TITLE

Biomarker evaluation of plasma microRNA-122, high-mobility group box-1 and keratin-18 in acute gallstone disease.

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ABSTRACT (244 words)

Background: A biomarker that stratifies patients with complications of gallstone disease from the denominator pool of people with acute biliary-type symptoms is needed. Circulating microRNA-122 (miRNA-122), high-mobility group box1 (HMGB1), full-length keratin-18 (flk-18) and caspase-cleaved keratin-18 (cck-18) are established hepatocyte injury biomarkers. The aim of this study was to evaluate the discriminatory power of these biomarkers in plasma to identify potential biliary complications that may require acute intervention.

Method: An observational biomarker cohort study was carried out in a University teaching hospital for 12 months beginning 3rd September 2014. Blood samples were collected from adults referred with acute biliary-type symptoms. miRNA-122 was measured by quantitative real-time polymerase chain reaction, and HMGB1, cck-18 and flk-18 by ELISA.

Results: 300 patients were screened and 289 patients were included. Plasma miRNA-122, cck-18 and flk-18 concentrations were increased in patients with gallstones compared to those without (*miRNA-122*: median: 2.89×10^4 copies/ml vs. 0.95×10^4 copies/ml [$P < 0.001$]; *cck-18*: 121.9 U/L vs. 104.6 U/L [$P = 0.041$]; *flk-18*: 252.4 U/L vs. 151.8 U/L [$P < 0.001$]). Uncomplicated gallstone disease was associated with higher miRNA-122 and cck-18 concentrations than complicated disease (*miRNA-122*: 5.72×10^4 copies/ml vs. 2.26×10^4 copies/ml [$P = 0.022$]; *cck-18*: 139.7 U/L vs. 111.4 U/L [$P = 0.049$]). There was no significant difference in HMGB1 concentration between patients with and without gallstones [$P = 0.480$]. Separation between groups for all biomarkers was modest.

Conclusion: microRNA-122 and keratin-18 plasma concentrations are elevated in patients with gallstones. However, these biomarkers were not sufficiently discriminatory to be progressed as clinically useful biomarkers in this context.

INTRODUCTION

Gallstone disease is one of the most common reasons for persons to present to hospital with gastrointestinal disease in Western countries¹. In the United Kingdom, there are approximately 330 people per 100,000 adult population admitted with an acute biliary disease annually². A point-of-care diagnostic test that can stratify patients with acute biliary-type symptoms according to their need for further investigation or intervention will be advantageous to patients and has the potential to reduce healthcare expenditure. Individuals with uncomplicated biliary colic and aseptic choledocholithiasis may be safely discharged without the need for inpatient management and can be managed in an outpatient or day-case setting. Conversely, patients with complications, for example cholecystitis, cholangitis and gallstone pancreatitis, are best managed as inpatients according to current national standards.

Standard available clinico-pathological data for patients presenting acutely with gallstones in the emergency setting include clinical history and examination, routine laboratory results, and ultrasound (USS) or magnetic resonance imaging (MRI) findings³. Currently, this set of investigations cannot always identify whether patients have complications of gallstones that necessitate admission. Therefore, a biomarker panel that could be used as an adjunct to existing investigations to guide treatment decisions would be extremely advantageous.

microRNA-122 (miRNA-122), high mobility group box 1 (HMGB1), full-length keratin-18 (flk-18) and caspase-cleaved keratin-18 (cck-18) have been identified as early acute hepatic injury biomarkers at presentation to hospital, particularly in the context of paracetamol overdose^{4,5}. miRNAs are single-stranded, small non-protein-coding ribonucleic acids (RNAs). miRNA-122 is expressed near exclusively in the hepatocyte and provides enhanced hepatic specificity over all current liver injury biomarkers⁶. HMGB1 is a nuclear

