Phylogenetic profiling identifies glucosyl-phosphoryl dolichol scramblase candidates

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1 Background

Protein glycosylation is essential for all eukaryotes, from disease-causing protists such as malaria and trypanosomes, to yeast and mammals. Secretory proteins are almost invariably *N*-glycosylated, *O*- and *C*-mannosylated, and/or GPI-anchored as they enter the lumen of the endoplasmic reticulum (ER).

All ER protein glycosylation reactions occur in the *lumen* and often involve luminal mannosylation and glucosylation steps in which mannose and glucose residues are sourced from the glycolipids mannosyl- and glucosyl-phosphoryl dolichol (MPD and GPD, respectively) [1, 2]. Paradoxically, these two lipids are synthesized on the *cytoplasmic* face of the ER and must therefore be flipped across the ER membrane to provide a source of *luminal* mannose and glucose. As the spontaneous rate of MPD and GPD flipping is extremely low, specific transporters are needed to facilitate the transbilayer movement of MPD and GPD across the ER membrane at a physiological rate. MPD and GPD transport activities have been demonstrated and characterized in ER microsomes, as well as in vesicles reconstituted with ER membrane proteins [3–6]. The transport proteins have been shown to be highly structure specific, discriminating between isomers of their lipid substrates, and facilitating lipid movement bidirectionally in an ATP-independent manner (the last point defines them as *scramblases*, whereas they were previously known as ATP-independent flippases). Although most of the enzymes and co-factors of ER protein glycosylation are known, the molecular identities of the critical dolichol glycolipid scramblases remain a mystery.

Unlike MPD scramblase that is required for all ER protein glycosylation reactions, GPD scramblase is needed exclusively for the synthesis of the glucosylated *N*-glycan precursor Glc₃Man₉GlcNAc₂-PP-dolichol (G3M9-DLO). Even though non-glucosylated *N*-glycan precursors are substrates for the protein *N*-glycosylation machinery, the presence of the tri-glucosyl cap – and hence GPD scramblase activity – is critically important for glycosylation efficiency in many eukaryotes, including yeast and humans [7]. Two points are noteworthy. (i) Not all organisms have glucose in their *N*-glycan precursor [8]. (ii) While the synthesis of glucosylated *N*-glycan precursors is not essential for the viability of yeast [9, 10], yeast cells that are deficient in G3M9-DLO synthesis display numerous phenotypes including under-glycosylation of proteins, abnormal cell shape and altered susceptibility to a variety of chemicals. In humans, optimization of oligosaccharide transfer efficiency provided by the glucosyl cap is critical as evinced by severe human diseases.

Taking advantage of the fact that not all *N*-glycosylation-competent organisms have glucose in their *N*-glycan precursor, we implemented a bioinformatics approach for assignment of protein function, namely phylogenetic profiling. Using this procedure, we identified a number of polytopic ER membrane proteins as GPD scramblase candidates in yeast.

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Figure 1: Role of GPD scramblase in G3M9-DLO synthesis in yeast. Glc₃Man₉GlcNAc₂-PP-dolichol (G3M9-DLO; DLO=dolichol-linked oligosaccharide) is synthesized via a series of reactions that begin on the cytoplasmic face of the ER to generate Man₅GlcNAc₂-PP-dolichol (M5-DLO). M5-DLO is translocated across the ER membrane by M5-DLO scramblase. In the lumen, M5-DLO is extended to Man₉GlcNAc₂-PP-dolichol (M9-DLO) through the action of MPD-dependent mannosyltransferases and eventually extended to G3M9-DLO. The glucosylation reactions require lumenally oriented GPD. GPD is synthesized from dolichyl-P and UDP-glucose on the cytoplasmic face of the ER by GPD synthase (Alg5), and then moved to the lumen by GPD scramblase. In the lumen, the glucosyltransferases Alg6, Alg8 and Alg10 use GPD to add glucose residues to M9-DLO to generate G3M9-DLO that provides the oligosaccharide used by oligosaccharyltransferase (OST) to *N*-glycosylate proteins. The synthesis, scrambling and consumption of GPD likely form a co-evolving metabolic unit. This premise is the basis of our phylogenetic profiling identification of GPD scramblase candidates.

2 Results

2.1 GPD scramblase candidates identified by phylogenetic profiling

Phylogenetic profiling predicts protein function based on patterns of protein presence or absence across multiple species (e.g. [11, 12]). If homologs are inherited or lost co-dependently, there is a high chance that they are functionally related or physically interacting, because they are likely to be subject to the same functional constraints, or lack thereof. Proteins with similar phylogenetic profiles tend to be part of a functional unit. A previous study [13] showed that glucosyltransferases are present or absent in organisms in sets. Here we used patterns generated by phylogenetic profiling to identify GPD scramblase candidates. As not all organisms have glucose in their *N*-glycan precursor [8] we hypothesized that the presence or absence of GPD scramblase in a particular organism will be highly correlated with, but not necessarily identical to, the presence or absence of other proteins of the glucosylation pathway, e.g. the glucosyltransferase Alg6 (Figure 1). We used yeast as a reference organism as it has a well-annotated genome and a complete *N*-glycosylation pathway.

2.2 Annotated genomes for phylogenetic profiling

We downloaded 687 annotated genomes from the Ensembl database: 408 fungal, 133 protist, 143 archaeal, 3 higher eukaryotes (dog, mouse, human). We downloaded all the fungal and all the protist genomes from release 27 of the Ensembl database, as well as 143 selected archaeal genomes (those which are also part of the OMA Browser database).

Within this set we identified organisms capable of *N*-glycosylation by using BLAST [14] (E-value threshold 1e-6) to probe for Alg7, the enzyme that initiates DLO synthesis, and Stt3, the catalytic subunit of oligosaccharyltransferase (we identified the Alg7 and Stt3 proteins in *Saccharomyces cerevisiae* from the SwissProt database as entries P07286.1 and P39007.2, respectively, which correspond to the YBR243C and YGL022W proteins in the Ensembl (SGD) annotation). Only genomes encoding both proteins were retained.

Phylogenetic profiling benefits from an increased amount of input data, but previous studies have shown diminishing returns as greater numbers of genomes are added [15]. Thus, we further reduced our dataset by picking a single exemplar for species that were represented by more than one strain. Our final list contained 337 genomes (see Appendix A).

2.3 Species tree of organisms capable of *N*-glycosylation

Several authors have suggested that phylogenetic profiles should be ordered by the phylogenetic relatedness of the constituent organisms, e.g., Cokus et al. [16]. The 337 organisms being used in this analysis were imported into

NCBI's Taxonomy Browser (<http://www.ncbi.nlm.nih.gov/Taxonomy/CommonTree/wwwcmt.cgi>). Some species were represented in NCBI under a different name, either because the name was spelled slightly differently in NCBI, or because the asexual and sexual stages have different names; these were accordingly changed before import. Table 1 indicates the species which are named differently in the Ensembl and NCBI databases.

Ensembl nomenclature	NCBI nomenclature
Dacryopinax sp	Dacryopinax sp. DJM-731 SS1
Microsporidia sp	Mitosporidium daphniae
Paracoccidioides sp	Paracoccidioides sp. 'lutzii' Pb01
Phytomonas sp	Phytomonas sp. EM1
Saccharomycetaceae sp	Saccharomycetaceae sp. 'Ashbya aceri'
Pyrenophora tritici-repentis	Pyrenophora tritici-repentis
Ashbya gossypii	Eremothecium gossypii
Canis familiaris	Canis lupus familiaris
Ceriporiopsis subvermispora	Gelatoporia subvermispora
Dothistroma septosporum	Mycosphaerella pini
Magnaporthe poae	Magnaporthiopsis poae
Melampsora laricipopulina	Melampsora larici-populina
Phaeosphaeria nodorum	Parastagonospora nodorum
Pythium vexans	Phytophythium vexans

Table 1: Species whose names differ in the Ensembl and NCBI databases.

The phylogenetic representation of the relationship of each of the 337 organisms to each other as determined by the NCBI's Taxonomic Browser was retrieved. The order of the organisms shown in Appendix A reflects the phylogenetic relationship to one another as determined by this method.

2.4 Profile generation

We used BLAST [14] to search the protein complement of each of the 337 genomes for homologs to all 6,692 yeast proteins. This was done three separate times, each time using a different E-value threshold cutoff (no threshold, 1e-2, and 1e-10 for our most stringent match). Three types of profiles were generated: (i) **BRH** (best reciprocal hit), a binary profile, where the presence or absence of each yeast protein is predicted for each organism. For a yeast protein to be marked as existing in organism O, the protein in organism O that is the highest scoring hit to the yeast protein must also find that same yeast protein as one of its top two hits when searched against the yeast proteome. (ii) **Score profile**, a quantitative homology measure, where the score of the top hit to the yeast protein is divided by the score of its self-match (i.e., the score when the yeast protein is aligned to itself). This normalized score is a continuous variable from 0 to 1 (0 = non-existence; 1 = perfect match). (iii) **E-value profile**, calculated by taking the $1/\log_{10}(\text{E-value score})$, and capping the maximum value at 1. The E-value profile is on a 0-1 scale, where 0 is a perfect match, and 1 is non-existence. The profile of Alg5 (GPD synthase) [9, 10] was more promiscuous than expected (see below), so as bait we instead used the GPD-dependent glucosyltransferase Alg6 [9]. Distances of all profiles were calculated relative to Alg6, using a Jaccard-like metric for the BRH profile, and the Canberra distance metric for the others.

