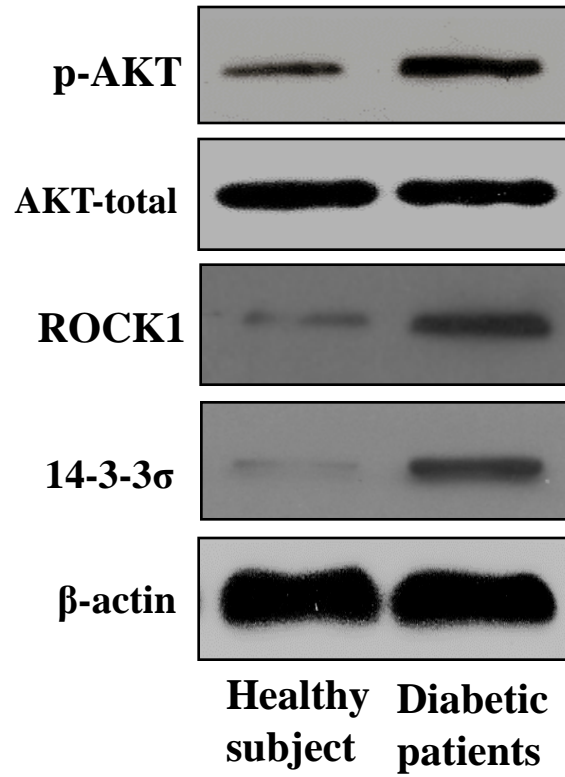


Fig. 6B

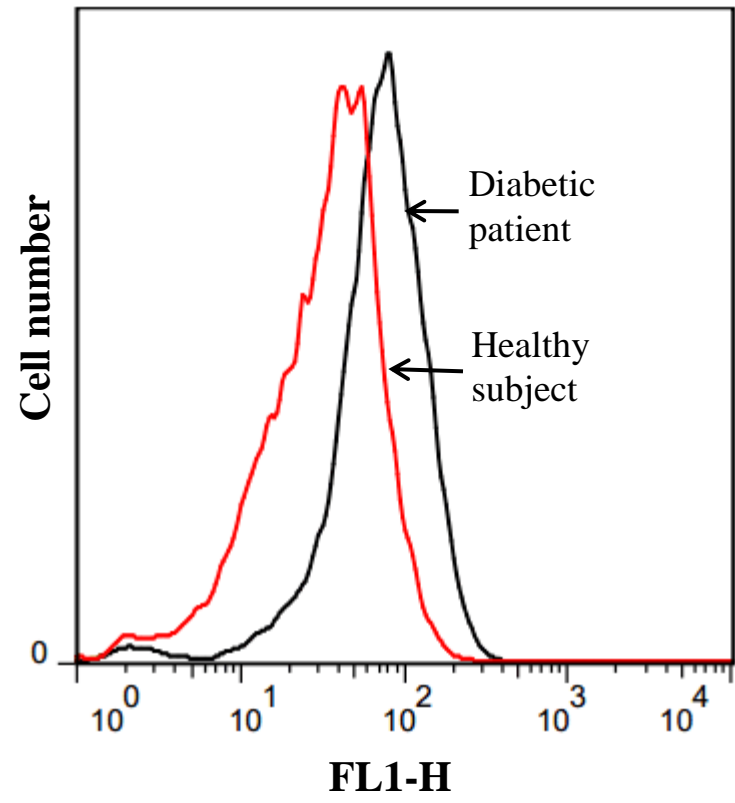


Fig. 6C

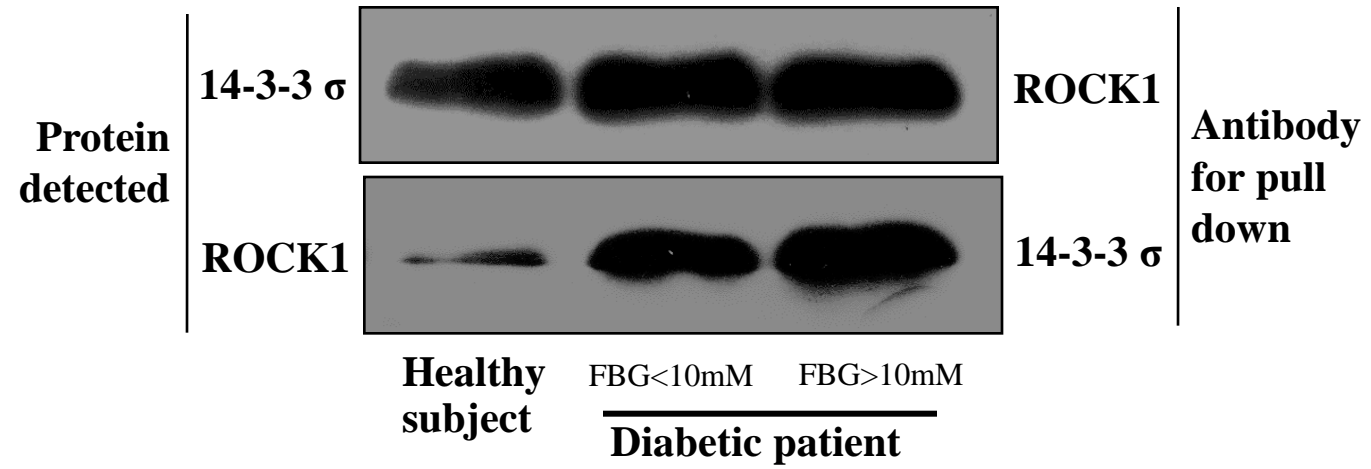


Fig. 6D

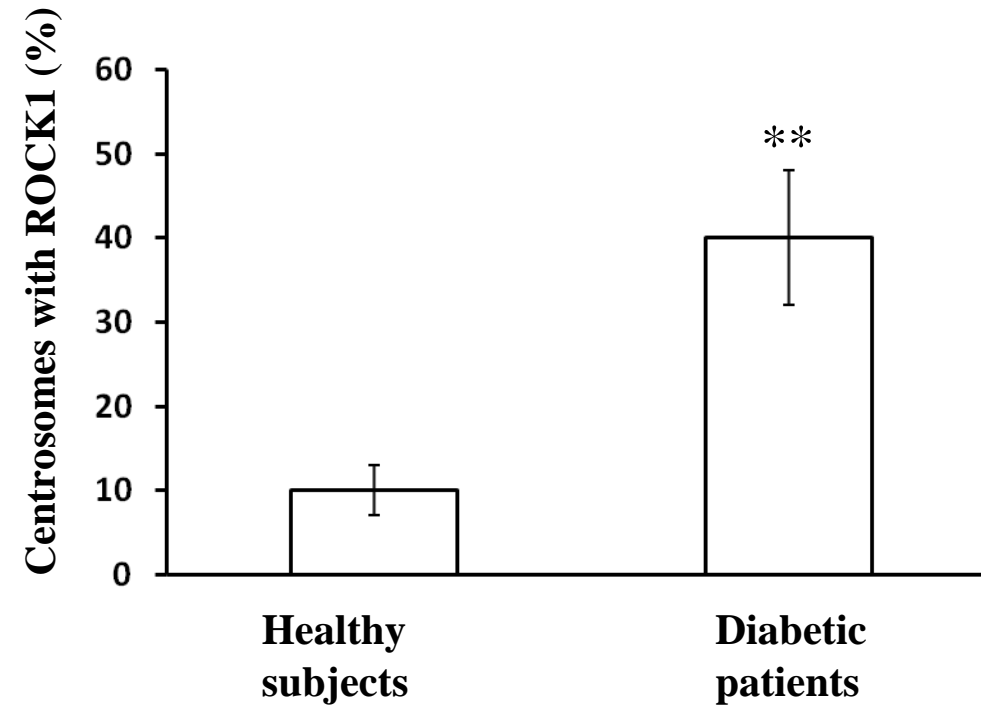


Fig. 6E

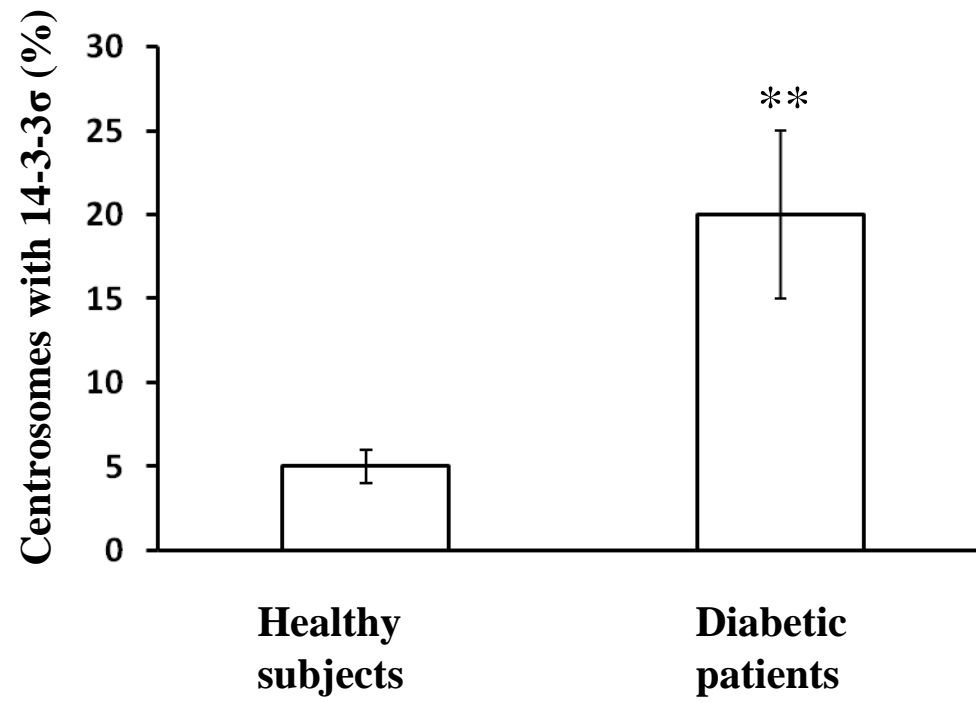


Table S1. Differentially expressed genes and KEGG pathway analysis

Metabolism			
Overview			
ko01200	Carbon metabolism		
	ko:K00036	G6PD, zwf; glucose-6-phosphate 1-dehydrogenase	T up
ko01210	2-Oxocarboxylic acid metabolism		
ko01212	Fatty acid metabolism		
	ko:K08765	CPT1; carnitine O-palmitoyltransferase 1	T up
ko00030	Pentose phosphate pathway		
	ko:K00036	G6PD, zwf; glucose-6-phosphate 1-dehydrogenase	T up
ko00040	Pentose and glucuronate interconversions		
	ko:K00011	E1.1.1.21, AKR1; aldehyde reductase	T up
ko00051	Fructose and mannose metabolism		
	ko:K00846	KHK; ketohexokinase	T up
	ko:K00011	E1.1.1.21, AKR1; aldehyde reductase	T up
ko00052	Galactose metabolism		
	ko:K00011	E1.1.1.21, AKR1; aldehyde reductase	T up
ko00500	Starch and sucrose metabolism		
	ko:K01513	ENPP1_3; ectonucleotide pyrophosphatase/phosphodiesterase family member 1/3	T up
ko00520	Amino sugar and nucleotide sugar metabolism		
	ko:K00820	E2.6.1.16, glmS; glucosamine--fructose-6-phosphate aminotransferase	T up
ko00620	Pyruvate metabolism		
	ko:K00011	E1.1.1.21, AKR1; aldehyde reductase	T up
ko00630	Glyoxylate and dicarboxylate metabolism		
	ko:K15788	GLYCTK; glycerate kinase	C up
Energy metabolism			
ko00190	Oxidative phosphorylation		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
ko00910	Nitrogen metabolism		
	ko:K01672	E4.2.1.1; carbonic anhydrase	T up
ko00920	Sulfur metabolism		
	ko:K17218	sqr; sulfide:quinone oxidoreductase	T up
Lipid metabolism			
ko00061	Fatty acid biosynthesis		
ko00062	Fatty acid elongation		
	ko:K10248	ELOVL3; elongation of very long chain fatty acids protein 3	T up
ko00071	Fatty acid degradation		
	ko:K08765	CPT1; carnitine O-palmitoyltransferase 1	T up
ko00140	Steroid hormone biosynthesis		
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1	T up
	ko:K04119	AKR1C3; aldo-keto reductase family 1 member C3	T up
	ko:K01015	SULT2B1; alcohol sulfotransferase	T up

ko00561	Glycerolipid metabolism		
	ko:K01080	PPAP2; phosphatidate phosphatase	T up
	ko:K11160	DGAT2; diacylglycerol O-acyltransferase 2	T up
	ko:K00011	E1.1.1.21, AKR1; aldehyde reductase	T up
	ko:K15788	GLYCTK; glycerate kinase	C up
ko00564	Glycerophospholipid metabolism		
	ko:K01080	PPAP2; phosphatidate phosphatase	T up
	ko:K06124	PHOSPHO1; phosphoethanolamine/phosphocholine phosphatase	T up
	ko:K01049	ACHE; acetylcholinesterase	T up
ko00565	Ether lipid metabolism		
	ko:K01080	PPAP2; phosphatidate phosphatase	T up
ko00600	Sphingolipid metabolism		
	ko:K04718	SPHK; sphingosine kinase	T up
	ko:K12350	SMPD1, ASM; sphingomyelin phosphodiesterase	T up
	ko:K01080	PPAP2; phosphatidate phosphatase	T up
ko00590	Arachidonic acid metabolism		
	ko:K00509	PTGS1, COX1; prostaglandin-endoperoxide synthase 1	T up
	ko:K04119	AKR1C3; aldo-keto reductase family 1 member C3	T up
	ko:K07418	CYP2J; cytochrome P450, family 2, subfamily J	T up
	ko:K01832	TBXAS1, CYP5A; thromboxane-A synthase	T up
ko00591	Linoleic acid metabolism		
