Title: The dark side of the mean: brain structural heterogeneity in schizophrenia and its polygenic risk.

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35 **Key Points**

36 **Question:** Is schizophrenia and its polygenic risk associated with brain structural
37 heterogeneity in addition to mean changes?

38 **Findings:** In a sample of 1151 patients and 2010 controls, schizophrenia was associated with
39 increased heterogeneity in fronto-temporal thickness, cortical, ventricle, and hippocampal
40 volumes, besides robust reductions in mean estimates. In an independent sample of 12,490
41 controls, polygenic risk for schizophrenia was associated with thinner fronto-temporal
42 cortices and smaller CA2/3 of the left hippocampus, but not with heterogeneity.

43 **Meaning:** Schizophrenia is associated with increased inter-individual differences in brain
44 structure, possibly reflecting clinical heterogeneity, gene-environment interactions, or
45 secondary disease factors.

46

47 **Abstract**

48 **Importance:** Between-subject variability in brain structure is determined by gene-
 49 environment interactions, possibly reflecting differential sensitivity to environmental and
 50 genetic perturbations. Magnetic resonance imaging (MRI) studies have revealed thinner
 51 cortices and smaller subcortical volumes in patients. However, such group-level comparisons
 52 may mask considerable within-group heterogeneity, which has largely remained unnoticed in
 53 the literature

54 **Objective:** To compare brain structural variability between individuals with SZ and healthy
 55 controls (HC) and to test if respective variability reflects the polygenic risk for SZ (PRS) in
 56 HC.

57 **Design, Setting, and Participants:** We compared MRI derived cortical thickness and
 58 subcortical volumes between 2,010 healthy controls and 1,151 patients with SZ across 16
 59 cohorts. Secondly, we tested for associations between PRS and MRI features in 12,490
 60 participants from UK Biobank.

61 **Main Outcomes and Measures:** We modeled mean and dispersion effects of SZ and PRS
 62 using double generalized linear models. We performed vertex-wise analyses for thickness,
 63 and region-of-interest analysis for cortical, subcortical and hippocampal subfield volumes.
 64 Follow-up analyses included within-sample analysis, controlling for intracranial volume and
 65 population covariates, test of robustness of PRS threshold, and outlier removal.

66 **Results:** Compared to controls, patients with SZ showed higher heterogeneity in cortical
 67 thickness, cortical and ventricle volumes, and hippocampal subfields. Higher PRS was
 68 associated with thinner frontal and temporal cortices, as well as smaller left CA2/3, but was
 69 not significantly associated with dispersion.

70 **Conclusion and relevance:** SZ is associated with substantial brain structural heterogeneity
 71 beyond the mean differences. These findings possibly reflect higher differential sensitivity to

72 environmental and genetic perturbations in patients, supporting the heterogeneous nature of
 73 SZ. Higher PRS for SZ was associated with thinner fronto-temporal cortices and smaller
 74 subcortical volumes, but there were no significant associations with the heterogeneity in these
 75 measures, i.e. the variability among individuals with high PRS were comparable to the
 76 variability among individuals with low PRS. This suggests that brain variability in SZ results
 77 from interactions between environmental and genetic factors that are not captured by the
 78 PGR. Factors contributing to heterogeneity in fronto-temporal cortices and hippocampus are
 79 thus key to further our understanding of how genetic and environmental factors shape brain
 80 biology in SZ.

81 Introduction

82 Schizophrenia (SZ) is a severe psychiatric disorder with a lifetime prevalence of about 1%,
 83 rendering it a leading cause of disability worldwide with 26 million people affected¹. While
 84 genetic and environmental factors contributing to disease risk have been identified, the
 85 pathophysiology still remains elusive^{2,3}. Patients diagnosed with SZ display substantial
 86 heterogeneity in terms of their clinical characteristics and symptoms⁴, treatment response⁵ and
 87 long term prognosis⁶. The notion that the observed heterogeneity stems at least partially from
 88 distinct subtypes of patients with differentially affected neurobiology and clinical and
 89 cognitive profiles⁷⁻⁹, has not been fully confirmed¹⁰, and the question of whether there is one
 90 unifying pathophysiological process shared across patients, or a multitude of disease
 91 processes leading to a similar clinical syndrome remains salient¹¹.

92 SZ is associated with widespread brain abnormalities, with the most robust group-
 93 level mean structural differences being ventricle enlargement, reduced thickness and area of
 94 frontal and temporal cortices, as well as reduced hippocampal and amygdala volumes¹²⁻¹⁴.
 95 However there is also substantial variability between patients^{7,8,15,16}, presenting a major
 96 challenge for achieving imaging based diagnostic predictions with any clinical utility^{17,18}.
 97 Rather than simply reflecting noise, this inter-individual variability in brain structure may
 98 possibly carry relevant information regarding gene-environment interactions related to the
 99 individual sensitivity to environmental and genetic perturbation. Only a few studies have
 100 investigated whether heterogeneity differs between healthy participants and SZ patients. One
 101 functional imaging study reported increased heterogeneity in both connectivity and spatial
 102 extent of functional brain networks in SZ¹⁹. Regions with altered spatial variance in functional
 103 networks included areas previously implicated in SZ, such as auditory and sensorimotor
 104 cortices and basal ganglia, and networks showing increased heterogeneity overlapped with
 105 those showing mean volume differences, implying that the mean and variance measures

provide complementary but converging results²⁰. A recent meta-analysis reported increased inter-individual volumetric variability in several cortical and subcortical structures, including the temporal lobe, thalamus, hippocampus and amygdala in SZ, and lower variability in the anterior cingulate cortex (ACC)¹⁵. These results point to the importance of modeling heterogeneity as well as mean changes. Detecting brain regions that are more homogenous in patients could point to a primary role in a shared underlying SZ pathophysiology, while regions of increased heterogeneity might be informative of putative subtypes of disease, or possibly reflect regional differences in the sensitivity to genetic and environmental perturbations.

