

1       **Testing Darwin’s hypothesis about the most wonderful plant in the**  
2       **world: The Venus flytrap’s marginal spikes are a ‘horrid prison’ for**  
3                               **moderate-sized insect prey**

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26 **Abstract**

27 Botanical carnivory is a novel feeding strategy associated with numerous physiological and  
28 morphological adaptations. However, the benefits of these novel carnivorous traits are rarely  
29 tested. Here, we used field observations and lab experiments to test the prey capture function of  
30 the marginal spikes on snap traps of the Venus flytrap (*Dionaea muscipula*). Our field and  
31 laboratory results suggested surprisingly inefficient capture success: fewer than 1 in 4 prey  
32 encounters led to prey capture. Removing the marginal spikes decreased the rate of prey capture  
33 success for moderate-sized cricket prey by 90%, but this effect disappeared for larger prey. The  
34 nonlinear benefit of spikes suggests that they provide a better cage for capturing more abundant  
35 insects of moderate and small sizes, but may also provide a foothold for rare large prey to  
36 escape. Our observations support Darwin's hypothesis that the marginal spikes form a 'horrid  
37 prison' that increases prey capture success for moderate-sized prey, but the decreasing benefit for  
38 larger prey is unexpected and previously undocumented. Thus, we find surprising complexity in  
39 the adaptive landscape for one of the most wonderful evolutionary innovations among all plants.  
40 These findings further enrich our understanding of the evolution and diversification of novel trap  
41 morphology in carnivorous plants.

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## 49 **Introduction**

50 The origins of novel structures remain an important and poorly understood problem in  
51 evolutionary biology (Mayr 1960, Mozcek 2008). Novel traits are often key innovations  
52 providing new ecological opportunities (Maia et al. 2013; Stroud and Losos 2016; Wainwright et  
53 al. 2012). Despite the importance of these traits, our understanding of the adaptive value of novel  
54 structures is often assumed, and rarely tested directly. Frequently, this is because it is difficult or  
55 impossible to manipulate the trait without impairing organismal function in an unintended way;  
56 however, many carnivorous plant traits do not present this obstacle.

57 Botanical carnivory is a novel feeding strategy that has evolved at least nine separate  
58 times in over 700 species of angiosperms, typically in areas with severely limited nitrogen and  
59 phosphorus (Ellison 2006; Givnish 2015; Givnish et al. 1984; Król et al. 2012, Roberts and  
60 Oosting 1958). Pitfall traps evolved independently at least 6 times and sticky traps 5 times.  
61 However, snap traps have most likely evolved only once in the ancestral lineage leading to the  
62 aquatic waterwheel (*Aldrovandra vesiculosa*) and Venus flytrap (*Dionaea muscipula*), which is  
63 sister to the sundews (*Drosera* spp.) and within the Caryophalles (Cameron 2002, Givnish 2015,  
64 Walker et al. 2017). Multiple hypotheses have been proposed for why snap traps evolved  
65 including the ability to capture larger prey, capture prey more quickly, or more completely digest  
66 prey (Darwin 1875; Gibson and Waller 2009). However, these hypotheses have never been  
67 tested except for a few field studies documenting the size and diversity of arthropod prey  
68 (Gibson 1991; Hutchens and Luken 2015; Youngsteadt et al. 2018).

69 The marginal spikes found in *Dionaea* are modified trichomes that extend from the margin of  
70 the trap lobes. These spikes are homologous to the trichomes of sundews, but do not exude any  
71 sticky resin and have lost the mucus glands (Gibson and Waller 2009). Darwin was the first to

72 document evidence for carnivory in flytraps and sundews in a series of careful experiments and  
73 proposed that the marginal spikes of flytraps enhance prey capture success by providing a cage-  
74 like structure around the top of the trap that contains the prey (Darwin 1875; Gibson and Waller  
75 2009). Darwin (1875) also hypothesized that while small insects will be able to escape between  
76 the spikes, a moderately sized insect will be “pushed back again into its horrid prison with  
77 closing walls” (page 312), and large, strong insects will be able to free themselves. Determining  
78 the function of the marginal spikes is important for understanding the rarity of mechanical snap  
79 traps whereas sticky and pitfall traps are ubiquitous across carnivorous plants.