	ko:K07418	CYP2J; cytochrome P450, family 2, subfamily J	T up
Nucleotide metabolism			
ko00230	Purine metabolism		
	ko:K01513	ENPP1_3; ectonucleotide pyrophosphatase/phosphodiesterase family member 1/3	T up
	ko:K01509	E3.6.1.3; adenosinetriphosphatase	T up
	ko:K01487	E3.5.4.3, guaD; guanine deaminase	C up
	ko:K13762	PDE5; cGMP-specific 3',5'-cyclic phosphodiesterase	T up
	ko:K08045	ADCY5; adenylate cyclase 5	T up
ko00240	Pyrimidine metabolism		
Amino acid metabolism			
ko00250	Alanine, aspartate and glutamate metabolism		
	ko:K00820	E2.6.1.16, glmS; glucosamine--fructose-6-phosphate aminotransferase (isomerizing)	T up
ko00260	Glycine, serine and threonine metabolism		
	ko:K15788	GLYCTK; glycerate kinase	C up
	ko:K00314	SARDH; sarcosine dehydrogenase	T up
ko00330	Arginine and proline metabolism		
	ko:K00472	E1.14.11.2; prolyl 4-hydroxylase	T up
ko00380	Tryptophan metabolism		
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1	T up
Metabolism of other amino acids			

ko00480	Glutathione metabolism		
	ko:K00799	GST, gst; glutathione S-transferase	T up
	ko:K00036	G6PD, zwf; glucose-6-phosphate 1-dehydrogenase	T up
Glycan biosynthesis and metabolism			
ko00513	Various types of N-glycan biosynthesis		
	ko:K00781	SIAT6; N-acetylglucosaminyl transferase (sialyltransferase 6)	C up
ko00512	Mucin type O-glycan biosynthesis		
	ko:K00727	GCNT1; beta-1,3-galactosyl-O-glycosyl-glycoprotein beta-1,6-N-acetylglucosaminyltransferase	T up
	ko:K00710	GALNT; polypeptide N-acetylgalactosaminyltransferase	T up
	ko:K00780	SIAT4A; beta-galactoside alpha-2,3-sialyltransferase (sialyltransferase 4A) [EC:2.4.99.4]	T up
ko00514	Other types of O-glycan biosynthesis		
	ko:K05948	FNG; fringe [EC:2.4.1.222]	T up
	ko:K00781	SIAT6; N-acetylglucosaminyl transferase (sialyltransferase 6) [EC:2.4.99.6]	C up
ko00532	Glycosaminoglycan biosynthesis - chondroitin sulfate / dermatan sulfate		
	ko:K00771	XYLT; protein xylosyltransferase [EC:2.4.2.26]	T up
ko00534	Glycosaminoglycan biosynthesis - heparan sulfate / heparin		
	ko:K00771	XYLT; protein xylosyltransferase [EC:2.4.2.26]	T up
ko00533	Glycosaminoglycan biosynthesis - keratan sulfate		
	ko:K00780	SIAT4A; beta-galactoside alpha-2,3-sialyltransferase (sialyltransferase 4A) [EC:2.4.99.4]	T up
	ko:K00741	B3GNT1, B3GNT2; N-acetylglucosaminyltransferase [EC:2.4.1.149]	T up
	ko:K09664	B3GNT7; beta-1,3-N-acetylglucosaminyltransferase 7 [EC:2.4.1.-]	T up
	ko:K04745	CHST2; carbohydrate 6-sulfotransferase 2 [EC:2.8.2.-]	T up
	ko:K00781	SIAT6; N-acetylglucosaminyl transferase (sialyltransferase 6) [EC:2.4.99.6]	C up
ko00601	Glycosphingolipid biosynthesis - lacto and neolacto series		
	ko:K00741	B3GNT1, B3GNT2; N-acetylglucosaminyltransferase [EC:2.4.1.149]	T up
	ko:K00781	SIAT6; N-acetylglucosaminyl transferase (sialyltransferase 6) [EC:2.4.99.6]	C up
ko00603	Glycosphingolipid biosynthesis - globo series		
	ko:K00780	SIAT4A; beta-galactoside alpha-2,3-sialyltransferase (sialyltransferase 4A) [EC:2.4.99.4]	T up
	ko:K01988	A4GALT; lactosylceramide 4-alpha-galactosyltransferase [EC:2.4.1.228]	T up
ko00604	Glycosphingolipid biosynthesis - ganglio series		
	ko:K03373	SIAT7C, ST6GalNAc III; N-acetylgalactosaminyltransferase (sialyltransferase 7C) [EC:2.4.99.-]	T up

