

1 **The algae community in taxon Haptophyceae at the early bloom stage of**
2 ***Phaeocystis globosa* in Northern Beibu Gulf in winter**

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19 **Abstract:**

20 *Phaeocystis globosa* (Order Phaeocystales, family Phaeocystaceae) caused significant impact on
21 aquaculture farming, global climate change and industry. Since the year 2010, intensive red tides
22 of *P. globosa* began to appear in Beibu Gulf, where previously free of harmful algal blooms, and
23 posed great threats to the cooling systems of a nuclear power plant in 2014 and 2015. In order to
24 discover the bloom mechanism, the community structure of marine microalgae, with a focus on
25 Haptophyceae taxa, in winter in the northern Beibu Gulf near the Qinzhou Bay, Sanniang Bay
26 (SNB) and Dafenjiang River Estuary (DRE), were explored via 18S ribosomal DNA analysis of the
27 V4 region using the Illumina-Based Sequencing platform. The correlation between the relative
28 abundance of five kinds of Haptophyceae algae and environmental factors of seawater were

29 analyzed. The most abundant Haptophyceae-related OTU in terms of number of reads was
30 identified as *Phaeocystis* and *Chrysochromulina*. The abundance for other Haptophyceae class
31 was relatively low, such as *Haptolina*, *Prymnesium* and *Isochrysis*. *Phaeocystis* was present in all
32 samples sites except S6, S11, S12, S14 and S15, and particularly abundant at S8, nearly 29 times
33 more than the second most abundant site. Most notably, the results showed that *Phaeocystis*
34 displayed highly positive linear correlation with the concentration of NO_3^- -N (Pearson $r=0.856$,
35 $p<0.01$). Linear regression analysis indicated that *Phaeocystis* was significantly linearly related to
36 the NO_3^- -N ($R^2=0.732$; $Y=-0.005 + 0.410 \cdot X$, Y is the relative abundance of *P.globosa*, X is the
37 concentration of NO_3^- -N; $F=38.227$, $P<0.05$) and NO_3^- -N has a significant positive effect on
38 *P.globosa* (regression coefficient is 0.410, $P=0.000$). Moreover, the relative abundance of
39 *Phaeocystis* was significant related to temperature of sea water (Pearson $r=-0.882$, $p<0.01$).
40 Water temperature can explain the 77.8% change reason for the *P.globosa* ($R^2=0.778$), and has a
41 significant effect on *P. globosa* ($Y=0.169-0.009 \cdot X$, $F=49.031$, $P<0.05$), and the regression
42 coefficient is -0.009 ($P=0.000$) which indicated a significant negative impact relationship between
43 them. Our high throughput sequencing (HTS) based research illustrated how the *P. globosa*
44 bloom generated and its relationship with NO_3^- -N and temperature of sea water in northern
45 Beibu Gulf for the first time, and bringing hope for solving this big problem.

46

47 **Key Words:**

48 *Phaeocystis globosa*, high throughput sequencing, nitrate, temperature, Haptophyceae, Beibu
49 Gulf

50

51 **Introduction:**

52 Haptophyceae (or Prymnesiophyceae), a class of the phylum Chrysophyta, contained the Order
53 Prymnesiales, Discoasterales, Phaeocystales, Isochrysidales. The bloom of some Haptophyceae
54 algae had occurred frequently worldwide and led to great ecological disaster and substantial
55 economic losses. For instance, *Prymnesium parvum*, a species of Haptophyceae algae (Order

56 Prymnesiales, family Prymnesiaceae), is capable of producing a toxin, prymnesin, and kills fish
57 [1]. Similarly, large area bloom of the *Chrysochromulina polylepis* (Order Prymnesiales, family
58 Prymnesiaceae) have resulted in mortality of trout and salmon in Scandinavian waters during
59 Spring 1988 [2]. Further more, the bloom of *Phaeocystis globosa* (Order Phaeocystales, family
60 Phaeocystaceae) caused mortality of cultured fish [3], Mussel Mortalities [4], higher
61 concentration of DMS [5] and Clogging of Cooling System of Power Plant [6], so both significantly
62 impact aquaculture farming, global climate change and industry.

63 Beibu Gulf is an important habitat for protection and propagation of marine organisms,
64 known for Indo-Pacific humpback dolphins [7], horseshoe crabs [8], and also an ecologically
65 sensitive region [9]. Recently, rapid economic development and human activities had already
66 resulted in great degradation of marine environment [10,11]. Most noticeably, human-induced
67 nutrient enrichment is becoming a serious problem for coastal marine areas of Beibu Gulf [9,12].
68 Since the year 2010, intensive red tides of *P. globosa* began to appear in Beibu Gulf, where
69 previously free of harmful algal blooms, and posed great threats to the cooling systems of a
70 nuclear power plant in 2014 and 2015 [6]. Up to now, knowledge on where the *P. globosa*
71 originated and the bloom mechanism is still quite limited. In our opinion, in order to solve the
72 great confusion above, special attention should be paid to the early bloom stage and the
73 environmental characteristics leading to the bloom. For this purpose, many methods had been
74 utilized, such as automatic monitoring buoy [13], satellite remote sensing [14], “molecular
75 probes” [15], methods to detect and quantify toxins [16] and a combination of methods above
76 [17].

