Genome sequence and description of *Blautia brookingsii* str SG772 nov., a new species of anaerobic bacterium isolated from healthy human gut.

Sudeep Ghimire^{1,2}, Roshan Kumar^{1,2}, Eric Nelson^{1,2}, Jane Christopher-Hennings^{1,2}, and Joy Scaria^{1,2*}

*Corresponding Author

Email: joy.scaria@sdstate.edu

¹Department of Veterinary and Biomedical Sciences, South Dakota State University, Brookings, SD, USA.

²South Dakota Center for Biologics Research and Commercialization, SD, USA.

Abstract

Strain SG-772 is a Gram positive, strictly anoxic bacterium isolated from the feces of a healthy human fecal donor. Based on 16S rRNA gene sequence, the strain showed maximum similarity (94.39%) with *Blautia stercoris* GAM6-1 in EZ-Taxon server and thus assigned the genus *Blautia*. The scanning electron micrograph of the bacterium revealed the characteristic coccobacillus shape as well as the complete absence of flagellum, suggesting its non-motile phenotype. This strain was found to utilize 27 substrates based on Biolog AN plates assay, with maximum preference for D-mannitol. Additionally, the strain was found to be resistant to tetracycline and streptomycin. Genome sequencing and analysis revealed an overall genome size of 3.49 Mbp and GC content of 43.97%. Based on RAST annotation server, the closest neighbor was *Blautia hansenii* DSM20583. Average Nucleotide Identity (ANI) of these strains were 81.69%, suggesting a high level of genomic variation. The comparative genome analysis of strain SG772 with *B. hansenii* DSM20583 revealed a total of 411 orthologous genes coding for basic metabolic functions. Furthermore, the genomes were functionally distinct based on COG categories. Thus, based on all these differences, we propose a novel species of genus *Blautia* named as *Blautia brookingsii* SG772.

Keywords: *Blautia*, *brookingsii* SG772, Human gut, Genome, Anaerobe

42

43 **Introduction**

44 The members of family *Lachnospiraceae* are commensal bacteria inhabiting the human
 45 gastrointestinal tract (1). These group of bacteria degrade complex polysaccharides to produce
 46 short chain fatty acids. Acetate, butyrate and propionate produced as the result of digestion of
 47 polysaccharides can be used as an energy source for the host (2). *Lachnospiraceae* in human gut
 48 microbiota plays important role in maintaining health and disease balance (3-7). Recently, a new
 49 genus *Blautia* has been formed under *Lachnospiraceae* family by reclassifying few members of
 50 genus *Clostridium* and *Ruminococcus* (8). *Blautia* has been reported to provide colonization
 51 resistance against *Enterococcus faecium* (9) and *Clostridium difficile* (5).

52 The members of genus *Blautia* are gram- positive, non-motile, coccoid shaped; obligate
 53 anaerobes which can produce a wide variety of compounds as the by-product of fermentation.
 54 Most of the members of this genus were either isolated from the human or mammalian fecal
 55 samples (8, 10). During the culturomics of human feces, a novel *Blautia*-like strain was isolated
 56 from Brain Heart Infusion (BHI) based medium and named as strain SG772. Based on 16S
 57 rRNA gene sequence phylogeny, it was taxonomically clustered with the genus *Blautia*.
 58 Additionally, using taxo-genomics approach, the bacterium SG772 showed distinct
 59 morphological, physiological and genomic attribute as compared to its closest neighbor. Based
 60 on all these differences, we propose a novel species of genus *Blautia* named as *Blautia*
 61 *brookingsii* SG772.

62 **Organism information**

63 **Growth conditions and biochemical properties**

During culturomics of healthy human fecal sample, strain SG772 was isolated from a healthy human fecal sample using Brain Heart Infusion (BHI) agar medium anaerobically at 37°C. Morphologically, colonies were round, whitish, convex and smooth after 48 hours incubation on BHI agar. Further, cellular morphology of strain SG772 was examined using a scanning electron microscope. Microscopic study revealed the complete absence of flagellum, indicating that strain SG772 is non-motile. The size of the bacterium varied between 0.5-0.8 x 1.8-2.5 µm (Figure 1). The Gram-stain test was performed using a Gram staining kit (HiMedia) and the strain was found to be Gram-positive. Optimum temperature and pH for growth of the bacteria was found to be 37°C and 6.8 respectively.