	ko:K00780	SIAT4A; beta-galactoside alpha-2,3-sialyltransferase (sialyltransferase 4A) [EC:2.4.99.4]	T up
	ko:K00725	B4GALNT1, GALGT; (N-Acetylneuraminyl)-galactosylglucosylceramide N-acetylgalactosaminyltransferase [EC:2.4.1.92]	T up
Metabolism of cofactors and vitamins			
ko00730	Thiamine metabolism		
ko00740	Riboflavin metabolism		
	ko:K01513	ENPP1_3; ectonucleotide pyrophosphatase/phosphodiesterase family member 1/3 [EC:3.1.4.1 3.6.1.9]	T up
ko00750	Vitamin B6 metabolism		
ko00760	Nicotinate and nicotinamide metabolism		
	ko:K01513	ENPP1_3; ectonucleotide pyrophosphatase/phosphodiesterase family member 1/3 [EC:3.1.4.1 3.6.1.9]	T up
ko00770	Pantothenate and CoA biosynthesis		
	ko:K01513	ENPP1_3; ectonucleotide pyrophosphatase/phosphodiesterase family member 1/3 [EC:3.1.4.1 3.6.1.9]	T up
ko00830	Retinol metabolism		
	ko:K12664	CYP26B; cytochrome P450, family 26, subfamily B	T up
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1 [EC:1.14.14.1]	T up
Metabolism of terpenoids and polyketides			
ko00980	Metabolism of xenobiotics by cytochrome P450		
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1 [EC:1.14.14.1]	T up
	ko:K00799	GST, gst; glutathione S-transferase [EC:2.5.1.18]	T up
ko00982	Drug metabolism - cytochrome P450		
	ko:K00799	GST, gst; glutathione S-transferase [EC:2.5.1.18]	T up
Enzyme families			
Genetic Information Processing			
Transcription			
ko03040	Spliceosome		
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
ko03041	Spliceosome		
Translation			
ko03010	Ribosome		
	ko:K02929	RP-L44e, RPL44; large subunit ribosomal protein L44e	C up
ko03011	Ribosome		
ko03016	Transfer RNA biogenesis		
ko00970	Aminoacyl-tRNA biosynthesis		
	ko:K01867	WARS, trpS; tryptophanyl-tRNA synthetase [EC:6.1.1.2]	C up
ko03013	RNA transport		

	ko:K03257	EIF4A; translation initiation factor 4A	C up
	ko:K03250	EIF3E, INT6; translation initiation factor 3 subunit E	C up
	ko:K00784	rnz; ribonuclease Z [EC:3.1.26.11]	C up
	ko:K03245	EIF3J; translation initiation factor 3 subunit J	C up
Folding, sorting and degradation			
ko03060	Protein export		
	ko:K09490	HSPA5, BIP; heat shock 70kDa protein 5	C up
ko04141	Protein processing in endoplasmic reticulum		
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
	ko:K04452	DDIT3, GADD153; DNA damage-inducible transcript 3	C up
	ko:K14019	PPP1R15A, GADD34; protein phosphatase 1 regulatory subunit 15A	C up
	ko:K09490	HSPA5, BIP; heat shock 70kDa protein 5	C up
	ko:K11863	ATXN3, MJD; Ataxin-3 [EC:3.4.22.-]	C up
	ko:K14027	HERPUD1, HERP; homocysteine-responsive endoplasmic reticulum-resident ubiquitin-like domain member 1 protein	C up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
ko04120	Ubiquitin mediated proteolysis		
	ko:K10605	BRCA1; breast cancer type 1 susceptibility protein	C up
ko03440	Homologous recombination		
	ko:K10865	MRE11; double-strand break repair protein MRE11	C up
	ko:K08775	BRCA2, FANCD1; breast cancer 2 susceptibility protein	C up
ko03450	Non-homologous end-joining		
	ko:K10865	MRE11; double-strand break repair protein MRE11	C up
	ko:K10886	XRCC4; DNA-repair protein XRCC4	C up
ko03460	Fanconi anemia pathway		
	ko:K10605	BRCA1; breast cancer type 1 susceptibility protein	C up
	ko:K08775	BRCA2, FANCD1; breast cancer 2 susceptibility protein	C up
Environmental Information Processing			
Membrane transport			
ko02000	Transporters		
ko02010	ABC transporters		
	ko:K05675	ABCD1, ALD; ATP-binding cassette, subfamily D (ALD), member 1	T up
	ko:K05679	ABCG1; ATP-binding cassette, subfamily G (WHITE), member 1	T up
	ko:K05680	ABCG4; ATP-binding cassette, subfamily G (WHITE), member 4	T up
Signal transduction			
ko04014	Ras signaling pathway		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K12361	RASGRP2; RAS guanyl-releasing protein 2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K05466	ANGPT2; angiopoietin 2	C up

	ko:K17635	RGL1, RGL; ral guanine nucleotide dissociation stimulator-like 1	T up
	ko:K04349	RASGRF1; Ras-specific guanine nucleotide-releasing factor 1	C up
	ko:K16859	PGF; placenta growth factor	T up
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
ko04015	Rap1 signaling pathway		
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K12361	RASGRP2; RAS guanyl-releasing protein 2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K05466	ANGPT2; angiopoietin 2	C up
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
	ko:K16859	PGF; placenta growth factor	T up
ko04010	MAPK signaling pathway		
	ko:K04434	MAPK8IP1, JIP1; mitogen-activated protein kinase 8 interacting protein 1	T up
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
	ko:K04415	MAP3K8, COT; mitogen-activated protein kinase kinase kinase 8 [EC:2.7.11.25]	T up
	ko:K04349	RASGRF1; Ras-specific guanine nucleotide-releasing factor 1	C up
	ko:K12361	RASGRP2; RAS guanyl-releasing protein 2	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
	ko:K04452	DDIT3, GADD153; DNA damage-inducible transcript 3	C up
	ko:K04854	CACNA1G; voltage-dependent calcium channel T type alpha-1G	T up
ko04012	ErbB signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04310	Wnt signaling pathway		
	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
	ko:K03213	NKD; naked cuticle	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K10162	BAMBI; BMP and activin membrane-bound inhibitor	T up
ko04330	Notch signaling pathway		
	ko:K06058	DTX; deltex	T up

	ko:K05948	FNG; fringe [EC:2.4.1.222]	T up
	ko:K06051	DLL; delta	T up
	ko:K02599	NOTCH; Notch	T up
ko04340	Hedgehog signaling pathway		
	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K16798	GLI2; zinc finger protein GLI2	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
ko04350	TGF-beta signaling pathway		
	ko:K04657	CHRD; chordin	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
	ko:K04677	SMAD6_7; mothers against decapentaplegic homolog 6/7	T up
	ko:K10162	BAMBI; BMP and activin membrane-bound inhibitor	T up
ko04390	Hippo signaling pathway		
	ko:K16798	GLI2; zinc finger protein GLI2	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K16819	AMOT; angiominin	T up
	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
ko04391	Hippo signaling pathway -fly		
	ko:K09851	RASSF2_4; Ras association domain-containing protein 2/4	T up
	ko:K16669	FAT4; protocadherin Fat 4	T up
ko04370	VEGF signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04718	SPHK; sphingosine kinase [EC:2.7.1.91]	T up
	ko:K08273	SH2D2A, VRAP; SH2 domain protein 2A	C up
ko04630	Jak-STAT signaling pathway		
	ko:K05134	IL10RA; interleukin 10 receptor alpha	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K11222	STAT4; signal transducer and activator of transcription 4	T up
	ko:K05138	IL22RA1; interleukin 22 receptor alpha 1	T up
	ko:K05433	IL15; interleukin 15	T up
ko04064	NF-kappa B signaling pathway		
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko04668	TNF signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K05141	TNFRSF1B, TNFR2; tumor necrosis factor receptor superfamily member 1B	T up