77 Environmental DNA-based Techniques (EDT) was utilized in monitoring Prokaryote and
78 Eukaryotes in water environments for many years and significantly gained impetus over
79 traditional approaches presently [18]. Recently, there are many EDT methods advented for this
80 object; for instance, DNA metabarcoding [19,20], Microsatellite DNA marker [21], and high
81 throughput sequencing (HTS) techniques based metagenomics [22,23] and amplicons [24,25]. In
82 recent years, the HTS techniques has been widely used and remarkably promoted the ecological
83 studies of bacteria [26], fungi [27], algae [28] and animals [29]. However, to date, molecular
84 diversity of microeukaryotes, including algae, in the Beibu Gulf marine region, remains

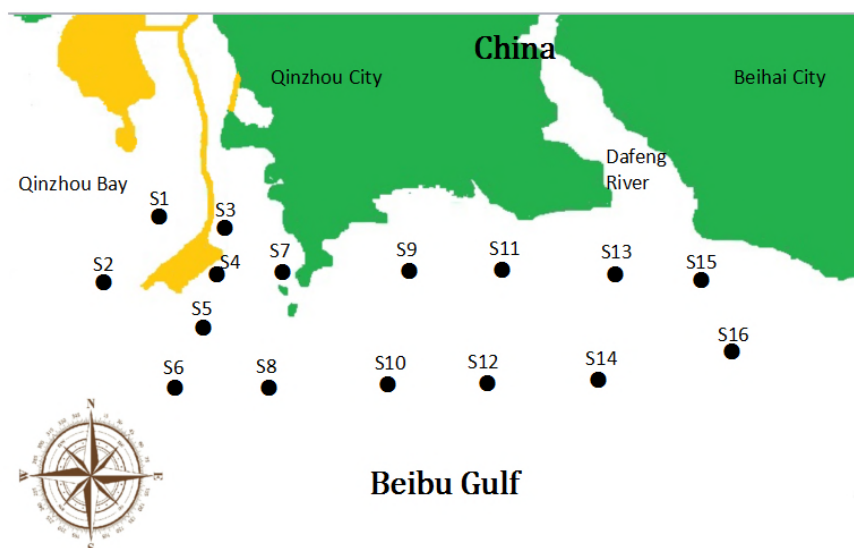
85 unexplored. Thus, the recent study about the community and diversity structure of marine
86 microalgae, with a focus on Haptophyceae taxa in winter in the northern Beibu Gulf near the
87 Qinzhou Bay, Sanniang Bay (SNB) and Dafenjiang River Estuary (DRE), were explored via 18S
88 ribosomal DNA analysis of the V4 region using the Illumina-Based Sequencing platform. This
89 study provides a valid taxonomic reference dataset for future microeukaryotic community
90 structure and diversity studies, aimed at monitoring environmental change in the northern Beibu
91 Gulf.

92

93 **Materials and Methods**

94 **Sample Collection**

95 The samples were collected on 27 Dec 2017, when large *Phaeocystis* bloom was observed
96 between eastern Qinzhou Bay (EQB) and Dafenjiang River Estuary (DRE), during the middle
97 several days of Jan to the end of Mar in 2018. Seawater was collected from surface, middle and
98 deep using a CTD Rosette Water Sampling System (Sea-Bird Electronics, USA) between eastern
99 Qinzhou Bay (EQB) and Dafenjiang River Estuary (DRE) (21°32'58''N, 108°39'56''E-21°37'39''N,
100 108°55'57''E) and mixed evenly. A total of 16 seawater samples (S1-S16, Figure 1) were collected
101 in 3L sterile polyethylene bottles, kept in the dark at 4-8 °C, and filtered at laboratory within 4h.
102 Each 3000 ml seawater sample was filtered through a 1.2 µm mixed cellulose membrane filter
103 (Advantec, Japan) and filtrates were frozen immediately at -20°C for subsequent molecular
104 analyses.



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106 Figure 1. Locations of the the 16 sites (S1-S16) where seawater samples were collected on 27 Dec
107 2017, including six sites near Sandun Dock (S1-S6), six sites near Sanniang Bay (SNB, S7-S12) and
108 four sites in the Dafengjiang River Estuary (DRE,S13-S16).

109 **DNA Extraction, PCR and illumina sequencing**

110 The PowerWater DNA isolation kit (MoBio Laboratories Inc., CA, USA) was used to extract the
111 DNA of the total organisms on the 1.2 μm filters following the manufacturer's protocol. The DNA
112 samples were detected by 1% agarose gels and NanoDrop One spectrophotometer (Thermo
113 Fischer Scientific Inc., USA), and then were amplified using the primers 528F and 706R [30,31]
114 which was designed to amplify the hypervariable region V4 of eukaryote 18S rRNA gene. Illumina
115 sequencing was carried out by the Novogene Company (Beijing, China). Sequencing libraries
116 were generated using TruSeq[®] DNA PCR-Free Sample Preparation Kit (Illumina, USA) following
117 manufacturer's recommendations and index codes were added. The library quality was assessed
118 on the Qubit@ 2.0 Fluorometer (Thermo Scientific) and Agilent Bioanalyzer 2100 system. At last,
119 the library was sequenced on an Illumina HiSeq2500 platform and 250 bp paired-end reads were
120 generated. The raw data sequences were assigned to samples by their unique barcodes. The 18S
121 rDNA primers and barcodes were cut off to generate pair-end (PE) reads. Paired-end reads were
122 merged using FLASH (V1.2.7) [32]; the raw tags were filtered to obtain the high-quality clean tags
123 using QIIME software package (V1.7.0) [33]. Sequences analysis were performed by Uparse
124 software (Uparse v7.0.1001) [34]. Sequences with $\geq 97\%$ similarity were assigned to the same

