Diversity loss selects for altered plant phenotypic responses to local arbuscular mycorrhizal communities

Terhi Hahl[1]§, Sofia J. van Moorsel[1]§, Marc W. Schmid[1,2], Debra Zuppinger-Dingley[1], Bernhard Schmid[1] and Cameron Wagg[1]*

[1]URPP Global Change and Biodiversity and Department of Evolutionary Biology and Environmental Studies, University of Zürich, Winterthurerstrasse 190, CH-8057, Switzerland

[2]MWSchmid GmbH, Möhrlistrasse 25, CH-8006 Zürich

§Equal contribution

*Author for correspondence:

Cameron Wagg, cameron.wagg@ieu.uzh.ch

Summary

1. Biodiversity loss not only impairs ecosystem functioning but can also alter the selection for traits in plant communities. At high diversity selection favours traits that allow for greater niche partitioning, whereas at low diversity selection may favour greater defence against pathogens. However, it is unknown whether changes in plant diversity also select for altered interactions with soil organisms.

2. We assessed whether the responses in plant growth and functional traits to their local arbuscular mycorrhizal fungal (AMF) communities have been altered by the diversity of the plant communities from which both plants and AMF communities were obtained. We grew plants with AMF communities that originated from either plant monocultures or mixtures in a fully factorial design that included both negative and positive controls, by inoculating no AMF or a foreign AMF respectively.

3. We found that AMF from plant mixtures were more beneficial than monoculture AMF for two out of five plant species. Plants from mixtures generally grew better than those from monocultures, but suffered greater damage by leaf pathogens. Although plant growth and phenotypic responses were dependent on the AMF communities with which they associated, we found little evidence for plant growth responses specific to their local AMF communities and results differed between species and traits.

4. Our results show that plants from mixtures were selected for increased growth at the expense of reduced defence and vice versa for plants from monocultures, providing evidence for plant diversity-dependent selection on competitive growth vs. defence. Furthermore, our study suggests that effects of a common history between plants and AMF do not follow a general pattern leading to increased or decreased mutualism.

5. *Synthesis*: Here we provide evidence that biodiversity loss can alter evolutionary trajectories of plant phenotypes and responses to their local AMF communities. However, the

selection for altered plant–AMF interactions differ between plant species. To understand how plant communities respond and evolve under a changing environment requires further knowledge about life strategies of plant species and their above–belowground interactions.

Key-words: biodiversity experiment, functional traits, growth–defence trade-off, mutualism–parasitism continuum**,** plant–AMF interactions, plant–soil feedbacks

1 | Introduction

How reciprocal interactions between plants and soil biota drive plant community structure and ecosystem functioning has been a focal point in ecology (Bever, 1994; Klironomos, 2002; van der Heijden et al., 2006; van der Putten et al., 2013). These plant–soil feedbacks (PSF) describe the relationships between plants and their associated soil pathogens and mutualists, which negatively or positively affect plant performance (Bever et al., 2010). Negative PSF are common and thought to be key to maintaining diversity in plant communities and to successional changes in plant community composition through time (Kardol, Martijn Bezemer, & van der Putten, 2006; Klironomos, 2002; Andrew Kulmatiski, Beard, Stevens, & Cobbold, 2008; Petermann, Fergus, Turnbull, & Schmid, 2008; van der Putten et al., 2013). Positive PSFs are known to improve plant performance by either enhancing pathogen defence (reducing negative PSF) or stimulating plant growth (van der Putten et al., 2013). Plants simultaneously interact with both pathogens and beneficial soil organisms that together determine plant performance. However, negative PSF and positive PSF have rarely been disentangled and only recently studies considered that both negative and positive PSF might change over ecological time-scales (van der Heijden, Bardgett, & van Straalen, 2008; terHorst et al. 2014). Here we assess the consequences of plant diversity loss on interactions between plants and associated communities of soil mutualists. We specifically tested whether changes in plant diversity result in intraspecific differences in plant phenotypic responses to their local arbuscular mycorrhizal fungi (AMF) communities over a short evolutionary time scale (11 years).

Arbuscular mycorrhizal fungi (AMF) are soil-borne fungi, which form symbiotic relationships with most land plants (Harley & Harley, 1987; Smith & Read, 1997; Wang & Qiu, 2006). These fungi are typically known for improving plant growth by providing enhanced soil nutrient uptake in exchange for plant photosynthates (Smith & Read 2008).

Furthermore, AMF diversity has been shown to be an important driver of aboveground plant diversity and ecosystem productivity (Francis & Read, 1994; van der Heijden et al., 1998; Wagg, Jansa, Stadler, Schmid, & van der Heijden., 2011a). Conversely, the composition of AMF communities can be strongly affected by the composition and diversity of plant communities, such that the diversity of plant communities is interlinked with the diversity of AMF communities (Burrows & Pfleger, 2002; Klironomos, McCune, Hart, & Neville, 2000; König et al., 2010; Milcu et al., 2013; Scherber et al., 2010). It is therefore conceivable that a more diverse plant community can sustain a more beneficial AMF community due to a positive effect of plant diversity on supporting a greater AMF diversity. On the other hand, plants growing in monocultures may also select for a more beneficial AMF community, as the greater abundance of a specific plant species could select for AMF that are particularly beneficial to that particular species (Arguello et al., 2016; Kiers et al., 2011; Werner & Kiers, 2015). Although there is evidence indicating that plants growing in monoculture for a prolonged period of time had evolved altered phenotypic responses to competition and positive PSF (Zuppinger-Dingley et al., 2014; Zuppinger-Dingley, Flynn, De Deyn, Petermann, & Schmid, 2016), we currently lack evidence as to whether changes in plant diversity alter plant responses to their local AMF communities over time. We therefore test the hypothesis that AMF communities originating from plant communities differing in diversity (plant monocultures vs. plant mixtures) would differ in their influence on plant growth and phenotypic traits (H1).

Plants can adapt to short-term selection pressure imposed by plant community diversity (van Moorsel, Schmid, Hahl, Zuppinger-Dingley, & Schmid, 2017; Zuppinger-Dingley et al., 2014). This rapid evolution in plant communities can occur via a sorting-out from standing genetic variation (Fakheran et al., 2010). In monocultures, plants may accumulate more specialized pathogens than in diverse plant communities, which potentially

reduces plant productivity in monocultures over time (Kulmatiski, Beard, & Heavilin, 2012; Marquard et al., 2013; Schnitzer et al., 2011; van der Putten et al., 2013). At higher plant diversity levels, however, specialized pathogens may become diluted (Eisenhauer, 2012; Latz et al., 2012), thus conferring lower pathogen-based selection pressure than in monocultures. Consequently, in plant mixtures, plants would likely trade-off reduced defence for increased competitive growth, whereas in monocultures plants would trade-off reduced growth for increased defence (Herms & Mattson, 1992). If plants are differentially selected for pathogen defence or competitive growth traits depending upon their neighbourhood diversity from which they originate (e.g. Zuppinger-Dingley et al. 2014, Zuppinger-Dingley et al. 2016), then we hypothesize that plants with different selection backgrounds exhibit different phenotypic traits related to competition and pathogens (H2).

Furthermore, in plant monocultures under a high pathogen load (Schnitzer et al., 2011), it may be particularly important for plants to associate with AMF as these fungi are known to protect plants against pathogens (Newsham, Fitter, & Watkinson, 1995; Pozo & Azcón-Aguilar, 2007). The increasing positive PSF on monocultures through time (e.g. Zuppinger-Dingley et al. 2016) could thus be potentially reflecting co-selection of plant–AMF relationships. Considering the potential for plant diversity-driven trade-offs between competitive growth and plant defence, it is likely that over time plants and their associated soil biota, such as AMF, may have altered PSF depending on the diversity of the plant community from which they have originated. We therefore finally hypothesize that differences in plant diversity have differentially altered plant phenotypic responses to their local AMF communities resulting in specific ‘home’ vs. ‘away’ plant–AMF community interactions (H3). Home vs. away effects have been used in reciprocal transplant experiments (Joshi et al., 2001) and reciprocal inoculation experiments (Klironomos, 2002; Wagg, Husband, Massicotte, & Peterson, 2011b) to describe a matching of locally co-occurring

121 populations. In the present study, we compared home-combinations (same history for both
122 AMF and plants) with away-combinations (different history for the two partners).