80 Traits that enhance prey capture ability are expected to be strongly selected for given the  
81 benefits of additional nutrients and the energetic and opportunity costs associated with a  
82 triggered trap missing its intended prey. Nutrients from insect prey increase the growth rate of  
83 Venus flytraps (Darwin 1878; Roberts and Oosting 1958) at a cost of lower photosynthetic  
84 efficiency of carnivorous plants compared to other plants (Ellison and Gotelli 2009; Pavlovic et  
85 al. 2009). The traps are triggered by an action potential when specialized trigger hairs are  
86 stimulated (Volkov et al. 2008, 2009) and close as quickly as 100 milliseconds forming a cage  
87 around the prey item (Poppinga et al. 2013). If the trap fails to capture an insect, it takes between  
88 two and three days for the trap to re-open, during which time it is unable to be used for prey  
89 capture. Beyond the energy expended to close a trap and the opportunity cost of a miss, there is a  
90 cost associated with declining trap performance and trap death. Traps that have closed and re-  
91 opened have lower subsequent trap closure speeds and trap gape angle (Stuhlman 1948).  
92 Additionally, after a few closings, traps rapidly die. The marginal spikes provide a novel and  
93 unique function that potentially increases prey capture rate and minimizes the costs associated  
94 with a failed trap closing event.

95           We measured prey capture efficiency and the effect of marginal spikes using field  
96 observations of wild Venus flytraps and laboratory experiments. By testing the prey capture  
97 ability of plants with intact spikes and ones with the spikes clipped off, we assessed the novel  
98 function of the marginal spike cage for prey capture.

99

## 100 **Methods:**

### 101 *Field Data Collection*

102 The Green Swamp Preserve, NC, USA is one of the last remaining eastern pine savanna habitats  
103 containing endemic flytraps. To estimate prey capture rates, we identified individual plants ( $n =$   
104 14) and recorded the number of traps that fell into four categories: alive and closed, dead and  
105 closed, alive and open, and dead and open. All closed traps ( $n = 100$ ) had their length, defined  
106 here as the widest point of the lobes on the long axis, recorded with digital calipers. We used a  
107 flashlight to illuminate the trap from behind making anything inside the trap visible as a  
108 silhouette. If the trap contained something it was assigned a value of 1 for “catch” and if it  
109 contained nothing it was assigned a 0 for “miss”. We also noted when a trap was closed on  
110 another trap or contained debris inside such as sticks or grass (these were considered a miss;  $n =$   
111 7). Logistic regression in R Studio (R Statistical Programming Group 2018; RStudio Team 2015)  
112 was used to determine if trap length had a significant effect on prey capture rate in the field.

113

### 114 *Laboratory prey capture experiments*

115 Plants used in lab experiments were tissue-cultured and purchased from commercial suppliers  
116 (bugbitingplants.com; stores.ebay.com/joelcarnivorousplants/). The plants were maintained in  
117 40 liter terraria under high-output fluorescent lighting (14-hour daylight cycle) with 8 cm pots

118 submerged in 1-4 cm of reverse osmosis water at all times. Throughout the duration of the  
119 experiments, the plants were kept at ambient temperatures under the lights, ranging from 35° C  
120 during the day to 22 C at night), and 50 – 90% humidity. Crickets were purchased from Petsmart  
121 and kept in 4-liter plastic containers with shelter, water, and a complete diet (Fluker’s cricket  
122 food).

123 To assess the adaptive role of marginal spikes, we set up prey capture arenas (Fig 1C).  
124 Each arena consisted of one plant in a petri dish of distilled water, one cricket of known length  
125 (range: 0.7 cm – 2.3 cm) and mass (range: 0.026 g – 0.420 g), cricket food, and a ramp from the  
126 dry bottom of the arena to the plant. Only healthy crickets with all six legs were used for prey  
127 capture trials. Crickets were chosen as the prey item because they represent one extreme of prey  
128 difficulty (large and able to jump) while still making up approximately 10% of the flytrap’s diet  
129 in the wild (Ellison and Gotelli 2009). All closed traps were initially marked. We checked the  
130 plants for closed traps after three days and after one week. Every closed, empty trap was  
131 recorded as a 0 for “miss” and every closed trap that contained prey was recorded as a 1 for  
132 “catch”. Following one unmanipulated trial with the spikes intact, we used scissors to clip the  
133 spikes from every trap on the plant (Fig 1). The plants were then allowed to recover for a week  
134 until the traps re-opened. After the traps re-opened, we placed each plant through a second trial  
135 with a new cricket. We performed 51 prey capture trials (34 plants total, 17 used only for  
136 unmanipulated trials, and 17 used once before and after spike removal). Only 1 trial resulted in  
137 no traps triggered over the full week. We also set up control trials (n = 5) with a newly dead  
138 cricket placed on the bottom of the tank and negative controls with no cricket at all (n = 2) to  
139 ensure that any experimental trap closures were triggered by the cricket and not spontaneous.

140 To analyze the relationship between prey mass, treatment, trap length, and prey capture  
141 success we used multiple logistic regression models in R and generalized linear mixed-effect  
142 models using the lme4 package (Bates et al. 2015). For the linear mixed effect models, we used  
143 Akaike information criteria with correction for small sample size (AICc) to compare models. We  
144 chose prey capture success as our proxy for performance and fitness due to the evidence that the  
145 growth rate of flytraps is greatly enhanced by ingesting insect prey (Schulze et al. 2001). We  
146 visualized changes in the performance landscape due to removing marginal spikes by estimating  
147 thin-plate splines for trials with and without spikes. We fit splines by generalized cross-  
148 validation using the Tps function in the Fields package (Nychka et al. 2015) in R (R Core Team  
149 (2017).