binding protein with pro-inflammatory activity⁷ that is released by immune and necrotic cells^{7,8}. Keratins are part of the intermediate filament system and they play an important role in cellular mechanisms. They are a hallmark of epithelial cells including hepatocytes and biliary cells^{9,10}. During apoptosis, full-length keratin-18 (flk-18) is cleaved by caspases to produce the fragment caspase-cleaved keratin-18 (cck-18)¹¹. flk-18 is released into blood during necrosis and cck-18 during apoptosis¹². Studies in pre-clinical models have demonstrated increased circulating miRNA-122, HMGB1 and keratin-18 following bile duct ligation¹³. In humans with cholecystitis, keratin-18 has been reported to be elevated in both bile and serum samples, when compared to a control group of healthy volunteers¹⁴. Shifeng *et al.*⁶ reported that circulating miRNA-122 has a high diagnostic value with regard to distinguishing patients with biliary calculi from normal controls. In that study, the area under the receiver operator curve (ROC-AUC) was 0.93, which suggests high accuracy. However, the controls were healthy subjects, whereas in clinical practice, the relevant comparison group will be patients with other aetiologies of abdominal pain. In the present study, we recruited a cohort of patients with acute abdominal pain.

The aim of this study was to evaluate whether circulating hepatic injury biomarkers discriminate between people with and without gallstone disease and uncomplicated from complicated gallstone disease during the first 24 hours of hospital admission.

METHODS

Ethics

This study was assessed by South East Scotland Research Ethics Service of National Health Service (NHS) Lothian and was given ethical approval under the terms of the Governance Arrangements for Research Ethics Committees (Harmonised Edition). Lothian R&D project number 2014/0224; REC number 14/EM/0211.

Study Design and Setting

This study was an observational cohort biomarker study conducted in the Emergency Department and Emergency General Surgery Department at the Royal Infirmary of Edinburgh, NHS Lothian, U.K.

Inclusion and Exclusion Criteria

All adult patients referred to the on-call General Surgeons with acute abdominal pain, irrespective of the characteristics of the pain, and with a differential diagnosis of gallstone disease upon index presentation were eligible for inclusion. Patients must have had presented to hospital within 24 hours prior to recruitment. Patients who were unable to give informed consent, or who refused to participate in the study, or who were younger than 16 years of age or transferred from a hospital outside the NHS regional trust (NHS Lothian) were excluded from the study.

Study Population, sample size and data collection

This proof-of-concept research study was designed to inform a power calculation for a multi-centre qualification study, and therefore the sample size was pragmatic. This study had been approved to recruit a total of 300 patients over a period of two years.

Recruitment was for 52 weeks: 3rd September 2014 to 2nd September 2015. Potential participants were identified at the on-call General Surgery team's daily handover. All patients recruited to the study had given informed consent. Clinical details, imaging reports, operation findings and pathology reports were obtained from hand written and electronic medical notes. Each patient was then followed up through the electronic record for a minimum of three months. Whole blood was sampled within 24 hours of hospital presentation into EDTA vacutainer tubes (S-Monovette, Sarstedt), centrifuged at 1000xg for 15 minutes at 4°C and the plasma layer retained. The supernatant was then separated into aliquots and stored at -80°C until analysis.

Variables and Definitions

Clinical data included patient demographics, routine laboratory blood and microbiology results, imaging investigations, and/or information on surgical interventions and pathology results. The final diagnoses of the patients were determined by histopathology results, operative findings, imaging investigations and/or discharge scripts, following that hierarchy. Patients with a diagnosis of non-specific abdominal pain (NSAP) had normal investigation results and a definitive cause of their abdominal pain was not found. Patients with gallstone diseases were grouped into 'Uncomplicated' and 'Complicated' categories: patients with 'uncomplicated' disease had either biliary colic (cholelithiasis in gallbladder) or aseptic choledocholithiasis (choledocholithiasis without clinical, laboratory or microbiological evidence of sepsis); patients with 'complicated' gallstone disease included those with cholangitis (septic choledocholithiasis), cholecystitis or gallstone pancreatitis.

Total RNA Extraction, Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR) and enzyme-linked immunosorbent assay (ELISA) Analysis

Laboratory analyses of all biomarkers were carried out blinded to participant clinical

data. Analysis of miRNA-122 was carried out in the Centre for Cardiovascular Sciences, University of Edinburgh); HMGB1, flk-18 and cck-18 laboratory analyses were carried out at the MRC Centre for Drug Safety Science, University of Liverpool.