SZ is highly heritable²¹, motivating the ongoing efforts to identify intermediate brain phenotypes associated with disease liability in order to elucidate the pathway from genes to illness manifestation. Several SZ risk loci have been identified²², but the individual contribution of each identified variant is weak and as of yet no common variants have been conclusively linked to the disease. Polygenic risk scores (PRS) for SZ, which represent a weighted sum of common genetic SZ risk alleles, have been proposed to account for the polygenic nature of disease risk and to increase predictive power²³. Beyond being predictive of case-control status²², SZ PRS have been associated with negative symptoms, anxiety and lower cognitive ability in adolescents²⁴. Polygenic burden has also been linked to reduced cortical thickness, as well as with prefrontal working memory-, and hippocampal encoding-related activation and connectivity in both patients and healthy participants²⁵⁻²⁸. This is in line with findings implicating both the frontal cortices and hippocampus as core regions in SZ pathophysiology²⁹. Polygenic risk for SZ is however only weakly associated with subcortical volumes³⁰. Importantly, risk alleles could also exert their effect by influencing the environmental sensitivity, which could be reflected in the phenotypic variability between individuals³¹.

Thus, revealing brain structures with higher or lower heterogeneity in SZ could facilitate discovery of intermediate brain phenotypes that may serve to identify putative subtypes^{8,32} of the disease, as well as by identifying phenotypes that are primary or common in the neurobiology of SZ¹⁵. Further, investigating how the genetic architecture of disease risk is related to brain heterogeneity could reveal regions in which the cumulative burden of common risk alleles influence the phenotypic variance³³. To this end, we directly compared the within-group dispersion in several key brain structural phenotypes, including cortical thickness, as well as in cortical, subcortical and hippocampal subfield volumes, between 1151 patients with SZ and 2010 healthy controls. Next, in order to test the whether between-subject variability is associated with the cumulative polygenic risk for SZ, we tested for associations between dispersion in the same brain features and PGR for SZ in 12,490 healthy individuals from the UK Biobank.

Material and methods

Samples: Sample characteristics are presented in Table 1 and eMethods (Samples), and cohort details are presented in eTable 1.

Image preprocessing

T1-images were processed using Freesurfer 5.3.0 (<http://surfer.nmr.mgh.harvard.edu>) for cortical reconstruction and volumetric segmentation³⁴⁻³⁷ and Freesurfer 6.0 for hippocampus subfield segmentation³⁸. Each participant's cortical thickness map was registered to the MNI305 fsaverage template and spatially smoothed (15 mm FWHM).

Genetic data

PRS were calculated using PRSice³⁹ v1.25, based on the European-Caucasian subset of the 2014 PGC2 schizophrenia GWAS²². The scores were calculated at a range of p-value

thresholds, from 0.001 to 0.5, with intervals of 0.001 (eMethods: Genetics). PRS based on a threshold of .05 were used for the main analysis since this threshold has been reported as optimal in terms of explaining case-control differences⁴⁰, but we also performed follow-up analysis to test the robustness of findings to threshold selection.

Statistical analysis

Statistical analyses were performed in R (3.2.2; <https://www.r-project.org/>) and MATLAB (2014a, The MathWorks, Inc., Natick, Massachusetts, United States). For all included measures in the multi-scanner case-control sample we used vertex/volume-wise generalized additive models (GAM, R-Package: mgcv⁴¹) to regress out scanner effects while accounting for age, sex and diagnosis. We then modeled vertex/volume-wise mean and dispersion effects using double generalized linear models (DGLMs), which iteratively fits a GLM modeling the mean parameter, and a second GLM modeling the dispersion parameter (eMethods: DGLMs; R-package: dglm⁴²). We then permuted the SZ and PRS column vector, respectively, and recalculated the mean and dispersion effects. For cortical thickness the true test statistics and the permuted statistical maps were then submitted to PALM⁴³ to correct for multiple comparisons using threshold-free cluster enhancement (TFCE) and tail approximation⁴⁷ (600 permutations: eMethods: Permutation). Maps were thresholded at a significance level of $p < .05$. For the cortical, subcortical, and hippocampal subfields volumes we performed 5000 permutations per volume and extracted the maximum t-value across ROIs when computing p-values in order to correct for multiple comparisons. Significance threshold was set at $p < .05$. We also performed a meta-analysis of the multi-scanner thickness data (R-Package: metafor⁴⁴) estimating case-control difference within each sample, and conducted analyses both with and without covarying for estimated intracranial volume (eTIV) for the volumetric measures. We performed follow-up analyses with more stringent exclusion criteria

(eMethods: Outliers). To assess the effect of PRS p-threshold selection, we performed a principal component analysis (R-package: prcomp, factoextra⁴⁵) on PRS scores calculated across several thresholds (eMethods: PRS-PCA), and reran cortical thickness analysis on PCA-scores.

Results

Vertex-wise thickness: SZ was associated with decreased mean thickness globally, with the exception of the visual cortex, as well as globally increased thickness dispersion (Figure 1, panel A and B, Figure 2A; eFigure 2). Meta-analysis of within-sample effects with more stringent exclusion criteria revealed significantly increased heterogeneity in SZ, indicating that dispersion effects are not simply explained by multi-site variability or a few extreme values (eFigure 3). PRS was associated with lower mean thickness in the right inferior frontal gyrus, the right lateral orbitofrontal cortex, the right pre-central gyrus, the right medial temporal cortex, and bilaterally in middle and superior temporal cortices (Figure 1, panels C and D; eFigure 4). Converging results were obtained upon re-analysis with the addition of the first four population components added as covariates (eFigure 5A), or with more stringent exclusion criteria (eFigure 5B). Follow-up analysis using the first component from the PRS principal component analysis (PRS-PC1) gave close to an identical pattern as the PRS-model based on a threshold of .05 (eFigure 5C; vertex-wise $r=.91$). There was no significant associations between polygenic risk and thickness dispersion, or between PRS-PC2 or PRS-PC3 and mean or dispersion of cortical thickness. Vertex-wise correlations between raw t-maps for SZ and polygenic risk were $r=.2$ and $r=.1$, for the mean and dispersion respectively (eFigure 6).

Cortical and subcortical volumes: SZ was associated with lower mean cortical volume, supratentorial volume, total and subcortical gray volume, cerebellar cortical volume, as well as brain stem, hippocampus, amygdala, thalamus and nucleus accumbens, and several white matter volumes, as well as increased ventricle, caudate nucleus, pallidum and putamen volumes. SZ was further associated with increased dispersion in mean cortical volume, total gray volume, left hippocampus and ventricle volumes (Figure 2B, Figure 3). Models without eTIV (eFigure 7A) revealed no significant differences in the mean volumes of caudate nucleus and left putamen, and resulted in an additional significant association with dispersion in supratentorial volume. Reanalysis of mean and dispersion models with more stringent exclusion criteria showed converging results (eTable 2). PGR was not associated with mean or dispersion in any of the subcortical volumes (Figure 3B), this was also true for models without adjustment for eTIV (eFigure 7B).