150

## 151 **Results:**

### 152 *Field Prey Capture Rates*

153 Only 24% of closed wild flytraps contained prey. This number represents a high-end estimate  
154 because anything inside the plants was counted as a catch, despite the possibility that the object  
155 was a piece of debris instead of an insect or spider. Of the 98 closed traps recorded, 8 were  
156 closed around obvious plant debris, and 2 contained identifiable prey (1 ant and 1 spider).  $55\% \pm$   
157  $5\%$  (mean  $\pm$  SE) of wild flytraps were open and alive, therefore able to capture prey.

158

### 159 *Laboratory Prey Capture Rates*

160 Similarly in the lab, only 16.5% of flytraps successfully captured prey out of all closed traps  
161 among unmanipulated plants. Only 5.8% of flytraps on these same plants with marginal spikes  
162 removed successfully captured prey. Tissue damage due to clipping marginal spikes quickly  
163 healed and clipped traps reopened within 4 days; thus, this disparity does not appear to be due to

164 any deleterious effect of tissue damage. Furthermore, no differences in trap closing speeds,  
165 health, or growth rates of manipulated traps were apparent. Indeed, marginal teeth began to  
166 regrow within approximately one week after removal, suggesting that we underestimated the  
167 effect of spike removal on prey capture since spikes were partially regrown by the end of each  
168 trial.

169  
170 Removing marginal spikes reduced the odds of prey capture by 90% relative to unmanipulated  
171 traps from the same plant while controlling for prey mass and trap length (effect of manipulation:  
172  $P = 0.002106$ ; linear mixed-effect model relative to model without treatment variable:  $\Delta AIC_c =$   
173 11). At large prey sizes and large trap lengths this effect disappears (note that spline SE crosses  
174 at large prey and trap sizes; Figs. 3b,c).

175  
176 *Effect of Prey Mass and Trap Length*

177 A linear mixed effect model with prey mass included provided a far better fit to the data than one  
178 without ( $\Delta AIC_c = 15$ ). In the full model, prey mass was a significant predictor of prey capture  
179 success ( $P = 0.000441$ ), with every 0.1 g increase in prey mass corresponding to a 73% decrease  
180 in prey capture performance (Fig 3).

181  
182 Larger trap size also increases the probability of successful prey capture after controlling for prey  
183 size, with every 1 cm increase in trap length increasing the odds of prey capture by 2.9-fold  
184 (Table 1). Larger trap size increased prey capture success for both manipulated and non-  
185 manipulated plants (Fig 3; logistic regression; manipulated:  $P = 0.02008$ ; non-manipulated:  $P =$



186 0.003007). A linear mixed effect model including trap length provided a much better fit for the  
187 data than one without ( $\Delta AIC_c = 31$ )

188

## 189 **Discussion:**

190 We provide the first direct test of how prey capture performance is affected by the presence of  
191 marginal spikes, trichomes which provide a novel function in Venus flytraps by forming what  
192 Darwin described as a “horrid prison”. We found that the marginal spikes are adaptive for prey  
193 capture of small and medium sized insects, but not larger insects. In controlled laboratory prey  
194 capture trials, 16.5% of trap closures resulted in successful prey capture whereas only 5.8% of  
195 trap closures successfully captured prey when marginal spikes were removed (Fig. 2b-c). We  
196 found similarly low prey capture rates in the Green Swamp Preserve, one of the natural habitats  
197 of the Venus flytrap: fewer than 25% of trap closures resulted in prey capture (Fig. 2a).  
198 Furthermore, only about half of the wild traps were open, alive, and available to catch prey.  
199 Given the documented tradeoff between photosynthetic efficiency and carnivory and costs  
200 associated with maintaining traps (Ellison and Gotelli 2009; Pavlovic et al. 2009), it is possible  
201 that the nutrients acquired from a relatively small number of traps are sufficient to maintain the  
202 plant. In support of this hypothesis, other carnivorous plants (*Sarracenia purpurea* and  
203 *Darlingtonia californica*) sustain themselves with prey capture rates as low as 2% for ants and  
204 wasps, respectively (Newell and Nastase 1998; Dixon et al., 2005). Alternatively, prey capture  
205 rates for tropical pitcher plants (*Nepenthes rafflesian*) may reach 100% for ants (Bauer et al.  
206 2008). Given that Venus flytraps fall in the middle of this range for pitfall traps, additional  
207 factors beyond prey capture rate may underlie the origins of mechanical snap traps.