As the strain SG772 exhibited no significant scores on Matrix-assisted desorption-ionization time-of-flight (MALDI-ToF) (Bruker Daltonics, Germany) (11), the genes encoding for 16S rRNA gene was amplified, sequenced using Applied Biosystems 3500xL, Genetic analyser (Applied Biosystems, MA, USA) at Department of Veterinary and Biomedical Sciences, South Dakota State University. The sequence was assembled using Sequencing Analysis version 5.1.1 Genious 10.2.3 (NJ, US). A continuous stretch of 1440 bp of the 16S rRNA gene of strain SG772 was obtained and this sequence was subjected to similarity search against EzTaxon- e-server (12). Analysis of 16S rRNA gene revealed highest sequence similarity (94.39%) to *Blautia stercoris* GAM6-1. Thereafter, the phylogenetic tree was constructed based on 16S rRNA gene sequence, together with top 30 taxonomically characterized strains. The evolutionary distance matrix was calculated using the distance model of Jukes & Cantor (1969) and an evolutionary tree was reconstructed using the neighbor joining method. The sequence of *Atobobium minutum* NCFB2751 was used as an outgroup (Figure 2). The resultant tree topology was evaluated by bootstrap analysis based on 1000 replicates (Felesenstein J, 1985) in Mega

version 5.2.2. Strain SG772 falls in the clade containing members belonging to the genus *Blautia*. (Figure. 2)

The growth pattern for the strain SG772 was determined using growth curve assay (Figure 3A). The biphasic growth pattern indicate its ability to use simple sugars in media initially with utilization of complex carbohydrates when simple sugars are depleted. Additionally, in order to elucidate the pattern of substrate oxidation, Biolog assay was performed using Biolog AN MicroPlate™ (Biolog Catalog # 70007) anaerobically. For this, 100 µl of overnight grown culture was plated onto 150 mm BHI agar plate and incubated for 48 hours anaerobically at 37⁰ C. Grown colonies were picked by a sterile cotton swab and inoculated to AN inoculating fluid (Biolog Catalog # 72007) until OD₆₅₀ reached 0.3. From this suspension, 100µl was pipetted into each well of 96 well biolog AN microplate in triplicate and incubated at 37⁰C anaerobically. OD₆₅₀ readings were taken at 0 hour and 24 hours post inoculation and results were analyzed and compared against *Blautia stercoris* GAM6-1 (Table 1). The strain was found to utilize 27 different substrates with maximum preference for D-mannitol (Figure 3B).

Antimicrobial susceptibility of the strain was tested using agar diffusion method in BHI agar medium. One hundred microliters of overnight grown culture of the strain was plated in BHI agar medium and antibiotic discs were placed. Zone of inhibition by antibiotics was measured after 24 hours of anaerobic incubation to determine the antibiotic sensitivity of the strain. The strain was found to be resistant to tetracycline and streptomycin. In contrast, the strain was susceptible to chloramphenicol, ampicillin, erythromycin and novobiocin.

Genome sequencing information

DNA isolation

Strain SG772 was sub-cultured on BHI agar and incubated at 37⁰C anaerobically for 48 hours. Further, it was cultured in 5 ml of BHI broth for 24 hours. Isolation of DNA was performed following E.Z.N.A DNA isolation kit (Omega, Biotek) protocol. Initially, 500µl of bacterial broth was pelleted by centrifuging at 10,000×g for 1 min. The pellet obtained was lysed in tris EDTA buffer using lysozyme for 1 hour and protein was digested using proteinase K overnight at 55⁰C. Subsequently, the mixture was pelleted by centrifuging at 8,000×g for 1 min and supernatant was spun in spin column followed by two washes of wash buffer. Finally, DNA was eluted using 50 µl of nuclease free water and kept at -20⁰C until use.