	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K04415	MAP3K8, COT; mitogen-activated protein kinase kinase kinase 8 [EC:2.7.11.25]	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K05433	IL15; interleukin 15	T up
ko04066	HIF-1 signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K16451	TIMP1; metalloproteinase inhibitor 1	T up
	ko:K05466	ANGPT2; angiopoietin 2	C up
ko04068	FoxO signaling pathway		
	ko:K08861	PLK2; polo-like kinase 2 [EC:2.7.11.21]	C up
	ko:K08863	PLK4; polo-like kinase 4 [EC:2.7.11.21]	C up
	ko:K13302	SGK1; serum/glucocorticoid-regulated kinase 1 [EC:2.7.11.1]	T up
	ko:K17845	KLF2; krueppel-like factor 2	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K06623	CDKN2D, P19, INK4D; cyclin-dependent kinase inhibitor 2D	T up
	ko:K08336	ATG12; ubiquitin-like protein ATG12	C up
ko04020	Calcium signaling pathway		
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
	ko:K04854	CACNA1G; voltage-dependent calcium channel T type alpha-1G	T up
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K04718	SPHK; sphingosine kinase [EC:2.7.1.91]	T up
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
ko04070	Phosphatidylinositol signaling system		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04151	PI3K-Akt signaling pathway		
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K16859	PGF; placenta growth factor	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K05466	ANGPT2; angiopoietin 2	C up
	ko:K10605	BRCA1; breast cancer type 1 susceptibility protein	C up
	ko:K13302	SGK1; serum/glucocorticoid-regulated kinase 1 [EC:2.7.11.1]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up

	ko:K06250	SPP1, BNSP, OPN; secreted phosphoprotein 1	T up
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K06591	ITGB8; integrin beta 8	C up
ko04150	mTOR signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04080	Neuroactive ligand-receptor interaction		
	ko:K04153	HTR1; 5-hydroxytryptamine receptor 1	T up
	ko:K04132	CHRM4; muscarinic acetylcholine receptor M4	T up
	ko:K04139	ADRA2B; adrenergic receptor alpha-2B	T up
	ko:K04309	LGR4, GPR48; leucine-rich repeat-containing G protein-coupled receptor 4	T up
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
	ko:K04295	S1PR5, EDG8; sphingosine 1-phosphate receptor 5	T up
	ko:K05205	GRIK5; glutamate receptor, ionotropic kainate 5	T up
	ko:K05206	GRID1; glutamate receptor delta-1 subunit	T up
	ko:K04607	GRM4; metabotropic glutamate receptor 4	T up
	ko:K05184	GABRD; gamma-aminobutyric acid receptor subunit delta	T up
	ko:K08362	NR1A2, THRB; thyroid hormone receptor beta	T up
	ko:K04265	ADORA1; adenosine A1 receptor	T up
ko04060	Cytokine-cytokine receptor interaction		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K05433	IL15; interleukin 15	T up
	ko:K05155	TNFRSF19, TROY; tumor necrosis factor receptor superfamily member 19	T up
	ko:K05141	TNFRSF1B, TNFR2; tumor necrosis factor receptor superfamily member 1B	T up
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K04175	IL8RA, CXCR1; interleukin 8 receptor alpha	T up
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up
	ko:K05134	IL10RA; interleukin 10 receptor alpha	T up
	ko:K05138	IL22RA1; interleukin 22 receptor alpha 1	T up
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
	ko:K05146	TNFRSF9, CD137; tumor necrosis factor receptor superfamily member 9	T up
	ko:K05480	EDA; ectodysplasin-A	T up

ko04052	Cytokines		
ko04512	ECM-receptor interaction		
	ko:K06250	SPP1, BNSP, OPN; secreted phosphoprotein 1	T up
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06267	HMMR, RHAMM; hyaluronan-mediated motility receptor	C up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
ko04514	Cell adhesion molecules (CAMs)		
	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K06759	CNTN1; contactin 1	T up
	ko:K16360	LRRC4B, NGL3; netrin-G3 ligand	T up
	ko:K06550	L1CAM; L1 cell adhesion molecule	T up
	ko:K16359	NTNG2; netrin-G2	T up
Cellular Processes			
Transport and catabolism			
ko04144	Endocytosis		
	ko:K13649	FOLR; folate receptor	T up
	ko:K04677	SMAD6_7; mothers against decapentaplegic homolog 6/7	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K05126	RET; proto-oncogene tyrosine-protein kinase Ret [EC:2.7.10.1]	T up
	ko:K04175	IL8RA, CXCR1; interleukin 8 receptor alpha	T up
	ko:K12958	CAV2; caveolin 2	C up
	ko:K06278	CAV1; caveolin 1	C up
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up
ko04147	Exosome		
ko04145	Phagosome		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
	ko:K03990	C3; complement component 3	T up
	ko:K07375	TUBB; tubulin beta	T up
	ko:K07375	TUBB; tubulin beta	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K06560	MRC; mannose receptor, C type	T up
	ko:K07898	RAB7B; Ras-related protein Rab-7B	T up
ko04142	Lysosome		
	ko:K12350	SMPD1, ASM; sphingomyelin phosphodiesterase [EC:3.1.4.12]	T up
	ko:K12383	GM2A; ganglioside GM2 activator	T up
	ko:K12397	AP3B; AP-3 complex subunit beta	T up
	ko:K12394	AP1S1_2; AP-1 complex subunit sigma 1/2	C up
ko04146	Peroxisome		
	ko:K05675	ABCD1, ALD; ATP-binding cassette, subfamily D (ALD), member 1	T up

	ko:K13339	PEX6, PXAAA1; peroxin-6	T up
ko04140	Regulation of autophagy		
	ko:K08336	ATG12; ubiquitin-like protein ATG12	C up
Cell motility			
ko04810	Regulation of actin cytoskeleton		
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K04132	CHRM4; muscarinic acetylcholine receptor M4	T up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
Cell growth and death			
ko04110	Cell cycle		
	ko:K06671	STAG1_2, SCC3, IRR1; cohesin complex subunit SA-1/2	C up
	ko:K06623	CDKN2D, P19, INK4D; cyclin-dependent kinase inhibitor 2D	T up
	ko:K06622	CDKN2C, P18, INK4C; cyclin-dependent kinase inhibitor 2C	T up
	ko:K06669	SMC3, CSPG6; structural maintenance of chromosome 3 (chondroitin sulfate proteoglycan 6)	C up
	ko:K06644	SFN; stratifin	T up
ko04114	Oocyte meiosis		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K06669	SMC3, CSPG6; structural maintenance of chromosome 3 (chondroitin sulfate proteoglycan 6)	C up
ko04210	Apoptosis		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04115	p53 signaling pathway		
	ko:K06509	KAI1, CD82, TSPAN27; CD82 antigen	T up
	ko:K10131	PMAIP1; phorbol-12-myristate-13-acetate-induced protein 1	C up
	ko:K10141	SESN; sestrin	T up
	ko:K06644	SFN; stratifin	T up
Cell communication			
ko04510	Focal adhesion		
	ko:K06250	SPP1, BNSP, OPN; secreted phosphoprotein 1	T up
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
	ko:K12958	CAV2; caveolin 2	C up
	ko:K06278	CAV1; caveolin 1	C up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up

	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K04349	RASGRF1; Ras-specific guanine nucleotide-releasing factor 1	C up
	ko:K16859	PGF; placenta growth factor	T up
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
ko04520	Adherens junction		
	ko:K06086	SORBS1, SH3D5, PONSIN, CAP; sorbin and SH3 domain containing 1	T up
ko04530	Tight junction		
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
ko04540	Gap junction		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K07375	TUBB; tubulin beta	T up
	ko:K07375	TUBB; tubulin beta	T up
Organismal Systems			
Immune system			
ko04640	Hematopoietic cell lineage		
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K04012	CR2, CD21; complement receptor type 2	T up
ko04610	Complement and coagulation cascades		
	ko:K03913	SERPINA5, PCI; protein C inhibitor	T up
	ko:K04012	CR2, CD21; complement receptor type 2	T up
	ko:K03909	TFPI; tissue factor pathway inhibitor	C up
	ko:K03990	C3; complement component 3	T up
	ko:K01314	F10; coagulation factor X [EC:3.4.21.6]	T up
	ko:K03984	SERPINA1, AAT; alpha-1-antitrypsin	T up
ko04611	Platelet activation		
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K01832	TBXAS1, CYP5A; thromboxane-A synthase [EC:5.3.99.5]	T up
	ko:K00509	PTGS1, COX1; prostaglandin-endoperoxide synthase 1 [EC:1.14.99.1]	T up
	ko:K12361	RASGRP2; RAS guanyl-releasing protein 2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up