miRNA was extracted from plasma using the miRNeasy Serum/Plasma Kit (Qiagen, Venlo, Netherlands) according to the manufacturer's protocol^{15,16}. Synthetic miR-39 (at 1.6×10^8 copies/ μ L) was spiked in as an internal control. miRNAs were measured with Taqman-based qPCR. Small RNA elutes were reverse transcribed using specific stem-loop reverse-transcription RT primers (Applied Biosystems, Foster City, CA, USA) for each target miRNA species, following the manufacturer's instructions. Specific stem loop rt primer targeting UGGAGUGUGACAAUGGUGUUUG and 5' UCACCGGGUGUAAAUCAGCUUG 3' was used. No template controls (NTCs) were included to test for miRNA contamination. The expression of miR-39 and miRNA-122 were analysed using the standard $2^{-\Delta\Delta C_t}$ method¹⁷ normalized to the miR-39 spiked internal control.

Plasma HMGB1 and keratin-18 were determined by ELISA according to the manufacturer's guidelines (Shino-Test/IBL International^{18,19} for HMGB1; and PEVIVA²⁰ for cck-18 and flk-18) and our previously published protocols^{5,21}.

Statistical analysis

Continuous variables are presented as medians and interquartile ranges (IQR). Categorical variables are presented as frequencies. The Pearson Chi-squared test was used to examine the association between categorical variables. The Kolmogorov-Smirnov test was used to compare the distribution of non-parametric data against the normal distribution. The Kruskal-Wallis test was used to compare non-parametric variables followed by Dunn's *post hoc* test for multiple comparisons. 95% confidence intervals (CI) were calculated for all normally distributed continuous variables. Positive predictive value (PPV) and negative

predictive value (NPV) were utilized as the most relevant measures of clinical utility. Receiver-operating characteristic (ROC) curve analyses were plotted for the different patient groups. Areas under the ROC curves were calculated with 95% CI. Sensitivity, PPV and NPV were obtained at 90% specificity. All statistical tests were based on a two-sided α -value of 0.05. Statistical analysis was performed using IBM SPSS Statistics Version 19.0 (IBM Corp., Armonk, NY, USA), and G*Power3.1 (Universität Düsseldorf, Germany). Figures were designed, and correlation analyses were carried out using GraphPad Prism Version 6.0 (GraphPad Software, Inc., La Jolla, CA, USA).

RESULTS

Study Population

A total of 300 patients were screened for possible inclusion in the study. The study CONSORT flow-chart is presented in **Figure 1**. Eleven patients who had consented to participate in the study were excluded: two were transferred to a different hospital prior obtaining blood samples; one was under 16 years old; five were admitted more than 24 hours prior to recruitment; two patients' blood samples were collected in the wrong sample tube; one blood sample tube did not have the required volume for biomarker testing. Of the 289 patients included in the study, as their final diagnosis, 47 (16.3%) patients had non-specific abdominal pain (NSAP), 64 patients (22.1%) had uncomplicated gallstone disease, 115 patients (39.8%) had a complicated gallstone disease and 63 patients (21.8%) had other non-gallstone related disease.

The demographics of the study subjects are illustrated in **Table 1**. There were more females compared to males between the four patient groups ($P < 0.001$). Patients with uncomplicated gallstone diseases (46.4 years [I29.8-61.8]) were significantly younger than patients with complicated diseases (56.6 years [44.6-68.5]; $P = 0.002$).

Among those who underwent a cholecystectomy ($n = 276$, 44.6%), there was no difference in the time period from patient's index hospital presentation to the day of operation between patient groups, even between those with (median: 3.0 days [2.0-4.8]) and without (median: 4.0 days [2.0-7.5]) complications of gallstones ($P = 0.12$). Interestingly, there was no statistical difference between the two gallstone disease subgroups (uncomplicated: $n = 11$, 17.2% vs. complicated: $n = 18$, 15.7%; $P = 0.790$) with regard to CT investigation, but there was a significantly different proportion of patients with complicated gallstone disease ($n = 43$, 37.4%) who underwent MRCP compared to those with an uncomplicated disease ($n = 34$, 53.1%) ($P = 0.041$) (**Supplementary Table 1**).

Serum ALT, AlkP and GGT activities were higher in patients with uncomplicated gallstone diseases compared to complicated diseases (**Table 1**).