Hippocampal subfields: Patients with SZ was associated with lower mean volume in both left and right whole hippocampus as well as in all hippocampal subfields, accompanied with larger right hippocampal fissures. There was increased dispersion in SZ in left whole hippocampus, as well as in left molecular layer, left granule cell layer of the dentate gyrus (GC-DG) and left CA4 (Figure 4A). Models without adjustment for eTIV gave the same results for mean volumes with the exception for left hippocampal fissure, which did not survive correction. Additional dispersion effects were observed for right whole hippocampus, and left CA1 (eFigure 8A). When reanalyzing SZ mean and dispersion models with more stringent exclusion criteria, we obtained similar results (eTable 3). PGR was associated with smaller left CA2/3. None of the subfields showed an association between volume dispersion and PRS (Figure 4B). Models without adjustment for eTIV revealed smaller left and right CA2/3, left granule cell layer of the dentate gyrus (GC-DG), and left CA4 (eFigure 8B) in

patients with SZ. Re-analysis with population covariates added to the models, re-analysis with stricter exclusion criteria, and modeling PRS using the first principal component, did not alter conclusions (eTable 4).

Discussion

In the current study we found that SZ is associated with increased brain heterogeneity in cortical thickness, as well as in cortical volumes, lateral and third ventricles and hippocampal volumes. The findings, based on harmonized analysis protocols for all included datasets, were robust to strict procedures for removing outliers, and follow-up meta-analysis confirmed that multi-site case-control difference cannot be explained by scanning site. These findings are largely in line with a recent meta-analytic study showing increased volumetric heterogeneity in the temporal lobe and lateral and third ventricles¹⁵. It also extends this meta-analysis by showing increased heterogeneity in cortical thickness as well as in specific hippocampal subfields. Further, increased polygenic risk in healthy individuals was associated with reductions in thickness in frontal and temporal regions, as well as in left dentate gyrus and CA4 and bilateral CA2/3, but not related to thickness dispersion.

We found widespread reductions in cortical thickness in SZ patients, with the characteristic pattern of stronger fronto-temporal effects, as well as global reductions in cortical volume⁴⁶. In addition to these mean changes, we found that SZ is also associated with increased thickness heterogeneity compared to healthy controls. No cortical region showed the opposite pattern of increased homogeneity among patients. As with previous studies we found mean reductions in several brain volumes, with the most robust effects for cortical volume, cerebellum and hippocampus, as well as ventricle enlargement. These regions additionally showed increased heterogeneity in patients compared to controls, and again no region showed increased homogeneity, as might result if a particular region was similarly

affected by a common pathophysiological mechanism¹⁵. Instead, the results are in line with previous studies suggesting substantial neurobiological heterogeneity in SZ¹⁶. They do however contrast with a recent report¹⁵ of increased volumetric homogeneity in SZ. One possible explanation is differing sample inclusion criteria, as the previous study included only first-episode psychosis patients. It is not unlikely that an earlier disease stage may offer a more direct window into core aspects of the pathophysiology which later shift towards increased inter-individual variability as patients vary across different illness stages and degrees of severity, as well as differences in treatment and medication status.

PRS reflect cumulative risk across multiple genetic loci, and SZ PRS is associated with several phenotypic traits, including liability for psychiatric disease such as bipolar disorder and schizoaffective disorder, negative symptoms, as well as IQ, working memory performance and brain activation^{25,47,48}. SZ-PRS has been associated with cortical gyrification in healthy participants⁴⁹, as well as reductions in global cortical thickness²⁷. The current results show that higher SZ-PRS in healthy participants are associated with mean decrease in thickness in fronto-temporal cortices. Interestingly, these shifts in mean thickness were not associated with changes in brain heterogeneity, as was found for patients, pointing to differential effects of genetic risk on mean and heterogeneity changes. A recent study found that SZ environmental risk scores and SZ-PRS scores are both independently related to frontotemporal cortical thinning in patients, but not controls²⁷. This underscores the importance of also investigating environmental risk factors, as well as gene-environment interplay, and their role in explaining the observed clinical and neurobiological heterogeneity

With regard to hippocampal volumes, we found that higher polygenic risk was associated with smaller volumes of the left CA2/3, in the absence of an effect on total hippocampal volume and after correcting for total intracranial volume, suggesting a specific effect of genetic risk on this region. The hippocampus has been hypothesized to play a

primary role in the pathophysiology of SZ, through progressive changes to its neural circuits as the disease evolves²⁹. Our results also complement recent studies reporting that polygenic risk for SZ is predictive of hippocampal activation during memory encoding²⁸, and of polygenic overlap between SZ and hippocampus volume⁵⁰. Also, while the patients showed increased hippocampal heterogeneity, only mean effects were associated with PRS, mirroring the findings on cortical thickness. Thus, the dentate gyrus and CA2/3 emerge as key regions both for the manifestation of, and the genetic risk for, SZ and potentially informative for the classification of sub-types and degrees of severity.

Despite reliable associations between SZ and brain morphometry¹⁴, PRS is only weakly associated with subcortical volumes, however a recent study found polygenic overlap for SZ and hippocampal, putamen and intracranial volumes⁵⁰. The lack of associations between PRS and subcortical volumes in the current study is in line with most previous reports on PRS³⁰.