123 To address our three hypotheses, we used five plant species that occurred in plant
124 mixtures and as monocultures within the Jena Biodiversity Experiment (Roscher et al., 2004)
125 and the AMF spore communities collected from their rhizospheres. Plants and AMF had thus
126 been co-occurring in natural field conditions for eleven years (van Moorsel et al., 2018). To
127 specifically address the home and away plant–AMF interactions, we isolated AMF spore
128 communities from the rhizosphere soil and propagated them in trap cultures. Our design was
129 a reciprocal inoculation experiment where plants with a history of occurring in monocultures
130 (monoculture-type plants) or diverse plant mixtures (mixture-type plants) were inoculated
131 with AMF communities isolated from the same monocultures (monoculture AMF) or
132 mixtures (mixture AMF). In addition, we included a negative AMF control treatment (no
133 AMF present) as well as a positive control where plants were inoculated with an external
134 AMF species (*Rhizoglomus irregulare* (Błaszk., Wubet, Renker & Buscot) Sieverd., G.A.
135 Silva & Oehl), which did not share any history with the plants.
136

137 **2 | Materials and methods**
138

139 2.1 | PLANT SELECTION HISTORIES

140 We used five common perennial European grassland species from three functional
141 groups (Roscher et al., 2004): three small herbs (*Plantago lanceolata* L., *Prunella vulgaris* L.
142 and *Veronica chamaedrys* L.), one tall herb (*Galium mollugo* L.) and one legume (*Lathyrus*
143 *pratensis* L.). The plants had a selection history from the Jena Experiment (Roscher et al.,
144 2004), where they were sown in either monoculture or mixture in 2002 (selection history
145 “monoculture-type plants” and “mixture-type plants”, respectively). The species

compositions in the experimental plots in Jena were maintained by weeding three times per year in spring, summer and autumn and by mowing twice per year at peak biomass times in spring and summer. Each of the studied plant species had undergone eleven years of selection from 2002 until 2014 in either plant monocultures (monoculture-type plants) or mixtures (mixture-type plants, Fig. 1). The five plant species are known to associate with arbuscular mycorrhizal fungi (Harley & Harley, 1987).

2.2 | FIRST CONTROLLED SEED PRODUCTION

In spring 2010, entire plant communities of 48 plots (12 monocultures, 12 two-species mixtures, 12 four-species mixtures and 12 eight-species mixtures) of the biodiversity experiment in Jena, Germany (the Jena Experiment) were collected as cuttings. Additionally, the top 30 cm soil of the 48 plots was pooled together, mixed and placed back into the excavated locations at the Jena Experiment. The cuttings were then transplanted to an experimental garden in Zurich, Switzerland, in identical plant composition and seeds were collected over summer 2010. The plots were filled with 30 cm of soil (1:1 mixture of garden compost and agricultural field soil, pH 7.4, Gartenhumus, RICOTER Erdaufbereitung AG, Aarberg, Switzerland), and fenced with netting to minimize cross-pollination with plants outside the plots (for details see Zuppinger-Dingley et al. 2014). In spring 2011, seedlings were propagated from the collected seeds in a glasshouse in Zurich and then transplanted back into the mixed soil in the same plots of the Jena Experiment from where their parents had originally been excavated. In the re-established plots, these newly established plant communities were maintained for three years until 2014 to allow them to become associated with their own microbial communities (van Moorsel et al., 2018).

2.3 | SEED PRODUCTION AND SOIL COLLECTION

In March 2014, entire plant communities from the re-established plots at the Jena Experiment were collected and planted in their respective communities in 1 x 1 m plots in the experimental garden in Zurich. The plots were filled with 30 cm of soil (1:1 mixture of garden compost and agricultural field soil, pH 7.4, Gartenhumus, RICOTER Erdaufbereitung AG, Aarberg, Switzerland), and fenced with netting to minimize cross-pollination. During the relocation of the plant communities, we collected rhizosphere soil samples attached to the roots of the plants (Fig. 1). By then, the soil communities had undergone three years of establishment and eight plus three years of potential co-selection with each of the five plant species in monocultures or mixtures. Seeds were collected from five monocultures and five mixtures (one four-species mixture and four eight-species mixtures). The seeds of the five plant species were stored at +4 ˚C for two months. Four weeks before the start of the experiment, the seeds were surface-sterilized in 7–14 % bleach for 10–45 min and subsequently germinated on 1% water-agar.

2.4 | INOCULUM PREPARATION

We created trap cultures using AMF spore isolation to minimize the potential that other unintended fungi were also propagated. To isolate AMF spore communities from the sampled rhizosphere soils, we passed deionized water and 25 g of soil sample through a series of sieves from 500 µm to 32 µm using a sugar gradient-centrifugation method (Sieverding, 1991). The AMF spores were manually collected with a pipet under a microscope at 200-fold magnification. To bulk up the isolated AMF communities for use as inoculum, we established trap cultures that consisted of 2 L of 4:1 sand-soil mixture, autoclaved at 120 °C for 99 min, and a monoculture of trap plants of one the five tested plant species (Fig. 1). All trap cultures received approximately 400 AMF spores in 30 ml of

deionized water, except for the negative control trap cultures, which received 30 ml of deionized water without AMF spores. For the trap-culture plants we used new seeds from a commercial seed supplier that provided the original seed material for the Jena Experiment (Rieger-Hofmann GmbH, Blaufelden-Raboldshausen, Germany). The seeds were surface-sterilized in 7–14 % bleach for 10–45 min and pre-germinated on 1 % water agar. Each AMF trap culture was replicated twice. After ten months of growth in the glasshouse, we collected root samples from each trap culture that were then fixed in 50 % ethanol, cleared in 10 % KOH, and stained in 5 % ink-vinegar (Vierheilig, Coughlan, Wyss, & Piché, 1998). Colonization by AMF was microscopically assessed for successful mycorrhizal establishment by quantifying AMF root colonization following the transect-intersect method (McGonigle, Miller, Evans, Fairchild, & Swan, 1990). We further quantified the concentration of AMF spores in trap cultures with fungal colonization. We isolated AMF spores from a 10-g soil sample with the same sieving and centrifugation methods used when setting up the AMF trap-culture pots. Trap-plant cultures that exhibited fungal root colonization and spore production were dried and the plants were harvested at ground level. The roots were harvested, cut into 3–5 cm fragments and the belowground content of the trap cultures was used as soil inoculum in the PSF experiment described below.

For the positive control AMF treatment, we used a trap culture substrate containing *Rhizoglomus irregulare* (Błaszk., Wubet, Renker & Buscot) Sieverd., G.A. Silva & Oehl as the inoculum. *Rhizoglomus irregulare* (previous names *Glomus intraradices* and *R. irregularis*; (Sieverding, da Silva, Berndt, & Oehl, 2015) is an AMF taxon common in natural grasslands. The culture was developed for nine months in a substrate of 15 % soil, 65 % sand and 20 % oil binder with *P. lanceolata* plants, which had no shared selection history with plants or soils from the Jena Experiment. The *R. irregulare* material was obtained from

M.G.A. van der Heijden's Ecological Farming Group (Agroscope Reckenholz-Tänikon, Zurich, Switzerland).

2.5 | EXPERIMENTAL DESIGN

To establish the AMF treatments, we filled 1-L pots with gamma-radiated (27–54 kGy) 1:1 (weight/weight) sand-soil mixture and added 9 % (volume/volume) of inoculum without AMF (negative control), inoculum of AMF isolated from plants grown in monoculture (monoculture AMF) or mixture (mixture AMF) or inoculum containing *R. irregular* (a positive control). One monoculture- or mixture-type plant of a single plant species was planted into each pot (Fig. 1, lower panel). To standardize the non-AMF microbial community within each pot, we created a microbial wash by filtering 1.2 L of a mixture of unsterilized field soil and the AMF trap culture substrates through a series of sieves and finally through filter paper (MN615, Macherey-Nagel GmbH & Co. KG) with 5 L of deionized water. Each pot received 10 ml of the microbial-wash filtrate. The experiment included four AMF treatments in total, two plant histories (monoculture- and mixture-type plants) and five plant species in a full factorial design (Table S1). Control and *R. irregulare* AMF treatments were replicated five times and the two other AMF treatments were replicated ten times (five times per trap-culture replicate, Table S1). For the mixture-type plants of *G. mollugo*, we did not have sufficient seedlings for the full design and the AMF treatments were thus only replicated 9 and 8 times, respectively (Table S1). The 297 pots were randomly arranged within five experimental blocks in a glasshouse compartment with each particular treatment combination and trap-culture replicate occurring only once in each block.

2.6 | DATA COLLECTION

We cut the plants to 4 cm aboveground three months after planting seedlings into the pots of the AMF treatments (referred to as first harvest). After five months of plant growth, maximum height and average leaf absorbance (SPAD-502Plus Chlorophyll Meter, KONICA MINOLTA, INC., Osaka, Japan) of three representative leaves of each plant were measured and the aboveground biomass was harvested at ground-level (referred to as second harvest). The biomass of each plant was dried at 70 °C for 48 h and then weighed. We assessed leaf mass per area (LMA) and leaf dry matter content (LDMC) at the second harvest by measuring the area of fresh leaves (LI-3100C Area Meter, LI-COR, Lincoln, USA) immediately after harvest and assessing the weight of the leaves before (fresh weight) and after drying (dry weight). Finally, we estimated the degree of damage on plant aboveground tissues due to powdery mildew (family Erysiphaceae) and two-spotted spider mites (*Tetranychus urticae* Koch). To determine AMF colonization, roots were washed free of adhering rhizosphere soil and cut into small 1–5 cm fragments and a random subsample of roots were then stored in 50 % ethanol for microscopic quantification of AMF using the same clearing, staining and scoring methods as described above. All measured traits are listed in Table S2.