HMGB1

There was no significant difference in plasma HMGB1 concentration between patients with and without gallstone disease (*gallstone diseases*: 2.37 ng/mL [1.40-3.53]; *non-gallstones*: 2.31 ng/mL [1.22-3.60]; $P=0.480$) or between the four different study groups ($P=0.584$) (**Table 2** and **Supplementary Figure 1**).

miRNA-122

miRNA-122 in gallstone diseases and non-gallstone diseases

Circulating miRNA-122 concentration was higher in patients with gallstone disease than in those without gallstones (2.89×10^4 copies/ml [0.88-9.94 $\times 10^4$] vs. 0.95×10^4 copies/ml [0.41-3.30 $\times 10^4$]; $P<0.001$) (**Figure 2A**). ROC analysis (**Figure 2B**) of miRNA-122 for patients with and without gallstone disease produced an AUC of 0.66. The calculated PPV and NPV at 90% specificity were 72.5% and 39.8% respectively (**Table 3**).

miRNA-122 in uncomplicated and complicated gallstone diseases

Patients with uncomplicated gallstone disease had a significantly higher miRNA-122 concentration than those with complicated gallstone disease (5.72×10^4 copies/ml [0.97-16.85 $\times 10^4$] vs. 2.26×10^4 copies/ml [0.75-7.86 $\times 10^4$]; $P=0.022$) (**Table 2** and **Figure 2C**). ROC analysis (**Figure 2D**) comparing uncomplicated and complicated gallstone diseases yielded an AUC of 0.60 (**Table 3**) (**Supplementary Figure 2** and **Supplementary Table 2** and **5** illustrate the gallstone disease subgroups).

cck-18

cck-18 in gallstone diseases and non-gallstone diseases

Plasma cck-18 concentration was significantly higher in patients with gallstone disease than those without (121.96 U/L [89.65-184.17]; vs. 104.57 U/L [84.23-150.55]; $P=0.041$) (**Figure 3A**). A ROC curve analysis of cck-18 in patients with and without gallstones yielded an AUC of 0.57 (**Figure 3B**), with a calculated PPV of 50.0% and NPV of 37.1% (**Table 3**).

cck-18 in uncomplicated and complicated gallstone diseases

cck-18 concentration was higher in patients with uncomplicated (139.71 U/L [98.15-228.71]) gallstone diseases than those with complications (111.35 U/L [87.17-171.83]) $P=0.049$) (**Table 2** and **Figure 3C**). This translated to an AUC of 0.59 (**Figure 3D**) with PPV of 45.0% and NPV of 65.4% at 90% specificity (**Table 3**) (**Supplementary Figure 3** and **Supplementary Table 3** and **5** illustrate gallstone disease subgroups).

flk-18

flk-18 in gallstone diseases and non-gallstone diseases

Similarly, flk-18 concentration was significantly higher in patients with gallstone disease than those without (252.38 U/L [136.63-525.67]; vs. 151.84 U/L [81.96-330.30]; $P<0.001$) (**Figure 4A**). A ROC curve analysis (**Figure 4B**) of flk-18 in patients with and without gallstones yielded an AUC of 0.63 (**Table 3**).

flk-18 in uncomplicated and complicated gallstone diseases

There was no significant difference in flk-18 concentration between the two subgroups of gallstone diseases ($P=0.160$) (**Table 2** and **Figure 4C**) (**Supplementary Figure 4** and **Supplementary Table 4** and **5** illustrate gallstone disease subgroups).

DISCUSSION

Early identification of individuals with potentially serious complications of gallstones from the denominator pool of all persons presenting with biliary-type symptoms is of translational importance. In this study, we report that plasma miRNA-122, cck-18 and flk-18 concentrations were significantly higher in patients with gallstone diseases than in non-gallstone diseases. However, the degree of overlap between groups was high, thus detracting from the utility of this approach to accurately stratify on an individual patient basis.

The biomarkers measured in this paper have robust evidence supporting their utility in preclinical models and the clinical setting for the early, sensitive and specific identification of hepatocellular injury following paracetamol overdose (an archetypal hepatocyte toxic agent) and also hypoxic hepatitis. Published papers have suggested this panel of biomarkers may also have utility with regard to acute cholestatic pathology induced by gall stones.