125 OTUs. For each representative sequence, the GreenGene Database [35] was used to annotate
126 taxonomic information.

127 **Physical and chemical analyses of seawater characteristics**

128 The seawater temperature ($^{\circ}\text{C}$), salinity were measured using the SBE 911 plus CTD (Sea-Bird
129 Electronics, USA). Dissolved oxygen (DO) concentrations (ml/l) were measured using the SeaBird
130 43 (Sea-Bird Electronics, USA). The pH values were measured by pH meter (METTLER, FE38-
131 Meter, 30254110). Chlorophyll-a fluorescence was measured using the WETStar (WET Labs,
132 USA). Inorganic nutrient concentrations (nitrate [NO_3^- -N], ammonia [NH_4^+ -N], nitrite [NO_2^-], and
133 phosphate [PO_4^{3-}]) were determined from 100 mL samples with an Alliance Integral Futura
134 Autoanalyzer II [36,37]. The TOC content was determined with TOC analyzer (Multi N/C 3100,
135 Analytik Jena AG, Jena, Germany) according to the procedure explained by Ali [38]. Spatial
136 distribution of seawater characteristics was interpolated by the measurements of the 16
137 sampling sites using Kriging method [39]. Interpolations outside the sampling area and over the
138 terrestrial landscape were subtracted.

139 **Statistical analysis**

140 Data were compiled and transformed in Microsoft Excel. Correlation between variables were
141 made using a linear Pearson's r coefficient. Linear regression analysis was facilitated and
142 conducted between closely related. Statistics were generated using the SigmaStat version 2.01
143 software package (SPSS, Inc., Chicago, Ill). All comparisons were performed at the 95%
144 confidence level.

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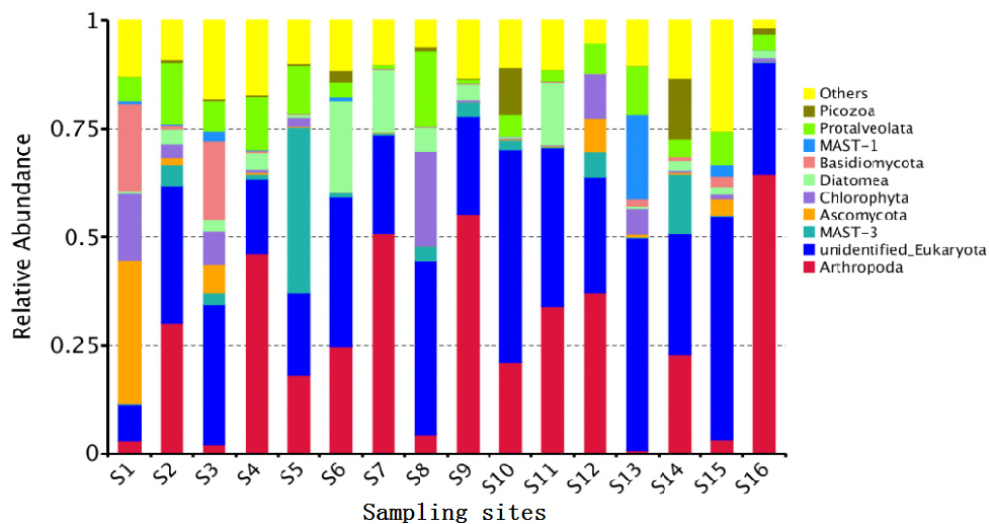
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147 **Result**

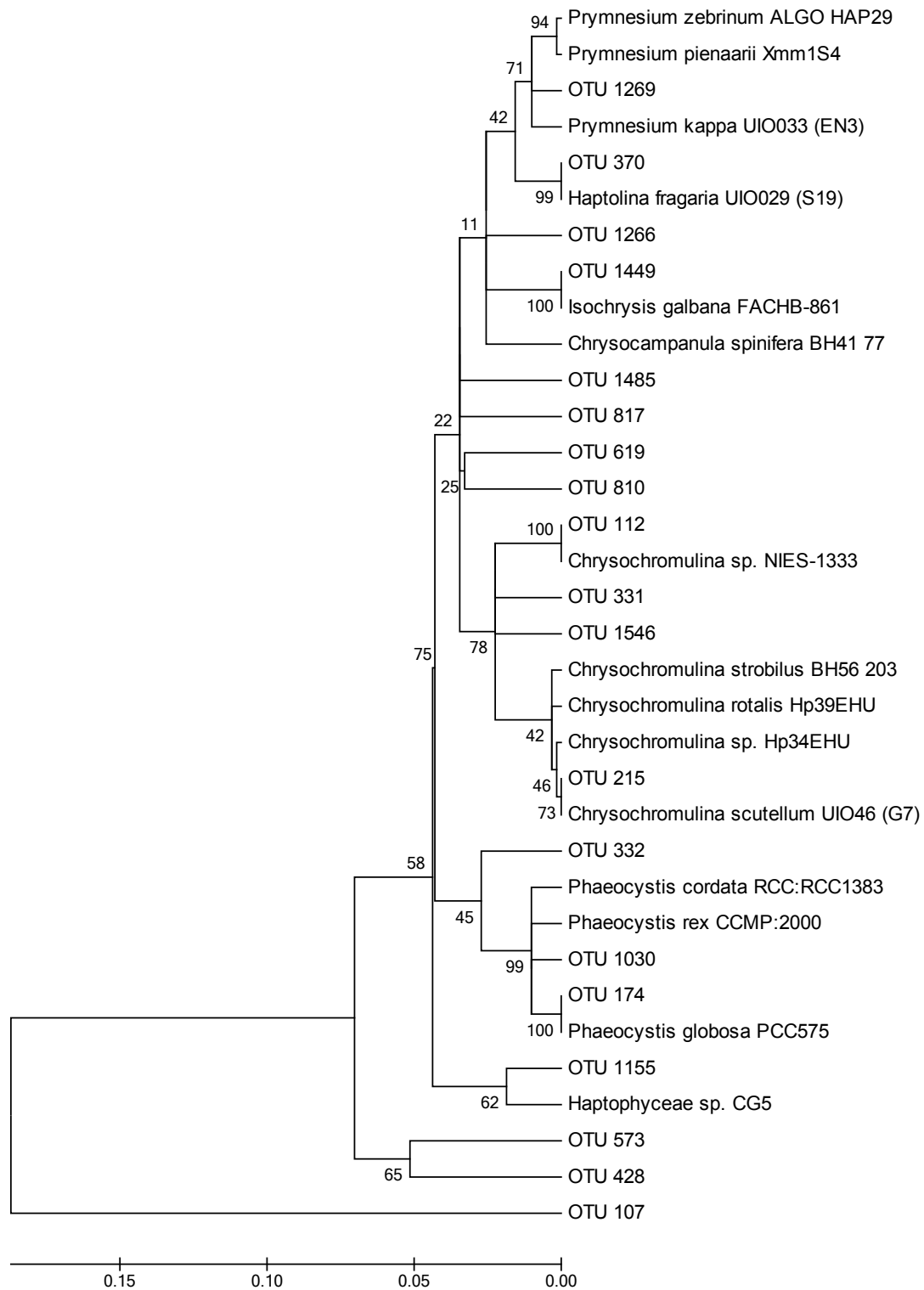
148 **OTUs profile based on 18s RNA amplicon analysis using high throughput sequencing (HTS)**