2.5 Filters

Annotations for each yeast protein were obtained from the Saccharomyces Genome Database (SGD) (http://downloads.yeastgenome.org/curation/chromosomal_feature/SGD_features.tab). Only records pertaining to ORFs were retained. Transmembrane domains were predicted for all yeast proteins using the TMHMM server (<http://www.cbs.dtu.dk/services/TMHMM/> [17]). The results were filtered to include only membrane proteins, and further refined to prioritize only multispansing proteins (≥ 3 transmembrane domains). We also filtered out proteins that were not found in the three higher eukaryotes. A final manual filter restricted the list to proteins that are known to be ER-localized, and those whose localization is ambiguous.

For each BLAST E-value threshold, we listed the top 50 hits that were identified in all three profile-generating methods (Appendix B) and ranked them according to the lowest rank that a protein has in any of the three lists. Although the lists differ slightly, the top candidates are common to all: (in rank order) Alg8, Mns1, Ale1, Erg24, Erc1, Ybr220C, Erg3, Gwt1, Alg2, Scs7, Ste24, Erg11, Sur2, Ydr338C, Alg3, Alg9, and Ykr051W. Depending on the

outcome of our biochemical analyses, we may pick additional candidates from the ranked lists that we generated, potentially including proteins that span the membrane only once. Such proteins may oligomerize to generate a pseudo-polytopic membrane protein that could have a transport function.

The presence of Alg8 as our top hit is unsurprising because it is required for synthesis of the glucose cap in G3M9-DLO and would be expected to co-evolve with Alg6. Three candidates (Ybr220C, Ydr338C, Ykr051W) are annotated as proteins of unknown function, two (Erc1, Ydr338C) are homologous to proteins that belong to the MOP exporter superfamily, and all except three (Gwt1, Alg2, Erg11) are non-essential for growth, consistent with glucosylation being non-essential in yeast. Interestingly, most have a lipid-related function, e.g. Ale1 is a lysophospholipid acyltransferase and Scs7 is a sphingolipid fatty acid α -hydroxylase, and three (Erg24, Ste24, Erg11) were identified in photocrosslinking studies using dolichyl phosphate analogs [18].

2.6 Notes on the profiling method and results

Phylogenetic profiling is a powerful method to identify functionally associated, co-evolving proteins such as enzymes that catalyze sequential steps in a biochemical pathway. While the method identifies the most significant co-evolutionary events, the generated profiles may not always match exactly. The most common causes for imperfect matches are a) incomplete protein annotation of a genome, b) different rates of evolution in independent lineages which may cause evolutionarily more distant proteins to outscore (in BLAST) the proteins which are actually closer, and (c) the possibility that in some organisms, a protein may have another, unrelated, function [11, 19].

Thus, Alg5 is found in a few organisms that do not synthesize glucosylated DLOs, suggesting novel roles for GPD [20] and/or an alternate function for Alg5 in these organisms. Likewise, the glucosyltransferase Alg8 is present in every Alg6-positive genome (268 of the 337 genomes we analyzed are Alg6-positive) as expected, but it is also present in 3 genomes (e.g. *Toxoplasma gondii*) that do not have Alg6. As Alg8 cannot add glucose unless Alg6 has first acted, and it has no other known function, its presence in these three genomes may be an evolutionary remnant. Another reason that profiles may not match exactly is if the GPD scramblase protein has another function, and is therefore subject to selective pressure unrelated to the glucosylation pathway. In this case, the candidate protein should be present if Alg6 is present, but may also be present when Alg6 is absent. For example, Erg24 is found in 19 of the 69 genomes that do not have Alg6. These genomes include those of trypanosomatids (e.g. *Trypanosoma brucei*) that do not synthesize glucosylated DLOs, but require the C14 sterol reductase activity of Erg24 to synthesize sterols. Thus, Erg24 may be a bifunctional protein with both GPD scramblase and sterol reductase activities. Despite the not-always-perfect profile matches, it is important to recognize that the power of the phylogenetic profiling approach used here lies in its ability to generate an unbiased, ranked list of candidates that allow us to prioritize our efforts to evaluate potential scramblase functions of the candidates.

3 Summary

Our top GPD scramblase candidates (in rank order, those in bold are essential for yeast growth) identified by phylogenetic profiling are Alg8, Mns1, Ale1, Erg24, Erc1, Ybr220C, Erg3, **Gwt1**, **Alg2**, Scs7, Ste24, **Erg11**, Sur2, Ydr338C, Alg3, Alg9, and Ykr051W. We are currently evaluating these candidates via *in vitro* and *in vivo* tests to identify the scramblase. This preprint will be updated once experimental data are available.

4 Acknowledgements

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References

- [1] B Schenk, F Fernandez, and CJ Waechter. The ins(ide) and out(side) of dolichyl phosphate biosynthesis and recycling in the endoplasmic reticulum. *Glycobiology*, 11(5):61R–70R, 2001.
- [2] S Sanyal and AK Menon. Flipping lipids: why an’ what’s the reason for? *ACS Chem Biol*, 4(11):895–909, 2009.
- [3] JS Rush and CJ Waechter. Transmembrane movement of a water-soluble analogue of mannosylphosphoryl-dolichol is mediated by an endoplasmic reticulum protein. *J Cell Biol*, 130(3):529–536, 1995.

- [4] JS Rush and CJ Waechter. Functional reconstitution into proteoliposomes and partial purification of a rat liver er transport system for a water-soluble analogue of mannosylphosphoryldolichol. *Biochemistry*, 43(23):7643–7652, 2004.
- [5] JS Rush, K van Leyen, O Ouerfelli, B Wolucka, and CJ Waechter. Transbilayer movement of glc-p-dolichol and its function as a glucosyl donor: protein-mediated transport of a water-soluble analog into sealed er vesicles from pig brain. *Glycobiology*, 8(12):1195–1205, 1998.
- [6] S Sanyal and AK Menon. Stereoselective transbilayer translocation of mannosyl phosphoryl dolichol by an endoplasmic reticulum flippase. *Proc Natl Acad Sci U S A*, 107(25):11289–11294, 2010.
- [7] SJ Turco, B Stetson, and PW Robbins. Comparative rates of transfer of lipid-linked oligosaccharides to endogenous glycoprotein acceptors in vitro. *Proc Natl Acad Sci U S A*, 74(10):4411–4414, 1977.
- [8] J Samuelson and PW Robbins. Effects of n-glycan precursor length diversity on quality control of protein folding and on protein glycosylation. *Semin Cell Dev Biol*, 41:121–128, 2015.
- [9] KW Runge, TC Huffaker, and PW Robbins. Two yeast mutations in glucosylation steps of the asparagine glycosylation pathway. *J Biol Chem*, 259(1):412–417, 1984.
- [10] S Heesen, L Lehle, A Weissmann, and M Aebi. Isolation of the alg5 locus encoding the udp-glucose:dolichyl-phosphate glucosyltransferase from *saccharomyces cerevisiae*. *Eur J Biochem*, 224(1):71–79, 1994.
- [11] M Pellegrini, EM Marcotte, MJ Thompson, D Eisenberg, and TO Yeates. Assigning protein functions by comparative genome analysis: protein phylogenetic profiles. *Proc Natl Acad Sci U S A*, 96(8):4285–4288, 1999.
- [12] G Dey, A Jaimovich, SR Collins, A Seki, and T Meyer. Systematic discovery of human gene function and principles of modular organization through phylogenetic profiling. *Cell Reports*, 15:S2211–1247, 2015.
- [13] J Samuelson, S Banerjee, P Magnelli, J Cui, DJ Kelleher, R Gilmore, and PW Robbins. The diversity of dolichol-linked precursors to asn-linked glycans likely results from secondary loss of sets of glucosyltransferases. *Proc Natl Acad Sci U S A*, 102(5):1548–1553, 2005.
- [14] SF Altschul, TL Madden, AA Schäffer, J Zhang, Z Zhang, W Miller, and DJ Lipman. Gapped blast and psi-blast: a new generation of protein database search programs. *Nucleic Acids Res*, 25(17):3389–3402, 1997.
- [15] N Škunca and C Dessimoz. Phylogenetic profiling: How much input data is enough? *PLoS One*, 10(2):e0114701, 2015.
- [16] S Cokus, S Mizutani, and M Pellegrini. An improved method for identifying functionally linked proteins using phylogenetic profiles. *BMC Bioinformatics*, 8(Suppl 4):S7, 2007.
- [17] A Krogh, B Larsson, G von Heijne, and ELL Sonnhammer. Predicting transmembrane protein topology with a hidden markov model: application to complete genomes. *J Mol Biol*, 305(3):567–580, 2001.
- [18] JS Rush, T Subramanian, KL Subramanian, FO Onono, CJ Waechter, and HP Spielmann. Novel citronellyl-based photoprobes designed to identify er proteins interacting with dolichyl phosphate in yeast and mammalian cells. *Curr Chem Biol*, 9(2):123–141, 2015.
- [19] MK Basu, JD Selengut, and DH Haft. Prophyl: partial phylogenetic profiling to guide protein family construction and assignment of biological process. *BMC Bioinformatics*, 12:434–447, 2011.
- [20] KA Grabinska, SK Ghosh, Z Guan, J Cui, CR Raetz, PW Robbins, and J Samuelson. Dolichyl-phosphate-glucose is used to make o-glycans on glycoproteins of *trichomonas vaginalis*. *Eukaryotic Cell*, 7(8):1344–1351, 2008.