Limitations

An important source of heterogeneity in the present case-control sample could be the large number of different scanning-sites included. However, in addition to residualizing for scanner-site in the main analysis, we also performed within-sample analysis and ran meta-analysis, which rule out scanner as a major contributor to the observed effects. Heterogeneity could be associated with differences in medication status and duration of illness. While we did find increased caudate and putamen volumes, indicative of medication effects⁵¹, these volumes did not show altered heterogeneity in SZ. Still, antipsychotic medication could possibly affect brain heterogeneity in other regions, in absence of a change in mean volume. Investigation of such effects requires carefully controlled settings, and is therefore difficult to address in large-scale multi-site studies. Another possible explanation is that the increased variability is caused by movement artefacts, which are typically greater in clinical

populations⁵², however running the analysis in a subset with stricter criteria for dataset exclusion did not alter the conclusions. One important consideration for case-control studies in general is the possibility that healthy controls are abnormally normal compared to the general healthy population due to selection bias and strict exclusion criteria⁵³, underscoring the importance of studying the full range of phenotypic variability in the population. Further, the validity of choosing a given p-threshold selection among several possible thresholds when calculating PRS is uncertain. We addressed this by performing follow-up analysis where we performed PCA across PRS calculated across a wide range of thresholds, to derive a more general score of polygenic risk. This approach yielded results converging with the main analysis using a threshold of $p < .05$. The lack of association between SZ-PRS and brain heterogeneity suggest that the current SZ-PRS scores do not strongly reflect variance-controlling variants. As a composite score, SZ-PRS likely also hides a substantial genetic heterogeneity. A PRS-score calculated using a variance-controlling trait loci (vQTL) approach would likely be more sensitive in detecting such effects. Lastly, SZ is increasingly understood as a neurodevelopmental disorder⁵⁴ and disentangling the sources of heterogeneity in the adult patient population likely requires investigation of the life-span trajectories and aberrant developmental paths.

Conclusion

There are ongoing efforts to account for brain heterogeneity by means of delineating patient subtypes^{8,55}, as well as characterizing patients by their differential degree of affectedness along one or multiple symptom-axes^{11,56}. Here we report that SZ is associated with widespread and increased heterogeneity in cortical thickness, and cortical as well as hippocampal volume, beyond the known mean differences, compared controls. The results support to the notion that SZ is a highly heterogeneous disorder, and suggests that important information may be overlooked when only assessing mean differences. In healthy adults SZ-

333 PRS were associated with mean changes in brain areas implicated in SZ, but not associated
 334 with altered brain heterogeneity. Taken together, these findings warrant future longitudinal
 335 studies which can disentangle the genetic and environmental factors contributing to diverging
 336 trajectories and neurobiological heterogeneity, and in particular how these factors contribute
 337 to heterogeneity in fronto-temporal cortices and hippocampus.

Collaborators

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Author contribution statement

D.A. and L.T.W conceived of the study; D.A. performed the analysis with contributions from L.T.W., T.K and D.M; A.C.P., T.M., J.R., F.B., I.A., D.M.B., A.B., C.L.B., S.C., S.D., N.T.D., S.E., H.F.B., L.F., A.D.G., B.H., E.G.J., P.K., M.J.L., A.M.L., G.P., E.S., O.B.S., T.Q., M.Z., and O.A.A contributed with data preprocessing, quality assurance and interpretation of results. D.A., T.K. and L.T.W. wrote the first draft of the paper, and all authors contributed to the final manuscript. D.A. and L.T.W. had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Disclosures

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Figure 1. Mean and dispersion of cortical thickness. All maps were thresholded using permutation testing, threshold free cluster enhancement, and fitting the tail of the permutation distribution to a generalized Pareto distribution (500 permutations, $p < .05$, FWE). **A:** t-map for the SZ mean model; cold colors represent areas with decreased mean thickness in SZ compared to healthy controls. SZ was associated with decreased thickness globally, with the exception of the visual cortex, and with strongest effects in frontal and temporal regions, compared to healthy controls. **B:** t-map for the SZ dispersion model. Warm colors represent areas with increased heterogeneity in SZ compared to healthy controls. Inter-individual variability in cortical thickness showed a spatially global increase for the SZ-group compared to healthy controls. **C:** In an independent sample of healthy adults, the mean model showed that higher polygenic risk for SZ was associated with lower cortical thickness, represented by cold colors, in frontal and parietal cortices. **D:** Polygenic risk was not associated with cortical thickness heterogeneity in any region.

Figure 2. Shift function plots. The top panels show jittered marginal distribution scatterplots for variance model associations, with overlaid shift function plots using deciles. 95% confidence intervals were computed using a percentile estimation of the standard error of the difference between quantiles on 1000 bootstrap samples. The bottom panels show linked deciles from shift functions in top panels. **A:** Cortical thickness, vertex values were extracted by masking the images by the SZ-dispersion significance map, and averaged across vertices and hemispheres. Residualized for scanner, sex and age. SZ is associated with reduced thickness, with larger differences between groups in the lower deciles. **B:** 3rd ventricle volume, residualized for scanner, sex and age and eTIV. SZ is associated with larger volumes compared to controls, with the largest difference between groups in the upper deciles. **C-D:** Left whole hippocampus, and left molecular layer (hippocampal subfield), values residualized for scanner, sex and age and eTIV. SZ is associated with larger volumes compared to controls, with the largest difference between groups in the upper deciles.

Figure 3. Mean and dispersion of cortical and subcortical volumes. t-statistic for both mean (outline in light blue) and dispersion (outline in dark blue), filled blue dots mark significant effects after correction for multiple comparisons across regions (5000 permutations, permuted $p < .05$, FWE, eTIV-adjusted). **A:** The SZ-group had both decreased cortical and subcortical volumes, as well as increased ventricles and putamen and pallidum volumes. Cortical, hippocampal and ventricle volume were more heterogeneous in the SZ-group compared to healthy controls. **B:** Polygenic risk for SZ was not associated with mean changes nor dispersion in any of the regions.

Figure 4. Mean and dispersion of hippocampus subfields volumes. t-statistic for both mean (outline in light blue) and dispersion (outline in dark blue), filled blue dots mark significant effects after correction for multiple comparisons across regions (5000 permutations, permuted $p < .05$, FWE, eTIV-adjusted). **A:** The SZ-group had decreased hippocampal volumes, and this decrease was also evident in all subfields, and accompanied with an increase of the hippocampal fissures. Hippocampal volumes were also more heterogeneous in the SZ-group, and among the subfields this effect was present in the left molecular layer, bilateral CA1, right dentate gyrus and left CA4. **B:** Polygenic risk for SZ was associated with mean reductions of left dentate gyrus, left CA4 and bilateral CA2/3. Neither total hippocampal volumes nor any of the subfields showed a significant association between polygenic risk and volume heterogeneity.