149 The eukaryote communities in the seawater were represented by 1,594 OTUs identified in our

150 present study. A good coverage of the eukaryotic diversity in all samples were illustrated by
151 rarefaction curves, which were calculated and reached a plateau in all cases. All OTUs were
152 classified into 44 main Phylum, and the most abundant were Arthropoda, Ascomycota,
153 Chlorophyta, Diatomea, Basidiomycota, Protalveolata and Picozoa (Fig 2). Among all the OTUs,
154 our most concerned OTU was that identified as class Haptophyceae, which some Harmful algae
155 (such as *P.globosa* and *Prymnesium*) causing serious ecological disasters represented in this
156 group. In our study, 19 OTUs were classified into Haptophyceae, belonging to the groups of
157 *Prymnesium*, *Haptolina*, *Isochrysis*, *Chrysochromulina* and *Phaeocystis* respectively (Fig 3).
158 *Phaeocystis* was represented by OTU 174 and OTU 1030, with OTU 174 identified as *P. globosa*
159 (100% similarity). *Prymnesium* was represented only by one OTU (OTU 1269), which closely
160 related to *Prymnesium zebrinum* (98% similarity) and *Prymnesium pienaarrii* (97% similarity). The
161 most biggest taxonomic group in Haptophyceae was identified as genus *Chrysochromulina* which
162 including 4 OTUs. The other taxonomic groups belonging to Haptophyceae were *Haptolina*
163 *fragaria* (100% similarity, OTU370), *Isochrysis galbana* (100% similarity, OTU1449),
164 *Haptophyceae* sp. (96% similarity, OTU1155) (Fig 3).



166 Figure 2. The top ten OTUs were identified based on the hypervariable region V4 of eukaryote
167 18S rRNA gene and sequences with $\geq 97\%$ similarity were assigned to the same OTUs.

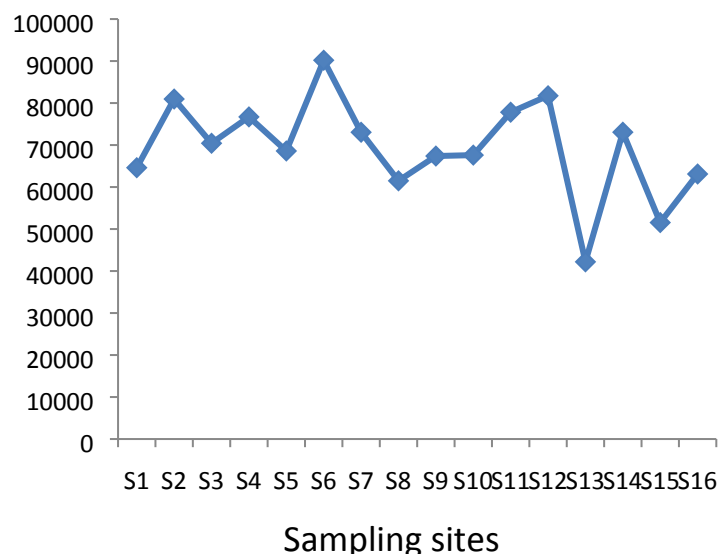


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169 Figure 3. The taxonomic relationship of 18S rDNA phylotypes of class Haptophyceae from the the
170 northern coast of Beibu Gulf. Kimura two-parameter model and midpoint rooting options were
171 used to reconstruct phylogenetic relationships. Numbers above branches indicate bootstraps for
172 NJ analysis (> 50).

173

174 The most abundant Haptophyceae-related OTU in terms of number of reads was identified
175 as *Phaeocystis* and *Chrysochromulina*. *Chrysochromulina* was the most abundant taxa containing
176 5353 reads, while *Phaeocystis* displayed a relatively lower number of 2298 reads. The abundance
177 for other Haptophyceae class was relatively low, such as *Haptolina* (228 reads), *Prymnesium* (9
178 reads), *Isochrysis* (6 reads) (Fig 4). The relative abundance of the five kinds of Haptophyceae at
179 different sampling sites showed different characteristics. The spatial distribution of *Phaeocystis*
180 and *Prymnesium* were similar. *Phaeocystis* was present in all samples sites except S6, S11, S12,
181 S14 and S15, and particularly abundant at S8 (2150 reads, 3.4%), nearly 29 times more than the
182 second most abundant site S2 (73 reads). *Prymnesium* showed the same pattern, recording the
183 highest percentage at site S8 (7 reads, 0.01%) and haven't appeared on the other sites. While
184 *Isochrysis* indicated very different property, with major peaks occurred at the sites S3 (5 reads)
185 and none at the other sites. Most of algal species *Chrysochromulina* were abundant and fairly
186 distributed at different stations with reads ranged from 158 to 899, except highest percentage
187 (2.05 %, Site 8) and relatively lower at S6 (0%), S7 (19 reads, 0.03%), S9 (46 reads, 0.07%), S11 (30
188 reads, 0.04%), S12 (0%) and S16 (54 reads, 0.08%). The relative abundance of *Haptolina* among
189 different sites displayed similar pattern with *Chrysochromulina*, and the major peaks occurred at
190 sites S8 (Fig 5).