2.7 | DATA ANALYSES

Due to a contamination of control soil with AMF, one pot with *L. pratensis* was excluded from all analyses. The biomass data, morphological trait measurements, leaf damage estimates and AMF colonization were analysed using linear models. Because the first measure assessed growth and the second regrowth, the harvests were analyzed separately. Plant survival and AMF presence/absence were analysed using analysis of deviance. The results were summarized in analysis of variance (ANOVA) and deviance (ANDEV) tables

(McCullagh & Nelder, 1998; Schmid, Baruffol, Wang, & Niklaus, 2017). The explanatory terms of the models were block, plant history (monoculture-type vs. mixture-type), AMF treatments (four AMF treatments or sequence of the following three orthogonal contrasts: control vs. AMF treatments, *R. irregulare* vs. monoculture or mixture AMF and monoculture vs. mixture AMF), species and interactions of these. Statistical analyses were conducted using the software product R, version 3.0.2 (R Core Team, 2013).

3 | Results

3.1 | ROOT COLONIZATION

The root colonization by AMF was highest in all five plant species with the inculcation of *R. irregulare* (the positive control). For two species, *G. mollugo* and *L. pratensis*, AMF colonization was higher in the treatment with AMF originating from the plant monocultures, whereas for two other species, *P. lanceolata* and *V. chamaedrys*, root colonization was greater with AMF originating from plant mixtures. In *P. vulgaris*, AMF colonization was equally low for both AMF community histories. An increase in AMF colonization was positively correlated with an increase in individual plant biomass for all three AMF treatments (Fig. S1).

3.2 | EFFECTS OF AMF COMMUNITIES ON PLANT PHENOTYPIC RESPONSES (H1)

The four AMF treatments changed the phenotypes of the plants, but the response varied between the five species. We found significant effects of the AMF treatments on five traits: plant aboveground biomass at the first harvest, plant aboveground biomass at the second harvest, plant height, LDMC and AMF colonization (Fig. 2, Table 1). The effects of the AMF treatments on plant biomass were relatively consistent between the two harvests (Fig. 2). Biomass was generally highest in the treatment with the positive AMF control

293 (inoculation of *R. irregulare*), with the exception of *V. chamaedrys*, for which the negative
294 control (no AMF inoculated) resulted in the highest biomass. Soil containing AMF from
295 mixture plots was the overall second most productive AMF treatment in terms of plant
296 biomass, but again with the exception of *V. chamaedrys* and also *P. lanceolata* which
297 exhibited higher biomass when inoculated with AMF from monoculture plots. The effects of
298 the AMF treatments on plant height also varied among plant species. For *G. mollugo*, soil
299 containing *R. irregulare* or soil containing mixture AMF resulted in the tallest plants. For
300 three species, *L. pratensis*, *P. lanceolata* and *P. vulgaris*, the AMF treatment did not have a
301 strong effect on plant height, except that the control treatment lacking AMF resulted in the
302 shortest plants. In line with the biomass results, plant height was increased in the control
303 treatment for the small herb *V. chamaedrys*. The LDMC was strongly increased in the control
304 treatment for *L. pratensis* and slightly increased for *G. mollugo*. For the three small herbs, *P.*
305 *lanceolata*, *P. vulgaris* and *V. chamaedrys*, the LDMC was increased for the two AMF
306 treatments containing AMF from either mixture or monoculture plots as opposed to
307 treatments without AMF or with the external *R. irregulare*.
308
309 3.3 | INFLUENCE OF PLANT DIVERSITY HISTORY ON PLANT PHENOTYPES (H2)
310 Mixture-type plants grew taller than monoculture-type plants in three out of five
311 species. Monoculture-type plants of the other two species, *L. pratensis* and of *P. lanceolata*,
312 grew taller than their mixture-type counterparts. The leaf traits LDMC and LMA were
313 generally increased in mixture-type plants, except for *V. chamaedrys*. For plant height, LMA,
314 leaf damage and leaf absorbance the plant history effect was also visible across species
315 (Table 1, e.g. the main effect of plant history).
316 As anticipated, mixture-type plants overall had more damaged leaves than
317 monoculture-type plants, in particular for *L. pratensis* and *P. lanceolata* (Fig. 3, Table 1). The

318 monoculture-type or mixture-type plants resulted in differential phenotypes between
319 individuals of the same species. However, the strength of the response to selection history
320 varied greatly between the five studied species (Fig. 3, Table 1). We found significant overall
321 effects of plant history on seven traits: plant survival, plant aboveground biomass at the first
322 harvest, plant aboveground biomass at the second harvest, plant height, LMA, LDMC and
323 leaf damage. Mixture-type plants had a greater survival in the case of the three small herbs,
324 whereas for the legume *L. pratensis* and the tall herb *G. mollugo*, monoculture-type plants
325 showed a higher survival (Fig. 3). Plant biomass was generally higher for mixture-type plants
326 (Fig. 3), but *L. pratensis* and *G. mollugo* showed the opposite pattern with increased biomass
327 for monoculture-type plants (for *L. pratensis* only at the first harvest). The difference in
328 biomass production between monoculture-and mixture-type plants was smaller at the second
329 harvest but still varied significantly among species (Fig. 3).
330
331 3.4 | EFFECTS OF MATCHING HOME AND AWAY PLANT AND AMF HISTORIES
332 (H3)
333 We did find overall home vs. away effects for LMA and leaf absorbance, but the
334 effects varied among the five plant species studied and responses differed in direction (Fig.
335 4). The home mixture combination (mixture-type plants and mixture AMF) resulted in
336 increased LMA in four out of five species, the exception being *V. chamaedrys* (Fig. 4a). The
337 home monoculture combination (monoculture-type plants and monoculture-AMF) only
338 increased LMA in *P. vulgaris* (Fig. 4a). Both away-combinations (monoculture-plant type
339 with mixture AMF and vice versa) increased LMA strongly in *V. chamaedrys* (Fig. 4a). The
340 away-combination between mixture-type plants and monoculture AMF also increased LMA
341 in *L. pratensis* (Fig. 4a). Both home-combinations increased leaf absorbance in the small

342 herbs *P. vulgaris* and *V. chamaedrys* (Fig. 4b). In contrast, both home-combinations reduced
343 leaf absorbance for the tall herb *G. mollugo* and the legume *L. pratensis*.
344
345 **4 | Discussion**
346 4.1. | EFFECTS OF AMF COMMUNITIES ON PLANT PHENOTYPIC RESPONSES (H1)
347 Here we assessed whether plant diversity loss not only alters plant traits related to
348 competitition, but also the phenotypic responses of plants to their AMF communities.
349 Specifically, we aimed to separate whether plant diversity loss alters the selection on plants
350 for altered responses to local AMF communities (H1), traits related to competitive growth
351 (H2) or both simultaneously (H3). For instance, although plants may be selected for altered
352 competitive growth characteristics in a more species diverse community (Zuppinger-Dingley
353 et al. 2014), it is also known that interactions with soil organisms alters how plants allocate
354 resources to traits, which affects plant growth strategies (Dudenhöffer, Ebeling, Klein, &
355 Wagg, 2018; Streitwolf-Engel et al., 1997). Conversely, plant traits have been thought to
356 influence interactions between plants and AMF (Baxendale, Orwin, Poly, Pommier, &
357 Bardgett, 2014). A recent study focusing on plant traits in relation to PSF effects found
358 evidence that a lower specific root length and a greater percentage colonization of the root
359 length by AMF resulted in more positive PSFs (Cortois et al. 2016). Furthermore,
360 mycorrhizal associations have been known to alter leaf chemical properties and alter
361 herbivore life-history traits via changes in leaf chemistry (Goverde, van der Heijden,
362 Wiemken, Sanders, & Erhardt, 2000; Smith & Read, 1997).
363 We assessed whether plant diversity can act as a selective environment on how AMF
364 communities influence plant growth and functional traits. In support of our first hypothesis
365 (H1) we found significant effects of the AMF treatments on plant phenotypes. Mixture AMF
366 were more beneficial than monoculture AMF for the taller species *G. mollugo* and *L.*

367 *pratensis*, but not for the three shorter species *P. lanceolata*, *P. vulgaris* and *V. chamaedrys*.
368 All studied plant species except *V. chamaedrys* showed increased biomass production in the
369 presence of AMF; and across species, plant biomass generally increased with increasing
370 AMF colonization. Interestingly, mixture AMF showed lower colonization than monoculture
371 AMF in the plant species that benefited from them and vice versa in the other three species.
372 These species-specific responses are in line with other studies showing context-dependent
373 AMF effects on plants (Burrows & Pfleger, 2002; Hoeksema et al., 2010). Although AMF
374 generally promote plant growth, the outcome of the interaction may vary from beneficial to
375 antagonistic for plant growth (Johnson, Graham, & Smith, 1997; Kiers & van der Heijden,
376 2006; Klironomos, 2003). The positive effect of mixture AMF on species with a low
377 colonization could indicate increased AMF efficiency in promoting plant growth. However,
378 lower growth at high colonization could also indicate that a shift from mutualism to
379 antagonism occurred in these species.