572

Case-control samples								
Sample	HC				SZ			
	n	mean age	std	females (n)	n	mean age	std	females (n)
BrainGluSchi	47	33,7	12,7	18	37	37,2	15,3	3
CIMH	43	30,6	11,5	14	53	31,2	8,6	17
CNP-1	102	31,7	8,9	47	25	36,9	9,3	8
CPN-1	23	30,7	8,6	12	25	36,0	8,6	4
COBRE	90	38,1	11,7	26	88	38,1	13,7	18
HUBIN	102	42,0	8,8	33	94	41,7	7,6	24
KaSP	45	25,7	5,2	21	37	28,5	7,5	12
MCIC-1	43	29,0	11,3	9	35	32,6	12,4	7
MCIC-2	26	33,1	12,4	11	32	32,4	10,3	8
MCIC-3	24	40,8	9,4	9	33	38,3	10,0	8
NUNA	44	31,3	8,5	22	44	33,0	6,8	14
NUSDAST	95	33,8	14,5	44	114	35,0	13,1	38
TOP-3T-1	294	31,8	7,6	121	112	28,0	8,1	38
TOP-3T-2	301	35,8	10,1	141	110	33,0	9,9	44
TOP-1.5	295	35,5	9,6	131	225	31,9	9,1	96
UNIBA	436	26,7	7,7	225	87	34,1	7,6	22
Total	2010	32,7	10,4	884	1151	33,8	10,6	361
PRS sample								
Sample	n	mean age	std	females (n)				
UKB	12490	55,9	7,5	6465				

573

574 **Table 1. Sample characteristics.** We included data from 16 samples for the case-control

575 analysis, each sample contained MR-scans from both patients with SZ and healthy control

576 participants. -analysis was performed on UKB participants, excluding those with a diagnosed

577 neurological or mental disorder. More detailed information about samples and

578 acknowledgements are provided in eTable 1.

Polygenic risk, principal component analysis

eFigure 1: Principal component analysis on PRS scores calculated at different p-thresholds. We performed PCA on PRS scores from the UK Biobank sample (n=12490), calculated from a threshold of $p < .001$, to $p < .5$ (500 thresholds, step of .001). **A:** Scree plot showing explained variance for the first 10 components. **B:** Cross-correlation matrix of first four MDS components, first five principal components, and PRS for thresholds from .001 to .5 (steps of .001 up to $p < .01$, then steps of .01). The first three principal components explained 92.5%, 3.1% and 1.8% of the variance, respectively. The first components showed high correlation with PRS across all thresholds, but relatively lower for the stricter compared to the more lenient p-thresholds. Component 2 correlated negatively with PRS from the stricter p-thresholds, and positive with more lenient p-thresholds. Component 3 was positively

correlated with scores calculated with the strictest and most lenient p-thresholds, while
correlating negatively with PRS based on the intermediate p-thresholds.

Figure 2: SZ Vertex-wise mean and dispersion models. **A:** unthresholded t-map for the SZ mean model; warm/cold colors represent areas with increased and decreased mean thickness in SZ compared to healthy controls. **B:** $-\log_{10}(p)$ map for the SZ-mean model. Thresholded at $-\log(p) > 1.6$ ($p < .05$, two tailed, FWE). Derived by comparing the vertex-wise values of the actual TFCE-map for the SZ mean-model with TFCE-maps derived from

500 random permutations of the diagnosis labels. **C:** $-\log_{10}(p)$ map derived from fitting a generalized Pareto distribution to the tail of the permutation distribution. **D:** unthresholded t-map for the SZ dispersion model; warm/cold colors represent areas with increased and decreased thickness dispersion in SZ compared to healthy controls. **E:** $-\log_{10}(p)$ map for the SZ-dispersion model. Thresholded at $-\log(p) > 1.6$ ($p < .05$, two tailed, FWE). Derived by comparing the vertex-wise values of the actual TFCE-map for the SZ dispersion-model with TFCE-maps derived from 500 random permutations of the diagnosis labels. **F:** $-\log_{10}(p)$ map derived from fitting a generalized Pareto distribution to the tail of the permutation distribution.

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612 **eFigure 3: Meta-analysis of SZ mean and dispersion effects with stricter inclusion criteria.** The above panel shows the within-sample
 613 effects of the mean-model in the vertices showing a significant group-level effect for the left and the right cortical hemisphere. The bottom panel
 614 displays the dispersion effects. Both hemispheres showed a significant meta-analytic thickness decrease, as well as a dispersion increase,
 615 suggesting that the results from the main analysis are not driven by site-related variance not accounted for in the multi-site regression models.
 616 The analysis also included removal of subjects with either eTIV or mean thickness-values at $|z| > 3$, or significant outlier test (corrected $p < .05$).
 617 For CNP-2, right hemisphere, the DGLM did not converge.
 618

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620

621 **eFigure 4: Polygenic risk, vertex-wise mean and dispersion models. A:** unthresholded t-
622 map for the PRS mean model; warm/cold colors represent areas with increased and decreased
623 mean thickness with increased PRS. **B:** $-\log_{10}(p)$ map for the PRS-mean model. Thresholded
624 at $-\log(p) > 1.6$ ($p < .05$, two tailed, FWE). Derived by comparing the vertex-wise values of
625 the actual TFCE-map for the PRS mean-model with TFCE-maps derived from 500 random
626 permutations of the PRS scores. **C:** $-\log_{10}(p)$ map derived from fitting a generalized Pareto
627 distribution to the tail of the permutation distribution. **D:** unthresholded t-map for the PRS

dispersion model; warm/cold colors represent areas with increased and decreased thickness dispersion with increased PRS. **E:** $-\log_{10}(p)$ map for the PRS-dispersion model. Thresholded at $-\log(p) > 1.6$ ($p < .05$, two tailed, FWE). Derived by comparing the vertex-wise values of the actual TFCE-map for the SZ dispersion-model with TFCE-maps derived from 500 random permutations of the PRS scores. **F:** $-\log_{10}(p)$ map derived from fitting a generalized Pareto distribution to the tail of the permutation distribution.

eFigure 5: Reanalysis of polygenic risk score models on cortical thickness. To test the robustness of the mean effect of PRS on cortical thickness, we performed several follow-up analysis: **A:** Uncorrected t-map for PRS-scores, including the four first population covariates derived from multidimensional scaling (MDS) analysis. **B:** Uncorrected t-map for reanalysis with extreme scoring individuals removed (mean thickness or eTIV, $|z| > 3$, or outlier test significant at $p < .05$, FWE). **C:** Uncorrected t-map for the analysis using the first principal component from the PCA on PRS scores calculated across a wide range of p-thresholds.