Genome sequencing, assembly and annotation

The genome was sequenced using Illumina MiSeq (Illumina Inc, CA) using 2x 250 paired end chemistry. Further, it was assembled using SPAdes 3.9.0 (13) and validated using Quast (14). Coding sequences were predicted using Glimmer 3.0 (15) and annotated with RAST 2.0 server (16) and the NCBI Prokaryotic Genomes Annotation Pipeline (PGAP) version 4.4. For core genome analysis, GenBank files were retrieved for strains SG772 along with its RAST neighbor *Blautia hansenii* DSM 20583 and were used for core genome analysis using OrthoMCL clustering algorithm (17) at 75% query coverage and sequence similarity (18). For functional annotation, the amino acid sequences were searched against the COG database to predict the abundance of COG gene families and heat map was constructed using Pearson correlation method and hierarchical clustering.

Genomic attributes

Assembly of the *B. brookingsii* SG772 genome produced 55 contigs with the genome size of 3.46 Mbp (N50 = 179,176) and 43.97% GC content (Table 2). The largest contig was of 401,827

bp while the smallest was of 624 bps. The genome contains 57 tRNA genes, 3343 coding sequences, a phage element and a CRISPR element (Table 2).

Insights from genome sequence

Insight into the genetic repertoire

In order to compare the newly sequenced genome of strain SG772 to its closest RAST neighbor *B. hansenii* DSM 20583 (8), we used BLAST Ring Image Generator (BRIG) (19) as a tool to show genome wide sequence similarity using *B. hansenii* DSM 20583 as a reference genome (Figure 4). Based on average nucleotide identity, these two genomes were 81.69% identical (19). Furthermore, this comparison revealed several differences in genome size, GC content and RNA copies (Table 2). For functional annotation, the amino acid sequences were searched against the COG database to predict the abundance of COG gene families and were compared against *B. hansenii* DSM 20583 (21). The gene families for energy production and conversion (C), amino acid transport and metabolism (E), carbohydrate transport and metabolism (G), transcription, ribosomal structure and biogenesis (J), transcription (K), replication, recombination and repair (L), cell wall/membrane/envelope biogenesis (M), inorganic ion transport and metabolism (P), defense mechanism (V) and unknown functions (S) were differentially enriched among these two strains (Figure 3C). Core genome analysis revealed a total of 411 core genes coding for basic metabolic functions. A total of 4,880 COG functions were annotated out of which only 70% were assigned to known categories. Additionally, resistance to cadmium, tetracycline, vancomycin, beta lactamase and fluoroquinolones along with dormancy and sporulation genes were relatively abundant in strain SG772. Furthermore, the genome was devoid of genes for motility and chemotaxis suggesting its non-motile phenotype.

Conclusions

This study presents the genome sequence for the strain SG772, with marked physiological and genomic differences from its neighbors *i.e.*, *B. stercoris* GAM6-1 and *B. hansenii* DSM20583. Based on the differences, we propose a novel species of genus *Blautia* named as *Blautia brookingsii* SG772.

Taxonomic and nomenclatural proposals

Blautia brookingsii (*brookingsii* referring to the isolation site of the type strain from Brookings, SD, USA). The cells are Gram stain positive, non-motile, coccobacillus, and 1.8-2.5 µm in length and 0.5-0.8 µm in diameter. Colonies were whitish, circular, smooth, and convex after 48 h of incubation in BHI agar. Optimal growth was observed at 37°C and pH of 6.8. It assimilates D-Mannitol, D-Melezitose, Adonitol, D-Glucosaminic acid, D-Arabitol, D- Sorbitol, D-Trehalose etc. Furthermore, the strain was resistant to tetracycline and streptomycin antibiotics whereas, it was susceptible to chloramphenicol, erythromycin, novobiocin and ampicillin. Genome sequence revealed the genome size of the strain SG772 to be 3.46Mbp with G+C content of 43.97 %. The type strain SG772 isolated from hlthy human fecal sample collected at Brookings, SD, USA and has been deposited in the Microbial Culture Collection at the National Centre for Cell Science, India.

Declarations

Authors declare no competing interests.

Acknowledgements

This work was supported in part by the USDA National Institute of Food and Agriculture, Hatch projects SD00H532-14 and SD00R540-15 to JS, and a grant from the South Dakota Governor's Office of Economic Development awarded to JS, EN and JH.