	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
ko04620	Toll-like receptor signaling pathway		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04415	MAP3K8, COT; mitogen-activated protein kinase kinase kinase 8 [EC:2.7.11.25]	T up
	ko:K06250	SPP1, BNSP, OPN; secreted phosphoprotein 1	T up
ko04621	NOD-like receptor signaling pathway		
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko04622	RIG-I-like receptor signaling pathway		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K08336	ATG12; ubiquitin-like protein ATG12	C up
ko04623	Cytosolic DNA-sensing pathway		
ko04650	Natural killer cell mediated cytotoxicity		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K07987	RAET1; retinoic acid early transcript 1	T up
ko04612	Antigen processing and presentation		
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
ko04660	T cell receptor signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K04415	MAP3K8, COT; mitogen-activated protein kinase kinase kinase 8 [EC:2.7.11.25]	T up
ko04662	B cell receptor signaling pathway		
	ko:K04012	CR2, CD21; complement receptor type 2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04664	Fc epsilon RI signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04666	Fc gamma R-mediated phagocytosis		
	ko:K01080	PPAP2; phosphatidate phosphatase [EC:3.1.3.4]	T up
	ko:K04718	SPHK; sphingosine kinase [EC:2.7.1.91]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04670	Leukocyte transendothelial migration		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up

	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
ko04672	Intestinal immune network for IgA production		
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up
	ko:K05433	IL15; interleukin 15	T up
ko04062	Chemokine signaling pathway		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K12361	RASGRP2; RAS guanyl-releasing protein 2	T up
	ko:K12365	PREX1; phosphatidylinositol-3,4,5-trisphosphate-dependent Rac exchanger 1 protein	T up
	ko:K04175	IL8RA, CXCR1; interleukin 8 receptor alpha	T up
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
Endocrine system			
ko04911	Insulin secretion		
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K04944	KCNN3; potassium intermediate/small conductance calcium-activated channel subfamily N member 3	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K16882	PCLO; protein piccolo	T up
ko04910	Insulin signaling pathway		
	ko:K07192	FLOT; flotillin	C up
	ko:K06086	SORBS1, SH3D5, PONSIN, CAP; sorbin and SH3 domain containing 1	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04920	Adipocytokine signaling pathway		
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K08765	CPT1; carnitine O-palmitoyltransferase 1 [EC:2.3.1.21]	T up
	ko:K05141	TNFRSF1B, TNFR2; tumor necrosis factor receptor superfamily member 1B	T up
ko03320	PPAR signaling pathway		
	ko:K06086	SORBS1, SH3D5, PONSIN, CAP; sorbin and SH3 domain containing 1	T up
	ko:K08765	CPT1; carnitine O-palmitoyltransferase 1 [EC:2.3.1.21]	T up
	ko:K08755	FABP6; fatty acid-binding protein 6, ileal (gastrotropin)	T up
ko04912	GnRH signaling pathway		

	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko04913	Ovarian Steroidogenesis		
	ko:K07418	CYP2J; cytochrome P450, family 2, subfamily J [EC:1.14.14.1]	T up
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1 [EC:1.14.14.1]	T up
	ko:K04119	AKR1C3; aldo-keto reductase family 1 member C3 [EC:1.1.1.64 1.1.1.188 1.1.1.213]	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko04915	Estrogen signaling pathway		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
ko04914	Progesterone-mediated oocyte maturation		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04917	Prolactin signaling pathway		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04921	Oxytocin signaling pathway		
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
	ko:K05005	KCNJ12; potassium inwardly-rectifying channel subfamily J member 12	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04918	Thyroid hormone synthesis		
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K09490	HSPA5, BIP; heat shock 70kDa protein 5	C up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
ko04919	Thyroid hormone signaling pathway		
	ko:K08362	NR1A2, THRB; thyroid hormone receptor beta	T up
	ko:K02599	NOTCH; Notch	T up
	ko:K17903	RCAN2, ZAKI4; calcipressin-2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up

	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04916	Melanogenesis		
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
ko04614	Renin-angiotensin system		
Circulatory system			
ko04260	Cardiac muscle contraction		
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
ko04261	Adrenergic signaling in cardiomyocytes		
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
	ko:K04845	SCN1B; voltage-gated sodium channel type I beta	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04270	Vascular smooth muscle contraction		
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
Digestive system			
ko04970	Salivary secretion		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko04971	Gastric acid secretion		
	ko:K00907	MYLK; myosin-light-chain kinase [EC:2.7.11.18]	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko04972	Pancreatic secretion		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K07884	RAB3D; Ras-related protein Rab-3D	T up
ko04976	Bile secretion		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko04973	Carbohydrate digestion and absorption		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up

ko04974	Protein digestion and absorption		
	ko:K13781	SLC7A8, LAT2; solute carrier family 7 (L-type amino acid transporter), member 8	T up
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K14207	SLC38A2, SNAT2; solute carrier family 38 (sodium-coupled neutral amino acid transporter), member 2	C up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
ko04975	Fat digestion and absorption		
	ko:K11160	DGAT2; diacylglycerol O-acyltransferase 2 [EC:2.3.1.20 2.3.1.75]	T up
	ko:K01080	PPAP2; phosphatidate phosphatase [EC:3.1.3.4]	T up
ko04977	Vitamin digestion and absorption		
ko04978	Mineral absorption		
Excretory system			
ko04962	Vasopressin-regulated water reabsorption		
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
ko04960	Aldosterone-regulated sodium reabsorption		
	ko:K13302	SGK1; serum/glucocorticoid-regulated kinase 1 [EC:2.7.11.1]	T up
	ko:K08555	NR3C2, MR; mineralocorticoid receptor	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04961	Endocrine and other factor-regulated calcium reabsorption		
ko04964	Proximal tubule bicarbonate reclamation		
ko04966	Collecting duct acid secretion		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
Nervous system			
ko04724	Glutamatergic synapse		
	ko:K14207	SLC38A2, SNAT2; solute carrier family 38 (sodium-coupled neutral amino acid transporter), member 2	C up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K15009	SHANK; SH3 and multiple ankyrin repeat domains protein	T up
	ko:K05205	GRIK5; glutamate receptor, ionotropic kainate 5	T up
	ko:K04607	GRM4; metabotropic glutamate receptor 4	T up
ko04727	GABAergic synapse		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K05184	GABRD; gamma-aminobutyric acid receptor subunit delta	T up
	ko:K04647	HAP1; huntingtin-associated protein 1	T up
	ko:K14207	SLC38A2, SNAT2; solute carrier family 38 (sodium-coupled neutral amino acid transporter), member 2	C up

ko04725	Cholinergic synapse		
	ko:K01049	ACHE; acetylcholinesterase [EC:3.1.1.7]	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K05005	KCNJ12; potassium inwardly-rectifying channel subfamily J member 12	T up
	ko:K04132	CHRM4; muscarinic acetylcholine receptor M4	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
ko04728	Dopaminergic synapse		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
ko04726	Serotonergic synapse		
	ko:K07418	CYP2J; cytochrome P450, family 2, subfamily J [EC:1.14.14.1]	T up
	ko:K00509	PTGS1, COX1; prostaglandin-endoperoxide synthase 1 [EC:1.14.99.1]	T up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K04153	HTR1; 5-hydroxytryptamine receptor 1	T up
ko04720	Long-term potentiation		
ko04730	Long-term depression		
ko04723	Retrograde endocannabinoid signaling		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K05184	GABRD; gamma-aminobutyric acid receptor subunit delta	T up
	ko:K13700	ABHD6; abhydrolase domain-containing protein 6 [EC:3.1.1.23]	T up
ko04721	Synaptic vesicle cycle		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
	ko:K15294	CPLX1_2; complexin-1/2	T up
ko04722	Neurotrophin signaling pathway		
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up