208           A second hypothesis for the evolution of mechanical snap traps is selection for capturing  
209 larger prey. In habitats where multiple carnivorous plant species coexist we would expect  
210 specialization and ecological partitioning (Schoener 1974). Sundews, which grow in sympatry  
211 with Venus flytraps in the Green Swamp Preserve, frequently allow prey items larger than 5mm  
212 to escape (Gibson 1991) whereas flytraps have been known to capture prey as large as 30mm  
213 with an estimated average of 9.3mm (Jones 1923; Ellison and Gotelli 2009). In this study, we  
214 found estimated prey capture rates as high as 80% for the largest flytrap sizes despite the average  
215 prey size (15.2 mm) being larger than what was reported by Jones (1923). This suggests that  
216 mechanical traps are capable of capturing much larger prey than sticky traps. Although some  
217 studies found no support for resource partitioning among sympatric species assemblies of  
218 carnivorous plants (Ellison and Gotelli 2009; Verbeek and Boasson 1993), others demonstrated  
219 differential prey distributions at the individual plant level and among species (Karlsson et al.  
220 1987; Gibson and Waller 2009; Thum 1986). Given the extreme differences in mean and  
221 maximum prey sizes between sticky traps and snap traps, it is likely that resource partitioning at  
222 least plays a role in the continued coexistence of sundews with flytraps throughout their limited  
223 range.

224           Surprisingly, the effect of removing the marginal spikes for medium-sized traps on prey  
225 capture success nearly disappears for larger traps. We observed a possible mechanistic  
226 explanation for this counterintuitive result. Crickets are often climbing on the marginal spikes of  
227 large traps, and when they trigger them they are able to push against the marginal spikes to pry  
228 themselves free. In contrast, when a cricket triggers a large trap with no spikes, it has nothing to  
229 use to free itself. Marginal spikes appear to provide leverage for larger insect prey to escape.  
230 There is also a possible physical explanation for the diminishing benefit of the marginal spikes at

231 large trap sizes. Stuhlman (1948) speculated that friction between the marginal spikes may slow  
232 down trap closure. Because the contact area over which friction matters is proportional to the  
233 length squared, we would expect disproportionately large frictional forces as the length of  
234 marginal spikes increases on larger traps.

235 In his writings on insectivorous plants, Darwin (1875) hypothesized that the marginal  
236 spikes allowed flytraps to capture larger insects while letting tiny insects go free. Later work has  
237 been mixed on whether snap traps are size-selective (Hutchens and Luken 2009; Hatcher and  
238 Hart 2014 (ontogenetic changes)) and we did not find any evidence for size-selection here. For  
239 medium and small insects, the cage formed by marginal spikes provided a drastic increase in  
240 prey capture rates, a finding that is compatible with Darwin's original hypothesis. At large prey  
241 sizes, however, the symmetry between our findings and his hypothesis begin to break down. We  
242 found diminishing returns at larger prey sizes, and while Darwin predicted large insects would  
243 break free from traps, the mechanism he outlines is different than the one we observe. We did  
244 not find that fully trapped insects were breaking free, as he notes in his book. Instead, we found  
245 insects that were partially trapped or trapped perpendicular to the trap's long axis were the ones  
246 to break free, potentially with the aid of the marginal spikes.

247 We demonstrated that the novel marginal spikes, forming a 'horrid prison', are an  
248 adaptation for prey capture with nonlinear effects at larger prey/trap sizes. Given the diversity of  
249 carnivorous plant traps, from the sticky traps of sundews to the rapid suction traps of bladderwort  
250 (Brown et al. 2012), we contend that carnivorous plants offer a rich system for investigating the  
251 adaptive value of novel traits, particularly within the context of prey capture. Furthermore, this  
252 system lends itself to tractable experimental work carried out by undergraduate researchers. This  
253 project was carried out during a one-semester course-based undergraduate research experience

254 (CURE) course taught at UNC, entitled ‘The Evolution of Extraordinary Adaptations’.  
255 Characterizing the role of these unique features aids our understanding of potential axes of  
256 selection that drive the evolution of different trap types and the rarity of mechanical traps. In  
257 turn, this tractable laboratory and field systems offers insights into the origins of one of the most  
258 wonderful evolutionary innovations among all plants.

259

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266

## 267 **Data accessibility**

268 All data and R scripts used for this study will be deposited in the Dryad Digital Repository.

269

## 270 **References**

271

272 Bauer U, Bohn HF, Federle W. 2008. Harmless nectar source or deadly trap: *Nepenthes* pitchers are activated by  
273 rain, condensation and nectar. *Proceedings of the Royal Society B: Biological Sciences* 275:259–265.