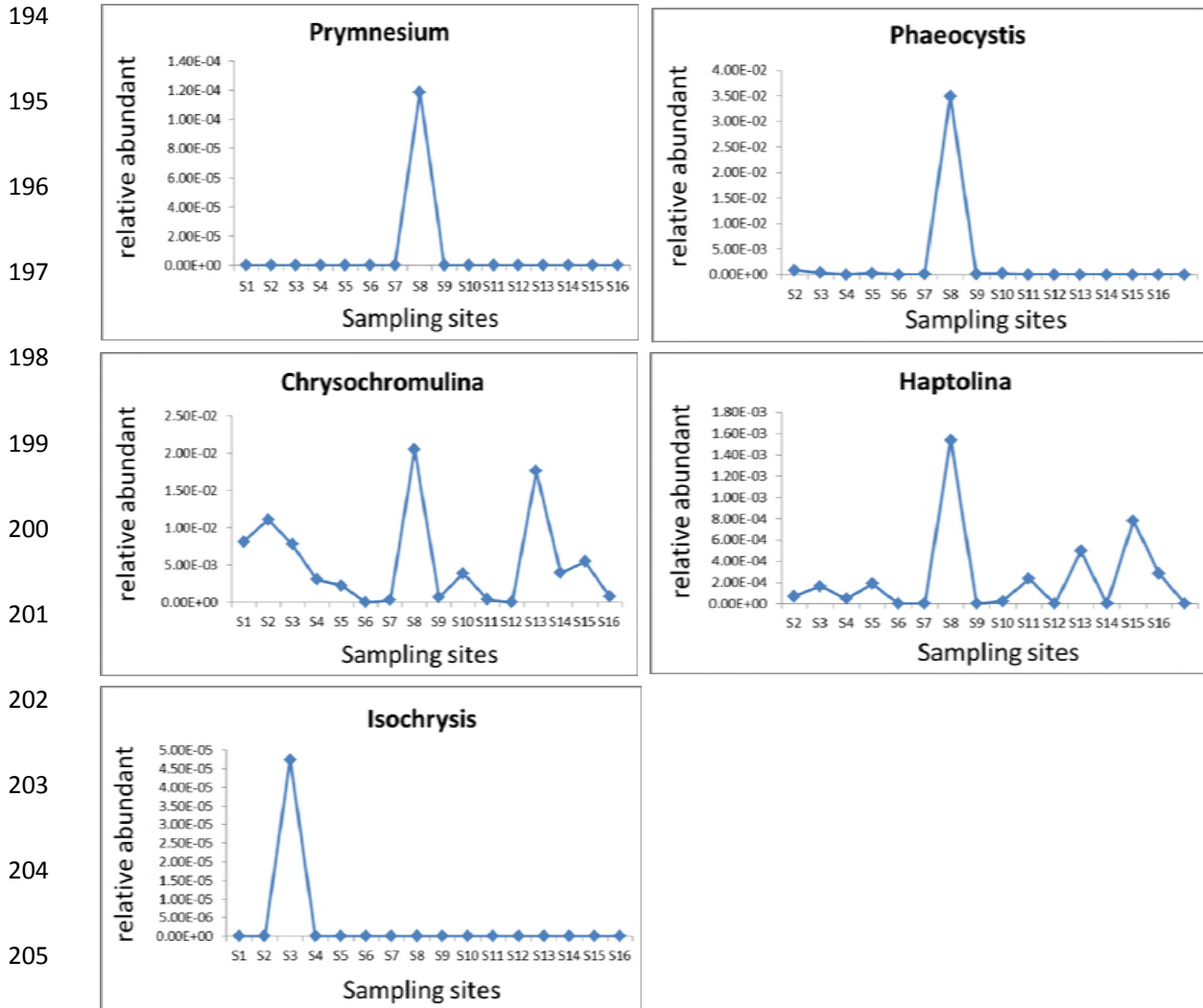


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192 Figure 4. The number of reads detected in 16 sampling sites using 18s RNA amplicon analysis.

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211 Figure 5. Spacial variations of relative abundance (%) of five Haptophyceae algae

212 (Prymnesium, Haptolina, Isochrysis, Chrysochromulina and Phaeocystis) at the different

213 stations in the northern coast of Beibu Gulf in Dec 2017.

214

215 **Physical and chemical analyses of seawater characteristics**

216 Physical and chemical characteristics of the seawater in the 16 sampling sites, including nutrient
217 concentrations (TOC, $\text{NH}_4^+\text{-N}$, $\text{NO}_3^-\text{-N}$, $\text{NO}_2^-\text{-N}$, and phosphate [PO_4^{3-}]) and other environmental
218 condition (chlorophyll-a, DO, salinity, pH, temperature of seawater), were detected. Just as our
219 above discovery that relative abundance of some Haptophyceae algae were obviously higher at
220 site S8 than others, some environmental factors at S8 were apparently special as well. Seawater
221 temperature during the period of the study ranged from 15.4 to 18.5 °C and the lowest
222 temperature appeared at site S8, while the salinity ranged from 25.7 to 37.3 ppt and S8
223 contained the lowest value 25.69 ppt. The highest value recorded for DO was 9.32 mg/L at site
224 S8 and lowest was 8.33 mg/L at site S13. The highest value of pH was 8.71 at site 12 and lowest
225 was 8.22 at S8. For the nutrient concentrations, the site S8 possessed the highest concentration
226 of $\text{NO}_3^-\text{-N}$ (0.066 mg/L). Spatial distribution of seawater characteristics over the study region,
227 including DO, $\text{NO}_3^-\text{-N}$, temperature of seawater and pH, were simply illustrated on the map by
228 using blue and red colour (Figure 6).