Appendix A: List of organisms used in phylogenetic profiling analysis, ordered by phylogenetic relatedness

1. *Guillardia theta*
2. *Bigelowiella natans*
3. *Reticulomyxa filosa*
4. *Perkinsus marinus* atcc 50983
5. *Oxytricha trifallax* gca 000295675
6. *Stylonychia lemnae*
7. *Tetrahymena thermophila*
8. *Paramecium tetraurelia*
9. *Babesia equi* strain wa
10. *Babesia bovis*
11. *Plasmodium reichenowi*
12. *Plasmodium falciparum*
13. *Plasmodium inui* san antonio 1
14. *Plasmodium vivax*
15. *Plasmodium knowlesi*
16. *Plasmodium cynomolgi* strain b
17. *Plasmodium yoelii* yoelii
18. *Plasmodium vinckei* petteri
19. *Plasmodium chabaudi*
20. *Plasmodium berghei*
21. *Gregarina niphandrodes*
22. *Hammondia hammondi*
23. *Toxoplasma gondii*
24. *Cryptosporidium muris* rn66
25. *Cryptosporidium parvum* iowa ii
26. *Eimeria necatrix*
27. *Eimeria tenella* gca 000499545
28. *Naegleria gruberi*
29. *Polysphondylium pallidum* pn500
30. *Dictyostelium fasciculatum*
31. *Dictyostelium purpureum*
32. *Entamoeba nuttalli* p19
33. *Entamoeba dispar* saw760
34. *Entamoeba invadens* ip1
35. *Entamoeba histolytica*
36. *Acanthamoeba castellanii* str neff
37. *Spironucleus salmonicida*
38. *Giardia intestinalis*
39. *Giardia lamblia*
40. *Trichomonas vaginalis* g3
41. *Phytomonas* sp isolate em1
42. *Angomonas deanei*
43. *Trypanosoma rangeli* sc58
44. *Trypanosoma cruzi*
45. *Trypanosoma brucei*
46. *Leishmania mexicana* mhom gt 2001 u1103
47. *Leishmania major*
48. *Leishmania infantum* jpcm5
49. *Leishmania donovani*
50. *Leishmania panamensis*
51. *Leishmania braziliensis* mhom br 75 m2904
52. *Salpingoeca rosetta*
53. *Monosiga brevicollis* mx1
54. *Canis familiaris*
55. *Mus musculus*
56. *Homo sapiens*
57. *Rhizophagus irregularis* daom 181602
58. *Microsporidia* sp ugp3
59. *Nematocida parisii* ertm1
60. *Rozella allomyces* csf55
61. *Batrachochytrium dendrobatidis* jam81
62. *Wallemia ichthyophaga* exf 994
63. *Wallemia sebi* cbs 633 66
64. *Mixia osmundae* iam 14324
65. *Melampsora laricipopulina*
66. *Puccinia triticina*
67. *Puccinia graminis*
68. *Rhodosporidium toruloides* np11
69. *Microbotryum violaceum*
70. *Tilletiaria anomala* ubc 951
71. *Pseudozyma brasiliensis*
72. *Pseudozyma hubeiensis* sy62
73. *Pseudozyma aphidis* dsm 70725
74. *Pseudozyma antarctica*
75. *Sporisorium reilianum*
76. *Ustilago hordei*
77. *Ustilago maydis*
78. *Dacryopinax* sp djm 731 ss1
79. *Heterobasidion irregulare* tc 32 1
80. *Rhizoctonia solani* 123e
81. *Botryobasidium botryosum* fd 172 ss1
82. *Tulasnella calospora* mut 4182
83. *Gloeophyllum trabeum* atcc 11539
84. *Serendipita vermifera* maff 305830
85. *Piriformospora indica* dsm 11827
86. *Fibroporia radiculosa*
87. *Phanerochaete carnosa* hhb 10118 sp
88. *Phlebiopsis gigantea* 11061 1 cr5 6
89. *Ceriporiopsis subvermispora* b
90. *Fomitopsis pinicola* fp 58527 ss1
91. *Trametes cinnabarina*
92. *Jaapia argillacea* mucl 33604
93. *Piloderma croceum* f 1598
94. *Paxillus rubicundulus* ve08 2h10
95. *Serpula lacrymans* var *lacrymans* s7 3
96. *Scleroderma citrinum* foug a
97. *Pisolithus microcarpus* 441
98. *Pisolithus tinctorius* marx 270
99. *Suillus luteus* uh slu lm8 n1
100. *Moniliophthora roreri* mca 2997
101. *Galerina marginata* cbs 339 88
102. *Hebeloma cylindrosporum* h7
103. *Amanita muscaria* koide bx008
104. *Laccaria amethystina* laam 08 1
105. *Laccaria bicolor* s238n h82
106. *Coprinopsis cinerea* okayama7 130
107. *Agaricus bisporus* var *bisporus* h97
108. *Schizophyllum commune* h4 8
109. *Pleurotus ostreatus* pc15
110. *Trichosporon asahii* var *asahii* cbs 2479
111. *Cryptococcus gattii* r265
112. *Cryptococcus neoformans*
113. *Tuber melanosporum*
114. *Dactylellina haptotyla* cbs 200 50
115. *Arthrobotrys oligospora* atcc 24927
116. *Pseudogymnoascus destructans* 20631 21

117. *Pseudogymnoascus pannorum* vkm f 103
118. *Oidiodendron maius* zn
119. *Marssonina brunnea* f sp multigermtubi mb m1
120. *Glarea lozoyensis* atcc 20868
121. *Sclerotinia borealis* f 4157
122. *Botrytis cinerea*
123. *Erysiphe necator*
124. *Blumeria graminis*
125. *Pestalotiopsis fici* w106 1
126. *Eutypa lata* ucrell
127. *Togninia minima* ucrpa7
128. *Grosmannia clavigera* kw1407
129. *Ophiostoma piceae* uamh 11346
130. *Sporothrix brasiliensis* 5110
131. *Sporothrix schenckii* atcc 58251
132. *Magnaporthe oryzae*
133. *Magnaporthe poae*
134. *Gaeumannomyces graminis*
135. *Myceliophthora thermophila* atcc 42464
136. *Chaetomium thermophilum* var *thermophilum* dsm 1495
137. *Chaetomium globosum* cbs 148 51
138. *Thielavia terrestris* nr1 8126
139. *Podospora anserina* s mat
140. *Sordaria macrospora*
141. *Neurospora tetrasperma* fgsc 2508
142. *Neurospora crassa*
143. *Scedosporium apiospermum*
144. *Verticillium alfalfae* vams 102
145. *Verticillium dahliae*
146. *Colletotrichum sublineola*
147. *Colletotrichum fioriniae* pj7
148. *Colletotrichum gloeosporioides*
149. *Colletotrichum higginsianum*
150. *Colletotrichum graminicola*
151. *Colletotrichum orbiculare*
152. *Stachybotrys chlorohalonata* ibt 40285
153. *Stachybotrys chartarum* ibt 40288
154. *Beauveria bassiana* arsef 2860
155. *Cordyceps militaris* cm01
156. *Trichoderma atroviride* imi 206040
157. *Trichoderma reesei*
158. *Trichoderma virens*
159. *Fusarium solani*
160. *Fusarium pseudograminearum*
161. *Fusarium graminearum*
162. *Fusarium oxysporum*
163. *Fusarium verticillioides*
164. *Fusarium fujikuroi*
165. *Torrubiella hemipterigena*
166. *Metarhizium majus* arsef 297
167. *Metarhizium robertsii*
168. *Metarhizium brunneum* arsef 3297
169. *Metarhizium guizhouense* arsef 977
170. *Metarhizium acridum* cqma 102
171. *Metarhizium album* arsef 1941
172. *Metarhizium anisopliae*
173. *Claviceps purpurea* 20 1
174. *Ustilagoideia virens*
175. *Acremonium chrysogenum* atcc 11550
176. *Endocarpon pusillum* z07020
177. *Cyphellophora europaea* cbs 101466
178. *Rhinocladiella mackenziei* cbs 650 93
179. *Cladophialophora psammophila* cbs 110553
180. *Cladophialophora immunda*
181. *Cladophialophora yegresii* cbs 114405
182. *Cladophialophora bantiana* cbs 173 52
183. *Cladophialophora carrionii* cbs 160 54
184. *Coniosporium apollinis* cbs 100218
185. *Fonsecaea pedrosoi* cbs 271 37
186. *Exophiala aquamarina* cbs 119918
187. *Exophiala sideris*
188. *Exophiala xenobiotica*
189. *Exophiala oligosperma*
190. *Exophiala mesophila*
191. *Exophiala spinifera*
192. *Exophiala dermatitidis* nih ut8656
193. *Capronia coronata* cbs 617 96
194. *Capronia epimyces* cbs 606 96
195. *Capronia semiimmersa*
196. *Byssoscleromyces spectabilis* no 5
197. *Talaromyces marneffeii* atcc 18224
198. *Talaromyces stipitatus* atcc 10500
199. *Neosartorya fischeri*
200. *Penicillium rubens* wisconsin 54 1255
201. *Penicillium oxalicum* 114 2
202. *Penicillium solitum*
203. *Penicillium italicum*
204. *Penicillium digitatum* pd1
205. *Penicillium expansum*
206. *Aspergillus fumigatus*
207. *Aspergillus ruber* cbs 135680
208. *Aspergillus nidulans*
209. *Aspergillus terreus*
210. *Aspergillus oryzae*
211. *Aspergillus niger*
212. *Aspergillus flavus*
213. *Aspergillus clavatus*
214. *Coccidioides posadasii* str silveira
215. *Paracoccidioides* sp lutzii pb01
216. *Paracoccidioides brasiliensis* pb03
217. *Uncinocarpus reesii* 1704
218. *Microsporium gypseum* cbs 118893
219. *Arthroderma otae* cbs 113480
220. *Arthroderma benhamiae* cbs 112371
221. *Trichophyton soudanense* cbs 452 61
222. *Trichophyton equinum* cbs 127 97
223. *Trichophyton verrucosum* hki 0517
224. *Trichophyton interdigitale* h6
225. *Trichophyton tonsurans* cbs 112818
226. *Trichophyton rubrum* d6
227. *Blastomyces dermatitidis* atcc 18188
228. *Histoplasma capsulatum* h88
229. *Verruconis gallopava*
230. *Neofusicoccum parvum* ucrnp2
231. *Macrophomina phaseolina* ms6
232. *Baudoinia compniacensis* uamh 10762
233. *Zymoseptoria tritici*
234. *Sphaerulina musiva* so2202
235. *Pseudocercospora fijiensis* cirad86
236. *Dothistroma septosporum*
237. *Aureobasidium subglaciale* exf 2481
238. *Aureobasidium melanogenum* cbs 110374
239. *Aureobasidium pullulans* exf 150