380 Colonization in our positive AMF control treatment of *R. irregulare*, which did not
381 share a common history with the experimental plants, was greater than in mixture or
382 monoculture AMF treatments and led to higher plant biomass especially at the first harvest. A
383 high colonization ability of this taxon is typical and has been shown to be not as beneficial to
384 plant hosts as less abundant colonization by other AMF taxa (Hart and Reader 2002, Wagg et
385 al., 2011a, Engelmoer, Behm, & Kiers 2014). This particularly productive and cosmopolitan
386 AMF taxon may have been more effective in colonizing plant roots and influencing plant
387 responses compared to the inoculation of AMF spore communities native to the field site
388 where a greater diversity of AMF taxa were likely present (Dassen et al., 2017). The co-
389 inoculation of several AMF taxa could have resulted in competition among AMF taxa for a
390 single host plant in our study, thus reducing their ability to colonize roots and influence plant
391 responses (Bennett & Bever, 2009; Engelmoer et al., 2014). These results are in line with

previous findings reporting that different AMF isolates vary in their effect on plant phenotypes (Klironomos, 2003; Koch, Antunes, Maherali, Hart, & Klironomos, 2017; Streitwolf-Engel, Boller, Wiemken, & Sanders, 1997), which may depend on co-infection with other AMF species (Arguello et al., 2016; Bennett & Bever, 2009).

4.2 | INFLUENCE OF PLANT DIVERSITY HISTORY ON PLANT PHENOTYPES (H2)

Our second hypothesis, that mixture-type plants grow faster whereas monoculture-type plants are better defended was generally supported by our experiment (see significant main effects of plant history in Table 1). But here again the plant species varied in their responses. Mixture-type plants of three species were more productive and mixture-type plants of three species grew taller than monoculture-type plants. Furthermore, three out of the five species showed higher LDMC and four species higher LMA for mixture-type plants in comparison with monoculture-type plants, indicating that mixture-type plants also invested more resources into leaf biomass production. In contrast, monoculture-type plants had less leaf damage, which confirmed the second part of our hypothesis 2, namely that monoculture-type plants evolved increased defence. We observed particularly severe infection by powdery mildew in mixture-type plants of *P. lanceolata*, suggesting, in agreement with Engelmoer et al. (2014), that monoculture-type plants of *P. lanceolata* may have been subjected to particularly strong selection pressure for pathogen defence in comparison with mixture-type plants. Previously it has been shown that community diversity as selective environment can alter the selection on plant traits favouring greater character displacement in more diverse plant communities (Zuppinger-Dingley et al., 2014), with the assumption that competitive growth traits trade off with selection for pathogen defence in plant monocultures (Herms & Mattson, 1992). The increased resource investment into leaves in mixture-type plants thus may have been at the cost of lowered pathogen defence. Specialized pathogens have indeed

been observed to become diluted at higher plant diversity levels in the Jena Experiment (Eisenhauer, Reich, & Scheu, 2012; Rottstock, Joshi, Kummer, & Fischer, 2014), thus conferring lower selection pressure in mixtures than in monocultures. To increase survival, plants in monocultures are expected to allocate more resources to defence (Bezemer & Vandam, 2005) or may enter more beneficial symbioses (Newsham et al., 1995). Consequently, survival in plant monocultures may depend on the ability of plants to allocate resources to these interactions whereas plants in mixtures can allocate these resources to growth.

4.3 | HOME VS. AWAY EFFECTS OF PLANT AND AMF HISTORIES (H3)

Our third hypothesis was that a common selection history of plants and their associated fungi in the field would lead to a matching between plants and their associated AMF communities, i.e. 'home' vs. 'away' AMF effects on plant phenotypic responses. We found limited evidence for this hypothesis. The main predictor for increased positive PSF, plant biomass, was not affected by home vs. away combination, which indicates that beneficial interactions did not strengthen over time. Instead, we found a home vs. away effect for the two functional traits LMA and leaf absorbance. For these two leaf properties, we found some evidence that monoculture-type plants were more influenced by monoculture AMF and mixture-type plants were more influenced by mixture AMF. Namely, leaf absorbance and LMA were increased in the small herb *P. vulgaris* if the AMF had a common selection history. Increased leaf absorbance and high LMA are related to higher area-based nitrogen content (Niinemets, 1997), suggesting that co-selected AMF may have improved the nitrogen uptake of these plants. The opposite response was observed for leaf absorbance in *L. pratensis* and for LMA in *V. chamaedrys*. Although for LMA and leaf absorbance the results would be in line with our hypothesis for a specific effect of 'matching' plant and AMF

community histories (indicating their co-selection), the common selection history between plants and AMF was rather detrimental to the plants' growth response. This would suggest that a common association history between plants and their AMF communities may have become more antagonistic, rather than mutualistic, over the eleven years of co-selection.

5 | Conclusions

Here we present evidence that the loss of plant diversity differentially alters how AMF communities influence phenotypic responses in plants. We found that AMF communities selected in plant mixtures were more beneficial than AMF selected in plant monocultures for two out of five plant species. Furthermore, mixture-type plants generally grew better, but suffered more leaf damage than monoculture-type plants, providing evidence for the differential selection on competitive growth vs. defence depending on the interspecific diversity of neighbouring plants. However, home vs. away effects between AMF and plants from mixtures or AMF and plants from monocultures were rare. When they did occur they were generally in the direction of increased antagonism by the AMF, leading to a reduced performance of the plant partner. We assert that conducting reciprocal inoculation experiments over longer ecological time-scales could contribute to a better understanding of plant–AMF interactions (Lekberg & Koide, 2014). Our results suggest that biodiversity loss can alter evolutionary trajectories in the interactions between plants and AMF through differential selection pressures on both partners at high and low diversity. Furthermore, our data suggest that a shared history between plants and AMF does not follow a general pattern leading to increased mutualism. Rather, changes in these interactions are context-dependent and may strengthen over time, even switching directionality. Further investigating the sustainability of AMF–plant mutualisms in a world with changing species composition and

466 biodiversity will be important when searching for applications for AMF communities to
467 support ecosystem functioning in the future.
468
469 **Acknowledgements**
470 We thank M. Furler, D. Topalovic, D. Trujillo Villegas and T. Zwimpfer for technical
471 assistance and A. Ferrari, S. Karbin, J. Moser Frias and Y. Xu for help with measurements.
472 Thanks to M. van der Heijden for assistance and the AMF isolates. This study was supported
473 by the Swiss National Science Foundation (grants number 147092 and 166457 to B. S.) and
474 the University Research Priority Program Global Change and Biodiversity of the University
475 of Zurich. The Jena Experiment is supported by the German Science Foundation (FOR 1451,
476 SCHM 1628/5-2).
477
478 **Authors' contributions**
479 T.H., C.W., S.J.V.M., D.Z.D and B.S. planned and designed the research, T.H. carried
480 out the experiment and T.H., C.W., M.W.S., S.J.V.M and B.S. analyzed the data. S.J.V.M,
481 T.H., C.W. and B.S. wrote the manuscript with the other authors contributing to revisions.
482
483 **Data accessibility**
484 Data will be archived on Pangaea upon acceptance of this article.
485
486 **References**
487 Arguello, A., O'Brien, M. J., van der Heijden, M. G. A., Wiemken, A., Schmid, B., &
488 Niklaus, P. A. (2016). Options of partners improve carbon for phosphorus trade in the
489 arbuscular mycorrhizal mutualism. *Ecology Letters*, *19*(6), 648–656.
490 doi:10.1111/ele.12601

491 Baxendale, C., Orwin, K. H., Poly, F., Pommier, T., & Bardgett, R. D. (2014). Are plant–soil
492 feedback responses explained by plant traits? *New Phytologist*, *204*(2), 408–423.
493 doi:10.1111/nph.12915
494 Bennett, A. E., & Bever, J. D. (2009). Trade-offs between arbuscular mycorrhizal fungal
495 competitive ability and host growth promotion in Plantago lanceolata. *Oecologia*,
496 *160*(4), 807–816. doi:10.1007/s00442-009-1345-6
497 Bever, J. D. (1994). Feeback between plants and their soil communities in an old field
498 community. *Ecology*, *75*(7), 1965–1977. doi:10.2307/1941601
499 Bever, J. D., Dickie, I. A., Facelli, E., Facelli, J. M., Klironomos, J., Moora, M., … Zobel, M.
500 (2010). Rooting theories of plant community ecology in microbial interactions.
501 *Trends in Ecology & Evolution*, *25*(8), 468–478. doi:10.1016/j.tree.2010.05.004
502 Bezemer, T., & Vandam, N. (2005). Linking aboveground and belowground interactions via
503 induced plant defenses. *Trends in Ecology & Evolution*, *20*(11), 617–624.
504 doi:10.1016/j.tree.2005.08.006
505 Burrows, R. L., & Pfleger, F. L. (2002). Host responses to AMF from plots differing in plant
506 diversity. *Plant and Soil*, *240*(1), 169–180.
507 Cadotte, M. W. (2017). Functional traits explain ecosystem function through opposing
508 mechanisms. *Ecology Letters*, *20*(8), 989–996. doi:10.1111/ele.12796
509 Cortois, R., Schröder-Georgi, T., Weigelt, A., van der Putten, W. H., & De Deyn, G. B.
510 (2016). Plant-soil feedbacks: role of plant functional group and plant traits. *Journal of*
511 *Ecology*, *104*(6), 1608–1617. doi:10.1111/1365-2745.12643
512 Dassen, S., Cortois, R., Martens, H., de Hollander, M., Kowalchuk, G. A., van der Putten, W.
513 H., & De Deyn, G. B. (2017). Differential responses of soil bacteria, fungi, archaea
514 and protists to plant species richness and plant functional group identity. *Molecular*
515 *Ecology*. doi:10.1111/mec.14175