274

275 Brown, M.D., Holzman, R., Berg, O. and Müller, U.K., 2012. Sub-millisecond flow fields induced by bladderwort,  
276 the fastest known suction feeder. *Society for Integrative and Comparative Biology*, January 3-7, 2012. Charleston,  
277 SC, USA.

278

279 Cameron, Kenneth M. Molecular Evidence for the Common Origin of Snap-Traps Among Carnivorous Plants.  
280 2002. *American Journal of Botany* 89:1503 – 1509.

281

282 Darwin, Charles. *Insectivorous Plants*. 1875. *The Complete Works of Charles Darwin Online*. Retrieved 2009-08-  
283 26. Chapter XIII: pages 286 – 320.

284

- 285 Darwin, F., 1878. Experiments on the Nutrition of *Drosera rotundifolia*. Botanical Journal of the Linnean  
286 Society, 98:17-31.  
287
- 288 Douglas Bates, Martin Maechler, Ben Bolker, Steve Walker (2015). Fitting Linear Mixed-Effects Models Using  
289 lme4. Journal of Statistical Software, 67(1), 1-48.<[doi:10.18637/jss.v067.i01](https://doi.org/10.18637/jss.v067.i01)>.  
290
- 291 Douglas Nychka, Reinhard Furrer, John Paige and Stephan Sain (2015). “fields: Tools for spatial data.”  
292 doi: [10.5065/D6W957CT](https://doi.org/10.5065/D6W957CT), R package version 9.0  
293
- 294 Dixon PM, Ellison AM, Gotelli NJ. 2005. Improving the precision of estimates of the frequency of rare events.  
295 Ecology 86: 1114–1123.  
296
- 297 Ellison, A. M. 2006. Nutrient Limitation and Stoichiometry of Carnivorous Plants. Plant Biology 8(6): 740-747  
298
- 299 Ellison, DM; Gotelli, NJ. 2009. “Energetics and the evolution of carnivorous plants—Darwin’s ‘Most Wonderful  
300 plants in the world’”. Experiment Botany. 60(1):19–42  
301
- 302 Gibson TC. 1991. Differential escape of insect from carnivorous plant traps. American Midland Naturalist 125:55–  
303 62  
304
- 305 Gibson, Thomas C. and Waller, D. M. 2009. Evolving Darwin’s ‘most wonderful’ plant: Ecological Steps to a Snap-  
306 trap. New Phytologist 183: 575–587.  
307
- 308 Givnish, T. J., Burkhardt, E. L., Happel, R. E., and Weintraub, J. D. 1984. Carnivory in the bromeliad *Brocchinia*  
309 *reducta*, with a cost/benefit model for the general restriction of carnivorous plants to sunny, moist, nutrient-poor  
310 habitats. American Naturalist 124:479 – 497.  
311
- 312 Givnish, T. J. 2015. “New evidence on the origin of carnivorous plants”. Proceedings of the National Academy of  
313 Sciences 112(1):10–11.  
314
- 315 Hutchens, John J. and Luken, James O. 2009. Prey Capture in the Venus Flytrap: Collection or Selection. Botany  
316 87:1007 – 1010.  
317
- 318 Jones FM. 1923. The most wonderful plant in the world. Natural History 23:589–596.  
319
- 320 Karlsson, P.S., Nordell, K.O., Eirefelt, S. and Svensson, A.N.D.A., 1987. Trapping efficiency of three carnivorous  
321 Pinguicula species. Oecologia, 73(4):518-521.  
322
- 323 Król, E., Płachno, B.J., Adamec, L., Stolarz, M., Dziubińska, H. and Trębacz, K., 2012. Quite a few reasons for  
324 calling carnivores ‘the most wonderful plants in the world’. Annals of Botany, 109(1):47-64.  
325
- 326 Luken, James O. 2005. *Dionaea muscipula* (Venus flytrap) establishment, release, and response of associated  
327 species in mowed patches on the rims of Carolina bays. Restoration Ecology 13(4):678-684.  
328
- 329 Maia, R., Rubenstein, D.R. and Shawkey, M.D., 2013. Key ornamental innovations facilitate diversification in an  
330 avian radiation. Proceedings of the National Academy of Sciences, 110(26):10687-10692.  
331
- 332 Mayr, E., 1960. The emergence of evolutionary novelties. Evolution after Darwin, 1:349-380.  
333
- 334 Müller, G.B. and Nitecki, M.H., 1990. Developmental mechanisms at the origin of morphological novelty: a side-  
335 effect hypothesis. Pages 99-130 in M. H. Nitecki, ed Evolutionary Innovations. University of Chicago Press.  
336
- 337 Newell SJ, Nastase AJ. 1998. Efficiency of insect capture by *Sarracenia purpurea* (Sarraceniaceae), the northern  
338 pitcher plant. American Journal of Botany 85:88–91  
339