240. *Phaeosphaeria nodorum*
241. *Leptosphaeria maculans*
242. *Setosphaeria turcica* et28a
243. *Pyrenophora teres*
244. *Pyrenophora tritici-repentis*
245. *Bipolaris oryzae* atcc 44560
246. *Bipolaris sorokiniana* nd90pr
247. *Bipolaris victoriae* fi3
248. *Bipolaris zeicola* 26 r 13
249. *Bipolaris maydis* atcc 48331
250. *Kuraishia capsulata* cbs 1993
251. *Clavispora lusitanae* atcc 42720
252. *Yarrowia lipolytica*
253. *Saccharomyces arboricola* h 6
254. *Saccharomycetaceae* sp ashbya aceri
255. *Kazachstania naganishii* cbs 8797
256. *Kazachstania africana* cbs 2517
257. *Lachancea lanzarotensis*
258. *Lachancea thermotolerans* cbs 6340
259. *Tetrapisispora blattae* cbs 6284
260. *Tetrapisispora phaffii* cbs 4417
261. *Vanderwaltozyma polyspora* dsm 70294
262. *Eremothecium cymbalariae* dbvpg 7215
263. *Ashbya gossypii*
264. *Kluyveromyces lactis*
265. *Naumovozyma dairenensis* cbs 421
266. *Naumovozyma castellii* cbs 4309
267. *Candida glabrata*
268. *Zygosaccharomyces rouxii*
269. *Zygosaccharomyces bailii* isa1307
270. *Torulaspora delbrueckii*
271. *Wickerhamomyces ciferrii*
272. *Komagataella pastoris*
273. *Spathaspora passalidarum* nr1 y 27907
274. *Candida tenuis* atcc 10573
275. *Lodderomyces elongisporus* nr1 yb 4239
276. *Candida orthopsilosis* co 90 125
277. *Candida dubliniensis* cd36
278. *Candida tropicalis* mya 3404
279. *Candida albicans* wo 1
280. *Debaryomyces hansenii* cbs767
281. *Meyerozyma guilliermondii* atcc 6260
282. *Scheffersomyces stipitis* cbs 6054
283. *Millerozyma farinosa* cbs 7064
284. *Ogataea parapolyomorpha* dl 1
285. *Pichia kudriavzevii*
286. *Pneumocystis murina* b123
287. *Schizosaccharomyces cryophilus*
288. *Schizosaccharomyces octosporus*
289. *Schizosaccharomyces japonicus*
290. *Schizosaccharomyces pombe*
291. *Aureococcus anophagefferens*
292. *Blastocystis hominis*
293. *Albugo laibachii*
294. *Aphanomyces invadans*
295. *Aphanomyces astaci*
296. *Saprolegnia diclina* vs20
297. *Saprolegnia parasitica* cbs 223 65
298. *Pythium iwayamai*
299. *Pythium arrhenomanes*
300. *Pythium ultimum*
301. *Pythium aphanidermatum*
302. *Pythium irregulare*
303. *Pythium vexans*
304. *Hyaloperonospora arabidopsidis*
305. *Phytophthora kernoviae*
306. *Phytophthora ramorum*
307. *Phytophthora lateralis*
308. *Phytophthora sojae*
309. *Phytophthora parasitica*
310. *Phytophthora infestans*
311. *Thalassiosira oceanica*
312. *Thalassiosira pseudonana*
313. *Phaeodactylum tricornutum*
314. *Galdieria sulphuraria*
315. *Chondrus crispus*
316. *Nitrosopumilus maritimus* scm1
317. *Caldisphaera lagunensis* dsm 15908
318. *Metallosphaera cuprina* ar 4
319. *Metallosphaera sedula*
320. *Sulfolobus tokodaii* str 7
321. *Sulfolobus islandicus* hve10 4
322. *Sulfolobus solfataricus* 98 2
323. *Sulfolobus acidocaldarius* dsm 639
324. *Hyperthermus butylicus* dsm 5456
325. *Thermogladius cellulolyticus* 1633
326. *Ignicoccus hospitalis* kin4 i
327. *Staphylothermus hellenicus* dsm 12710
328. *Staphylothermus marinus* fl
329. *Desulfurococcus kamchatkensis* 1221n
330. *Desulfurococcus mucosus* dsm 2162
331. *Thermophilum pendens* hrk 5
332. *Aciduliprofundum boonei* t469 gca 000025665 1
333. *Methanosaeta thermophila* pt
334. *Methanothermobacter marburgensis* str marburg
335. *Methanobrevibacter ruminantium* m1
336. *Methanosphaera stadtmanae* dsm 3091
337. *Methanothermobacter fervidus* dsm 2088

Appendix B: Ranked top 50 genes for each BLAST E-value threshold

Ranked list, Top50 (TM-containing, ER-localized only), BLAST threshold: 0

Name	Rank	#TMs	SGD name	Description
ALG6	1	10	YOR002W	dolichyl-P-Glc:Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase Alpha 1,3 glucosyltransferase; involved in transfer of oligosaccharides from dolichyl pyrophosphate to asparagine residues of proteins during N-linked protein glycosylation; mutations in human ortholog are associated with disease
ALG8	2	12	YOR067C	dolichyl-P-Glc:Glc1Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase—YOR29-18