516 Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., … Gorné, L. D.
517 (2016). The global spectrum of plant form and function. *Nature*, *529*(7585), 167–171.
518 doi:10.1038/nature16489
519 Dudenhöffer, J.-H., Ebeling, A., Klein, A.-M., & Wagg, C. (2018). Beyond biomass: Soil
520 feedbacks are transient over plant life stages and alter fitness. *Journal of Ecology*,
521 *106*(1), 230–241. doi:10.1111/1365-2745.12870
522 Eisenhauer, N. (2012). Aboveground–belowground interactions as a source of
523 complementarity effects in biodiversity experiments. *Plant and Soil*, *351*(1–2), 1–22.
524 doi:10.1007/s11104-011-1027-0
525 Eisenhauer, N., Reich, P. B., & Scheu, S. (2012). Increasing plant diversity effects on
526 productivity with time due to delayed soil biota effects on plants. *Basic and Applied*
527 *Ecology*, *13*(7), 571–578. doi:10.1016/j.baae.2012.09.002
528 Engelmoer, D. J. P., Behm, J. E., & Toby Kiers, E. (2014). Intense competition between
529 arbuscular mycorrhizal mutualists in an *in vitro* root microbiome negatively affects
530 total fungal abundance. *Molecular Ecology*, *23*(6), 1584–1593.
531 doi:10.1111/mec.12451
532 Fakheran, S., Paul-Victor, C., Heichinger, C., Schmid, B., Grossniklaus, U., & Turnbull, L.
533 A. (2010). Adaptation and extinction in experimentally fragmented landscapes.
534 *Proceedings of the National Academy of Sciences*, *107*(44), 19120–19125.
535 doi:10.1073/pnas.1010846107
536 Francis, R., & Read, D. J. (1994). The contributions of mycorrhizal fungi to the
537 determination of plant community structure. *Plant and Soil*, *159*(1), 11–25.
538 doi:10.1007/BF00000091

539 Goverde, M., van der Heijden, M. G. A., Wiemken, A., Sanders, I. R., & Erhardt, A. (2000).
540 Arbuscular Mycorrhizal Fungi Influence Life History Traits of a Lepidopteran
541 Herbivore. *Oecologia*, *125*(3), 362–369.
542 Harley, J. L., & Harley, E. L. (1987). A check-list of Mycorrhiza in the British flora. *New*
543 *Phytologist*, *105*, 1–102.
544 Hart, M. M., & Reader, R. J. (2002). Taxonomic Basis for Variation in the Colonization
545 Strategy of Arbuscular Mycorrhizal Fungi. *The New Phytologist*, *153*(2), 335–344.
546 Herms, D. A., & Mattson, W. J. (1992). The Dilemma of Plants: To Grow or Defend. *The*
547 *Quarterly Review of Biology*, *67*(3), 283–335.
548 Hoeksema, J. D., Chaudhary, V. B., Gehring, C. A., Johnson, N. C., Karst, J., Koide, R. T.,
549 … Umbanhowar, J. (2010). A meta-analysis of context-dependency in plant response
550 to inoculation with mycorrhizal fungi. *Ecology Letters*, *13*(3), 394–407.
551 doi:10.1111/j.1461-0248.2009.01430.x
552 Johnson, N. C., Graham, J. H., & Smith, F. A. (1997). Functioning of Mycorrhizal
553 Associations Along the Mutualism-Parasitism Continuum. *The New Phytologist*,
554 *135*(4), 575–586.
555 Joshi, J., Schmid, B., Caldeira, M. C., Dimitrakopoulos, P. G., Good, J., Harris, R., … others.
556 (2001). Local adaptation enhances performance of common plant species. *Ecology*
557 *Letters*, *4*(6), 536–544.
558 Kardol, P., Martijn Bezemer, T., & van der Putten, W. H. (2006). Temporal variation in
559 plant-soil feedback controls succession: Plant-soil feedback controls succession.
560 *Ecology Letters*, *9*(9), 1080–1088. doi:10.1111/j.1461-0248.2006.00953.x
561 Kiers, E. T., Duhamel, M., Beesetty, Y., Mensah, J. A., Franken, O., Verbruggen, E., …
562 Bücking, H. (2011). Reciprocal Rewards Stabilize Cooperation in the Mycorrhizal
563 Symbiosis. *Science*, *333*(6044), 880. doi:10.1126/science.1208473

564 Kiers, E. T., & van der Heijden, M. G. (2006). Mutualistic stability in the arbuscular
565 mycorrhizal symbiosis: exploring hypotheses of evolutionary cooperation. *Ecology*,
566 *87*(7), 1627–1636.
567 Klironomos, J. N. (2002). Feedback with soil biota contributes to plant rarity and
568 invasiveness in communities. *Nature*, *417*(6884), 67–70. doi:10.1038/417067a
569 Klironomos, J. N. (2003). Variation in plant response to native and exotic arbuscular
570 mycorrhizal fungi. *Ecology*, *84*(9), 2292–2301.
571 Klironomos, J. N., McCune, J., Hart, M., & Neville, J. (2000). The influence of arbuscular
572 mycorrhizae on the relationship between plant diversity and productivity. *Ecology*
573 *Letters*, *3*(2), 137–141.
574 Koch, A. M., Antunes, P. M., Maherali, H., Hart, M. M., & Klironomos, J. N. (2017).
575 Evolutionary asymmetry in the arbuscular mycorrhizal symbiosis: conservatism in
576 fungal morphology does not predict host plant growth. *New Phytologist*, *214*(3),
577 1330–1337. doi:10.1111/nph.14465
578 König, S., Wubet, T., Dormann, C. F., Hempel, S., Renker, C., & Buscot, F. (2010). TaqMan
579 real-time PCR assays to assess arbuscular mycorrhizal responses to field manipulation
580 of grassland biodiversity: effects of soil characteristics, plant species richness, and
581 functional traits. *Applied and Environmental Microbiology*, *76*(12), 3765–3775.
582 Kulmatiski, A., Beard, K. H., & Heavilin, J. (2012). Plant-soil feedbacks provide an
583 additional explanation for diversity-productivity relationships. *Proceedings of the*
584 *Royal Society B: Biological Sciences*, *279*(1740), 3020–3026.
585 doi:10.1098/rspb.2012.0285
586 Kulmatiski, A., Beard, K. H., Stevens, J. R., & Cobbold, S. M. (2008). Plant-soil feedbacks: a
587 meta-analytical review. *Ecology Letters*, *11*(9), 980–992. doi:10.1111/j.1461-
588 0248.2008.01209.x

589 Latz, E., Eisenhauer, N., Rall, B. C., Allan, E., Roscher, C., Scheu, S., & Jousset, A. (2012).
590 Plant diversity improves protection against soil-borne pathogens by fostering
591 antagonistic bacterial communities: *Plant diversity improves protection against soil-*
592 *borne pathogens*. *Journal of Ecology*, *100*(3), 597–604. doi:10.1111/j.1365-
593 2745.2011.01940.x
594 Lau, J. A., & Lennon, J. T. (2011). Evolutionary ecology of plant-microbe interactions: soil
595 microbial structure alters selection on plant traits. *New Phytologist*, *192*(1), 215–224.
596 doi:10.1111/j.1469-8137.2011.03790.x
597 Lekberg, Y., & Koide, R. T. (2014). Integrating physiological, community, and evolutionary
598 perspectives on the arbuscular mycorrhizal symbiosis. *Botany*, *92*(4), 241–251.
599 doi:10.1139/cjb-2013-0182
600 Marquard, E., Schmid, B., Roscher, C., De Luca, E., Nadrowski, K., Weisser, W. W., &
601 Weigelt, A. (2013). Changes in the Abundance of Grassland Species in Monocultures
602 versus Mixtures and Their Relation to Biodiversity Effects. *PLoS ONE*, *8*(9), e75599.
603 doi:10.1371/journal.pone.0075599
604 McCullagh, P., & Nelder, J. A. (1998). *Generalized Linear Models* (Second Edition).
605 Chapman and Hall. Retrieved from
606 http://www.utstat.toronto.edu/~brunner/oldclass/2201s11/readings/glmbook.pdf
607 McGonigle, T.P., Miller, M.H., Evans, D.G., Fairchild, G. L., & Swan, J. A. (1990). A new
608 method which gives an objective measure of colonization of roots by vesicular—
609 arbuscular mycorrhizal fungi. *New Phytologist*, *115*(3), 495–501. doi:10.1111/j.1469-
610 8137.1990.tb00476.x
611 Milcu, A., Allan, E., Roscher, C., Jenkins, T., Meyer, S. T., Flynn, D., … Eisenhauer, N.
612 (2013). Functionally and phylogenetically diverse plant communities key to soil biota.
613 *Ecology*, *94*(8), 1878–1885. doi:10.1890/12-1936.1