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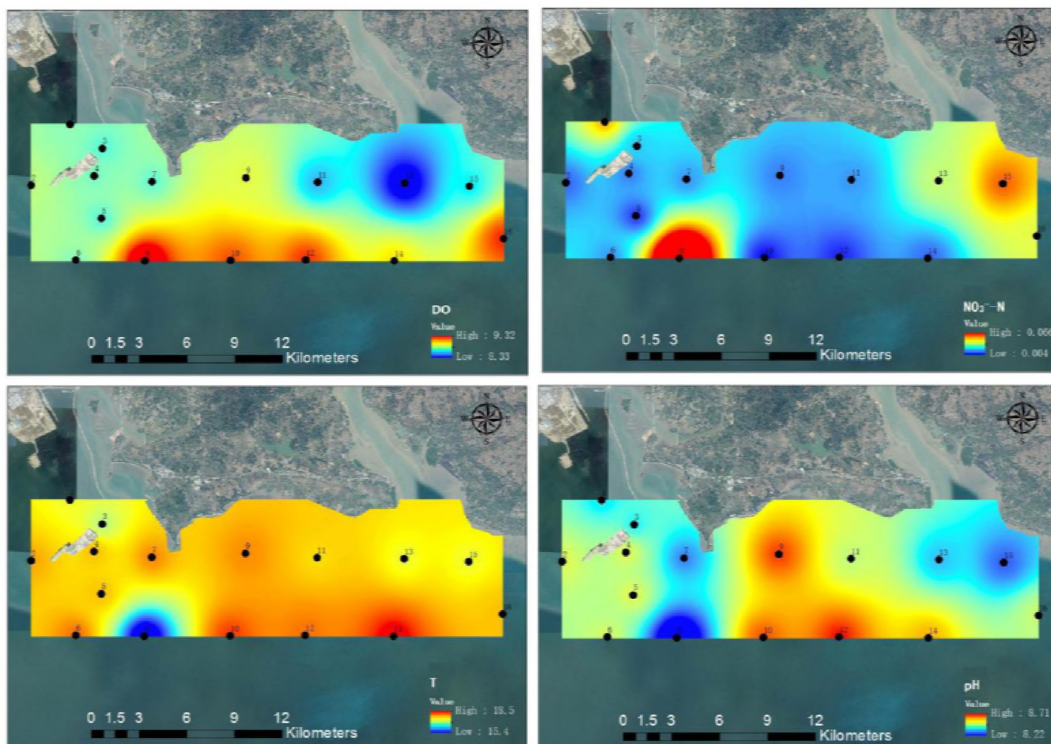
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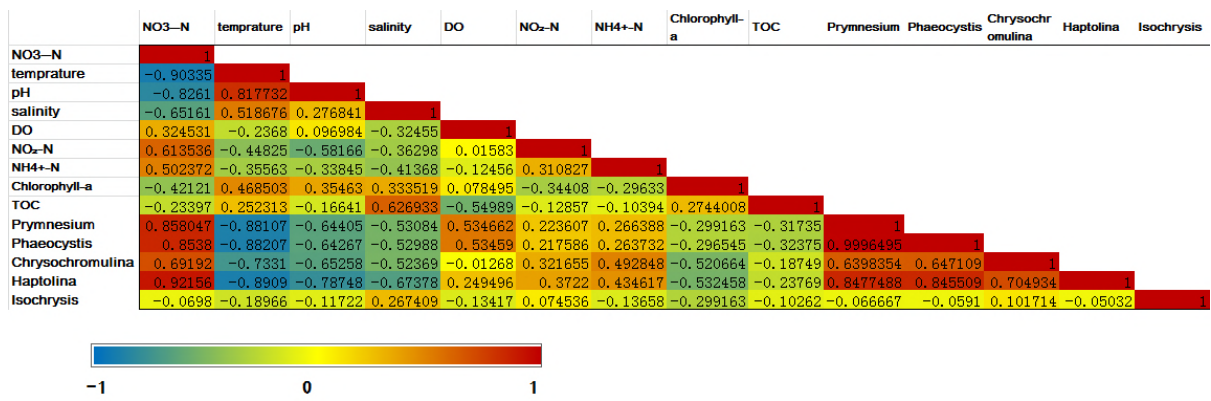
242 Figure 6. Spatial distribution of seawater characteristics over the study region, including DO (a),
243 NO_3^- -N (b), T-temperature of seawater (c) and pH (d).