Name	Rank	#TMs	SGD name	Description
MNS1	4	1	YJR131W	Glucosyl transferase; involved in N-linked glycosylation; adds glucose to the dolichol-linked oligosaccharide precursor prior to transfer to protein during lipid-linked oligosaccharide biosynthesis; similar to Alg6p mannosyl-oligosaccharide 1,2- α -mannosidase
ALE1	14	7	YOR175C	Alpha-1,2-mannosidase; involved in ER-associated protein degradation (ERAD); catalyzes the removal of one mannose residue from a glycosylated protein, converting the modification from Man9GlcNAc to Man8GlcNAc; catalyzes the last step in glycoprotein maturation in the ER and is critical for ER protein degradation lysophospholipid acyltransferase—LCA1—LPT1—SLC4
ERG24	18	8	YNL280C	Broad-specificity lysophospholipid acyltransferase; part of MBOAT family of membrane-bound O-acyltransferases; key component of Lands cycle; may have role in fatty acid exchange at sn-2 position of mature glycerophospholipids delta(14)-sterol reductase
AVT3	35	10	YKL146W	C-14 sterol reductase; acts in ergosterol biosynthesis; mutants accumulate the abnormal sterol ignosterol (ergosta-8,14 dienol), and are viable under anaerobic growth conditions but inviable on rich medium under aerobic conditions YKL146W
CSC1	39	11	YLR241W	Vacuolar transporter; exports large neutral amino acids from the vacuole; member of a family of seven <i>S. cerevisiae</i> genes (AVT1-7) related to vesicular GABA-glycine transporters YLR241W
BOR1	40	10	YNL275W	Calcium permeable gated cation channel; may be involved in detoxification; similar to Arabidopsis CSC1 YNL275W
ERG11	45	2	YHR007C	Boron efflux transporter of the plasma membrane; binds HCO ₃ ⁻ , I ⁻ , Br ⁻ , NO ₃ ⁻ and Cl ⁻ ; has similarity to the characterized boron efflux transporter <i>A. thaliana</i> BOR1 sterol 14-demethylase—CYP51
ERV46	46	1	YAL042W	Lanosterol 14- α -demethylase; catalyzes C-14 demethylation of lanosterol to form 4,4'-dimethyl cholesta-8,14,24-triene-3- β -ol in ergosterol biosynthesis pathway; transcriptionally down-regulated when ergosterol is in excess; member of cytochrome P450 family; associated and coordinately regulated with the P450 reductase Ncp1p FUN9
ERG3	50	3	YLR056W	Protein localized to COPII-coated vesicles; forms a complex with Erv41p; involved in the membrane fusion stage of transport C-5 sterol desaturase—PSO6—SYR1
YPR003C	52	10	YPR003C	C-5 sterol desaturase; glycoprotein that catalyzes the introduction of a C-5(6) double bond into episterol, a precursor in ergosterol biosynthesis; transcriptionally down-regulated when ergosterol is in excess; mutants are viable, but cannot grow on non-fermentable carbon sources; substrate of HRD ubiquitin ligase YPR003C
STE24	64	4	YJR117W	Putative sulfate permease; physically interacts with Hsp82p; green fluorescent protein (GFP)-fusion protein localizes to the ER; YPR003C is not an essential gene zinc metalloprotease—PIO2—AFC1
PMT5	65	11	YDL093W	Highly conserved zinc metalloprotease; functions in two steps of a-factor maturation, C-terminal CAAX proteolysis and the first step of N-terminal proteolytic processing; contains multiple transmembrane spans; human homolog ZMPSTE24 implicated in mandibuloacral dysplasia (MAD), and can complement yeast null mutant putative dolichyl-phosphate-mannose-protein mannosyltransferase PMT5
ALG1	67	1	YBR110W	Protein O-mannosyltransferase; transfers mannose residues from dolichyl phosphate-D-mannose to protein serine/threonine residues; acts in a complex with Pmt3p, can instead interact with Pmt2p in some conditions; target for new antifungals chitobiosyldiphosphodolichol beta-1,4 mannosyltransferase
AUR1	67	6	YKL004W	Mannosyltransferase; involved in asparagine-linked glycosylation in the endoplasmic reticulum (ER); essential for viability; human homolog ALG1 complements yeast null mutant inositol phosphorylceramide synthase
ZRT2	72	7	YLR130C	Phosphatidylinositol:ceramide phosphoinositol transferase; required for sphingolipid synthesis; can mutate to confer aureobasidin A resistance; also known as IPC synthase low-affinity Zn(2+) transporter ZRT2
ZRT1	80	8	YGL255W	Low-affinity zinc transporter of the plasma membrane; transcription is induced under low-zinc conditions by the Zap1p transcription factor high-affinity Zn(2+) transporter ZRT1
ATG15	83	1	YCR068W	High-affinity zinc transporter of the plasma membrane; responsible for the majority of zinc uptake; transcription is induced under low-zinc conditions by the Zap1p transcription factor triglyceride lipase ATG15—CVT17—AUT5
				Putative lipase required for lysis of autophagic and Cvt bodies; targeted to intravacuolar vesicles during autophagy via the multivesicular body (MVB) pathway; required for the maintenance of lipid droplet quantity after the diauxic shift; regulates lipolysis

Name	Rank	#TMs	SGD name	Description
ERG4	84	7	YGL012W	delta(24(24(1)))-sterol reductase
LAC1	85	7	YKL008C	C-24(28) sterol reductase; catalyzes the final step in ergosterol biosynthesis; mutants are viable, but lack ergosterol sphingosine N-acyltransferase LAC1—DGT1 Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lag1p; LAC1 has a paralog, LAG1, that arose from the whole genome duplication
ERG9	86	1	YHR190W	bifunctional farnesyl-diphosphate farnesyltransferase/squalene synthase Farnesyl-diphosphate farnesyl transferase (squalene synthase); joins two farnesyl pyrophosphate moieties to form squalene in the sterol biosynthesis pathway
NHA1	86	9	YLR138W	YLR138W Na ⁺ /H ⁺ antiporter; involved in sodium and potassium efflux through the plasma membrane; required for alkali cation tolerance at acidic pH
ALG3	90	9	YBL082C	dolichyl-P-Man:Man(5)GlcNAc(2)-PP-dolichol alpha-1,3-mannosyltransferase—RHK1 Dolichol-P-Man dependent alpha(1-3) mannosyltransferase; involved in the synthesis of dolichol-linked oligosaccharide donor for N-linked glycosylation of proteins
MEP3	92	11	YPR138C	ammonium permease MEP3 Ammonium permease of high capacity and low affinity; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation ammonia permease; MEP3 has a paralog, MEP1, that arose from the whole genome duplication
DGA1	101	1	YOR245C	diacylglycerol O-acyltransferase Diacylglycerol acyltransferase; catalyzes the terminal step of triacylglycerol (TAG) formation, acylates diacylglycerol using acyl-CoA as an acyl donor; Lro1p and Dgalp can O-acylate ceramides; localized to lipid particles
VCX1	101	11	YDL128W	MNR1—HUM1 Vacuolar membrane antiporter with Ca ²⁺ /H ⁺ and K ⁺ /H ⁺ exchange activity; involved in control of cytosolic Ca ²⁺ and K ⁺ concentrations; has similarity to sodium/calcium exchangers, including the bovine Na ⁺ /Ca ²⁺ ,K ⁺ antiporter
YKR051W	103	7	YKR051W	YKR051W Putative protein of unknown function
MEP1	104	11	YGR121C	ammonium permease MEP1—AMT1 Ammonium permease; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation; human homolog RHCG complements yeast null mutant; mutations in human homolog RHCG implicated in metabolic acidosis; MEP1 has a paralog, MEP3, that arose from the whole genome duplication
CHS7	105	7	YHR142W	YHR142W Protein of unknown function; may be involved in chitin biosynthesis by regulation of Chs3p export from the ER; relocalizes from bud neck to ER upon DNA replication stress
YVC1	107	7	YOR087W	TRPY1—YOR088W Vacuolar cation channel; mediates release of Ca(2+) from the vacuole in response to hyperosmotic shock
GWT1	108	12	YJL091C	YJL091C Protein involved in the inositol acylation of GlcN-PI; the inositol acylation of glucosaminyl phosphatidylinositol (GlcN-PI) forms glucosaminyl(acyl)phosphatidylinositol (GlcN(acyl)PI), an intermediate in the biosynthesis of glycosylphosphatidylinositol (GPI) anchors
ERG1	110	2	YGR175C	squalene monooxygenase Squalene epoxidase; catalyzes the epoxidation of squalene to 2,3-oxidosqualene; plays an essential role in the ergosterol-biosynthesis pathway and is the specific target of the antifungal drug terbinafine
HFD1	110	1	YMR110C	YMR110C Hexadecenal dehydrogenase; involved in the conversion of sphingosine 1-phosphate breakdown product hexadecenal to hexadecenoic acid; located in the mitochondrial outer membrane and also in lipid particles; similar to ALDH3A2, a human fatty aldehyde dehydrogenase (FALDH) mutated in Sjogren-Larsson syndrome, a neurocutaneous disorder
PGA3	110	2	YML125C	NQR1 Putative cytochrome b5 reductase, localized to the plasma membrane; may be involved in regulation of lifespan; required for maturation of Gas1p and Pho8p, proposed to be involved in protein trafficking; PGA3 has a paralog, AIM33, that arose from the whole genome duplication
CBR1	112	1	YIL043C	CBR5 Cytochrome b reductase; not essential for viability; also detected in mitochondria; mutation in conserved NADH binding domain of the human ortholog results in type I methemoglobinemia
LAG1	112	7	YHL003C	sphingosine N-acyltransferase LAG1

Name	Rank	#TMs	SGD name	Description
ERV29	114	7	YGR284C	Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lac1p; forms ER foci upon DNA replication stress; homolog of human CERS2, a tumor metastasis suppressor gene whose silencing enhances invasion/metastasis of prostate cancer cells; LAG1 has a paralog, LAC1, that arose from the whole genome duplication
HNMI	114	12	YGL077C	CTR1
OLE1	114	4	YGL055W	stearoyl-CoA 9-desaturase—MDM2
FTH1	115	7	YBR207W	YBR207W
SCS7	115	3	YMR272C	fatty acid alpha-hydroxylase—FAH1
SAC1	116	2	YKL212W	phosphatidylinositol-3-phosphatase SAC1—RSD1
TNA1	117	12	YGR260W	YGR260W
YGR149W	118	7	YGR149W	YGR149W
TPO1	119	12	YLL028W	YLL028W
UBC6	119	1	YER100W	E2 ubiquitin-conjugating protein UBC6—DOA2
FTR1	120	7	YER145C	high-affinity iron permease FTR1
GAB1	122	8	YLR459W	CDC91
MEP2	122	11	YNL142W	ammonium permease MEP2