Our finding that plasma miRNA-122 concentration was increased in patients with gallstones agrees with that of Shifeng *et al.*⁶, who compared their disease cohort with ‘healthy’ control groups. However, their study yielded a much higher AUC on the ROC analysis, 0.93 with 77.4% sensitivity and 96.4%. The probable explanation for this is that our comparison group was patients admitted acutely under general surgery with a non-gallstone related diagnosis, who therefore may have had deranged inflammatory markers and/or LFTs due to another pathological process. This demonstrates the importance of clinically relevant control groups in biomarker discovery and validation studies.

In the present study, there was no significant difference in plasma HMGB1 concentration across all comparisons. This is consistent with a study by Shi *et al.*⁷ which demonstrated HMGB1 over expression in gallbladder cancers but not in benign GB tissues or cholelithiasis.

Plasma cck-18 and flk-18 were significantly higher in patients with gallstones, in

agreement with studies by others^{14,22} who examined gallbladder tissue, bile and serum. In our study, patients with cholecystitis were not further subdivided into active/inactive and acute/chronic subtypes due to sample sizing. Nevertheless, Simopoulos *et al.*¹⁴ demonstrated no difference in total k-18 and cck-18 between active and inactive chronic cholecystitis.

We recognize some important limitations in this study. This was a pilot study to determine whether there is a signal of biomarker utility in this context of use to take forward into larger, multi-centre studies. We believe that the results presented in this paper are clear in their failure to support utility with regard to patient stratification at hospital presentation. We did not proceed to perform multi-variable analysis given the near total overlap between the primary comparisons. Serial patient blood sampling for biomarker analyses may have also been useful to determine the true nature of biomarker concentration changes during the disease progress.

In conclusion, microRNA-122 and cleaved cytokeratin-18 plasma concentrations are elevated in patients with common bile duct complications of gallstones but are not sufficiently discriminatory to be progressed as clinically useful biomarkers in this context of use. Their clinical utility is in the context of acute hepatocyte injury.

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Figure 1

Figure 1: CONSORT diagram for the main study. The number of patients who underwent cholecystectomy within each group is illustrated in brackets with an asterisk). GS, gallstones; GB, gall bladder; CBD, common bile duct; NSAP, non-specific abdominal pain; UGI, upper gastrointestinal; HPB, hepatopancreaticobiliary; RCC, renal cell cancer; UTI, urinary tract infection; LRTI, lower respiratory tract infection; MSK, musculoskeletal; SMV, superior mesenteric vein; WCC, white blood cell count; CRP, C-reactive protein; LFT, liver function test.

Figure 2

Figure 2A: Plasma miRNA-122 values in gallstone and non-gallstone diseases normalised to miRNA-39. Each data point represents an individual. The horizontal line in each patient group represents the median value. Statistical significances (P value) by Kruskal Wallis test and Dunn's multiple comparison test is shown in the figure for between group comparison. **B.** Receiver Operating Characteristic (ROC) curve of miRNA-122 for gallstone versus non-gallstone diseases. **C.** miRNA-122 values for the four main patient groups. **D.** ROC curve of miRNA-122 values for uncomplicated versus complicated gallstone diseases.

Figure 3

Figure 3A: Plasma concentrations of cck-18 protein in gallstone and non-gallstone diseases. Each data point represents an individual. The horizontal line in each patient group represents the median value. Statistical significances (P value) Kruskal Wallis test and Dunn's multiple comparison test is shown in the figure for between group comparison. **B.** ROC curve of cck-18 for gallstone versus non-gallstone diseases. **C.** cck-18 values for the four main patient groups. **D.** ROC curve of cck-18 values for uncomplicated versus complicated gallstone diseases.

Figure 4

Figure 4A: Plasma flk-18 protein concentrations in gallstone and non-gallstone diseases. Each data point represents an individual. The horizontal line in each patient group represents the median value. Statistical significances (P value) by Kruskal Wallis test and Dunn's multiple comparison test is shown in the figure for between group comparison. **B.** ROC curve of flk-18 for gallstone versus non-gallstone diseases. **C.** flk-18 concentrations for the four main patient groups. **D.** ROC curve of flk-18 values for uncomplicated versus complicated gallstone diseases.

Table 1. Demographics of the study cohort and routine blood test results.