References

1. McLellan SL, Newton RJ, Vandewalle JL, Shanks OC, Huse SM, Eren AM, Sogin ML. 2013. Sewage reflects the distribution of human faecal Lachnospiraceae. *Environ Microbiol* 15:2213-27.
2. Biddle A, Stewart L, Blanchard J, Leschine S. 2013. Untangling the Genetic Basis of Fibrolytic Specialization by Lachnospiraceae and Ruminococcaceae in Diverse Gut Communities. *Diversity* 5:627-640.
3. Young VB. 2017. The role of the microbiome in human health and disease: an introduction for clinicians. *BMJ* 356:j831.
4. Million M, Diallo A, Raoult D. 2017. Gut microbiota and malnutrition. *Microb Pathog* 106:127-138.
5. Takahashi K, Nishida A, Fujimoto T, Fujii M, Shioya M, Imaeda H, Inatomi O, Bamba S, Sugimoto M, Andoh A. 2016. Reduced Abundance of Butyrate-Producing Bacteria Species in the Fecal Microbial Community in Crohn's Disease. *Digestion* 93:59-65.
6. Reeves AE, Koenigsnecht MJ, Bergin IL, Young VB. 2012. Suppression of *Clostridium difficile* in the gastrointestinal tracts of germfree mice inoculated with a murine isolate from the family Lachnospiraceae. *Infect Immun* 80:3786-94.
7. Kameyama K, Itoh K. 2014. Intestinal colonization by a Lachnospiraceae bacterium contributes to the development of diabetes in obese mice. *Microbes Environ* 29:427-30.
8. Liu C, Finegold SM, Song Y, Lawson PA. 2008. Reclassification of *Clostridium coccoides*, *Ruminococcus hansenii*, *Ruminococcus hydrogenotrophicus*, *Ruminococcus luti*, *Ruminococcus productus* and *Ruminococcus schinkii* as *Blautia coccoides* gen. nov., comb. nov., *Blautia hansenii* comb. nov., *Blautia hydrogenotrophica* comb. nov., *Blautia luti* comb. nov., *Blautia producta* comb. nov., *Blautia schinkii* comb. nov. and description of *Blautia wexlerae* sp. nov., isolated from human faeces. *Int J Syst Evol Microbiol* 58:1896-902.
9. Caballero S, Kim S, Carter RA, Leiner IM, Susac B, Miller L, Kim GJ, Ling L, Pamer EG. 2017. Cooperating Commensals Restore Colonization Resistance to Vancomycin-Resistant *Enterococcus faecium*. *Cell Host Microbe* 21:592-602 e4.
10. Eren AM, Sogin ML, Morrison HG, Vineis JH, Fisher JC, Newton RJ, McLellan SL. 2015. A single genus in the gut microbiome reflects host preference and specificity. *ISME J* 9:90-100.
11. Seng P, Abat C, Rolain JM, Colson P, Lagier JC, Gouriet F, Fournier PE, Drancourt M, La Scola B, Raoult D. 2013. Identification of rare pathogenic bacteria in a clinical microbiology laboratory: impact of matrix-assisted laser desorption ionization-time of flight mass spectrometry. *J Clin Microbiol* 51:2182-94.
12. Chun J, Lee JH, Jung Y, Kim M, Kim S, Kim BK, Lim YW. 2007. EzTaxon: a web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequences. *Int J Syst Evol Microbiol* 57:2259-61.
13. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA.

2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455-77.
14. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072-5.
15. Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673-9.
16. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: rapid annotations using subsystems technology. BMC genomics 9:75.
17. Li L, Stoeckert CJ, Jr., Roos DS. 2003. OrthoMCL: identification of ortholog groups for eukaryotic genomes. Genome Res 13:2178-89.
18. Contreras-Moreira B, Vinuesa P. 2013. GET_HOMOLOGUES, a versatile software package for scalable and robust microbial pangenome analysis. Appl Environ Microbiol 79:7696-701.
19. Konstantinidis KT, Tiedje JM. 2005. Genomic insights that advance the species definition for prokaryotes. Proc Natl Acad Sci U S A 102:2567-72.