Ranked list, Top50 (TM-containing, ER-localized only), BLAST threshold: 1e-2

Name	Rank	#TMs	SGD name	Description
ALG6	1	10	YOR002W	dolichyl-P-Glc:Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase Alpha 1,3 glucosyltransferase; involved in transfer of oligosaccharides from dolichyl pyrophosphate to asparagine residues of proteins during N-linked protein glycosylation; mutations in human ortholog are associated with disease
ALG8	2	12	YOR067C	dolichyl-P-Glc:Glc1Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase—YOR29-18 Glucosyl transferase; involved in N-linked glycosylation; adds glucose to the dolichol-linked oligosaccharide precursor prior to transfer to protein during lipid-linked oligosaccharide biosynthesis; similar to Alg6p
MNS1	4	1	YJR131W	mannosyl-oligosaccharide 1,2-alpha-mannosidase Alpha-1,2-mannosidase; involved in ER-associated protein degradation (ERAD); catalyzes the removal of one mannose residue from a glycosylated protein, converting the modification from Man9GlcNAc to Man8GlcNAc; catalyzes the last step in glycoprotein maturation in the ER and is critical for ER protein degradation
ALE1	8	7	YOR175C	lysophospholipid acyltransferase—LCA1—LPT1—SLC4 Broad-specificity lysophospholipid acyltransferase; part of MBOAT family of membrane-bound O-acyltransferases; key component of Lands cycle; may have role in fatty acid exchange at sn-2 position of mature glycerophospholipids
ERG24	18	8	YNL280C	delta(14)-sterol reductase C-14 sterol reductase; acts in ergosterol biosynthesis; mutants accumulate the abnormal sterol ignosterol (ergosta-8,14 dienol), and are viable under anaerobic growth conditions but inviable on rich medium under aerobic conditions
ERG3	23	3	YLR056W	C-5 sterol desaturase—PSO6—SYR1 C-5 sterol desaturase; glycoprotein that catalyzes the introduction of a C-5(6) double bond into episterol, a precursor in ergosterol biosynthesis; transcriptionally down-regulated when ergosterol is in excess; mutants are viable, but cannot grow on non-fermentable carbon sources; substrate of HRD ubiquitin ligase
BOR1	39	10	YNL275W	YNL275W Boron efflux transporter of the plasma membrane; binds HCO ₃ ⁻ , I ⁻ , Br ⁻ , NO ₃ ⁻ and Cl ⁻ ; has similarity to the characterized boron efflux transporter A. thaliana BOR1
CSC1	39	11	YLR241W	YLR241W Calcium permeable gated cation channel; may be involved in detoxification; similar to Arabidopsis CSC1
AVT3	40	10	YKL146W	YKL146W Vacuolar transporter; exports large neutral amino acids from the vacuole; member of a family of seven S. cerevisiae genes (AVT1-7) related to vesicular GABA-glycine transporters
ERG11	45	2	YHR007C	sterol 14-demethylase—CYP51 Lanosterol 14-alpha-demethylase; catalyzes C-14 demethylation of lanosterol to form 4,4"-dimethyl cholesta-8,14,24-triene-3-beta-ol in ergosterol biosynthesis pathway; transcriptionally down-regulated when ergosterol is in excess; member of cytochrome P450 family; associated and coordinately regulated with the P450 reductase Ncp1p
ERV46	49	1	YAL042W	FUN9 Protein localized to COPII-coated vesicles; forms a complex with Erv41p; involved in the membrane fusion stage of transport
YPR003C	55	10	YPR003C	YPR003C Putative sulfate permease; physically interacts with Hsp82p; green fluorescent protein (GFP)-fusion protein localizes to the ER; YPR003C is not an essential gene
ERG4	59	7	YGL012W	delta(24(24(1)))—sterol reductase C-24(28) sterol reductase; catalyzes the final step in ergosterol biosynthesis; mutants are viable, but lack ergosterol
PMT5	64	11	YDL093W	putative dolichyl-phosphate-mannose-protein mannosyltransferase PMT5 Protein O-mannosyltransferase; transfers mannose residues from dolichyl phosphate-D-mannose to protein serine/threonine residues; acts in a complex with Pmt3p, can instead interact with Pmt2p in some conditions; target for new antifungals
ALG1	67	1	YBR110W	chitobiosyldiphosphodolichol beta-1,4 mannosyltransferase Mannosyltransferase; involved in asparagine-linked glycosylation in the endoplasmic reticulum (ER); essential for viability; human homolog ALG1 complements yeast null mutant
AUR1	68	6	YKL004W	inositol phosphorylceramide synthase Phosphatidylinositol:ceramide phosphoinositol transferase; required for sphingolipid synthesis; can mutate to confer aureobasidin A resistance; also known as IPC synthase
STE24	70	4	YJR117W	zinc metalloprotease—PIO2—AFC1 Highly conserved zinc metalloprotease; functions in two steps of a-factor maturation, C-terminal CAAX proteolysis and the first step of N-terminal proteolytic processing; contains multiple transmembrane spans; human homolog ZMPSTE24 implicated in mandibuloacral dysplasia (MAD), and can complement yeast null mutant
ZRT2	73	7	YLR130C	low-affinity Zn(2+) transporter ZRT2

Name	Rank	#TMs	SGD name	Description
MEP3	74	11	YPR138C	Low-affinity zinc transporter of the plasma membrane; transcription is induced under low-zinc conditions by the Zap1p transcription factor ammonium permease MEP3 Ammonium permease of high capacity and low affinity; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation ammonia permease; MEP3 has a paralog, MEP1, that arose from the whole genome duplication
ZRT1	80	8	YGL255W	high-affinity Zn(2+) transporter ZRT1 High-affinity zinc transporter of the plasma membrane; responsible for the majority of zinc uptake; transcription is induced under low-zinc conditions by the Zap1p transcription factor
ATG15	83	1	YCR068W	triglyceride lipase ATG15—CVT17—AUT5 Putative lipase required for lysis of autophagic and Cvt bodies; targeted to intravacuolar vesicles during autophagy via the multivesicular body (MVB) pathway; required for the maintenance of lipid droplet quantity after the diauxic shift; regulates lipolysis
MEP1	84	11	YGR121C	ammonium permease MEP1—AMT1 Ammonium permease; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation; human homolog RHCG complements yeast null mutant; mutations in human homolog RHCG implicated in metabolic acidosis; MEP1 has a paralog, MEP3, that arose from the whole genome duplication
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NHA1	86	9	YLR138W	YLR138W Na ⁺ /H ⁺ antiporter; involved in sodium and potassium efflux through the plasma membrane; required for alkali cation tolerance at acidic pH
ALG3	88	9	YBL082C	dolichyl-P-Man:Man(5)GlcNAc(2)-PP-dolichol alpha-1,3-mannosyltransferase—RHK1 Dolichol-P-Man dependent alpha(1-3) mannosyltransferase; involved in the synthesis of dolichol-linked oligosaccharide donor for N-linked glycosylation of proteins
LAC1	93	7	YKL008C	sphingosine N-acyltransferase LAC1—DGT1 Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lag1p; LAC1 has a paralog, LAG1, that arose from the whole genome duplication
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DGA1	100	1	YOR245C	diacylglycerol O-acyltransferase Diacylglycerol acyltransferase; catalyzes the terminal step of triacylglycerol (TAG) formation, acylates diacylglycerol using acyl-CoA as an acyl donor; Lro1p and Dgalp can O-acylate ceramides; localized to lipid particles
VCX1	101	11	YDL128W	MNR1—HUM1 Vacuolar membrane antiporter with Ca ²⁺ /H ⁺ and K ⁺ /H ⁺ exchange activity; involved in control of cytosolic Ca ²⁺ and K ⁺ concentrations; has similarity to sodium/calcium exchangers, including the bovine Na ⁺ /Ca ²⁺ ,K ⁺ antiporter
YKR051W	102	7	YKR051W	YKR051W Putative protein of unknown function
GWT1	108	12	YJL091C	YJL091C Protein involved in the inositol acylation of GlcN-PI; the inositol acylation of glucosaminyl phosphatidylinositol (GlcN-PI) forms glucosaminyl(acyl)phosphatidylinositol (GlcN(acyl)PI), an intermediate in the biosynthesis of glycosylphosphatidylinositol (GPI) anchors
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LAG1	112	7	YHL003C	sphingosine N-acyltransferase LAG1 Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lac1p; forms ER foci upon DNA replication stress; homolog of human CERS2, a tumor metastasis suppressor gene whose silencing enhances invasion/metastasis of prostate cancer cells; LAG1 has a paralog, LAC1, that arose from the whole genome duplication