	Overall	NSAP ¹	Uncomplicated ²	Complicated ³	Others	P-value
N (%)	289 (100)	47 (16.3)	64 (22.1)	115 (39.8)	63 (21.8)	
Gender (n, (%))						
Male	88 (30.4)	3 (6.4)	9 (14.1)	53 (46.1)	23 (36.5)	<0.001
Female	201 (69.6)	44 (93.6)	55 (85.9)	62 (53.9)	40 (63.5)	
Age (Years)						
Median	51.3	41.3	46.4	56.6	52.2	<0.001
IQR	37.5-67.4	26.7-57.3	29.8-61.8	44.6-68.5	39.3-67.5	
95% CI	49.3,53.5	37.9,49.4	42.7,51.7	53.1,59.4	48.2,56.6	
Underwent Operation (n, (%))						
Yes	276 (44.6)	2 (4.3)	37 (57.8)	88 (76.5)	2 (3.2)	<0.001
No	160 (55.4)	45 (95.7)	27 (42.2)	27 (23.5)	61 (96.8)	
Duration from index presentation to Operation (Days)						
Median	3.0	1.5	4.0	3.0	24.0	0.067
IQR	2.0-5.5	1.0- -	2.0-7.5	2.0-4.8	5.0- -	
95% CI	7.6,17.6	-4.9,7.9	4.1,25.2	5.8,17.6	-217.4,265.4	
Total bilirubin (umol/L)						
Median	14.0	11.0	22.5	17.0	11.0	<0.001
IQR	8.0-31.5	7.0-15.0	10.3-46.8	9.0-35.0	7.0-33.0	
95% CI	24.3,32.7	10.2,14.2	25.9,44.1	23.3,35.6	19.8,44.6	
Alanine aminotransferase, ALT (U/L)						
Median	36.0	23.0	135.0	38.0	41.0	<0.001
IQR	19.0-169.5	14.0-29.0	23.5-359.0	20.0-178.0	18.0-100.0	
95% CI	119.2,174.1	20.9,32.2	175.7,336.5	106.6,185.2	71.4,181.5	
Alkaline Phosphatases, AlkP (U/L)						
Median	96.0	76.0	139.5	94.0	94.0	<0.001
IQR	71.0-172.5	60.0-103.0	86.0-264.0	70.0-158.0	74.0-238.0	
95% CI	137.9,172.0	75.2,101.3	153.1,223.0	121.7,170.5	135.8,238.7	
Gamma-glutamyl transpeptidase, GGT (U/L)						
Median	87.0	25.0	238.0	114.5	74.0	<0.001
IQR	30.0-389.5	14.0-54.0	55.0-457.5	39.3-392.5	30.0-497.8	
95% CI	211.4,312.7	24.9,54.5	236.5,422.1	178.9,358.3	187.4,437.3	

¹ Non-Specific Abdominal Pain

² Biliary colic, Aseptic Choledocholithiasis

³ Cholecystitis, Cholangitis, Gallstone Pancreatitis

⁴ Pearson Chi-Square test (For 'Gender' and 'Underwent Operation')

⁵ Kruskal-Wallis test and Dunn's multiple comparison test (For 'Age', 'Duration from index presentation to Operation', 'Total bilirubin', 'ALT', 'AlkP' and 'GGT')

Table 2. Biomarker concentrations.

	Overall	NSAP ¹	Uncomplicated ²	Complicated ³	Others	P-value ⁴
miRNA-122 (x10 ⁴ copies/ml)						
Median	1.81	0.90	5.72	2.26	1.13	
IQR	0.54-7.0	0.41-2.23	0.97-16.85	0.75-7.86	0.42-4.42	<0.001
95% CI	6.21,16.40	4.32,26.42	9.51,22.24	1.54,22.82	2.26,8.23	
HMGB1 (ng/ml)						
Median	2.34	2.08	2.60	2.29	2.70	
IQR	1.34-3.55	1.26-3.50	1.30-4.67	1.44-3.37	1.18-4.16	0.584
95% CI	2.13,2.60	1.50,2.59	2.05,3.01	2.02,2.56	1.90,3.10	
cck-18 (U/L)						
Median	114.60	100.33	139.71	111.35	114.84	
IQR	87.40-171.53	75.05-135.40	98.15-228.71	87.17-171.83	87.12-181.26	0.009
95% CI	105.02,123.67	87.23,112.47	121.82,168.56	100.31,134.24	95.27,125.78	
flk-18 (U/L)						
Median	202.99	107.08	322.55	236.34	205.54	
IQR	102.63-450.55	66.39-168.43	133.43-662.04	136.63-429.38	101.81-462.56	<0.001
95% CI	170.60,237.24	78.90,149.70	202.10, 479.32	184.91,304.06	151.78,325.69	