614 Newsham, K. K., Fitter, A. H., & Watkinson, A. R. (1995). Arbuscular Mycorrhiza Protect an
615 Annual Grass from Root Pathogenic Fungi in the Field. *Journal of Ecology*, *83*(6),
616 991–1000. doi:10.2307/2261180
617 Niinemets, U. (1997). Role of Foliar Nitrogen in Light Harvesting and Shade Tolerance of
618 Four Temperate Deciduous Woody Species. *Functional Ecology*, *11*(4), 518–531.
619 Petermann, J. S., Fergus, A. J., Turnbull, L. A., & Schmid, B. (2008). Janzen-Connell effects
620 are widespread and strong enough to maintain diversity in grasslands. *Ecology*, *89*(9),
621 2399–2406.
622 Powell JR, Parrent JL, Hart MM, Klironomos JN, Rillig MC, Maherali H (2009)
623 Phylogenetic trait conservatism and the evolution of functional trade-offs in
624 arbuscular mycorrhizal fungi. *Proceedings of the Royal Society B,* 276, 4237–4245.
625 doi:10.1098/rspb.2009.1015
626
627 Pozo, M. J., & Azcón-Aguilar, C. (2007). Unraveling mycorrhiza-induced resistance. *Current*
628 *Opinion in Plant Biology*, *10*(4), 393–398. doi:10.1016/j.pbi.2007.05.004
629 Reich, P. B. (2014). The world-wide 'fast–slow' plant economics spectrum: a traits
630 manifesto. *Journal of Ecology*, *102*(2), 275–301. doi:10.1111/1365-2745.12211
631 Roscher, C., Schumacher, J., Baade, J., Wilcke, W., Gleixner, G., Weisser, W. W., …
632 Schulze, E.-D. (2004). The role of biodiversity for element cycling and trophic
633 interactions: an experimental approach in a grassland community. *Basic and Applied*
634 *Ecology*, *5*(2), 107–121.
635 Rottstock, T., Joshi, J., Kummer, V., & Fischer, M. (2014). Higher plant diversity promotes
636 higher diversity of fungal pathogens, while it decreases pathogen infection per plant.
637 *Ecology*, *95*(7), 1907–1917.
638 Scherber, C., Eisenhauer, N., Weisser, W. W., Schmid, B., Voigt, W., Fischer, M., …
639 Tscharntke, T. (2010). Bottom-up effects of plant diversity on multitrophic

640 interactions in a biodiversity experiment. *Nature*, *468*(7323), 553–556.
641 doi:10.1038/nature09492
642 Schmid, B., Baruffol, M., Wang, Z., & Niklaus, P. A. (2017). A guide to analyzing
643 biodiversity experiments. *Journal of Plant Ecology*, *10*(1), 91–110.
644 doi:10.1093/jpe/rtw107
645 Schnitzer, S. A., Klironomos, J. N., HilleRisLambers, J., Kinkel, L. L., Reich, P. B., Xiao, K.,
646 … others. (2011). Soil microbes drive the classic plant diversity–productivity pattern.
647 *Ecology*, *92*(2), 296–303.
648 Sieverding, E., da Silva, G. A., Berndt, R., & Oehl, F. (2015). Rhizoglomus, a new genus of
649 the Glomeraceae. *Mycotaxon*, *129*(2), 373–386. doi:10.5248/129.373
650 Smith, S. E., & Read, D. J. (2008). Mycorrhizal symbiosis (Third Edition). Academic Press,
651 Amsterdam.
652 Streitwolf-Engel, R., Boller, T., Wiemken, A., & Sanders, I. R. (1997). Clonal Growth Traits
653 of Two Prunella Species are Determined by Co-Occurring Arbuscular Mycorrhizal
654 Fungi from a Calcareous Grassland. *Journal of Ecology*, *85*(2), 181–191.
655 doi:10.2307/2960650
656 terHorst, C. P., Lennon, J. T., & Lau, J. A. (2014). The relative importance of rapid evolution
657 for plant-microbe interactions depends on ecological context. *Proceedings of the*
658 *Royal Society B: Biological Sciences*, *281*(1785), 20140028–20140028.
659 doi:10.1098/rspb.2014.0028
660 van der Heijden, M. G. A., Streitwolf-Engel, R., Riedl, R., Siegrist, S., Neudecker, A.,
661 Ineichen, K., … Sanders, I. R. (2006). The mycorrhizal contribution to plant
662 productivity, plant nutrition and soil structure in experimental grassland. *New*
663 *Phytologist*, *172*(4), 739–752. doi:10.1111/j.1469-8137.2006.01862.x

664 van der Heijden, M. G., Klironomos, J. N., Ursic, M., Moutoglis, P., Streitwolf-Engel, R.,
665 Boller, T., … Sanders, I. R. (1998). Mycorrhizal fungal diversity determines plant
666 biodiversity, ecosystem variability and productivity. *Nature*, *396*, 69–72.
667 van der Putten, W. H., Bardgett, R. D., Bever, J. D., Bezemer, T. M., Casper, B. B., Fukami,
668 T., … Wardle, D. A. (2013). Plant-soil feedbacks: the past, the present and future
669 challenges. *Journal of Ecology*, *101*(2), 265–276. doi:10.1111/1365-2745.12054
670 van Moorsel, S. J., Hahl, T., Wagg, C., De Deyn, G. B., Flynn, D. F. B., Zuppinger-Dingley,
671 D., & Schmid, B. (2018). Community evolution increases plant productivity at low
672 diversity. *Ecology Letters*, *21*, 128–137. doi:10.1111/ele.12879
673 van Moorsel, S. J., Schmid, M. W., Hahl, T., Zuppinger-Dingley, D., & Schmid, B. (2017).
674 Selection in response to community diversity alters plant performance in newly
675 assembled test communities. *BioRxiv*. doi:10.1101/158709
676 Vierheilig, H., Coughlan, A. P., Wyss, U., & Piché, Y. (1998). Ink and Vinegar, a Simple
677 Staining Technique for Arbuscular-Mycorrhizal Fungi. *Applied and Environmental*
678 *Microbiology*, *64*(12), 5004–5007.
679 Wagg, C., Jansa, J., Stadler, M., Schmid, B., & van der Heijden, M. G. A. (2011a).
680 Mycorrhizal fungal identity and diversity relaxes plant—plant competition. *Ecology*,
681 *92*(6), 1303–1313.
682 Wagg, C., Husband, B.C., Massicotte, H.B., & Peterson, R.L. (2011b) Soil microbial
683 communities from an elevational cline differ in their effect on conifer seedling
684 growth. *Plant and Soil* 340: 491-504.
685 Wang, B., & Qiu, Y.-L. (2006). Phylogenetic distribution and evolution of mycorrhizas in
686 land plants. *Mycorrhiza*, *16*(5), 299–363. doi:10.1007/s00572-005-0033-6
687 Werner, G. D. A., & Kiers, E. T. (2015). Partner selection in the mycorrhizal mutualism. *New*
688 *Phytologist*, *205*(4), 1437–1442. doi:10.1111/nph.13113

689 Zuppinger-Dingley, D., Flynn, D. F. B., De Deyn, G. B., Petermann, J. S., & Schmid, B.
690 (2016). Plant selection and soil legacy enhance long-term biodiversity effects.
691 *Ecology*, *97*(4), 918–928. doi:10.1890/15-0599.1
692 Zuppinger-Dingley, D., Schmid, B., Petermann, J. S., Yadav, V., De Deyn, G. B., & Flynn,
693 D. F. B. (2014). Selection for niche differentiation in plant communities increases
694 biodiversity effects. *Nature*, *515*(7525), 108–111. doi:10.1038/nature13869
695

696 **Supporting information**

697 Additional Supporting Information may be found online in the Supporting Information tab for
698 this article:

699 **Methods S1.** Assessment of soil N and P content at the beginning of the experiment.

700 **Fig. S1** Aboveground biomass production of monoculture- and mixture-type plants in control
701 soil in dependence of different levels of AMF colonization.

702 **Table S1.** Experimental design.

703 **Table S2.** Plant traits measured at the experimental plant age.