Vertex-correlations - Schizophrenia and polygenic risk

Figure 6: Spatial overlap between SZ and PRS. The vertex-wise t-values for the SZ-models are shown on the x-axis and the vertex-wise t-values for the PRS-models on the Y-axis. The left plot shows the mean-model (vertex-wise correlation of .2) and the right plot shows the dispersion models (vertex-wise correlation of .1). The heatmaps represent the count of vertices at a value.

Cortical & subcortical volumes, mean and dispersion effects

Figure 7: Cortical and subcortical volumes, without adjustment for eTIV. t-statistic for both mean (light blue) and dispersion (dark blue), filled dots mark significant effects after

correction for multiple comparisons across regions (5000 permutations, permuted $p < .05$, FWE). **A:** The SZ-group showed decreased cortical and subcortical volumes, as well as increased ventricles and putamen and pallidum volumes. Cortical, right hippocampal and ventricle volumes were more heterogeneous in the SZ-group compared to healthy controls. **B:** Polygenic risk for SZ was not associated with mean changes nor dispersion in any of the regions.

Figure 8: Hippocampus subfields volumes, without adjustment for eTIV. t-statistic for both mean (light blue) and dispersion (dark blue), filled dots mark significant effects after correction for multiple comparisons across regions (5000 permutations, permuted $p < .05$, FWE). **A:** The SZ-group had decreased hippocampal volumes, and this decrease was also evident in all subfields, accompanied with a decrease of the left hippocampal fissure. Bilateral hippocampal volumes were also more heterogeneous in the SZ-group, and among the subfields this effect was present in the left molecular layer, left CA1, left GC-DG and left CA4. **B:** Polygenic risk for SZ was associated with mean reductions of the left GC-DG, left

673 CA4, and bilateral CA2/3. Neither total hippocampal volumes nor any of the subfields
674 showed a significant association between polygenic risk and volume heterogeneity.

Sample	Source	Comment	Reference
BrainGluSchi	http://schizconnect.org/ SchizConnect was funded by NIMH cooperative agreement 1U01 MH097435.	Data obtained from COINS source. Supported by NIMH-grant R01MH084898-01A1.	1
CIMH	Authors	CIMH was supported by the Deutsche Forschungsgesellschaft (DFG, projects KI 576/14-2, ZI1253/3-1, ZI1253/3-2) and the European Community's Seventh Framework Programme (FP7/2007–2013) grant agreement #602450 (IMAGEMEND)	2-4
CNP 1 & 2 (DS000030)	https://openfmri.org/	Data sets were obtained from the OpenfMRI database. Data from 2 imaging sites. DS000030 was supported by the Consortium for Neuropsychiatric Phenomics (NIH Roadmap for Medical Research grants UL1-DE019580, RL1MH083268, RL1MH083269, RL1DA024853, RL1MH083270, RL1LM009833, PL1MH083271, and PL1NS062410).	5,6
COBRE	http://schizconnect.org/	Data obtained from COINS source. Supported by grant 5P20RR021938 /P20GM103472 from the NIH to Dr. Vince Calhoun.	7
HUBIN	Authors	HUBIN was supported by the Swedish Research Council (2006-2992, 2006-986, K2007-62X-15077-04-1, 2008-2167, 2008-7573, K2010-62X-15078-07-2, K2012-61X-15078-09-3, 14266-01A,02-03, 2017-949), the regional agreement on medical training and clinical research between Stockholm County Council and the Karolinska Institutet, the Knut and Alice Wallenbergs Foundation and the HUBIN project	8,9
KaSP	Authors	KaSP was supported by grants from the Swedish Medical Research Council (SE: 2009-7053; 2013-2838; SC: 523-2014-3467), the Swedish Brain Foundation, Ahlén-stiftelsen, Svenska Läkaresällskapet, Petrus och Augusta Hedlunds Stiftelse, Torsten Söderbergs Stiftelse, the AstraZeneca-Karolinska Institutet Joint Research Program in Translational Science, Söderbergs Konungliga Stiftelse, Professor Bror Gadelius Minne, Knut och Alice Wallenbergs stiftelse, Stockholm County Council (ALF and PPG), Centre for Psychiatry Research, KID-funding from the Karolinska Institutet.	10,11
MCIC-1/2/3	http://schizconnect.org/	Data obtained from COINS source. MCIC was supported by the Department of Energy under Award Number DE- FG02-08ER64581.	12
NUNDA	http://schizconnect.org/	Data obtained from schizconnect.org. Source: NUNDA Supported by NIMH grant MH056584.	13
NUSDAST	http://schizconnect.org/	Data obtained from schizconnect.org. Source: NU_REDCAP. Supported by NIMH Grant 1R01 MH084803	14,15
TOP3T 1 & 2, TOP-1.5	Authors	The work was funded by the Research Council of Norway (213837, 223273, 204966/F20, 213694, 229129, 249795/F20, 248778), the South-Eastern Norway Regional Health Authority (2013-123, 2014-097, 2015-073, #2017-112) and Stiftelsen Kristian Gerhard Jebsen.	16-19
UKB	https://www.ukbiobank.ac.uk	This research has been conducted using the UK Biobank Resource (access code 27412)	20
UNIBA	Authors	This work was supported by a “Capitale Umano ad Alta Qualificazione” grant by Fondazione Con Il Sud awarded to Alessandro Bertolino and by a Hoffmann-La Roche Collaboration Grant awarded to Giulio Pergola. This project has received funding from the European Union Seventh Framework Programme for research, technological development and demonstration under grant agreement no. 602450 (IMAGEMEND). This paper reflects only the author's views and the European Union is not liable for any use that may be made of the information contained therein.	21

eTable1: Summary of included samples. We included publicly available data as well as data provided by co-authors. Sample reference list contains publications related to the sample as well as data sources.