- 340 Pavlovič, Andrej, and Michaela Saganová. 2015. A novel insight into the cost–benefit model for the evolution of  
341 botanical carnivory. *Annals of botany* 115(7):1075-1092.  
342
- 343 Poppinga, S., Masselter, T. and Speck, T. 2013. Faster than their prey: new insights into the rapid movements of  
344 active carnivorous plants traps. *BioEssays*, 35(7):649-657.  
345
- 346 R Core Team (2017). R: A language and environment for statistical computing. R Foundation for Statistical  
347 Computing, Vienna, Austria. URL <https://www.R-project.org/>.  
348
- 349 RStudio Team (2015). RStudio: Integrated Development for R. RStudio, Inc., Boston, MA  
350 URL <http://www.rstudio.com/>.  
351
- 352 Roberts, Patricia R., and Henry John Oosting. 1958. Responses of Venus flytrap (*Dionaea muscipula*) to factors  
353 involved in its endemism. *Ecological Monographs* 28(2):193-218.  
354
- 355 Schulze, W., Schulze, E.D., Schulze, I. and Oren, R., 2001. Quantification of insect nitrogen utilization by the venus  
356 flytrap *Dionaea muscipula* catching prey with highly variable isotope signatures. *Journal of Experimental*  
357 *Botany* 52(358):1041-1049.  
358
- 359 Schoener, T.W., 1974. Resource partitioning in ecological communities. *Science* 185(4145):27-39.  
360
- 361 Stroud, J.T. and Losos, J.B., 2016. Ecological opportunity and adaptive radiation. *Annual Review of Ecology,*  
362 *Evolution, and Systematics* 47.  
363
- 364 Stuhlman, O., 1948. A physical analysis of the opening and closing movements of the lobes of Venus' fly-  
365 trap. *Bulletin of the Torrey Botanical Club* 75:22-44.  
366
- 367 Thum, M., 1986. Segregation of habitat and prey in two sympatric carnivorous plant species, *Drosera rotundifolia*  
368 and *Drosera intermedia*. *Oecologia* 70(4):601-605.  
369
- 370 Volkov, A. G., Adesina, T., Markin, V.S., Jovanov, E. 2008. Kinetics and Mechanism of *Dionaea muscipula* Trap  
371 Closing. *American Society of Plant Biologists*. 146(2):694 – 702.  
372
- 373 Volkov, A. G. Carrell, H., Markin, V.S. 2009. Biologically Closed Electrical Circuits in Venus Flytrap. 2009. *Plant*  
374 *Physiol.* Vol. 149(4):1661 – 1667  
375
- 376 Verbeek NAM, Boasson R. 1993. Relationship between types of prey captured and growth form in *Drosera* in  
377 southwestern Australia. *Australian Journal of Ecology* 18:203–207.  
378
- 379 Wainwright, P.C., Smith, W.L., Price, S.A., Tang, K.L., Sparks, J.S., Ferry, L.A., Kuhn, K.L., Eytan, R.I. and Near,  
380 T.J., 2012. The evolution of pharyngognath: a phylogenetic and functional appraisal of the pharyngeal jaw key  
381 innovation in labroid fishes and beyond. *Systematic Biology*, 61(6):1001-1027.  
382
- 383 Walker, J.F., Yang, Y., Moore, M.J., Mikenas, J., Timoneda, A., Brockington, S.F. and Smith, S.A., 2017.  
384 Widespread paleopolyploidy, gene tree conflict, and recalcitrant relationships among the carnivorous  
385 Caryophyllales. *American journal of botany*, 104(6):858-867.  
386
- 387 H. Wickham. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2009.  
388
- 389 Youngsteadt, E., Irwin, R.E., Fowler, A., Bertone, M.A., Giacomini, S.J., Kunz, M., Suiter, D. and Sorenson, C.E.,  
390 2018. Venus Flytrap Rarely Traps Its Pollinators. *The American Naturalist* 191(4):539-546.  
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393 **Table 1.** Generalized linear mixed-effect model showing the effect of removing the marginal  
394 spikes (manipulation), trap length, and prey mass on prey capture performance (logistic  
395 regression). Significant *P*-values are bolded.