¹ Non-Specific Abdominal Pain

² Biliary colic, Aseptic Choledocholithiasis

³ Cholecystitis, Cholangitis, Gallstone Pancreatitis

⁴ Kruskal-Wallis test and Dunn's multiple comparison test

Table 3. Biomarker evaluation by receiver-operator characteristics.

	ROC AUC (95% CI)	Sensitivity (%) (95% CI)	PPV (%) ¹	NPV (%) ²	P-value ³
miRNA-122					
Gallstone vs. Non-gallstone ⁴	0.66(0.59-0.72)	16(10-25)	72.5	39.8	<0.001
Uncomplicated ⁵ vs. Complicated ⁶	0.60(0.51-0.69)	22(15-30)	56.0	67.5	0.022
cck-18					
Gallstone vs. Non-gallstone ⁴	0.57(0.50-0.64)	6(3-13)	50.0	7.1	0.041
Uncomplicated ⁵ vs. Complicated ⁶	0.59(0.50-0.68)	14(8-22)	45.0	65.4	0.049
flk-18					
Gallstone vs. Non-gallstone ⁴	0.63(0.56-0.70)	20(13-29)	76.6	40.9	<0.001
Uncomplicated ⁵ vs. Complicated ⁶	0.56(0.47-0.65)	12(7-20)	42.1	65.0	0.160

¹ Positive predictive value

² Negative predictive value

³ Kruskal-Wallis test and Dunn's multiple comparison test

⁴ NSAP and Other diseases

⁵ Biliary colic, Aseptic choledocholithiasis

⁶ Cholecystitis, Cholangitis, Gallstone pancreatitis

REFERENCES

1. Portincasa P, Moschetta A, Palasciano G. Cholesterol gallstone disease. *The Lancet*. 2006 Jul 21; 368(9531): 230-239.
2. NHS Digital. Hospital Episode Statistics, Admitted Patient Care – England, 2011-12 [NS]. Published 2012 Nov 1 [Assessed 15th June 2017]. Available from: <http://content.digital.nhs.uk/searchcatalogue?productid=9161&q=gallbladder&infotype=0%2fOfficial+statistics&sort=Relevance&size=50&page=1#top>.
3. Borzellino G, Steccanella F, Mantovani W, Genna M. Predictive factors for the diagnosis of severe acute cholecystitis in an emergency setting. *Surgical Endoscopy*. 2013 Sep 1; 27(9): 3388-3395.
4. Dear JW, Antoine DJ, Starkey-Lewis P, Goldring CE, Park BK. Early detection of paracetamol toxicity using circulating liver microRNA and markers of cell necrosis. *British Journal of Clinical Pharmacology*. 2014 May 1; 77(5): 904-995.
5. Antoine DJ, Jenkins RE, Dear JW, Williams DP, McGill MR, Sharpe MR, Craig DG, Simpson KJ, Jaeschke H, Park BK. Molecular forms of HMGB1 and keratin-18 as mechanistic biomarkers for mode of cell death and prognosis during clinical acetaminophen hepatotoxicity. *Journal of Hepatology*. 2012 May 31; 56(5):1070-1079.
6. Shifeng H, Danni W, Pu C, Ping Y, Ju C, Liping Z. Circulating liver-specific miR-122 as a novel potential biomarker for diagnosis of cholestatic liver injury. *PloS One*. 2013 Sep 27; 8(9): e73133.
7. Shi Z, Huang Q, Chen J, Yu P, Wang X, Qiu H, Chen Y, Dong Y. Correlation of HMGB1 expression to progression and poor prognosis of adenocarcinoma and squamous cell/adenosquamous carcinoma of gallbladder. *American Journal of Translational Research*. 2015; 7(10).