Name	Rank	#TMs	SGD name	Description
ERV29	114	7	YGR284C	YGR284C Protein localized to COPII-coated vesicles; involved in vesicle formation and incorporation of specific secretory cargo; protein abundance increases in response to DNA replication stress
OLE1	114	4	YGL055W	stearoyl-CoA 9-desaturase—MDM2 Delta(9) fatty acid desaturase; required for monounsaturated fatty acid synthesis and for normal distribution of mitochondria
CHS7	115	7	YHR142W	YHR142W Protein of unknown function; may be involved in chitin biosynthesis by regulation of Chs3p export from the ER; relocalizes from bud neck to ER upon DNA replication stress
SCS7	115	3	YMR272C	fatty acid alpha-hydroxylase—FAH1 Sphingolipid alpha-hydroxylase; functions in the alpha-hydroxylation of sphingolipid-associated very long chain fatty acids, has both cytochrome b5-like and hydroxylase/desaturase domains, not essential for growth
SAC1	116	2	YKL212W	phosphatidylinositol-3-phosphatase SAC1—RSD1 Phosphatidylinositol phosphate (PtdInsP) phosphatase; involved in hydrolysis of PtdIns[4]P in the early and medial Golgi; regulated by interaction with Vps74p; ER localized transmembrane protein which cycles through the Golgi; involved in protein trafficking and processing, secretion, and cell wall maintenance; regulates sphingolipid biosynthesis through the modulation of PtdIns(4)P metabolism
YVC1	117	7	YOR087W	TRPY1—YOR088W Vacuolar cation channel; mediates release of Ca(2+) from the vacuole in response to hyperosmotic shock
TPO1	119	12	YLL028W	YLL028W Polyamine transporter of the major facilitator superfamily; member of the 12-spanner drug:H(+) antiporter DHA1 family; recognizes spermine, putrescine, and spermidine; catalyzes uptake of polyamines at alkaline pH and excretion at acidic pH; during oxidative stress exports spermine, spermidine from the cell, which controls timing of expression of stress-responsive genes; phosphorylation enhances activity and sorting to the plasma membrane
RSN1	120	11	YMR266W	YMR266W Membrane protein of unknown function; overexpression suppresses NaCl sensitivity of sro7 mutant cells by restoring sodium pump (Ena1p) localization to the plasma membrane
UBC6	120	1	YER100W	E2 ubiquitin-conjugating protein UBC6—DOA2 Ubiquitin-conjugating enzyme involved in ERAD; located at the cytosolic side of the ER membrane; tail region contains a transmembrane segment at the C-terminus; substrate of the ubiquitin-proteasome pathway; ER-associated protein degradation is also known as ERAD
GAB1	121	8	YLR459W	CDC91 GPI transamidase subunit; involved in attachment of glycosylphosphatidylinositol (GPI) anchors to proteins; may have a role in recognition of the attachment signal or of the lipid portion of GPI
MEP2	122	11	YNL142W	ammonium permease MEP2 Ammonium permease involved in regulation of pseudohyphal growth; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation
ALG2	124	4	YGL065C	GDP-Man:Man(1)GlcNAc(2)-PP-dolichol alpha-1,3-mannosyltransferase Mannosyltransferase in the N-linked glycosylation pathway; catalyzes two consecutive steps in the N-linked glycosylation pathway; alg2 mutants exhibit temperature-sensitive growth and abnormal accumulation of the lipid-linked oligosaccharide Man2GlcNAc2-PP-Dol
YGL114W	125	12	YGL114W	OPT3 Putative protein of unknown function; predicted member of the oligopeptide transporter (OPT) family of membrane transporters
HFD1	126	1	YMR110C	YMR110C Hexadecenal dehydrogenase; involved in the conversion of sphingosine 1-phosphate breakdown product hexadecenal to hexadecenoic acid; located in the mitochondrial outer membrane and also in lipid particles; similar to ALDH3A2, a human fatty aldehyde dehydrogenase (FALDH) mutated in Sjogren-Larsson syndrome, a neurocutaneous disorder
GAP1	127	12	YKR039W	amino acid permease GAP1 General amino acid permease; Gap1p senses the presence of amino acid substrates to regulate localization to the plasma membrane when needed; essential for invasive growth
ORM1	127	3	YGR038W	YGR038W Protein that mediates sphingolipid homeostasis; evolutionarily conserved, required for resistance to agents that induce unfolded protein response; Orm1p and Orm2p together control membrane biogenesis by coordinating lipid homeostasis with protein quality control; ORM1 has a paralog, ORM2, that arose from the whole genome duplication

Ranked list, Top50 (TM-containing, ER-localized only), BLAST threshold: 1e-10

Name	Rank	#TMs	SGD name	Description
ALG6	1	10	YOR002W	dolichyl-P-Glc:Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase Alpha 1,3 glucosyltransferase; involved in transfer of oligosaccharides from dolichyl pyrophosphate to asparagine residues of proteins during N-linked protein glycosylation; mutations in human ortholog are associated with disease
ALG8	2	12	YOR067C	dolichyl-P-Glc:Glc1Man(9)GlcNAc(2)-PP-dolichol alpha-1,3-glucosyltransferase—YOR29-18 Glucosyl transferase; involved in N-linked glycosylation; adds glucose to the dolichol-linked oligosaccharide precursor prior to transfer to protein during lipid-linked oligosaccharide biosynthesis; similar to Alg6p
MNS1	4	1	YJR131W	mannosyl-oligosaccharide 1,2-alpha-mannosidase Alpha-1,2-mannosidase; involved in ER-associated protein degradation (ERAD); catalyzes the removal of one mannose residue from a glycosylated protein, converting the modification from Man9GlcNAc to Man8GlcNAc; catalyzes the last step in glycoprotein maturation in the ER and is critical for ER protein degradation
ALE1	12	7	YOR175C	lysophospholipid acyltransferase—LCA1—LPT1—SLC4 Broad-specificity lysophospholipid acyltransferase; part of MBOAT family of membrane-bound O-acyltransferases; key component of Lands cycle; may have role in fatty acid exchange at sn-2 position of mature glycerophospholipids
ERG24	17	8	YNL280C	delta(14)-sterol reductase C-14 sterol reductase; acts in ergosterol biosynthesis; mutants accumulate the abnormal sterol ignosterol (ergosta-8,14 dienol), and are viable under anaerobic growth conditions but inviable on rich medium under aerobic conditions
AVT3	21	10	YKL146W	YKL146W Vacuolar transporter; exports large neutral amino acids from the vacuole; member of a family of seven <i>S. cerevisiae</i> genes (AVT1-7) related to vesicular GABA-glycine transporters
NHX1	24	8	YDR456W	bifunctional K ⁺ /Na ⁺ :H antiporter NHX1—VPL27—NHA2—VPS44 Na ⁺ /H ⁺ and K ⁺ /H ⁺ exchanger; required for intracellular sequestration of Na ⁺ and K ⁺ ; located in the vacuole and late endosome compartments; required for osmotolerance to acute hypertonic shock and for vacuolar fusion; ortholog of human NHE9, which is linked to autism
YPR003C	27	10	YPR003C	YPR003C Putative sulfate permease; physically interacts with Hsp82p; green fluorescent protein (GFP)-fusion protein localizes to the ER; YPR003C is not an essential gene
CSC1	35	11	YLR241W	YLR241W Calcium permeable gated cation channel; may be involved in detoxification; similar to Arabidopsis CSC1
BOR1	42	10	YNL275W	YNL275W Boron efflux transporter of the plasma membrane; binds HCO ₃ ⁻ , I ⁻ , Br ⁻ , NO ₃ ⁻ and Cl ⁻ ; has similarity to the characterized boron efflux transporter <i>A. thaliana</i> BOR1
ALG2	43	4	YGL065C	GDP-Man:Man(1)GlcNAc(2)-PP-dolichol alpha-1,3-mannosyltransferase Mannosyltransferase in the N-linked glycosylation pathway; catalyzes two consecutive steps in the N-linked glycosylation pathway; alg2 mutants exhibit temperature-sensitive growth and abnormal accumulation of the lipid-linked oligosaccharide Man2GlcNAc2-PP-Dol
ERG3	44	3	YLR056W	C-5 sterol desaturase—PSO6—SYR1 C-5 sterol desaturase; glycoprotein that catalyzes the introduction of a C-5(6) double bond into episterol, a precursor in ergosterol biosynthesis; transcriptionally down-regulated when ergosterol is in excess; mutants are viable, but cannot grow on non-fermentable carbon sources; substrate of HRD ubiquitin ligase
STE24	48	4	YJR117W	zinc metalloprotease—PIO2—AFC1 Highly conserved zinc metalloprotease; functions in two steps of a-factor maturation, C-terminal CAAX proteolysis and the first step of N-terminal proteolytic processing; contains multiple transmembrane spans; human homolog ZMPSTE24 implicated in mandibuloacral dysplasia (MAD), and can complement yeast null mutant
SCS7	56	3	YMR272C	fatty acid alpha-hydroxylase—FAH1 Sphingolipid alpha-hydroxylase; functions in the alpha-hydroxylation of sphingolipid-associated very long chain fatty acids, has both cytochrome b5-like and hydroxylase/desaturase domains, not essential for growth
ERV46	58	1	YAL042W	FUN9 Protein localized to COPII-coated vesicles; forms a complex with Erv41p; involved in the membrane fusion stage of transport
ERG11	63	2	YHR007C	sterol 14-demethylase—CYP51 Lanosterol 14-alpha-demethylase; catalyzes C-14 demethylation of lanosterol to form 4,4'-dimethyl cholesta-8,14,24-triene-3-beta-ol in ergosterol biosynthesis pathway; transcriptionally down-regulated when ergosterol is in excess; member of cytochrome P450 family; associated and coordinately regulated with the P450 reductase Ncp1p
ALG5	64	1	YPL227C	dolichyl-phosphate beta-glucosyltransferase