244

245 **The Correlation between environmental factor and Haptophyceae algae**

246 The correlation between the relative abundance of five kinds of Haptophyceae algae and
247 environmental factors of seawater (temperature, pH, salinity, DO, NO_3^- -N, NO_2^- -N, NH_4^+ -N,
248 Chlorophyll a and TOC) were analyzed. The results showed that Pymnesium, Phaeocystis and
249 Haptolina displayed highly positive linear correlation with the concentration of NO_3^- -N (Pearson
250 $r=0.85\sim 0.92$, $p<0.01$). Isochrysis and Chrysochromulina indicated high negative correlation with
251 temperature of sea water (Pearson $r=-0.88\sim -0.89$, $p<0.01$). Except for this, there was a significant
252 negative correlation just for Pymnesium, Haptolina, Chrysochromulina and Phaeocystis with pH
253 and salinity (Pearson $r=-0.52\sim -0.78$, $p<0.01$). Chrysochromulina revealed significant negative
254 correlation with seawater temperature (Pearson $r=-0.787$, $p<0.01$). Haptolina and
255 Chrysochromulina also exhibited significant negative correlation with Chlorophyll-a (Pearson
256 $r=-0.52\sim -0.53$, $p<0.01$). There was a significant positive correlation of Pymnesium and
257 Phaeocystis with DO (Pearson $r=0.534$, $p<0.01$), and also the same relationship of
258 Chrysochromulina with NO_3^- -N (Pearson $r=0.69$, $p<0.01$). There was no other obvious correlation
259 between other environmental factors and those five kinds of Haptophyceae algae (Fig 7).

260



261

262 Figure 7. The correlation between the relative abundance of five kinds of Haptophyceae algae
 263 and environmental factor (NO₃⁻-N, temprature, H, salinity, DO, NO₂⁻-N, NH₄⁺-N, Chlorophyll a
 264 and TOC) was indicated by Correlation matrix (Pearson's product moment correlation
 265 coefficient). The more the colure of shaded cells was close to red, the more significantly the
 266 correlation was positively related (P < 0.05); inversely, the more the colure of shaded cells was
 267 close to blue, the more significantly the correlation was negatively related (P < 0.05).

268

269 Chrysochromulina was the most abundant taxon of Haptophyceae algae in this study. The
 270 environmental factors NO₃⁻-N, pH and seawater temperature on the relative abundance of
 271 Chrysochromulina were studied by regression analysis. The concentration of NO₃⁻-N possessed
 272 significant positive impact on the relative abundance of Chrysochromulina (Pearson r=0.692,
 273 p<0.01; R²=0.479; Y=-0.001 + 0.281*X, Y is the relative abundance of Chrysochromulina, X is the
 274 concentration of NO₃⁻-N; F=12.859, P<0.05; regression coefficient is 0.281, P=0.003). Conversely,
 275 the temperature of seawater indicated obvious negative influence on the relative abundance of
 276 Chrysochromulina (Pearson r=-0.733, p<0.01; R²=0.537; Y=0.123-0.007*X, Y is the relative
 277 abundance of Chrysochromulina, X is the value of seawater temperature; F=16.266, P<0.05;
 278 regression coefficient is -0.007, P=0.001), and pH showed the same features on it (Pearson
 279 r=-0.653, p<0.01; R²=0.426; Y=0.282-0.032*X, Y is the relative abundance of Chrysochromulina, X
 280 is the value of seawater pH; F=10.384, P<0.05; regression coefficient is -0.032, P=0.006).

281 There is consistent relation between the relative abundance of Phaeocystis and
 282 environmental factor NO₃⁻-N, DO, pH and temperature of sea water. The relative abundance of

283 Phaeocystis has obvious correlation with NO_3^- -N (Pearson $r=0.856$, $p<0.01$). Linear regression
284 analysis indicated that Phaeocystis was significantly linearly related to the NO_3^- -N ($R^2=0.732$;
285 $Y=-0.005 + 0.410*X$, Y is the relative abundance of *P.globosa*, X is the concentration of NO_3^- -N;
286 $F=38.227$, $P<0.05$) and NO_3^- -N has a significant positive effect on *P.globosa* (regression
287 coefficient is 0.410, $P=0.000$). The relative abundance of Phaeocystis had a positive correlation
288 with DO (Pearson $r=0.535$, $p<0.01$; $R^2=0.286$; $Y=-0.140 + 0.016*DO$, $F=5.616$, $P<0.05$; regression
289 coefficient is 0.016, $P<0.05$). The relative abundance of Phaeocystis was significant related to
290 temperature of sea water (Pearson $r=-0.882$, $p<0.01$). Water temperature can explain the 77.8%
291 change reason for the *P.globosa* ($R^2=0.778$), and has a significant effect on *P. globosa*
292 ($Y=0.169-0.009*X$, $F=49.031$, $P<0.05$), and the regression coefficient is -0.009 ($P=0.000$) which
293 indicated a significant negative impact relationship between them. The relative abundance of
294 Phaeocystis also has negative correlation with pH of sea water (Pearson $r=-0.643$, $p<0.01$).

295 Prymnesium exhibited very similar condition with Phaeocystis, concerning relationship
296 between their relative abundance and environmental characteristics. For example, the relative
297 abundance of Prymnesium showed highly correlation with NO_3^- -N (Pearson $r=0.858$, $p<0.01$)
298 and NO_3^- -N had a significant positive impact on the relative abundance of Prymnesium
299 ($R^2=0.736$; $Y=-0.000 + 0.002*X$, Y is the relative abundance of Prymnesium, X is the concentration
300 of NO_3^- -N; $F=39.079$, $P<0.05$; regression coefficient is 0.002, $P=0.000$). In contrast, the
301 temprature of seawater exhibited obvious negative effect on Prymnesium ($R^2=0.776$; $Y=0.001 -$
302 $0.000*X$, Y is the relative abundance of Prymnesium, X is the value of temperature of seawater;
303 $F=48.579$, $P<0.05$; regression coefficient is - 0.000, $P=0.000$).

304

305 **Discussion**

306 **NGS: a promising approach to study the community of algae in marine environments**

307 *P.globosa* bloom occurred in winter from the year of 2014-2015 near the coast of Beibu Gulf
308 annually. In order to discover the bloom mechanism, *P. globosa* was monitored between eastern
309 Qinzhou Bay (EQB) and Dafenjiang River Estuary (DRE) during Nov 15th 2017 to Feb 15th 2018.