Volume	SZ mean (t)	SZ Disp (t)	SZ mean outlier (t)	SZ disp outlier (t)
Total GrayVol	-18,04	3,41	-17,97	2,19
lhCortexVol	-17,20	3,15	-17,15	2,03
CortexVol	-17,05	3,24	-17,07	1,63
rhCortexVol	-16,65	3,20	-16,68	1,23
Right-Hippocampus	-13,86	2,43	-12,49	1,83
Left-Hippocampus	-13,83	2,39	-11,66	2,51
SupraTentorialVolNotVentVox	-12,85	2,40	-13,69	0,17
SupraTentorialVolNotVent	-12,41	1,99	-13,65	0,17
Left-Cerebellum-Cortex	-11,69	2,92	-11,20	-0,65
SupraTentorialVol	-11,43	-0,09	-12,63	0,03
Right-Cerebellum-Cortex	-10,69	-0,09	-10,53	-0,19
Brain-Stem	-9,64	-0,91	-9,20	-1,08
Left-Amygdala	-8,82	1,86	-8,60	1,58
Right-Thalamus-Propor	-8,35	-1,32	-7,00	1,96
Left-Thalamus-Propor	-7,53	0,45	-6,83	2,12
rhCorticalWhiteMatterVol	-7,11	-0,97	-6,12	0,75
CC_Mid_Posterior	-7,09	0,93	-8,25	-1,81
CorticalWhiteMatterVol	-6,97	-1,17	-6,26	1,49
Left-Cerebellum-White-Matter	-6,82	2,27	-7,38	-1,42
lhCorticalWhiteMatterVol	-6,64	2,24	-6,15	1,89
Right-VentralDC	-6,35	2,22	-5,87	0,12
Right-Cerebellum-White-Matter	-6,25	-1,47	-6,34	-1,30
CC_Central	-6,18	-0,14	-7,15	-0,95
Right-Amygdala	-6,16	0,81	-6,03	1,51
Left-VentralDC	-5,82	1,13	-4,44	-1,19
Right-Accumbens-area	-5,78	0,91	-5,18	0,76
CC_Mid_Anterior	-4,81	2,37	-6,31	-0,27
SubCortGrayVol	-4,63	-1,30	-2,72	1,57
CC_Posterior	-3,28	-0,04	-4,57	1,44
Left-Accumbens-area	-3,26	2,04	-2,80	-0,20
CC_Anterior	-2,33	1,05	-2,14	-0,18
Left-vessel	1,37	2,03	0,79	0,09
Right-vessel	1,69	-0,39	1,94	-0,03
Right-Caudate	2,16	1,56	4,60	1,57
4th-Ventricle	2,29	-0,05	2,51	0,66
Left-Caudate	4,00	2,20	4,76	1,57
5th-Ventricle	4,80	1,89	0,68	0,09
Left-choroid-plexus	5,05	1,74	4,07	1,72
Left-Putamen	5,48	0,62	5,62	0,06
Right-choroid-plexus	5,67	-0,91	6,52	2,43
Right-Putamen	6,44	4,48	7,94	0,20
Optic-Chiasm	6,48	2,17	5,83	-0,86
Left-Lateral-Ventricle	7,03	2,70	7,50	1,74
Right-Lateral-Ventricle	7,09	1,88	6,94	2,18
non-WM-hypointensities	7,40	1,54	8,19	1,69
CSF	8,19	1,95	5,31	2,04
Right-Inf-Lat-Vent	8,82	3,18	9,83	1,53
Right-Pallidum	9,55	2,33	9,70	1,99
Left-Pallidum	9,57	2,21	9,60	3,87
Left-Inf-Lat-Vent	9,77	5,57	12,03	4,19
3rd-Ventricle	12,24	5,78	9,741	4,190

eTable2: SZ cortical and subcortical analysis with and without outlier removal. We

excluded participants based on extreme values (described in Methods) and reran mean and

dispersion models without these participants.

Hippocampus subfield	SZ mean (t)	SZ Disp (t)	SZ mean outlier (t)	SZ disp outlier (t)	
Right Molecular Layer	-12,98	1,71	-12,53	1,54	
Right Whole Hippocampus	-12,86	2,16	-12,58	1,74	
Left Whole Hippocampus	-12,23	3,45	-12,28	2,30	
Left Molecular Layer	-11,78	3,45	-11,70	2,43	
Right GC-DG	-11,48	1,85	-11,11	1,61	
Right CA1	-11,37	1,18	-10,89	1,00	
Left CA1	-11,00	2,18	-11,10	1,04	
Right CA4	-10,99	1,55	-10,67	1,39	
Right presubiculum	-10,81	0,71	-10,45	0,46	
Left GC-DG	-10,72	3,22	-10,62	2,33	
Left HATA	-10,57	1,04	-10,57	0,56	
Left presubiculum	-10,36	2,18	-10,44	1,27	
Left CA4	-9,80	3,00	-9,69	2,30	s
Right subiculum	-9,36	1,71	-9,23	1,12	
Right HATA	-9,23	2,01	-8,81	0,90	
Left Hippocampal tail	-8,99	0,89	-9,06	1,14	
Left subiculum	-8,34	2,22	-8,30	1,22	
Right Hippocampal tail	-7,81	0,91	-7,36	0,43	
Right CA2/3	-7,31	0,02	-6,87	0,23	
Right parasubiculum	-7,18	-0,55	-7,15	-0,57	
Left parasubiculum	-6,56	0,78	-6,37	0,35	
Left CA2/3	-6,14	2,14	-6,20	1,47	
Left fimbria	-4,66	0,30	-4,52	0,65	
Right fimbria	-4,36	0,51	-4,23	1,10	
Left hippocampal fissure	3,99	0,83	4,71	0,23	
Right hippocampal fissure	5,93	0,57	6,33	-0,28	

eTable3: SZ hippocampal subfields analysis with and without outlier removal. We excluded participants based on extreme values (described in Methods) and reran mean and dispersion models without these participants.