Sensory system			
ko04744	Phototransduction		
	ko:K13765	RGS9; regulator of G-protein signaling 9	T up
ko04740	Olfactory transduction		
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
ko04742	Taste transduction		
	ko:K04607	GRM4; metabotropic glutamate receptor 4	T up
ko04750	Inflammatory mediator regulation of TRP channels		
	ko:K04971	TRPV2; transient receptor potential cation channel subfamily V member 2	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K07418	CYP2J; cytochrome P450, family 2, subfamily J [EC:1.14.14.1]	T up
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
Development			
ko04320	Dorso-ventral axis formation		
	ko:K02599	NOTCH; Notch	T up
ko04360	Axon guidance		
	ko:K06820	PLXNA; plexin A	T up
	ko:K16359	NTNG2; netrin-G2	T up
	ko:K06838	SLIT1; slit 1	T up
	ko:K06842	SEMA6; semaphorin 6	T up
	ko:K06842	SEMA6; semaphorin 6	T up
	ko:K06550	L1CAM; L1 cell adhesion molecule	T up
	ko:K05114	EPHB6; Eph receptor B6 [EC:2.7.10.1]	T up
	ko:K06521	SEMA4; semaphorin 4	T up
	ko:K06843	NTN1; netrin 1	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K05462	EFNA; ephrin-A	T up
	ko:K04189	CXCR4; C-X-C chemokine receptor type 4	T up
	ko:K07529	DPYSL5, CRAM; dihydropyrimidinase-like 5	T up
	ko:K06840	SEMA3; semaphorin 3	T up
	ko:K06840	SEMA3; semaphorin 3	T up
	ko:K05112	EPHB3, HEK2, ETK2; Eph receptor B3 [EC:2.7.10.1]	T up
ko04380	Osteoclast differentiation		
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko04713	Circadian entrainment		
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up

	ko:K04854	CACNA1G; voltage-dependent calcium channel T type alpha-1G	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
ko04626	Plant-pathogen interaction		
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
Human Diseases			
Cancers			
ko05200	Pathways in cancer		
	ko:K00572	WNT7; wntless-type MMTV integration site family, member 7	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K08775	BRCA2, FANCD1; breast cancer 2 susceptibility protein	C up
	ko:K00445	WNT6; wntless-type MMTV integration site family, member 6	T up
	ko:K01384	WNT11; wntless-type MMTV integration site family, member 11	T up
	ko:K16859	PGF; placenta growth factor	T up
	ko:K05126	RET; proto-oncogene tyrosine-protein kinase Ret [EC:2.7.10.1]	T up
	ko:K09348	NKX3-1; homeobox protein Nkx-3.1	T up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K16798	GLI2; zinc finger protein GLI2	T up
	ko:K04514	Rock1; Rho-associated protein kinase 1 [EC:2.7.11.1]	T up
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
ko05202	Transcriptional misregulation in cancers		
	ko:K09355	PBX1; pre-B-cell leukemia transcription factor 1	T up
	ko:K15187	MLLT1_3, ENL, AF9; YEATS domain-containing protein 1/3	T up
	ko:K04452	DDIT3, GADD153; DNA damage-inducible transcript 3	C up
	ko:K02583	NGFR; nerve growth factor receptor (TNFR superfamily member 16)	T up
	ko:K06622	CDKN2C, P18, INK4C; cyclin-dependent kinase inhibitor 2C	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko05206	MicroRNAs in cancer		
	ko:K16866	TIMP3; metalloproteinase inhibitor 3	T up
	ko:K17460	BMF; Bcl-2-modifying factor	T up
	ko:K16865	PDCD4; programmed cell death protein 4	C up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
	ko:K10605	BRCA1; breast cancer type 1 susceptibility protein	C up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K02599	NOTCH; Notch	T up
ko05205	Proteoglycans in cancer		
	ko:K01384	WNT11; wntless-type MMTV integration site family, member 11	T up
	ko:K00445	WNT6; wntless-type MMTV integration site family, member 6	T up

	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K16865	PDCD4; programmed cell death protein 4	C up
	ko:K06278	CAV1; caveolin 1	C up
	ko:K12958	CAV2; caveolin 2	C up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K16866	TIMP3; metalloproteinase inhibitor 3	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
ko05204	Chemical carcinogenesis		
	ko:K00799	GST, gst; glutathione S-transferase [EC:2.5.1.18]	T up
	ko:K07408	CYP1A1; cytochrome P450, family 1, subfamily A, polypeptide 1 [EC:1.14.14.1]	T up
ko05203	Viral carcinogenesis		
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K03990	C3; complement component 3	T up
	ko:K10131	PMAIP1; phorbol-12-myristate-13-acetate-induced protein 1	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05210	Colorectal cancer		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05212	Pancreatic cancer		
	ko:K08775	BRCA2, FANCD1; breast cancer 2 susceptibility protein	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05214	Glioma		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05216	Thyroid cancer		
	ko:K05126	RET; proto-oncogene tyrosine-protein kinase Ret [EC:2.7.10.1]	T up
ko05221	Acute myeloid leukemia		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05220	Chronic myeloid leukemia		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05217	Basal cell carcinoma		
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K16798	GLI2; zinc finger protein GLI2	T up

	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
ko05218	Melanoma		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05211	Renal cell carcinoma		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05219	Bladder cancer		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K04445	MSK1, RPS6KA5; ribosomal protein S6 kinase alpha-5 [EC:2.7.11.1]	C up
ko05215	Prostate cancer		
	ko:K09487	HSP90B, TRA1; heat shock protein 90kDa beta	C up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K09348	NKX3-1; homeobox protein Nkx-3.1	T up
ko05213	Endometrial cancer		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05222	Small cell lung cancer		
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
ko05223	Non-small cell lung cancer		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
Immune diseases			
ko05322	Systemic lupus erythematosus		
	ko:K03990	C3; complement component 3	T up
ko05323	Rheumatoid arthritis		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K05453	CSF1, MCSF; macrophage colony-stimulating factor 1	T up
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
	ko:K05433	IL15; interleukin 15	T up
ko05320	Autoimmune thyroid disease		
ko05321	Inflammatory bowel disease (IBD)		
	ko:K11222	STAT4; signal transducer and activator of transcription 4	T up
Neurodegenerative diseases			
ko05010	Alzheimer's disease		

	ko:K04550	LRP1, CD91; low-density lipoprotein receptor-related protein 1 (alpha-2-macroglobulin receptor)	T up
ko05012	Parkinson's disease		
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko05014	Amyotrophic lateral sclerosis (ALS)		
	ko:K05141	TNFRSF1B, TNFR2; tumor necrosis factor receptor superfamily member 1B	T up
ko05016	Huntington's disease		
	ko:K04647	HAP1; huntingtin-associated protein 1	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K10408	DNAH; dynein heavy chain, axonemal	C up
ko05020	Prion diseases		
	ko:K09490	HSPA5, BIP; heat shock 70kDa protein 5	C up
Substance dependence			
ko05030	Cocaine addiction		
	ko:K13765	RGS9; regulator of G-protein signaling 9	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
ko05031	Amphetamine addiction		
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
ko05032	Morphine addiction		
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K04265	ADORA1; adenosine A1 receptor	T up
	ko:K05184	GABRD; gamma-aminobutyric acid receptor subunit delta	T up
ko05033	Nicotine addiction		
	ko:K05184	GABRD; gamma-aminobutyric acid receptor subunit delta	T up
ko05034	Alcoholism		
	ko:K04266	ADORA2A, ADOR; adenosine receptor A2a	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K04543	GNG7; guanine nucleotide-binding protein G(I)/G(S)/G(O) subunit gamma-7	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
Cardiovascular diseases			
ko05410	Hypertrophic cardiomyopathy (HCM)		

	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K07610	DES; desmin	T up
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
ko05412	Arrhythmogenic right ventricular cardiomyopathy (ARVC)		
	ko:K06591	ITGB8; integrin beta 8	C up
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K07610	DES; desmin	T up
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
ko05414	Dilated cardiomyopathy (DCM)		
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K07610	DES; desmin	T up
	ko:K04859	CACNA2D2; voltage-dependent calcium channel alpha-2/delta-2	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K10351	MYL2; myosin regulatory light chain 2	C up
	ko:K06591	ITGB8; integrin beta 8	C up
ko05416	Viral myocarditis		
	ko:K06278	CAV1; caveolin 1	C up
Endocrine and metabolic diseases			
ko04930	Type II diabetes mellitus		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K04854	CACNA1G; voltage-dependent calcium channel T type alpha-1G	T up
ko04950	Maturity onset diabetes of the young		
	ko:K08027	NR5A2, FTF; nuclear receptor subfamily 5 group A member 2	T up
ko04932	Non-alcoholic fatty liver disease (NAFLD)		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K04452	DDIT3, GADD153; DNA damage-inducible transcript 3	C up
Infectious diseases			
ko05110	Vibrio cholerae infection		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
ko05120	Epithelial cell signaling in Helicobacter pylori infection		
	ko:K02148	ATPeV1C, ATP6C; V-type H ⁺ -transporting ATPase subunit C	T up
	ko:K04175	IL8RA, CXCR1; interleukin 8 receptor alpha	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko05130	Pathogenic Escherichia coli infection		
	ko:K07375	TUBB; tubulin beta	T up
	ko:K07375	TUBB; tubulin beta	T up
ko05132	Salmonella infection		
	ko:K07898	RAB7B; Ras-related protein Rab-7B	T up