704

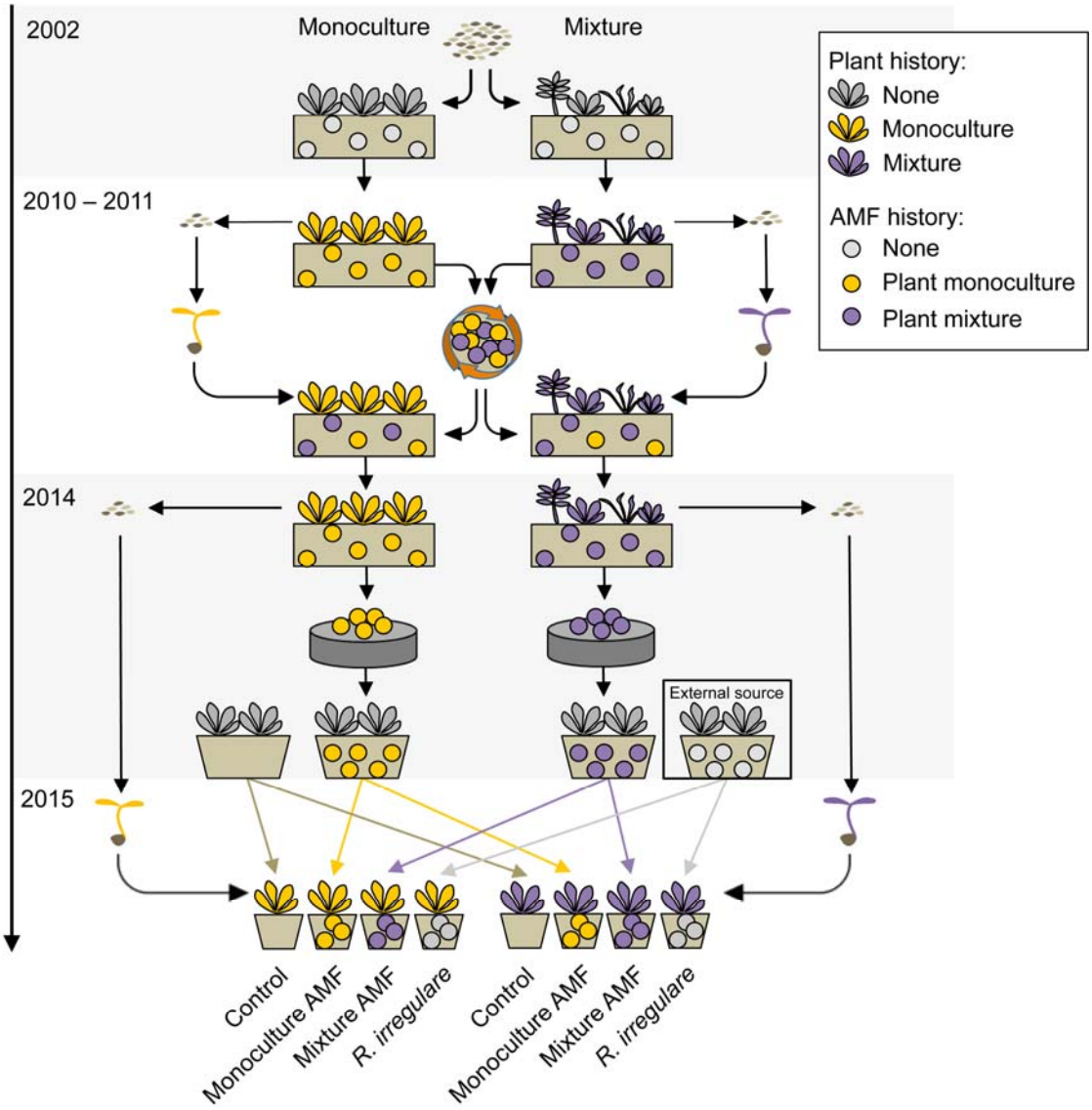
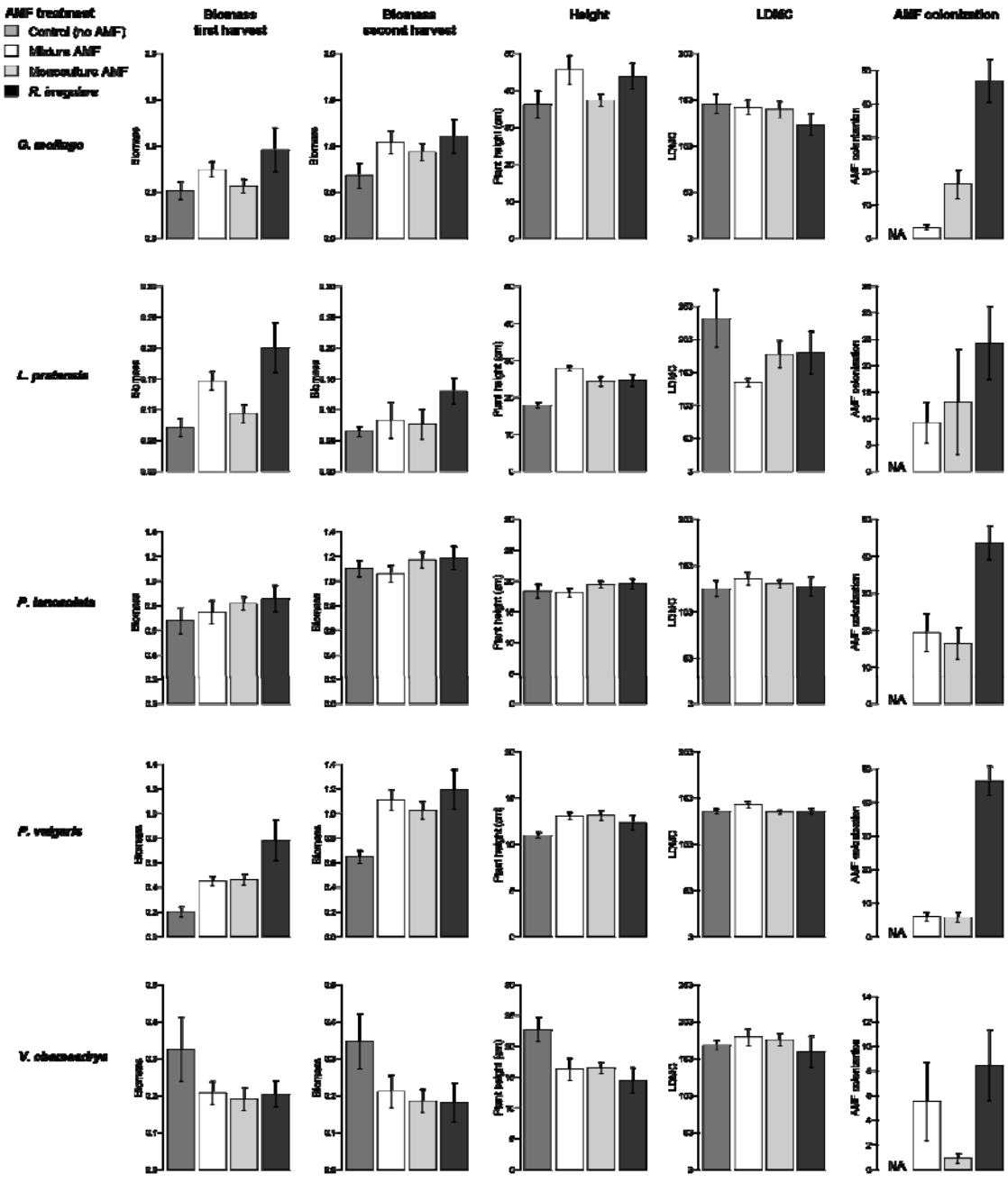


Figure 1. Experimental design. Plant monocultures and mixtures in the Jena Experiment were sown in 2002 and maintained until 2010. In 2010, the plants of 48 plots underwent a first controlled seed production event and the soil of the plots was pooled, mixed and placed back to the excavated locations. In spring 2011, the seedlings produced were transplanted back to the mixed soil in the same plots from which their parents were excavated. The plant communities could then again associate with their own microbial communities potentially co-assembling and co-evolving until 2014. In spring 2014, the plants underwent a second controlled seed production event, and the AMF spores from their rhizosphere soil were isolated. The isolated AMF communities accumulated in trap-cultures for ten months with

715 trap plants lacking a common selection history with the AMF spores. Control trap-cultures
716 without AMF spores were established as negative control. Four treatments were created: pots
717 with sterile soil and 1) 9 % inoculum without AMF, 2) inoculum of AMF isolated from plants
718 grown in monoculture, 3) inoculum of AMF isolated from plants grown in mixture and 4)
719 inoculum containing *Rhizoglomus irregulare*. Finally, the plants with a selection history in
720 either monoculture (monoculture-type plants) or mixture (mixture-type plants) were planted
721 individually into the prepared pots.
722