Name	Rank	#TMs	SGD name	Description
				UDP-glucose:dolichyl-phosphate glucosyltransferase; involved in asparagine-linked glycosylation in the endoplasmic reticulum
ALG1	66	1	YBR110W	chitobiosyldiphosphodolichol beta-1,4 mannosyltransferase
				Mannosyltransferase; involved in asparagine-linked glycosylation in the endoplasmic reticulum (ER); essential for viability; human homolog ALG1 complements yeast null mutant
GWT1	70	12	YJL091C	YJL091C
				Protein involved in the inositol acylation of GlcN-PI; the inositol acylation of glucosaminyl phosphatidylinositol (GlcN-PI) forms glucosaminyl(acyl)phosphatidylinositol (GlcN(acyl)PI), an intermediate in the biosynthesis of glycosylphosphatidylinositol (GPI) anchors
LAC1	72	7	YKL008C	sphingosine N-acyltransferase LAC1—DGT1
				Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lag1p; LAC1 has a paralog, LAG1, that arose from the whole genome duplication
ZRT2	73	7	YLR130C	low-affinity Zn(2+) transporter ZRT2
				Low-affinity zinc transporter of the plasma membrane; transcription is induced under low-zinc conditions by the Zap1p transcription factor
PMT5	77	11	YDL093W	putative dolichyl-phosphate-mannose-protein mannosyltransferase PMT5
				Protein O-mannosyltransferase; transfers mannose residues from dolichyl phosphate-D-mannose to protein serine/threonine residues; acts in a complex with Pmt3p, can instead interact with Pmt2p in some conditions; target for new antifungals
VCX1	79	11	YDL128W	MNR1—HUM1
				Vacuolar membrane antiporter with Ca ²⁺ /H ⁺ and K ⁺ /H ⁺ exchange activity; involved in control of cytosolic Ca ²⁺ and K ⁺ concentrations; has similarity to sodium/calcium exchangers, including the bovine Na ⁺ /Ca ²⁺ ,K ⁺ antiporter
AUR1	82	6	YKL004W	inositol phosphorylceramide synthase
				Phosphatidylinositol:ceramide phosphoinositol transferase; required for sphingolipid synthesis; can mutate to confer aureobasidin A resistance; also known as IPC synthase
CBR1	82	1	YIL043C	CBR5
				Cytochrome b reductase; not essential for viability; also detected in mitochondria; mutation in conserved NADH binding domain of the human ortholog results in type I methemoglobinemia
SAC1	83	2	YKL212W	phosphatidylinositol-3-phosphatase SAC1—RSD1
				Phosphatidylinositol phosphate (PtdInsP) phosphatase; involved in hydrolysis of PtdIns[4]P in the early and medial Golgi; regulated by interaction with Vps74p; ER localized transmembrane protein which cycles through the Golgi; involved in protein trafficking and processing, secretion, and cell wall maintenance; regulates sphingolipid biosynthesis through the modulation of PtdIns(4)P metabolism
ZRT1	83	8	YGL255W	high-affinity Zn(2+) transporter ZRT1
				High-affinity zinc transporter of the plasma membrane; responsible for the majority of zinc uptake; transcription is induced under low-zinc conditions by the Zap1p transcription factor
ATG15	85	1	YCR068W	triglyceride lipase ATG15—CVT17—AUT5
				Putative lipase required for lysis of autophagic and Cvt bodies; targeted to intravacuolar vesicles during autophagy via the multivesicular body (MVB) pathway; required for the maintenance of lipid droplet quantity after the diauxic shift; regulates lipolysis
ERG4	92	7	YGL012W	delta(24(24(1)))-sterol reductase
				C-24(28) sterol reductase; catalyzes the final step in ergosterol biosynthesis; mutants are viable, but lack ergosterol
NHA1	92	9	YLR138W	YLR138W
				Na ⁺ /H ⁺ antiporter; involved in sodium and potassium efflux through the plasma membrane; required for alkali cation tolerance at acidic pH
LAG1	93	7	YHL003C	sphingosine N-acyltransferase LAG1
				Ceramide synthase component; involved in synthesis of ceramide from C26(acyl)-coenzyme A and dihydrosphingosine or phytosphingosine, functionally equivalent to Lac1p; forms ER foci upon DNA replication stress; homolog of human CERS2, a tumor metastasis suppressor gene whose silencing enhances invasion/metastasis of prostate cancer cells; LAG1 has a paralog, LAC1, that arose from the whole genome duplication
MEP3	94	11	YPR138C	ammonium permease MEP3
				Ammonium permease of high capacity and low affinity; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation ammonia permease; MEP3 has a paralog, MEP1, that arose from the whole genome duplication
LRO1	95	1	YNR008W	phospholipid:diacylglycerol acyltransferase
				Acyltransferase that catalyzes diacylglycerol esterification; one of several acyltransferases that contribute to triglyceride synthesis; Lro1p and Dga1p can O-acylate ceramides; putative homolog of human lecithin cholesterol acyltransferase

Name	Rank	#TMs	SGD name	Description
ALG3	96	9	YBL082C	dolichyl-P-Man:Man(5)GlcNAc(2)-PP-dolichol alpha-1,3-mannosyltransferase—RHK1 Dolichol-P-Man dependent alpha(1-3) mannosyltransferase; involved in the synthesis of dolichol-linked oligosaccharide donor for N-linked glycosylation of proteins
ERG9	96	1	YHR190W	bifunctional farnesyl-diphosphate farnesyltransferase/squalene synthase Farnesyl-diphosphate farnesyl transferase (squalene synthase); joins two farnesyl pyrophosphate moieties to form squalene in the sterol biosynthesis pathway
MEP1	102	11	YGR121C	ammonium permease MEP1—AMT1 Ammonium permease; belongs to a ubiquitous family of cytoplasmic membrane proteins that transport only ammonium (NH ₄ ⁺); expression is under the nitrogen catabolite repression regulation; human homolog RHCG complements yeast null mutant; mutations in human homolog RHCG implicated in metabolic acidosis; MEP1 has a paralog, MEP3, that arose from the whole genome duplication
YKR051W	102	7	YKR051W	YKR051W Putative protein of unknown function
DGA1	106	1	YOR245C	diacylglycerol O-acyltransferase Diacylglycerol acyltransferase; catalyzes the terminal step of triacylglycerol (TAG) formation, acylates diacylglycerol using acyl-CoA as an acyl donor; Lro1p and Dga1p can O-acylate ceramides; localized to lipid particles
PGA3	107	2	YML125C	NQR1 Putative cytochrome b5 reductase, localized to the plasma membrane; may be involved in regulation of lifespan; required for maturation of Gas1p and Pho8p, proposed to be involved in protein trafficking; PGA3 has a paralog, AIM33, that arose from the whole genome duplication
YGR149W	112	7	YGR149W	YGR149W Putative protein of unknown function; predicted to be an integral membrane protein
CHS7	113	7	YHR142W	YHR142W Protein of unknown function; may be involved in chitin biosynthesis by regulation of Chs3p export from the ER; relocates from bud neck to ER upon DNA replication stress
ERG1	114	2	YGR175C	squalene monooxygenase Squalene epoxidase; catalyzes the epoxidation of squalene to 2,3-oxidosqualene; plays an essential role in the ergosterol-biosynthesis pathway and is the specific target of the antifungal drug terbinafine
RSN1	115	11	YMR266W	YMR266W Membrane protein of unknown function; overexpression suppresses NaCl sensitivity of sro7 mutant cells by restoring sodium pump (Ena1p) localization to the plasma membrane
SLC1	115	1	YDL052C	1-acylglycerol-3-phosphate O-acyltransferase SLC1 1-acyl-sn-glycerol-3-phosphate acyltransferase; catalyzes the acylation of lysophosphatidic acid to form phosphatidic acid, a key intermediate in lipid metabolism; enzymatic activity detected in lipid particles and microsomes
ERV29	117	7	YGR284C	YGR284C Protein localized to COPII-coated vesicles; involved in vesicle formation and incorporation of specific secretory cargo; protein abundance increases in response to DNA replication stress
OLE1	118	4	YGL055W	stearoyl-CoA 9-desaturase—MDM2 Delta(9) fatty acid desaturase; required for monounsaturated fatty acid synthesis and for normal distribution of mitochondria
TPO1	119	12	YLL028W	YLL028W Polyamine transporter of the major facilitator superfamily; member of the 12-spanner drug:H(+) antiporter DHA1 family; recognizes spermine, putrescine, and spermidine; catalyzes uptake of polyamines at alkaline pH and excretion at acidic pH; during oxidative stress exports spermine, spermidine from the cell, which controls timing of expression of stress-responsive genes; phosphorylation enhances activity and sorting to the plasma membrane
GAB1	122	8	YLR459W	CDC91 GPI transamidase subunit; involved in attachment of glycosylphosphatidylinositol (GPI) anchors to proteins; may have a role in recognition of the attachment signal or of the lipid portion of GPI
YNR048W	122	2	YNR048W	putative aminophospholipid translocase regulatory protein—CRF1 Potential noncatalytic subunit for phospholipid translocase Dnf3p; YNR048W has a paralog, CDC50, that arose from the whole genome duplication
PHO84	124	9	YML123C	phosphate transporter PHO84—phoT High-affinity inorganic phosphate (Pi) transporter; also low-affinity manganese transporter; regulated by Pho4p and Spt7p; mutation confers resistance to arsenate; exit from the ER during maturation requires Pho86p; cells overexpressing Pho84p accumulate heavy metals but do not develop symptoms of metal toxicity