310 The beginning of the *P. globosa* bloom appeared from the middle several days of Jan in 2018,
311 and disappearing occurred at the end of Mar in 2018. All the above findings were based on
312 microscopy observations and cell counts, and it seems that the bloom appeared suddenly and
313 instantly. We believed that finding out what happened before the emergence of *P. globosa*, not
314 only by observations of microscopy and naked eyes, was very significant for our understanding of
315 bloom mechanism of *P. globosa* in Beibu Gulf.

316 Classically, the algae ecological value is weighted based on the relative abundance of
317 morphologically identified species. This traditional method is costly, time-consuming, and
318 requires excellent taxonomic expertise, which is not always available [40]. Comparatively
319 speaking, the eDNA and NGS approach for identification and quantification of algae open a new
320 avenues for assessing and monitoring of aquatic ecosystems [41]. In our present work, by
321 utilizing NGS based eDNA detection, not only the *P. globosa*, but also all the algae species
322 belonging to Haptophyceae, taxonomically including *P. globosa*, were discovered and analyzed.
323 The two most abundant OTUs in taxon Haptophyceae were affiliated with *Chrysochromulina*
324 (5353 reads) and *Phaeocystis* (2298 reads), significantly more reads than *Haptolina* (228 reads),
325 *Prymnesium* (9 reads) and *Isochrysis* (6 reads). The genus *Chrysochromulina* include two species
326 *Chrysochromulina scutellum* and *Chrysochromulina* sp., both not very clear about their
327 significance in ecosystem. But for the genus *Phaeocystis*, *P. globosa* was obviously detected
328 among the samples, and this was in accord with our traditional monitoring approaches for the
329 findings of *P.globosa* bloom in Beibu Gulf. Our result convincingly implicated that algae
330 community reflected by eDNA and 18S ribosomal DNA analysis of the V4 region using the
331 Illumina-Based Sequencing platform was suitable for monitoring the harmful algae in the early
332 bloom stage. Many scientist had utilized HTS or NGS to detect algae in aquatic ecosystems and
333 got some attractive results; for instance, NGS had been employed to microalgal diversity in the
334 lichen *Ramalina farinacea* [42], Diatom resting stages in surface sediments [43], Diatom
335 biomonitoring [40] and detection of harmful algal bloom species [44]. All the results show that
336 eDNA and HTS sequencing is a promising approach to explore the community of algae in aquatic
337 environments [45].

338 **Hypothesis about the bloom mechanism of *P.globosa* in Beibu Gulf**

339 Some mechanisms about the bloom of *P. globosa* could be proposed from our present work.

340 The first and most interesting finding was the bloom mode of *P. globosa*. In this study, at the
341 early bloom stage, *P. globosa* was only obviously detected at site S8 with relatively much higher
342 reads (2150) than other sixteen sites (148 in all). Therefore, the bloom of *P. globosa* may
343 originate from a point of site (S8) and then spread to other regions, generally speaking, just as
344 “diffusion from point to face”. Several evidences supported our opinion. Firstly, the marine
345 environment in the Beibu Gulf was protected relatively better than other coastal zones in China
346 [46]. Large area pollution, especially excessive nutrient concentration, had never appeared and
347 been reported previously. Inversely, the probability of point source pollution in the coastal zone
348 was even greater. The first appearance and flourish of *P. globosa* only at site S8 was probably
349 because of its special condition, such as aquaculture farming nearby and consequent
350 eutrophication phenomenon, rich nutrients brought by bottom-up stream and terrestrial
351 drainage. Then the Phaeocystis was carried by transport of some water and drifted along the
352 coastline under the influence of stream in Beibu Gulf. Some previous research could support our
353 opinion. For instance, during the Phaeocystis bloom of the year 1957 around the coast of North
354 Wales, Phaeocystis was only able to proliferate in Liverpool Bay, and then spread to other regions
355 [47]; when Phaeocystis bloom in the coastal of north-western English Channel in 1990,
356 Phaeocystis bloom emanated from near-shore, then spread towards the south-east in accord
357 with the wind direction [48]. If our hypothesis is correct, releasing effective bio-agents and
358 environmental controls on certain region to inhibit harmful algal blooms (HABs) *P. globosa* will
359 be promising [49-51]. In this way, we only need to paid more attention to the early stage of *P.*
360 *globosa* bloom, not after outbreaks in large scale, and this should be extremely effective.

361 The second interesting finding about the bloom of *P. globosa* was in relevant to the
362 concentration of NO_3^- -N, pH, temperature and DO of seawater. More specifically, the bloom of *P.*
363 *globosa* has a significant positive correlation with NO_3^- -N and negatively related to temperature
364 of seawater. In the Beibu Gulf, the average temperature of seawater was above 15-16 °C and
365 the most suitable temperature for growth of *P. globosa* is 15-16 °C [52], so the most relative
366 abundance of *P. globosa* appeared first at site S8 may originate from its appropriate temperature
367 of seawater. Nutrient elements, especially nitrogen and phosphorus, have obvious influence on

368 Phaeocystis blooms [52,53]. However, the impact of different forms of N-sources to the bloom of
369 Phaeocystis had never been discovered previously. In our present work, it was the first time that
370 the great correlation between NO_3^- -N and Phaeocystis bloom was illustrated. This result provided
371 a helpful reference to our government on how to manage marine environment and control the
372 *P.globosa* bloom. They should pay more attention to reduce the emission of nitrogen, especially
373 NO_3^- -N, to the coastal zone of northern Beibu Gulf.

374

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380

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391 **Reference:**

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