	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko05131	Shigellosis		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K06484	ITGA5; integrin alpha 5	T up
ko05133	Pertussis		
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K03990	C3; complement component 3	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko05134	Legionellosis		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K03990	C3; complement component 3	T up
ko05150	Staphylococcus aureus infection		
	ko:K03990	C3; complement component 3	T up
ko05152	Tuberculosis		
	ko:K04718	SPHK; sphingosine kinase [EC:2.7.1.91]	T up
	ko:K03990	C3; complement component 3	T up
	ko:K06560	MRC; mannose receptor, C type	T up
	ko:K05134	IL10RA; interleukin 10 receptor alpha	T up
	ko:K10049	CEBPG; CCAAT/enhancer binding protein (C/EBP), gamma	C up
ko05100	Bacterial invasion of epithelial cells		
	ko:K06484	ITGA5; integrin alpha 5	T up
	ko:K16938	SEPT3_9_12; septin 3/9/12	T up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06278	CAV1; caveolin 1	C up
	ko:K12958	CAV2; caveolin 2	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05166	HTLV-I infection		
	ko:K09341	MSX; homeobox protein MSX	T up
	ko:K09341	MSX; homeobox protein MSX	T up
	ko:K05433	IL15; interleukin 15	T up
	ko:K08045	ADCY5; adenylate cyclase 5 [EC:4.6.1.1]	T up
	ko:K00572	WNT7; wingless-type MMTV integration site family, member 7	T up
	ko:K00445	WNT6; wingless-type MMTV integration site family, member 6	T up
	ko:K01384	WNT11; wingless-type MMTV integration site family, member 11	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02354	FZD4, fz4; frizzled 4	T up
	ko:K06622	CDKN2C, P18, INK4C; cyclin-dependent kinase inhibitor 2C	T up
ko05162	Measles		
	ko:K18052	PRKCQ; novel protein kinase C theta type [EC:2.7.11.13]	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up

	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
ko05164	Influenza A		
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K01349	FURIN, PCSK3; furin [EC:3.4.21.75]	T up
	ko:K15046	NS1BP; influenza virus NS1A-binding protein	C up
ko05161	Hepatitis B		
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K09048	CREB3; cyclic AMP-responsive element-binding protein 3	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K11222	STAT4; signal transducer and activator of transcription 4	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
ko05160	Hepatitis C		
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K03250	EIF3E, INT6; translation initiation factor 3 subunit E	C up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K14217	IFIT1; interferon-induced protein with tetratricopeptide repeats 1	T up
ko05168	Herpes simplex infection		
	ko:K05433	IL15; interleukin 15	T up
	ko:K14217	IFIT1; interferon-induced protein with tetratricopeptide repeats 1	T up
	ko:K03990	C3; complement component 3	T up
ko05169	Epstein-Barr virus infection		
	ko:K05134	IL10RA; interleukin 10 receptor alpha	T up
	ko:K04012	CR2, CD21; complement receptor type 2	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
ko05146	Amoebiasis		
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K05717	FN1; fibronectin 1	T up
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K06236	COL1A5; collagen, type I/II/III/V/XI/XXIV/XXVII, alpha	T up
	ko:K07898	RAB7B; Ras-related protein Rab-7B	T up
	ko:K06237	COL4A; collagen, type IV, alpha	T up
ko05144	Malaria		

	ko:K10030	IL8, CXCL8; interleukin 8	T up
	ko:K04550	LRP1, CD91; low-density lipoprotein receptor-related protein 1 (alpha-2-macroglobulin receptor)	T up
ko05145	Toxoplasmosis		
	ko:K06240	LAMA3_5; laminin, alpha 3/5	T up
	ko:K05134	IL10RA; interleukin 10 receptor alpha	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K03283	HSPA1_8; heat shock 70kDa protein 1/8	T up
ko05140	Leishmaniasis		
	ko:K03990	C3; complement component 3	T up
ko05142	Chagas disease (American trypanosomiasis)		
	ko:K03990	C3; complement component 3	T up
	ko:K04633	GNAL; guanine nucleotide-binding protein G(olf) subunit alpha	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K02649	PIK3R; phosphoinositide-3-kinase, regulatory subunit	T up
	ko:K10030	IL8, CXCL8; interleukin 8	T up

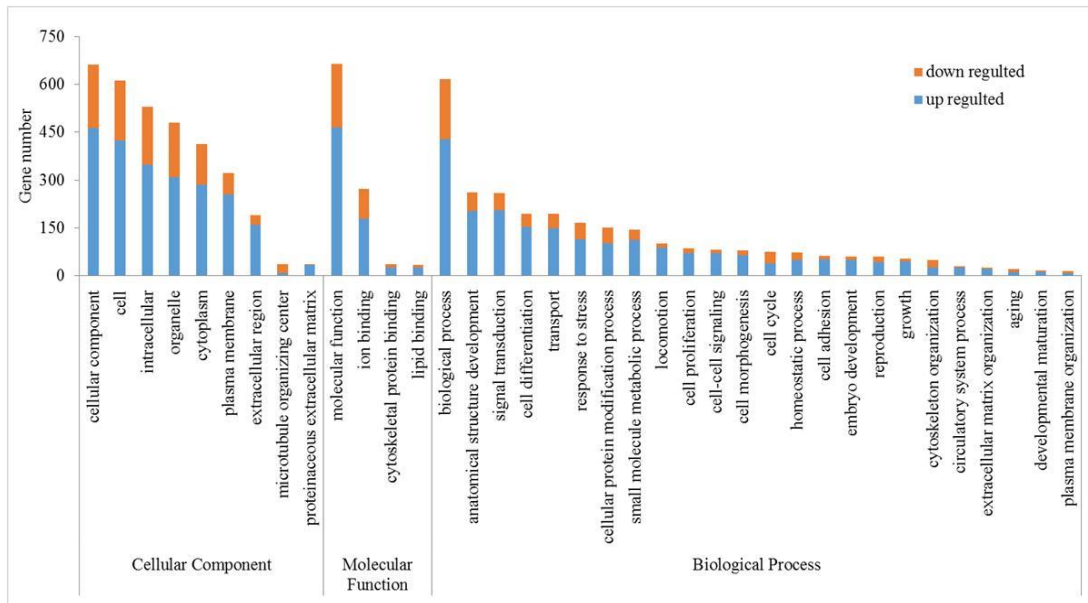


Fig. S1. GO annotation of the differentially expressed genes. The genes are enriched in different terms in three functional categories.

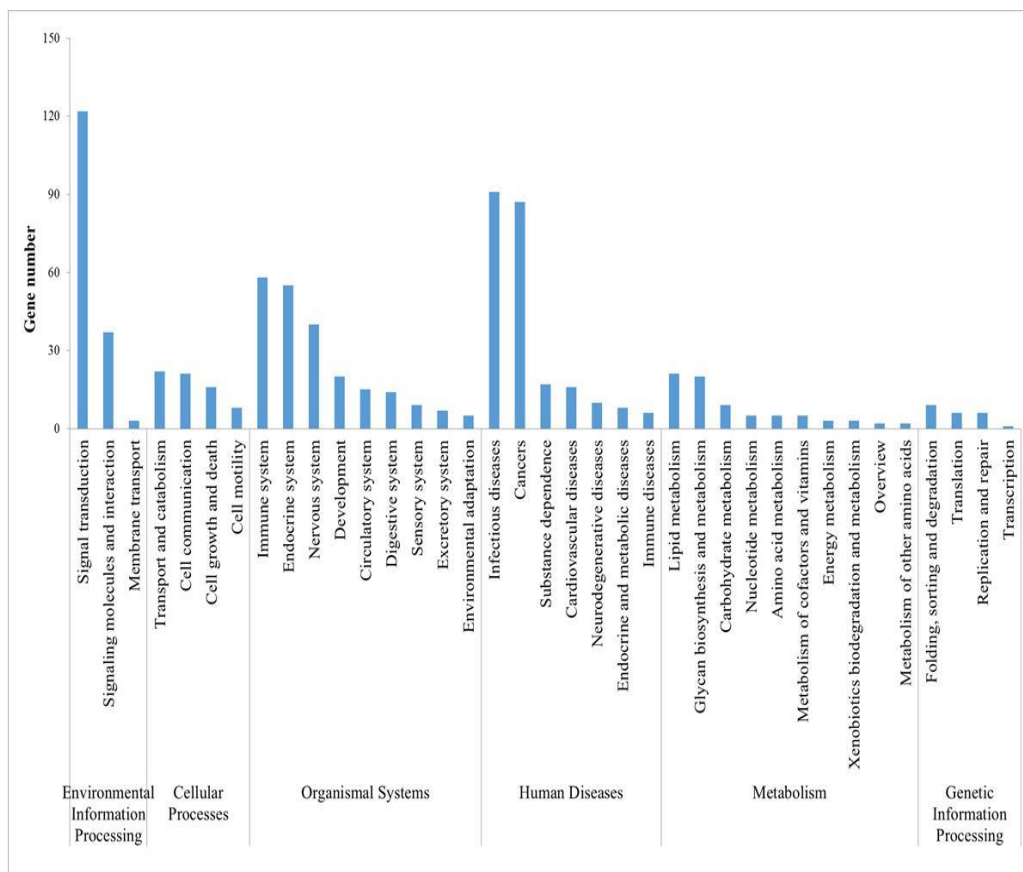


Fig. S2. KEGG annotation of the differentially expressed genes. The genes are enriched in different pathways in different functional groups.