Table 1: Physiological features of strain SG-772 compared to phylogenetic neighbor *Blautia stercoris* GAM6-1.

Characteristics	SG772	<i>Blautia stercoris</i> GAM6-1*
Habitat	Human gut	Human gut
Classification	Domain: Bacteria	Domain: Bacteria
	Phylum: Firmicutes	Phylum: Firmicutes
	Class: Clostridia	Class: Clostridia
	Order: Clostridiales	Order: Clostridiales
	Family: Lachnospiraceae	Family: Lachnospiraceae
	Genus: <i>Blautia</i>	Genus: <i>Blautia</i>
	Species: <i>brookingsii</i>	Species: <i>stercoris</i>
Optimum temperature	37°C	37°C
Shape	Coccobacilli	Cocci
Optimum pH	6.8	6.2
α-Galactosidase activity	-	+
β-Galactosidase activity	-	+
α-Glucosidase	-	+
β –Glucosidase	-	+
N-Acetyl-D-glucosamine	-	+
Turanose	-	-
Erythritol	-	+
Salicin	-	+
L-Glutamine	-	+
α-methyl-D-glucoside	-	NA
Glycyl-L- Proline	-	NA

Glycyl-L- Glutamine	-	-
Dextrin	+	NA
Arbutin	+	NA
D-Saccharic Acid	+	NA
a-D-Glucose	+	NA
L-Valine	+	NA
D-Cellobiose	+	NA
L-Rhamnose	+	NA
Succinic Acid	+	NA
D-Galactose	+	-
D-Mannose	+	NA
Palatinose	+	NA
Uridine-5'- Mono- phosphate	+	-
L-Fucose	+	NA
D-Glucose- 6-Phosphate	+	NA
Lactulose	+	+
D-Trehalose	+	NA
Succinic Acid Mono-Methyl Ester	+	NA
b-Cyclodextrin	+	NA
D-Sorbitol	+	NA
3-Methyl-D- Glucose	+	NA
D-Arabitol	+	NA
D-Glucosaminic Acid	+	NA
Adonitol	+	+
D-Melezitose	+	NA
D-Mannitol	+	NA

(*Data adopted from Park et al., 2012)

Table 2: General attributes of *Blautia brookingsii* SG772 with *Blautia hansenii* DSM20583

Characteristics	<i>Blautia brookingsii</i> SG772	<i>Blautia hansenii</i> DSM20583
Genome size (bp)	3,459,071	3,065,946
Number of contigs	55	1
Coding sequences	3343	3073
%GC content	43.97	39.04
tRNA	57	65
rRNAs (5S, 16S, 23S)	1, 8, 7	5, 5, 5

ncRNAs	4	4
Pseudogenes	152	68
CRISPR elements	1	2
Phage	1	0

Figure 1: Scanning electron micrograph of strain SG772 grown on BHI agar at 37°C for 24 hours. Bar, 2.0 μm

Figure 2: 16s rRNA sequence based phylogenetic tree constructed using neighbor-joining method. The 16S rRNA gene sequence was compared against the EZ-taxon, which showed the

highest similarity (94.39%) with *Blautia stercoris* GAM-61. The sequence of *Atobobium minutum* NCFB2751 was used as an outgroup. Numbers at nodes indicate bootstrap values expressed as percentages of 1000 replications. Bar, 0.02 accumulated changes per nucleotide. GenBank accession numbers are shown in parentheses.

Figure 3: Phenotypic and genotypic profiling of *Blautia sp. SG772*- (A) Growth curve of the strain *Blautia sp. SG772*, (B) Bar plot depicting different patterns of substrate utilization, (C) Bar graph depicting comparative abundance of COG gene families between the two genomes.

Figure 4: Comparative genome map of *Blautia* sp. SG772 using *B. hansenii* DSM 20583 as a reference. From the inside out, circle 1 represents the mean centered G+C content; circle 2 shows GC skew calculated as $(G - C) / (G + C)$; circle 3 represents the genome sequence similarity of *Blautia* sp. SG772 based on color gradient against the reference genome of *B. hansenii* DSM 20583; while circle 4 represents the genome of *B. hansenii* DSM 20583.