723



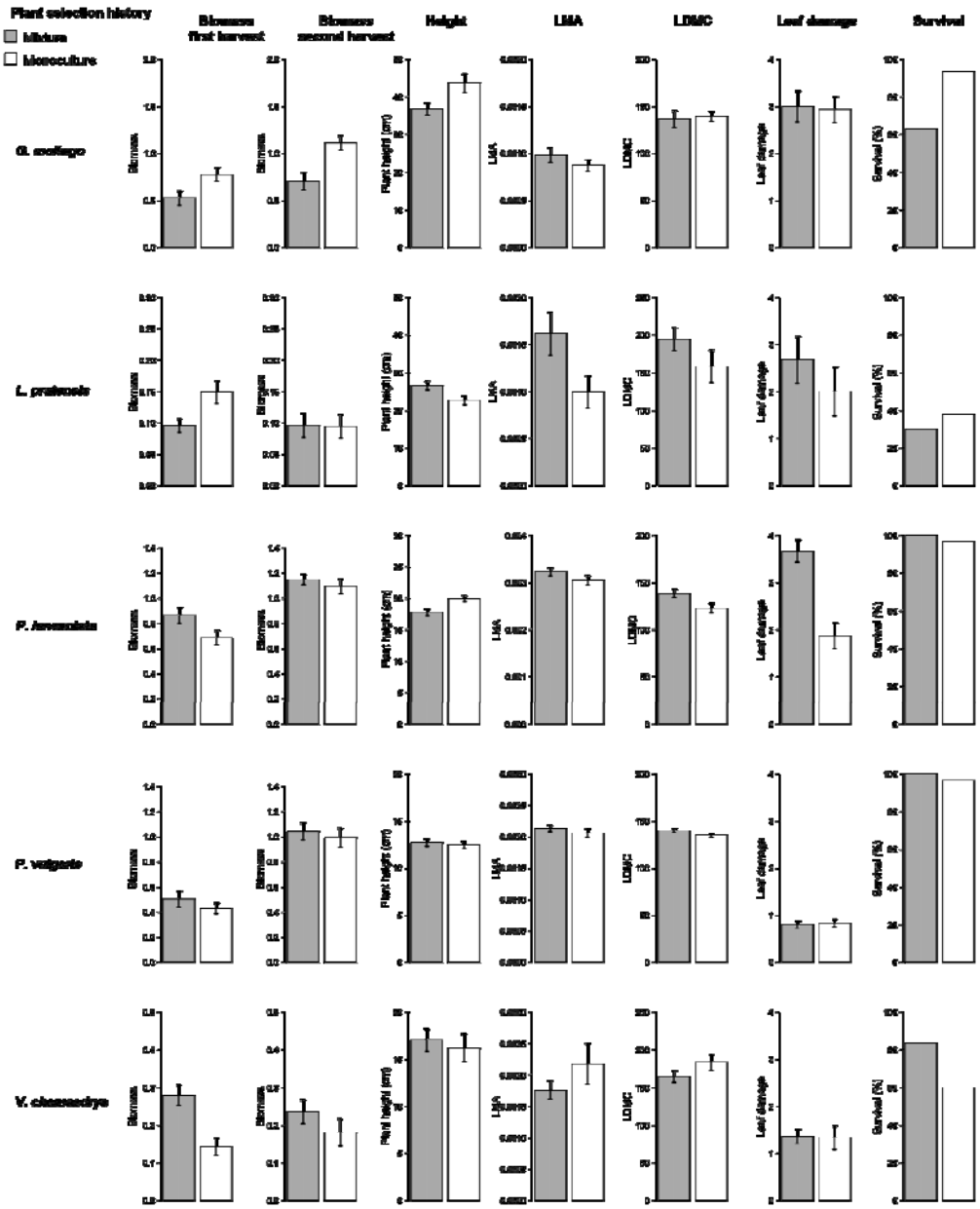
724

725 **Figure 2** Functional traits in responses to the AMF treatments are shown separately for each

726 of the five plant species assessed. Bars are means with standard errors calculated from raw

727 data. See Table 1 for significance of effects.

728



729

730 **Figure 3.** Functional traits in response to plant selection history for five species. Grey bars
731 refer to plants with a history of growing in species mixtures at the Jena Experiment, white
732 bars refers to plants with a history of growing in monoculture at the same field site. Shown
733 are means and standard errors calculated from raw data. See Table 1 for significance of
734 effects.

735

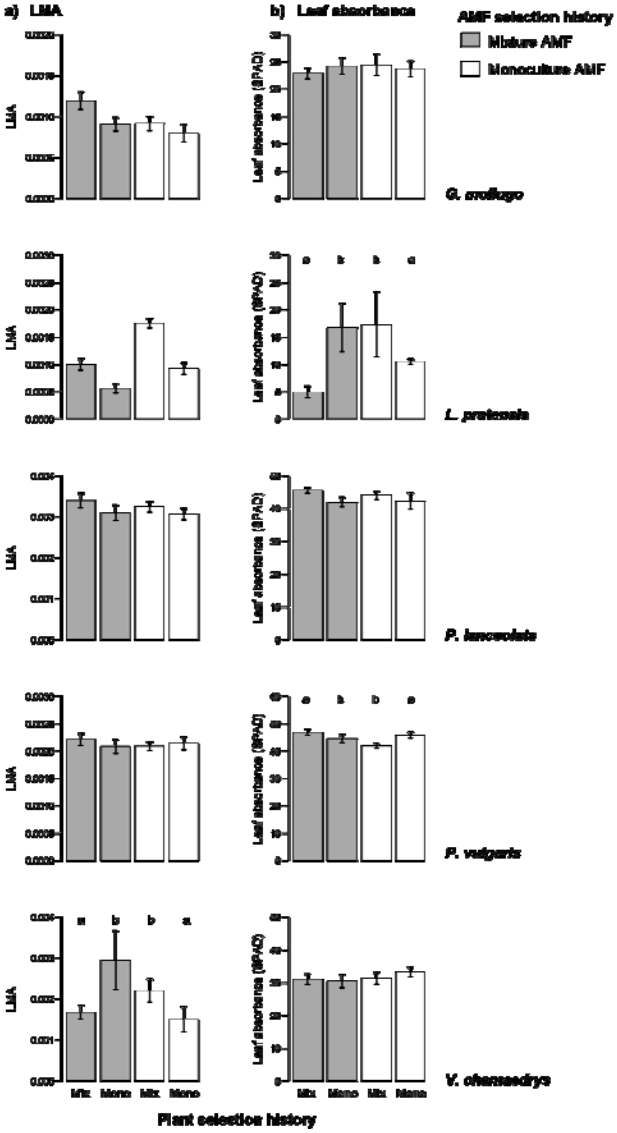


Figure 4 (a) LMA and (b) leaf absorbance in response to home and away combinations of plant selection history and AMF selection history for five species. Grey bars refer to AMF from mixed plant communities; white bars refer to AMF from monocultures. The first and last bar of each plot corresponds to home combinations; the two middle bars correspond to away combinations. Shown are means and standard errors calculated from raw data. Significant home effects are indicated by letters "a" and "b" on top of bars: $P < 0.01$ for leaf absorbance in *L. pratense*, $P < 0.05$ for leaf absorbance in *P. vulgaris* and $P < 0.05$ for LMA in *V. chamaedrys*.

Table 1. ANOVA results for eight traits and ANDEV results for plant survival in response to the design variables.

	Plant survival			Biomass first harvest			Biomass second harvest			Plant height			LMA			LDMC			Leaf damage			Leaf absorbance			AMF colonization		
Source of variation	***Df***	**% Dev**	***P***	***Df***	***% SS***	***P***	***Df***	***% SS***	***P***	***Df***	***% SS***	***P***	***Df***	***% SS***	***P***	***Df***	***% SS***	***P***	***Df***	**% SS**	***P***	***Df***	**% SS**	***P***	***Df***	**% SS**	***P***
Block	4	0.81	0.448	4	6.25	< 0.001	4	6.63	< 0.001	4	1.31	0.018	4	4.85	< 0.001	4	5.58	0.002	4	5.68	< 0.001	4	3.40	< 0.001	4	1.70	0.130
Plant history (PH)	1	0.03	0.689	1	0.03	0.656	1	0.47	0.050	1	2.26	< 0.001	1	0.88	0.014	1	0.63	0.164	1	1.43	0.016	1	0.39	0.032	1	0.10	0.510
AMF treatment (ST)	3	1.44	0.088	3	3.52	< 0.001	3	0.67	0.139	3	0.44	0.258	3	0.95	0.090	3	0.25	0.852	3	0.57	0.503	3	1.58	0.000	3	38.21	< 0.001
Species (Sp)	4	31.48	< 0.001	4	45.11	< 0.001	4	62.74	< 0.001	4	69.90	< 0.001	4	59.82	< 0.001	4	20.53	< 0.001	4	35.70	< 0.001	4	75.64	< 0.001	4	7.21	< 0.001
PH x ST	3	0.34	0.668	3	0.06	0.935	3	0.20	0.646	3	0.38	0.315	3	0.47	0.357	3	0.72	0.524	3	0.04	0.982	3	0.04	0.916	3	0.11	0.923
PH x Sp	*4*	*5.06*	*< 0.001*	4	3.58	< 0.001	4	2.55	< 0.001	4	1.67	0.005	4	1.60	0.028	4	3.70	0.024	4	6.70	< 0.001	4	0.31	0.446	4	0.56	0.665
ST x Sp	12	3.85	0.131	*12*	*4.44*	*0.005*	*12*	*2.94*	*0.024*	*12*	*3.37*	*0.003*	12	2.18	0.245	12	7.09	0.045	12	3.07	0.396	12	1.46	0.147	12	9.02	< 0.001
PH x ST x Sp	12	2.35	0.546	12	3.00	0.081	11	1.73	0.224	11	1.03	0.568	*11*	*3.30*	*0.024*	11	3.61	0.429	11	2.59	0.470	11	1.78	0.038	11	2.00	0.671
Residuals	252	54.65		224	34.01		183	22.06		183	19.63		180	25.94		180	57.88		183	44.21		183	15.38		174	41.08	

746