Hippocampus subfield	PRS mean (t)	PRS Disp (t)	PRS mean MDS (t)	PRS disp MDS (t)	PRS mean outlier (t)	PRS disp outlier (t)	PCA1 mean (t)	PCA1 disp (t)
Left CA2/3	-3,00	-1,08	-2,99	-0,90	-2,67	-1,07	-2,76	-0,48
Left GC-DG	-2,26	-0,93	-2,15	-0,70	-2,10	-1,43	-1,95	-0,12
Right CA2/3	-2,20	-0,56	-2,10	-0,15	-1,98	-0,11	-2,34	0,21
Left CA4	-1,95	-0,87	-1,86	-0,65	-1,70	-1,35	-1,65	-0,13
Left Whole hippocampus	-1,83	-0,61	-1,62	-0,34	-1,68	-1,12	-1,36	-0,19
Right HATA	-2,04	-0,81	-1,96	-0,64	-2,18	-0,92	-2,39	-0,82
Left HATA	-2,15	-0,65	-2,14	-0,37	-2,03	-0,87	-2,06	-0,47
Left CA1	-1,79	-0,70	-1,55	-0,45	-1,87	-1,38	-1,41	-0,32
Left molecular layer	-1,71	-0,86	-1,47	-0,65	-1,48	-1,58	-1,31	-0,47
Right GC-DG	-1,61	-1,18	-1,39	-0,86	-1,59	-0,87	-1,51	-0,43
Right CA4	-1,54	-1,31	-1,33	-0,94	-1,44	-0,78	-1,45	-0,63
Right Whole hippocampus	-0,90	-1,17	-0,56	-0,82	-0,82	-1,03	-0,37	-0,63
Left parasubiculum	-1,37	-1,13	-1,35	-0,88	-1,09	-0,70	-1,50	-0,65
Right molecular layer	-0,81	-1,17	-0,46	-0,83	-0,84	-0,84	-0,35	-0,60
Left fimbria	-1,25	0,73	-1,29	0,89	-1,73	0,78	-1,90	0,55
Right CA1	-0,82	-0,76	-0,54	-0,39	-0,81	-0,66	-0,36	-0,11
Left subiculum	-0,79	-0,76	-0,58	-0,59	-0,72	-1,62	-0,57	-0,69
Left Hippocampal tail	-0,82	0,58	-0,75	0,77	-0,91	0,99	0,04	0,74
Right Hippocampal tail	-0,53	0,47	-0,41	0,70	-0,46	0,48	0,66	0,42
Left presubiculum	-0,32	-0,52	-0,07	-0,37	0,07	-0,60	0,17	-0,35
Right presubiculum	-0,15	-1,67	0,23	-1,40	0,49	-1,29	0,62	-0,85
Right fimbria	-0,15	0,24	-0,12	0,50	-0,78	0,12	-0,67	0,64
Right subiculum	0,54	-0,63	0,85	-0,27	0,57	-0,45	0,93	0,05
Right parasubiculum	0,21	-0,98	0,36	-0,80	0,74	0,03	0,40	-0,27
Left hippocampal fissure	0,46	-0,16	0,60	-0,21	0,41	-0,52	0,64	-0,09
Right hippocampal fissure	2,12	0,73	2,30	0,83	2,19	0,88	2,35	0,60

eTable4: SZ-PRS hippocampal subfields analysis with and without outlier removal. We excluded participants based on extreme values (described in Methods) and reran mean and dispersion models without these participants.

eMethods:

Samples: For the case-control analysis of cortical thickness and cortical and subcortical volumes we included MRI-scans from 16 cross-sectional study samples with a total of 3161 participants of which 2010 were healthy controls, and 1151 patients diagnosed with SZ. The case-control hippocampus subfield analysis was performed in a sub-sample of 2870 participants. The polygenic risk analysis was performed in a non-overlapping sample consisting of 12490 participants from the UK Biobank (UKB) that had MRI scans and genetic data available, all scanned on the same scanner³⁵. All UKB participants diagnosed with any ICD-10 mental or neurological disorder were excluded.

Genetics: Filtering include removal of SNPs with ambiguous alleles (AT or CG) and of variants in the major histocompatibility complex (MHC; chromosome 6, 26-33Mb), and pruning of SNPs based on linkage disequilibrium (LD) and p-value. All variants in LD with the local SNP with the smallest p-value were removed (the SNP with the smallest p-value within a 250kb window was retained, and all neighbours with a LD $r^2 > 0.1$ were removed; a step known as clumping).

DGLM: Modeling the dispersion is important for obtaining correct mean parameter estimates if dispersion varies as a function of the predictor, and also allows for systematic investigation into factors associated variability in observations⁴⁵. DGLMs were fitted using the following model specifications for both the mean and dispersion part; case control: $Y \sim Age + Sex + SZ$, and for polygenic risk: $Y \sim Age + Sex + PRS$, where Y is the mean in the first step, and the dispersion in the second step. For the group comparison we then obtain an estimate of the mean difference between groups, as well as an estimate of the difference in dispersion around the mean between SZ and controls for the outcome measure. For the PRS, we obtain an

estimate of the linear relationship with PRS and an estimate of the relationship between PRS and the dispersion of the outcome measure.

Permutation: Due to the computational demand of the vertex-wise analysis (~5 minutes pr. data chunk, one permutation totals 328 chunks) we ran 600 permutations for each model (case-control, polygenic risk) before fitting a generalized Pareto distribution to the tail of the permutation distribution⁴⁷. For transparency both the raw and fitted permuted p-values are provided.

Outliers: Participants with a z-score for either eTIV or for volume of interest at $|z| > 3$ were removed, as well as participants with extreme values based on a statistical outlier-test (Rpackage: car⁴⁹, corrected $p < .05$ for the linear models $Y \sim Age + Sex + SZ$ and $Y \sim Age + Sex + PRS$).

PRS-PCA: PCA was performed on SZ-PRS scores from p-thresholds $p < .001$ to $p < .05$ (500 thresholds, steps of .001), before extracting subject loadings from the first three principal components for associations with cortical thickness. The first three principal components explained 92.5%, 3.1% and 1.8% of the variance, respectively (eFigure 1A). The first component (PRS-PC1) showed high correlation with PRS across all thresholds, but relatively lower correlations for the PRS calculated at the stricter p-thresholds compared to the PRS calculated using more lenient p-thresholds. Component 2 (PRS-PC2) correlated negatively with PRS from the stricter p-thresholds, and positive with more lenient p-thresholds. Component 3 (PRS-PC3) was positively correlated with scores calculated with the strictest and most lenient p-thresholds, while correlating negatively with PRS based on the intermediate p-thresholds (eFigure 1B).