396

Model Term	Estimate $\pm$ SE	<i>P</i>
Manipulation	-2.32 $\pm$ 0.75	<b>0.002107</b>
Trap Length	4.74 $\pm$ 1.08	<b>0.000011</b>
Prey Mass	-13.36 $\pm$ 3.80	<b>0.000441</b>

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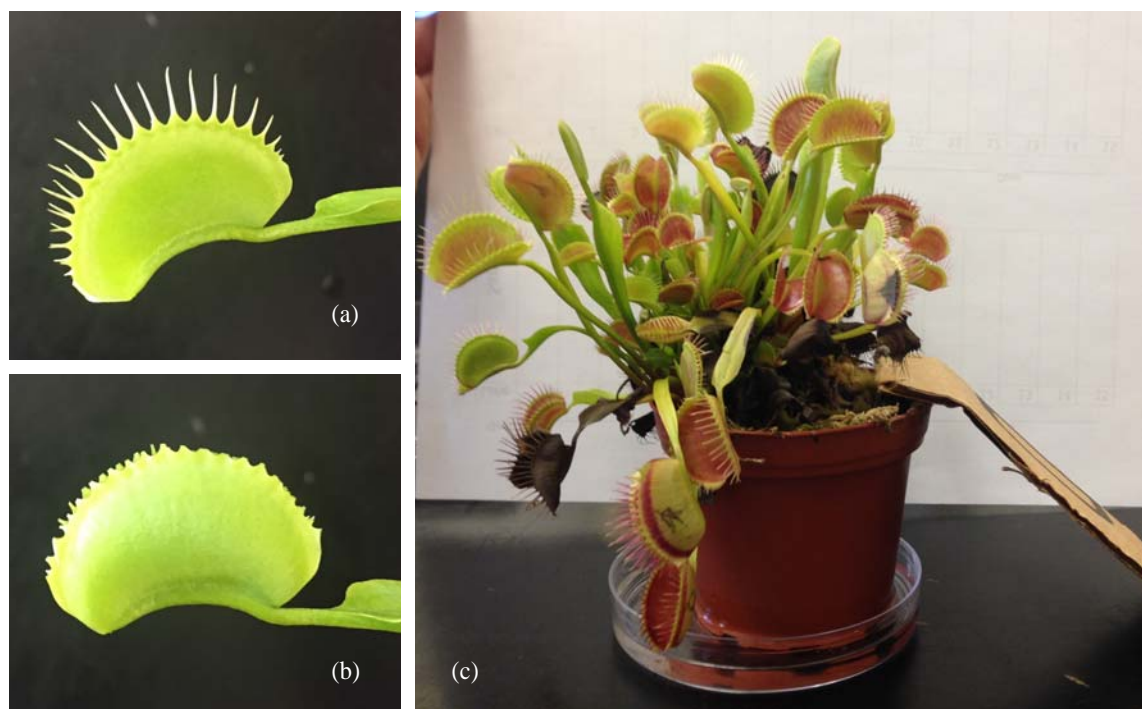
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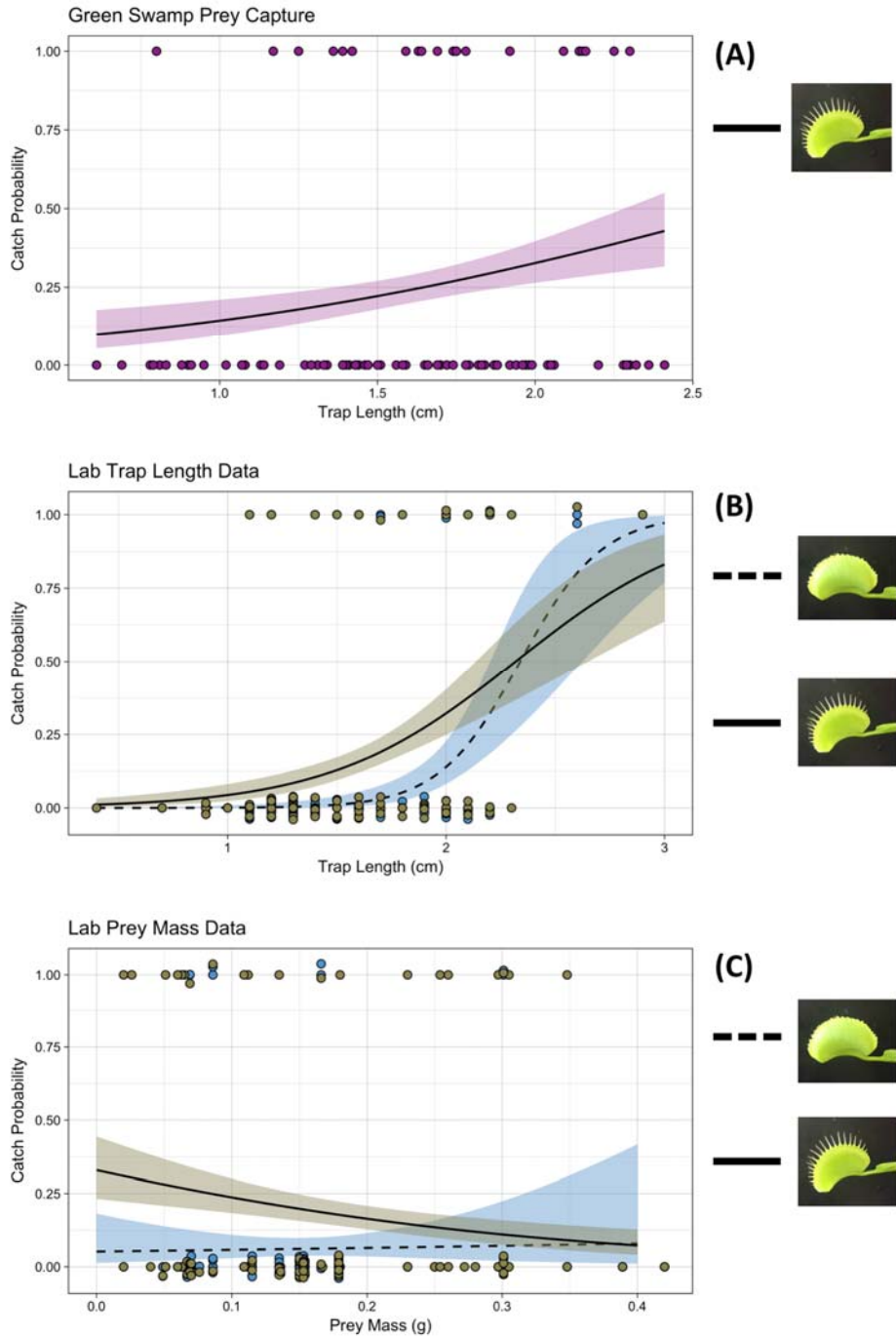