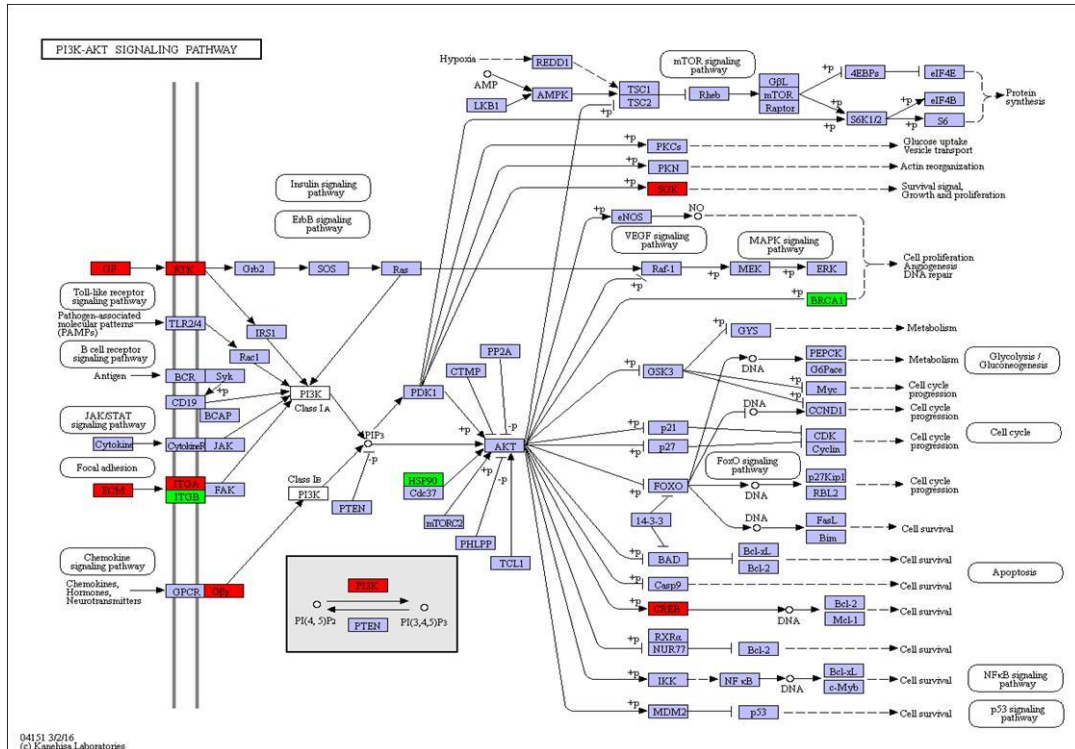


Fig. S3. KEGG analysis reveals that AKT pathway is activated by high glucose, insulin and palmitic acid.

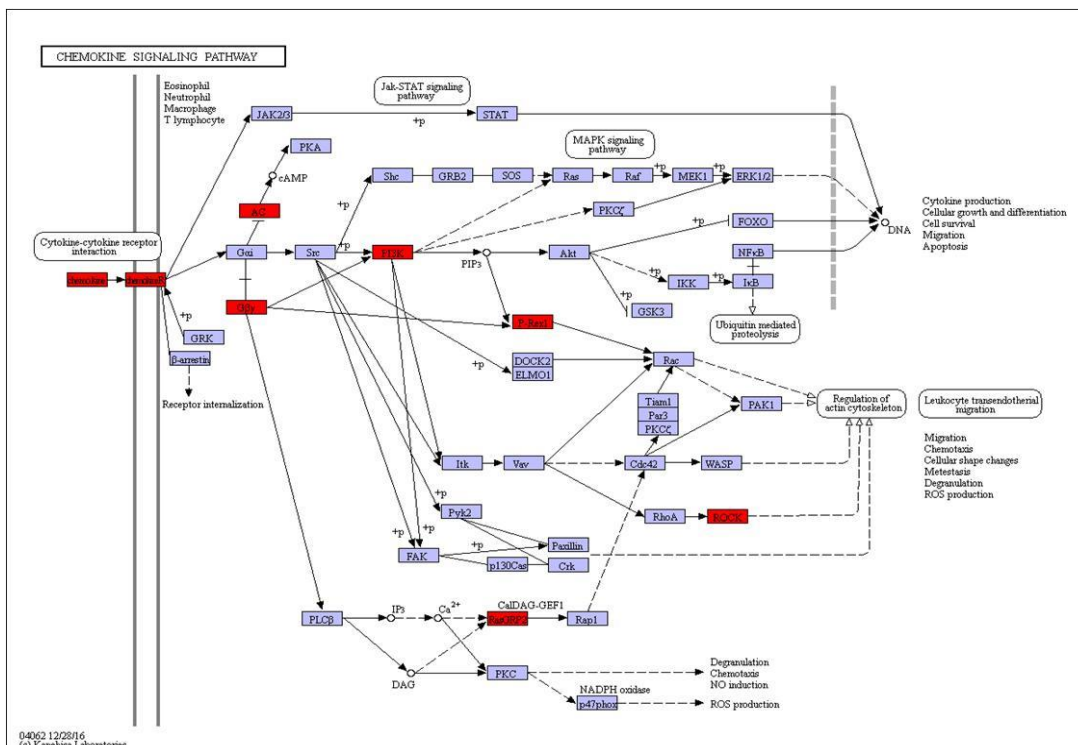


Fig. S4. KEGG analysis reveals that ROCK pathway is activated by high glucose, insulin and palmitic acid.

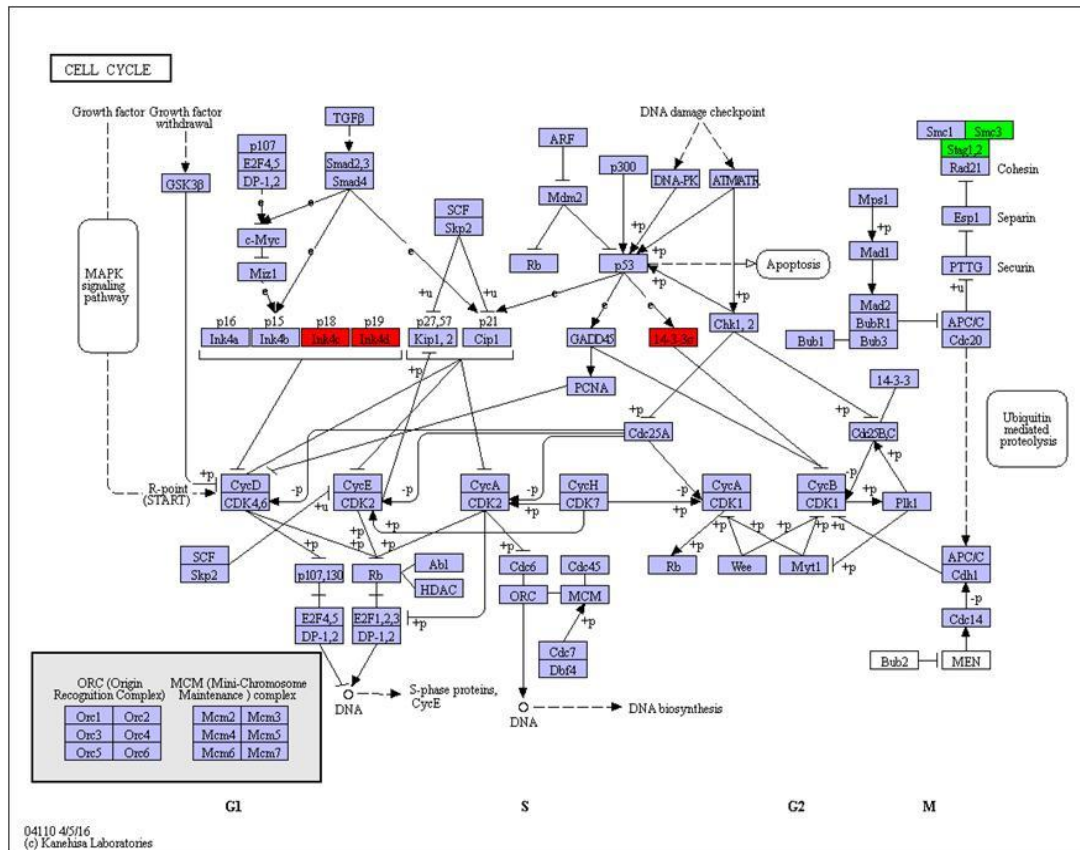


Fig. S5. KEGG analysis reveals that 14-3-3 σ pathway is activated by high glucose, insulin and palmitic acid.