8. Antoine DJ, Williams DP, Kipar A, Jenkins RE, Regan SL, Sathish JG, Kitteringham NR, Park BK. High-mobility group box-1 protein and keratin-18, circulating serum proteins informative of acetaminophen-induced necrosis and apoptosis in vivo. *Toxicological Sciences*. 2009 Sep 25; 112(2): 521-531.
9. Chu PG, Weiss LM. Keratin expression in human tissues and neoplasms. *Histopathology*. 2002 May 1; 40(5): 403-439.
10. Wilkins BJ, Gong W, Pack M. A novel keratin18 promoter that drives reporter gene expression in the intrahepatic and extrahepatic biliary system allows isolation of cell-type specific transcripts from zebrafish liver. *Gene Expression Patterns*. 2014 Mar 31; 14(2):62-68.
11. Caulín C, Salvesen GS, Oshima RG. Caspase cleavage of keratin 18 and reorganization of intermediate filaments during epithelial cell apoptosis. *The Journal of Cell Biology*. 1997 Sep 22; 138(6): 1379-1394.
12. Lorente L, Martín MM, González-Rivero AF, Ferreres J, Solé-Violán J, Labarta L, Díaz C, Jiménez A, Borreguero-León JM. Serum levels of caspase-cleaved cytokeratin-18 and mortality are associated in severe septic patients: pilot study. *PloS One*. 2014 Oct 7; 9(10): e109618.
13. Woolbright BL, Antoine DJ, Jenkins RE, Bajt ML, Park BK, Jaeschke H. Plasma biomarkers of liver injury and inflammation demonstrate a lack of apoptosis during obstructive cholestasis in mice. *Toxicology and Applied Pharmacology*. 2013 Dec 15; 273(3): 524-331.
14. Simopoulos C, Tsaroucha AK, Asimakopoulos B, Giatromanolaki A, Gavriilidis P, Polychronidis A, Karayiannakis A. Total and caspase-cleaved cytokeratin 18 in chronic cholecystitis: A prospective study. *BMC Gastroenterology*. 2008 May 6; 8(1): 14.

15. Qiagen. miRNeasy Serum/Plasma Handbook: miRNeasy Serum/Plasma Kit For purification of total RNA, including miRNA, from animal and human plasma and serum; miRNeasy Serum/Plasma Spike-In Control; For normalization of miRNA purification from serum or plasma. Qiagen: Sample and Assay Technologies. [Online] 2012. Available from:
<http://www.qiagen.com/aw/resources/download.aspx?id=710c0168-e408-408b-95af-91df5b5b1dd6&lang=en> [Assessed on 15th June 2017].
16. Applied Biosystems. TaqMan Small RNA Assays (TaqMan microRNA assays; TaqMan siRNA assays; Custom TaqMan small RNA assays) Protocol. Applied Biosystems by Life Technologies. [Online] 2011. Available from:
https://tools.lifetechnologies.com/content/sfs/manuals/cms_042167.pdf [Assessed on 15th June 2017]
17. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ CT method. Methods. 2001 Dec 1; 25(4): 402-408.
18. SHINO-TEST Corporation. HMGB1 ELISA Kit: Assay Principle. [Online] 2017. Available from: <http://www.hmgb1.net/en/products/elisa.html> [Assessed on 15th June 2017]
19. IBL International. HMGB1 ELISA. [Online] 2011. Available from:
<http://novamedline.com/downloads/instructions/en/ST51011.pdf> [Assessed on 15th June 2017]
20. PEVIVA Products. M65 ELISA. [Online] 2014. Available from:
http://diapharma.com/wp-content/uploads/2017/03/P10020_Peviva_M65_ELISA_2014-12-19_Rev90097_insert.pdf [Assessed on 15th June 2017]

21. Antoine DJ, Dear JW, Lewis PS, Platt V, Coyle J, Masson M, Thanacoody RH, Gray AJ, Webb DJ, Moggs JG, Bateman DN. Mechanistic biomarkers provide early and sensitive detection of acetaminophen-induced acute liver injury at first presentation to hospital. *Hepatology*. 2013 Aug 1; 58(2): 777-787.
22. Kasprzak A, Malkowski W, Seraszek A, Surdyk-Zasada J, Szmeja J, Rogacki K, Kaczmarek E, Zabel M. Cytokeratin 8 and 18 tissue expression in gallbladder mucosa of patients with cholelithiasis. *Polish Journal of Pathology*. 2011 Dec 1; 62(4): 241-249.