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421 **Figure 1:** (a) Intact trap; (b) trap with the marginal spines removed; (c) representative prey

422 capture arena containing one plant, one cricket, a ramp, and a petri dish of water.





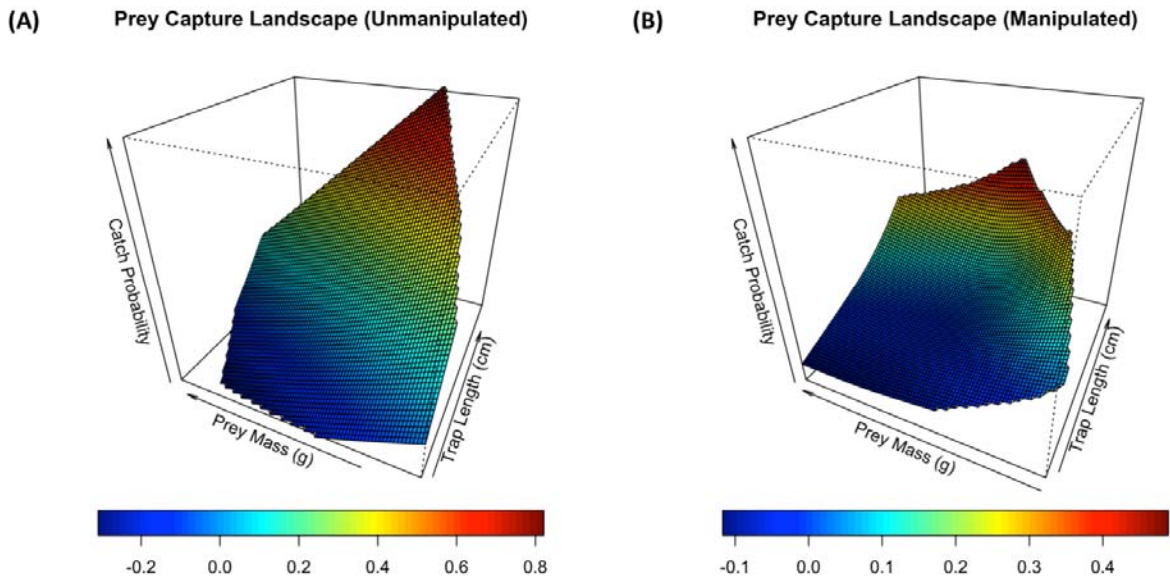
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425 **Fig. 2:** (A) Prey capture success of wild plants in the Green Swamp Preserve, NC as a function  
426 of trap length (measured to the nearest 0.01"). (B) Prey capture success of laboratory plants as a  
427 function of trap length (measured to the nearest 0.1") (C) Prey capture success of laboratory  
428 plants as a function of prey mass. Lines of best fit were estimated using logistic regression with

429 shaded areas corresponding to  $\pm 1$  SE. Each point represents one successful (1) or unsuccessful  
430 (0) capture by a flytrap, often resulting in multiple failed captures per cricket mass.

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436 **Fig. 3:** Prey capture landscapes for intact plants (left) and manipulated plants (right). Catch  
437 probability is on the z axis and represented by the heat colors relative to insect prey mass and  
438 trap length plotted in the x-y plane. The performance landscape for plants without marginal  
439 spikes is greatly depressed at small trap sizes, but is similar at large trap/prey sizes.

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